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Third generation EGFR TKIs in EGFR-mutated NSCLC: Where are we now and where are we going



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ABSTRACT

The therapeutic landscape of Non Small Lung Cancer (NSCLC) has been profoundly changed over the last decade with the clinical introduction of Epidermal Growth Factor Receptor (EGFR) tyrosine kinase inhibitors (TKIs) and the discovery of EGFR activating mutations as the major predictive factor to these agents. Despite impressive clinical activity against EGFR-mutated NSCLCs, the benefit seen with 1st and 2nd generation EGFR TKIs is usually transient and virtually all patients become resistant. Several different mechanisms of acquired resistance have been reported to date, but the vast majority of patients develop a secondary exon 20 mutation in the ATP-binding site of EGFR, namely T790M. The discovery of mutant-selective EGFR TKIs that selectively inhibit EGFR-mutants, including T790M-harboring NSCLCs, while sparing EGFR wild type, provide the opportunity for overcoming the major mechanism of acquired resistance to 1st and 2nd generation EGFR TKIs, with a relatively favorable toxicity profile. The development of this novel class of EGFR inhibitors poses novel challenges in the rapidly evolving therapeutic paradigm of EGFR—mutated NSCLCs and the next few years will witness the beginning of a new era for EGFR inhibition in lung cancer. The aim of this paper is to provide a comprehensive overview of the increasing body of data emerging from the ongoing clinical trials with this promising novel therapeutic class of EGFR inhibitors.

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1. Introduction

The therapeutic landscape of Non Small Lung Cancer (NSCLC) has been profoundly changed over the last decade with the clinical introduction of Epidermal Growth Factor Receptor (EGFR) tyrosine kinase inhibitors (TKIs) and the discovery of EGFR activating

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mutations as the major predictive factor to these agents (Russo et al., 2015). Despite impressive clinical activity against EGFRmutated NSCLCs and proved superiority over chemotherapy in the 1st line setting in molecularly-selected patients, the benefit seen with 1st and 2nd generation EGFR TKIs is usually transient and virtually all patients become resistant after approximately 9-13 months (Mok et al., 2009; Zhou et al., 2011; Wu et al., 2015; Wu et al., 2014; Sequist et al., 2013). Several different mechanisms of acquired resistance have been reported to date, but the vast majority of patients (41-62%) develop a secondary exon 20 mutation in the ATP-binding site of EGFR, namely T790M (Hata et al., 2013; Kuiper et al., 2014; Sequist et al., 2011; Yu et al., 2013). Various strategies have been studied to overcome T790M mutation, but only few have reported clinical meaningful benefit. The discovery of mutant-selective EGFR TKIs that selectively inhibit EGFR-mutants, including T790M-harboring NSCLCs, while sparing EGFR wild type, provide the opportunity for overcoming the major mechanism of acquired resistance to 1st and 2nd generation EGFR TKIs, with a relatively favorable toxicity profile. To date, three different 3rd generation EGFR TKIs are in advanced development stage, with the recent FDA and EMA approval of Osimertinib and the FDA breakthrough designation of Rociletinib and Olmutinib. The development of this novel class of EGFR inhibitors poses novel challenges in the rapidly evolving therapeutic paradigm of EGFRmutated NSCLCs and the next few years will witness the beginning of a new era for EGFR inhibition in lung cancer. The aim of this paper is to provide a comprehensive overview of the increasing body of data emerging from the ongoing clinical trials with this promising novel therapeutic class of EGFR inhibitors.

2. Mechanisms of acquired resistance to 1st and 2nd generation EGFR inhibitors

After treatment with first or second generation EGFR TKIs, virtually all patients after approximately 12 months develop acquired resistance (AR). Different mechanisms of AR to EGFR TKIs have been reported to date and may broadly divided in two subgroups: pharmacological and biological mechanisms, including alterations in the drug target, bypass track mechanisms, phenotypic/histologic changes and downstream signaling pathways alterations (Camidge et al., 2014; Lovly and Shaw, 2014). In 2005 two different groups first reported the development of a secondary mutation in the EGFR TK domain causing a substitution of a threonine with a methionine at the 790 site of the exon 20 (T790M mutation), leading to resistance to first generation EGFR TKI Gefitinib after initial response (Kobayashi et al., 2005a; Pao et al., 2005). The T790M mutation is the major mechanism of AR, since it is detected in vivo in approximately 41-62% of the cases at re-biopsy after acquired resistance to 1st generation EGFR TKIs (Hata et al., 2013; Kuiper et al., 2014; Sequist et al., 2011; Yu et al., 2013). Following preclinical ies reporting the development of T790M mutation even after irreversible EGFR TKIs (Godin-Heymann et al., 2008), a recent retrospective analysis confirmed T790M mutations as the major mechanism of AR, even in patients became resistant to the irreversible EGFR inhibitor Afatinib, with an overall frequency of ~50% (Wu et al., 2016). The development of T790M is associated with AR to both 1st and 2nd generation EGFR TKIs by increasing the receptor affinity to the ATP and consequently limiting the potency of these ATP-competitive kinase inhibitors (Yun et al., 2008). The emergence of T790M mutations in brain metastases is less frequent than extracranial sites (17% vs. 41% in a retrospective analysis), suggesting that other mechanisms of AR may be more relevant in the central nervous system (CNS), such as poor drug exposure (Hata et al., 2013).

The identification of T790M mutation after AR to EGFR TKIs raised the question whether these mutations were the results of a selection process of pre-existing clones T790M+ due to the selective pressure of EGFR TKIs or were acquired during treatment because of novel genetic and epigenetic alterations (Suda et al., 2012). In favor of the "selection model" there are some preclinical and clinical evidences of the presence of pre-existing resistant clones before treatment, including T790M+, albeit their exact frequency and clinical significance is not yet fully understood (Maheswaran et al., 2008; Costa et al., 2014; Su et al., 2012; Ye et al., 2013; Yu et al., 2014; Lee et al., 2014). Indeed, preclinical data indicate that tumors with AR likely harbor mixed populations of drugsensitive and drug-resistant cells with differential growth rates, with the T790M+ cells showing slower growth than T790M- ones (Chmielecki et al., 2011). These data are further supported by clinical evidences indicating a more indolent natural history and a longer post-progression survival among T790M harboring patients (Hata et al., 2013; Oxnard et al., 2011; Uramoto et al., 2012).

In few cases the development of acquired resistance may involve the development of other rarer secondary mutations, including D761Y (Balak et al., 2006), L747S (Costa et al., 2007) and L854A (Bean et al., 2008).

Acquisition of a secondary mutation is not the solely mechanism of AR to EGFR TKIs, but different preclinical and clinical studies, but EGFR-mutated tumors may escape the EGFR blockage in several other ways, including: Small Cell Lung Cancer (SCLC) transformation (2–14%) (Kuiper et al., 2014; Sequist et al., 2011; Yu et al., 2013; Niederst et al., 2015a), Epithelial to Mesenchimal Transition (EMT) (Sequist et al., 2011; Byers et al., 2013), MET amplification (4–5%) (Sequist et al., 2011; Yu et al., 2013; Takezawa et al., 2012), HER2 amplification (12–13%) (Yu et al., 2013; Takezawa et al., 2012), PIK3CA mutations (5%) (Sequist et al., 2011), AXL activation (Zhang et al., 2012), BRAF mutations (Ohashi et al., 2012), and NFkB activation (Bivona et al., 2011). In some instances, multiple mechanisms may operate simultaneously, albeit in the majority of cases seems to be mutually exclusive (Yu et al., 2013).

The better knowledge of the inherited mechanisms of AR to EGFR TKIs contributed to the development of rationally strategies to overcome resistance.

Given its high frequency, it is not surprising that different strategies have been developed to restore the sensitivity of EGFR T790M-mutant cells to EGFR inhibition. To date, several different strategies have been evaluated in both preclinical and clinical models, but only few have been proved effective *in vivo*. Indeed, some promising *in vitro* strategies, such as the use of irreversible EGFR inhibitors ("second generation" EGFR TKIs) against T790M-mutants (Kobayashi et al., 2005b; Kwak et al., 2005; Engelman et al., 2007), failed *in vivo* due to a narrow therapeutic window (Sequist et al., 2010; Landi et al., 2014; Reckamp et al., 2014; Miller et al., 2012). Moreover, the use of a vertical blockage with an EGFR TKI and a monoclonal anti-EGFR antibody (i.e. Afatinib and Cetuximab), albeit proved effective against T790M-harboring tumors, is limited by the unfavorable toxicity profile (Janjigian et al., 2014).

The use of mutant-selective, EGFR wild type sparing, "third generation" inhibitors is an emerging therapeutic strategy in patients with AR to 1st and 2nd generation inhibitors. These newer agents differ from quinazoline-based reversible (Gefitinib and Erlotinib) and irreversible EGFR TKIs (Afatinib, Dacomitinib, Neratinib), because of their aminopyrimidine scaffold and have been specifically developed to target EGFR mutations, including T790M, with only minimal activity against wild type EGFR. The first-in-class third generation EGFR TKI reported was WZ4002, which did not progress to clinical trials (Zhou et al., 2009), but now a few different 3rd generation EGFR TKIs are in active clinical development in NSCLC. In the following sections we will provide an overview of the

latest preclinical and clinical data on three of the most promising agents of this novel class of EGFR TKIs.

