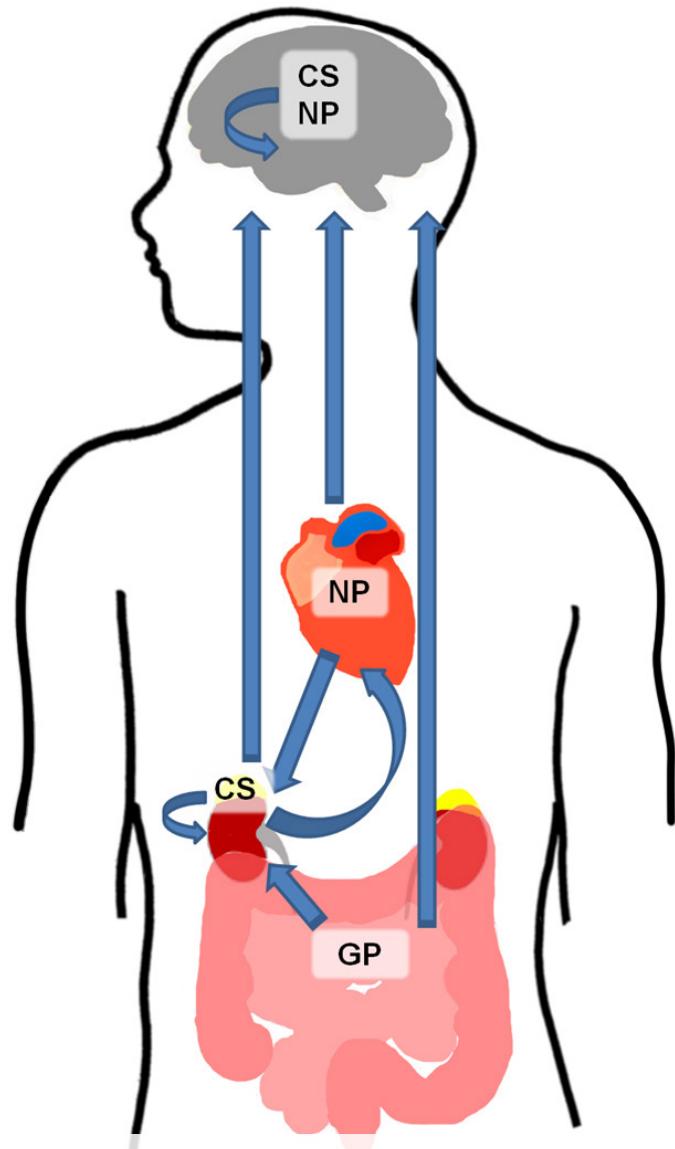


# THE NATRIURETIC HORMONES

EDITED BY: Harvey Craig Gonick and Vardaman M. Buckalew  
PUBLISHED IN: Frontiers in Endocrinology





## **Frontiers Copyright Statement**

© Copyright 2007-2015 Frontiers Media SA. All rights reserved.

All content included on this site, such as text, graphics, logos, button icons, images, video/audio clips, downloads, data compilations and software, is the property of or is licensed to Frontiers Media SA ("Frontiers") or its licensees and/or subcontractors. The copyright in the text of individual articles is the property of their respective authors, subject to a license granted to Frontiers.

The compilation of articles constituting this e-book, wherever published, as well as the compilation of all other content on this site, is the exclusive property of Frontiers. For the conditions for downloading and copying of e-books from Frontiers' website, please see the Terms for Website Use. If purchasing Frontiers e-books from other websites or sources, the conditions of the website concerned apply.

Images and graphics not forming part of user-contributed materials may not be downloaded or copied without permission.

Individual articles may be downloaded and reproduced in accordance with the principles of the CC-BY licence subject to any copyright or other notices. They may not be re-sold as an e-book.

As author or other contributor you grant a CC-BY licence to others to reproduce your articles, including any graphics and third-party materials supplied by you, in accordance with the Conditions for Website Use and subject to any copyright notices which you include in connection with your articles and materials.

All copyright, and all rights therein, are protected by national and international copyright laws.

The above represents a summary only. For the full conditions see the Conditions for Authors and the Conditions for Website Use.

**ISSN 1664-8714**

**ISBN 978-2-88919-709-5**

**DOI 10.3389/978-2-88919-709-5**

## **About Frontiers**

Frontiers is more than just an open-access publisher of scholarly articles: it is a pioneering approach to the world of academia, radically improving the way scholarly research is managed. The grand vision of Frontiers is a world where all people have an equal opportunity to seek, share and generate knowledge. Frontiers provides immediate and permanent online open access to all its publications, but this alone is not enough to realize our grand goals.

## **Frontiers Journal Series**

The Frontiers Journal Series is a multi-tier and interdisciplinary set of open-access, online journals, promising a paradigm shift from the current review, selection and dissemination processes in academic publishing. All Frontiers journals are driven by researchers for researchers; therefore, they constitute a service to the scholarly community. At the same time, the Frontiers Journal Series operates on a revolutionary invention, the tiered publishing system, initially addressing specific communities of scholars, and gradually climbing up to broader public understanding, thus serving the interests of the lay society, too.

## **Dedication to quality**

Each Frontiers article is a landmark of the highest quality, thanks to genuinely collaborative interactions between authors and review editors, who include some of the world's best academicians. Research must be certified by peers before entering a stream of knowledge that may eventually reach the public - and shape society; therefore, Frontiers only applies the most rigorous and unbiased reviews.

Frontiers revolutionizes research publishing by freely delivering the most outstanding research, evaluated with no bias from both the academic and social point of view.

By applying the most advanced information technologies, Frontiers is catapulting scholarly publishing into a new generation.

## **What are Frontiers Research Topics?**

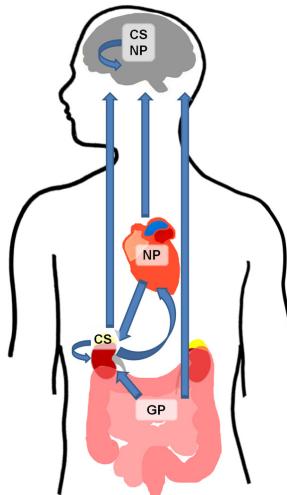
Frontiers Research Topics are very popular trademarks of the Frontiers Journals Series: they are collections of at least ten articles, all centered on a particular subject. With their unique mix of varied contributions from Original Research to Review Articles, Frontiers Research Topics unify the most influential researchers, the latest key findings and historical advances in a hot research area! Find out more on how to host your own Frontiers Research Topic or contribute to one as an author by contacting the Frontiers Editorial Office: [researchtopics@frontiersin.org](mailto:researchtopics@frontiersin.org)

# THE NATRIURETIC HORMONES

Topic Editors:

**Harvey Craig Gonick**, University of California, Berkeley, USA

**Vardaman M. Buckalew**, Wake Forest School of Medicine, USA



Adapted from: Hodes A and Lichtstein D (2014) Natriuretic hormones in brain function. *Front. Endocrinol.* 5:201. doi: 10.3389/fendo.2014.00201

The title follows from the original demonstration by Dr. Hugh de Wardener in 1961 that a humoral agent is produced after extracellular volume expansion which results in a vigorous diuresis and natriuresis. Thus the name of “natriuretic hormone” was coined. In the years that followed several investigators pursued the search for the hormone. What resulted, however, was the discovery of several hormones with different characteristics, all of which were natriuretic. Initially it was found that the hormone was similar in action to ouabain or digoxin, hence the appellation of ouabain-like or digoxin-like. The hormone was found to be an inhibitor of Na-K-ATPase, which would fit with it being a cardiotonic steroid. On the other hand, neither ouabain or digoxin migrated on Sephadex gel filtration in the same locus as the hormone. Other investigators claim to have identified the hormone-initially as a vanadium-diascorbate, later as bufadienolides such as marinobufagenin, yet later as a macrocyclic derivative of inorganic carbon suboxide with a molecular weight of 408 Da. Some support for the latter finding was derived from an earlier report that a semi-purified Sephadex-derived compound was found to have a molecular weight of about 12,000 Da but the active compound, when split from its carrier protein, had a molecular weight of exactly 408 Da. This compound had not been further identified. As further development was the demonstration by Bricker and colleagues that a natriuretic substance could be purified from uremic urine. This turned out to be a xathurenic acid derivative. Meanwhile the focus began to turn to natriuretic peptides derived from heart (ANF and BNP). These peptides have a shorter duration of action than the cardiotonic steroid-like hormone and ANF has proved to be most useful as a measure of heart failure. It should also be stressed that marinobufagenin, like ANF, is elevated in congestive heart failure, whereas the steroid-like hormone is depressed or absent in this state. This review will attempt to describe and contrast the properties of each of the proposed natriuretic hormones, including their locus on Sephadex separation, potency, duration of action, chemical structure (if known), behavior in hypertension, renal failure, heart failure, and brain disease. As most recent work has focussed on marinobufagenin, this hormone will be brought up to date by investigators in the field.

**Citation:** Gonick, H. C., Buckalew, V. M., eds. (2015). The natriuretic hormones. Lausanne: Frontiers Media. doi: 10.3389/978-2-88919-709-5

# Table of Contents

- 04 Editorial: Natriuretic hormones**  
Harvey Craig Gonick and Vardaman M. Buckalew
- 06 Early stages of the natriuretic hormone story**  
Branislav Lichardus
- 09 Endogenous digitalis-like factors: an overview of the history**  
Vardaman M. Buckalew
- 18 The trade-off between dietary salt and cardiovascular disease; a role for Na/K-ATPase signaling?**  
Joe X. Xie, Anna Pearl Shapiro and Joseph Isaac Shapiro
- 26 Natriuretic hormone: the ultimate determinant of the preservation of external sodium balance**  
Neal S. Bricker, Christopher D. Cain and Stewart Shankel
- 32 Natriuretic hormones, endogenous ouabain, and related sodium transport inhibitors**  
John M. Hamlyn
- 42 Natriuretic hormones in brain function**  
Anastasia Hodes and David Lichtstein
- 55 Identification of putative natriuretic hormones isolated from human urine**  
Herbert J. Kramer
- 59 Evidence for a 12 kDa “carrier protein” for natriuretic hormone**  
Harvey C. Gonick
- 66 Spherical oligo-silicic acid SOSA disclosed as possible endogenous digitalis-like factor**  
Franz Kerek and Victor A. Voicu

# Editorial: Natriuretic hormones

Harvey Craig Gonick<sup>1</sup> and Vardaman M. Buckalew<sup>2\*</sup>

<sup>1</sup> Division of Nephrology, Department of Medicine, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA, USA, <sup>2</sup> Section of Nephrology, Department of Medicine, Wake Forest School of Medicine, Winston-Salem, NC, USA

**Keywords:** natriuretic hormones, atrial natriuretic peptides, cardiotonic steroids, hypertension, endogenous ouabain, bufodienolides

The link between renal sodium excretion and body fluid volumes has been a topic of interest since the early days of renal physiology (1). Glomerular filtration rate (GFR) and aldosterone were the first two controllers of renal sodium excretion to be recognized. A “third factor” was discovered when de Wardener and colleagues showed that volume expansion natriuresis still occurred in dogs given supramaximal doses of mineralocorticoids and without an increase in GFR (2). de Wardener et al. suggested that the natriuresis was due to a “natriuretic hormone” (NH), launching a new field of investigation. NH was thus defined as a compound that circulates in blood, the level of which is regulated appropriately by changes in sodium and water balance.

The nine chapters that follow highlight several areas in which the NH field has subsequently developed. First, de Wardener’s original experiments were refined to control potential factors other than GFR and aldosterone. These studies, reviewed by Lichardus (3), confirmed the original observation, and also led to the discovery of the so-called “physical factors” that affect sodium excretion (4). Second, attempts to purify NH from various tissues and body fluids, as summarized in Table 1 by Hamlyn (5), led to the discovery of two important families of factors, atrial natriuretic peptides (ANP), and endogenous cardiotonic steroids (CTS).

The discovery of ANP led rapidly to its characterization as a family of peptides with three major components: ANP derived from cardiac tissue, BNP from brain, and CNP from endothelium (6). This complex system of natriuretic vasodilators, discussed in the chapters by Hamlyn (5), and Hodes and Lichstein (6), continues to be of interest as an NH, a neurotransmitter (6), and a factor with many other functions (7).

In sharp contrast, the identification of endogenous CTS as circulating Na, K ATPase inhibitors has been the source of considerable controversy (8). Two structural classes of CTS have been identified: authentic ouabain, and the bufodienolides, originally identified in toads (9). As discussed by Hamlyn, a bufodienolide is the more likely candidate NH, whereas ouabain in physiologic concentration appears to be antinatriuretic (5). CTS cause vasoconstriction by a mechanism proposed by Blaustein et al. (10) and endogenous CTS have been implicated in the pathophysiology of hypertension (9). Despite overwhelming evidence to the contrary, however, difficulty identifying ouabain in biological fluids continues to be reported (11, 12).

At least three other natriuretic compounds related to control of sodium balance have been identified. As discussed by Gonick (13), a natriuretic protein associated with various forms of human hypertension consists of a 408 Da compound and a 12 kDa carrier protein. The identity of these compounds has not been completed (see below). Two xanthurenic acid derivatives (MW 368 and 284), discussed by Bricker et al. (14), are potential regulators of sodium balance in chronic renal failure. Dietary sodium releases two natriuretic factors from the gastrointestinal tract identified as guanylin and uroguanylin (MW 10.3 and 1.7 kDa), discussed in the paper by Hodes and Lichstein (6).

Two other types of natriuretic compounds that inhibit Na, K ATPase, but are not natriuretic hormones as defined above, are two vanadium diascorbates (MW approximately 400), and two spherical oligo silicic acids (SOSA) (MW 408). These interesting compounds of uncertain significance are

## OPEN ACCESS

### Edited and reviewed by:

Hubert Vaudry,  
University of Rouen, France

### \*Correspondence:

Vardaman M. Buckalew  
buckalew@wakehealth.edu

### Specialty section:

This article was submitted to  
Neuroendocrine Science, a section of  
the journal Frontiers in Endocrinology

Received: 22 June 2015

Accepted: 29 June 2015

Published: 16 July 2015

### Citation:

Gonick HC and Buckalew VM (2015)  
Editorial: Natriuretic hormones.  
*Front. Endocrinol.* 6:108.  
doi: 10.3389/fendo.2015.00108

described in papers by their discoverers Kramer (15) and Kerek and Voicu (16), respectively. The apparent identical molecular weight of SOSA and the compound isolated by Gonick is possibly a coincidence. Alternatively, it may be related to the trace amounts of silicon in human plasma (17) and have significance beyond that implied by the serendipitous discovery of SOSA.

A third area is studies of the physiological mechanism(s) by which these factors might function as natriuretic hormones. ANP release is controlled by stretch of the cardiac atria (18), and in that regard is a classic, volume controlled, NH system as originally conceived (19). Although evidence indicates that CTS are released in response to increased sodium intake (5), no causal mechanism connecting sodium intake and CTS release has been demonstrated. As noted by Hamlyn (5), cerebrospinal fluid sodium concentration controls the release of an unidentified natriuretic factor from brain, causing "CNS natriuresis." The possible role of CTS or some other NH in this phenomenon should be explored.

A fourth area of investigation is studies of collateral, pathophysiological, effects of the various factors. Both ANP and CTS

are vasoactive and have been implicated in the pathophysiology of hypertension (5, 6, 9). Since both appear to be neurotransmitters or neuromodulators (6), their role in hypertension may have both central and peripheral components. Pathological effects of excess CTS in the CNS may also include an etiologic role in mood disorders, including depression and bipolar disorder (6).

Finally, the signaling effects of CTS on Na<sup>+</sup>, K<sup>+</sup> ATPase not dependent on ion pumping, mediated by activation of the tyrosine kinase Src and other signaling molecules, is discussed by Xie et al. (20). This pathway is involved in the natriuretic effect of bufadienolides (21), and is implicated in pathophysiological processes, such as oxidative stress, and organ fibrosis (20).

de Wardener's original hypothesis has led to the discovery of a rich array of factors with multiple biologic activities. ANP is the only classical NH described so far. Future work, including the development of methods for antagonizing these factors, should increase our understanding of the regulation of renal sodium excretion and blood pressure and offer novel treatments for several clinical disorders.

## References

- Epstein FH. Renal excretion of sodium and the concept of volume receptor. *Yale J Biol Med* (1956) 29(3):282–98.
- de Wardener HE, Mills IH, Clappham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intravenous saline in the dog. *Clin Sci* (1961) 21:249–58.
- Lichardus B. Early stages of the natriuretic hormone story. *Front Endocrinol* (2014) 5:180. doi:10.3389/fendo.2014.00180
- Martino JA, Earley LE. Demonstration of a role of physical factors as determinants of the natriuretic response to volume expansion. *J Clin Invest* (1967) 46(12):1963–78. doi:10.1172/JCI105686
- Hamlyn J. Natriuretic hormones, endogenous ouabain, and related sodium transport inhibitors. *Front Endocrinol* (2014) 5:199. doi:10.3389/fendo.2014.00199
- Hodes A, Lichstein D. Natriuretic hormones in brain function. *Front Endocrinol* (2014) 5:201. doi:10.3389/fendo.2014.00201
- Rubattu S, Sciarretta S, Volpe M. Atrial natriuretic peptide gene variants and circulating levels: implications in cardiovascular diseases. *Clin Sci (Lond)* (2014) 127(1):1–13. doi:10.1042/CS20130427
- Blaustein MP. Why isn't endogenous ouabain more widely accepted? *Am J Physiol Heart Circ Physiol* (2014) 307(5):H635–9. doi:10.1152/ajpheart.00404.2014
- Buckalew V. Endogenous digitalis-like factors: an overview of the history. *Front Endocrinol* (2015) 6:49. doi:10.3389/fendo.2015.00049
- Blaustein MP. Sodium ions, calcium ions, blood pressure regulation, and hypertension: a reassessment and a hypothesis. *Am J Physiol* (1977) 232(5):C165–73.
- Baecher S, Kroiss M, Fassnacht M, Vogeser M. No endogenous ouabain is detectable in human plasma by ultra-sensitive UPLC-MS/MS. *Clin Chim Acta* (2014) 431C:87–92. doi:10.1016/j.cca.2014.01.038
- Lewis LK, Yandle TG, Hilton PJ, Jensen BP, Begg EJ, Nicholls MG. Endogenous ouabain is not ouabain. *Hypertension* (2014) 64(4):680–3. doi:10.1161/HYPERTENSIONAHA.114.03919
- Gonick HC. Evidence for a 12 kDa "carrier protein" for natriuretic hormone. *Front Endocrinol* (2014) 5:196. doi:10.3389/fendo.2014.00196
- Bricker NS, Cain CD, Shankel S. Natriuretic hormone: the ultimate determinant of the preservation of external sodium balance. *Front Endocrinol* (2014) 5:212. doi:10.3389/fendo.2014.00212
- Kramer HJ. Identification of putative natriuretic hormones isolated from human urine. *Front Endocrinol* (2015) 6:66. doi:10.3389/fendo.2015.00066
- Kerek F, Voicu VA. Spherical oligo-silicic acid SOSA disclosed as possible endogenous digitalis-like factor. *Front Endocrinol* (2015) 5:233. doi:10.3389/fendo.2014.00233
- Saldanha LF, Gonick HC, Rodriguez HJ, Marmelzat JA, Repique EV, Marcus CL. Silicon-related syndrome in dialysis patients. *Nephron* (1997) 77(1):48–56. doi:10.1159/000190246
- Edwards BS, Zimmerman RS, Schwab TR, Heublein DM, Burnett JC Jr. Atrial stretch, not pressure, is the principal determinant controlling the acute release of atrial natriuretic factor. *Circ Res* (1988) 62(2):191–5. doi:10.1161/01.RES.62.2.191
- de Wardener HE, Clarkson EM. Concept of natriuretic hormone. *Physiol Rev* (1985) 65(3):658–759.
- Xie JX, Shapiro AP, Shapiro JL. The trade-off between dietary salt and cardiovascular disease: a role for Na/K-ATPase signaling? *Front Endocrinol* (2014) 5:97. doi:10.3389/fendo.2014.00097
- Arnaud-Batista FJ, Costa GT, de Oliveira IMB, Costa PPC, Santos CF, Fonteles MC, et al. Natriuretic effect of bufalin in isolated rat kidneys involves activation of the Na<sup>+</sup>-K<sup>+</sup>-ATPase-Src kinase pathway. *Am J Physiol Renal Physiol* (2012) 302(8):F959–66. doi:10.1152/ajprenal.00130.2011

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Gonick and Buckalew. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Early stages of the natriuretic hormone story

Branislav Lichardus<sup>1,2\*</sup>

<sup>1</sup> School of Management, City University of Seattle Programs, Bratislava, Slovakia

<sup>2</sup> Formerly affiliated with The Institute of Experimental Endocrinology, Slovak Academy of Sciences, Bratislava, Slovakia

**Edited by:**

Harvey Craig Gonick, University of California Berkeley, USA

**Reviewed by:**

Vardaman Buckalew, Wake Forest School of Medicine, USA

Mordecai P. Blaustein, University of Maryland School of Medicine, USA

**\*Correspondence:**

Branislav Lichardus, School of Management, Panónska cesta 17, Bratislava 851 04, Slovakia  
e-mail: blichardus@vsm.sk

The paper reviews the early stages of the research on natriuretic hormone. The described experimental work was designed and accomplished in several internationally recognized laboratories where the author was invited to extend his projects. The cross-circulation experiments in animals with acutely increased extracellular fluid volume documented, that in the mechanism of natriuresis – besides a series of the physical natriuretic factors – there is still room for an active humoral natriuretic substance. This substance inhibited the sodium transporting enzyme, Na,K-ATPase, in the frog skin. Analogous inhibition of the renal Na,K-ATPase may be partly responsible for the increased sodium excretion. It was further shown that the extent of natriuresis is positively modulated by the concentration of sodium in the cerebrospinal fluid detected in the anterior-third ventricle region (AV3V) in the brain.

