

Genome variation and population structure in three African malaria vector species within the *Anopheles gambiae* complex

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Abstract

Population Sampling

The third and final phase of the Ag1000g project data resource contains wild-caught *Anopheles* mosquito genomes collected from a total of 124 sites across 19 countries, six of which are novel, in Sub-Saharan Africa.

Collections from Malawi increase the density of coverage in West Africa, Central African Republic and Democratic Republic of Congo begin to fill the gap previously present in Central Africa while Malawi, Mozambique and Tanzania provide much more power to analyse East African malaria vectors.

New quality control and filtering pipelines were developed for this phase, so all 4,693 samples submitted across the three project phases were evaluated with these new methods.

High quality whole genome sequences from 2784 unique individuals passed all sample filters, 2532 females, 223 males and 29 samples with unknown sex (see Methods).

1823 of these wild-caught individuals are novel to phase 3.

181 samples from phase 2 did not pass the new quality thresholds are not included in this data release (@@ either - see Supplementary table OR explain 172 were lost via FILTER_second_rep_hi_skew <- then need to explain what this is?).

Alongside sampling from natural populations, 300 individuals from 15 lab crosses, parents and offspring, have been sequenced.

Four of these crosses are novel to phase 3, adding more power to measure the quality of wild sample variation and to methods validation more generally.

Parents of the new crosses were drawn from colonies where founders were collected in Nagongera (Uganda) and Kisumu (Kenya) .

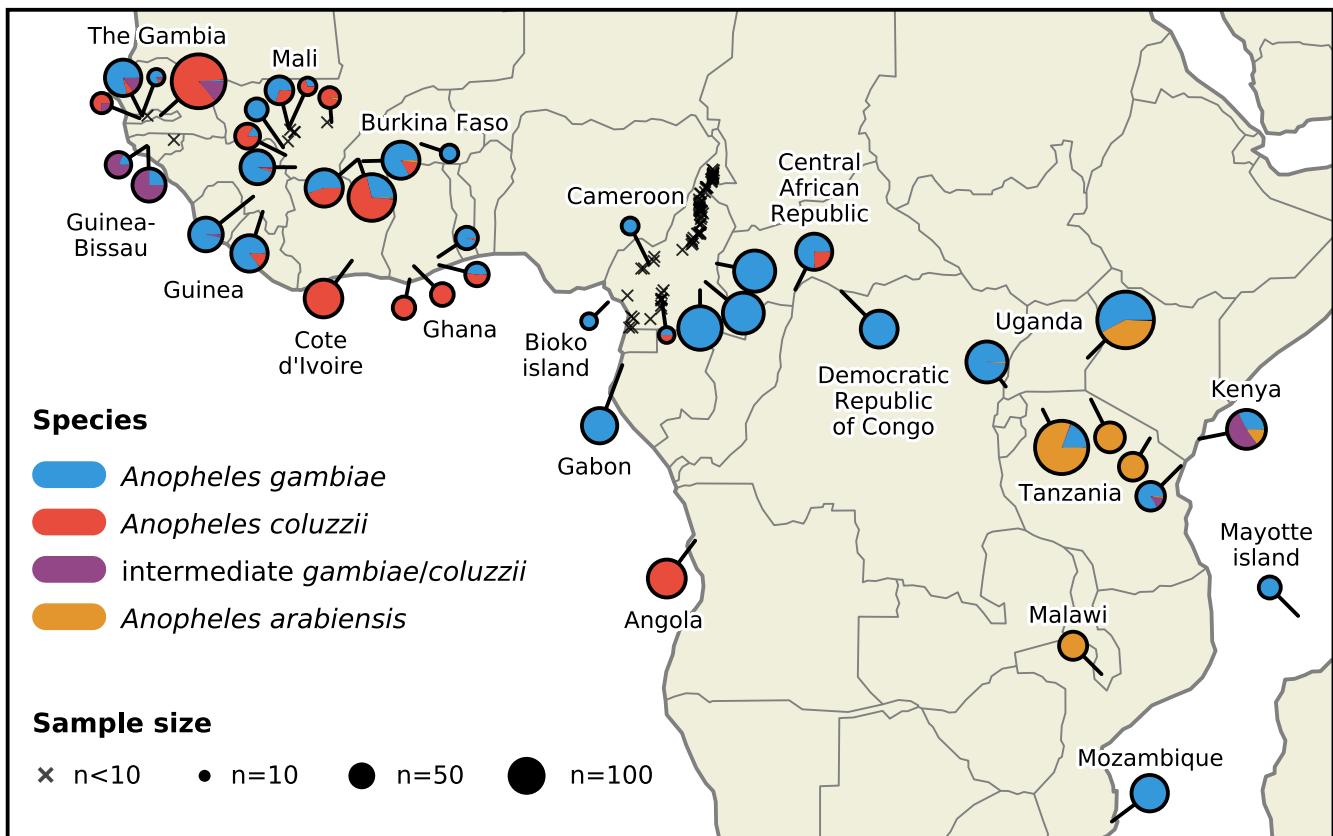


Figure 1: Sample Collection Map

Whole Genome Sequencing and Alignment

4,693 individual mosquitoes were sequenced on either Illumina HiSeq2000 (n=3,130) or Illumina HiSeqX (n=1,563) to a target coverage of 30X.

Between machine types the median number of bases sequenced per sample was 9.76Gb and 10.33Gb respectively, representing a difference in yield (two-tailed mann-whitney U p < 0.0001).

These values correspond to a yield per reference base (vs AgamP4) of 35.76X and 37.82X.

91.9% of HiSeqX runs and 80.5% of HiSeq2000 runs met the target yield of 30X.

Reads were aligned to the AgamP4 reference and Single Nucleotide Polymorphisms (SNPs) called using GATK UnifiedGenotyper.

All samples successfully completed the pipeline and entered the sample quality control (QC) process.

Sample QC

The sample QC process was composed of three stages, sequence quality assurance, replicate handling, and anomaly detection.

668 samples were removed where sequencing was of insufficient quality to accurately call genotypes across the whole genome.

Exclusions were due to poor coverage (n=410), potential contamination (n=229), and an ambiguous sex call (n=29).

Where technical replicates were available, we excluded 4 pairs (8 samples) with low genotype concordance.

Where pairs met the concordance threshold we excluded the lower quality sample (n=407).

Samples were also screened pairwise within submission sets for unexpected pairs, though none were detected.

The AG1000G-X submission set, made up of laboratory experimental crosses, was exempted from this step due to familial similarity and high levels of inbreeding.

The third stage used principal component analysis (PCA) to identify and exclude individual samples that were outliers based on available metadata.

A review process identified samples that could not be explained parsimoniously, and were therefore likely to be sample mix ups or instances of mislabelling.

28 samples were excluded as they respectively dominated the first principal components, indicating high divergence from all other samples and therefore likely members of other Anopheline species.

A further 82 samples were excluded as potential sample mix ups.

Following all sample QC steps, 3,483 samples (74.2%) were retained from the original cohort for analysis.

Coverage

Summary of site coverage post QC exclusions.

Species assignment and sex calling

The *Anopheles gambiae* complex is a cryptic group of sibling species, with no single locus offering unambiguous resolution of species.

To identify species we looked beyond the conventional set of PCR based markers and applied a wider set of ancestry informative markers (AIMs).

Species were not assigned to samples from laboratory colony crosses due to inbreeding and high levels of genetic drift.