3. Osimertinib (AZD9291)

Osimertinib (AZD9291; AstraZeneca) is a novel irreversible, small molecule inhibitor, developed to target both sensitizing and resistant mutant forms of the EGFR while sparing the wild type form of the receptor (Osimertinib, 2016).

This mono-anilino-pyrimidine compound is structurally and pharmacologically distinct from all other TKIs. Osimertinib binds irreversibly to the EGFR kinase by targeting the cysteine-797 residue in the ATP binding site via covalent bond formation (Ward et al., 2013; Finlay et al., 2014). The drug exhibits nearly 200 times greater potency against L858R/T790M than wild type EGFR. Studies conducted in vivo, revealed that Osimertinib is metabolized to produce at least two circulating metabolite species, AZ5104 and AZ7550. AZ7550 had a comparable potency and selectivity profile, while AZ5104 is more potent against mutant and wild type EGFR forms (Cross et al., 2014; Yosaatmadja et al., 2015).

In vitro studies indicate that Osimertinib has activity against mutant EGFR, including T790M+, but selectivity margin against wild type EGFR. The drug has minimal off-target activity, with a limited number of non-HER kinases inhibited, but conserves activity against HER2/4. In contrast, Osimertinib is not effective against lines harboring non-T790M resistance, such NRAS mutations, MET amplification, and epithelial-to-mesenchymal transition (EMT) (Cross et al., 2014).

Studies conducted *in vivo* xenograft models demonstrated good bioavailability of Osimertinib and moderate clearance with half-life of 3 h after oral dosing in the mouse. The circulating metabolites have a similar half-life and the total exposure level (AUC) were approximately 68% and 33% compared with parent compound for AZ7550 and AZ5104 respectively. Quantitative whole body autoradiography (QWBA) studies in rat brain indicate that Osimertinib has a brain-to-blood ratio of up to 2 over the first 24 h, suggesting the potential of AZD9291 to penetrate the brain (Cross et al., 2014). Studies in mouse models indicate that Osimertinib is distributed in the CNS at greater extent than Gefitinib, Rociletinib and Afatinib (Ballard et al., 2016) and has potential activity also against leptomeningeal metastases (LMs) in both EGFR TKI-naïve and TKI-resistant tumors (Nanjo et al., 2016).

The AURA trial was a phase I/II study evaluating the safety, tolerability and activity of Osimertinib in patients with EGFR-mutated NSCLC progressing after a previous EGFR TKI. The phase I component of the study included a dose escalation part (Osimertinib 20 mg to 240 mg), using a rolling six design, and an expansion part with multiple biomarker-guided cohorts of patients. Before enrollment in the expansion cohort, the T790M status of the tumor was mandatory. The recommended phase II dose (RP2D) was 80 mg once daily. The phase II (extension part) component of the study enrolled T790M+ NSCLCs progressing on a previous EGFR TKI at the dose of 80 mg/d. A total of 253 patients (31 in the dose-escalation cohorts and 222 in the expansion cohort) were enrolled. EGFR T790M was detected in 62% of patients enrolled in the expansion cohort (Jänne et al., 2015). A maximum tolerated dose (MTD) was not found, however at the 160 mg and 240 mg dose levels there were an increase in the incidence and severity of adverse events (AEs) associated with EGFR wild type inhibition (namely, skin and gastrointestinal AEs), therefore the dose of 80 mg once daily was considered the RP2D dose. Of the 239 patients evaluated for response 51% had PR or CR, 33% SD and 14% PD. The disease control rate (DCR) was 84% (95% CI, 79-88). Among patients with EGFR T790M, the objective response rate (ORR) was 61% (78 of the 127 patients; 95% CI, 52-70), and the DCR was 95% (121 of the 127 patients; 95% CI, 90–98). Although the

data published were immature, the median PFS was 8.2 months in the overall population, 9.6 months in the T790M+ group (95% CI, 8.3 to not reached) and 2.8 months (95% CI, 2.1–4.3) in patients with no detectable EGFR T790M (Jänne et al., 2015). These promising results were recently confirmed in the updated analysis of the phase I expansion cohort and the pooled analysis of the phase II studies AURA extension and AURA2, evaluating Osimertinib at 80 mg/daily in T790M+ TKI-pretreated NSCLCs. Osimertinib was associated with a 71% and 66% ORR, 9.7 months and 11.0 months PFS, respectively. The safety profile was relatively favorable with the most common AEs being skin rash (37–41% of all grades, 0–1% G3/4) and diarrhea (35–38% of all grades, 1–2% G3/4) (Yang et al., 2016a). Moreover, quality of life (QoL) analysis of the AURA2 trial revealed a significant improvement in functioning and health status/QoL from baseline assessments up to the first 36 weeks (Rudell et al., 2016).

The intriguing preclinical intracranial activity of Osimertinib prompted the development of the phase I BLOOM trial, evaluating Osimertinib and AZD3759 in EGFR-mutated NSCLC patients with BMs and LMs. At the recent 2016 ASCO annual meeting, the preliminary results of the Cohort 1 of the study (EGFR-mutated NSCLCs with LMs) were presented, reporting a promising activity with Osimertinib 160 mg/d in this unfavorable prognostic subgroup of patients (Yang et al., 2016b). The Cohort 2 of the study is enrolling T790M+ NSCLC patients with LMs.

Osimertinib is being evaluated in multiple clinical trials across different settings and combinations, to understand its potential benefit for overcoming newly identified forms of resistance. The ongoing phase III trials are summarized in Table 1.

A phase III, open Label, randomized study (AURA3, NCT02151981) is ongoing. Designed to assess the efficacy of Osimertinib compared with platinum-based doublet chemotherapy for patients with locally advanced or metastatic NSCLC whose disease has progressed with previous EGFR TKIs and whose tumors harbor a T790M mutation. Secondary outcome measurements are ORR, Duration of Response (DoR), DCR, and OS. On 18 July 2016 AstraZeneca announced that the AURA3 Phase III trial met its primary endpoint, demonstrating superior PFS compared to standard platinum-based doublet chemotherapy. The randomized double blind phase III FLAURA trial (NCT02296125) is evaluating Osimertinib in the first line setting versus Gefitinib or Erlotinib.

The ongoing TATTON study (NCT02143466), a multi-arm Phase Ib study, is designed to assess the safety, tolerability, pharmacokinetics and preliminary anti-tumor activity of Osimertinib in combination with ascending doses of novel therapeutics, including Durvalumab (anti-PDL1 inhibitor), Selumetinib (anti-MEK inhibitor) and Savolitinib (anti-MET inhibitor), in patients with EGFR-mutated advanced NSCLC who have progressed following therapy with an EGFR TKI. Finally, the phase III study CAURAL study (NCT02454933) will investigate Osimertinib in combination with Durvalumab as a potential second-line treatment for EGFR-mutated NSCLC patients carrying the T790M mutation to assess the efficacy and safety of Osimertinib in combination with Durvalumab versus Osimertinib monotherapy.

On 13 November 2015, Osimertinib received FDA approval for patients carrying a T790M mutation and whose disease has progressed after treatment with other EGFR TKI, followed by EMA approval on 2 February 2016, making Osimertinib the first new drug approved under the EMA expedited process (Osimertinib, 2016)

4. Rociletinib (CO-1686)

Rociletinib (CO-1686; Clovis Oncology Inc.) is a novel, oral, mutant-selective, covalent EGFR inhibitor studied for the treatment of NSCLC.

Table 1 Clinical data of efficacy with Osimertinib in T790M-positive NSCLC.

Study	Phase	n (T790M+ pts)	Osimertinib Dose	Line(s) of treatment	ORR	DCR	Median PFS
AURA (Jänne et al., 2015)	1	138	20-240 mg	≥2	62%	95%	9.6 mos
AURA extension (Yang et al., 2016a)	2	201	80 mg	≥2	62%	90%	12.3 mos
AURA 2 (Rudell et al., 2016)	2	199	80 mg	≥2	70%	92%	9.9 mos
AURA 3 (Yang et al., 2016b)	3	279	80 mg	2	71%	93	10.1 mos

Rociletinib is a potent 2,4-disubstituted pyrimidine molecule that covalently modify the Cys797 in the ATP-binding pocket of the EGFR kinase domain. Studies conduced *in vitro* demonstrated that CO-1686 is a potent inhibitor of L858R/T190M and approximately 22-fold more selective than *wild type* EGFR. Twenty-three targets inhibited more than 50% at 0.1 µmol/L; EGFR del19-, T790M-, L858R/T790M-, and L858Rmutant kinases have the highest degree of inhibition, however, other kinase targets were observed to be inhibited at lower potency, including focal adhesion kinase (FAK), CHK2, ERBB4, and Janus-activated kinase 3 (JAK3) (Walter et al., 2013; Tjin Tham Sjin et al., 2014).