**Keywords:** natriuretic hormone, cross-circulation experiments, Na,K-ATPase, anterior-third ventricle region-AV3V, cerebrospinal fluid sodium concentration

The first International “Symposium on Natriuretic Hormone” was held at the Smolenice Castle, the Congress Center of the Slovak Academy of Sciences in 1969, 12 years after the suggestion of Homer Smith that some such factor could exist (1) and 8 years since the first corroborative experimental data were presented by the team of Hugh de Wardener (2). At the next symposium organized by the same Institution, a decade later we dealt rather with “Natriuretic Hormones” (3). This review is a selected account on the elaboration of the early stages of the hypothesis on the existence of a natriuretic hormone in which I was privileged to participate.

## CROSS-CIRCULATION EXPERIMENTS AND “rein au cou”

In the experiments of de Wardener et al. (2), evidence was advanced for the transfer of a natriuretic material from the donor dog with expanded extracellular fluid volume (ECFV) by the infusion of saline to the cross-perfused recipient dog. The experiments were arranged in such a way that the provoked natriuresis and urine excretion in the recipient animal was neither a result of an increase of the glomerular filtration rate nor of the decrease of known circulating hormones, anti-natriuretic steroids, and vasopressin. Consequently, a third factor was suggested to be involved.

Our group, in pursuing this provocative idea, modified the cross-perfusion experiment in dogs. Only one kidney *in situ* in the recipient dog was cross-perfused under constant perfusion pressure by the donor’s blood in order to expose the recipient kidney to a larger concentration of a natriuretic material than in cross-perfusion of the whole animals. To prevent a possible natriuretic effect of blood dilution by saline infusion rather the blood volume of the donor dog was expanded by an “artificial blood” (suspension of homologous erythrocytes in 6% bovine albumin in Ringer–Locke solution). The cross-perfused kidney increased sodium and urine excretion following the blood volume expansion of the donor dog. However, this “transferred natriuresis” was less pronounced than natriuresis provoked by infusion of saline in the previous experiment of de Wardener. It was concluded that

even if the natriuretic effect of blood dilution is eliminated the cross-perfusion experiments may reveal the appearance of a natriuretic factor in blood of the donor dog following its blood volume expansion (4). It was established in other experiments that the bovine albumin in the “artificial blood” *as such* was not critical for natriuresis evoked by blood volume expansion (5).

Yet, in another experimental set up, the homologous kidney was transplanted to the neck of a dog (“*rein au cou*”) in which vasopressin and creatinine were infused and DOCA was administered intramuscularly at least 3 h before the urine collection started. Subsequently, the blood volume of the dog with the transplanted kidney was expanded by infusion of homologous blood. The experimental conditions assured constant renal arterial and venous pressures in the transplanted kidney, a constant plasma oncotic pressure and constant hematocrit, glomerular filtration rate, and renal blood flow. Moderate, when compared with the renal output of the other kidney *in situ*, but significant increase in urine output and sodium excretion was observed by the transplanted kidney. Since non-hormonal and hormonal factors modulating sodium and water excretion were kept under control in the transplanted kidney, the results indicated more specifically than in previous experiments that a natriuretic humoral material might play a role in the mechanism of natriuresis provoked by blood volume expansion (6).

The operation of a blood-borne natriuretic factor in rat cross-circulation experiment was shown only when so-called sustained fluid volume expansion was achieved by urine reinfusion in the expanded donor animal. This procedure apparently intensified the natriuretic signal to the recipient animal (7).

## A NATRIURETIC OR A DILUTION OF AN ANTI-NATRIURETIC SUBSTANCE?

Our next attempt was to challenge the question that emerged from the cross-circulation experiments, namely, whether the

appearance of a putative blood-borne natriuretic factor in animals with expanded ECFV was the result of an increased concentration of a natriuretic or a dilution of an anti-natriuretic substance. The experiments were performed on un-anesthetized cows, from which substantial volume of blood can be removed without reversing the effect induced by the expansion of their ECFV with 6% dextran in physiological saline (30 ml/kg b.w. at a rate of 100 ml/min). A blood sample of 1000 ml was withdrawn before the infusion and another one at the end of the infusion. The deproteinized plasma was applied intravenously to the assay rats in a volume of 0.2 ml. A non-significant tendency of control samples rather to decrease both the rate of urine flow and sodium excretion in the assay animals, was observed. On the other hand, the samples withdrawn during the ECFV expansion in cows produced statistically significant increases in urine flow, sodium excretion, and the tubular fractional sodium excretion in the assay rats. The sensitivity of the biological assay (i.e., the natriuretic activity applied in 0.2 ml of deproteinized plasma) might indicate that a separate low-molecular weight natriuretic factor might be involved, and that we are not dealing with a mere dilution effect of infusion on an anti-natriuretic activity (8).

The same plasma sample also decreased the short-circuit current representing active sodium transport in the frog skin by a transporting enzyme Na,K-ATPase (9, 10). Buckalew et al. (11) showed that an ultrafiltrate of blood from volume expanded dogs inhibited sodium transport in the toad bladder. It may be another indication for the presence of a natriuretic material in the tested plasma sample as it was proposed that the renal mechanism of natriuresis is via inhibition of the transporting enzyme in the nephron (12).

Further elaboration of the concept of an endogenous inhibitor of the Na,K-ATPase resulted in an attractive hypothesis linking the inhibitor to digoxin-like activity found in some organs, blood, and urine and to the pathogenesis of essential and low-renin arterial hypertension (13–16). Indeed, it was subsequently found that anti-digoxin serum decreased blood pressure in young rats with DOCA-salt hypertension (17). The same anti-digoxin serum, however, was ineffective in suppressing natriuresis induced by ECFV expansion with saline in rats (18). This finding illustrated at that time that the number and nature of substances represented by the endogenous digoxin-like activity had not been satisfactorily answered (19–21), which is true even now.

## INVOLVEMENT OF PERIPHERAL AND CENTRAL NERVOUS SYSTEM IN THE SODIUM BALANCE

The reflex regulation of renal water excretion originating in the stretch receptors of heart atria having the vagal nerves as the afferent limb and vasopressin as an efferent limb (Gauer–Henry reflex) was proposed to be applicable also to the mechanism of renal regulation of sodium excretion. The efferent limb of the reflex was presumed to be a natriuretic factor produced in the brain. However, we found that natriuresis induced by infusion of artificial blood in dogs with either innervated or denervated kidneys was not abolished by bilateral vagotomy. Thus, the analogy of reflex renal sodium control with a modified Gauer–Henry reflex did not seem to be primarily at play (22). We, unfortunately, did not offer at that time a more creative conclusion from these experiments.

It is obvious today that – 14 years before the discovery of atrial natriuretic peptides – we missed a possibility to speculate about the role of *the heart as such* in the renal sodium excretion during the ECFV expansion. Our shortcoming in judgment was partly due to the fact that others found the afferent signal to travel along afferent sympathetic nerves and the spinal cord which, of course, were not interrupted by vagotomy.

It has been shown in various species of anesthetized or conscious experimental animals that increased concentration or dilution of sodium salts in the cerebrospinal fluid (CSF) interfere, respectively, with renal sodium excretion (23). In our studies on conscious sheep, the increase in the CSF sodium concentration and the simultaneous expansion of the ECFV resulted in a much higher increase in renal sodium excretion in comparison to the effect of ECFV expansion in animals with normal CSF sodium concentration. Dilution of CSF sodium prevented completely natriuresis following the ECFV expansion. This is thus another indication of that the brain may be involved in the control of renal regulation of ECFV by monitoring sodium concentration in CSF (24).

The periventricular organs that lack the blood–brain barrier seem to be critical for changes in CSF and also in the systemic ECF sodium concentration. A critical brain area where the sodium concentration or osmolality is monitored is probably the anterior wall of the third ventricle (AV3V). This conclusion is supported by experiments in conscious sheep with ablated AV3V. The non-lesioned control sheep were in a spontaneous water balance, whereas water balance in the animals with chronically ablated AV3V region was re-established by forced application of drinking water through intraruminal tube. It was found that ablation of AV3V region blocks natriuresis to hypertonic but not isotonic NaCl load provided the lesioned sheep is in water balance. McKinley et al. (25, 26) suggested that the AV3V region has a role in regulation of renal Na excretion in conditions where the plasma Na concentration increases. It was further proposed that increased renal Na excretion in response to hypernatremia is another cerebrally mediated osmoregulatory response. However, the AV3V region does not seem to be critical for the mechanism of natriuresis induced by ECFV expansion with isotonic saline (24, 26, 27).

The posterior hypothalamus was also found to be involved in the sodium and ECFV balance. Lesions in the posterior nucleus of the hypothalamus are followed by a renal salt wasting syndrome in rats, cats, and man. In both animal experiments and clinical cases, it was shown that this type of negative sodium balance could not be corrected by adrenal steroids. We tried to identify in rats whether the nuclei in the posterior hypothalamus react to the disturbed sodium balance. The rats were given only a 2% saline to drink for 10 days. A control group drank tap water. The hypothalami were then sectioned and the volume of 200 cell nuclei was determined in each animal of the following nuclei: posterior, ventromedialis, dorsomedialis, and arcuate. The distribution of cell nuclear volume size showed a statistically significant decrease only in the posterior hypothalamic nucleus, which might suggest that it could have been specifically influenced by increased sodium concentration in blood and/or in CSF. At the time of this experiment, changes of volume of cell nuclei in the nucleus posterior hypothalami were taken for an indication of their neuroendocrine activity in connection with salt loading (28, 29).

## CONCLUSION

Using modified cross-perfusion experiments, in which the blood was not diluted, a methodology was put forward to exclude the natriuretic effect of physical factors. It was confirmed that during blood volume expansion a natriuretic or dilution of anti-natriuretic material was revealed.

It was shown, in an attempt to isolate the presumed substance playing a role in natriuresis following the blood or the whole ECF volume expansion that it may be a substance with a natriuretic activity.

The isolated substance also decreased the short-circuit current in the frog skin, which is an indication for its potential to decrease the activity of the transporting enzyme Na,K-ATPase. This action of the natriuretic substance could be its contribution to the renal mechanism of natriuresis following ECFV expansion.

It was shown in conscious sheep that the brain is involved in the control of renal regulation of ECFV by monitoring sodium concentration in CSF. A critical brain area, as indicated by others, where the sodium concentration or osmolality is monitored is the anterior wall of the third ventricle (AV3V).

Indirect evidence was presented for a neuroendocrine activity of the nucleus posterior hypothalami related to the sodium balance in rats.

## REFERENCES

- Cort JH, Lichardus B. Introductory remarks. In: Cort JH, Lichardus B, editors. *Regulation of Body Fluid Volumes by the Kidney—Symposium on Natriuretic Hormone*. Basel: S. Karger (1970). p. 1–10.
- de Wardener HE, Mills IH, Clapham WF, Hayter CJ. Studies on efferent mechanism of sodium diuresis which follows administration of intravenous saline. *Clin Sci* (1961) **21**:249–58.
- Lichardus B. Natriuretic hormones. In: Lichardus B, Schrier RW, Ponec J, editors. *Hormonal Regulation of Sodium Excretion*. Amsterdam: Elsevier/North Holland Biomedical Press (1980). p. 1–7.
- Lichardus B, Pearce JW. Evidence for a humoral natriuretic factor released by blood volume expansion. *Nature* (1966) **209**:407–9. doi:10.1038/209407a0
- Sonnenberg H, Pearce JW. Renal response to measured blood volume expansion in differently hydrated dogs. *Am J Physiol* (1962) **203**:344–52.
- Lichardus B, Nizet A. Water and sodium excretion after blood volume expansion under condition of constant arterial, venous and plasma oncotic pressures and constant haematocrit. *Clin Sci* (1972) **42**:701–9.
- Pearce JW, Sonnenberg H, Lichardus B, Veress AT. Interaction of extrarenal and intrarenal factors in volume natriuresis. In: Cort JH, Lichardus B, editors. *Regulation of Body Fluid Volumes by the Kidney—Symposium on Natriuretic Hormone*. Basel: S. Karger (1970). p. 72–92.
- Lichardus B, Pliska V, Uhrin V, Barth T. The cow as a model for investigating natriuretic activity. *Lancet* (1968) **291**:127–9. doi:10.1016/S0140-6736(68)92728-1
- Sedlakova E, Lichardus B, Cort JH. Plasma saluretic activity: its nature and relation to oxytocin analogs. *Science* (1969) **64**:580–2. doi:10.1126/science.164.3879.580
- Lichardus B, Pliska V, Uhrin V, Barth T. Natriuretic and antinatriuretic activities in deproteinised bovine plasma after dextran infusion. In: Cort JH, Lichardus B, editors. *Regulation of Body Fluid Volumes by the Kidney—Symposium on Natriuretic Hormone*. Basel: S. Karger (1970). p. 114–21.
- Buckalew VM Jr, Martinez FJ, Green WJ. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**:926–35. doi:10.1172/JCI106312
- Kramer HJ, Gonick HC. Effect of extracellular volume expansion on renal Na,K-ATPase and cell metabolism. *Nephron* (1974) **12**:281–96. doi:10.1159/000180341
- Haddy F, Overbeck H. The role of humoral agents in volume expanded hypertension. *Life Sci* (1976) **19**:935–47. doi:10.1016/0024-3205(76)90284-8
- Blaustein MP. Sodium ions, calcium ions, blood-pressure regulation and hypertension – reassessment and hypothesis. *Am J Physiol* (1977) **232**:C165–73.
- Gruber KA, Buckalew VM Jr. Evidence that natriuretic factor is a cascading peptide hormone system. In: Lichardus B, Schrier RW, Ponec J, editors. *Hormonal Regulation of Sodium Excretion*. Amsterdam: Elsevier/North Holland Biomedical Press (1980). p. 342–8.
- de Wardener HE, MacGregor G. The natriuretic hormone and essential hypertension. In: Lichardus B, Schrier RW, Ponec J, editors. *Hormonal Regulation of Sodium Excretion*. Amsterdam: Elsevier/North Holland Biomedical Press (1980). p. 387–90.
- Zicha J, Kunes J, Stolba P. Endogenous digoxin-like factor contributes to the elevation of systemic resistance in rats exposed to high-salt intake from prepuberty. *J Hypertens Suppl* (1985) **3**:S17–9.
- Lichardus B, Zicha J, Ponec J, Stolba P, Pohlova I. Anti-digoxin serum with antipressor properties is ineffective in attenuating natriuresis during extracellular fluid volume expansion. *Physiol Bohemoslov* (1986) **35**:361.
- Lichardus B. Natriuretic hormones in volume natriuresis. *Physiol Res* (1991) **40**:161–8.
- Lichardus B, Kovacs L. Natriuretic hormone as a circulating inhibitor of Na,K-ATPase. In: Bittar E, Bittar N, editors. *Molecular and Cellular Endocrinology*. London: JAI Press Inc (1997). p. 501–15.
- Buckalew VM. Endogenous digitalis-like factors. An historical overview. *Front Biosci* (2005) **10**:2325–34. doi:10.2741/1701
- Pearce JW, Lichardus B. Effects of vagotomy and renal denervation on renal response to blood volume expansion. *J Physiol Pharmacol* (1967) **45**:689–703. doi:10.1139/y67-082
- McKinley MJ, Congiu M, Denton DA, Lichardus B, Park E, Tarjan E, et al. Cerebrospinal fluid composition and homeostatic responses to dehydration. In: Schrier RW, editor. *Vasopressin*. New York, NY: Raven Press (1985). p. 299–309.
- Lichardus B, Ponec J, McKinley MJ, Okolicany J, Gabauer I, Styk J, et al. The anterior third ventricle region is a receptor site for composition rather than volume of body fluids. In: Porter C, Jezova D, editors. *Circulating Regulatory Factors and Neuroendocrine Function*. New York, NY: Plenum Press (1990). p. 211–26.
- McKinley MJ, Congiu M, Denton DA, Park RG, Penschow J, Simpson JB, et al. The anterior wall of the third cerebral ventricle and homeostatic responses to dehydration. *J Physiol (Paris)* (1984) **79**:421–7.
- McKinley MJ, Lichardus B, McDougal JG, Weisinger RS. Periventricular lesions block natriuresis to hypertonic but not isotonic NaCl loads. *Am J Physiol* (1992) **262**:F98–107.
- Lichardus B, McKinley MJ, Denton DA, Ponec J. Brain involvement in the regulation of renal sodium excretion. *Klin Wochenschr* (1987) **65**:33–9.
- Lichardus B, Mitro A, Cort JH. Size of cell nuclei in the hypothalamus of the rat as a function of salt loading. *Am J Physiol* (1965) **208**:1075–7.
- Bajusz E. Modern trends in neuroendocrinology with special reference to clinical problems. A concluding review. In: Bajusz E, editor. *An Introduction to Clinical Neuroendocrinology*. New York, NY: S. Karger (1967). p. 428–534.

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Received:** 26 August 2014; **paper pending published:** 19 September 2014; **accepted:** 07 October 2014; **published online:** 11 November 2014.

**Citation:** Lichardus B (2014) Early stages of the natriuretic hormone story. *Front. Endocrinol.* **5**:180. doi: 10.3389/fendo.2014.00180

This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Lichardus. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Endogenous digitalis-like factors: an overview of the history

**Vardaman M. Buckalew\***

Medical Center Boulevard, Wake Forest School of Medicine, Winston Salem, NC, USA

**Edited by:**

Pierrette Gaudreau, Centre Hospitalier de l'Université de Montréal Research Center, Canada

**Reviewed by:**

Ricardo Borges, University of La Laguna, Spain

Pierrette Gaudreau, Centre Hospitalier de l'Université de Montréal Research Center, Canada

Mordecai P. Blaustein, University of Maryland School of Medicine, USA

**\*Correspondence:**

Vardaman M. Buckalew, Medical Center Boulevard, Wake Forest School of Medicine, Winston Salem, NC 27157, USA

e-mail: buckalew@wakehealth.edu

The sodium pump is a ubiquitous cell surface enzyme, a Na<sub>+</sub> K ATPase, which maintains ion gradients between cells and the extracellular fluid (ECF). The extracellular domain of this enzyme contains a highly conserved binding site, a receptor for a plant derived family of compounds, the digitalis glycosides. These compounds inhibit the enzyme and are used in the treatment of congestive heart failure and certain cardiac arrhythmias. The highly conserved nature of this enzyme and its digitalis receptor led to early suggestions that endogenous regulators might exist. Recent examination of this hypothesis emerged from research in two separate areas: the regulation of ECF volume by a natriuretic hormone (NH), and the regulation of peripheral vascular resistance by a circulating inhibitor of vascular Na<sub>+</sub> K ATPase. These two areas merged with the hypothesis that NH and the vascular Na<sub>+</sub> K ATPase inhibitor were in fact the same entity, and that it played a causative role in the pathophysiology of certain types of hypertension. The possibility that multiple endogenous digitalis-like factors (EDLFs) exist emerged from efforts to characterize the circulating enzyme inhibitory activity. In this review, the development of this field from its beginnings is traced, the current status of the structure of EDLFs is briefly discussed, and areas for future development are suggested.

**Keywords:** natriuretic hormone, digitalis-like factor, ouabain, marinobufagenin, bufadienolides, cardenolides

## BACKGROUND

The regulation of salt and water excretion by the kidneys has occupied investigators since at least from the beginning of the 20th century. Highlights of these investigations are documented in early reviews of the subject, notably those by Epstein (1) and Smith (2) in the 1950s. These reviews supported the existence of a receptor-integrator-effector reflex by which changes in some component of the extracellular fluid (ECF) volume (“volume receptors”) caused appropriate changes in renal sodium excretion. Both “efferent factors,” the numerous hemodynamic, humoral, and neural factors known to directly affect renal sodium excretion, and “afferent factors,” the stimuli that activate the efferent factors, were reviewed.

Smith, separating the factors influencing free water excretion from those affecting sodium excretion, considered the mechanism of the latter as being similar to the former. Based on these and other evolutionary considerations, he postulated that the proposed effector for sodium excretion, which he called “Hormone X”, was an anti-natriuretic hormone, analogous to antidiuretic hormone, which had evolved to conserve sodium as our primitive ancestors made their “ascent through the brackish waters of the estuary/to the salt poor lakes and ponds” (Strauss) (2). Aldosterone had been identified in the early 1950s, so Smith’s Hormone X was clearly proposed as an additional volume sensitive sodium retaining hormone, decreased levels of which would cause natriuresis in response to increased ECF volume.

At the time of Smith’s review, it was well established that two factors were preeminent in controlling renal sodium excretion, glomerular filtration rate (GFR), and aldosterone. Thus, Smith’s

review set the stage for exploration for a “third factor,” a term which did not originate with Smith. The earliest investigators to use the term in print, if not the first, were Bricker et al., who were searching for the mechanisms contributing to the progressive increase in the absolute rate of sodium excretion per nephron as the nephron population decreased in chronic renal failure (3). Bricker et al. were the first to recognize that similar mechanisms might contribute to both volume expansion natriuresis and the renal adaptation to chronic renal failure.

## THE CONCEPT OF NATRIURETIC HORMONE

Four years after Smith’s review, the mechanism of “volume expansion natriuresis” was addressed in a classic paper by deWardener et al. published in 1961 (4). They showed that natriuresis caused by saline infusion in dogs given large doses of mineralocorticoid was not abolished when GFR was reduced below initial levels by constriction of the aorta above the renal arteries. Furthermore, they showed that blood circulated from volume expanded dogs (donor) to euolemic dogs (recipient) caused natriuresis in the recipient. Based on these studies, deWardener et al. suggested that volume expansion increased the circulating level of some natriuretic substance, and the concept of “natriuretic hormone” was born.

