

Multi-modal image registration: matching MRI with histology

Lejla Alic^{*ab}, Joost C. Haeck^b, Stefan Klein^{ab}, Karin Bol^{ab}, Sandra T. Van Tiel^b,
Piotr A. Wielepolski^b, Magda Bijster^c, Wiro J. Niessen^{abd}, Monique Bernsen^b, Jifke F Veenland^{ab},
Marion de Jong^c

^a BIGR – Biomedical Imaging Group Rotterdam, Departments of Radiology^b and Nuclear Medicine^c,
Erasmus MC, Rotterdam, P.O. Box 2040, 3000 CA Rotterdam, The Netherlands

^dFaculty of Applied Sciences, Delft University of Technology, Delft, The Netherlands

ABSTRACT

Spatial correspondence between histology and multi sequence MRI can provide information about the capabilities of non-invasive imaging to characterize cancerous tissue. However, shrinkage and deformation occurring during the excision of the tumor and the histological processing complicate the co registration of MR images with histological sections. This work proposes a methodology to establish a detailed 3D relation between histology sections and *in vivo* MRI tumor data. The key features of the methodology are a very dense histological sampling (up to 100 histology slices per tumor), mutual information based non-rigid B-spline registration, the utilization of the whole 3D data sets, and the exploitation of an intermediate *ex vivo* MRI.

In this proof of concept paper, the methodology was applied to one tumor. We found that, after registration, the visual alignment of tumor borders and internal structures was fairly accurate. Utilizing the intermediate *ex vivo* MRI, it was possible to account for changes caused by the excision of the tumor: we observed a tumor expansion of 20%. Also the effects of fixation, dehydration and histological sectioning could be determined: 26% shrinkage of the tumor was found. The annotation of viable tissue, performed in histology and transformed to the *in vivo* MRI, matched clearly with high intensity regions in MRI. With this methodology, histological annotation can be directly related to the corresponding *in vivo* MRI. This is a vital step for the evaluation of the feasibility of multi-spectral MRI to depict histological ground-truth.

Keywords: multi-modal registration, MRI, histology, tumor, tissue characterization

1. INTRODUCTION

Multi-modal MR imaging is often used for non-invasive tumor characterization. To assess what information can be derived from the different MRI sequences, determining the spatial relation between MR images and ground truth, e.g. histology, would be highly beneficial. The accuracy with which this registration can be achieved directly affects this analysis. Although the registration among *in vivo* imaging modalities is well established, accurate registration involving histology remains difficult and has been explored during the last decade. The early non-rigid registration approaches consist of registering MRI images using surface matching to cryosections¹, or volume marching to histological sections². At present, results that have been reported have been limited to global matching of MRI volume to small numbers of histological sections. To create histological images, tumor tissue undergoes excision, fixing by formalin consisting in a precipitation of the tissue proteins followed by dehydration. An important side effect of the precipitation is a decreasing ability of proteins to keep the water resulting in a deformation of the tissues and tearing artefacts. All these actions inevitably change the tumor morphology and thereby complicate the registration of *in vivo* MRI to histological sections.

Simple visual colocalization based on local tissue features³ does not take into account the 3D deformations of the histology slices and is prone to observer bias. In more advanced experimental⁴ and clinical studies⁵⁻⁷, point based registration derived from manual placement of control points along the boundary has been used. The limited number (usually up to 7) of control points restricts thereby the capability of this method to model complex elastic deformations due to tissue processing.

The aim of this work is to develop and implement a method for establishing a detailed 3D relation between *in vivo* MRI tumor data and histological sections. The following features needs to be taken into account:

- The large and complex deformations between *in vivo* MRI and histological sections.
- The artefacts due to the manual steps during histological processing, e.g. orientation differences, staining variations, non-tissue noise, damaged and missing sections.

The key features of our methodology are a very dense histologic sampling, up to 120 histology slices per tumor are acquired, the exploitation of intermediate *ex vivo* MRI, and a mutual information based non rigid B-spline registration based on whole 3D datasets.