In vitro studies in NSCLC cell lines expressing EGFR mutants (T790M mutation, exon 19 deletion E746-A750, L858R/T790M double mutation), demonstrated that Rociletinib potently inhibits proliferation in the mutant EGFR NSCLC cells with Growth inhibition (Gl $_{50}$) values ranging from 7 to 32 nmol/L. In comparison, the Gl $_{50}$ value for wild type EGFR cells was 547 nmol/L. CO-1686 also inhibits NRAS and KRAS mutations in WT EGFR cells at concentrations of 4275 and 1806 nmol/L, respectively. The efficacy of CO-1686 was also examined against other EGFR mutants including the exon 18 mutation G719S, an exon 19 insertion mutant (I744-K745insKIPVAI), an exon 20 insertion (H773-V774HVdup), and the exon 21 mutation L861Q. Rociletinib was active against these rare mutants also, with the exception of exon 20 insertions (Walter et al., 2013).

The TIGER program was an accelerated clinical development program for Rociletinib in patients with mutant EGFR NSCLC in different therapeutic settings (Table 2).

The phase I/II dose-finding TIGER-X evaluated Rociletinib in EGFR-mutated NSCLCs progressing after previous 1st/2nd generation EGFR TKIs. The study consisted of two parts, the phase I dose-escalation component followed by the phase 2 expansion part (requiring T790M status assessment before enrollment) to assess efficacy at 500 mg twice daily, 625 mg twice daily, and 750 mg twice daily. Two different formulations of Rociletinib were initially developed, the free-base form (entered the clinic in March 2013) and the hydrogen bromide salt (HBr), designed to improve the pharmacokinetic profile, introduced during dose escalation from August 2013 that replaced the free-base form. Primary objectives of the phase I were safety, toxicity profile, and pharmacokinetic characteristics of Rociletinib. Secondary end points included ORR, DoR, PFS, and QoL. Primary end points of the phase II part were ORR and DoR, whereas secondary and exploratory end points were the same as in the phase I component. A MTD was not defined in the phase I portion. Preliminary results, based on 130 patients enrolled, reported an ORR among 46 patients with centrally confirmed T790M-positive tumors 59%, with a DCR of 93%. Response rates were similar between patients with exon deletion 19 or L858R EGFR mutation. The estimated median PFS was 13.1 months. The

ORR among 17 T790M-negative patients was 29%, with a DCR of 59% and a median PFS of 5.6 months. Among 20 patients whose tumors were not assessable for T790M the ORR was 15%. The predominant grade 3 AE was hyperglycemia (22%), which was generally manageable and did not result in treatment discontinuation (Sequist et al., 2015). Preclinical studies suggest that hyperglycemia is caused by a Rociletinib metabolite that inhibits the type I insulin-like growth factor receptor (IGF-IR) and induces activation of the IGFIR pathway, a proposed resistance mechanism for EGFR inhibition (Cortot et al., 2013), although the contribution of the IGF-IR inhibitory effect of Rociletinib to its antitumor activity is currently unknown.

These preliminary results prompted the establishment of a rapid developmental program (Table 2) and granted, in May 2014, the U.S. FDA Breakthrough Therapy designation for the treatment of mutant NSCLCs with the T790M mutation after progression on EGFR-directed therapy.

Unfortunately, on May 2016 Clovis inc. announced the interruption of clinical development of Rociletinib and terminated enrollment in all ongoing sponsored clinical studies based on the unsatisfactory results of a pooled analysis of TIGER-X and TIGER-2 trials submitted to the US FDA, reporting a 32% ORR (95% CI 25, 40) with a median DoR of 8.8 months in T790M+ patients treated with Rociletinib at the doses of 625 mg BID and 23% ORR (95% CI 14, 34) with a median DoR of 9.1 months in patients who received 500 mg BID (Anon, 2016).

5. Olmutinib (BI1482694/HM61713)

Olmutinib (BI 1482694/HM61713; Hanmi Pharmaceuticals and Boehringer Ingelheim) is a 3rd generation EGFR TKI, developed to specifically inhibit EGFR mutants, including T790M, while sparing wild type EGFR.

The safety and pharmacokinetic profile of this agent was evaluated in the phase I/II HM-EMSI-101 study, a multicenter trial conducted in Korean patients previously treated with at least one EGFR TKI. At the recommended phase II dose (RP2D: 800 mg qd), all eligible patients had to have confirmed T790M mutation in the tumor. The primary endpoint was ORR; secondary endpoints included DOR, DCR, PFS and safety. Data of patients treated at the RP2D of 800 mg qd were recently presented: 54% confirmed ORR by independent review and 90% DCR in T790M-positive TKI-pretreated NSCLCs with a favorable safety profile (Park et al., 2016a).

ELUXA 1 (HM-EMSI-202) is an ongoing pivotal Phase II global clinical trial, designed to further investigate the efficacy and safety of Olmutinib in patients T790M-positive NSCLC with acquired resistance after first-line EGFR TKIs. Primary endpoint is ORR according to RECIST 1.1, while secondary endpoints are DCR, DoR, PFS, OS, TTP, tumor shrinkage, patients reported outcomes (PROs), and safety

Table 2Ongoing phase III trials evaluating Osimertinib in NSCLC.

Study	Phase	Setting/population	Treatment(s)	Primary endpoint
FLAURA	III	EGFR-mut+ 1st line	Osimertinib vs. Gefitinib/Erlotinib	PFS
ADAURA	III	EGFR-mut+ Adjuvant setting	Osimertinib vs.Placebo	DFS
CAURAL	III	T790M+, EGFR-TKI-pretreated	Osimertinib + Durvalumab vs. Osimertinib	PFS
ASTRIS	III	T790M+, EGFR-TKI-pretreated	Osimertinib	Efficacy and safety in a real world setting

Table 3Initial development program of Rociletinib.

Study	Phase	Setting population	Treatment(s)	Primary endpoint(s)
TIGER-1	II/III	EGFR-mut+ TKI-naive	Rociletinib vs. Erlotinib	PFS
TIGER-2	II	T790M+ prior TKI	Rociletinib	ORR
TIGER-3	III	T790M+ prior TKI and platinum doublet	Rociletinib vs. mono-chemotherapy	PFS
NCT02630186	Ib/II	EGFR-mut+ 1st line and pretreated	Rociletinib + Atezolizumab	Safety and activity
NCT02580708	I/II	EGFR-mut+ 1st line and pretreated	Rociletinib + Trametinib	Safety and activity

(Park et al., 2016b). The ELUXA pivotal trial program (Table 3) will include also the initiation of two Phase III studies in 2016 comparing Olmutinib with chemotherapy in previously treated T790M+NSCLCs (ELUXA 2) and in the first line setting versus Afatinib (ELUXA 3).

On December 2015, the US FDA granted breakthrough therapy designation to Olmutinib based on the promising results from the Phase I/II HM-EMSI-101 clinical trial.

6. Other 3rd Generation EGFR TKIs

EGF816 (Novartis Pharmaceuticals) is a covalent, irreversible, EGFR TKI with high *in vitro* activity against EGFR mutants, including T790M. Preliminary results of the multi-arm phase I/II study (NCT02108964) in advanced NSCLC patients harboring T790M mutation showed a manageable safety profile, with the most common grade 3/4 AEs were maculo-papular rash (14%), anemia (6%), and diarrhea (6%), and an intriguing clinical activity with a confirmed 44% ORR and a 91% DCR. Median PFS was 9.2 months (95% CI 9.0-NE) (Tan et al., 2016).

ASP8273 (Astellas Pharma Inc) is another small molecule mutant-selective, irreversible EGFR inhibitor with higher *in vitro* affinity against EGFR-mutants than wild type that is being evaluated in phase II/III studies in EGFR-mutated NSCLCs. At the recommended phase II dose of 300 mg/d, ASP8273 showed a robust antitumor activity in both subjects with pre-treated EGFR mutation-positive NSCLC and T790M+. For the 45 subjects treated with ASP8273 300 mg with evaluable data, DCR was 62%, with 16 patients achieving a PR and 12 a SD. For the 40 T790M+ subjects with evaluable data, DCR was 65% with 15 PR and 11 SD. Preliminary median PFS was 6.7 months in both overall population and T790M+ patients (Yu HA et al., 2016). The ongoing phase III trial SOLAR will compare the activity of ASP8273 in the first line setting versus 1st generation EGFR TKIs.

Brigatinib (ARIAD Pharmaceuticals), also known as AP26113, is dual ALK/EGFR with preclinical activity against the oncogenic ALK fusion protein and mutants resistant to Crizotinib, but also against activated and T790M-mutant EGFR (Rivera et al., 2012). Brigatinib received Breakthrough Therapy designation from the FDA in October 2014 for the treatment of patients with ALK-rearranged NSCLC with acquired resistance to Crizotinib and data from the phase I/II study (NCT01449461) in EGFR-mutated NSCLC are awaited.

Finally, PF-06747775 (Pfizer) and Avitinib (Hangzhou ACEA Pharmaceutical Research) are other two novel 3rd generation EGFR TKIs in early development phase and results of phase the I/II studies are awaited.

7. Emerging mechanisms of acquired resistance to 3rd generation EGFR TKIs

Despite promising activity against EGFR mutant T790M+ NSCLCs, acquired resistance to 3rd generation EGFR TKIs ultimately occurs through different and, only partially, known mechanisms.

Prior studies, evaluating the structural analogue tool compound WZ4002, reported activation of MAPK and IGF1R pathways as

possible mechanisms of acquired resistance to irreversible mutantselective EGFR TKIs (Cortot et al., 2013; Ercan et al., 2012), but no additional mutations, a largely anticipated mechanism of resistance, were reported until recently.