To distinguish *A. arabiensis* from *A. gambiae s.l* a set of novel markers was derived from data from the 16 genomes project (ref).

Using cut offs based on agreement with the established PCR marker, 368 individuals were classed as *A. arabiensis* and 2415 as *A. gambiae s.l*.

A single individual collected in Tororo, Uganda is classed as intermediate- given the majority (XX%) of AIM SNPs in the genome are heterozygous between the *gambiae*-like and *arabiensis*-like alleles, this individual is likely to be an F1 hybrid.

To resolve the *A. gambiae s.l*/individuals as *A. gambiae* and *A. coluzzii* we applied 729 AIMs identified by Neafsey et al (ref), and used by us in previous analyses (ref paper2, paper1).

Of the 2415 *A. gambiae s.l*/individuals, 1571 were called as *A. gambiae s.s*, 675 as *A. coluzzii* and 169 as intermediate (ref collection map).

Many intermediate samples were sampled from the Western coast of West Africa (particularly The Gambia and Guinea Bissau), and given distinct populations of *A. gambiae s.l*. and *A. coluzzii* are also found in this region, highlights the complexity of species relationships here.

SNP filtering and quality

Site filtering is necessary to ensure that reported variation is of high quality.

Features of specific regions of the *Anopheles* genome contribute to calling errors in short-read technologies; such features include regions of high divergence from the reference, high homology between regions, copy number variation, or the presence of transposable elements.

Where previously we have used manually curated cutoffs based on observed mendelian error rates to filter sites (ref phase1, phase2), here we built a statistical model where cohort level summary statistics were used to identify sites likely to contain genotyping errors.

Using the 15 available *Anopheles* pedigrees previously described, we used the presence of mendelian error at sites as a proxy for genotype discordance.

10 of the 15 crosses were used to train the model while 5 were held out for validation.

Each of the 5 pedigrees represent independent evaluation sets.

Before applying the site filters, the false discovery rate (FDR) of the 5 crosses over all autosomal sites ranged between 0.74% and 1.10% (table XXX).

The application of the site filters defines the accessible fraction of the genome at 72.58%, and the range of false discovery rates is 0.04% to 0.10%.

The median fold change of FDR was -3.71.

On the hemizygous X chromosome we used the more direct measure of heterozygote calls in males to ascertain mendelian error.

In the dataset are 220 *A. gambiae* s./male samples, each of which represent an independent proxy for genotype discordance.

Pre-application of the site filters, subject to a Genotype Quality (GQ) threshold of 30, the median heterozygosity rate was 0.244%, and post filtering this drops to 0.023% (table XX).

The median fold change in error rate was -3.33, with 69.97% of the X chromosome passing site filters.

The new model based method represents a marked improvement over the site filters generated as part of phase 2.

All 5 evaluation pedigrees showed a small reduction in FDR, but coupled with the lower rate of accessibility in phase 2 (62.05%) resulted in a significant improvement in Youden score (Table XXX) across autosomes.

The X chromosome showed a similar pattern, the heterozygosity rate is similar to the new site filters (0.028%), but the higher accessibility in the updated filter set (69.97% vs 62.46%) yields improved sensitivity.

As genomic features vary between species, different sets of site filters were generated to allow high quality analyses both within and between species.

The `gamb_colu` site filters were generated as above, and are appropriate for analyses that include *gambiae* and *coluzzii* samples only.

The `arab` site filters were generated following application of the model to the summary statistics from *arabiensis* samples in the cohort (n=XXX), this set of site filters are appropriate when working with *A. arabiensis* samples only.

Finally, the `gamb_colu_arab` site filters allow analyses across all three species and are the intersection of the `gamb_colu` and the `arab` site filters.

Place holders for tables.

- Table A: Mendel errors per cross per autosome. row indices: chromosome and raw/filtered column indexes: crosses + frac accessible. ie 8 rows, and 6 columns.
- Table B: comparison of cross and X row indices: raw/filtered column indices: MER, frac accessible, Youden, each for 2 and 3. column indexes: crosses + frac accessible. ie 2 rows, and 6 columns.

```
---  
caption: 'Result of heterozygote calls on male X chromosome'  
alignment: LLLLLLL  
include: content/tables/mer_X.csv  
csv-kwargs:  
    dialect: unix  
width: [0.2, 0.1, 0.1, 0.1, 0.1, 0.1, 0.1, 0.1]  
---
```

Genome accessibility

We define accessibility as the fraction of sites in a region passing the appropriate set of site filters.

Overall, 70% of the genome, and ??% of the exome are considered accessible in the `gamb_colu` set.

This is an improvement from phase 2, where XXX of the genome, and YYY of the exome was considered accessible.

As expected, accessibility was generally lower around the centromeres, and in regions of heterochromatin (table ref).

One notable region of low accessibility spans 40-41Mbp of chromosome 3R, this corresponds to ??.

Accessibility of the `arab` site filters closely follows that of `gamb_colu`, with the exception of the X chromosome where we see substantially lower values.

This appears to be driven by high divergence between AgamP4 and our *A. arabiensis* samples, particularly around the Xag inversion at Q-Q Mbp, ref figure.

On the autosomes the divergence from the reference is comparable between *A. arabiensis* and *A. gambiae*/ *A. coluzzii* samples, suggesting a strong basis for comparison across species.

The median divergence (Dxy) of 100kbp windows is XXX (5%/95% TTT/SSS) for gambiae/coluzzii and YYY (TTT/YYY) for arabiensis.

On the X chromosome these values are XXX (/) for gambiae/coluzzii and YYY (/) for arabiensis.

SNP discovery

Overall, we report XX,XXX,SSS single nucleotide polymorphisms (SNPs) segregating in this cohort that pass filters, of which XX,XXX (%) are multiallelic.

12,223 SNPs are segregating in both species groups, while XXX are private to *gambiae*/*coluzzii* and YYY to *arabiensis* [fig ref].

This phase of the study reports an additional XXX SNPs from phase 2.

In XXX *gambiae* and *coluzzii* individuals we report 12,222,222 SNPs (Q% multiallelic), corresponding to a SNP every 1.6 accessible bases.

In XXX *arabiensis* individuals we identify 10,000,000 SNPs (Q% multiallelic), a SNP every 2.5 accessible bases.

Population Structure

Genetic Diversity within Populations

Insecticide Resistance

Gene Drive

Methods

Population Sampling

Mosquitoes, from natural populations, were collected at 124 sites (unique latitude/longitudes) in 19 sub-Saharan African countries (Figure 1; Supplemental Table @@??).

95 of these sites are novel to Ag1000G phase 3, including 19 sites in six newly sampled countries, the remainder were previously sampled in phases 1 and 2 of the project ([??]; @[??] 2 doi when published).

New samples present in phase 3 comprised the following:

Burkina Faso

Two new submissions of collections from Burkina Faso are included here. The first added collections made in three villages separated by at most 30km: Bana (11.233, -4.472), Souroukoudinga (11.235, -4.535) and Pala (11.150, -4.235).

These collections were made in July and October 2014, and January, February and April 2015. The area is agricultural, with rice-growing areas near Bana and Souroukoudinga, and a large mango grove near Pala.

Female mosquitoes were collected by human landing catch, pyrethrum spray collection or aspiration; males were collected by swarm netting. Both *An. gambiae* and *An. coluzzii* ([1](#)) were collected.