2. METHODS

2.1 Study design

The study design consists of a number of independent imaging processes as shown in Fig. 1-A. First the rats were inoculated subcutaneously with CA20948 tumor and, when the tumor reached approximately 10 mm in diameter, imaged using a clinical 3T MRI scanner and a standard clinical wrist coil. After completing the MR examination, animals were sacrificed, and the tumors were gently dissected. During the dissection, the adjacent tumor surfaces were dyed using four tissue dyes in order to track its *in vivo* orientation. Subsequently, tumors were chemically fixed followed by slicing off a small portion along the longest tumor axis, and embedded in agar solution to prevent tumor movement during *ex vivo* MRI scanning. The plane for slicing histological sections and a reference plane for *ex vivo* MRI is defined as follows. The knowledge of the reference plane within *in vivo* MRI was used to resample *in vivo* data (shown on the top of Fig. 1-A.) resembling the *ex vivo* MRI orientation. Additionally, the reference plane limits the degrees of freedom during separate registration steps. Following the *ex vivo* MR examination, tumors were processed in histokinette, and subsequently embedded in paraffin from which 4 μ m sections were sliced and stained. Additionally, the stained sections were digitized at 20x magnification. The digital histology sections were rigidly registered afterwards to the adjacent section and stacked to a 3D set assuming the section thickness to be equal to the distance between the sections, in this case 60 μ m.

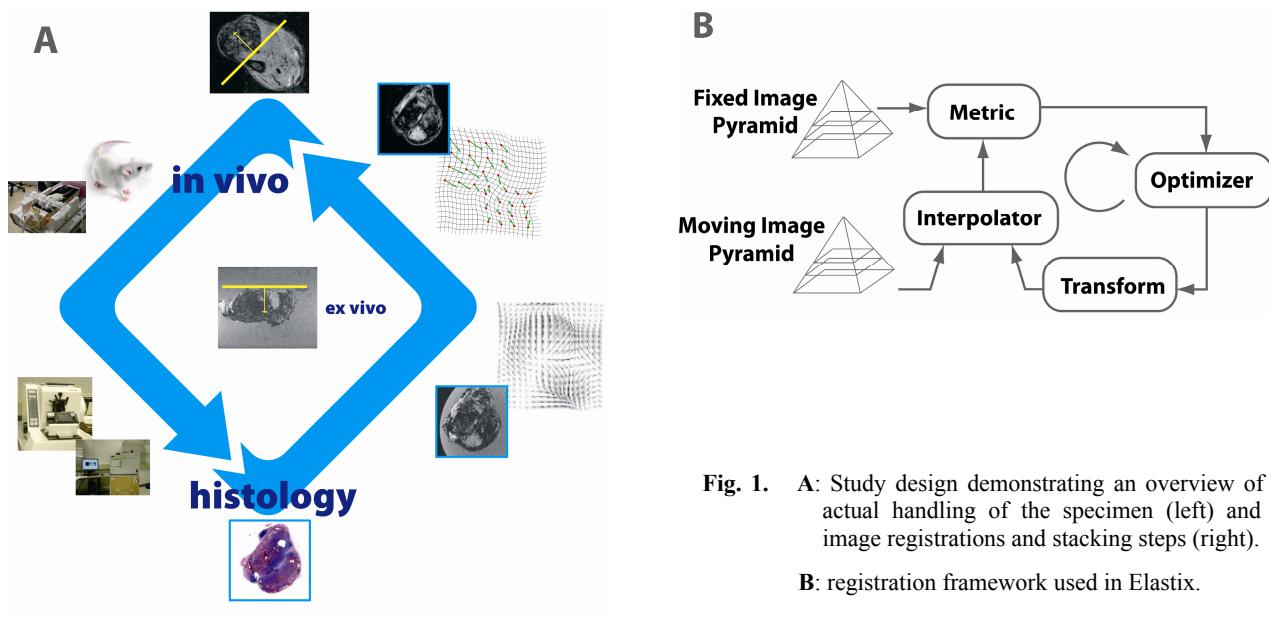


Fig. 1. A: Study design demonstrating an overview of the actual handling of the specimen (left) and the image registrations and stacking steps (right).