This class of agents has been developed to inhibit EGFR T790M mutations, binding covalently to the cysteine residue 797 (Zhou et al., 2009). Therefore, it is not surprising that EGFR T790M+ cells acquiring resistance to mutant selective EGFR TKIs, due to the selective pressure of these agents, may develop a tertiary mutation that results in a cysteine to serine change at position 797 (C797S) in a region of the ATP-binding pocket of EGFR that is opposite to that of T790M. The EGFR C797S mutation resembles the acquired C481S mutation to the irreversible BTK (Bruton tyrosine kinase) inhibitor Ibrutinib in patients with chronic lymphocytic leukemia (Woyach et al., 2014), suggesting that this mutation type may represent a common mechanism of AR to covalent kinase inhibitors.

Indeed, preclinical studies, using cultured patient-derived EGFR exon19del/T790M cell lines (MGH121) and through mutagenesis screen in EGFR mutant cells with or without T790M, reported the development of three tertiary mutations (C797S, L844V and L718Q) in the EGFR after acquiring resistance to the 3rd generation compound WZ4002 (Niederst et al., 2015b; Ercan et al., 2015). The presence of these mutations and their allelic/genomic context has been reported to influence their sensitivity to the different EGFR TKIs available. Indeed, C797S conferred resistance to all the 3rd generation EGFR TKIs evaluated (WZ4002, Osimertinib and Rociletinib), but was sensitive to 1st and 2nd generation inhibitors (with reduced sensitivity in the context of L858R/C797S cells), L844V was associated with resistance to WZ4002 and Rociletinib, but not to Osimertinib (independently of the presence of T790M mutation), Gefitinib and Afatinib, while L718O exhibited resistance to WZ4002 and Rociletinib, but not to Osimertinib, only in the context of exon19 del/L718Q (Ercan et al., 2015). A possible explanation may be found in the inherited differences in the molecular structure of Rociletinib and Osimertinib, since the first is structurally more similar to WZ4002 than the latter. Moreover, Niederst et al. reported that the allelic context of this acquired resistance may be essential for the therapeutic strategy of patients with EGFR mutation, since the development in the same alleles of T790M and C797S mutations (i.e. in cis mutations) confers resistance to all the EGFR TKIs known, but the development of these mutations in different alleles (i.e. in trans) may pave the way to a combinatory approach of both 1st and 3rd generation TKIs. Moreover, the front-line use of mutant-selective inhibitors and the subsequent development of C797S mutation may raise the opportunity of the use of 1st generation agents, since del19/C797S mutations are still sensitive to Gefitinib (Niederst et al., 2015b).

These findings suggest that different sequential and combinatory approaches may necessary in EGFR-mutated NSCLCs patients when treated with different classes of EGFR TKIs, since the mechanisms of AR may vary based on the molecular and therapeutic context.

The development of C797S mutation has been reported *in vivo* in a small cohort (15 subjects) of EGFR T790M positive patients progressing after Osimertinib: this tertiary mutation was found, using a droplet digital PCR (ddPCR) assay, in 40% of patients after AR to

this agent, while 33% of the cases exhibited persistence of the secondary mutation T790M (without C797S) and 27% showed neither T790M nor C797S mutations (Thress et al., 2015). The development of C797S has been recently confirmed *in vivo* in tumor samples obtained after re-biopsy in patients with acquired resistance to Osimertinib (Yu et al., 2015) and Olmutinib (Song et al., 2015) and in plasma samples from patients with AR to Rociletinib, albeit with a less frequency than observed with Osimertinib (2% vs. 32%), suggesting a distinct pattern of resistance mechanisms (Chabon et al., 2016).

Jia et al. recently identified an investigational compound, EAI045, that targets selected drug-resistant EGFR mutants but spares the wild-type receptor. Unlike 3rd generation mutant-selective EGFR TKIs, this compound binds to T790M-mutant EGFR in an allosteric site created by the displacement of the regulatory C-helix in an inactive conformation of the kinase. This different mechanism of action do not allow an efficient EGFR blockage as single agent, but a marked synergy was observed with the anti-EGFR antibody Cetuximab in mouse models of lung cancer driven by L858R/T790M EGFR and, interestingly, by L858R/T790M/C797S EGFR (Jia et al., 2016).

Studying 32 cell lines with acquired resistance to several EGFR TKIs, including Gefitinib, Afatinib, WZ4002 and Osimertinib, Eberlein et al. reported that resistance to AZD9291 and other EGFR inhibitors in vitro is often associated with increased dependence on RAS signaling, through NRAS mutations (including the novel E63K NRAS mutation) and KRAS gain, and sensitivity to the MEK inhibitor Selumetinib (Eberlein et al., 2015). These data are in line with previous reports indicating the RAS-MAPK signaling pathway as an alternative mechanisms of acquired resistance to EGFR inhibitors in both lung and colon cancer (Misale et al., 2014) in many different ways, including loss of NF1 (de Bruin et al., 2014a), CRKL amplification (Cheung et al., 2011), MAPK1 amplification (Ercan et al., 2012). Moreover, the combination of Osimertinib and Selumetinib delayed the emergence of resistance in EGFRm and EGFRm/T790M cells (Eberlein et al., 2015). These data provide the rationale for concomitant use of Osimertinib and Selumetinib in the ongoing multi-arm phase Ib trial TATTON [NCT02143466].

Activation of alternative signaling pathways in an EGFR-independent way, may also occur after acquired resistance to Osimertinib. Planchard et al. recently reported the amplification of HER2 and MET (not present in the pre-treatment specimens), respectively, in two patients enrolled into the phase I AURA trial in re-biopsy specimens collected at the AR to this mutant-selective TKI with disappearance of the T790M mutation and no other additional mutations (Planchard et al., 2015).

In a small case series of 4 EGFR exon 19 del/T790M NSCLCs patients with acquired resistance to Osimertinib enrolled into the AURA trial, re-biopsy after AR revealed the presence of different mechanisms of resistance: none of the 4 patients developed C797S mutation, but loss of EGFR T790M mutation was observed in addition to alternative pathways activation, including FGFR1 amplification, PTEN deletion, MAPK1 and AKT3 overexpression, and histologic transformation (i.e. SCLC transition) (Kim et al., 2015), confirming previous in vitro findings (Cross et al., 2014; Ercan et al., 2012; Tabara et al., 2012; Ware et al., 2013). Similarly to a previous report (Niederst et al., 2015a), SCLC transformation was associated with RB loss. This small case series conducted on patients progressing after both 1st and 3rd generation EGFR TKIs confirms the complexity and substantial overlay of mechanisms of acquired resistance, under the selective pressure of different classes EGFR TKIs.

The loss of secondary mutation T790M mutation after AR and SCLC transformation was also reported for Rociletinib. Piotrowska et al. using the MGH NGS platform recently reported in 13 biopsies among 12 EGFR T790M+ patients progressing after treatment

with Rociletinib the loss of EGFR T790M mutation in 6 (with evidence of SCLC transformation plus RB1 loss in 2/6 cases) and persistence of T790M mutation in 7 samples (with EGFR amplification in 3/7 tissue biopsies), respectively. No C797S positive cases were reported, probably due to the small sample size and the population evaluated in the study: both primary and secondary resistant patients to Rociletinib were included and mostly with poor response to the treatment. Interestingly, using a patientderived cell line from an Afatinib-resistant NSCLC, the authors also demonstrated the intratumor heterogeneity of T790M mutations within the same sample "T790M positive", providing a possible explanation for the apparent "loss" of this mutation at the AR to Rociletinib: it is likely expression of selection of pre-existing T790M-wild type clones rather than mutation/deletion of T790M alleles (Piotrowska et al., 2015). Moreover, a recent study using a CAPP-seq ctDNA, analysis, allowing simultaneously study of singlenucleotide variants (SNVs), insertions/deletions, rearrangements, and somatic copy-number alterations (SCNAs), revealed a largely underestimate heterogeneity of mechanisms of acquired resistance to 3rd generation EGFR TKIs, with co-existence of multiple mechanisms in the same patient at higher frequency than previously reported (46% in T790M-mutant vs. 5-15%) (Chabon et al., 2016). These findings have important clinical implications since intrapatient heterogeneity may negatively affect clinical response to 3rd generation EGFR TKis.

8. Future perspectives

The development of 3rd generation EGFR TKIs poses novel challenges in the therapeutic management of EGFR-mutant NSCLCs (Fig. 1). One of the possible future scenarios is the use of mutant-selective EGFR TKIs in the 1st line setting.

Front-line use of use of these agents might provide several advantages over the current approved EGFR TKIs: activity against EGFR mutants, including T790M, while sparing EGFR wild type and delay of acquired resistance, as demonstrated in *in vitro* models (Ramalingam et al., 2014), with a better central nervous system (CNS) penetration. Indeed, Ballard et al. recently reported in a mouse model that Osimertinib is distributed in the CNS at greater extent than Gefitinib, Rociletinib and Afatinib (Ballard et al., 2016), providing the rationale for the use of Osimertinib in EGFR-mutant patients with brain metastases as first line option.

Several different clinical trials are evaluating the role of EGFR-mutant selective EGFR TKIs in TKI-naïve patients [Table 4].

