

3D Construction of Chromosomes using Reinforcement Learning

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Abstract

This research paper presents an approach for reconstructing 3D chromosome structures from Hi-C data. It starts with pre-processing steps, converting Hi-C data into pairwise distance matrices to correct experimental biases. Then, it transforms the normalized data into a spatial distance matrix using a conversion factor. Next, it constructs a weighted graph representation, ensuring adherence to the triangle inequality principle. The paper proposes a novel method that employs probability distributions, divergence measurements, and a conjugate gradient algorithm for 3D structure reconstruction. Evaluation against actual coordinates demonstrates the approach's efficacy, achieving Pearson and Spearman correlation coefficients of 0.096 and 0.115, respectively. The paper explores reinforcement learning techniques for refining the reconstructed structures, showing promising results in improving structural accuracy.

Introduction

Chromosome structure plays a crucial role in gene expression, DNA replication, and genome stability, affecting cellular processes and organismal development. Understanding the 3D arrangement of chromosomes is vital for grasping genome function and regulation. Recent advancements, especially in Hi-C data, have offered significant insights into how chromatin is organized within the nucleus.

Reconstructing chromosome structures from Hi-C data poses challenges, demanding sophisticated computational methods. Traditional approaches rely on optimization techniques to deduce 3D chromatin structures from interaction frequencies. However, they might struggle to accurately capture complex spatial arrangements.

Research on how chromosome structure influences gene expression has gained traction due to its potential in combating genetic diseases. Fluorescence in Situ Hybridization (FISH) technology visualizes chromosome positions in 3D, while Hi-C records interactions between fragments. Researchers employ various models and methods, such as distance-based, contact-based, and probability-based reconstruction techniques, to rebuild chromosome structures from Hi-C data. Despite obstacles, scientists are actively working on high-resolution reconstruction algorithms to precisely rebuild chromosome structures.

Reinforcement learning (RL) presents a promising approach for advancing chromosome structure reconstruction.

RL enables agents to learn optimal behavior through interaction with the environment and feedback as rewards. Applying RL to chromosome modeling allows for adaptive learning to refine reconstructed 3D chromatin structures iteratively.

This research explores state-of-the-art techniques for constructing chromosome structures using reinforcement learning. We examine existing methods like the 3DMax algorithm, the GEM framework, and the Orca model architecture, assessing their strengths and limitations in reconstructing chromosome structures from Hi-C data. Furthermore, we propose a novel RL-based approach for 3D chromosome modeling, discussing its potential benefits and implications for understanding genome organization and function.

By integrating the latest developments in reinforcement learning into chromosome structure reconstruction, this research aims to uncover new insights into the genome's spatial organization and its influence on gene regulation and cellular processes.

Background

Understanding the three-dimensional (3D) folding of the genome is vital for understanding various cellular processes. Reconstructing the 3D spatial arrangements of chromosomes from interaction frequencies derived from chromosome conformation capture (3C)-based data is key to this effort.

However, modeling the 3D chromatin structure faces significant challenges due to uncertain and sparse experimental data, as well as the dynamic and stochastic nature of chromatin organization. Hi-C data provides a matrix where each element represents the interaction frequency between pairs of genomic loci. The goal is to reconstruct the 3D organization of the genome and obtain spatial coordinates for all genomic loci.

Integrating additional constraints, such as nuclear shape and size, with Hi-C data can improve modeling reliability. Methods like multidimensional scaling (MDS), ChromSDE, ShRec3D, and miniMDS address these challenges.

Converting interaction frequencies (F) to spatial distances (D) is crucial but often relies on implicit assumptions, leading to biases and inaccuracies in reconstructed chromatin structures.

Advances in genomic technologies, particularly high-throughput chromosomal conformation capture (Hi-C), have transformed chromatin architecture studies. Reinforcement learning (RL) offers a promising approach, allowing computational agents to learn optimal strategies from Hi-C data. Current methodologies like the 3DMax algorithm, the genomic organization reconstructor based on the conformational energy and manifold learning (GEM) framework, and the Orca model architecture aim to overcome limitations in accurately capturing chromosomal organization.

This research aims to develop a novel approach that integrates reinforcement learning techniques to iteratively refine 3D chromatin models from Hi-C data.