B: registration framework used in Elastix.

The registration consists of two steps: A) registration of 3D *in vivo* MRI to 3D *ex vivo* MRI, and B) registration of 3D stacked histology to 3D *ex vivo* MRI. All registration steps and the concatenation of the acquired transformations were performed using Elastix (<http://elastix.isi.uu.nl/>)⁸ which is implemented using the publicly available Insight Tool Kit (ITK)⁹⁻¹⁰ see also Fig. 1-B. To account for non-rigid deformations, a high-dimensional deformation model was used for all registrations with the displacement fields parameterized using B-spline models. Automatic 3D registrations were performed by maximization of the mutual information measure with a stochastic gradient descent algorithm¹¹.

2.2 Animal and tumor model

Lewis rats (Harlan-CPB, Austerlitz, The Netherlands) weighing 250-300g were inoculated subcutaneously in the right hind limb just above the ankle with human pancreatic tumor cells (CA20948 tumor). The animals were inspected daily for tumor growth and general appearance. The tumors were imaged when they reached approximately 10mm in diameter. Before the MR imaging, the animals were anesthetized by administration of Hypnorm (Janssen Pharmaceutica, Beerse, Belgium). Analgesia was achieved by intraperitoneal injection of ketamine (Alfasan, Woerden, The Netherlands). During the imaging, the animals were kept at a temperature of 38-39°C with a warm water mattress. After completing the MR examination, animals were sacrificed by subcutaneous injection of ketamine, and the tumors were gently dissected. The study was conducted with the approval of the Committee for Animal Research at our institution. In this paper the tumor of one rat is used.

2.3 Magnetic resonance imaging

Magnetic resonance imaging was performed using a clinical 3T MRI scanner (Signa Excite, GE Healthcare, USA). A standard clinical wrist coil (Flick Engineering Solutions B.V, The Netherlands) with an internal diameter of 5 cm was used for both *in vivo* MRI and *ex vivo* MRI acquisitions. The 3D susceptibility weighted (T2*W) images were used in the registration process because of the excellent structural detail of the tumor content, and a good signal to noise and contrast to noise ratio. *In vivo* MRI settings: Tr/Te = 23.18/8.88 ms, flip angle of 10°, and XYZ- resolution of 0.098 x 0.098 x 0.2 mm, and *ex vivo* MRI settings: Tr/Te = 42.17/20.88 ms, flip angle of 15°, and XYZ- resolution of 0.059 x 0.059 x 0.1 mm.

2.4 Specimen processing

During the dissection, adjacent tumors planes were dyed using four different tissue colours to define the origin of the tumor and track its orientation with the respect to *in vivo* position. The upper side (underneath the skin), two adjacent sides, and tail side were marked consistently. Immediately after dissection, tumors were placed in 200 cc 10% buffered formalin (Boom, The Netherlands). After the fixation process was completed, a thin section of the whole tumor volume was subsequently sliced off along the longest tumor axis in order to define the plane for histological sectioning and thereby create a reference plane for *ex vivo* MRI, see Fig 2. The specimen was then gently washed and transferred into a small test tube on a 1% agar solved in Phosphate Buffered Saline (PBS, AbD Serotec, MorphoSys, Munich, Germany) with the cutted plane down and subsequently embedded into agar to prevent tumor movement and to provide a water rich environment for *ex vivo* MRI. After the *ex vivo* MR examination, tumors placed in histokinette for 24h processing, and embedded in paraffin, from which two axial slices of 4µm were slices at 60µm intervals using a microtome. Serial sections were mounted on glass slide and stained with hematoxylin and eosin. The position of the individual slices was consequently recorded in a logbook. The slides were digitized using the NanoZoomer Digital Pathology (C9600, Hamamatsu, Japan) at 20x magnification. For registration purposes RGB images are converted to grayscale by eliminating the hue and saturation information while retaining the luminance.