Preliminary data from two cohorts of TKI-naïve patients enrolled into the phase I AURA trial treated with Osimertinib at 80 mg or 160 mg daily were recently reported, showing a promising activity in this subset of patients: 67% ORR in the 80 mg cohort and 77% in the 160 mg cohort, 97% DCR overall and a median PFS of 19.3 months in the overall population (median not reached in the 80 mg cohort and 19.3 months in the 160 mg cohort) (Ramalingam et al., 2016).

The FLAURA trial (NCT02296125) is an ongoing double blind, randomized, phase III study comparing Osimertinib 80 mg/d with standard 1st generation EGFR TKIs, Gefitinib and Erlotinib. Rociletinib is being evaluating in both treatment-naïve and pretreated patients in the phase II part of TIGER1 trial and in combination with the anti-PDL1 inhibitor Atezolizumab in a phase Ib/II (NCT02630186). ASP8273 is also being compared with first generation EGFR TKIs in the 1st line setting in the open label phase III trial SOLAR (NCT02588261).

Osimertinib is also being studied in the adjuvant setting (stage IB-IIIA NSCLC with EGFR mutations after surgical resection with/without chemotherapy) in the phase III trial ADAURA (NCT02511106).

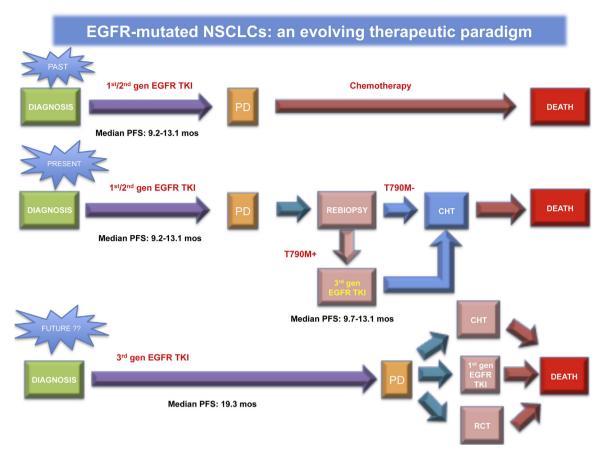


Fig. 1. Evolving therapeutic algorithm in EGFR mutated NSCLCs. Legend: CHT, chemotherapy; RCT, randomized clinical trial.

Table 4 Initial development program of Olmutinib.

Name	Phase	Population	Primary Objective (s)	Control Arm
HM-EMSI-101	I/II	T790M+ TKI-pretreated (expanded cohort)	Safety, Tolerability and Pharmacokinetic	-
ELUXA 1 (HM-EMSI-202)	II	T790M+ TKI-pretreated	ORR	-
ELUXA-2	III	T790M+ TKI-pretreated	PFS	Platinum doublet CHT
ELUXA-3	III	EGFR-mut+ 1st line	PFS	Afatinib
ELUXA-4	I/II	Japanese T790M+ TKI-pretreated	ORR	-
ELUXA-6	II	NSCLC for whom a needle biopsy may not be appropriate	Prospectively use blood-based biomarker testing to select patients with EGFR T790M+	-

Another challenge posed by 3rd generation EGFR TKIs is the rebiopsy of patients with acquired resistance to 1st/2nd generation EGFR TKIs. Albeit tumor tissue genotyping is the gold standard, a re-biopsy is not always feasible in clinical practice because of scheduling problems, costs, risks of complications and issues related to tissue acquisition and preservation (LA Jr and Bardelli, 2014). Moreover, tumor heterogeneity may represent an obstacle for tumor genotyping, since single region sampling may underestimate the genomic complexity of a tumor (Gerlinger et al., 2012 8; de Bruin et al., 2014b). Liquid biopsy may overcome some of the limits of traditional tissue biopsy and in particular plasma genotyping of circulating tumor DNA (ctDNA) has shown promising results.

In the phase I AURA trial, patients with EGFR activating mutations and centrally confirmed tumor and/or plasma genotyping (BEAMing) T790M result were enrolled. Among 216 patients with both plasma and tissue genotyping results, the concordance rate was 82% for exon 19 deletions, 86% for L858R and 70% for T790M. Outcomes were similar for 179 patients T790M+ in tumor (62% ORR and 9.7 months PFS) and for 167 patients T790M+ in plasma

(63% ORR, 9.7 months), but unexpectedly differed for T790M- in plasma (46% ORR, 8.2 months PFS) and T790M- in tumor (26% ORR, 3.4 months PFS). These data suggest that patients with a T790M negative results in plasma genotyping may be divided in two subgroups with different outcome: T790M undetected (38% ORR, 4.4 months PFS) and T790M uninformative (64% ORR, 15.2 months PFS) (Oxnard et al., 2016). These data suggest that plasma and tissue genotyping can have complementary roles and that liquid biopsy should be offered early to identify T790M+ candidates for Osimertinib therapy, reserving traditional tissue biopsy to T790M- cases in plasma.

The favorable toxicity profile of 3rd generation EGFR TKIs pave the way to combinatory schedules with other agents in order to provide a more comprehensive anti-tumor activity and/or to prevent the emergence of resistant clones. Several clinical trials are ongoing evaluating different combinations in EGFR-mutated NSCLCs in various setting [Table 5].

Recently, safety concerns raised from the ongoing multi-arm phase Ib study TATTON evaluating different combinations and

Table 5Selected studies with 3rd generation EGFR TKIs in the 1st line setting.

Treatment(s)	Phase	Population	Study Name
ASP8273 vs. Gefitinib/Erlotinib	III	EGFR-mut+	NCT02588261 (SOLAR)
Osimertinib vs. Gefitinib/Erlotinib	III	EGFR-mut+	NCT02296125 (FLAURA)
Osimertinib vs. Gefitinib	II	EGFR-mut+	APPLE/EORTC 1613
Osimertinib	II	EGFR-mut+	NCT02841579 (AZENT)
Osimertinib	II	EGFR-mut+ (liquid biopsy)	NCT02769286 (LiquidLung-O)
Osimertinib + Bevacizumab	I/II	EGFR-mut+	NCT02803203
EGF816	II	de novo T790M (Arm 3)	NCT02108964
ASP8273	II	EGFR-mut+	NCT02500927

Table 6Ongoing trials combining 3rd generation EGFR TKIs with other agents in EGFR-mutated NSCLCs.

Drug	Combination(s)	Phase	Study
Osimertinib	Selumetinib (anti-MEK) Durvalumab (anti-PDL1) Savolitinib (anti-MET)	Ib	NCT02143466 (TATTON)
Osimertinib	Durvalumab (anti-PDL1)	III	NCT02454933 (CAURAL)
Osimertinib	Bevacizumab (anti-VEGF)	II	NCT03133546 (BOOSTER)
Osimertinib	Navitoclax (anti-Bcl-2)	I	NCT02520778
Osimertinib	Itacitanib (anti-JAK1)	I/II	NCT02917993
Osimertinib	INK-128 (anti-TORC1/2)	I	NCT02503722
Osimertinib	Necitumumab (anti-EGFR mAb)	I	NCT02496663
EGF-816	Nivolumab (anti-PD1)	II	NCT02323126
EGF-816	PDR001 (anti-PD1)	I	NCT02900664
EGF-816	Capmatinib (anti-MET)	I/II	NCT02335944

schedules of Osimertinib with other investigational agents, since 38% of patients treated with Osimertinib plus the anti-PDL1 inhibitor Durvalumab developed an interstitial lung disease (ILD) at an unexpectedly higher frequency compared with both single agents (2.9% and 2.0%, respectively) (Ahn et al., 2016) (Table 6).

9. Conclusions

Preclinical and clinical data with mutant-selective EGFR TKIs are promising and are changing the therapeutic landscape of EGFR-mutated NSCLCs (Fig. 1). The rapid FDA approval of Osimertinib (only 2.8 months after first patient treated) and the fast development of Rociletinib and Olmutinib are profoundly influencing the therapeutic algorithm of EGFR mutant NSCLCs, with a new and effective option after acquired resistance to 1st and 2nd generation EGFR TKIs. Ongoing trials will produce the definitive evidences of the best in class EGFR TKI and the optimal therapeutic sequence of these agents.

Conflicts of interest

No potential conflicts of interest declared.

