#### Network Science

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Network Dynamical Process: SDS II

# 6.1 Functional Linkage Networks

### 6.1.1 Preliminary

Two genes may have the same function if their protein products interact or if they have very similar patterns of gene expression.

#### **Definition 1** FLN

An FLN [1, 2, 3] is a powerful medium for inferring gene function by integrating the evidence captured by protein protein interactions and gene-expression data. An FLN is a graph in which each node represents a protein or a gene. A node is labelled by the set of functions that annotate the gene; an edge in an FLN connects two genes if some experimental or computational procedure suggests that these genes might share the same function. Each edge in the FLN has a real-valued weight; the sign of the weight indicates whether the connected genes share or do not share the function, while the magnitude of the weight reflects our confidence in the edge.

#### 6.1.2 Gene Annotation Using Functional Linkage Networks

Goal: Determine if a collection of genes have some set of biological functions.

**Idea:** Use existing knowledge of what genes express what functions to predict functions of new genes.

#### Modeling approach:

- 1. Represent genes as vertices and connect genes if there is sufficient experimental/computational evidence indicating that they share biological functions as in Figure 6.1.
- 2. Each edge has a weight  $w \in [-1, 1]$  encoding the degree of of co-expression.
- 3. A new biological function is given, and it is known that some subset of the genes expresses this function.

4. Assign a state +1 to genes that express the new function and -1 to genes that do not express function f. Assign state 0 to states of remaining vertices.

$$k = \begin{cases} -1 & -\text{ not expressed;} \\ 0 & -\text{ no idea;} \\ 1 & -\text{ expressed.} \end{cases}$$

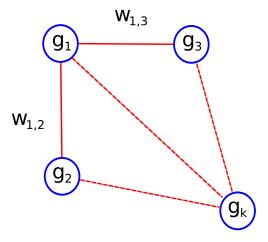


Figure 6.1: Gene function prediction.

Given the functional linkage network (FLN) and the initial configuration  $x = \{x_i\}$ , assign states  $\pm 1$  to all the genes with state 0 so as to minimize the energy function

$$E(x) = -\sum_{i,j} w_{i,j} x_i x_j$$

**Observation:** Locally, neighbor genes with identical states should be connected by a positive edge and neighbor genes with opposite states should be connected by a negative edge.

The algorithm in [3] computes an approximation to such a minimal state through an iterative sequential scheme.

- 1. Select (at random) a permutation  $\pi$  of the set V of vertices initially in state 0.
- 2. Repeat: Update the state of each of vertex  $v \in V$  asynchronously using the update sequence  $\pi$  and the vertex function

$$x_i \xrightarrow{f_i} sign \sum_{i,j} w_{j,i} x_i.$$

The algorithm stops when successive configurations are identical, i.e., "convergence".

**Observation:** This algorithm corresponds to a fixed point computation of a threshold SDS.

**Fact:** The SDS has only fixed points as attractors (i.e. no periodic orbits of period no smaller than 2).

## 6.2 NOR Permutation SDS

**Definition 2** NOR Function

$$nor(x_1, x_2, ..., x_k) = \begin{cases} 1 & \text{if all x's are 0;} \\ 0 & \text{otherwise.} \end{cases}$$

Can we have a fixed point?

For sequence  $\pi$ , function  $nor_{\pi}$ , x is a fixed point if nor(x) = x.

There is no fixed point.

**Note:**  $transient\ length$  is at most 1 for any point. Periodic points are in a one to one correspondence with the independent sets of the graph Y.

What characterizes a periodic point for *nor* permutation SDS?

Initial state:  $x = (x_1, x_2, \dots, x_k)$ .

To be periodic, a state can only contain isolated 1's.

## 6.3 Invertibility

When is  $F_{\pi} = F_{\pi(n)} \circ F_{\pi(n-1)} \circ \dots \circ F_{\pi(2)} \circ F_{\pi(1)}$  invertible? Assume  $k = \{0, 1\}$ .

#### **Definition 3** Invertible

take y, there is a unique x, such that F(x) = y. Each  $F_i$  should be invertible.

$$F_i(x) = (x_1, x_2, \dots x_{i-1}, f_i(x[i]), x_k)$$

That happens if  $g_i = f(-; \cap x_i)$ ,  $K \to K$  is a bijection function for each fixed  $\cap x_i$ . There are 4 functions.

**Implication:** If  $k \in \{0,1\}$ , and  $F_{\pi}$  is invertible,  $F_{\pi}^{-1} = F_{inv(\pi)}$ . In general,  $(F_{\pi}|_p)^{-1} = F_{inv(\pi)}|_p$ .

Fixed point ada independent.

#### Proposition 1.

Let  $F_{\pi}$  be a permutation SDS map and assume  $x \in Fix(F_{\pi})$ . Then  $x \in Fix(F_{\pi'})$  for all  $\pi \in S_Y$ .

#### Proposition 2.

A threshold SDS only has fixed points as attractors/limit sets.

Threshold function:  $f_k\{0,1\}^m \to \{0,1\}.$ 

$$F_{inv(\pi)} \circ F_{\pi}(x) = x$$

## 6.4 Schedule Equivalence

### 6.4.1 Functional Equivalence

Let  $\pi, \sigma \in S_Y$ , where  $F_{\pi} = F_{\sigma}$ .

$$F_{\pi} = F_{\pi(n)} \circ \ldots \circ F_{\pi(1)}$$

If  $\pi$  and  $\sigma$  differ by a transportation of consequtive, non-adjacent orientations, they induce the same SDS. Alternatively,  $\pi$  and  $\sigma$  induce the same SDS map if  $O_Y^{\pi}$  and  $O_Y^{\sigma}$  are the same acyclic orientations.

**Example:** in a  $Circ_4$ ,  $F_{\pi} = F_{\pi(1)} \circ F_{\pi(2)} \circ F_{\pi(4)} \circ F_{\pi(3)}$  and  $F_{\sigma} = F_{\sigma(1)} \circ F_{\sigma(4)} \circ F_{\sigma(2)} \circ F_{\sigma(3)}$  are functional equivalent.

The largest number of distinct permutation SDS maps  $\alpha(Y) = |Acyc(Y)|$ . Since

$$\alpha(Circ_n) = 2^n - 2,$$

we have  $\leq 2^n - 2$  SDS maps.

 $\alpha$  is turtle canonical.  $\alpha(Y) = \alpha(Y'_e) + \alpha(Y''_e)$ , where  $\alpha(Y'_e) = Y \setminus \{e\}$  and  $\alpha(Y''_e) = Y/\{e\}$ .

## 6.4.2 Other Equivalence

Dynamical Equivalence

Orbit Equivalence

# 6.5 List of Supplemental Notes

- circ4\_nor\_equivalences.pdf
- SDS\_Application.pdf

# References

- [1] N. Massjouni, C. Rivera, and T. M. Murali, "Virgo: Computational prediction of gene functions," *Nucleic Acids Research*, vol. 34, pp. W340–W344, 2006.
- [2] M. Henning, "Graph dynamical systems a mathematical framework for interactionbased systems, their analysis and simulations," in *Discrete Models in Systems Biology Workshop*, December 2008.
- [3] U. Karaoz, T. M. Murali, S. Letovsky, Y. Zheng, C. Ding, C. R. Cantor, and S. Kasif, "Whole-genome annotation by using evidence integration in functional-linkage networks," *Proceedings of the National Academy of Sciences*, pp. 2888–2893, 2004.