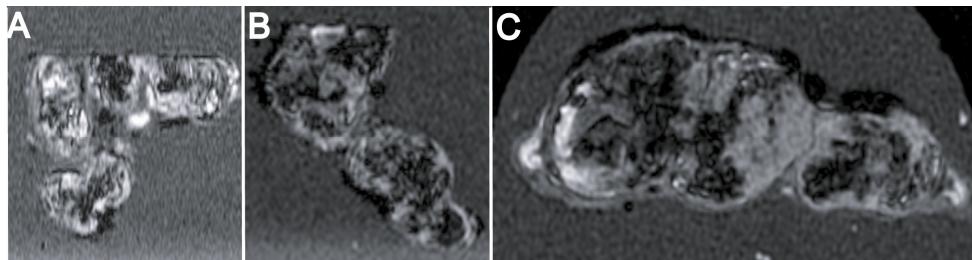


Fig. 2. An example of *ex vivo* MRI of dissected specimen showing coronal (A), sagittal (B), and axial (C) slices.

2.5 Registration

The overall goal is to register *in vivo* MRI images of the whole tumor to its histological sections. The study design in Fig. 1-A (with on the right hand side registration flow) shows a generalized registration approach. To achieve the goal, the registration is performed using an intermediate *ex vivo* MRI.

We perform registration as an optimization process that searches for an image transform allowing the closest similarity between consecutive images. The input is a pair of fixed (F) and moving (M) images. The framework iteratively passes M through 4 stages: *transform*, *metric*, *optimizer*, and *interpolator* as shown in Fig. 1-B. At a given iteration, the *transform* stage applies transform T_n on M which is then resampled onto a grid in the *interpolator* stage to yield $T_n(M)$. The *metric* stage computes the goodness of fit between F and $T_n(M)$ based on the similarity metric values from current and prior iterations allowing the *optimizer* to refine transform T_{n+1} .

Prior to the registrations 2D histology sections were first rigidly registered to its adjacent section, and then stacked to a 3D set. The global transformation consists of two distinct parts: A) registration of 3D *ex vivo* MRI to 3D *in vivo* MRI (referred to as ex2in), and B) registration of stacked 2D histological tissue sections to 3D *ex vivo* MRI (referred to as st2ex). The separate transformations were subsequently concatenated in order to accomplish coregistration of *in vivo* MRI to histological sections, referred to as st2in. The displacement field was parameterized with a B-spline model with mutual information of a 32-bin histogram similarity measure. Optimisation was performed using adaptive stochastic gradient descent algorithm (11). The registration steps are described in more detail below.

Stacking. Prior to 3D registrations the gray scale images of histological sections were stacked in a 3D data set. Owing the manual character of the process, sections were mounted in different orientations on glass slides. Therefore, in plane rigid registration was needed for each pair of histological sections before stacking. The registration was carried out using a multi resolution approach with a five level Gaussian image pyramid (16, 8, 4, 2, 1) for both the fixed and moving image. At the last level, the original images are used for registration. The number of iterations per section was set to 1000, and computation time per section varied between 60s and 70s. The obtained rigid transformations were then applied to the separate image channels, and the rgb-stack was constructed using MeVisLab application (MeVis Research GmbH, Bremen, Germany) with the adjustment of the slice thickness to 60 μ m which resembles the actual distance between the sections.

Registration of histology stack to *ex vivo* MRI (st2ex). Starting from the defined surface (as shown in Fig. 2) in *ex vivo* MRI, and carefully tracking the progress of the tissue sectioning facilitates registration of 3D stacked histology to *ex vivo* 3D-T2*^w MR image. The registration started as rigid, followed by an affine transformation allowing isotropic scaling to account for shrinkage, and finalized by Bspline registration. All registration steps were carried out using a multi resolution approach with a four level Gaussian image pyramid (8, 4, 2, 1) for both fixed (*ex vivo* 3D-T2*^w MRI) and moving (3D histology stack) images. The final grid spacing was specified to 4mm in all dimensions. The moving image derivative in Z-direction was set to 0 in order to favour the in plane registration and therefore enable managing distortions due to the manual processing of the histological sections. The number of iterations was set to 500, and computation time of the separate registration steps was: 145s, 127s and 367s, respectively.