Specimens were stored in 80% ethanol and DNA was extracted using the DNeasy Tissue Kit (Qiagen).

The second new submission from Burkina Faso added collections of indoor resting adults made by spray catch from Monomtenga in central Burkina Faso (12.06, -1.17).

These specimens were sorted morphologically to *An. gambiae* s.l.

Ovaries of half-gravid females were dissected and placed in numbered individual micro-tubes containing modified Carnoy's solution (1:3 glacial acetic acid: 100% ethanol).

Carcasses were placed in correspondingly numbered micro-tubes over desiccant. Genomic DNA was isolated from individual mosquitoes using one of the following: DNeasy Extraction Kit (Qiagen, Valencia, CA), Puregene kit (Gentra Systems, Inc., Minneapolis, MN), DNAzol kit (Molecular Research Center, Inc., Cincinnati, OH.) or Easy-DNA kit (Invitrogen, Carlsbad, CA).

An. gambiae s.s. and its molecular forms were identified using one of two rDNA-based PCR/RFLP assays ([2](#); [3](#)).

Ovaries from specimens of the desired species were subject to polytene chromosome analysis.

Democratic Republic of the Congo

Samples were collected from Gbadolite (4.283, 21.017), a town located in the far north of the Democratic Republic of Congo (DRC) near the border with the Central African Republic, surrounded by forest.

In common with much of DRC, malaria transmission rates are high, and the samples are *An. gambiae* s.s., which is the dominant vector.

Samples were collected as larvae from temporary pools within and around the town by dipping in early August 2015. All larvae were reared to adults and females preserved over silica for DNA extraction using Qiagen DNAEasy kits.

Central African Republic

Collections were carried out in Bangui (4.367, 18.583), during December 1993, by indoor resting aspiration or pyrethrum spray catch.

Cameroon

Two new submissions of samples from Cameroon are included in this phase.

In the first submission anopheline mosquitoes were taken from 64 locations covering a 1,500 km north-to-south transect that crossed all eco-geographical areas of Cameroon ([4](#)).

Mosquito collection involved spraying aerosols of pyrethroid insecticides inside human dwellings, dead mosquitoes were retrieved from white sheets that were laid on the floor.

Anopheline mosquitoes were identified using morphological identification keys (>>Gillies and De Meillon 1968<<@[???] reference books with no ISBN?; [5](#)).

Ovaries from half-gravid *An. gambiae* s.l. females were dissected and stored in Carnoy's fixative solution (absolute ethanol:glacial acetic acid 3:1) for cytogenetic analyses.

Carcasses were stored individually in tubes containing a desiccant and kept at -20°C until they were molecularly processed. All half-gravid specimens collected in each village were identified to species and molecular forms using PCR-RFLP ([2](#)).

The second Cameroonian submission came from pyrethrum spray collections, larval sampling and human landing catches conducted in twenty locations during October 2013.

These villages are scattered throughout the country and reflect a gradient of human-dominated environments, for example, forest (Manda: 5.726, 10.868 and Campo: 2.367, 9.817); forest/savanna transition (Tibati: 6.469, 12.629); savanna (Lagdo: 9.049, 13.656); suburban area (Nkolondom: 3.972, 11.516) and urban areas (Douala: 4.055, 9.721 and Yaoundé: 3.880, 11.506).

Contributed specimens were *An. gambiae* and *An. coluzzii* ([2](#)).

Population genomics studies indicated the presence of relatively differentiated subgroups within both species as well as clusters thriving in polluted breeding sites in large cities ([???]).

Specimens were stored on silica gel, and DNA was extracted using a Zymo research kit for adults and a Qiagen kit for larvae.

Mayotte

Phase 3 adds collections from three novel sites on the island of Mayotte.

Samples were collected as larvae during March-April 2011 in temporary pools by dipping in Tsounzou (-12.797, 45.185), Tsinkoura (-12.936, 45.138) and aerogare (-12.803, 45.283).

Larvae were stored in 80% ethanol prior to DNA extraction.

All specimens contributed were *An. gambiae* ([3](#)) with the standard $2L^{+a}/2L^{+a}$ or inverted $2L^a/2L^a$ karyotype as determined by the molecular PCR diagnostics ([6](#)).

The samples were identified as males or females by the sequencing read coverage of the X chromosome using LookSeq ([7](#)).

Gabon @[???] this submission "GA-B" included in the release?? I can't see it in the meta data, delete?

Anopheles sampling was carried out in three new locations in Gabon @[???]?:

Adult *An. gambiae* females were collected in Benguia (-1.633, 13.492) by human landing catches (National Research Ethics Committee of Gabon n°. 0031/2014/SG/CNE) in September 2015.

Benguia is a forest-savanna village with less than 300 inhabitants. Malaria is endemic across the year.

The specimens were stored immediately at -80°C.

An. coluzzii larvae were sampled in Libreville (0.390, 9.454) and Cocobeach (0.992, 9.576) by dipping in natural breeding sites in January 2016.

Libreville is the capital of the country and it is urban and polluted site, Cocobeach is a coastal village north of Libreville.

Across both malaria is endemic across the year.

The specimens were stored in alcohol at -20°C.

Total genomic DNA was extracted for all the mosquitoes using the CTAB protocol ([8](#)).

The Gambia

Two new submissions of samples from The Gambia are present in phase 3.

The first were collected along the Gambia River from the western coastal region of The Gambia, (Low River Area; Caputo et al. 2008), in August 2006.

An. gambiae and *An. coluzzii* specimens were identified to species following the PCR-RFLP protocol ([2](#)) using DNA extracted from the mosquito leg.

Only *An. coluzzii* specimens were collected from villages of Tankular (13.417, -16.033) and Kalataba (13.550, -15.617).

An. gambiae and *An. coluzzii* specimens were found in sympatry and collected from villages of Yallal Tankonjala (13.550, -15.700), Sare Samba Sowe (13.583, -15.900) and Hamdalai (13.567, -16.0167).

PCR-RFLP protocol also revealed the presence of mosquitoes with hybrid *An. gambiae*/*An. coluzzii* genotype in Yallal Tankonjala and Sare Samba Sowe.

Collections of indoor daytime-resting half gravid mosquitoes were carried out mainly in human dwellings and, in few cases, in animal shelters.

Collections were carried out by pyrethroid and/or paper-cup mouth aspirators from 12 AM to sunset, and kept in vials with desiccant.

Ovaries were dissected, maintained into Carnoy fixative (three parts pure ethanol:one part glacial acetic acid) and stored at -20°C before polytene chromosome preparations ([9](#)).

Chromosome scoring was carried out under a phase-contrast optical microscope.

Paracentric inversion karyotypes were scored according to the nomenclature and conventions of Coluzzi et al. ([[????](#)](79)90036-1) and Touré et al. ([10](#)).

The second new submission consists of adult mosquitoes collected at Wali Kunda in the rural, central river region of The Gambia (13.567, -14.917).

The area is 180 km from the sea, on the south bank of the River Gambia, in flat Sudan savannah with a small fishing village (and a research field station) as well as rice fields and swamplands.

The dominant *Anopheles* vector species is *An. coluzzii* ([11](#)).

Mosquitoes were captured using human landing collections both inside and outside huts for 19 days in October and November 2012. Mosquitoes were stored in RNAlater or dried over silica gel and stored at -20°C.