Key Terminology

High-throughput chromosome conformation capture (HiC) is a method in biology to study how chromosomes are structured in a cell. It helps scientists see how different parts of an organism's DNA interact within the cell's nucleus.

Chromosome arrangement isn't random in a cell's nucleus; it's organized in specific ways that affect cell functions like gene expression and DNA copying. HiC experiments explore these arrangements.

Genomic loci are specific spots on a chromosome, like genes or control areas. HiC experiments check how these spots interact with each other, showing spatial relationships and possible functions.

Pairwise interactions in HiC data show where two genomic loci physically touch. If one spot on chromosome 1 often touches another spot on chromosome 2, that's a pairwise interaction.

Interaction frequency shows how often two genomic loci meet in the nucleus. Lots of meetings suggest proximity, while few meetings mean more separation.

HiC file is a format to save HiC experiment data. It includes how often different genomic loci interact, plus details like experiment settings and the exact locations of analyzed spots on chromosomes.

Spatial arrangements of chromosomes mean how they're laid out in a cell's nucleus. It's like arranging furniture in a room, and it affects how genes work and cells function. HiC experiments help us understand these arrangements by seeing how different chromosome parts connect, revealing the 3D shape of DNA in cells.

Related Work

Several significant methodologies and algorithms have been developed for reconstructing chromosome structures from Hi-C data. One notable approach is the Oluwadare et al. (2018), which utilizes gradient ascent optimization to reconstruct chromosome structures. This algorithm optimizes the log-likelihood objective function by incorporating factors such as contact maps, resolution, and interaction frequencies derived from the Hi-C data. Another notable study

by Trieu et al. (2014) focuses on the large-scale reconstruction of the 3D structures of human chromosomes from chromosomal contact data. This approach involves preprocessing Hi-C data to reduce biases, removing contacts with low likelihood ratios, and representing chromosome structures based on observed contacts and non-contacts. These methodologies provide essential insights into the reconstruction of chromosome structures from Hi-C data, contributing to advancements in genomics and biomedicine research.

The GEM framework, as outlined by Zhou (2022), offers a method for reconstructing 3D spatial organizations of chromosomes through manifold learning techniques. The framework embeds neighboring affinities from Hi-C space into 3D Euclidean space, aiming to preserve local structure while mapping from Hi-C space. The Orca model architecture, as described by Zhu et al. (2018), presents a hierarchical sequence encoder and multilevel cascading decoder for multiscale 3D genome prediction. Trained on processed micro-C datasets for H1-ESCs and HFF cells, the model predicts interaction matrices representing pairwise genome interactions at varying resolutions. These diverse methodologies and algorithms contribute to the growing body of research in chromosome structure reconstruction, providing valuable insights and applications for understanding genomic organization and function.

The research paper J Li et al. (2016) outlines a method to reconstruct chromosome 3D structures from Hi-C data using computational techniques and reinforcement learning principles. The dataset includes both simulated and real Hi-C data from mouse embryonic stem cells (mESC) and human GM06990 cells. The methodology entails representing states using 3D coordinates, implementing actions like the shortest-path method and multidimensional scaling (MDS), and optimizing rewards by minimizing the discrepancy between predicted and experimental contact frequencies.

In Gong et al. (2023), the NeRV-3D-DC framework is presented for reconstructing 3D chromosome structures from Hi-C data. The dataset encompasses simulated and actual Hi-C data from GM12878 and IMR90 cell lines. The methodology includes converting contact matrices into distance matrices, reconstructing structures using divide-and-conquer strategies, and evaluating structures using metrics such as root mean square error (RMSE) and Pearson correlation coefficient (PCC).

Zhang et al. (2024) explores the analysis of 3D genomic mapping data, particularly focusing on Hi-C data to identify structural features like compartments, topologically associating domains (TADs), and loops. The paper emphasizes the challenges of achieving consistent feature identification across different methods and datasets.

Wang et al. (2023) introduces the EVRC algorithm for reconstructing chromosome 3D structures from Hi-C data. The dataset comprises simulation data and published Hi-C datasets. The methodology involves converting interaction frequencies into spatial distances, integrating the co-clustering coefficient, and refining structure fit using reinforcement learning.