Three problems quickly emerged after this ground-breaking study was published. First, although the cross circulation studies were careful to control the volume of the recipient dog, the possible effects of blood dilution by the saline infusion in the donor dog were not. In addition, the possibility that the natriuresis in the

recipient dog might be due to suppression of an anti-natriuretic factor as suggested by Smith had not been definitively eliminated. Each of these issues was addressed in the burst of work in other laboratories that followed the original paper by de Wardener et al. Importantly, these investigators effectively dealt with the dilution issue in a subsequent cross circulation study in which the recipient dog was infused with blood from a reservoir in which blood from both donor and recipient was in equilibrium (5). Other studies addressing the issue of dilution were published by Lichardus et al. and others (6). Studies of the effects of blood dilution on renal sodium excretion subsequently led to exploration of the so called “physical factors” on renal tubular sodium reabsorption by a number of laboratories (7).

In essentially all refinements of the cross circulation studies, the natriuresis in the recipient was much less than that in the donor animal, a finding that was never entirely explained, but some interesting observations were made. For example, response in the recipient was increased by infusing blood from the donor into the aorta just above the renal arteries (8), suggesting a short biologic half-life of the circulating natriuretic factor (9). Also, recipient response was enhanced by preventing the donor from excreting the administered volume load, suggesting some effect of “sustained” volume expansion, an interesting but poorly defined concept that has not been explored further (10).

### MECHANISMS OF NATRIURESIS

The cross circulation studies did not distinguish between the presence of a natriuretic substance versus suppression of an anti-natriuretic substance. To make that distinction, a number of laboratories reported natriuretic activity in plasma, urine, and/or kidney tissue of volume expanded animals (11–13), the mechanism of which drew immediate interest. The question was whether the factor caused changes in renal hemodynamics or directly inhibited tubular sodium transport systems. The first studies suggesting the latter were performed by Bricker et al. in which inhibition of p-aminohippurate (PAH) transport by rabbit kidney cortical slices was inhibited by plasma from volume expanded subjects (14). Inhibition of transport in renal tubular epithelium was subsequently shown in isolated tubular cells (15).

Other early studies utilized anuran membranes as models of renal tubular sodium transport. Cort and Lichardus reported inhibition of sodium transport as measured by Ussing's short circuit current (SCC) technique in isolated frog skin by deproteinized, concentrated plasma extracts with very high sodium concentrations (16). In more extensive studies using plasma ultrafiltrates with physiological salt concentrations from volume expanded dogs, Buckalew et al. in 1970 showed similar effects on toad bladder SCC of *bufo marinus* (17). Ussing and others had demonstrated that the SCC in anuran membrane was due to active sodium transport, and could be inhibited by ouabain. The demonstration that the putative natriuretic hormone inhibited tubular transport and SCC set the stage for investigation of the effect of this factor or factors on Na, K ATPase. The initial attempts to relate natriuretic hormone (NH) to Na, K ATPase inhibition were unsuccessful. In the best documented studies, Katz et al. were unable to show inhibition of the enzyme in renal cortical microsomes from volume expanded dogs and rats, or an effect of plasma dialyzates

from these animals on renal microsomal Na, K ATPase isolated from euvolemic animals (18). However, Gonick et al. subsequently reported that a natriuretic fraction extracted from renal tissue and plasma of volume expanded animals inhibited SCC in frog skin and ouabain sensitive Na, K ATPase isolated from whole rat kidney (19, 20).

Studies of the effect of plasma, and extracts of plasma and urine of volume expanded subjects on sodium excretion in assay animals, usually rats, demonstrated two basic patterns that differed primarily in time to peak and duration of effect. The shorter acting pattern showed an immediate onset, a peak effect in 40–60 min, and duration of about 120 min (21, 22). The longer-acting pattern exhibited a delay in onset of 10–60 min, a peak effect in 2–3 h, and duration longer than 3 h (22). Some initial purification studies indicated that the more rapidly acting factor was found in fractions containing low molecular weight substances, and the longer acting factor appeared in fractions containing high molecular weight substances (23).

### NATRIURETIC HORMONE AS INHIBITOR OF Na, K ATPase

Two major developments in the late 1970s and early 1980s caused a shift in the direction of NH research. The discovery in 1981 of atrial natriuretic factor (ANF) by DeBold et al. (24) and its subsequent characterization as a peptide signaling cascade present in many organs displaced most other lines of investigation with regard to the existence and nature of a NH. Early studies did not show an effect of ANF on Na, K ATPase (25, 26); however, subsequent studies revealed a more complex situation (27–29). Nevertheless, it was clear from the early work that ANF and the natriuretic inhibitor of renal epithelial transport systems dependent on Na, K ATPase were two entirely different systems. However, very little further work on the non-ANF NH hypothesis was performed. Instead, the focus shifted to the second major development, namely, that NH might be an inhibitor of vascular Na, K ATPase that could also be a causative factor in certain types of hypertension.

The suggestion that some types of hypertension, especially those associated with ECF volume expansion, might be due to a circulating inhibitor of vascular Na, K ATPase evolved from studies of the phenomenon of potassium-induced vasodilation. Overbeck et al. showed that the dilator response to potassium, but not to other agents, was suppressed in the forelimb of the rat with two kidney, one clip hypertension and the dog with one-kidney, one-wrap hypertension (30). Subsequent studies showed that potassium-induced vasodilation was completely blocked by ouabain, leading to the hypothesis that the vasodilation was due to stimulation of vascular smooth muscle Na, K ATPase. According to this hypothesis, stimulation of the electrogenic sodium pump led to hyperpolarization, decreased voltage sensitive influx of calcium, and hence vascular relaxation (31).

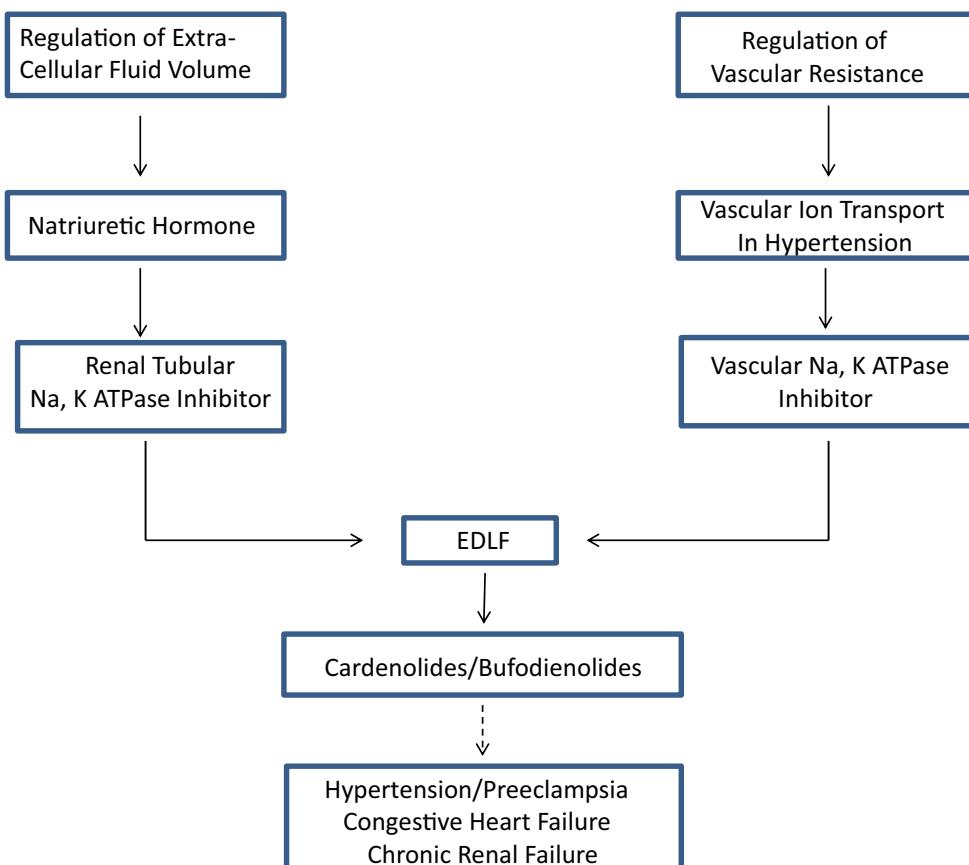
Reduced serum potassium produced identical effects in the opposite direction. That is, hypokalemia was associated with vasoconstriction and suppressed Na pump activity, suggesting a cause and effect relationship. As predicted by this paradigm, vascular depolarization was found in several volume expanded hypertension models. Thus, the hypothesis was proposed that vasoconstriction leading to hypertension might be caused by generalized inhibition of vascular Na, K ATPase activity (32). In a further

refinement of the hypothesis in 1976, based on a review of then existing evidence for a humoral factor that slowly increased blood pressure in both animal models and humans with hypertension, Haddy et al. proposed that Na, K ATPase inhibition in vascular tissue, and hence vasoconstriction, might be due to a circulating factor (33). They, in fact, proposed in that review that the postulated circulating inhibitor of Na, K ATPase might be “natriuretic hormone.” Thus, the two fields of ECF volume regulation and regulation of vascular tone in volume expanded models of hypertension were brought together in the search for a common, explanatory factor (**Figure 1**).

A unifying explanation for the connection between vascular tone, intracellular calcium concentration, and the sodium pump was proposed by Blaustein in 1977 (34). The model was based on the presence of a Na–Ca exchanger located in the plasma membrane, driven by the intracellular–extracellular sodium gradient. According to the hypothesis, supported by kinetic calculations (34), inhibition of the sodium pump by the NH would cause increased vasoconstriction by inhibiting the outward transport of calcium by the Na–Ca exchanger.

## VOLUME EXPANDED MODELS OF HYPERTENSION

The NH hypothesis of hypertension raised the question of how volume regulation by a potentially vasoconstrictor NH occurred in normal versus hypertensive subjects. Volume expanded models of hypertension involved some manipulation that reduced the ability of the kidney to excrete sodium. This approach was based on the concept proposed by Guyton et al. (35) that all hypertension was caused by an abnormal relationship between blood pressure and renal sodium excretion. According to this hypothesis, in normal subjects, renal adaptation to increases and decreases in sodium intake occur without any or with only small changes in systemic blood pressure. However, increased blood pressure is required to maintain ECF volume regulation in the presence of impaired renal sodium excretion through the phenomenon of “pressure diuresis.” Guyton postulated the rise in pressure was due to a volume induced increase in cardiac output and the consequent “long term autoregulation” (i.e., vasoconstriction) that ensued. Thus, ECF volume is maintained at the expense of increased peripheral vascular resistance and high blood pressure.



**FIGURE 1 |**The concept of an endogenous digitalis-like factor (EDLF) that inhibits Na, K ATPase in a manner similar to the cardiac glycosides developed from two lines of investigation (see text), the response of renal sodium excretion to extracellular fluid volume expansion, and the regulation of peripheral vascular resistance in hypertension. Research on the identity of EDLF

indicates that mammalian species synthesize two classes of steroids that are either identical to, or analogs of those found in plants (cardenolides) and toads (bufodienolides). Evidence suggests that one or more of these compounds may be involved in the pathophysiology of various hypertensive disorders, chronic renal failure, and congestive heart failure.

Based on this theory, several investigators proposed a unifying hypothesis incorporating NH that explained many observations then existing in the literature (36, 37). According to this formulation, the defect in renal response to increases in sodium and water intake in hypertensive subjects leads to increases in NH, vascular Na, K ATPase inhibition, vasoconstriction, and increased blood pressure. Volume homeostasis is maintained in the presence of a defect in renal sodium excretion by both the rise in blood pressure through the mechanism of pressure natriuresis, and by the effect of the NH to inhibit renal tubular sodium reabsorption. The difference between hypertensive and normotensive subjects was, as suggested by deWardener and MacGregor, that the former would be in a “state of continuous correction of a slightly expanded extracellular volume,” resulting in a sustained elevation of NH (37). While this concept may have some general validity, the role of EDLF differs in various forms of hypertension (see below).

### ENDOGENOUS DIGITALIS-LIKE FACTOR

Because of the suggestion that NH might be an inhibitor of Na, K ATPase, it was subsequently referred to as “ouabain-like” or “digitalis-like.” This terminology became more than nomenclature as the field turned to proving the true digitalis-like nature of the circulating factor.

### THE CONCEPT OF ENDOGENOUS DRUG-LIKE COMPOUNDS

The demonstration that the specificity and actions of some drugs were due to drug binding to stereospecific receptors had led to speculation that naturally occurring endogenous compounds existed that bound specifically to these receptors (38). The discovery of endogenous opioids was a direct result of this hypothesis (39). In 1976, Ginzler et al. proposed, as an extension of this concept, that antigen–antibody binding specificity might be analogous to drug-receptor binding specificity (40). That is, an antibody specific for a drug might recognize the same structure as the specific receptor for that drug, and could act as a “surrogate” receptor. This hypothesis had at least two interesting implications. First, antibodies to drugs might recognize endogenous compounds that utilize the same receptor as the drug; and second, antibodies to drugs (or endogenous compounds) might be used to block the effects of those compounds by displacing them from their receptor. The second possibility had already been anticipated by a number of investigators including the demonstration that digoxin antibodies would reverse the clinical manifestations of digoxin intoxication (41).

Based on these concepts, Gruber et al. showed in 1980 that plasma of volume expanded dogs but not euvolemic dogs contained a factor that cross reacted with digoxin antibodies in a specific fashion; i.e., the dose response curve in the digoxin radioimmunoassay (RIA) of the endogenous factor was parallel to that of authentic digoxin (42). Furthermore, plasma extracts containing the digoxin immunoreactive compound inhibited Na, K ATPase, providing further evidence for a true EDLF that had some structural and functional similarity to digoxin. The finding also suggested that digoxin RIAs could be used to study plasma levels of this factor and numerous studies of mammalian “digoxin-like” factor were soon published (43). However, studies using this approach are subject to non-specific cross reactivity of various

interfering substances in the digoxin RIA and have led to some confusion (44, 45). Interestingly, the first demonstration of an endogenous substance that cross reacts with digoxin antibodies was in newborns, who were suspected of having been poisoned with digoxin (46).

### CHARACTERIZATION OF EDLF

Subsequent to the work briefly described above, numerous attempts have been made to purify and identify the principal factor responsible for the digitalis-like factor demonstrated in volume expanded subjects. According to the initial hypothesis, a truly endogenous, natriuretic, hypertension promoting digitalis-like factor would have the following characteristics. First of all, it would be synthesized endogenously, and secreted under the control of relevant physiological or pathophysiological stimuli. Second, it would inhibit renal and vascular Na, K ATPase in a “ouabain-like” fashion; that is, it would bind to the same receptor and have similar effects on the enzyme as ouabain. Thirdly, its inhibition of renal tubular and vascular Na, K ATPase would cause natriuresis and vasoconstriction, respectively. Unfortunately, attempts to identify such a factor have been complicated by the fact that inhibition of the enzyme in various assay systems is a non-specific effect of many diverse compounds (47). As a result, numerous candidate structures have been identified, including steroids, lipids, peptides, and a variety of other novel compounds (45, 48, 49). A complete review of these reports is beyond the scope of this paper. Rather, we have chosen to focus on a class of compounds that are “digitalis-like” steroids, and which meet the theoretical criteria outlined above, with one exception. They have not been shown unequivocally to be “endogenous” since their synthetic pathway has not been completely elucidated, although preliminary studies suggest they are synthesized in the adrenal gland (50–53).

In 1991, Hamlyn et al. reported purification of a compound indistinguishable from ouabain by mass spectroscopy from 3001 of human plasma (54). Subsequent work seemed to confirm this observation and indicated that mammalian ouabain is present in multiple body fluids and tissues. However, the issue of whether mammalian tissues contain authentic ouabain has remained highly controversial (55–57) despite substantial evidence in support of this finding (58).

Amphibian species have been known for many years to synthesize a number of different steroids called bufadienolides that inhibit Na, K ATPase in a manner similar to the cardenolides (59). Dienolides differ from cardenolides in the structure of the lactone ring, which contains six members and two unsaturated double bonds compared to five members and one double bond in the cardenolides (43). Both cardenolides and dienolides have a 14  $\beta$  hydroxyl group and a cis tertiary configuration of the C/D ring junction. Lichstein et al. identified a bufadienolide in toad skin and plasma as resibufogenin (60). They also demonstrated that the concentration of dienolides in toad skin was regulated by the salt content and osmolality of its aquatic environment (61, 62).

Bagrov et al. purified a digitalis-like compound from toad venom (63), which they subsequently identified as a previously described bufadienolide marinobufagenin (MBG) (64). Subsequently, purification of a substance from urine of patients after an acute myocardial infarction by high pressure liquid

chromatography confirmed a structure indistinguishable from authentic MBG (65). Using a polyclonal antibody to toad MBG, they demonstrated increased concentration of a compound recognized by that antibody in plasma of volume expanded dogs (66) and rats (67), and patients with preeclampsia (68). Using antibodies specific for ouabain and MBG, they demonstrated that mammalian plasma contains both ouabain-like and MBG-like compounds (66). Subsequent work has demonstrated that the MBG-like compound meets essentially all the criteria originally postulated for the EDLF-type NH described above (69, 70).

Yoshika et al. have shown that MBG immunoreactivity secreted by adrenomedullary derived cells in tissue culture is composed of at least two compounds, MBG and a related compound marinobufotoxin (MBT) (71). MBT was shown to increase blood pressure when administered intraperitoneally to rats (71).

### EDLF AND HYPERTENSION

Using multiple assays for EDLF, numerous studies have attempted to show some correlation between plasma EDLF levels and the blood pressure in human and experimental hypertension, details of which have been previously reviewed and are beyond the scope of this paper (69, 72–80). Many of these studies have relied on measurements using RIA technology with antibodies raised against the compound(s) of interest. As noted, these studies are subject to cross reactivity with compounds other than those to which the antibody was raised. Despite these problems, it seems likely that some EDLF(s) are elevated in some forms of human and experimental hypertension and may play a role in its pathophysiology.

Although most studies of the role of EDLF in hypertension have focused on the circulating factors, it seems likely that endogenous ouabain plays a role in certain types of hypertension through a pathway in the central nervous system (CNS). EDLF has been demonstrated in hypothalamic and pituitary extracts of rats, a compound (or compounds) that crossreacts with a polyclonal anti-ouabain antibody (81). Extensive studies by Huang et al. have shown increases in this compound in the hypothalamus of Dahl salt-sensitive rats (82), spontaneously hypertensive rats (SHR) (83), and normal rats in which blood pressure is increased by an increase in cerebrospinal fluid sodium concentration (84). The critical role of brain EDLF in each of these models was demonstrated by prevention of the rise in blood pressure by CNS administration of a commercially available antigen binding fragment (FAB) of an antidigoxin antibody known to cross react with EDLF (Digibind®) (see below). A further complexity in the hypertension promoting CNS EDLF system has been demonstrated by studies of central infusion of angiotensin II in rats. This hypertension provoking maneuver causes an increase in circulating endogenous ouabain through activation of a neuronal pathway involving central aldosterone (85).

An integrated role for both endogenous cardenolides and bufadienolides in hypertension in Dahl salt-sensitive rats is suggested by studies showing that release of MBG is controlled by the CNS ouabain pathway discussed above (86). Further studies on the role of CNS pathways in the pathophysiology of hypertension, and in controlling circulating endogenous digitalis-like factors (EDLFs) and blood pressure should be of interest.

### REVERSAL OF EDLF EFFECTS BY FUNCTIONAL ANTAGONISTS

Several functional antagonists of EDLF have been reported to reverse the effects of Na, K ATPase inhibition in various clinical and experimental situations, among which are anti-dogoxin and anti-ouabain antiserum, and two steroid compounds, rostafuroxin and resibufogenin, that may be receptor antagonists of one or another component of EDLF.

Digibind® is a purified FAB of a sheep anti-digoxin antibody developed for the treatment of digoxin intoxication that is no longer available commercially. Studies using Digibind® as a probe to assess the possible role of EDLF in hypertensive subjects assume that it will cross react with EDLF, and that in large enough doses will displace EDLF from its receptor, analogous to its effect in digoxin toxicity. A number of studies are compatible with this formulation. Krep et al. showed that Digibind® reduced blood pressure in the DOCA-salt rat model (87). Kaide et al. obtained the same results in a 5/6 reduced renal mass model (88). In the latter study, no effect of Digibind® on blood pressure was observed in sham-operated controls, suggesting that the blood pressure reduction was not due to some non-specific or toxic effect of Digibind® such as an anaphylactoid reaction. Mann et al. had suggested the latter, but their studies were done with commercial preparations other than Digibind® (89). In addition to these *in vivo* studies, Krep et al. showed that Digibind® reversed the contraction response of isolated aorta to an EDLF isolated from peritoneal dialysis fluid (90). Digibind® has also been reported to reduce blood pressure in several hypertension models when given directly into the CNS (84, 91), to block the natriuresis of saline infusion in dogs (66), and to improve neonatal outcomes in fetuses born to patients with preeclampsia (92).

Antibodies against other glycosides have also been shown to lower blood pressure in animal models. Anti-ouabain antibodies had no effect on blood pressure in normal rats (93). However, immunization against ouabain prevented the development of hypertension in Dahl salt-sensitive rats (94), and reduced sodium excretion in normal rats (93). Also, administration of MBG antibodies lowered blood pressure in Dahl salt-sensitive rats (95). These studies suggest that whatever EDLF might be, whether single or multiple compounds, it cross reacts with antibodies against several candidate EDLFs.

In addition to the work with antibodies, two possible receptor antagonists of EDLF have been reported. Rostafuroxin® is a digitoxigenen derivative that selectively displaces ouabain from the Na, K ATPase receptor (96). The compound lowered blood pressure in Milan hypertensive rats (97), but failed to lower blood pressure in clinical trials in essential hypertension in humans (77). Resibufogenin (RBG) is a bufadienolide isolated from toad skin (60) and the traditional Chinese medication Chan Su made from dried toad venom (98). RBG has a structure that only differs from MBG by one oxygen atom on the 5-position of the steroid nucleus (99). Although RBG has “digitalis-like activity” (inhibits *in vitro* Na, K ATPase activity and ouabain binding) (60), RBG has been shown to lower blood pressure in rat models of preeclampsia and DOCA-salt hypertension (100), both of which have elevated levels of MBG. These data suggest RBG antagonizes at least some effects of MBG, but the exact mechanism has not been elucidated.