Guinea and Mali

A novel sample submission to phase 3 included collections from both Guinea and Mali.

Mosquitoes were collected from four different study sites at the border with Mali and in Guinea Conakry. Takan (11.47, -8.33) and Toumani Oulena (10.83, -7.81) are both small villages in the Yanfolila district of southern Mali and represent the Sudanian savannah ecological zone.

Takan is arid savannah, while Toumani Oulena is humid savannah.

In Guinea Conakry, we sampled in Koraboh, (9.28, -10.03) a small village in the Kissidougou district in the Faranah region representing a semi-forest site with intermediate ecology, a mix of savannah and forest, and in Koundara, (8.48, -9.53), a small village in the Macenta district in the Nzerekore region representing deep forest ecology.

All reported collections occurred in October and November in 2012.

At each site, mosquitoes were collected using three different methods: human-landing capture, indoor manual aspirator or pyrethroid spray catch, and larval capture - where the first and second instar larvae were raised to adult in a field insectary under standard insectary conditions prior to DNA isolation from the adults, and the third and fourth instar larvae were preserved directly for DNA isolation, without rearing in the insectary.

The two distinct methods of larval collection were used to control for possible genetic bias inherent in lab rearing of captured larvae.

Across sites, all types of larval sites were sampled, including both temporary and permanent sites.

Human-landing captures were performed both inside dwellings and outside (>10 m from dwelling) at night between 18:00 and 06:30.

The indoor aspirator or spray catches were done in the morning between 06:00 and 12:00.

Adult specimens or third and fourth instar larvae were preserved immediately in 80% ethanol until later DNA extraction.

First and second instar larvae were raised to adults in nearby field insectaries and upon emergence were preserved in 80% ethanol.

DNA was extracted from mosquitoes using DNAzol by the provided protocol (Invitrogen, CA, USA).

Further details on sampling and basic Southern Mali/Guinea vector biology are presented in Coulibaly et al. (doi:10.1186/s12936-016-1242-5).

Guinea Bissau

Two new Guinea Bissau collection sites are added here, both performed in October 2010 by indoor CDC light traps.

Samples were also collected from Ga-Mbana (12.052, -14.902) and Leibala (12.272, -14.222).

Ga-Mbana is a rural village located along a main road in central Guinea Bissau, while Leibala is a neighbourhood of the eastern town of Gabu.

The samples of Ga-Mbana comprised *An. coluzzii* whereas the samples of Leibala comprised *An. gambiae*, all being identified by IGS and SINEX markers as described in Vicente et al. ([12](#)).

The *kdr* pyrethroid target site resistance mutation L1014F occurs at high frequency in Leibala but at very low frequency in Ga-Mbana ([12](#)).

Malaria is meso-hyperendemic ([13](#)) and sporozoite rates are below 1% in the region.

Specimens were stored on silica gel and DNA extraction was performed by a phenol-chloroform protocol described in Donnelly et al. ([14](#)).

Kenya

New Kenyan specimens were obtained from villages located in Kilifi County near the Kenyan coast between 2000 and 2014.

All Anopheles mosquito sampling was conducted indoors using CDC light traps.

An. gambiae, *An. funestus*, *An. arabiensis* and *An. merus* were present at sampling locations ([15](#)).

Sporozoite rates for the area during previous studies were 1.47% ([16](#)).

Mali

Two further submissions containing collections from 11 sites in Mali are new for phase 3.

In the first, collections were made in four villages in the Koulikoro region; Tieneguebougou (12.810, -8.080) approximately 20 km north of Bamako, and Kababougou (12.890, -8.150), Ouassorola (12.900, -8.160), Sogolombougou (12.880, -8.140), approximately 30 km north of Bamako.

The collections were made in August 2014 by human landing catch and pyrethrum spray catch.

Both *An. gambiae* and *An. coluzzii* ([2](#)) were collected.

Specimens were stored in 80% ethanol.

In the second submission, collections of indoor resting adults were made by spray catch from seven villages in the southern part of Mali in August-September 2004: Banambani (12.800, -8.050), Bancoumana (12.200, -8.200), Douna (13.210, -5.900), Fanzana (13.200, -6.130), Kela (11.880, -8.450), Moribobougou (12.690, -7.870) and N'Gabakoro (12.680, -7.840).

Specimens were sorted morphologically to *An. gambiae* s.l.

Ovaries of half-gravid females were dissected and placed in numbered individual micro-tubes containing modified Carnoy's solution (1:3 glacial acetic acid: 100% ethanol).

Carcasses were placed in correspondingly numbered micro-tubes over desiccant.

Genomic DNA was isolated from individual mosquitoes using one of the following: DNeasy Extraction Kit (Qiagen, Valencia, CA), Puregene kit (Genta Systems, Inc., Minneapolis, MN), DNAzol kit (Molecular Research Center, Inc., Cincinnati, OH.) or Easy-DNA kit (Invitrogen, Carlsbad, CA).

An. gambiae s.s. and its molecular forms were identified using one of two rDNA-based PCR/RFLP assays ([2](#); [3](#)).

Ovaries from specimens of the desired species were subject to polytene chromosome analysis.

Malawi

Specimens were obtained from villages within the catchment of the Majete Malaria Project, Chikhwawa District, Malawi (-15.933, 34.755) ([17](#)).

Mosquitoes were collected indoors and outdoors by Suna light trap in May 2015.

Chikhwawa District is an area with perennial and intense malaria transmission ([18](#)).

All specimens were *An. arabiensis* ([2](#)).

Additional details of vector population bionomics may be found in ([17](#); [19](#)).

Specimens were stored over silica and DNA was extracted using the Qiagen plate protocol.

Mozambique

Mosquito samples were collected in Furvela (-23.716, 35.299), Mozambique, by CDC light traps between December 2003 and April 2004.

Specimens were stored on silica gel and DNA was extracted according to Collins et al. ([20](#)).

Contributed specimens consisted of *An. gambiae* individuals identified according to Fanello et al. (2).

Furvela is a rural village located in Inhambane Province, where malaria is transmitted mainly by *An. gambiae* and *An. funestus* (21).

An. arabiensis and *An. merus* are also found at low frequency.

Sporozoite rates around 4% have been reported in *An. gambiae* from Furvela (21).

Tanzania

Tanzanian samples were collected from four distinct locations.

Moshi samples came from lower Mabogini (-3.400, 37.350), rice fields near lower Moshi on the southern slope of Mount Kilimanjaro, a region shown to have increasing resistance to pyrethroids (22).

Mosquitoes were collected as larvae, during the rice growing season in August-September 2012, raised to adults and females bioassayed in WHO tubes for one hour with 0.05% lambda cyhalothrin ([??]).

Alive and dead mosquitoes were preserved over silica.

In Tanzanian samples screened in Kabula et al. (23), Moshi was the most pyrethroid resistant population, they were found to be completely DDT susceptible, only in one out of 642 mosquitoes assayed by Matowo et al. (22) was found to carry a *kdr* resistance mutation (Vgsc-995F).

Tarime collections took place in the village of Komaswa (-1.417, 34.183) about 410 km north west of Moshi, during August 2012.

Mosquito larvae were collected, raised to adults and females bioassayed with a range of insecticides in WHO tubes for one hour ([??]), finding almost complete multi-insecticide susceptibility: permethrin (100% mortality), lambda cyhalothrin (97%), fenitrothion (100%), DDT (100%) and bendiocarb (100%) (Nyka, T. unpublished data – Insecticide Resistance Monitoring Report 2012. NIMR Tanzania).