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References

- Ahn, M.J., Yang, J., Yu, H., Saka, H., Ramalingam, S., Goto, K., et al., 2016. 1360: Osimertinib combined with durvalumab in EGFR-mutant non-small cell lung cancer: results from the TATTON phase lb trial. J. Thorac. Oncol. 11 (Suppl. 4), \$115
- Anon, 2016. FDA Briefing Document Oncologic Drugs Advisory Committee Meeting April 12, 2016. http://www.fda.gov/downloads/AdvisoryCommittees/CommitteesMeetingMaterials/Drugs/OncologicDrugsAdvisoryCommittee/UCM494782.pdf.
- Balak, M.N., Gong, Y., Riely, G.J., Somwar, R., Li, A.R., Zakowski, M.F., et al., 2006. Novel D761Y and common secondary T790M mutations in epidermal growth

- factor receptor-mutant lung adenocarcinomas with acquired resistance to kinase inhibitors. Clin. Cancer Res. 12 (21), 6494–6501.
- Ballard, P., Yates, J.W., Yang, Z., Kim, D.W., Yang, J.C., Cantarini, M., et al., 2016. Preclinical comparison of osimertinib with other EGFR-TKIs in EGFR-Mutant NSCLC brain metastases models, and early evidence of clinical brain metastases activity. Clin. Cancer Res., pii: clincanres.0399.2015. [Epub ahead of print].
- Bean, J., Riely, G.J., Balak, M., Marks, J.L., Ladanyi, M., Miller, V.A., Pao, W., 2008. Acquired resistance to epidermal growth factor receptor kinase inhibitors associated with a novel T854A mutation in a patient with EGFR-mutant lung adenocarcinoma. Clin. Cancer Res. 14 (22), 7519–7525.
- Bivona, T.G., Hieronymus, H., Parker, J., Chang, K., Taron, M., Rosell, R., et al., 2011. FAS and NF-κB signalling modulate dependence of lung cancers on mutant EGFR. Nature 471 (7339), 523–526.
- Byers, L.A., Diao, L., Wang, J., Saintigny, P., Girard, L., Peyton, M., et al., 2013. An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies Axl as a therapeutic target for overcoming EGFR inhibitor resistance. Clin. Cancer Res. 19 (1), 279–290.
- Camidge, D.R., Pao, W., Sequist, L.V., 2014. Acquired resistance to TKIs in solid tumours: learning from lung cancer. Nat. Rev. Clin. Oncol. 11 (8), 473–481.
- Chabon, J.J., Simmons, A.D., Lovejoy, A.F., Esfahani, M.S., Newman, A.M., Haringsma, H.J., et al., 2016. Circulating tumour DNA profiling reveals heterogeneity of EGFR inhibitor resistance mechanisms in lung cancer patients. Nat. Commun. 7, 11815.
- Cheung, H.W., Du, J., Boehm, J.S., He, F., Weir, B.A., Wang, X., et al., 2011. Amplification of CRKL induces transformation and epidermal growth factor receptor inhibitor resistance in human non-small cell lung cancers. Cancer Discov. 1 (7), 608–625.
- Chmielecki, J., Foo, J., Oxnard, G.R., Hutchinson, K., Ohashi, K., Somwar, R., et al., 2011. Optimization of dosing for EGFR-mutant non-small cell lung cancer with evolutionary cancer modeling. Sci. Transl. Med. 3 (90), 90ra59.
- Cortot, A.B., Repellin, C.E., Shimamura, T., Capelletti, M., Zejnullahu, K., Ercan, D., et al., 2013. Resistance to irreversible EGF receptor tyrosine kinase inhibitors through a multistep mechanism involving the IGF1R pathway. Cancer Res. 73 (2), 834–843.
- Costa, D.B., Halmos, B., Kumar, A., Schumer, S.T., Huberman, M.S., Boggon, T.J., et al., 2007. BIM mediates EGFR tyrosine kinase inhibitor-induced apoptosis in lung cancers with oncogenic EGFR mutations. PLoS Med. 4 (10), 1669–1679 (discussion 1680).
- Costa, C., Molina, M.A., Drozdowskyj, A., Giménez-Capitán, A., Bertran-Alamillo, J., Karachaliou, N., et al., 2014. The impact of EGFR T790M mutations and BIM mRNA expression on outcome in patients with EGFR-mutant NSCLC treated with erlotinib or chemotherapy in the randomized phase III EURTAC trial. Clin. Cancer Res. 20 (7), 2001–2010.
- Cross, D.A., Ashton, S.E., Ghiorghiu, S., Eberlein, C., Nebhan, C.A., Spitzler, P.J., et al., 2014. AZD9291, an irreversible EGFR TKI, overcomes T790M-mediated resistance to EGFR inhibitors in lung cancer. Cancer Discov. 4 (9), 1046–1061.
- Eberlein, C.A., Stetson, D., Markovets, A.A., Al-Kadhimi, K.J., Lai, Z., Fisher, P.R., et al., 2015. Acquired resistance to the mutant-selective EGFR inhibitor AZD9291 is associated with increased dependence on RAS signaling in preclinical models. Cancer Res. 75 (12), 2489–2500.
- Engelman, J.A., Zejnullahu, K., Gale, C.M., Lifshits, E., Gonzales, A.J., Shimamura, T., et al., 2007. PF00299804, an irreversible pan-ERBB inhibitor, is effective in lung

- cancer models with EGFR and ERBB2 mutations that are resistant to gefitinib. Cancer Res. 67 (24), 11924–11932.
- Ercan, D., Xu, C., Yanagita, M., Monast, C.S., Pratilas, C.A., Montero, J., et al., 2012. Reactivation of ERK signaling causes resistance to EGFR kinase inhibitors. Cancer Discov. 2 (10), 934–947.
- Ercan, D., Choi, H.G., Yun, C.H., Capelletti, M., Xie, T., Eck, M.J., et al., 2015. EGFR mutations and resistance to irreversible pyrimidine-based EGFR inhibitors. Clin. Cancer Res. 21 (17), 3913–3923.
- Finlay, M.R., Anderton, M., Ashton, S., Ballard, P., Bethel, P.A., Box, M.R., et al., 2014. Discovery of a potent and selective EGFR inhibitor (AZD9291) of both sensitizing and T790M resistance mutations that spares the wild type form of the receptor. J. Med. Chem. 57 (20), 8249–8267.
- Gerlinger, M., Rowan, A.J., Horswell, S., Larkin, J., Endesfelder, D., Gronroos, E., et al., 2012 8. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. N. Engl. J. Med. 366 (10), 883–892.
- Godin-Heymann, N., Ulkus, L., Brannigan, B.W., McDermott, U., Lamb, J., Maheswaran, S., et al., 2008. The T790M gatekeeper mutation in EGFR mediates resistance to low concentrations of an irreversible EGFR inhibitor. Mol. Cancer Ther. 7 (4), 874–879.
- Hata, A., Katakami, N., Yoshioka, H., Takeshita, J., Tanaka, K., Nanjo, S., et al., 2013. Rebiopsy of non-small cell lung cancer patients with acquired resistance to epidermal growth factor receptor-tyrosine kinase inhibitor: comparison between T790M mutation-positive and mutation-negative populations. Cancer 119 (24), 4325–4332.
- Jänne, P.A., Yang, J.C., Kim, D.W., Planchard, D., Ohe, Y., Ramalingam, S.S., et al., 2015. AZD9291 in EGFR inhibitor-resistant non-small-cell lung cancer. N. Engl. J. Med. 372 (18), 1689–1699.
- Janjigian, Y.Y., Smit, E.F., Groen, H.J., Horn, L., Gettinger, S., Camidge, D.R., et al., 2014. Dual inhibition of EGFR with afatinib and cetuximab in kinase inhibitor-resistant EGFR-mutant lung cancer with and without T790M mutations. Cancer Discov. 4 (9), 1036–1045.
- Jia, Y., Yun, C.H., Park, E., Ercan, D., Manuia, M., Juarez, J., et al., 2016. Overcoming EGFR(T790M) and EGFR(C797S) resistance with mutant-selective allosteric inhibitors. Nature 534 (7605), 129–132.
- Kim, T.M., Song, A., Kim, D.W., Kim, S., Ahn, Y.O., Keam, B., et al., 2015. Mechanisms of acquired resistance to AZD9291: a mutation-Selective, irreversible EGFR inhibitor. J. Thorac, Oncol. 10 (12), 1736–1744.
- Kobayashi, S., Boggon, T.J., Dayaram, T., Jänne, P.A., Kocher, O., Meyerson, M., et al., 2005a. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. N. Engl. J. Med. 352 (8), 786–792.
- Kobayashi, S., Ji, H., Yuza, Y., Meyerson, M., Wong, K.K., Tenen, D.G., Halmos, B., 2005b. An alternative inhibitor overcomes resistance caused by a mutation of the epidermal growth factor receptor. Cancer Res. 65 (16), 7096–7101.
- Kuiper, J.L., Heideman, D.A., Thunnissen, E., Paul, M.A., van Wijk, A.W., Postmus, P.E., Smit, E.F., 2014. Incidence of T790M mutation in (sequential) rebiopsies in EGFR-mutated NSCLC-patients. Lung Cancer 85 (1), 19–24.
- Kwak, E.L., Sordella, R., Bell, D.W., Godin-Heymann, N., Okimoto, R.A., Brannigan, B.W., et al., 2005. Irreversible inhibitors of the EGF receptor may circumvent acquired resistance to gefitinib. Proc. Natl. Acad. Sci. U. S. A. 102 (21), 7665–7670.
- LA Jr, Diaz, Bardelli, A., 2014. Liquid biopsies: genotyping circulating tumor DNA. J. Clin. Oncol. 32 (6), 579–586.
- Landi, L., Tiseo, M., Chiari, R., Ricciardi, S., Rossi, E., Galetta, D., et al., 2014. Activity of the EGFR-HER2 dual inhibitor afatinib in EGFR-mutant lung cancer patients with acquired resistance to reversible EGFR tyrosine kinase inhibitors. Clin. Lung Cancer 15 (6), 411–417, e4.
- Lee, Y., Lee, G.K., Lee, Y.S., Zhang, W., Hwang, J.A., Nam, B.H., et al., 2014. Clinical outcome according to the level of preexisting epidermal growth factor receptor T790M mutation in patients with lung cancer harboring sensitive epidermal growth factor receptor mutations. Cancer 120 (14), 2090–2098.
- Lovly, C.M., Shaw, A.T., 2014. Molecular pathways: resistance to kinase inhibitors and implications for therapeutic strategies. Clin. Cancer Res. 20 (9), 2249–2256.
- Maheswaran, S., Sequist, L.V., Nagrath, S., Ulkus, L., Brannigan, B., Collura, C.V., et al., 2008. Detection of mutations in EGFR in circulating lung-cancer cells. N. Engl. I. Med. 359 (4), 366–377.
- Miller, V.A., Hirsh, V., Cadranel, J., Chen, Y.M., Park, K., Kim, S.W., et al., 2012. Afatinib versus placebo for patients with advanced, metastatic non-small-cell lung cancer after failure of erlotinib, gefitinib, or both, and one or two lines of chemotherapy (LUX-Lung 1): a phase 2b/3 randomised trial. Lancet Oncol. 13 (5). 528–538.
- Misale, S., Arena, S., Lamba, S., Siravegna, G., Lallo, A., Hobor, S., Russo, M., Buscarino, M., Lazzari, L., Sartore-Bianchi, A., Bencardino, K., Amatu, A., Lauricella, C., Valtorta, E., Siena, S., Di Nicolantonio, F., Bardelli, A., 2014. Blockade of EGFR and MEK intercepts heterogeneous mechanisms of acquired resistance to anti-EGFR therapies in colorectal cancer. Sci. Transl. Med. 6 (224), 224ra26.
- Mok, T.S., Wu, Y.L., Thongprasert, S., Yang, C.H., Chu, D.T., Saijo, N., et al., 2009. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. N. Engl. J. Med. 361 (10), 947–957.
- Nanjo, S., Ebi, H., Arai, S., Takeuchi, S., Yamada, T., Mochizuki, S., et al., 2016. High efficacy of third generation EGFR inhibitor AZD9291 in a leptomeningeal carcinomatosis model with EGFR-mutant lung cancer cells. Oncotarget 7 (4), 3847–3856.
- Niederst, M.J., Sequist, L.V., Poirier, J.T., Mermel, C.H., Lockerman, E.L., Garcia, A.R., et al., 2015a. RB loss in resistant EGFR mutant lung adenocarcinomas that transform to small-cell lung cancer. Nat. Commun. 6, 6377.