Although similar in structure, different cardenolides and bufadienolides have surprising and unpredictable species and tissue differences in their biological actions (101–103), including antagonism of each other's effects (104–107). The mechanism of the latter phenomenon has been extensively studied by Song et al. (105). They proposed a complex set of models in which  $\alpha$  and  $\beta$  subunits of Na, K ATPase can function as tetraprotomers with varying degrees of aggregation and pump inhibition. This concept may lead to an entirely new method of manipulating sodium pump function that could have clinical implications.

It should also be noted that ACTH induced hypertension in rats can be prevented by making the  $\alpha$ -2 Na, K ATPase receptor for cardiac glycosides resistant to those compounds through genetic manipulation (108). This clearly implicates endogenous Na, K ATPase inhibitor(s) in the etiology of this type of experimental hypertension.

## SUMMARY IN RETROSPECT AND FUTURE DIRECTIONS

The search for a factor that regulates renal sodium excretion in response to increased blood volume, a NH, stimulated by the experiments of deWardener et al. (4) has produced a huge body of literature, which can no longer be reviewed in a single article. This review, an update of an earlier one (109), emphasizes how the search for a NH converged with studies of the mechanism of increased vascular resistance in hypertension, resulting in the discovery of EDLF(s) (Figure 1). This important discovery has widespread physiologic and pathophysiologic implications and explains, at least in part, the highly conserved nature of the ouabain binding site on membrane Na, K ATPase.

The fact is that, after all the work briefly summarized here, an amazing degree of complexity to a relatively simple if naïve concept has emerged. Regarding the original NH proposal of deWardener et al., no single entity has emerged that fits their hypothesis, and new physiologically relevant natriuretic factors may yet be discovered (9). Atrial natriuretic peptides are clearly volume sensitive natriuretic factors that likely play some role in the renal response to acute volume expansion. ANPs have multiple effects including vasodilation, and one or more of these peptides probably play some role in the pathophysiology of hypertension and congestive heart failure (110). Interactions between ANP and several EDLFs have been demonstrated, which have potentially important physiologic and pathophysiologic implications (27, 111, 112), and further work in the area can be anticipated.

Ouabain, MBG, and other bufadienolides continue to be investigated as putative physiological regulators of renal and cardiovascular function, but no clear integrating hypothesis has yet emerged. MBG appears to fit the criteria for the circulating factor proposed by the original NH hypothesis better than ouabain (72), but ouabain is clearly involved in the regulation of sodium excretion by the CNS and further work on this system is anticipated (9, 73). The intrarenal mechanism by which bufadienolides cause natriuresis, which involves the recently discovered signaling function of Na, K ATPase, should be of ongoing interest (113). The synthetic pathways and tissue(s) origin for the various EDLFs have not yet been completely determined, and high priority should be given to this project (58). If EDLFs play a role in normal physiology and some hypertensive states, as current evidence

indicates, interference with their synthesis or antagonism of their effects should provide further insights, and possibly new targets for antihypertensive drugs.

Anti-digoxin antibodies interact with a broad range of EDLFs (114) and have been utilized in both experimental animals and man, with some interesting results (87, 88, 115, 116). Although the commercial preparation used in most of these studies is no longer available, another commercially available preparation of digoxin antibodies (DigiFab<sup>®</sup>) has similar if not identical cross reactivity with EDLF (116–118). Finally, the ability of individual cardiotonic steroids (CTS) to interfere with the effects of other CTS has added another layer of complexity and suggests another novel approach to the study of and possible therapy of various conditions in which CTS may be involved (105).

Despite ongoing controversy regarding some of the details (58), the hypothesis that an endogenous regulator(s) of the ouabain binding site on the Na, K ATPase enzyme is involved in control of the cardiovascular system has proved to be immensely fertile. Further investigation of these endogenous regulators holds great promise for a better understanding of cardiovascular physiology, the pathophysiology of a diverse set of clinical disorders, including hypertension, preeclampsia, chronic renal failure, congestive heart failure, and cancer (70), and the intricate complexities of the ouabain binding site (108, 119, 120).

## REFERENCES

- Epstein FH. Renal excretion of sodium and the concept of volume receptor. *Yale J Biol Med* (1956) **29**(3):282–98.
- Smith HW. Salt and water volume receptors: an exercise in physiologic apologetics. *Am J Med* (1957) **23**(4):623–52. doi:10.1016/0002-9343(57)90232-2
- Bricker NS. The control of sodium excretion with normal and reduced nephron populations. The pre-eminence of third factor. *Am J Med* (1967) **43**(3):313–21. doi:10.1016/0002-9343(67)90188-X
- de Wardener HE, Mills IH, Clappham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intravenous saline in the dog. *Clin Sci* (1961) **21**:249–58.
- Bahlmann J, McDonald SJ, Ventom MG, de Wardener HE. The effect on urinary sodium excretion of blood volume expansion without changing the composition of blood in the dog. *Clin Sci* (1967) **32**(3):403–13.
- Lichardus B. Early stages of the natriuretic hormone story. *Front Endocrinol (Lausanne)* (2014) **5**:180. doi:10.3389/fendo.2014.00180
- Martino JA, Earley LE. Demonstration of a role of physical factors as determinants of the natriuretic response to volume expansion. *J Clin Invest* (1967) **46**(12):1963–78. doi:10.1172/JCI105686
- Blythe WB, D'Avila D, Gitelman HJ, Welt LG. Further evidence for a humoral natriuretic factor. *Circ Res* (1971) **28**(5):21–31. doi:10.1161/01.RES.28.5.II-21
- Hamlyn J. Natriuretic hormones, endogenous ouabain, and related sodium transport inhibitors. *Front Endocrinol (Lausanne)* (2014) **5**:199. doi:10.3389/fendo.2014.00199
- Sonnenberg H, Veress AT, Pearce JW. A humoral component of the natriuretic mechanism in sustained blood volume expansion. *J Clin Invest* (1972) **51**(10):2631–44. doi:10.1172/JCI107081
- Kruck F. Influence of humoral factors on renal tubular sodium handling. *Nephron* (1969) **6**(3):205–16. doi:10.1159/000179729
- Sealey JE, Kirshman JD, Laragh JH. Natriuretic activity in plasma and urine of salt-loaded man and sheep. *J Clin Invest* (1969) **48**(12):2210–24. doi:10.1172/JCI106187
- Viskoper JR, Czaczkes JW, Schwartz N, Ullmann TD. Natriuretic activity of a substance isolated from human urine during the excretion of a salt load. Comparison of hypertensive and normotensive subjects. *Nephron* (1971) **8**(6):540–8. doi:10.1159/000179959
- Bricker NS, Klahr S, Purkerson M, Schultze RG, Avioli LV, Birge SJ. In vitro assay for a humoral substance present during volume expansion and uraemia. *Nature* (1968) **219**(5158):1058–9. doi:10.1038/2191058a0

15. Clarkson EM, Talner LB, de Wardener HE. The effect of plasma from blood volume expanded dogs on sodium, potassium and PAH transport of renal tubule fragments. *Clin Sci* (1970) **38**(6):617–27.
16. Cort JH, Dousa T, Pliska V, Lichardus B, Safarova J, Vranesic M, et al. Saluretic activity of blood during carotid occlusion in the cat. *Am J Physiol* (1968) **215**(4):921–7.
17. Buckalew VM Jr, Martinez FJ, Green WE. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**(5):926–35. doi:10.1172/JCI106312
18. Katz AI, Genant HK. Effect of extracellular volume expansion on renal cortical and medullary Na K ATPase. *Pflugers Arch* (1971) **330**(2):136–48. doi:10.1007/BF00643030
19. Hillyard SD, Lu E, Gonick HC. Further characterization of the natriuretic factor derived from kidney tissue of volume-expanded rats. Effects on short-circuit current and sodium-potassium-adenosine triphosphatase activity. *Circ Res* (1976) **38**(4):250–5. doi:10.1161/01.RES.38.4.250
20. Gonick HC, Kramer HJ, Paul W, Lu E. Circulating inhibitor of sodium-potassium-activated adenosine triphosphatase after expansion of extracellular fluid volume in rats. *Clin Sci Mol Med* (1977) **53**(4):329–34.
21. Buckalew VM Jr, Nelson DB. Natriuretic and sodium transport inhibitory activity in plasma of volume-expanded dogs. *Kidney Int* (1974) **5**(1):12–22. doi:10.1038/ki.1974.2
22. Pearce JW, Veress AT, Sonnenberg H. Time course of onset and decay of humoral natriuretic activity in the rat. *Can J Physiol Pharmacol* (1975) **53**(5):734–41. doi:10.1139/y75-102
23. Buckalew VMGKA. Natriuretic hormone. 2nd ed. In: Epstein M, editor. *The Kidney in Liver Disease*. New York: Elsevier (1983). p. 479–99.
24. De Bold AJ, Borenstein HB, Veress AT, Sonnenberg H. A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats. *Life Sci* (1981) **28**(1):89–94. doi:10.1016/0024-3205(81)90370-2
25. Sagnella GA, Nolan DA, Shore AC, MacGregor GA. Effects of synthetic atrial natriuretic peptides on sodium-potassium transport in human erythrocytes. *Clin Sci (Lond)* (1985) **69**(2):223–6.
26. Thibault G, Garcia R, Cantin M, Genest J. Atrial natriuretic factor. Characterization and partial purification. *Hypertension* (1983) **5**(2 Pt 2):175–80. doi:10.1161/01.HYP.5.2\_Pt\_2.175
27. Buckalew VM. Atrial peptides modify the effect of marinobufagenin on sodium pumps: implications for blood pressure control. *Hypertension* (2006) **48**(6):1029–30. doi:10.1161/01.HYP.0000248119.54493.18
28. Fedorova OV, Agalakova NI, Morrell CH, Lakatta EG, Bagrov AY. ANP differentially modulates marinobufagenin-induced sodium pump inhibition in kidney and aorta. *Hypertension* (2006) **48**(6):1160–8. doi:10.1161/01.HYP.0000248129.20524.d0
29. Correa AH, Choi MR, Gironacci M, Valera MS, Fernandez BE. Signaling pathways involved in atrial natriuretic factor and dopamine regulation of renal Na, K ATPase activity. *Regul Pept* (2007) **138**(1):26–31. doi:10.1016/j.regpep.2006.08.001
30. Overbeck HW. Vascular responses to cations, osmolality, and angiotensin in renal hypertensive dogs. *Am J Physiol* (1972) **223**(6):1358–64.
31. Chen WT, Brace RA, Scott JB, Anderson DK, Haddy FJ. The mechanism of the vasodilator action of potassium. *Proc Soc Exp Biol Med* (1972) **140**(3):820–4. doi:10.3181/00379727-140-36560
32. Haddy FJ. Potassium and blood vessels. *Life Sci* (1975) **16**(10):1489–97. doi:10.1016/0024-3205(75)90065-X
33. Haddy FJ, Overbeck HW. The role of humoral agents in volume expanded hypertension. *Life Sci* (1976) **19**(7):935–47. doi:10.1016/0024-3205(76)90284-8
34. Blaustein MP. Sodium ions, calcium ions, blood pressure regulation, and hypertension: a reassessment and a hypothesis. *Am J Physiol* (1977) **232**(5):C165–73.
35. Guyton AC. Abnormal renal function and autoregulation in essential hypertension. *Hypertension* (1991) **18**(5 Suppl):III49–53. doi:10.1161/01.HYP.18.5\_Suppl.III49
36. Haddy F, Pamnani M, Clough D. The sodium-potassium pump in volume expanded hypertension. *Clin Exp Hypertens* (1978) **1**(3):295–336. doi:10.3109/10641967809068611
37. de Wardener HE, MacGregor GA. Dahl's hypothesis that a saluretic substance may be responsible for a sustained rise in arterial pressure: its possible role in essential hypertension. *Kidney Int* (1980) **18**(1):1–9. doi:10.1038/ki.1980.104
38. Gruber KA. Endogenous druglike substances: implications and approaches to their study. *Perspect Biol Med* (1982) **26**(1):51–61. doi:10.1353/pbm.1982.0017
39. Hughes J, Smith TW, Kosterlitz HW, Fothergill LA, Morgan BA, Morris HR. Identification of two related pentapeptides from the brain with potent opiate agonist activity. *Nature* (1975) **258**(5536):577–80. doi:10.1038/258577a0
40. Gintzler AR, Levy A, Spector S. Antibodies as a means of isolating and characterizing biologically active substances: presence of a non-peptide, morphine-like compound in the central nervous system. *Proc Natl Acad Sci U S A* (1976) **73**(6):2132–6. doi:10.1073/pnas.73.6.2132
41. Curd J, Smith TW, Jaton JC, Haber E. The isolation of digoxin-specific antibody and its use in reversing the effects of digoxin. *Proc Natl Acad Sci U S A* (1971) **68**(10):2401–6. doi:10.1073/pnas.68.10.2401
42. Gruber KA, Whitaker JM, Buckalew VM Jr. Endogenous digitalis-like substance in plasma of volume-expanded dogs. *Nature* (1980) **287**(5784):743–5. doi:10.1038/287743a0
43. Schoner W. Endogenous digitalis-like factors. *Prog Drug Res* (1993) **41**:249–91.
44. Graves SW, Williams GH. Endogenous digitalis-like natriuretic factors. *Annu Rev Med* (1987) **38**:433–44. doi:10.1146/annurev.me.38.020187.002245
45. Goto A, Yamada K, Yagi N, Yoshioka M, Sugimoto T. Physiology and pharmacology of endogenous digitalis-like factors. *Pharmacol Rev* (1992) **44**(3):377–99.
46. Woolfson RG, Poston L, de Wardener HE. Digoxin-like inhibitors of active sodium transport and blood pressure: the current status. *Kidney Int* (1994) **46**(2):297–309. doi:10.1038/ki.1994.275
47. Shlevin HH, Na, K ATPase inhibitors: implications for new drug discovery. *Drug Dev Res* (1984) **4**(3):275–84. doi:10.1002/ddr.430040305
48. Buckalew VM, Haddy FJ. Circulating natriuretic factors in hypertension. In: Laredo J, Brenner BM, editors. *Hypertension: Pathophysiology, Diagnosis and Management*. New York, NY: Raven Press, Ltd (1990). p. 939–54.
49. Wechter WJ, Benaksas EJ. Natriuretic hormones. *Prog Drug Res* (1990) **34**:231–60.
50. Dmitrieva RI, Bagrov AY, Lalli E, Sassone-Corsi P, Stocco DM, Doris PA. Mammalian bufadienolide is synthesized from cholesterol in the adrenal cortex by a pathway that is independent of cholesterol side-chain cleavage. *Hypertension* (2000) **36**(3):442–8. doi:10.1161/01.HYP.36.3.442
51. Dmitrieva RI, Lalli E, Doris PA. Regulation of adrenocortical cardiotonic steroid production by dopamine and PKA signaling. *Front Biosci* (2005) **10**:2489–95. doi:10.2741/1713
52. Hamlyn JM, Laredo J, Shah JR, Lu ZR, Hamilton BP. 11-hydroxylation in the biosynthesis of endogenous ouabain: multiple implications. *Ann NY Acad Sci* (2003) **986**:685–93. doi:10.1111/j.1749-6632.2003.tb07283.x
53. Laredo J, Shah JR, Lu ZR, HAMILTON BP, Hamlyn JM. Angiotensin II stimulates secretion of endogenous ouabain from bovine adrenocortical cells via angiotensin type 2 receptors. *Hypertension* (1997) **29**(1):401–7. doi:10.1161/01.HYP.29.1.401
54. Hamlyn JM, Blaustein MP, Bova S, DuCharme DW, Harris DW, Mandel F, et al. Identification and characterization of a ouabain-like compound from human plasma. *Proc Natl Acad Sci U S A* (1991) **88**(14):6259–63. doi:10.1073/pnas.88.21.9907-d
55. Baecher S, Kroiss M, Fassnacht M, Vogeser M. No endogenous ouabain is detectable in human plasma by ultra-sensitive UPLC-MS/MS. *Clin Chim Acta* (2014) **431C**:87–92. doi:10.1016/j.cca.2014.01.038
56. Lewis LK, Yandle TG, Lewis JG, Richards AM, Pidgeon GB, Kaaja RJ, et al. Ouabain is not detectable in human plasma. *Hypertension* (1994) **24**(5):549–55. doi:10.1161/01.HYP.24.5.549
57. Lewis LK, Yandle TG, Hilton PJ, Jensen BP, Begg EJ, Nicholls MG. Endogenous ouabain is not ouabain. *Hypertension* (2014) **64**(4):680–3. doi:10.1161/HYPERTENSIONAHA.114.03919
58. Blaustein MP. Why isn't endogenous ouabain more widely accepted? *Am J Physiol Heart Circ Physiol* (2014) **307**(5):H635–9. doi:10.1152/ajpheart.00404.2014
59. Flier J, Edwards MW, Daly JW, Myers CW. Widespread occurrence in frogs and toads of skin compounds interacting with the ouabain site of Na, K ATPase. *Science* (1980) **208**(4443):503–5. doi:10.1126/science.6245447
60. Lichtstein D, Kachalsky S, Deutsch J. Identification of a ouabain-like compound in toad skin and plasma as a bufodienolide derivative. *Life Sci* (1986) **38**(14):1261–70. doi:10.1016/0024-3205(86)90418-2
61. Lichtstein D, Gati I, Babila T, Haver E, Katz U. Effect of salt acclimation on digitalis-like compounds in the toad. *Biochim Biophys Acta* (1991) **1073**(1):65–8. doi:10.1016/0304-4165(91)90183-H