Song et al. (2021) presents the DQN x-drop algorithm for local sequence alignment utilizing deep reinforcement

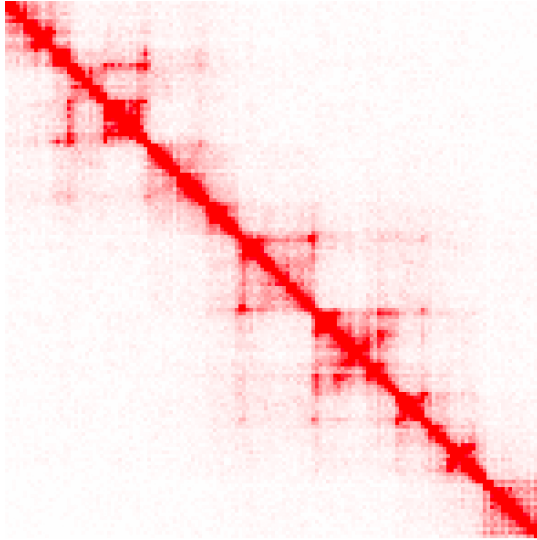


Figure 1: HiC contact map. Image source: Homer et al., retrieved from <http://homer.ucsd.edu/homer/interactions2/HiCmatrices.html>

learning. The dataset consists of DNA sequence pairs. The methodology includes representing alignment states, selecting actions based on alignment directions, and enhancing alignment scores using reinforcement learning.

Z Li et al. (2023) discusses methods for reconstructing 3D chromosome structures from Hi-C data, considering various techniques such as distance-based and contact-based methods. The paper underscores rewards based on the accuracy of reconstructed structures compared to experimental data and outlines methodologies involving state initialization, action selection, reward calculation, policy update, iteration, and validation.

Methodology

HiC Data

A High-Throughput Chromosome Conformation Capture (HiC) file is used in biology to store data from experiments that look at how chromosomes are arranged inside a cell. The data helps infer the proximity and interaction frequency between different genomic loci. HiC files typically contain information about the frequency of interactions between pairs of genomic loci as well as additional metadata such as experimental parameters and genomic coordinates.

Hi-C Data Preprocessing

Hi-C data, which reflects the likelihood of spatial contact between different genomic regions, is first converted into a pairwise distance matrix. This is achieved using the following formula:

$$distance_{ij} = \sqrt{Hi - Cdata_{ij}}$$

Where $distance_{ij}$ represents the pairwise distance between genomic regions i and j , and $Hi - Cdata_{ij}$ denotes the Hi-C data reflecting the likelihood of spatial contact between these regions.

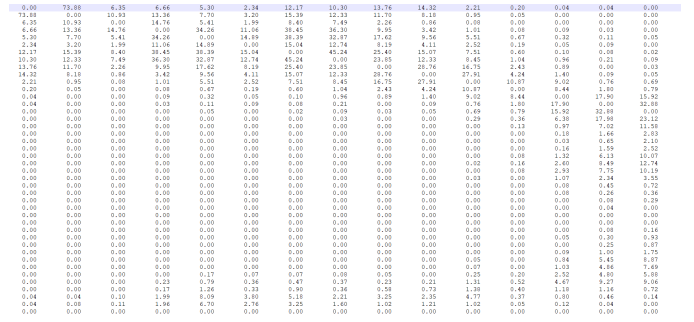


Figure 2: Snapshot of a HiC file

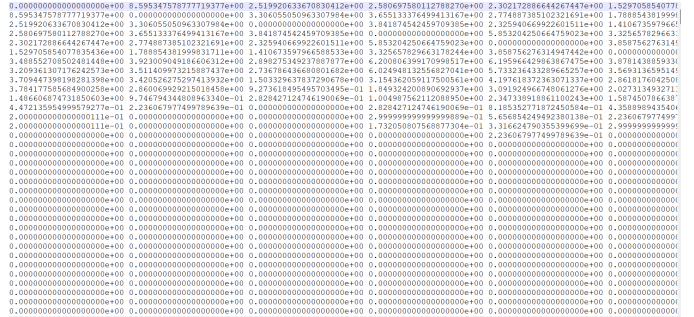


Figure 3: Snapshot of the converted pairwise distance matrix

Distance Matrix

A distance matrix is a mathematical tool used in different fields, including molecular biology. Studies like Hi-C experiments show the spatial relationships between genomic loci. Each cell in the matrix holds a value showing the distance or interaction frequency between pairs of genomic regions. Analyzing this matrix gives insights into how chromosomes are organized in a cell's nucleus.