Mulheza samples were collected from Zeneti village (-5.217, 38.650), northeast Tanzania.

Malaria is intense and perennial with transmission peaking after the rainy season in May and June (23). Mosquitoes were sampled between November 2012 and May 2013.

Indoor resting collections were used to obtain live females for deltamethrin susceptibility testing and pyrethrum spray catches were used for mosquitoes that were collected for blood meal analysis.

Collections were conducted between 06:00 and 09:00 from randomly selected houses.

Live mosquitoes collected for susceptibility testing were provided with 10% glucose solution and transported to the field insectary.

Mosquitoes were sorted and morphologically identified to species, carcasses were stored individually over desiccant for laboratory processing. Muleba (1.750, 31.667), the final collection region, is in the North-western part of Tanzania.

The district is known to be a malaria epidemic prone area with unstable transmission of varying seasonality.

The highest peak of malaria transmission is usually reached between May-July and November-January, which results from proceeding rain seasons.

There have been malaria vector control efforts since 2007 when indoor residual spraying using Lambdacyhalothrin was introduced.

Insecticide resistance in this district is coupled with high frequency of *kdr* pyrethroid target site mutations in the *An. gambiae* s.s population ([23](#); [24](#)).

Sampling was conducted over six months, which include both dry and rainy season and covers 6 villages selected to represent all major ecological systems in the district.

Uganda

In Uganda, a single new site is sampled in phase 3.

In Kihiihi subcounty, Kanungu District (-0.751, 29.701), resting mosquitoes were collected during October and November 2012.

Kihiihi is located in an upland area with seasonal malaria transmission ([25](#)).

All specimens were *An. gambiae* ([2](#)).

Additional details of vector population bionomics may be found in ([??]).

Specimens were stored in 80% ethanol and DNA was extracted using the Qiagen plate protocol.

Details of natural population samples previously described in phases 1 and 2 can be found in the Supplementary Information of The Anopheles gambiae 1000 Genomes Consortium ([??]; @[??] 2 doi when published - respectively).

Colony Crosses

15 crosses were contributed to Ag1000G phase 3.

The crosses were generated using parents from eight different colonies: G3(MRA-112); Kisumu(MRA-762); Pimperena (canonical representative of *An. gambiae* species; MRA-861); Ghana (recent colony of *An. coluzzii* from Okyereko, southern Ghana ([26](#))); Mali-NIH (canonical representative of *An. coluzzii* species; Niono, MRA-860); (P)Akron (Benin, MRA-913); Nagongera (Tororo, Uganda); and Tiassalé (southern Cote d'Ivoire ([26](#))).

The labels, e.g. "29-2", are identifiers used for each of the crosses within the project and have no special meaning.

Full details of cross production, sequencing and quality control is described in the Supplementary Information of The Anopheles gambiae 1000 Genomes Consortium ([??]).

The only difference between the production of the four crosses that are novel to phase 3: B4, K2, K4 and K6, and those from earlier phases of the project, is that multiple males and multiple females were placed together for mating, then all male and all egg laying female mosquitoes were sequenced.

This necessitated matching up potential fathers of crosses with the correct mother and offspring.

For each cross for which the father was in doubt, the list of potential parental pairs was computed.

For each of these pairs, for each chromosome, the Mendelian error was computed for every sample of the progeny and the median value (along samples) was plotted for every computation.

If one pair yielded median Mendelian errors significantly lower for every chromosome than any other pair (except X which was consistent no matter the parents), that pair was chosen as the parsimonious parents.

In four crosses (B4, K2, K4 and K6) parental pairs could be clearly identified and these crosses could, therefore, be included in the phase 3 data release.

Two of the novel crosses, K4 and K6, were found to be fathered by the same male, AC0398.

Whole Genome Sequencing and Alignment

4,693 individual mosquitoes were sequenced using the Illumina HiSeq2000 (n=3,130) and the Illumina HiSeqX (n=1,563) to a target coverage of 30X.

Reads were aligned to the AgamP4 reference genome using `bwa` version `0.7.15`.

Indel realignment was performed using GATK `v3.7-0` RealignerTargetCreator and IndelRealigner.

Single nucleotide polymorphisms were called against AgamP4 using GATK UnifiedGenotyper `v3.7-0`.

Sample genotypes were called independently, in genotyping mode, given all possible alleles at each site, allowing parallelisation over samples.

Coverage considered at individual sites was capped at 250X by random downsampling.

Full details of pipelines including all parameter settings are available in supplementary information.

Following successfull completion of the pipeline samples entered the sample quality control (QC) process.

Sample QC

The sample QC process was composed of three distinct stages: sequence quality assurance, replicate handling, and anomaly detection.

To meet the requirements of sequence quality assurance median coverage had to be at least 10X, and minumum 50% of the genome covered by at least 1X.

We also implemented the test for contamination in NGS alignments described in Jun et al (<https://doi.org/10.1016/j.ajhg.2012.09.004>).

Briefly the method estimates the likelihood of the observed alternate and reference allele counts under different contamination fractions given population allele frequencies.

Population allele frequencies were estimated from the Ag1000G phase 2 data.

The model computes a maximum likelihood value for a parameter representing percentage contamination (alpha).

Where this parameter was 4.5% or greater the sample was excluded.

We also made sex calls based on the modal coverage ratio X:3R.

3R was selected as representative of autosomal coverage as it is free from inversions and large regions of heterochromatin.

The modal coverage was used owing to concerns around the high skewness of coverage distributions.

To mitigate this further, when computing the modal coverage we only considered sites where coverage was at least 2X.

The sample was classed as male where the coverage ratio was between 0.4-0.6, and female between 0.8-1.2.

Where the ratio was outside these limits, the sex call was not made and the sample dropped.

One of the submission sets from The Gambia, was composed entirely of Whole Genome Amplified (WGA) samples.

These received a sex call where possible but the decision was made not to exclude based on ratio, due to the inherent value of this submission.

The fact this set is entirely WGA was considered throughout the analysis of these samples.

The sample QC process also included assessment of technical replicates.

We computed pairwise genetic distance between all sample pairs within a submission set.

The metric used was Hamming between alleles, so a genotype of 0/1 records a value of 1 against the genotype 1/2, allowing straightforward handling of multiallelic SNPs.

Only sites where both samples had a genotype call were included, mean distance was calculated over a denominator of the number of assessed sites multiplied by 2 for diploidy.

Computations were initially carried out on a downsampled set of 10 x 100,000 contiguous sites genome wide to be computationally feasible.

This use of chunks was convenient to leverage the underlying storage of data in `zarr` format.

Where a pair of samples fell beneath a conservative threshold of 0.012, the true genetic distance (i.e. without downsampling) was computed across all sites.

For each pair of technical replicates, we excluded both members of the pair where genetic distance was above 0.006.

Where replicate pairs met the concordance threshold we excluded the lower quality sample.

Quality was determined based on the skewdness of mean to median, i.e. $1 - |mean_{cov}/median_{cov}|$.

The sample with the lower value was preferred as it suggests a more normal coverage distribution.

To identify unknown replicate pairs as a result of sample mix ups or mislabelling, we screened within submission sets for unexpected pairs, using the genetic distance cut off of 0.006 as above.