62. Lichtstein D, Gati I, Haver E, Katz U. Digitalis-like compounds in the toad *Bufo viridis*: tissue and plasma levels and significance in osmotic stress. *Life Sci* (1992) **51**(2):119–28. doi:10.1016/0024-3205(92)90005-A
63. Bagrov AY, Roukoyatkina NI, Fedorova OV, Pinaev AG, Ukhanova MV. Digitalis-like and vasoconstrictor effects of endogenous digoxin-like factor(s) from the venom of *Bufo marinus* toad. *Eur J Pharmacol* (1993) **234**(2–3):165–72. doi:10.1016/0014-2999(93)90950-M
64. Bagrov AY, Roukoyatkina NI, Pinaev AG, Dmitrieva RI, Fedorova OV. Effects of two endogenous Na, K ATPase inhibitors, marinobufagenin and ouabain, on isolated rat aorta. *Eur J Pharmacol* (1995) **274**(1–3):151–8. doi:10.1016/0014-2999(94)00735-P
65. Bagrov AY, Fedorova OV, Dmitrieva RI, Howald WN, Hunter AP, Kuznetsova EA, et al. Characterization of a urinary bufadienolide Na, K ATPase inhibitor in patients after acute myocardial infarction. *Hypertension* (1998) **31**(5):1097–103. doi:10.1161/01.HYP.31.5.1097
66. Bagrov AY, Fedorova OV, Dmitrieva RI, French AW, Anderson DE. Plasma marinobufagenin-like and ouabain-like immunoreactivity during saline volume expansion in anesthetized dogs. *Cardiovasc Res* (1996) **31**(2):296–305. doi:10.1016/S0008-6363(95)00208-1
67. Fedorova OV, Doris PA, Bagrov AY. Endogenous marinobufagenin-like factor in acute plasma volume expansion. *Clin Exp Hypertens* (1998) **20**(5–6):581–91. doi:10.3109/10641969809053236
68. Lopatin DA, Ailamazian EK, Dmitrieva RI, Shpen VM, Fedorova OV, Doris PA, et al. Circulating bufadienolide and cardenolide sodium pump inhibitors in preeclampsia. *J Hypertens* (1999) **17**(8):1179–87. doi:10.1097/00004872-199917080-00018
69. Bagrov AY, Shapiro JI. Endogenous digitalis: pathophysiologic roles and therapeutic applications. *Nat Clin Pract Nephrol* (2008) **4**(7):378–92. doi:10.1038/ncpneph0848
70. Bagrov AY, Shapiro JI, Fedorova OV. Endogenous cardiotonic steroids: physiology, pharmacology, and novel therapeutic targets. *Pharmacol Rev* (2009) **61**(1):9–38. doi:10.1124/pr.108.00071
71. Yoshika M, Komiyama Y, Konishi M, Akizawa T, Kobayashi T, Date M, et al. Novel digitalis-like factor, marinobufotoxin, isolated from cultured Y-1 cells, and its hypertensive effect in rats. *Hypertension* (2007) **49**(1):209–14. doi:10.1161/01.HYP.0000250433.64202.78
72. Bagrov AY, Fedorova OV. Cardenolide and bufadienolide ligands of the sodium pump. How they work together in NaCl sensitive hypertension. *Front Biosci* (2005) **10**:2250–6. doi:10.2741/1694
73. Hamlyn JM, Blaustein MP. Salt sensitivity, endogenous ouabain and hypertension. *Curr Opin Nephrol Hypertens* (2013) **22**(1):51–8. doi:10.1097/MNH.0b013e32835b36ec
74. Hauck C, Fishman WH. Systemic hypertension: the roles of salt, vascular Na, K ATPase and the endogenous glycosides, ouabain and marinobufagenin. *Cardiol Rev* (2012) **20**(3):130–8. doi:10.1097/CRD.0b013e31823c835c
75. Manunta P, Hamlyn JM, Simonini M, Messaglio E, Lanzani C, Bracale M, et al. Endogenous ouabain and the renin-angiotensin-aldosterone system: distinct effects on Na handling and blood pressure in human hypertension. *J Hypertens* (2011) **29**(2):349–56. doi:10.1097/HJH.0b013e32833ea821
76. Schoner W, Scheiner-Bobis G. Endogenous cardiac glycosides: hormones using the sodium pump as signal transducer. *Semin Nephrol* (2005) **25**(5):343–51. doi:10.1016/j.semnnephrol.2005.03.010
77. Staessen J, Thijss L, Stolarz-Skrzypek K, Bacchieri A, Barton J, Espositi E, et al. Main results of the ouabain and adducin for specific intervention on sodium in hypertension trial (OASIS-HT): a randomized placebo-controlled phase-2 dose-finding study of rostafuroxin. *Trials* (2011) **12**(1):13. doi:10.1186/1745-6215-12-13
78. Manunta P, Messaglio E, Ballabeni C, Sciarrone MT, Lanzani C, Ferrandi M, et al. Plasma ouabain-like factor during acute and chronic changes in sodium balance in essential hypertension. *Hypertension* (2001) **38**(2):198–203. doi:10.1161/01.HYP.38.2.198
79. Wang JG, Staessen JA, Messaglio E, Nawrot T, Fagard R, Hamlyn JM, et al. Salt, endogenous ouabain and blood pressure interactions in the general population. *J Hypertens* (2003) **21**(8):1475–81. doi:10.1097/00004872-200308000-00010
80. Tomaschitz A, Piecha G, Ritz E, Meinitzer A, Haas J, Pieske B, et al. Marinobufagenin in essential hypertension and primary aldosteronism: a cardiotonic steroid with clinical and diagnostic implications. *Clin Exp Hypertens* (2015) **21**(2):108–15. doi:10.3109/10641963.2014.913604
81. Ferrandi M, Manunta P, Balzan S, Hamlyn JM, Bianchi G, Ferrari P. Ouabain-like factor quantification in mammalian tissues and plasma: comparison of two independent assays. *Hypertension* (1997) **30**(4):886–96. doi:10.1161/01.HYP.30.4.886
82. Huang BS, Leenen FH. Brain “ouabain” mediates the sympathoexcitatory and hypertensive effects of high sodium intake in Dahl salt-sensitive rats. *Circ Res* (1994) **74**(4):586–95. doi:10.1161/01.RES.74.4.586
83. Huang BS, Leenen FH. Brain ‘ouabain,’ sodium, and arterial baroreflex in spontaneously hypertensive rats. *Hypertension* (1995) **25**(4 Pt 2):814–7. doi:10.1161/01.HYP.25.4.814
84. Huang BS, Harmsen E, Yu H, Leenen FH. Brain ouabain-like activity and the sympathoexcitatory and pressor effects of central sodium in rats. *Circ Res* (1992) **71**(5):1059–66. doi:10.1161/01.RES.71.5.1059
85. Hamlyn JM, Linde CI, Gao J, Huang BS, Golovina VA, Blaustein MP, et al. Neuroendocrine humoral and vascular components in the pressor pathway for brain angiotensin II: a new axis in long term blood pressure control. *PLoS One* (2014) **9**(10):e108916. doi:10.1371/journal.pone.0108916
86. Fedorova OV, Agalakova NI, Talan MI, Lakatta EG, Bagrov AY. Brain ouabain stimulates peripheral marinobufagenin via angiotensin II signalling in NaCl-loaded Dahl-S rats. *J Hypertens* (2005) **23**(8):1515–23. doi:10.1097/01.hjh.0000174969.79836.8b
87. Krep H, Price DA, Soszynski P, Tao QF, Graves SW, Hollenberg NK. Volume sensitive hypertension and the digoxin-like factor. Reversal by a Fab directed against digoxin in DOCA-salt hypertensive rats. *Am J Hypertens* (1995) **8**(9):921–7. doi:10.1016/0895-7061(95)00181-N
88. Kaide J, Ura N, Torii T, Nakagawa M, Takada T, Shimamoto K. Effects of digoxin-specific antibody Fab fragment (Digibind) on blood pressure and renal water-sodium metabolism in 5/6 reduced renal mass hypertensive rats. *Am J Hypertens* (1999) **12**(6):611–9. doi:10.1016/S0895-7061(99)00029-1
89. Mann JF, Miemietz R, Ganter U, Ritz E. Haemodynamic effects of intact digoxin antibody and its Fab fragments in experimental hypertension. *J Hypertens* (1987) **5**(5):543–9. doi:10.1097/00004872-198710000-00006
90. Krep HH, Graves SW, Price DA, Lazarus M, Ensign A, Soszynski PA, et al. Reversal of sodium pump inhibitor induced vascular smooth muscle contraction with digibind. Stoichiometry and its implications. *Am J Hypertens* (1996) **9**(1):39–46. doi:10.1016/0895-7061(95)00260-X
91. Huang BS, Leenen FH. Blockade of brain “ouabain” prevents sympathoexcitatory and pressor responses to high sodium in SHR. *Am J Physiol* (1996) **271**(1 Pt 2):H103–8.
92. Lam GK, Hopoate-Sitake M, Adair CD, Buckalew VM, Johnson DD, Lewis DF, et al. Digoxin antibody fragment, antigen binding (Fab), treatment of preeclampsia in women with endogenous digitalis-like factor: a secondary analysis of the DEEP Trial. *Am J Obstet Gynecol* (2013) **209**(2):e1–6. doi:10.1016/j.ajog.2013.04.010
93. Nesher M, Dvela M, Igboekwe VU, Rosen H, Lichtstein D. Physiological roles of endogenous ouabain in normal rats. *Am J Physiol Heart Circ Physiol* (2009) **297**(6):H2026–34. doi:10.1152/ajpheart.00734.2009
94. Gomez-Sanchez EP, Gomez-Sanchez CE, Fort C. Immunization of Dahl SS/jr rats with an ouabain conjugate mitigates hypertension. *Am J Hypertens* (1994) **7**(7 Pt 1):591–6.
95. Fedorova OV, Talan MI, Agalakova NI, Lakatta EG, Bagrov AY. Endogenous ligand of alpha(1) sodium pump, marinobufagenin, is a novel mediator of sodium chloride-dependent hypertension. *Circulation* (2002) **105**(9):1122–7. doi:10.1161/hc0902.104710
96. Quadri L, Bianchi G, Cerri A, Fedrizzi G, Ferrari P, Gobbini M, et al. 17 beta-(3-furyl)-5 beta-androstan-3 beta, 14 beta, 17 alpha-triol (PST 2238). A very potent antihypertensive agent with a novel mechanism of action. *J Med Chem* (1997) **40**(11):1561–4. doi:10.1021/jm970162e
97. Ferrari P, Ferrandi M, Valentini G, Bianchi G. Rostafuroxin: an ouabain antagonist that corrects renal and vascular Na, K ATPase alterations in ouabain and adducin-dependent hypertension. *Am J Physiol Regul Integr Comp Physiol* (2006) **290**(3):R529–35. doi:10.1152/ajpregu.00518.2005
98. Xu W, Luo H, Zhang Y, Shan L, Li H, Yang M, et al. Simultaneous determination of five main active bufadienolides of Chan Su in rat plasma by liquid chromatography tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* (2007) **859**(2):157–63. doi:10.1016/j.jchromb.2007.09.026
99. Uddin MN, Allen SR, Jones RO, Zawieja DC, Kuehl TJ. Pathogenesis of pre-eclampsia: marinobufagenin and angiogenic imbalance as biomarkers of the syndrome. *Transl Res* (2012) **160**(2):99–113. doi:10.1016/j.trsl.2012.01.005

100. Puschett JB, Agunanne E, Uddin MN. Emerging role of the bufadienolides in cardiovascular and kidney diseases. *Am J Kidney Dis* (2010) **56**(2):359–70. doi:10.1053/j.ajkd.2010.01.023
101. Dvela M, Rosen H, Feldmann T, Nesher M, Lichtstein D. Diverse biological responses to different cardiotonic steroids. *Pathophysiology* (2007) **14**(3–4):159–66. doi:10.1016/j.pathophys.2007.09.011
102. Julian A, Linde CI, Pulina MV, Baryshnikov SG, Papparella I, Hamlyn JM, et al. Activation of c-SRC underlies the differential effects of ouabain and digoxin on Ca<sup>2+</sup> signaling in arterial smooth muscle cells. *Am J Physiol Cell Physiol* (2013) **304**(4):C324–33. doi:10.1152/ajpcell.00337.2012
103. Manunta P, Hamilton J, Rogowski AC, Hamilton BP, Hamlyn JM. Chronic hypertension induced by ouabain but not digoxin in the rat: antihypertensive effect of digoxin and digitoxin. *Hypertens Res* (2000) **23**:S77–85. doi:10.1291/hypres.23.Supplement\_S77
104. Feldmann T, Glukmann V, Medvedev E, Shpolansky U, Galili D, Lichtstein D, et al. Role of endosomal Na<sup>+</sup>-K<sup>+</sup>-ATPase and cardiac steroids in the regulation of endocytosis. *Am J Physiol Cell Physiol* (2007) **293**(3):C885–96. doi:10.1152/ajpcell.00602.2006
105. Song H, Karashima E, Hamlyn JM, Blaustein MP. Ouabain-digoxin antagonism in rat arteries and neurones. *J Physiol* (2014) **592**(Pt 5):941–69. doi:10.1113/jphysiol.2013.266866
106. Nesher M, Shpolansky U, Viola N, Dvela M, Buzaglo N, Ben-Ami HC, et al. Ouabain attenuates cardiototoxicity induced by other cardiac steroids. *Br J Pharmacol* (2010) **160**(2):346–54. doi:10.1111/j.1476-5381.2010.00701.x
107. Huang BS, Kudlac M, Kumarathasan R, Leenen FH. Digoxin prevents ouabain and high salt intake-induced hypertension in rats with sinoaortic denervation. *Hypertension* (1999) **34**(4):733–8. doi:10.1161/01.HYP.34.4.733
108. Dostanic-Larson I, Van Huysse JW, Lorenz JN, Lingrel JB. The highly conserved cardiac glycoside binding site of Na<sup>+</sup>, K<sup>+</sup>-ATPase plays a role in blood pressure regulation. *Proc Natl Acad Sci U S A* (2005) **102**(44):15845–50. doi:10.1073/pnas.0507358102
109. Buckalew VM. Endogenous digitalis-like factors. An historical overview. *Front Biosci* (2005) **10**:2325–34. doi:10.2741/1701
110. Rubattu S, Calvieri C, Pagliaro B, Volpe M. Atrial natriuretic peptide and regulation of vascular function in hypertension and heart failure: implications for novel therapeutic strategies. *J Hypertens* (2013) **31**(6):1061–72. doi:10.1097/HJH.0b013e32835ed5eb
111. Fedorova OV, Kashkin VA, Zakharkova IO, Lakatta EG, Bagrov AY. Age-associated increase in salt sensitivity is accompanied by a shift in the atrial natriuretic peptide modulation of the effect of marinobufagenin on renal and vascular sodium pump. *J Hypertens* (2012) **30**(9):1817–26. doi:10.1097/HJH.0b013e328356399b
112. Liu LP, Hong L, Yu L, Li HY, Ding DZ, Jin SJ, et al. Ouabain stimulates atrial natriuretic peptide secretion via the endothelin-1/ET(B) receptor-mediated pathway in beating rabbit atria. *Life Sci* (2012) **90**(19–20):793–8. doi:10.1016/j.lfs.2012.04.008
113. Arnaud-Batista FJ, Costa GT, de Oliveira IMB, Costa PPC, Santos CF, Fontelles MC, et al. Natriuretic effect of bufalin in isolated rat kidneys involves activation of the Na<sup>+</sup>, K<sup>+</sup>-ATPase-Src kinase pathway. *Am J Physiol Renal Physiol* (2012) **302**(8):F959–66. doi:10.1152/ajpregnol.00130.2011
114. Pullen MA, Brooks DP, Edwards RM. Characterization of the neutralizing activity of digoxin-specific Fab toward ouabain-like steroids. *J Pharmacol Exp Ther* (2004) **310**(1):319–25. doi:10.1124/jpet.104.065250
115. Huang BS, Leenen FH. Sympathoexcitatory and pressor responses to increased brain sodium and ouabain are mediated via brain Ang II. *Am J Physiol* (1996) **270**(1 Pt 2):H275–80.
116. Hopoate-Sitake ML, Adair CD, Mason LA, Torres C, Kipikasa J, Graves SW. Digibind reverses inhibition of cellular rb+ uptake caused by endogenous sodium pump inhibitors present in serum and placenta of women with preeclampsia. *Reprod Sci* (2011) **18**(2):190–9. doi:10.1177/1933719110385133
117. Ishkaraeva-Yakovleva VV, Fedorova OV, Solodovnikova NG, Frolova EV, Bzhelyansky AM, Emelyanov IV, et al. DigiFab interacts with endogenous cardiotonic steroids and reverses preeclampsia-induced Na<sup>+</sup>, K<sup>+</sup>-ATPase inhibition. *Reprod Sci* (2012) **19**(12):1260–7. doi:10.1177/1933719112447124
118. Pullen MA, Harpel MR, Danoff TM, Brooks DP. Comparison of non-digitalis binding properties of digoxin-specific Fabs using direct binding methods. *J Immunol Methods* (2008) **336**(2):235–41. doi:10.1016/j.jim.2008.05.005
119. Gable ME, Abdallah SL, Najjar SM, Liu L, Askari A. Digitalis-induced cell signaling by the sodium pump: on the relation of Src to Na<sup>+</sup>, K<sup>+</sup>-ATPase. *Biochem Biophys Res Commun* (2014) **446**(4):1151–4. doi:10.1016/j.bbrc.2014.03.071
120. Lorenz JN, Loreaux EL, Dostanic-Larson I, Lasko V, Schnetzer JR, Paul RJ, et al. ACTH-induced hypertension is dependent on the ouabain-binding site of the {alpha}2-Na,K ATPase subunit. *Am J Physiol Heart Circ Physiol* (2008) **295**(1):H273–80. doi:10.1152/ajpheart.00183.2008

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Received: 09 October 2014; accepted: 24 March 2015; published online: 13 April 2015.*

*Citation: Buckalew VM (2015) Endogenous digitalis-like factors: an overview of the history. *Front. Endocrinol.* **6**:49. doi: 10.3389/fendo.2015.00049*

*This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.*

*Copyright © 2015 Buckalew. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*



# The trade-off between dietary salt and cardiovascular disease; a role for Na/K-ATPase signaling?

**Joe X. Xie<sup>1</sup>, Anna Pearl Shapiro<sup>2</sup> and Joseph Isaac Shapiro<sup>3\*</sup>**

<sup>1</sup> Department of Medicine, University of Colorado School of Medicine, Aurora, CO, USA

<sup>2</sup> Department of Medicine, University of Toledo College of Medicine, Toledo, OH, USA

<sup>3</sup> Department of Medicine, Joan C. Edwards School of Medicine, Marshall University, Huntington, WV, USA

**Edited by:**

Harvey Craig Gonick, University of California Berkeley, USA

**Reviewed by:**

Gert Jansen, Erasmus MC, Netherlands

Harvey Craig Gonick, University of California Berkeley, USA

Neal S. Bricker, University of California Los Angeles School of Medicine, USA

**\*Correspondence:**

Joseph Isaac Shapiro, Department of Medicine, Joan C. Edwards School of Medicine, Marshall University, 1600 Medical Center Drive Suite 3408, Huntington, WV 25701, USA  
e-mail: shapiroj@marshall.edu

It has been postulated for some time that endogenous digitalis-like substances, also called cardiotonic steroids (CTS), exist, and that these substances are involved in sodium handling. Within the past 20 years, these substances have been unequivocally identified and measurements of circulating and tissue concentrations have been made. More recently, it has been identified that CTS also mediate signal transduction through the Na/K-ATPase, and consequently been implicated in profibrotic pathways. This review will discuss the mechanism of CTS in renal sodium handling and a potential “trade-off” effect from their role in inducing tissue fibrosis.

**Keywords:** cardiotonic steroids, digitalis-like factors, fibrosis, sodium pump, signaling, renal failure, hypertension

## INTRODUCTION

Increased dietary sodium chloride (NaCl) intake has been implicated in cardiovascular and renal diseases for some time (1), and this implication has recently become fairly solid (2). This relationship between dietary sodium intake and cardiovascular disease is demonstrated in several large scale studies, such as the international study of salt and blood pressure (INTERSTALT) (3) and the dietary approaches to stop hypertension (DASH) (4). With this relationship so demonstrated, understanding the specific mechanisms underlying the deleterious effects of NaCl becomes timely and relevant to clinical management.

This review will focus on one of the factors linking dietary NaCl to cardiovascular and renal disease. We will specifically discuss the role of digitalis-like factors, also known as endogenous cardiotonic steroids (CTS), which function as innate inhibitors of the Na/K-ATPase (5). Although the existence of these endogenous factors has been controversial (6–8), this is no longer the case. Some of these recent breakthroughs include the chemical identification of specific CTS in experimental animals and humans (9, 10), establishment of normal and pathological concentrations for

these substances as well as defining possible roles for CTS in animal models of and human disease states (11–13). We would also stress that the discovery of the cell signaling functions of the Na/K-ATPase and its role in molecular cellular biology (14–16) has also been quite relevant to this field. Here, we will emphasize the role of trade-off with respect to CTS signaling and Na homeostasis.

## RENAL SALT REABSORPTION AND THE EVIDENCE FOR “THIRD FACTOR”

The microscopic architecture of the kidney involves the attachment of vascular filtering units called glomeruli with tubules that modulate the quantity, electrolytes, and acid-base content of tubular fluid, which ultimately becomes urine. Simplistically, the tubules can be roughly broken down into proximal, where 60–80% of all Na and water reabsorption occur and distal, the nephron segments responsible for the fine tuning of what is excreted as urine.

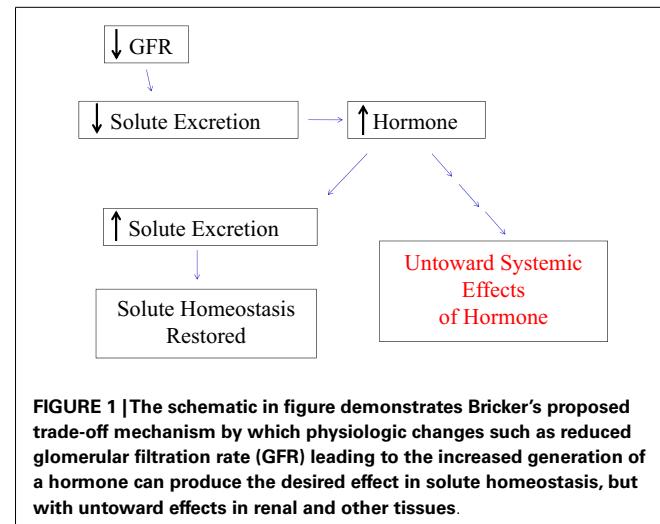
Clearly the renin–angiotensin–aldosterone system, vasopressin and the sympathetic nervous system are critically important in mammalian volume regulation as well as to the maintenance of blood pressure in the face of a hypovolemic insult (17). However, it is very clear that perturbations in these systems cannot explain natriuretic responses to acute or chronic expansion of blood volume (18). This point was first demonstrated in 1961 in a classic paper by de Wardener and colleagues (19). This study showed that natriuresis induced by saline infusion occurred even if renal perfusion pressure and glomerular filtration rate (GFR, factor 1) and aldosterone concentrations (factor 2) were prevented from changing. This so called “third factor,” which we now understand is (are) CTS, was a “hot” topic in the 1960s and 1970s, and was even incorporated into Guyton’s model for circulatory

**Abbreviations:** Ca, calcium; Cl, chloride; CTS, cardiotonic steroid; EGFR, epidermal growth factor receptor; EMT, epithelial-mesenchymal transformation; ERK, extracellular signal-regulated kinase; FAK, focal adhesion kinase; Fli-1, friend leukemia integration 1 transcription factor; GFR, glomerular filtration rate; Grb2, growth factor receptor-bound protein-2; MBG, marinobufagenin; Na, sodium; Na/K-ATPase, sodium potassium ATPase; NAC, N-acetyl cysteine; NHE3, sodium-hydrogen exchanger 3; PI(3)K, phosphoinositide 3-kinase; PKC, protein kinase C; PLC, phospholipase C; ROS, reactive oxygen species; SERCA, sarcoplasmic endoplasmic reticulum calcium ATPase; Shc, Src homology-2 domain containing protein; SOS, Son of Sevenless protein; TCB, telecinobufagin; TGF, transforming growth factor.

homeostasis (20). Cort and Lichardus observed that a circulating substance in animals subjected to carotid artery occlusion induced natriuresis in different mammals and inhibited sodium transport in frog skin (21). Buckalew showed that an ultrafiltrate of volume-expanded dogs inhibited sodium transport in toad bladders. They went on to propose that the active substance was an inhibitor of the Na/K-ATPase (22). Gonick and coworkers showed that volume expansion in rats, in fact, produced a chemical which did inhibit the ATPase activity of rat kidneys (11). In 1980, Gruber and Buckalew noted that elevated levels of circulating digoxin-like material was seen in volume-expanded dogs (23). Other important contributions were made in the laboratory of Schrier and de Wardener over the next decade (24–26). However, doubt as to the validity of Na/K-ATPase inhibitors developed during the 1980s and 1990s because of inconsistencies in the reported results. In particular, prevailing CTS assays were based on cross-reactivity of CTS with antibodies to digoxin. This cross-reactivity of the commercially employed anti-digoxin antibodies to CTS varied considerably (27–32). Probably, the most important inconsistency was that digitalis did not appear to be natriuretic in normal subjects (33). On this background, atrial (and brain) natriuretic peptide(s) were discovered, were obviously natriuretic, and their concentrations (which could be easily measured) were increased in volume-expanded states (34–38). Undoubtedly, these points deflected interest from the study of CTS. However, enthusiasm was renewed in the recent past for the following reasons. First, several CTS have been isolated from experimental animals and humans and chemically characterized. Specifically, marinobufagenin (MBG) as well as telecinobufagin (TCB) have been isolated from plasma and urine (9). Ouabain has also been identified although there is still some debate as to whether this is ouabain or something distinct, which also reacts to anti-ouabain antibodies (10, 39). The concentrations of ouabain (or ouabain like compound) and MBG appear to be in the range of 200–2700/min in humans, depending on whether disease is present (5, 40, 41). Plasma levels of TCB and bufalin are less well defined at present. Also, quite importantly, a signal cascade has been identified, which does not appear to involve enzymatic inhibition of the Na/K-ATPase. This signaling pathway involves CTS binding of the caveolar Na/K-ATPase in the company of Src and the EGFR and the elaboration of a signal cascade, which involves the generation of reactive oxygen species (ROS) (14, 16). Both of these concepts have been extensively reviewed (42–44).