Conversion to Distance Matrix

The normalized Hi-C contact matrix is then converted into a spatial pairwise distance matrix. This matrix represents the distances between different genomic regions, with a conversion factor (α) determining how these distances are calculated. The conversion is performed using the formula:

$$pairwiselistance_{ij} = (normalizedHi - Cdata_{ij})^{-\alpha}$$

Where $pairwiselistance_{ij}$ represents the spatial distance between genomic regions i and j , and $normalizedHi - Cdata_{ij}$ denotes the normalized Hi-C data.

Normalization of Hi-C Data

The Hi-C contact matrix is normalized using the Knight-Ruiz (KR) algorithm to eliminate systematic biases. This normalization method ensures that the resulting matrix has the same patterns as the original Hi-C data but with reduced noise and more distinct features. The KR normalization process involves:

1. Calculating row and column sums with a small constant added to avoid division by zero.
2. Determining row and column scaling factors.
3. Applying these scaling factors to normalize the Hi-C data.

Floyd Marshall's Algorithm

Floyd-Warshall's algorithm is a fundamental method in graph theory and computer science for finding the shortest paths between all pairs of vertices in a weighted graph. It uses a dynamic programming approach to update a distance matrix and refine the shortest paths. It is particularly useful for negative edge weights and dense graphs with a time complexity of $O(V^3)$, making it an efficient choice for computing the shortest paths in such scenarios.

Graph Representation

The distance matrix is treated as a weighted graph, where genomic regions are vertices and distances between them are weights. This graph is processed to ensure that all pairs of genomic regions meet the triangle inequality, using the Floyd-Warshall shortest path algorithm.

3D Structure Reconstruction

The processed distance matrices are then used as input to implement a method for reconstructing 3D chromosome structures from Hi-C data. This method calculates probability distributions based on distances, measures their divergence, and optimizes 3D coordinates using a conjugate gradient algorithm. The process involves the following steps:

1. **Probability Distribution Calculation:** Probability distributions are calculated based on the distance matrix D . The probability $p_{j|i}$ of a genomic region j being in contact with region i is computed using:

$$p_{j|i} = \frac{\exp(-D_{ij})}{\sum_k \exp(-D_{ik})}$$

2. **Euclidean Distance Calculation:** Euclidean Distance, named after the Greek mathematician Euclid, measures the straight-line distance between two points in Euclidean space. It's used in machine learning and pattern recognition as a similarity measure, forming the basis for algorithms like k-nearest neighbors and k-means.

The Euclidean Distance between two points $P(x_1, y_1)$ and $Q(x_2, y_2)$ in a two-dimensional Cartesian coordinate system is calculated using the Pythagorean theorem:

$$Distance = \sqrt{(x_2 - x_1)^2 + (y_2 - y_1)^2}$$

For points in higher-dimensional spaces, the Euclidean distance formula can be generalized accordingly.

Euclidean distances between genomic regions in 3D space are computed.

3. **Neighbor Probability Calculation:** The probability $q_{j|i}$ of two genomic regions being neighbors of each other in 3D space is calculated based on Euclidean distances.

4. **KL Divergence Calculation:** The Kullback-Leibler (KL) divergence between probability distributions $p_{j|i}$ and $q_{j|i}$ is computed.
5. **Cost Function Calculation:** A cost function is formulated using forward and reverse KL divergences.
6. **Model Training:** The model is trained to obtain 3D coordinates using the conjugate gradient algorithm. Initial 3D coordinates are randomly generated and iteratively optimized to minimize the cost function.
7. **Evaluation:** The reconstructed 3D coordinates are evaluated against actual coordinates using Pearson and Spearman correlation coefficients. These coefficients measure the similarity between the predicted and actual 3D structures.

Methodology for Reinforcement Learning in 3D Chromosome Structure Reconstruction

State Representation:

- The state is represented as a set of 3D coordinates (S) for each genomic bin in the Hi-C data. These coordinates define the spatial positions of genomic regions in a simulated 3D space.

Action Space:

- The action space consists of small perturbations applied to the current set of 3D coordinates. Each action corresponds to a small adjustment in the spatial position of the genomic bins.