Registration of *ex vivo* MRI to *in vivo* MRI (ex2in). The second stage in the registration process was to perform the coregistration of *ex vivo* 3D-T2*^w image and *in vivo* 3D-T2*^w image. The registration started as rigid, followed by affine allowing isotropic scaling to account for shrinkage, and finally by an elastic registration using a multi resolution approach with a recursive Gaussian image pyramid of four (8, 4, 2, 1), for both fixed (3D *in vivo* MRI) and moving (3D-

$T2^*$ w *ex vivo* MRI) image. The final grid spacing was specified to 4mm in all dimensions. The number of iterations was set to 500, and computation time of the separate registration steps was: 152s, 145s and 392s, respectively.

2.6 Experiments

As a first step, the accuracy of both registrations steps were visually inspected for all slices. To study the volume changes due to the affine and elastic transformations, the determinant of the affine transformation and the determinant of the Jacobian operator of the elastic transformations are computed for the ex2in and st2ex transformations separately. To evaluate the effect of registration, a volume of interest (VOI) representing ground-truth viable and necrotic cancerous regions were segmented in the 3D histology stack. Automatic segmentations were generated by transforming the manual segmentations from the histology images to the *ex vivo* domain and consecutively from the *ex vivo* domain to the *in vivo* domain using the transformations computed by the two registration steps. The distribution of the intensities in both transformed VOI's were compared with the distribution of the intensities when only a rigid transformation was applied. As a last step, a segmentation was performed based on the joint histogram distribution of the co-registered *ex vivo* MRI and 3D stacked histology and the joint histogram distribution of the *in vivo* MRI and *ex vivo* MRI and the correspondence with the ground-truth in histology was evaluated.

3. RESULTS

Images illustrating both registration steps (st2ex and ex2in) as well as the concatenation of those registrations (st2in) are shown in Figure 3. The first column shows the st2ex registration, the second column shows ex2in, and the third column shows the concatenated st2in registration. The registrations were visually inspected using a checker board approach as shown in the last row of Figure 3. The alignment of the tumor borders and of internal structures was evaluated as fairly accurate.

The overall volume preservation was estimated by the determinant of affine part of the ex2in and st2ex registrations. It turns out that a significant decrease in volume was observed during ex2in registration between *ex vivo* 3D- $T2^*$ w MRI and *in vivo* 3D- $T2^*$ w MRI, on average 20.6%. This is the inverted effect of the expansion of the tumor after excision from the surrounding tissue, in2ex. Quite the opposite, an increase of the tumor volume was observed during st2ex registration between 3D histology stack and *ex vivo* 3D- $T2^*$ w on average 26.9%. This corresponds to the inverted effect of dehydration of the specimen (ex2st). This provides an overall increase of tumor volume of 6.3%. To characterise the properties of the elastic deformation, the Jacobian operator is applied to the separate transformations (ex2in and st2ex), and its determinant is used to estimate the volume change. Table 1 summarizes the results of the elastic transformation. Concisely, although elastic transformation in ex2in preserves the volume while elastic transformation in st2ex show small shrinkage (on average), there is unambiguous difference in the behaviour of the different volume of interest (VOI). Moreover, st2ex transformation has a larger range in the Jacobian determinant, as can be observed in Table 1.

Table 1. Results of the elastic registration showing the range (and mean) of the Jacobian determinant for both transformations.

registration	Whole tumor	Viable VOI	Necrotic VOI
ex2in	1.00 (0.99-1.03)	1.00 (0.99-1.02)	1.01 (1.00-1.01)
st2ex	0.97 (0.50-1.00)	0.95 (0.50-1.00)	0.89 (0.50-1.00)

To evaluate the effect of registration, a volume of interest (VOI) representing ground-truth viable and necrotic cancerous regions were annotated in the 3D histology stack. In figure 4, the distributions of *ex vivo* MRI (first column) and *in vivo* MRI (second column) intensities segmented by the warped ground-truth volumes of interest (VOI) after rigid (first row) and B-spline (second row) registration. It can clearly be seen, that due to B-spline registration the 'viable VOI' intensity distribution shifts from Gaussian mixture to a more single Gaussian distribution.