- Niederst, M.J., Hu, H., Mulvey, H.E., Lockerman, E.L., Garcia, A.R., Piotrowska, Z., et al., 2015b. The allelic context of the C797S mutation acquired upon treatment with third-Generation EGFR inhibitors impacts sensitivity to subsequent treatment strategies. Clin. Cancer Res. 21 (17), 3924–3933.
- Ohashi, K., Sequist, L.V., Arcila, M.E., Moran, T., Chmielecki, J., Lin, Y.L., et al., 2012. Lung cancers with acquired resistance to EGFR inhibitors occasionally harbor BRAF gene mutations but lack mutations in KRAS, NRAS, or MEK1. Proc. Natl. Acad. Sci. U. S. A. 109 (31), E2127–33.
- Osimertinib, Yver A., 2016. (AZD9291)-a science-driven, collaborative approach to rapid drug design and development. Ann. Oncol. 27 (6), 1165–1170.
- Oxnard, G.R., Arcila, M.E., Sima, C.S., Riely, G.J., Chmielecki, J., Kris, M.G., et al., 2011. Acquired resistance to EGFR tyrosine kinase inhibitors in EGFR-mutant lung cancer: distinct natural history of patients with tumors harboring the T790 M mutation. Clin. Cancer Res. 17 (6), 1616–1622.
- Oxnard, G.R., Thress, K.S., Alden, R.S., Lawrance, R., Paweletz, C.P., Cantarini, M., et al., 2016. Association between plasma genotyping and outcomes of treatment with osimertinib (AZD 9291) in advanced non-small-cell lung cancer. J. Clin. Oncol., pii: JCO667162. [Epub ahead of print].
- Pao, W., Miller, V.A., Politi, K.A., Riely, G.J., Somwar, R., Zakowski, M.F., et al., 2005. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. PLoS Med. 2 (3), e73.
- Park, K., Lee, J.S., Lee, K.H., Kim, J.H., Cho, B.C., Min, Y.J., et al., 2016a. BI 1482694 (HM61713), an EGFR mutant-specific inhibitor, in T790M+ NSCLC: efficacy and safety at the RP2D. J. Clin. Oncol. 34 (suppl; abstr 9055).
- Park, K.M., Lee, J.S., Han, J.Y., Lee, K.H., Kim, J.H., Cho, E.K., et al., 2016b. Efficacy and safety of BI 1482694 (HM61713), an EGFR mutant-specific inhibitor, in T790M-positive NSCLC at the recommended phase II dose. J. Thorac. Oncol. 11 (Suppl. 4S), S113–S142.
- Piotrowska, Z., Niederst, M.J., Karlovich, C.A., Wakelee, H.A., Neal, J.W., Mino-Kenudson, M., et al., 2015. Heterogeneity underlies the emergence of EGFRT790 wild-type clones following treatment of T790M-positive cancers with a third-generation EGFR inhibitor. Cancer Discov. 5 (7), 713–722.
- Planchard, D., Loriot, Y., André, F., Gobert, A., Auger, N., Lacroix, L., Soria, J.C., 2015. EGFR-independent mechanisms of acquired resistance to AZD9291 in EGFR T790M-positive NSCLC patients. Ann. Oncol. 26 (10), 2073–2078.
- Ramalingam, S.S., Ohe, Y., Nogami, N., Yang, J.C., Eberlein, C., Ashton, S., et al., 2014. Pre-clinical and clinical evaluation of AZD9291, a mutation-specific inhibitor, in treatment-naïve EGFR mutated NSCLC. Ann. Oncol. 25 (Suppl. 4), iv146-iv164.
- Ramalingam, S., Yang, J.C., Lee, C.K., Kurata, T., Kim, D.W., John, T., et al., 2016. LBA1_PR: Osimertinib as first-line treatment for EGFR mutation-positive advanced NSCLC: updated efficacy and safety results from two Phase I expansion cohorts. J. Thorac. Oncol. 11 (Suppl. 4), S152.
- Reckamp, K.L., Giaccone, G., Camidge, D.R., Gadgeel, S.M., Khuri, F.R., Engelman, J.A., et al., 2014. A phase 2 trial of dacomitinib (PF-00299804), an oral, irreversible pan-HER (human epidermal growth factor receptor) inhibitor, in patients with advanced non-small cell lung cancer after failure of prior chemotherapy and erlotinib. Cancer 120 (8), 1145–1154.
- Rivera, V.M., Wang, F., Anjum, R., Zhang, S., Squillace, R., Keats, J., et al., 2012. AP26113 is a dual ALK/EGFR inhibitor: characterization against EGFR T79 M in cell and mouse models of NSCLC. Cancer Res. (Suppl. 8), 72, Abstract nr 1794. Rudell, K., Papadakis, K., Bodnar, C., Hoyle, C., Ghiorghiu, S., Ryden, A., 2016. The
- Rudell, K., Papadakis, K., Bodnar, C., Hoyle, C., Ghiorghiu, S., Ryden, A., 2016. The impact of osimertinib on function and health status for patients with EGFR mutation-positive advanced non-small cell lung cancer. J. Clin. Oncol. 34 (Suppl; abstr e20548).
- Russo, A., Franchina, T., Ricciardi, G.R., Picone, A., Ferraro, G., Zanghì, M., et al., 2015. A decade of EGFR inhibition in EGFR-mutated non small cell lung cancer (NSCLC): Old successes and future perspectives. Oncotarget 6 (29), 26814–26825.
- Sequist, L.V., Besse, B., Lynch, T.J., Miller, V.A., Wong, K.K., Gitlitz, B., et al., 2010. Neratinib, an irreversible pan-ErbB receptor tyrosine kinase inhibitor: results of a phase II trial in patients with advanced non-small-cell lung cancer. J. Clin. Oncol. 28 (18), 3076–3083.
- Sequist, L.V., Waltman, B.A., Dias-Santagata, D., Digumarthy, S., Turke, A.B., Fidias, P., et al., 2011. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. Sci. Transl. Med. 3 (75), 75ra26.
- Sequist, L.V., Yang, J.C., Yamamoto, N., O'Byrne, K., Hirsh, V., Mok, T., et al., 2013. Phase III study of afatinib or cisplatin plus pemetrexed in patients with metastatic lung adenocarcinoma with EGFR mutations. J. Clin. Oncol. 31 (27), 3327–3334
- Sequist, L.V., Soria, J.C., Goldman, J.W., Wakelee, H.A., Gadgeel, S.M., Varga, A., et al., 2015. Rociletinib in EGFR-mutated non-small-cell lung cancer. N. Engl. J. Med. 372 (18), 1700–1709.
- Song, H.N., Jung, K.S., Yoo, K.H., Cho, J., Lee, J.Y., Lim, S.H., et al., 2015. Acquired C797S mutation upon treatment with a T790M-specific third-generation EGFR inhibitor (HM61713) in non-small cell lung cancer. J Thorac Oncol., 00209, pii: \$1556-0864(15)
- Su, K.Y., Chen, H.Y., Li, K.C., Kuo, M.L., Yang, J.C., Chan, W.K., et al., 2012. Pretreatment epidermal growth factor receptor (EGFR) T790M mutation predicts shorter EGFR tyrosine kinase inhibitor response duration in patients with non-small-cell lung cancer. J. Clin. Oncol. 30 (4), 433–440.
- Suda, K., Mizuuchi, H., Maehara, Y., Mitsudomi, T., 2012. Acquired resistance mechanisms to tyrosine kinase inhibitors in lung cancer with activating epidermal growth factor receptor mutation–diversity, ductility, and destiny. Cancer Metastasis Rev. 31 (3–4), 807–814.