Reward Function:

- The reward is calculated based on the closeness of the generated 3D structure to the actual chromosome structure derived from experimental data. It penalizes deviations between the pairwise distances in the generated structure and the actual distances observed in the experimental data.

Policy:

- The policy is learned using the Proximal Policy Optimization (PPO) algorithm. It learns a mapping from states to actions by optimizing the expected cumulative reward over multiple episodes of interaction with the environment.

Environment:

- The RL environment, named `CoordinateGenerationEnv`, simulates the process of generating 3D chromosome structures from Hi-C data. It takes as input the initial set of 3D coordinates and the actual chromosome structure derived from experimental data.

Training Process:

- The RL agent interacts with the environment by taking actions (adjusting the 3D coordinates) and receiving rewards based on the closeness of the generated structure to the actual structure. - The PPO algorithm is trained using a set of trajectories generated by the RL agent. It updates the policy parameters to maximize the expected cumulative reward. - The training process continues for a specified number of iterations or until convergence criteria are met.

Results Analysis:

- The correlation coefficients provide insights into the similarity between the generated 3D structures and the actual chromosome structures. Higher correlation values indicate better agreement between the generated and actual structures.

Evaluation

Correlation Coefficients:

Pearson coefficient

The Pearson coefficient, also known as the Pearson correlation coefficient, measures the strength and direction of the linear relationship between two variables. It ranges from -1 to 1, with 1 indicating a perfect positive relationship, -1 indicating a perfect negative relationship, and 0 indicating no linear relationship. In our research, it is a crucial metric for assessing the correlation between reconstructed chromosome structures and ground-truth data.

Spearman coefficient

The Spearman coefficient, named after Charles Spearman, is a non-parametric measure of rank correlation that assesses the directionality of the association between variables rather than their linearity. It ranges from -1 to 1, with -1 indicating a perfect negative monotonic relationship, 1 indicating a perfect positive monotonic relationship, and 0 indicating no monotonic relationship. In our study, the Spearman coefficient complements the Pearson correlation by providing insights into the ordinal association between reconstructed chromosome structures and reference data.

Results

3D Structure Reconstruction

The reconstructed 3D chromosome structures were obtained through a multi-step process, including the calculation of probability distributions based on distances, the computation of Kullback-Leibler divergence to measure the dissimilarity between the reconstructed and actual structure coordinates, the optimization of a cost function using a conjugate gradient algorithm, and the assessment of the quality of the reconstructed structures using correlation coefficients. The Pearson correlation coefficient quantified the linear relationship between pairwise distances in the reconstructed and actual structures, while the Spearman correlation coefficient measured the monotonic relationship, providing insights into the accuracy and consistency of the reconstructed 3D chromosome structures.

Reinforcement Learning for Coordinate Generation

RL is used to automate 3D structure reconstruction by generating coordinates in genomic regions. A custom RL environment was created, where the agent's actions correspond to adjustments in 3D coordinates. The Proximal Policy Optimization (PPO) algorithm was chosen for its effectiveness in continuous action spaces. The PPO model was trained using

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5.828009927596567330e+00 -2.197268913611325480e+00 2.955282790941653470e+00

Figure 4: Snapshot of the x,y, and z coordinates of the predicted structure

the RL environment and learned to adjust coordinates iteratively to minimize discrepancies. However, the RL agent showed limited success, with low Pearson and Spearman correlation coefficients, indicating it struggled to capture complex spatial relationships in 3D chromosome structures. Despite RL's potential, further research is needed to improve the performance and robustness of RL algorithms in this domain. Addressing the complexities of chromosome organization presents significant challenges that require careful consideration in RL-based approaches.

Table 1: Correlation Coefficients

Statistic	Value
Pearson correlation coefficient	0.0001944
Spearman correlation coefficient	0.0001944

Conclusion

In summary, the research employs RL for 3D chromosome reconstruction, utilizing data preprocessing to ensure the quality and representation of chromosome structures. RL algorithms such as DDPG or PPO are trained to optimize objective functions for reconstructing biologically plausible models. Parameter tuning and optimization improve reconstruction performance, which is evaluated using correlation coefficients, RMSE, and visual inspection. This methodology presents a promising approach to understanding chromatin organization and gene regulation.

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