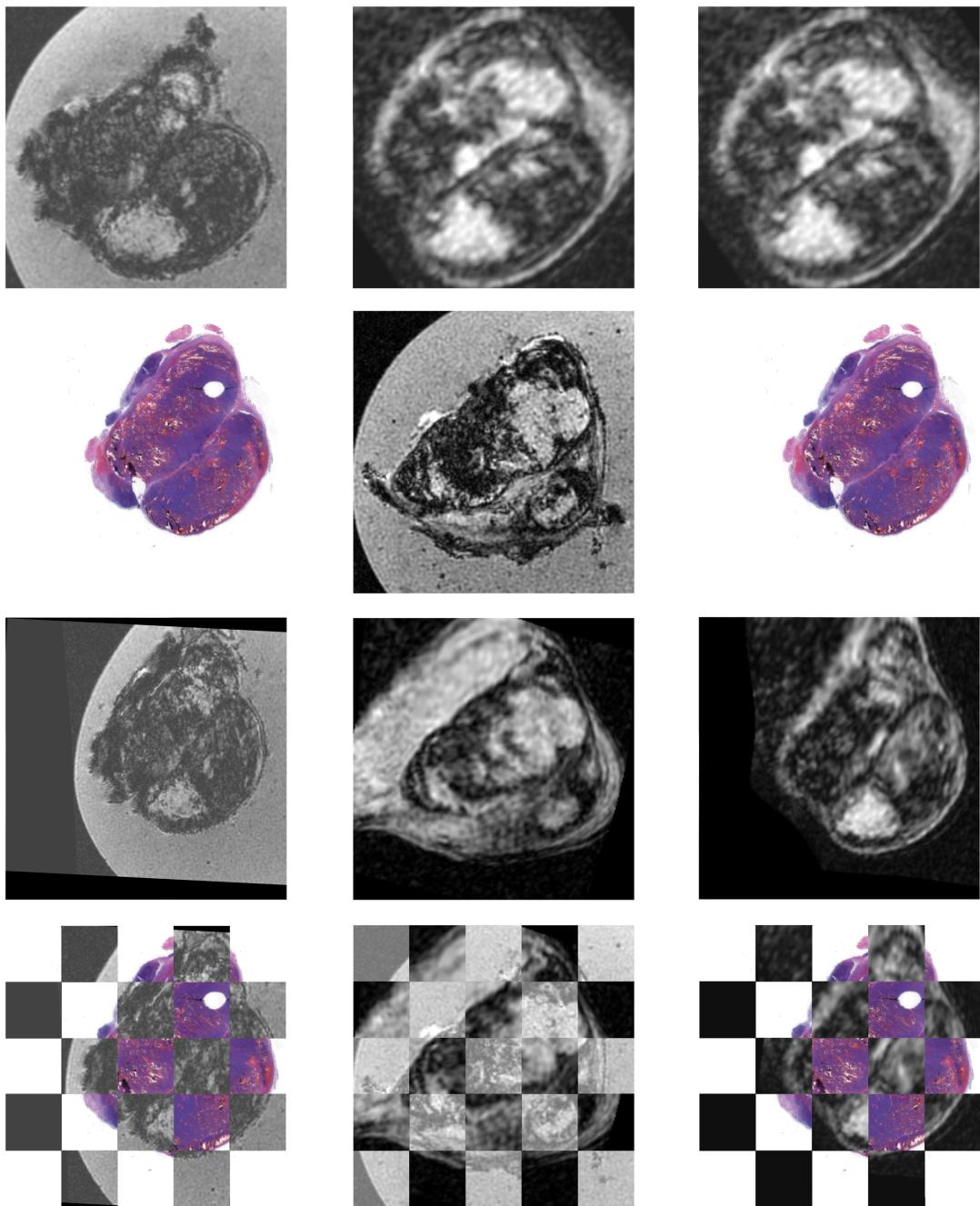


Fig. 3. Illustration of the registration process showing intermediate registration results (column one – st2ex and column two - ex2in), and the whole concatenated st2in registration (third column). *First row:* Fixed images. *Second row:* Moving images. *Third row:* Moving images in register with the fixed images. *Forth row:* Checkerboard view of corresponding fixed image (first row) and registered moving image (third row).

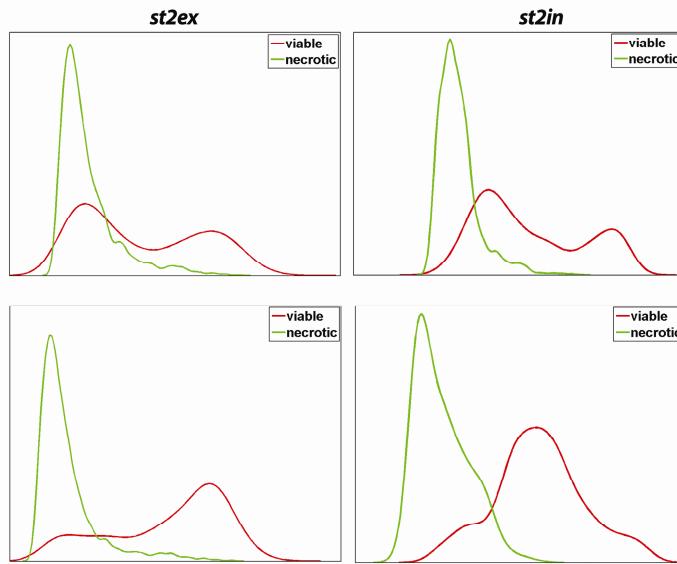


Figure 4. The distributions of *ex vivo* MRI (first column) and *in vivo* MRI (second column) intensities segmented by the warped ground-truth volumes of interest (VOI) after rigid (first row) and B-spline (second row) registration. VOIs were annotated in the histological sections. The necrotic VOI is shown in green, while the viable VOI is shown in red.

Figure 5 shows the co-registered *ex vivo* 3D-T2*w MRI (first column) and 3D stacked histology (second column). The third column shows the joint histogram distribution with horizontally the *ex vivo* intensities and vertically the histology gray-intensities. It can be seen that the gray-histology image is not suitable for the discrimination between viable and other tissue: both tissue types show similar gray-values, whereas in the *ex vivo* image viable tissue shows higher intensity values than non-viable tissue.

Figure 6 shows the co-registered *in vivo* 3D-T2*w MRI (first column) and *ex vivo* 3D-T2*w MRI (second column). The third column shows the joint histogram distribution with horizontally the *in vivo* intensities and vertically the *ex vivo* gray-intensities. The correspondence between the intensity values of the *in vivo* and *ex vivo* MRI can clearly be observed.