We did not attempt to identify unknown replicate pairs in the AG1000G-X submission set, made up of laboratory experimental crosses, due to familial similarity and high levels of inbreeding.

The third stage used principal component analysis (PCA) to identify and exclude individual samples that were outliers based on available metadata.

A review process identified samples that could not be explained parsimoniously, and were therefore likely to be sample mix ups or instances of mislabelling.

Using the PCA implementation in `scikit-allel v2.1.0` we downsampled to 100,000 segregating non-singleton sites from chromosomes 3R and 3L, to avoid regions complicated by known introgression loci or paracentric inversions.

Multiallelic sites were included as dummy rows by melting the data structure.

A careful review process identified: a) samples that dominated single principal components, either individually or in very small numbers.

This suggests an individual belonging to another Anopheline species, or some inherent problem with the sample.

Or b) samples that clustered with other samples inconsistently with metadata.

This was a subjective assessment, but bore in mind the given collection location, time, and PCR species assignment (where available) of the sample.

Multiple geographical sites submitted by the same partner were also considered, where sample mix ups formed the most parsimonious explanations of incongruent clustering.

Species assignment and sex calling

Ancestry informative markers were used to assign species in our cohort.

To derive markers informative between *A. arabiensis* and *A. gambiae s.l.*, we used publicly available data from the 16 genomes project (ref).

Whole genome SNP calls called against the AgamP3 reference for 12 *A. arabiensis* and 38 *A. gambiae s.l.* individuals were available.

Alleles were mapped onto the same alternate allele space, the frequencies of which were computed across both groups.

Sites that were multiallelic in either group were excluded, as well as sites where any genotypes were missing.

565,329 SNPs were identified as potentially informative where no shared alleles were present between groups.

These were spread throughout the genome, but were concentrated on the X chromosome (63.2%), particularly around the Xag inversion.

The full AIM set of positions and alleles are available as part of the phase 3 data release.

Called genotypes for each individual in the dataset were cross referenced against a random subset of 50,000 ancestry informative marker alleles, genotype alleles were accordingly classified as gambiae-like or arabiensis-like.

AIM fractions were cross referenced against PCR results available from a subset of individuals.

Given the relatively small number of *A. arabiensis* samples in the 16 genomes project- it was clear that a significant proportion of putative AIMs were not likely to be truly informative.

Therefore, classification requirements are less rigorous than other sets of validated markers; individuals were classed as *A. arabiensis* where a fraction >0.6 of alleles were arabiensis-like (n=368), and as *A. gambiae* s.l where this value was < 0.03 (n=2415).

Species were not assigned to samples from the AG1000G-X submission due to inbreeding and high levels of genetic drift.

A single individual collected in Tororo, Uganda is classed as intermediate- given the majority (XX%) of sites in the genome are heterozygous between the gambiae and arabiensis allele, this individual is likely to be an F1 hybrid.

To resolve the *A. gambiae* s.l individuals into *A. gambiae* and *A. coluzzii* we applied the 729 AIMs previously identified by Neafsey et al (ref).

?? Need to find code that creates gambiae from neafsey set

Cutoffs were made at <0.12 (gambiae) and >0.9 (coluzzii), with individuals between classed as intermediate.

Of the 2415 s.l individuals, 1571 were called as gambiae, 675 as coluzzii and 169 as intermediate (ref collection map).

SNP filtering and quality

Site filtering ensures that reported variation is of high quality.

As genomic features vary between species, different sets of site filters were generated to allow high quality analyses both within and between species.

The `gamb_colu` site filters were generated using a decision tree model, and are appropriate for analyses that include *gambiae* and *coluzzii* samples only.

Inputs to the decision tree model are summary statistics from the set of SNP calls and genotype alignments.

The `arab` site filters were generated following application of the resulting model to the summary statistics from *arabiensis* samples in the cohort, this set of site filters are appropriate when working with *A. arabiensis* samples only.

Finally, the `gamb_colu_arab` site filters allow analyses across all three species and are the intersection of the `gamb_colu` and the `arab` site filters.

Using the 15 available *Anopheles* pedigrees previously described, we used the presence of mendelian error at sites as a proxy for genotype discordance.

Where previously we have used manually curated cutoffs based on observed mendelian error rates to filter sites (ref phase1, phase2), here we built a statistical model where cohort level summary statistics were used to predict the presence or absence of mendelian error, becoming a binary classification problem.

Summary statistics used as input to the model are presented in table (ref).

Pedigrees included *A. gambiae* and *A. coluzzii* mosquitoes only, and summary statistics to build the initial site filters model came from females of these species (n=2415).

Males were excluded, so that the model could also be applied without modification to the X chromosome.

5 of the 15 crosses were held out for validation, so performance could be evaluated objectively.

Sites were defined as PASS where all genotypes across all 10 remaining crosses were called, and no mendelian inconsistencies were observed.

Sites were defined as FAIL where a mendelian inconsistency was observed in any pedigree.

All other sites were not considered eligible for inclusion in model training.

A balanced training set was generated from the remaining 10 crosses containing XXX autosomal(?) sites.

We applied a decision tree, as it provides decisions with clear explanations, and is similar in concept to the set of hard thresholds commonly used in SNP calling in non-model organisms.

A set of trees with different parameter settings were learned, exploring the depth of trees and the number of samples allowed at a terminal node.

Parameter settings were evaluated on an unbalanced evaluation set, consisting of XXX sites randomly sampled from the whole genome.

Leaves of the trained models contain different proportions of PASS sites, by increasing the cutoff for these proportions required to label a leaf as PASS, we were able to compute the area under the receiver operating curve (AUROC) for each parameter set.

The best performing parameter set based on AUROC was selected as the final model, the classification cutoff used was optimised based on the Youden statistic.

The resulting model was a decision tree of depth 8, with a maximum of XX (CHECK NUMBERS) terminal nodes, where leaves were assigned to PASS where > 0.533 of training data in that leaf were PASS.

All sites in the genome were then assigned to PASS or FAIL given the model inputs.

The 5 remaining cross pedigrees were used to perform a final evaluation of the approach.

Above definitions of PASS sites were retained, but independently within pedigrees, providing 5 distinct evaluation sets.

To evaluate performance on the hemizygous X chromosome we use the more direct measure of heterozygote calls in males.

In the dataset are 220 *A. gambiae s.l.* male samples, each of which represent an independent proxy for genotype discordance.

Male error rates were estimated from genotype calls with a minimum Genotype Quality (GQ) value of 30.

A non heterozygote call is labelled PASS, and a heterozygote call FAIL.

Error rates were computed for all crosses, over all chromosomes before and after application of site filters.

We report the false positive rate (FPR), (i.e proportion of the genome considered accessible), and the Youden statistic (ratio of sensitivity to specificity).