- Tabara, K., Kanda, R., Sonoda, K., Kubo, T., Murakami, Y., Kawahara, A., et al., 2012. Loss of activating EGFR mutant gene contributes to acquired resistance to EGFR tyrosine kinase inhibitors in lung cancer cells. PLoS One 7 (7), e41017.
- Takezawa, K., Pirazzoli, V., Arcila, M.E., Nebhan, C.A., Song, X., de Stanchina, E., et al., 2012. HER2 amplification: a potential mechanism of acquired resistance to EGFR inhibition in EGFR-mutant lung cancers that lack the second-site EGFRT790M mutation. Cancer Discov. 2 (10), 922–933.
- Tan, D.S., Yang, J.C., Leighl, N.B., Riely, G.J., Sequist, L.V., Felip, E., et al., 2016. Updated results of a phase 1 study of EGF816, a third-generation, mutant-selective EGFR tyrosine kinase inhibitor (TKI), in advanced non-small cell lung cancer (NSCLC) harboring T790 M. J. Clin. Oncol. 34 (suppl; abstr 2004)
- Thress, K.S., Paweletz, C.P., Felip, E., Cho, B.C., Stetson, D., Dougherty, B., et al., 2015. Acquired EGFR C797S mutation mediates resistance to AZD9291 in non-small cell lung cancer harboring EGFR T790M. Nat. Med. 21 (6), 560–562.
- Tjin Tham Sjin, R., Lee, K., Walter, A.O., Dubrovskiy, A., Sheets, M., Martin, T.S., et al., 2014. In vitro and in vivo characterization of irreversible mutant-selective EGFR inhibitors that are wild-type sparing. Mol. Cancer Ther. 13 (6), 1468–1479.
- Uramoto, H., Yano, S., Tanaka, F., 2012. T790M is associated with a favorable prognosis in Japanese patients treated with an EGFR-TKI. Lung Cancer 76 (1), 129–130
- Walter, A.O., Sjin, R.T., Haringsma, H.J., Ohashi, K., Sun, J., Lee, K., et al., 2013. Discovery of a mutant-selective covalent inhibitor of EGFR that overcomes T790M-mediated resistance in NSCLC. Cancer Discov. 3 (12), 1404–1415.
- Ward, R.A., Anderton, M.J., Ashton, S., Bethel, P.A., Box, M., Butterworth, S., et al., 2013. Structure- and reactivity-based development of covalent inhibitors of the activating and gatekeeper mutant forms of the epidermal growth factor receptor (EGFR). J. Med. Chem. 56 (17), 7025–7048.
- Ware, K.E., Hinz, T.K., Kleczko, E., Singleton, K.R., Marek, L.A., Helfrich, B.A., Cummings, C.T., Graham, D.K., Astling, D., Tan, A.C., Heasley, L.E., 2013. A mechanism of resistance to gefitinib mediated by cellular reprogramming and the acquisition of an FGF2-FGFR1 autocrine growth loop. Oncogenesis 2, e39. Woyach, J.A., Furman, R.R., Liu, T.M., Ozer, H.G., Zapatka, M., Ruppert, A.S., et al.,
- Woyach, J.A., Furman, R.R., Liu, T.M., Ozer, H.G., Zapatka, M., Ruppert, A.S., et a 2014. Resistance mechanisms for the Bruton's tyrosine kinase inhibitor ibrutinib. N. Engl. J. Med. 370 (24), 2286–2294.
- Wu, Y.L., Zhou, C., Hu, C.P., Feng, J., Lu, S., Huang, Y., et al., 2014. Afatinib versus cisplatin plus gemcitabine for first-line treatment of Asian patients with advanced non-small-cell lung cancer harbouring EGFR mutations (LUX-Lung 6): an open-label, randomised phase 3 trial. Lancet Oncol. 15 (2), 213–222.
- Wu, Y.L., Zhou, C., Liam, C.K., Wu, G., Liu, X., Zhong, Z., et al., 2015. First-line erlotinib versus gemcitabine/cisplatin in patients with advanced EGFR mutation-positive non-small-cell lung cancer: analyses from the phase III, randomized, open-label, ENSURE study. Ann. Oncol. 26 (9), 1883–1889.
- Wu, S.G., Liu, Y.N., Tsai, M.F., Chang, Y.L., Yu, C.J., Yang, P.C., et al., 2016. The mechanism of acquired resistance to irreversible EGFR tyrosine kinase inhibitor-afatinib in lung adenocarcinoma patients. Oncotarget 7 (11), 12404–12413.
- Yang, J., Ramalingam, S.S., Jänne, P.A., Cantarini, M., Mistudomi, T., 2016a. Osimertinib (AZD9291) in pre-treated pts with T790M-positive advanced

- NSCLC: updated phase 1 (P1) and pooled phase 2 (P2) results. J. Thorac. Oncol. 11 (Suppl. 4S), S152–S155.
- Yang, J.C., Kim, D.W., Kim, S.W., Cho, B.C., Lee, J.S., Ye, X., et al., 2016b. Osimertinib activity in patients (pts) with leptomeningeal (LM) disease from non-small cell lung cancer (NSCLC): Updated results from BLOOM, a phase I study. J. Clin. Oncol. 34 (suppl; abstr 9002).
- Ye, X., Zhu, Z.Z., Zhong, L., Lu, Y., Sun, Y., Yin, X., et al., 2013. High T790M detection rate in TKI-naive NSCLC with EGFR sensitive mutation: truth or artifact? J. Thorac. Oncol. 8 (9), 1118–1120.
- Yosaatmadja, Y., Silva, S., Dickson, J.M., Patterson, A.V., Smaill, J.B., Flanagan, J.U., et al., 2015. Binding mode of the breakthrough inhibitor AZD9291 to epidermal growth factor receptor revealed. J. Struct. Biol. 192 (3), 539–544.
- Yu HA, Spira AI, Horn, L., Weiss, J., West, H.J., Giaccone, G., et al., 2016. Antitumor activity of ASP8273 300 mg in subjects with EGFR mutation-positive non-small cell lung cancer: interim results from an ongoing phase 1 study. J. Clin. Oncol. 34 (suppl; abstr 9050).
- Yu, H.A., Arcila, M.E., Rekhtman, N., Sima, C.S., Zakowski, M.F., Pao, W., et al., 2013. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. Clin. Cancer Res. 19 (8), 2240–2247.
- Yu, H.A., Arcila, M.E., Hellmann, M.D., Kris, M.G., Ladanyi, M., Riely, G.J., 2014. Poor response to erlotinib in patients with tumors containing baseline EGRR T790M mutations found by routine clinical molecular testing. Ann. Oncol. 25 (2), 423–428.
- Yu, H.A., Tian, S.K., Drilon, A.E., Borsu, L., Riely, G.J., Arcila, M.E., Ladanyi, M., 2015. Acquired resistance of EGFR-mutant lung cancer to a T790M-specific EGFR inhibitor: emergence of a third mutation (C797S) in the EGFR tyrosine kinase domain. JAMA Oncol. 1 (7), 982–984.
- Yun, C.H., Mengwasser, K.E., Toms, A.V., Woo, M.S., Greulich, H., Wong, K.K., et al., 2008. The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. Proc. Natl. Acad. Sci. U. S. A. 105 (6), 2070–2075.
- Zhang, Z., Lee, J.C., Lin, L., Olivas, V., Au, V., LaFramboise, T., et al., 2012. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. Nat. Genet. 44 (8), 852–860.
- Zhou, W., Ercan, D., Chen, L., Yun, C.H., Li, D., Capelletti, M., et al., 2009. Novel mutant-selective EGFR kinase inhibitors against EGFR T790M. Nature 462 (7276), 1070–1074.
- Zhou, C., Wu, Y.L., Chen, G., Feng, J., Liu, X.Q., Wang, C., et al., 2011. Erlotinib versus chemotherapy as first-line treatment for patients with advanced EGFR mutation-positive non-small-cell lung cancer (OPTIMAL, CTONG-0802): a multicentre, open-label, randomised, phase 3 study. Lancet Oncol. 12 (8), 735–742.
- de Bruin, E.C., Cowell, C., Warne, P.H., Jiang, M., Saunders, R.E., Melnick, M.A., et al., 2014a. Reduced NF1 expression confers resistance to EGFR inhibition in lung cancer. Cancer Discov. 4 (5), 606–619.
- de Bruin, E.C., McGranahan, N., Mitter, R., Salm, M., Wedge, D.C., Yates, L., et al., 2014b. Spatial and temporal diversity in genomic instability processes defines lung cancer evolution. Science 346 (6206), 251–256.