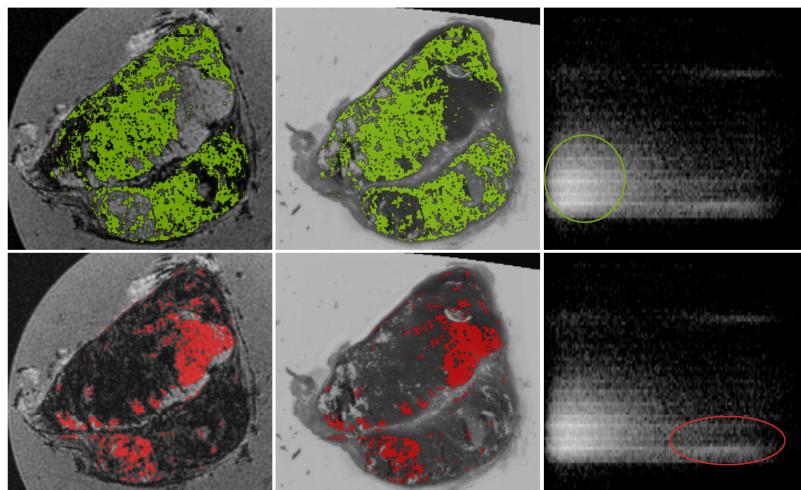


Fig. 5. The joint histogram distribution (third column) of co-registered *ex vivo* 3D-T2*W MRI (first column-fixed image) and 3D stacked histology (second column), referred to as st2ex. Green/red overlay in the images corresponds to the ellipses in the joint histogram distribution matching with non-viable (green) and viable (red) tissue.

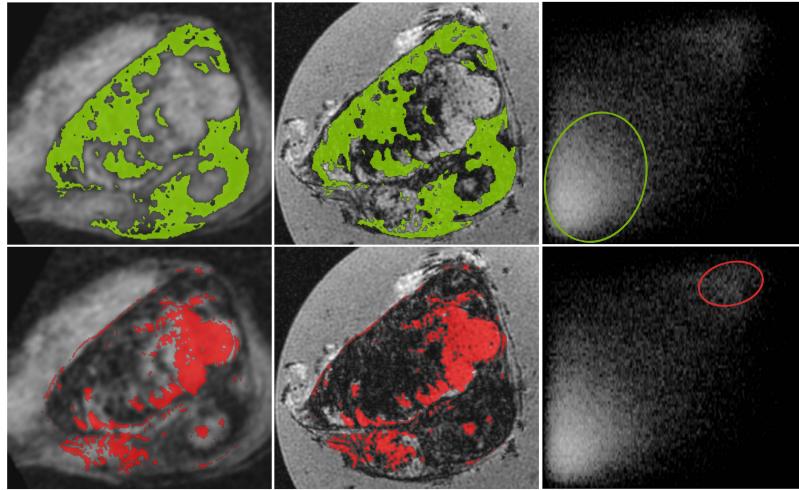


Fig. 6. The joint histogram distribution (third column) of co-registered *in vivo* 3D-T2*W MRI (first column-fixed image) and *ex vivo* 3D-T2*W MRI (second column), referred to as ex2in. Green/red overlay in the images corresponds to the ellipses in the joint histogram distribution matching with not viable (green) and viable (red) tissue.

4. DISCUSSION AND FUTURE DIRECTIONS

Non-rigid registration is essential in the process of correlating *in vivo* MRI data with histology. Matching of histological tissues sections to MR images of tumors is facilitated by using a multi-step registration process involving an *ex vivo* MRI scan of the tumor. Using this intermediate step, it is possible to account for changes caused by the excision of the tumor, the process of chemical fixation and the histological slicing.

Wang et al. (3) already showed connection between histology and MRI by verifying global tissue traits. Our preliminary results suggest that local registration between *in vivo* MRI as imaged by 3D-T2*W and histological sections is feasible when using a very dense histological sampling, a mutual information based non-rigid B-spline registration and the exploitation of an intermediate *ex vivo* MRI.

However, a more extensive study is needed in order to validate the registrations. Moreover, there are several possible modifications envisioned which should be explored because they can further increase the robustness and accuracy of the technique without significant increasing process time. For example, using the full colour range of the histology images could help automatic segmentation. Acquiring images with a stereotactic marker system¹² could facilitate the rigid registration of images. This will speed up the registrations and make manual annotations easier. The use of contrast agent in order to alter the tumor structural detail should be pursued as well.

We have proposed a methodology by which histological annotation can be related back to the corresponding *in vivo* MRI. This work is of crucial importance to build-up a database of multi-spectral MRI images and multi stained 3D histology which will serve as an essential and valuable source for understanding MR images and the feasibility of MRI to depict biological processes.

PUBLICATION ELSEWHERE

The work presented in this paper is not presented or published earlier.

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