References

1.:{unav}

Elien E Wilkins, Paul I Howell, Mark Q Benedict
Malaria Journal (2006) <https://doi.org/dbtfkr>
DOI: [10.1186/1475-2875-5-125](https://doi.org/10.1186/1475-2875-5-125) · PMID: [17177993](#) · PMCID: [PMC1769388](#)

2. Simultaneous identification of species and molecular forms of the *Anopheles gambiae* complex by PCR-RFLP

C. Fanello, F. Santolamazza, A. Della Torre
Medical and Veterinary Entomology (2002-12) <https://doi.org/ds5pmz>
DOI: [10.1046/j.1365-2915.2002.00393.x](https://doi.org/10.1046/j.1365-2915.2002.00393.x) · PMID: [12510902](#)

3. SHORT REPORT: A NEW POLYMERASE CHAIN REACTION-RESTRICTION FRAGMENT LENGTH POLYMORPHISM METHOD TO IDENTIFY ANOPHELES ARABIENSIS FROM AN. GAMBIAE AND ITS TWO MOLECULAR FORMS FROM DEGRADED DNA TEMPLATES OR MUSEUM SAMPLES

ALESSANDRA DELLA TORRE, ADALGISA CACCONE, FEDERICA SANTOLAMAZZA
The American Journal of Tropical Medicine and Hygiene (2004-06-01) <https://doi.org/ghcqgn>
DOI: [10.4269/ajtmh.2004.70.604](https://doi.org/10.4269/ajtmh.2004.70.604)

4. Ecological niche partitioning between *Anopheles gambiae* molecular forms in Cameroon: the ecological side of speciation

Frédéric Simard, Diego Ayala, Guy Kamdem, Marco Pombi, Joachim Etouna, Kenji Ose, Jean-Marie Fotsing, Didier Fontenille, Nora J Besansky, Carlo Costantini
BMC Ecology (2009) <https://doi.org/bd8bz5>
DOI: [10.1186/1472-6785-9-17](https://doi.org/10.1186/1472-6785-9-17) · PMID: [19460146](#) · PMCID: [PMC2698860](#)

5. The Anophelinae of Africa south of the Sahara. Suppl: Afrotropical region

M. T. Gillies, Botha de Meillon
Publications of the South African Institute for Medical Research (1987)
ISBN: [9780620103213](#)

6. MOLECULAR KARYOTYPING OF THE 2LA INVERSION IN ANOPHELES GAMBIAE

NORA J. BESANSKY, OLGA GRUSHKO, ALESSANDRA DELLA TORRE, IGOR SHARAKHOV, CECILE BRENGUES, JONATHAN K. KAYONDO, FEDERICA SANTOLAMAZZA, FREDERIC SIMARD, MARCO POMBI, MAMADOU COULIBALY, ... KARINE MOULINE

The American Journal of Tropical Medicine and Hygiene (2007-02-01) <https://doi.org/ghcqgg>
DOI: [10.4269/ajtmh.2007.76.334](https://doi.org/10.4269/ajtmh.2007.76.334)

7. LookSeq: A browser-based viewer for deep sequencing data

H. M. Manske, D. P. Kwiatkowski
Genome Research (2009-08-13) <https://doi.org/b5hmbh>
DOI: [10.1101/gr.093443.109](https://doi.org/10.1101/gr.093443.109) · PMID: [19679872](#) · PMCID: [PMC2775587](#)

8. POPULATION STRUCTURE OF ANOPHELES ARABIENSIS ON LA RÉUNION ISLAND, INDIAN OCEAN

FRÉDÉRIC SIMARD, RICHARD HUNT, DIDIER FONTENILLE, ISABELLE MORLAIS, ROMAIN GIROD
The American Journal of Tropical Medicine and Hygiene (2005-12-01) <https://doi.org/ghcqgp>
DOI: [10.4269/ajtmh.2005.73.1077](https://doi.org/10.4269/ajtmh.2005.73.1077)

9. Anopheles gambiae complex along The Gambia river, with particular reference to the molecular forms of An. gambiae s.s

Beniamino Caputo, Davis Nwakanma, Musa Jawara, Majidah Adiamoh, Ibrahima Dia, Lassana

Konate, Vincenzo Petrarca, David J Conway, Alessandra della Torre

Malaria Journal (2008) <https://doi.org/b97c6c>

DOI: [10.1186/1475-2875-7-182](https://doi.org/10.1186/1475-2875-7-182) · PMID: [18803885](https://pubmed.ncbi.nlm.nih.gov/18803885/) · PMCID: [PMC2569043](https://pubmed.ncbi.nlm.nih.gov/PMC2569043/)

10. The distribution and inversion polymorphism of chromosomally recognized taxa of the *Anopheles gambiae* complex in Mali, West Africa.

YT Touré, V Petrarca, SF Traoré, A Coulibaly, HM Maiga, O Sankaré, M Sow, MA Di Deco, M Coluzzi
Parassitologia (1998-12) <https://www.ncbi.nlm.nih.gov/pubmed/10645562>

PMID: [10645562](https://pubmed.ncbi.nlm.nih.gov/10645562/)

11. Does insecticide resistance contribute to heterogeneities in malaria transmission in The Gambia?

Kevin Ochieng' Opondo, David Weetman, Musa Jawara, Mathurin Diatta, Amfaal Fofana, Florence Crombe, Julia Mwesigwa, Umberto D'Alessandro, Martin James Donnelly
Malaria Journal (2016-03-15) <https://doi.org/ghcqgh>

DOI: [10.1186/s12936-016-1203-z](https://doi.org/10.1186/s12936-016-1203-z) · PMID: [26980461](https://pubmed.ncbi.nlm.nih.gov/26980461/) · PMCID: [PMC4793517](https://pubmed.ncbi.nlm.nih.gov/PMC4793517/)

12. Massive introgression drives species radiation at the range limit of *Anopheles gambiae*

José L. Vicente, Christopher S. Clarkson, Beniamino Caputo, Bruno Gomes, Marco Pombi, Carla A. Sousa, Tiago Antao, João Dinis, Giordano Bottà, Emiliano Mancini, ... João Pinto
Scientific Reports (2017-04-18) <https://doi.org/f93m36>

DOI: [10.1038/srep46451](https://doi.org/10.1038/srep46451) · PMID: [28417969](https://pubmed.ncbi.nlm.nih.gov/28417969/) · PMCID: [PMC5394460](https://pubmed.ncbi.nlm.nih.gov/PMC5394460/)

13. TRANSMISSION OF MIXED PLASMODIUM SPECIES AND PLASMODIUM FALCIPARUM GENOTYPES

VIRGÍLIO E. DO ROSÁRIO, KATINKA PÅLSSON, THOMAS G. T. JAENSON, GEORGES SNOUNOU, JOÃO PINTO, ANA PAULA AREZ

The American Journal of Tropical Medicine and Hygiene (2003-02-01) <https://doi.org/ghcqgm>

DOI: [10.4269/ajtmh.2003.68.2.0680161](https://doi.org/10.4269/ajtmh.2003.68.2.0680161)

14. Population structure in the malaria vector, *Anopheles arabiensis* Patton, in East Africa

MJ Donnelly, N Cuamba, JD Charlwood, FH Collins, H Townson

Heredity (1999-10-01) <https://doi.org/bg4xmm>

DOI: [10.1038/sj.hdy.6885930](https://doi.org/10.1038/sj.hdy.6885930) · PMID: [10583542](https://pubmed.ncbi.nlm.nih.gov/10583542/)

15. Identification of Single Specimens of the *Anopheles Gambiae* Complex by the Polymerase Chain Reaction

Julie A. Scott, William G. Brogdon, Frank H. Collins

The American Journal of Tropical Medicine and Hygiene (1993-10-01) <https://doi.org/ghcqgk>

DOI: [10.4269/ajtmh.1993.49.520](https://doi.org/10.4269/ajtmh.1993.49.520) · PMID: [8214283](https://pubmed.ncbi.nlm.nih.gov/8214283/)