### “TRADE-OFF” CONCEPT, A HISTORICAL PERSPECTIVE

The concept of “trade-off” plays an extremely powerful role in physiology. This is perhaps best described by Neal Bricker who postulated that in renal disease, the hormonal forces driving nephrons to maintain fluid and electrolyte homeostasis would be complicated by the untoward consequences of these elevated hormones mediating other effects, essentially creating the signs, symptoms, and pathophysiologic changes associated with the uremic syndrome (45, 46). As sodium (Na) handling is so critical to volume balance, electrolyte homeostasis, and acid-base status, it is not surprising that Bricker formulated this hypothesis to involve the Na/K-ATPase.

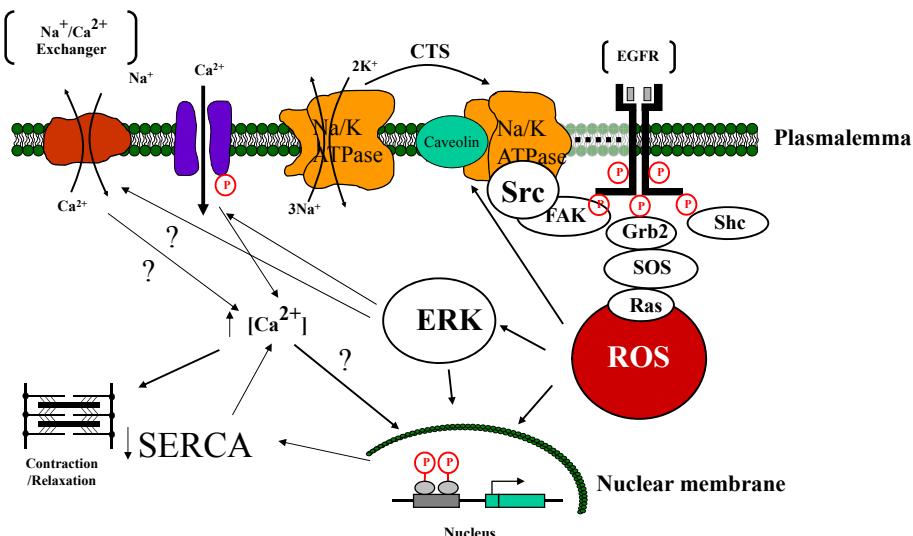


**FIGURE 1 |**The schematic in figure demonstrates Bricker's proposed trade-off mechanism by which physiologic changes such as reduced glomerular filtration rate (GFR) leading to the increased generation of a hormone can produce the desired effect in solute homeostasis, but with untoward effects in renal and other tissues.

Bricker speculated that an inhibitor of the Na/K-ATPase would circulate in increased concentration as a response to decreased GFR in order to maintain Na homeostasis (45). This inhibition would subsequently lead to decreased renal Na reabsorption, hence the maintenance of Na homeostasis (Figure 1). Unintended effects of higher concentrations of this Na/K-ATPase inhibitor would be responsible for some of the symptoms, signs, and abnormal laboratory results seen with chronic renal failure as well as potentially contribute to the progressive nature of chronic kidney disease (45, 47–50). As we will detail in this review, a potential consequence of increases in natriuretic hormone levels, specifically elevated CTS levels may be the profibrotic effects of these molecules (51). Before we address this, however, it may be useful to briefly discuss the evolution of our understanding of the Na/K-ATPase (45, 46), which had been described and characterized several decades before (52).

### DISCOVERY OF THE Na/K-ATPase, ITS ROLE IN SIGNALING CASCADES VS. ION TRANSPORTATION

The Na/K-ATPase was discovered by Skou in 1957 (53). This protein was demonstrated to be responsible for the electrogenic exchange of sodium and potassium (54). The Na/K-ATPase, also called the sodium pump, is present in all living cells (55). Although there has been some evolutionary modification of the sodium pump, in all multicellular animal cells, the sodium pump consists of (at least) a dimer of an alpha and beta subunit and is considered a member of P-type ATPases (43). Different isoforms of the alpha and beta subunits have been identified and are believed to have functional differences, a topic which has been extensively reviewed (56). Genes encoding the alpha-1 and alpha-2 isoforms reside on the chromosome 1 whereas alpha-3 appears to be coded for on chromosome 19 and alpha-4 (present only in sperm) is mapped to chromosome 13 in humans (57). The act of pumping sodium and potassium is accompanied by changes in conformation and phosphorylation state (43). It also requires energy provided by the hydrolysis of ATP as was initially identified also by Skou (58). The work of Skou was ultimately matured into the currently accepted Post-Albers model



**FIGURE 2 | A schematic illustrating the involvement of cardiotonic steroid (CTS) – induced Na/K-ATPase signal cascade initiated by the Na/K-ATPase mediated activation of Src tyrosine kinase and subsequent downstream targets eventually leading to the development of reactive oxygen species (ROS). Specifically, we postulate that in the microdomain of caveolae, the Na/K-ATPase functions as a scaffolding protein, interacting with CTS and changing conformation so as to active Src. Src then trans-activates the EGFR which leads to a signal cascade involving FAK, Shc, Grb2, and SOS resulting in the generation of ROS which in turn activates additional Na/K-ATPase molecules as well as**

causes downstream activation of ERK as well as effects on the nuclear transcription (43). ERK activation has effects on both L-type channels and possibly the Na/Ca exchanger with net effect to increase cytosolic Ca in some tissues (15). Nuclear effects in myocardial tissue include downregulation of SERCA transcription and translation (70). Abbreviations: EGFR, epidermal growth factor receptor; FAK, focal adhesion kinase; Shc, Src homology-2 domain containing protein; Grb2, growth factor receptor-bound protein-2; SOS, son of sevenless protein; ERK, extracellular-signal-regulated kinase; SERCA, sarcoplasmic/endoplasmic reticulum calcium ATPase.

for Na/K-ATPase pumping function (43). The alpha 1 subunit of the Na/K-ATPase has 11 transmembrane domains as well as several well defined cytosolic regions referred to as the N, P, catalytic, and A domains (43). Interestingly, the development and maintenance on an evolutionary scale of caveolin and Src binding motifs, which are scattered throughout these cytosolic domains appeared to occur between single celled animal structures and slime mold (59).

In the late 1990s, the laboratory of Dr. Zijian Xie added a significant wrinkle to this understanding. While it is certainly possible that some signaling does occur through the chemical inhibition of the plasmalemmal Na/K-ATPase, it does appear that other mechanisms must be proposed to explain the signaling. In fact, it appears that the specific Na/K-ATPase molecules responsible for the greatest amount of signaling in response to the binding of CTS are actually not involved in pumping sodium or potassium (60). In the late 1990s, Dr. Xie and colleagues observed that in neonatal cardiac myocytes, ouabain caused increases in ROS measured with CMDCF (14). It was further noted that some of the downstream effects of ouabain were blocked by *N*-acetyl cysteine (NAC) or vitamin E. These increases in ROS could be demonstrated even when cytosolic calcium was maintained low by removal of extracellular calcium (16). It was further noted that Ras activation appeared to be necessary to see increases in ROS (16). Other studies determined that interactions between the Na/K-ATPase and Src appeared to initiate the signal cascade. The alpha 1 subunit of the Na/K-ATPase binds Src and appears to maintain it in an inactive state. However,

binding a CTS appears to alter the Na/K-ATPase structure allowing Src to become activated which, in turn, trans-activates the EGFR, and begins the signal cascade which causes increases in ROS (61–64). The Na/K-ATPase–Src complex appears to function similar to a receptor tyrosine kinase. Downstream activation of PLC, PI(3)K, and PKC has also been established (15, 65–68) (Figure 2). The role of ROS in pump signaling has been extensively reviewed elsewhere (14, 16, 51, 69).

Although inhibition of the Na/K-ATPase is certainly one possible mechanism by which digitalis and related molecules might “signal,” it is important to emphasize that even transporting epithelia typically have a redundancy of Na/K-ATPase pumping units given that cytosolic Na levels live within a range ideally suited to regulate Na/K-ATPase activity. While it is possible that certain compartments of the cell see higher local concentrations of Na with modest inhibition of Na/K-ATPase pump activity, we emphasize that physiological and even pharmacological concentrations of digitalis do not demonstrably increase cytosolic Na concentrations in physiologically relevant preparations (42). We would further point out that most studies, including those from our lab, which demonstrate inhibition of the Na/K-ATPase by circulating substances do so with strategies to control for the cytosolic Na concentration (71–74).

Approximately one decade ago, a further analogy of Na/K-ATPase signaling to the signaling of receptor tyrosine kinases was established with the observation that CTS binding to the Na/K-ATPase in renal tissues triggers endocytosis of

the CTS-Na/K-ATPase complex (75). Subsequent studies have demonstrated that this internalization is associated with endosomal accumulation of the Na/K-ATPase and its caveolar signaling partners, and that the process requires both caveolin (and caveolar structure) and clathrin (76, 77). We have gone on to demonstrate that this process appears to also regulate the expression of the apical sodium transporter, NHE3, as well as impact renal salt excretion *in vivo* (78–80). Recent data from the laboratory of Dr. Lingrel utilizing novel genetic manipulations of the different alpha 1 isoforms in mice indicate that it is the alpha 1 subunit, which can be considered the functional receptor for these CTS. Interestingly, the amount of Na/K-ATPase alpha 1 subunit as well as its affinity for CTS appear to both positively correlate with the magnitude of the signaling effect (81–84).

Recently, we have made several observations that bring the consideration of ROS in the context of Na pump signaling in a new light. First, we found that the Dahl salt-resistant (R) strain of rats had a natriuretic response to a high salt diet, which did not require substantial increases in blood pressure (hence the term “salt resistant”) and was accompanied by activation of Src and ERK as well as redistribution in the renal proximal tubule cells of the basolateral Na/K-ATPase and apical NHE3. This was previously observed with the wild type Sprague Dawley animals (which were used as a founder population to generate Dahl R and salt sensitive, S, rats). In contrast, the Dahl S rats did not have this redistribution. Isolated proximal tubules from young Dahl R and S rats maintained on a low salt diet demonstrated ouabain sensitivity and insensitivity, respectively, in terms of Src and ERK activation as well as redistribution of the NaK-ATPase and NHE3 (85). Moving back to LLC-PK1 cells, we noted that the signaling observed with ouabain or other CTS could be duplicated by exposure to an ROS generation system (Glucose Oxidase + Glucose), blocked by anti-oxidants (e.g., *N*-acetyl cysteine) and was accompanied by specific carbonylation of two amino acids in the A domain portion of the alpha 1 subunit (86). Given that the proximal tubules of Dahl S rats demonstrate considerable carbonylation of plasma proteins including the Na/K-ATPase prior to exposure to high salt *in vivo* or ouabain *in vitro* (unpublished data), this suggests that chronic oxidation of the Na/K-ATPase may lead to impaired signal transduction in the proximal tubule and a form of oxidant “fatigue.” Perhaps of even greater importance, the protein oxidation seen with both ouabain and glucose oxidase/glucose was found to be reversible in a biochemical rather than a physiological sense since removing ouabain or glucose oxidase/glucose led to the return to non-carbonylated proteins regardless of whether new protein synthesis or protein degradation were inhibited. In addition, signaling through the Na/K-ATPase appeared to impact the amount and degree of protein carbonylation induced by glucose oxidase/glucose suggesting a role for the Na/K-ATPase as both a receptor and amplifier of ROS (86). We had seen *in vivo* data supporting this concept in earlier studies discussed below. Although a feed-forward system (which this appears to be) suggests ongoing amplification, it seems clear that endocytosis of this molecular machinery would be an effective termination mechanism (87). Whether the oxidatively modified Na/K-ATPase is a trigger for endocytosis is a topic we are actively investigating at present.

On this background, it is useful to consider whether a CTS is effectively natriuretic *in vivo*. This discussion began many years ago regarding the CTS pharmacological agent, digoxin, or digitalis, which was noted to effect natriuresis in patients with congestive heart failure but not normal subjects (88). Currently, there remains debate as to whether a CTS such as ouabain is, in fact, natriuretic (89). Although clearly this is important in understanding the physiological relevance of the molecular mechanisms described above, we would caution the reader that the answer to this question may be different depending on the physiological state of the experimental animal or subject at the time of the study (80, 85, 90). That said, we would certainly concede that a correlation between renal Na/K-ATPase signaling or inhibition and natriuresis may not always be present.

## ROLE IN CARDIAC AND RENAL FIBROSIS WITH EXPERIMENTAL RENAL FAILURE

Concern that CTS signaling through the Na/K-ATPase might be profibrotic grew from several studies. First, we observed that experimental renal failure produced cardiac fibrosis in both rat and mouse (91). We would stress that human uremic cardiomyopathy is believed to also be complicated by fibrosis. When we performed active immunization prior to induction of experimental renal failure, the cardiac fibrosis was markedly attenuated. In a separate group of animals, infusion of MBG designed to achieve similar plasma levels of MBG as seen with experimental renal failure also caused cardiac fibrosis. Evidence for Na/K-ATPase signaling (e.g., Src and ERK activation) was seen in both animals subjected to experimental renal failure or MBG infusion whereas active immunization against the MBG-Albumin conjugate attenuated this in the experimental renal failure group (51, 70, 91, 92). In addition, blockade of Na/K-ATPase signaling with active (or passive) immunization as well as pharmacologic blockade (see below) dramatically attenuated the oxidant stress in tissues seen with experimental renal failure (51, 91, 93, 94). Based on these animal studies, we next examined how CTS affected fibroblasts grown in culture. We noted that CTS (e.g., MBG, ouabain) induced increases in fibroblast collagen production as evidenced by either increased labeled proline incorporation or procollagen expression determined with Western blot. Evidence for Na/K-ATPase signaling (e.g., Src or ERK activation) could be observed as well. Moreover, ROS scavenging or pharmacological or molecular biological Src inhibition prevented increases in proline incorporation and collagen production seen with CTS. An increase in transcription was identified as we saw substantial increases in both mRNA for collagen as well as luciferase in cells transfected with a reporter construct following exposure to CTS. However, we did not see evidence for increased TGF beta signaling in these cells although pharmacological antagonism of the TGF beta system did block CTS stimulated collagen production (51). We next examined how CTS affected Fli-1 expression, stimulated by work performed by Watson and colleagues. Fli-1 is a negative regulator of collagen synthesis (95), and we noted that CTS induce decreases in Fli-1 expression in several types of fibroblasts (cardiac, renal, and dermal). We also observed that decreases in Fli-1 appear to be necessary for MBG to induce increases in collagen. Additional work showed that CTS induce translocation of PKCdelta from the cytosol to the nucleus in a PLC

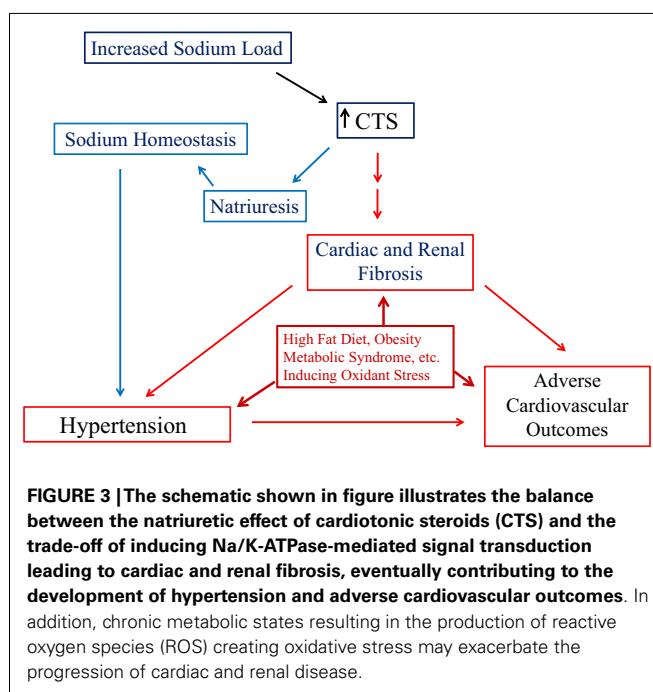
dependent manner. It appears that the translocation of PKC $\delta$  causes Fli-1 phosphorylation and subsequent degradation (94).

These studies next led to work examining the effects of mineralocorticoid antagonists. We should first say that Finotti and colleagues reported 30 years ago that spironolactone and canrenone were antagonists of ouabain binding to the Na/K-ATPase (96). We looked at whether this observation was applicable to our system. *In vitro*, we saw that both spironolactone and canrenone could attenuate MBG-induced increases in collagen production in cardiac fibroblasts. Interestingly, we could not see a substantial effect of aldosterone on cardiac collagen production. Our *in vitro* observations were extended to *in vivo* studies where we saw that administration of spironolactone to rats with experimental renal failure markedly attenuated the observed cardiac fibrosis (94). This suggests the Na/K-ATPase signaling cascade may be a useful target for therapeutic drug development.

Further studies have demonstrated that the effects of MBG (and other CTS) are not specific for cardiac fibroblasts. We have noted that renal fibroblasts have a very similar response as cardiac fibroblasts, suggesting a potential pathological role for MBG in producing renal fibrosis and progressive renal failure. Using MBG infusion in the rat, we saw that such infusion was associated with the induction of Snail, a transcription factor known to be involved in epithelial–mesenchymal transformation (EMT). In LLC-PK1 cells grown in culture, MBG induces EMT in a dose and time dependent way (97).

## TRADE-OFF WITH RESPECT TO CTS

With the aforementioned data, we would suggest that the CTS signal cascade through the Na/K-ATPase fits the concept of "trade-off." Specifically, CTS concentrations increase in response to volume expansion and/or salt loading. These CTS mediate increases in urinary Na excretion, maintaining Na homeostasis, but the



endocytosis machinery may fatigue with ongoing stimulation. Moreover, there are other consequences of the elevated CTS concentrations, namely vasoconstriction and hypertension along with fibrosis, which was described above (**Figure 3**). The fibrosis may lead to further renal insensitivity in terms of natriuresis, and the combination of events cascading to produce progressive cardiovascular disease.

FUTURE DIRECTIONS

As we better understand the role of CTS signaling through the Na/K-ATPase, several therapeutic targets come to mind, which may provide novel and effective therapy for different chronic diseases. First, there is the interaction of the CTS with the Na/K-ATPase. This has been addressed experimentally in our laboratory with both active and passive immunization (51, 91, 93, 98) as well as pharmacologically with several different approaches (94, 99). Other groups have developed different substances which can loosely describe as "ouabain antagonists" which we have recently reviewed (5). Rostafuroxin has been very well characterized and appears to have potential for the treatment of hypertension (100, 101). Recently, our laboratory has begun to develop strategies to alter the interaction between the Na/K-ATPase alpha 1 subunit and Src (102). However, it is clear that the aforementioned signaling cascade affords a number of possible sites for intervention including but not limited to the generation of ROS (69), activation of Src and activation of ERK. Unfortunately, these molecular targets will also fit under the general rubric of "trade-off." Although some aspects of CTS and signaling through the Na/K-ATPase may be maladaptive as we have discussed in this review, it is almost certain that inhibition of this CTS-Na/K-ATPase pathway may have deleterious effects which need to be navigated.