16. Wind direction and proximity to larval sites determines malaria risk in Kilifi District in Kenya

Janet T. Midega, Dave L. Smith, Ally Olotu, Joseph M. Mwangangi, Joseph G. Nzovu, Juliana Wambua, George Nyangweso, Charles M. Mbogo, George K. Christophides, Kevin Marsh, Philip Bejon

Nature Communications (2012-02-14) <https://doi.org/ghcqgd>

DOI: [10.1038/ncomms1672](https://doi.org/10.1038/ncomms1672) · PMID: [22334077](https://pubmed.ncbi.nlm.nih.gov/22334077/) · PMCID: [PMC3292715](https://pubmed.ncbi.nlm.nih.gov/PMC3292715/)

17. Assessment of the effect of larval source management and house improvement on malaria transmission when added to standard malaria control strategies in southern Malawi: study protocol for a cluster-randomised controlled trial

Robert S. McCann, Henk van den Berg, Peter J. Diggle, Michèle van Vugt, Dianne J. Terlouw, Kamija S. Phiri, Aurelio Di Pasquale, Nicolas Maire, Steven Gowelo, Monicah M. Mburu, ... Willem Takken
BMC Infectious Diseases (2017-09-22) <https://doi.org/ggr5gZ>
DOI: [10.1186/s12879-017-2749-2](https://doi.org/10.1186/s12879-017-2749-2) · PMID: [28938876](https://pubmed.ncbi.nlm.nih.gov/28938876/) · PMCID: [PMC5610449](https://pubmed.ncbi.nlm.nih.gov/PMC5610449/)

18. Mapping Malaria Transmission Intensity in Malawi, 2000–2010

Adam Bennett, Lawrence Kazembe, Don P. Mathanga, Damaris Kinyoki, Doreen Ali, Robert W. Snow, Abdisalan M. Noor

The American Journal of Tropical Medicine and Hygiene (2013-11-06) <https://doi.org/f5h9zz>
DOI: [10.4269/ajtmh.13-0028](https://doi.org/10.4269/ajtmh.13-0028) · PMID: [24062477](https://pubmed.ncbi.nlm.nih.gov/24062477/) · PMCID: [PMC3820324](https://pubmed.ncbi.nlm.nih.gov/PMC3820324/)

19. Entomological indices of malaria transmission in Chikhwawa district, Southern Malawi

Themba Mzilahowa, Ian M Hastings, Malcolm E Molyneux, Philip J McCall

Malaria Journal (2012-11-21) <https://doi.org/gbcb3h>

DOI: [10.1186/1475-2875-11-380](https://doi.org/10.1186/1475-2875-11-380) · PMID: [23171123](https://pubmed.ncbi.nlm.nih.gov/23171123/) · PMCID: [PMC3536595](https://pubmed.ncbi.nlm.nih.gov/PMC3536595/)

20. A Ribosomal RNA Gene Probe Differentiates Member Species of the *Anopheles gambiae* Complex

Frank H. Collins, Nora J. Besansky, M. Alina Mendez, Melissa O. Rasmussen, Victoria Finnerty, Philip C. Mehaffey

The American Journal of Tropical Medicine and Hygiene (1987-07-01) <https://doi.org/ghcqgi>
DOI: [10.4269/ajtmh.1987.37.37](https://doi.org/10.4269/ajtmh.1987.37.37) · PMID: [2886070](https://pubmed.ncbi.nlm.nih.gov/2886070/)

21. Analysis of the sporozoite ELISA for estimating infection rates in Mozambican anophelines

J. D. CHARLWOOD, E. V. E. TOMÁS, N. CUAMBA, J. PINTO

Medical and Veterinary Entomology (2015-03) <https://doi.org/f62ckd>

DOI: [10.1111/mve.12084](https://doi.org/10.1111/mve.12084) · PMID: [25088021](https://pubmed.ncbi.nlm.nih.gov/25088021/)

22. Genetic basis of pyrethroid resistance in a population of *Anopheles arabiensis*, the primary malaria vector in Lower Moshi, north-eastern Tanzania

Johnson Matowo, Christopher M Jones, Bilali Kabula, Hilary Ranson, Keith Steen, Franklin Mosha, Mark Rowland, David Weetman

Parasites & Vectors (2014) <https://doi.org/ghcqgg>

DOI: [10.1186/1756-3305-7-274](https://doi.org/10.1186/1756-3305-7-274) · PMID: [24946780](https://pubmed.ncbi.nlm.nih.gov/24946780/) · PMCID: [PMC4082164](https://pubmed.ncbi.nlm.nih.gov/PMC4082164/)

23. Susceptibility status of malaria vectors to insecticides commonly used for malaria control in Tanzania

Bilali Kabula, Patrick Tungu, Johnson Matowo, Jovin Kitau, Clement Mweya, Basiliiana Emidi, Denis Masue, Calvin Sindato, Robert Malima, Jubilate Minja, ... William Kisinza

Tropical Medicine & International Health (2012-06) <https://doi.org/f3zsbt>

DOI: [10.1111/j.1365-3156.2012.02986.x](https://doi.org/10.1111/j.1365-3156.2012.02986.x) · PMID: [22519840](https://pubmed.ncbi.nlm.nih.gov/22519840/)

24. High level of resistance in the mosquito *Anopheles gambiae* to pyrethroid insecticides and reduced susceptibility to bendiocarb in north-western Tanzania

Natacha Protopopoff, Johnson Matowo, Robert Malima, Reginald Kavishe, Robert Kaaya, Alexandra Wright, Philippa A West, Immo Kleinschmidt, William Kisinza, Franklin W Mosha, Mark Rowland

Malaria Journal (2013-05-02) <https://doi.org/ghcqgf>

DOI: [10.1186/1475-2875-12-149](https://doi.org/10.1186/1475-2875-12-149) · PMID: [23638757](https://pubmed.ncbi.nlm.nih.gov/23638757/) · PMCID: [PMC3655935](https://pubmed.ncbi.nlm.nih.gov/PMC3655935/)

25. Estimating the annual entomological inoculation rate for *Plasmodium falciparum* transmitted by *Anopheles gambiae* s.l. using three sampling methods in three sites in Uganda

Maxwell Kilama, David L Smith, Robert Hutchinson, Ruth Kigozi, Adoke Yeka, Geoff Lavoy, Moses R

Kamya, Sarah G Staedke, Martin J Donnelly, Chris Drakeley, ... Steve W Lindsay
Malaria Journal (2014-03-21) <https://doi.org/gdkzs>
DOI: [10.1186/1475-2875-13-111](https://doi.org/10.1186/1475-2875-13-111) · PMID: [24656206](#) · PMCID: [PMC4001112](#)

26. CYP6 P450 Enzymes and ACE-1 Duplication Produce Extreme and Multiple Insecticide Resistance in the Malaria Mosquito *Anopheles gambiae*

Constant V. Edi, Luc Djogbénou, Adam M. Jenkins, Kimberly Regna, Marc A. T. Muskavitch, Rodolphe Poupartdin, Christopher M. Jones, John Essandoh, Guillaume K. Kétoh, Mark J. I. Paine, ... David Weetman

PLoS Genetics (2014-03-20) <https://doi.org/f56k77>

DOI: [10.1371/journal.pgen.1004236](https://doi.org/10.1371/journal.pgen.1004236) · PMID: [24651294](#) · PMCID: [PMC3961184](#)