## REFERENCES

1. Luft FC, Weinberger MH. Sodium intake and essential hypertension. *Hypertension* (1982) **4**:III14–9. doi:10.1161/01.HYP.4.5\_Pt\_2.III14
  2. Ritz E. Salt and hypertension. *Nephrology* (2010) **15**(Suppl 2):49–52. doi:10.1111/j.1440-1797.2010.01311.x
  3. Elliott P. The INTERSALT study: an addition to the evidence on salt and blood pressure, and some implications. *J Hum Hypertens* (1989) **3**:289–98.
  4. Appel LJ, Moore TJ, Obarzanek E, Vollmer WM, Svetkey LP, Sacks FM, et al. A clinical trial of the effects of dietary patterns on blood pressure. DASH Collaborative Research Group. *N Engl J Med* (1997) **336**:1117–24. doi:10.1056/NEJM199704173361601
  5. Bagrov AY, Shapiro JI, Fedorova OV. Endogenous cardiotonic steroids: physiology, pharmacology, and novel therapeutic targets. *Pharmacol Rev* (2009) **61**:9–38. doi:10.1124/pr.108.000711
  6. Kelly RA. Excretion of artifactual endogenous digitalis-like factors. *Am J Physiol* (1986) **251**:H205–9.
  7. Kelly RA, O'Hara DS, Canessa ML, Mitch WE, Smith TW. Characterization of digitalis-like factors in human plasma. Interactions with NaK-ATPase and cross-reactivity with cardiac glycoside-specific antibodies. *J Biol Chem* (1985) **260**:11396–405.
  8. Hansen O. Do putative endogenous digitalis-like factors have a physiological role? *Hypertension* (1994) **24**:640–4. doi:10.1161/01.HYP.24.5.640
  9. Komiyama Y, Dong XH, Nishimura N, Masaki H, Yoshika M, Masuda M, et al. A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin Biochem* (2005) **38**:36–45. doi:10.1016/j.clinbiochem.2004.08.005
  10. Hamlyn JM, Blaustein MP, Bova S, DuCharme DW, Harris DW, Mandel F, et al. Identification and characterization of a ouabain-like compound from human plasma. *Proc Natl Acad Sci U S A* (1991) **88**:6259–63. doi:10.1073/pnas.88.21.9907.d

11. Gonick HC, Kramer HJ, Paul W, Lu E. Circulating inhibitor of sodium-potassium-activated adenosine triphosphatase after expansion of extracellular fluid volume in rats. *Clin Sci Mol Med* (1977) **53**:329–34.
12. Schoner W, Scheiner-Bobis G. Endogenous and exogenous cardiac glycosides and their mechanisms of action. *Am J Cardiovasc Drugs* (2007) **7**:173–89. doi:10.2165/00129784-200707030-00004
13. Kolmakova EV, Haller ST, Kennedy DJ, Isachkina AN, Budny GV, Frolova EV, et al. Endogenous cardiotonic steroids in chronic renal failure. *Nephrol Dial Transplant* (2011) **26**:2912–9. doi:10.1093/ndt/gfq772
14. Xie Z, Kometiani P, Liu J, Li J, Shapiro JI, Askari A. Intracellular reactive oxygen species mediate the linkage of Na+/K+-ATPase to hypertrophy and its marker genes in cardiac myocytes. *J Biol Chem* (1999) **274**:19323–8. doi:10.1074/jbc.274.27.19323
15. Tian J, Liu J, Garlid KD, Shapiro JI, Xie Z. Involvement of mitogen-activated protein kinases and reactive oxygen species in the inotropic action of ouabain on cardiac myocytes. A potential role for mitochondrial K(ATP) channels. *Mol Cell Biochem* (2003) **242**:181–7. doi:10.1023/A:1021114501561
16. Liu J, Tian J, Haas M, Shapiro JI, Askari A, Xie Z. Ouabain interaction with cardiac Na+/K+-ATPase initiates signal cascades independent of changes in intracellular Na+ and Ca2+ concentrations. *J Biol Chem* (2000) **275**:27838–44. doi:10.1074/jbc.M002950200
17. Zhou HZ, Shapiro JI, Chan L, Schrier RW. Atrial natriuretic peptide protects against cold ischemic injury in the isolated and in situ rat kidney. *J Am Soc Nephrol* (1990) **1**:927–8.
18. Abraham WT, Schrier RW. Body fluid volume regulation in health and disease. *Adv Intern Med* (1994) **39**:23–47.
19. de Wardener H, Mills IH, Clapham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intra-venous saline in the dog. *Clin Sci* (1961) **21**:249–58.
20. Guyton AC, Coleman TG, Granger HJ. Circulation: overall regulation. *Annu Rev Physiol* (1972) **34**:13–46. doi:10.1146/annurev.ph.34.030172.000305
21. Cort JH, Rudinger J, Lichardus B, Hagemann I. Effects of oxytocin antagonists on the saluresis accompanying carotid occlusion. *Am J Physiol* (1966) **210**:162–8.
22. Buckalew VM Jr, Martinez FJ, Green WE. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**:926–35. doi:10.1172/JCI106312
23. Gruber KA, Whitaker JM, Buckalew VM Jr. Endogenous digitalis-like substance in plasma of volume-expanded dogs. *Nature* (1980) **287**:743–5. doi:10.1038/287743a0
24. Schrier RW, McDonald KM, Marshall RA, Laufer DP. Absence of natriuretic response to acute hypotonic intravascular volume expansion in dogs. *Clin Sci* (1968) **34**:57–72.
25. De Wardener HE, Fabian M, Jones JJ, Lee J, Schrier RW, Verroust PJ. The effect of acute extracellular fluid volume expansion and acute haemorrhage on plasma antidiuretic hormone and oxytocin levels in dogs. *J Physiol* (1968) **196**:122.
26. de Wardener HE, Clarkson EM, Nutbourne DM, Schrier RW, Talner LB, Venstrom MG, et al. Evidence for a hormone other than aldosterone which controls urinary sodium excretion. *Adv Nephrol Necker Hosp* (1971) **1**:97–111.
27. Bergdahl B, Dahlstrom G, Molin L, Bertler A. Inter and intra laboratory variation of digoxin radioimmunoassay in Sweden. *Acta Pharmacol Toxicol (Copenh)* (1979) **45**:66–72. doi:10.1111/j.1600-0773.1979.tb02362.x
28. Bergdahl B, Molin L. Precision of digoxin radioimmunoassays and matrix effects: four kits compared. *Clin Biochem* (1981) **14**:67–71. doi:10.1016/S0009-9120(81)90704-9
29. Gusdon JP Jr, Buckalew VM Jr, Hennessy JF. A digoxin-like immunoreactive substance in preeclampsia. *Am J Obstet Gynecol* (1984) **150**:83–5. doi:10.1016/S0002-9378(84)80114-3
30. Pleasants RA, Gadsden RH Sr, McCormack JP, Piveral K, Sawyer WT. Interference of digoxin-like immunoreactive substances with three digoxin immunoassays in patients with various degrees of renal function. *Clin Pharm* (1986) **5**:810–6.
31. Miyashita H, Sato T, Tamura T, Tamura O, Tazawa H. The problems of digitalis therapy from the viewpoint of serum concentration with special reference to the sampling time, to the overlapping range of serum concentration where intoxicated and non-intoxicated patients are located and to atrial fibrillation. *Jpn Circ J* (1986) **50**:628–35. doi:10.1253/jcj.50.628
32. Ebara H, Suzuki S, Nagashima K, Koizumi T, Nishida A, Kanbe Y, et al. Digoxin- and digitoxin-like immunoreactive substances in amniotic fluid, cord blood, and serum of neonates. *Pediatr Res* (1986) **20**:28–31. doi:10.1203/00006450-198601000-00007
33. Hauptman PJ, Kelly RA. Digitalis. *Circulation* (1999) **99**:1265–70. doi:10.1161/01.CIR.99.9.1265
34. de Bold AJ, Borenstein HB, Veress AT, Sonnenberg H. A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats. *Life Sci* (1981) **28**:89–94. doi:10.1016/0024-3205(81)90370-2
35. de Bold AJ, Flynn TG. Cardionatrikin I – a novel heart peptide with potent diuretic and natriuretic properties. *Life Sci* (1983) **33**:297–302. doi:10.1016/0024-3205(83)90390-9
36. Pamani MB, Clough DL, Chen JS, Link WT, Haddy FJ. Effects of rat atrial extract on sodium transport and blood pressure in the rat. *Proc Soc Exp Biol Med* (1984) **176**:123–31. doi:10.3181/00379727-176-41851
37. Nakamoto M, Shapiro JI, Shanley PF, Chan L, Schrier RW. In vitro and in vivo protective effect of atriopeptin III on ischemic acute renal failure. *J Clin Invest* (1987) **80**:698–705. doi:10.1172/JCI113124
38. Buckalew VM, Morris M, Hamilton RW. Atrial natriuretic factor. *Adv Intern Med* (1987) **32**:1–25.
39. Baecher S, Kroiss M, Fassnacht M, Vogeser M. No endogenous ouabain is detectable in human plasma by ultra-sensitive UPLC-MS/MS. *Clin Chim Acta* (2014) **431**:87–92. doi:10.1016/j.cca.2014.01.038
40. Bagrov AY, Shapiro JI. Endogenous digitalis: pathophysiologic roles and therapeutic applications. *Nat Clin Pract Nephrol* (2008) **4**:378–92. doi:10.1038/ncpneph0848
41. Fedorova OV, Shapiro JI, Bagrov AY. Endogenous cardiotonic steroids and salt-sensitive hypertension. *Biochim Biophys Acta* (2010) **1802**:1230–6. doi:10.1016/j.bbadi.2010.03.011
42. Xie Z, Askari A. Na(+)/K(+)-ATPase as a signal transducer. *Eur J Biochem* (2002) **269**:2434–9. doi:10.1046/j.1432-1033.2002.02910.x
43. Pierre SV, Xie Z. The Na, K-ATPase receptor complex: its organization and membership. *Cell Biochem Biophys* (2006) **46**:303–16. doi:10.1385/CBB:46:3:303
44. Tian J, Xie ZJ. The Na-K-ATPase and calcium-signaling microdomains. *Physiology* (2008) **23**:205–11. doi:10.1152/physiol.00008.2008
45. Bricker NS. On the pathogenesis of the uremic state. An exposition of the “trade-off hypothesis”. *N Engl J Med* (1972) **286**:1093–9. doi:10.1056/NEJM197205182862009
46. Bricker NS, Fine LG. Uremia: formulations and expectations. The trade-off hypothesis: current status. *Kidney Int Suppl* (1978) **8**:S5–8.
47. Bricker NS, Schmidt RW, Favre H, Fine L, Bourgoignie JJ. On the biology of sodium excretion: the search for a natriuretic hormone. *Yale J Biol Med* (1975) **48**:293–303.
48. Bourgoignie JJ, Hwang KH, Ipakchi E, Bricker NS. The presence of a natriuretic factor in urine of patients with chronic uremia. The absence of the factor in nephrotic uremic patients. *J Clin Invest* (1974) **53**:1559–67. doi:10.1172/JCI107706
49. Licht A, Stein S, Bricker NS. Hormonal changes and transport adaptation in chronic renal failure: the possible role of a natriuretic hormone. *Biochem Soc Trans* (1978) **6**:837–9.
50. Licht A, Stein S, McGregor CW, Bourgoignie JJ, Bricker NS. Progress in isolation and purification of an inhibitor of sodium transport obtained from dog urine. *Kidney Int* (1982) **21**:339–44. doi:10.1038/ki.1982.27
51. Elkrehy J, Kennedy DJ, Yashawhi B, Vetteth S, Shidyak A, Kim EG, et al. Marinobufagenin stimulates fibroblast collagen production and causes fibrosis in experimental uremic cardiomyopathy. *Hypertension* (2007) **49**:215–24. doi:10.1161/01.HYP.0000252409.36927.05
52. Skou JC, Zerahn K. Investigations on the effect of some local anaesthetics and other amines on the active transport of sodium through the isolated short-circuited frog skin. *Biochim Biophys Acta* (1959) **35**:324–33. doi:10.1016/0006-3002(59)90381-6
53. Skou JC. The influence of some cations on an adenosine triphosphatase from peripheral nerves. *Biochim Biophys Acta* (1957) **23**:394–401. doi:10.1016/0006-3002(57)90343-8
54. Skou JC. Nobel Lecture. The identification of the sodium pump. *Biosci Rep* (1998) **18**:155–69. doi:10.1023/A:1020196612909

55. Skou JC, Esmann M. The Na<sub>+</sub>K-ATPase. *J Bioenerg Biomembr* (1992) **24**:249–61.
56. McDonough AA, Azuma KK, Lescalle-Matys L, Tang MJ, Nakhoul F, Hensley CB, et al. Physiologic rationale for multiple sodium pump isoforms. Differential regulation of alpha 1 vs alpha 2 by ionic stimuli. *Ann NY Acad Sci* (1992) **671**:156–68. doi:10.1111/j.1749-6632.1992.tb43793.x
57. Yang-Feng TL, Schneider JW, Lindgren V, Shull MM, Benz EJ Jr, Lingrel JB, et al. Chromosomal localization of human Na<sub>+</sub>,K-ATPase alpha- and beta-subunit genes. *Genomics* (1988) **2**:128–38. doi:10.1016/0888-7543(88)90094-8
58. Skou JC. The identification of the sodium pump. *Biosci Rep* (2004) **24**:436–51. doi:10.1007/s10540-005-2740-9
59. Xie Z, Xie J. The Na/K-ATPase-mediated signal transduction as a target for new drug development. *Front Biosci* (2005) **10**:3100–9. doi:10.2741/1766
60. Liang M, Tian J, Liu L, Pierre S, Liu J, Shapiro J, et al. Identification of a pool of non-pumping Na/K-ATPase. *J Biol Chem* (2007) **282**:10585–93. doi:10.1074/jbc.M609181200
61. Haas M, Askari A, Xie Z. Involvement of Src and epidermal growth factor receptor in the signal-transducing function of Na<sub>+</sub>/K-ATPase. *J Biol Chem* (2000) **275**:27832–7. doi:10.1074/jbc.M002951200
62. Haas M, Wang H, Tian J, Xie Z. Src-mediated inter-receptor cross-talk between the Na<sub>+</sub>/K-ATPase and the epidermal growth factor receptor relays the signal from ouabain to mitogen-activated protein kinases. *J Biol Chem* (2002) **277**:18694–702. doi:10.1074/jbc.M111357200
63. Wang H, Haas M, Liang M, Cai T, Tian J, Li S, et al. Ouabain assembles signaling cascades through the caveolar Na<sub>+</sub>/K-ATPase. *J Biol Chem* (2004) **279**:17250–9. doi:10.1074/jbc.M313239200
64. Tian J, Cai T, Yuan Z, Wang H, Liu L, Haas M, et al. Binding of Src to Na<sub>+</sub>/K-ATPase forms a functional signaling complex. *Mol Biol Cell* (2006) **17**:317–26. doi:10.1091/mbc.E05-08-0735
65. Elkareh J, Periyasamy SM, Shidyak A, Vetteth S, Schroeder J, Raju V, et al. Marinobufagenin induces increases in procollagen expression in a process involving protein kinase C and Fli-1: implications for uremic cardiomyopathy. *Am J Physiol Renal Physiol* (2009) **296**:F1219–26. doi:10.1152/ajprenal.90710.2008
66. Chen Y, Cai T, Yang C, Turner DA, Giovannucci DR, Xie Z. Regulation of inositol 1,4,5-trisphosphate receptor-mediated calcium release by the Na/K-ATPase in cultured renal epithelial cells. *J Biol Chem* (2008) **283**:1128–36. doi:10.1074/jbc.M708025200
67. Pierre SV, Yang C, Yuan Z, Seminero J, Mouas C, Garlid KD, et al. Ouabain triggers preconditioning through activation of the Na<sub>+</sub>, K-ATPase signaling cascade in rat hearts. *Cardiovasc Res* (2007) **73**:488–96. doi:10.1016/j.cardiores.2006.11.003
68. Yuan Z, Cai T, Tian J, Ivanov AV, Giovannucci DR, Xie Z. Na/K-ATPase tethers phospholipase C and IP<sub>3</sub> receptor into a calcium-regulatory complex. *Mol Biol Cell* (2005) **16**:4034–45. doi:10.1091/mbc.E05-04-0295
69. Priyadarshi S, Valentine B, Han C, Fedorova OV, Bagrov AY, Liu J, et al. Effect of green tea extract on cardiac hypertrophy following 5/6 nephrectomy in the rat. *Kidney Int* (2003) **63**:1785–90. doi:10.1046/j.1523-1755.2003.00914.x
70. Kennedy DJ, Malhotra D, Shapiro JI. Molecular insights into uremic cardiomyopathy: cardiotonic steroids and Na/K ATPase signaling. *Cell Mol Biol* (2006) **52**:3–14.
71. Xie ZJ, Wang YH, Ganjeizadeh M, McGee R Jr, Askari A. Determination of total (Na<sub>+</sub> + K<sub>+</sub>)-ATPase activity of isolated or cultured cells. *Anal Biochem* (1989) **183**:215–9. doi:10.1016/0003-2697(89)90470-3
72. Jack-Hays MG, Xie Z, Wang Y, Huang WH, Askari A. Activation of Na<sub>+</sub>/K(+)-ATPase by fatty acids, acylglycerols, and related amphiphiles: structure-activity relationship. *Biochim Biophys Acta* (1996) **1279**:43–8. doi:10.1016/0005-2736(95)00245-6
73. Periyasamy SM, Chen J, Cooney D, Carter P, Omran E, Tian J, et al. Effects of uremic serum on isolated cardiac myocyte calcium cycling and contractile function. *Kidney Int* (2001) **60**:2367–76. doi:10.1046/j.1523-1755.2001.00053.x
74. Xie Z. Ouabain interaction with cardiac Na/K-ATPase reveals that the enzyme can act as a pump and as a signal transducer. *Cell Mol Biol* (2001) **47**:383–90.
75. Liu J, Periyasamy SM, Gunning W, Fedorova OV, Bagrov AY, Malhotra D, et al. Effects of cardiac glycosides on sodium pump expression and function in LLC-PK1 and MDCK cells. *Kidney Int* (2002) **62**:2118–25. doi:10.1046/j.1523-1755.2002.00672.x
76. Liu J, Liang M, Liu L, Malhotra D, Xie Z, Shapiro JI. Ouabain-induced endocytosis of the plasmalemmal Na/K-ATPase in LLC-PK1 cells requires caveolin-1. *Kidney Int* (2005) **67**:1844–54. doi:10.1111/j.1523-1755.2005.00283.x
77. Liu J, Kesiry R, Periyasamy SM, Malhotra D, Xie Z, Shapiro JI. Ouabain induces endocytosis of plasmalemmal Na/K-ATPase in LLC-PK1 cells by a clathrin-dependent mechanism. *Kidney Int* (2004) **66**:227–41. doi:10.1111/j.1523-1755.2004.00723.x
78. Cai H, Wu L, Qu W, Malhotra D, Xie Z, Shapiro JI, et al. Regulation of apical NHE3 trafficking by ouabain-induced activation of the basolateral Na<sub>+</sub>-K<sub>+</sub>-ATPase receptor complex. *Am J Physiol Cell Physiol* (2008) **294**:C555–63. doi:10.1152/ajpcell.00475.2007
79. Oweis S, Wu L, Kiela PR, Zhao H, Malhotra D, Ghishan FK, et al. Cardiac glycoside downregulates NHE3 activity and expression in LLC-PK1 cells. *Am J Physiol Renal Physiol* (2006) **290**:F997–1008. doi:10.1152/ajprenal.00322.2005
80. Periyasamy SM, Liu J, Tanta F, Kabak B, Wakefield B, Malhotra D, et al. Salt loading induces redistribution of the plasmalemmal Na/K-ATPase in proximal tubule cells. *Kidney Int* (2005) **67**:1868–77. doi:10.1111/j.1523-1755.2005.00285.x
81. Wansapura AN, Lasko V, Xie Z, Fedorova OV, Bagrov AY, Lingrel JB, et al. Marinobufagenin enhances cardiac contractility in mice with ouabain-sensitive alpha1 Na<sub>+</sub>-K<sub>+</sub>-ATPase. *Am J Physiol Heart Circ Physiol* (2009) **296**:H1833–9. doi:10.1152/ajpheart.00285.2009
82. Loreaux EL, Kaul B, Lorenz JN, Lingrel JB. Ouabain-sensitive alpha1 Na<sub>+</sub>-K<sub>+</sub>-ATPase enhances natriuretic response to saline load. *J Am Soc Nephrol* (2008) **19**:1947–54. doi:10.1681/ASN.2008020174
83. Dostanic-Larson I, Lorenz JN, Van Huysse JW, Neumann JC, Moseley AE, Lingrel JB. Physiological role of the alpha1- and alpha2-isoforms of the Na<sub>+</sub>-K<sub>+</sub>-ATPase and biological significance of their cardiac glycoside binding site. *Am J Physiol Regul Integr Comp Physiol* (2006) **290**:R524–8. doi:10.1152/ajpregu.00838.2005
84. Dostanic I, Schultz Jel J, Lorenz JN, Lingrel JB. The alpha 1 isoform of Na<sub>+</sub>, K-ATPase regulates cardiac contractility and functionally interacts and co-localizes with the Na/Ca exchanger in heart. *J Biol Chem* (2004) **279**:54053–61. doi:10.1074/jbc.M410737200
85. Liu J, Yan Y, Liu L, Xie Z, Malhotra D, Joe B, et al. Impairment of Na/K-ATPase signaling in renal proximal tubule contributes to Dahl salt-sensitive hypertension. *J Biol Chem* (2011) **286**:22806–13. doi:10.1074/jbc.M111.246249
86. Yan Y, Shapiro AP, Haller S, Katragadda V, Liu L, Tian J, et al. Involvement of reactive oxygen species in a feed-forward mechanism of Na/K-ATPase-mediated signaling transduction. *J Biol Chem* (2013) **288**:34249–58. doi:10.1074/jbc.M113.461020
87. Liu J, Shapiro JI. Regulation of sodium pump endocytosis by cardiotonic steroids: molecular mechanisms and physiological implications. *Pathophysiology* (2007) **14**:171–81. doi:10.1016/j.pathophys.2007.09.008
88. Eichna LW, Farber SJ, Berger AR, Earle DP, Rader B, Pellegrino E, et al. The interrelationships of the cardiovascular, renal and electrolyte effects of intravenous digoxin in congestive heart failure. *J Clin Invest* (1951) **30**:1250–61. doi:10.1172/JCI102545
89. Buckalew V. Is endogenous ouabain a physiological regulator of cardiovascular and renal function? *Am J Physiol Heart Circ Physiol* (2009) **297**:H1972–3. doi:10.1152/ajpheart.01002.2009
90. Nesher M, Dvela M, Igbokwe VU, Rosen H, Lichtstein D. Physiological roles of endogenous ouabain in normal rats. *Am J Physiol Heart Circ Physiol* (2009) **297**:H2026–34. doi:10.1152/ajpheart.00734.2009
91. Kennedy DJ, Vetteth S, Periyasamy SM, Kanj M, Fedorova L, Khouri S, et al. Central role for the cardiotonic steroid marinobufagenin in the pathogenesis of experimental uremic cardiomyopathy. *Hypertension* (2006) **47**:488–95. doi:10.1161/01.HYP.0000202594.82271.92
92. Kennedy DJ, Elkareh J, Shidyak A, Shapiro AP, Smaili S, Mutgi K, et al. Partial nephrectomy as a model for uremic cardiomyopathy in the mouse. *Am J Physiol Renal Physiol* (2008) **294**:F450–4. doi:10.1152/ajprenal.00472.2007
93. Haller ST, Kennedy DJ, Shidyak A, Budny GV, Malhotra D, Fedorova OV, et al. Monoclonal antibody against marinobufagenin reverses cardiac fibrosis in rats with chronic renal failure. *Am J Hypertens* (2012) **25**:690–6. doi:10.1038/ajh.2012.17

94. Tian J, Shidyak A, Periyasamy SM, Haller S, Taleb M, El-Okdi N, et al. Spironolactone attenuates experimental uremic cardiomyopathy by antagonizing marinobufagenin. *Hypertension* (2009) **54**:1313–20. doi:10.1161/HYPERTENSIONAHA.109.140038
95. Czuwara-Ladykowska J, Shirasaki F, Jackers P, Watson DK, Trojanowska M. Fli-1 inhibits collagen type I production in dermal fibroblasts via an Sp1-dependent pathway. *J Biol Chem* (2001) **276**:20839–48. doi:10.1074/jbc.M010133200
96. Finotti P, Palatini P. Canrenone as a partial agonist at the digitalis receptor site of sodium-potassium-activated adenosine triphosphatase. *J Pharmacol Exp Ther* (1981) **217**:784–90.
97. Fedorova LV, Raju V, El-Okdi N, Shidyak A, Kennedy DJ, Vetteth S, et al. The cardiotonic steroid hormone marinobufagenin induces renal fibrosis: implication of epithelial-to-mesenchymal transition. *Am J Physiol Renal Physiol* (2009) **296**:F922–34. doi:10.1152/ajprenal.90605.2008
98. Haller ST, Drummond CA, Yan Y, Liu J, Tian J, Malhotra D, et al. Passive immunization against marinobufagenin attenuates renal fibrosis and improves renal function in experimental renal disease. *Am J Hypertens* (2013) **27**(4):603–9. doi:10.1093/ajh/hpt169
99. Zhang Z, Li Z, Tian J, Jiang W, Wang Y, Zhang X, et al. Identification of hydroxyxanthones as Na/K-ATPase ligands. *Mol Pharmacol* (2010) **77**:961–7. doi:10.1124/mol.110.063974
100. Ferrari P, Ferrandi M, Valentini G, Bianchi G. Rostafuroxin: an ouabain antagonist that corrects renal and vascular Na<sup>+</sup>-K<sup>+</sup>-ATPase alterations in ouabain and adducin-dependent hypertension. *Am J Physiol Regul Integr Comp Physiol* (2006) **290**:R529–35. doi:10.1152/ajpregu.00518.2005
101. Ferrari P. Rostafuroxin: an ouabain-inhibitor counteracting specific forms of hypertension. *Biochim Biophys Acta* (2010) **1802**:1254–8. doi:10.1016/j.bbadi.2010.01.009
102. Li Z, Zhang Z, Xie JX, Li X, Tian J, Cai T, et al. Na/K-ATPase mimetic pNaK-tide peptide inhibits the growth of human cancer cells. *J Biol Chem* (2011) **286**:32394–403. doi:10.1074/jbc.M110.207597

**Conflict of Interest Statement:** Neither Dr. Joe Xie nor Ms. Anna Pearl Shapiro has any conflicts to report. Dr. Joseph Isaac Shapiro currently receives grant support from the NIH concerning this review topic (HL109015 as principal investigator, HL071556 and HL105649 as Co-investigator). Dr. Joseph Isaac Shapiro also holds some awarded patents related to this work (US Patent 8,283,441, Canadian Patents 2641303, 2667251, 2774486, 2360383).

Received: 11 April 2014; accepted: 07 June 2014; published online: 17 July 2014.

Citation: Xie JX, Shapiro AP and Shapiro JI (2014) The trade-off between dietary salt and cardiovascular disease; a role for Na/K-ATPase signaling? *Front. Endocrinol.* **5**:97. doi: 10.3389/fendo.2014.00097

This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Xie, Shapiro and Shapiro. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Natriuretic hormone: the ultimate determinant of the preservation of external sodium balance

Neal S. Bricker<sup>1\*</sup>, Christopher D. Cain<sup>2</sup> and Stewart Shankel<sup>3</sup>

<sup>1</sup> School of Medicine, University of California at Los Angeles, Los Angeles, CA, USA

<sup>2</sup> Phytoanalytics, Grand Terrace, CA, USA

<sup>3</sup> Department of Medicine, School of Medicine, University of California at Riverside, Riverside, CA, USA

## Edited by:

Harvey Craig Gonick, University of California, Berkeley, USA

## Reviewed by:

Joao Carlos Dos Reis Cardoso,  
University of Algarve, Portugal  
Vardaman Buckalew, Wake Forest  
School of Medicine, USA

## \*Correspondence:

Neal S. Bricker, 727 South Orange  
Grove Blvd., Suite 6, Pasadena, CA  
91105, USA

e-mail: nealbricker@ca.rr.com

The present manuscript focuses on a putative natriuretic hormone. It includes the history of a long-term search for the pure molecule, ranging from partial purification to synthesis. It includes a description of seven different bioassay systems used, a resume of the sequential steps in purification, and a summary of a series of experimental protocols employed in the effort to define the biologic properties of the inhibitor of sodium (Na) transport. Two closely related molecules were purified and synthesized. Both are xanthurenic acid derivatives (xanthurenic acid 8-O-β-D-glucoside and xanthurenic acid 8-O-sulfate). It is concluded that one or both of these two low molecular weight compounds (MW: 368 and 284) meet many of the criteria for the final modulator of Na excretion.

**Keywords:** natriuretic hormone, sodium transport, ENaC, xanthurenic acid derivatives, synthesized NH

## INTRODUCTION

It is our assumption that there is a single natriuretic hormone that serves to modulate the renal excretion of sodium (Na) so as to preserve an ongoing equality between consecutive 24 h Na excretion by the kidneys and the contemporaneous intake of Na. The focus on what we presume to be a hormone was initiated from the classic experiments of DeWardener et al. (1–3). The present symposium provides a contemporary review of the state-of-the-art of natriuretic hormone research by key investigators who have pursued the identity and biologic properties of several different putative natriuretic hormones.

Because of the design of DeWardener's protocol, it appeared likely that all of the factors then known to affect the renal excretion of Na could be excluded as the definitive control element. These included changes in glomerular filtration rate (GFR) (so-called "first factor") and changes in mineralocorticoid hormone activity ("second factor"). We coined the term "third factor" (4), which subsequently was replaced by "natriuretic factor."

Throughout a long period of time, a multinational group of investigators has sought to isolate and characterize the natriuretic hormone. To this point in time, despite considerable progress, the ultimate goal remains elusive. Hence, the present symposium.

Our interest in an endogenous natriuretic hormone arose out of long-term studies on the pathologic physiology of chronic progressive renal disease (CRD) (5). It was the pattern of Na excretion that evolved as nephron destruction proceeded.

What is a remarkable change in the estimated Na excretion rate per nephron from the beginning to the end of CRD is presented in Table 1. In constructing this table, the intake of Na per 24 h was maintained constant (e.g., at 120 mEq/day) and at all levels of GFR, external Na balance was preserved (the latter is not unusual down to a GFR as low as 10% of normal).

At a normal GFR of 120 ml/min in an 80 kg person, Na balance is achieved by the excretion of 1/2 of 1% of the filtered load of

Na. A fall in GFR to 60 ml/min mandates the excretion of 1% of the filtered Na. A further reduction of GFR (to 30 ml/min) due to the progression of the underlying renal disease requires the excretion of 2% of the filtered Na. And at a GFR of 15 ml/min, external Na balance is achieved by the excretion of 4% of the filtered Na. Thus, per unit of Na entering the extracellular fluid on a constant Na intake, the excretion rate per milliliter of residual GFR with no time delay increases by eightfold. Finally, in many forms of CRD, single nephron GFR (SNGFR) doubles as nephron loss advances. Na excretion rate per residual nephron per milliequivalents of Na intake, approaches 16 times the value in the normal subject.

## BIOASSAY SYSTEMS

Seven different bioassay systems were used in both the sequential steps in the isolation, purification, and synthesis of NH and the studies of the biologic effects of the test materials. Each assay system allowed for a quantitative measure of the inhibitory effects of a test sample on Na transport either *in vitro* or *in vivo*.

The primary *in vitro* systems (frog skin and toad bladder) (6, 7) involve polar epithelial cells that transport Na from the serosal surface of the membrane across the mucosal surface. Na transport is quantified by the short-circuit current across the isolated membrane (in an Ussing chamber). Activity of partially purified to pure NH was detectable only when the inhibitor was added to the serosal surface of the membrane.

In micropuncture studies (8), isolated cortical collecting tubules, dissected from normal rabbits were perfused with partially purified test material from the urine of uremic patients. The perfusate was delivered into the lumen of each nephron segment and the peritubular surface was bathed in a solution of known composition (see Experimental Data).

The *in vivo* assays were performed in rats (9). The test materials were delivered intravenously, intraarterially, or via a gastric tube.

**Table 1 | Adaptation in Na excretion per nephron in advancing CRD on a constant salt intake.**

GFR ml/ mn	Na in mEq/ day	Na out mEq/ day	$\mu\text{l}$ SNGFR % of normal	Na out per nephron nEq	Magnification per nephron
120	120	120	100	700	1
60	120	120		1,400	2
30	120	120		2,800	4
15	120	120	200	5,600	8
7 1/2	120	120		11,212	16

See text in reference to the increase in SNGFR. Column 6 depicts the exponential increase in sodium excretion per nephron per unit of sodium intake.

In the rat assay, the rats employed had: (1) two normal kidneys; (2) one remnant kidney (75% reduction of renal mass) and a contralateral normal kidney; or (3) a solitary remnant kidney following removal of the normal kidney.

### PURIFICATION OF NH

Twenty-four hour collections of urine were reduced from their original volume to approximately 20 ml of sludge by lyophilization. The sludge was then redissolved in isotonic saline to 25 ml components each equal to 6 h of original urine. Each sample then was chromatographed through Sephadex G-25 with online monitoring of UV absorption at 290 nm and electrical conductivity (10).

Biologic activity was limited to the “post-salt” peaks using the frog skin assay (11). Short-circuit current decreased from 40–50 to 20–30  $\mu\text{Amp}/\text{cm}^2$ . These samples were purified using high performance liquid chromatography (HPLC) runs. The active fractions were purified further by two consecutive HPLC runs. The eluate was monitored by fluorescence and UV absorbance.

After further concentration, a bioassay was performed by infusion of the test material into the uremic rat (11). A strong and sustained natriuresis was recorded.

The natriuretic fractions, consistently showed two peaks with strong fluorescence (excitation 332 nm; emission 430 nm) and characteristic UV absorption (UV max at 338 nm). These spectroscopic signatures were used as the main pooling criteria for further HPLC-based purification of NH (11). Additional steps in isolation, purification, and synthesis of the two molecules: xanthurenic acid 8-O- $\beta$ -D-glucoside and xanthurenic acid 8-O-sulfate are described in a separate publication (11).

### EXPERIMENTAL DATA

#### UREMIC VS. CONTROL SUBJECTS

Based on the adaptation in Na excretion in advancing CRD (see Table 1), bioassays were performed on normal rats using partially purified urine samples from 17 patients with advanced CRD (mean GFR 8.7 ml/min) and 14 normal control subjects. The assays from the normal subjects were negative [i.e., no significant increase in either absolute Na excretion ( $U_{\text{Na}}V$ ), or the fraction of filtered Na excreted ( $FE_{\text{Na}}$ ) (12)]. The uremic fractions produced a highly significant increase from baseline levels in both parameters

of Na excretion. With more concentrated samples of the uremic urine fractions, values for  $FE_{\text{Na}}$  rose to levels as high as 12%.

#### ADVANCED CRD WITH A SUPERIMPOSED EDEMA-FORMING STATE: (THE NEPHROTIC SYNDROME)

Eight patients with advanced CRD and the nephrotic syndrome were studied (12). Assays were performed on normal rats using partially purified fractions of serum in all eight studies. Fractions of urine were also used in three studies. Values for  $U_{\text{Na}}V$  decreased from control levels (by an average of 0.97  $\mu\text{Eq}/\text{min}$ ). The mean value for  $FE_{\text{Na}}$  also decreased (1.35%).

Danovitch et al. (13) studied a group of five patients with far advanced CRD (GFR 5.2–16.0 ml/min) who initially were in Na balance on controlled metabolic diets containing from 58 to 342 mEq of Na per day. In each patient, while under close observation (clinical and laboratory), the Na content of the diet was reduced at intervals of 1 week or longer over 4–14 weeks. Four of the patients exhibited a salt-losing state, wherein Na excretion exceeded Na intake. Two of these patients required intravenous salt replacement. At the completion of the studies, all patients maintained external Na balance while ingesting a mean of  $5.0 \pm 2.9$  mEq of Na/day. Thus, in contrast to patients with advancing CRD who maintain Na balance on a constant salt intake and a progressively decreasing nephron population, these patients were subjected to a progressive reduction of Na intake with an unchanging nephron population. (GFR remained constant throughout the studies.) The adaptive increase in Na excretion in advancing CRD (Table 1), thus was reversed with slow (and cautious) serial reductions in salt intake. The salt-losing tendency of CRD was reversed.

#### MICROPUNCTURE STUDIES

As described under Bioassay Systems, partially purified natriuretic factor was infused into the lumen of isolated cortical collecting tubules of normal rabbits. No effect was observed with intraluminal infusions. However, when the natriuretic material was added to the solution bathing the peritubular surface, the effects were rapid in onset and highly significant. Net Na flux (measured isotopically) from luminal to peritubular surface of the nephrons, decreased from 6.29 to 3.20 pmol/s ( $p < 0.001$ ) with no change in Na flux in the opposite direction. Potential difference rose rapidly from  $-22.5$  to  $-12$  mV. Control studies, using the same fraction from normal subjects, had no effect (8).

#### STUDIES ON NORMAL HUMAN BEINGS EXPOSED TO WATER IMMERSION

Epstein et al. (14) have established water immersion to the neck as a reliable and reproducible stimulus evoking natriuresis in normal subjects. The experiments were performed on 12 normal adults (15). Each was studied twice – once under control conditions sitting in a chair and again during water immersion. The paired studies were performed at the same time of day. Partially purified urine samples were assayed for natriuretic activity in normal rats.

$U_{\text{Na}}V$  and  $FE_{\text{Na}}$  did not change from the baseline values in the control studies. During water immersion  $U_{\text{Na}}V$  rose ( $1.27 \pm 0.28 \mu\text{Eq}/\text{min}$ ) and  $FE_{\text{Na}}$  increased from pre-immersion baseline values by  $1.29 \pm 0.21\%$ . Both changes were highly significant ( $p < 0.001$ ) (15).

## STUDIES IN DOGS

Normal dogs were maintained on a Na intake varying from 3 to 258 mEq/day, with and without fludrocortisone. Urine was partially purified and assayed for natriuretic effect in normal rats and for inhibition of Na transport in isolated toad bladders. In dogs on the 258 mEq Na diet and 0.2 mg fludrocortisone/day, both a statistically significant natriuretic response and inhibition of short-circuit current ( $p < 0.001$ ) were observed. In dogs fed 3 mEq of Na per day with fludrocortisone, no significant effect was found in either assay system (16).

## END-ORGAN RESPONSIVENESS

The effects of nephron loss on the natriuretic response (9) to partially purified natriuretic factor were studied in three groups of rats, each on a normal salt diet.

1. Group 1: normal rats: intraarterial infusion into one renal artery produced a unilateral natriuresis.
2. Group 2: intraarterial infusion into the renal artery of a unilateral remnant kidney in rats with a contralateral normal kidney produced an increase in  $\text{FE}_{\text{Na}}$ , which was equal bilaterally. Intravenous infusion of the natriuretic fraction also produced comparable increments of  $\text{FE}_{\text{Na}}$  in the remnant and the normal kidneys.
3. Group 3: in uremic rats with a solitary remnant kidney (no contralateral kidney), the intraarterial infusion of natriuretic factor produced an increase in  $\text{FE}_{\text{Na}}$  that was significantly greater than in remnant kidneys of group 2 rats or normal kidneys of group 1.

## NATRIURETIC RESPONSE TO SYNTHESIZED (PURE) NH

Synthesized preparations of xanthurenic acid 8-O- $\beta$ -D-glucoside (NH) and xanthurenic acid 8-O-sulfate (NH-1) were bioassayed in normal rats (11).

The intravenous infusion of NH (range 0.14–16.4 nmol) and NH-1 (0.7–4.21 nmol) was studied in eight and five normal rats, respectively. In the NH group,  $\Delta U_{\text{Na}}V$  averaged  $3.68 \pm 0.55 \mu\text{Eq}/\text{min}$ ; in the five experiments in which NH-1 was the test substance,  $\Delta U_{\text{Na}}V$  averaged  $4.33 \pm 0.71 \mu\text{Eq}/\text{min}$  (1).

In five additional studies, a combination of NH (1.4–3.41 nmol) and NH-1 (1.75–5.44 nmol) was assayed.  $\Delta U_{\text{Na}}V$  averaged  $5.0 \pm 0.89 \mu\text{Eq}/\text{min}$ .

## STUDIES BY HOFFMAN AND ASSOCIATES

In a recent publication, Hoffman et al. (17) studied the effects of xanthurenic acid 8-O- $\beta$ -D-glucoside on Na excretion in adult male Sprague-Dawley rats. Each rat was given two consecutive incremental doses (6.3 + 31.5 nmol) of NH (designated as XAG). Values for  $\Delta U_{\text{Na}}V$  were  $3.21 \pm 1.12$  and  $3.99 \pm 0.95$  at the two dosages, respectively. Values for  $\Delta \text{FE}_{\text{Na}}$  increased significantly ( $1.63 \pm 0.46$ ) during the second dose of XAG.

In these studies, GFR (inulin clearance) remained unchanged. Mean arterial pressure (MAP) and total renal blood flow were recorded electronically every 5 min for 60 s during the control periods and for 30–40 min periods during the XAG infusions. All hemodynamic values remained stable (17).

Two important new observations were made by Hoffman and coworkers:

1. In rats pretreated with amiloride, an inhibitor of  $\text{E}_{\text{Na}}\text{C}$ , the epithelial Na channel in the distal tubule, the natriuretic effects of XAG were completely abolished (17).
2. In rats subjected to chronic blockade of the NO system, the natriuretic response to XAG was diminished suggesting to these investigators that the renal effects of XAG could be mediated in part by activation of the renal NO system (17).

## DISCUSSION

A large number of factors, both humoral and physical are known to influence the renal tubular transport of Na. But none of these, including changes in GFR or mineralocorticoid hormone activity (see Introduction) is believed to be the final modulator of net Na transport and thus of Na excretion. We believe that natriuretic hormone fulfills this role. It thus would serve as the definitive element of a sophisticated biologic control system that is charged with the preservation of Na balance and with the constancy of the extracellular fluid volume. But, while there may be virtual unanimity of opinion about the existence of natriuretic hormone, there is no such unanimity about the nature of this hormone.

In this manuscript, we have reviewed the properties of a natriuretic factor, which we have pursued for a long period of time. The activity was obtained from both serum (or plasma) and urine using material that ranged from partially purified to pure, chemically synthesized molecules (11). We believe that the experimental data reviewed in the manuscript meet at least some of the criteria for natriuretic hormone. These include:

1. The rapid and reversible inhibition of net Na transport across polar epithelial cell systems, including the distal portion of the nephron.
2. The foregoing biologic activity is present only when the inhibitor is added to the peritubular surface of the nephron or the serosal surface of equivalent *in vitro* models.
3. The natriuretic effect of the purified and synthesized material is completely blocked by prior administration of amiloride, an inhibitor of  $\text{E}_{\text{Na}}\text{C}$  activity in the distal portion of the nephron.
4. Inhibition of Na transport in the distal nephron associated with a rapid shift in transepithelial electric potential difference (i.e., a less negative intraluminal potential). This change would favor the excretion of Cl with Na as opposed to increased secretion of K.
5. No evidence for a fixed coupling ratio between the inhibition of Na transport and K secretion.
6. Increase in natriuretic activity in advancing CRD in purified serum or urine samples.
7. An increase in end-organ responsiveness to the inhibitor associated with nephron loss.
8. Lack of natriuretic effect of the inhibitor in patients with advanced CRD and a superimposed edema-forming state (the nephrotic syndrome).
9. Reversal of salt-losing state in advanced CRD by progressive slow reduction of salt intake to very low levels.

**Table 2 | A comparison of several natriuretic substances.**

	<b>Ouabain (OLS)</b>	<b>Marinobufogenin (MFG)</b>	<b>Vanadium diascorbate (VD)</b>	<b>Atrial natriuretic peptides</b>			<b>Xanthurenic acid 8-O-<math>\beta</math>-D-glucoside (XAG)</b>
				<b>ANP</b>	<b>BNP</b>	<b>CNP</b>	
Isolation	Plasma and adrenal cortex and hypothalamus (18–23)	Plasma, urine, and adrenal cortex (24, 25)	Urine and plasma (26)	Atria, heart, and kidney (27)	Brain, heart, and kidney (27)	Brain, heart, and vasculature (27, 28)	Plasma and urine from uremic patients (6, 8, 10, 29)
Stimulus	All, ACTH, ↑BP positive sodium balance or intake; ↑ serum K+ up to 5 mEq/l	Same as OLS	Salt loading, aldosteronism, and volume expansion (26)	Stretch of cardiac wall – especially the atria, ET adrenergic stimuli (30)	Same as ANP (30)	Same? as ANP (30)	Normal and uremic patients and animals (10, 15)
M.W.	584.6 (31)	387? 600? (31)	403	2,000–3,000± (32)	2,000–3,000± (32)	2,000–3,000± (32)	368 (glucoside) 284 (sulfate) (11)
Structure	Steroid (31)	Steroid (31)	Vanadate diascorbate from ascorbic acid (26)	28 AA (32)	32 AA (32)	22 AA (32)	8-O-(D-glucoside) 8-O-sulfate of xanthurenic acid from tryptophan (11)
Site of Action	α2α3 NaK ATPase primarily (18–23, 25, 26, 33–43) acts on basolateral membrane of PCT and NHE3 in PCT (36, 37)	α1 NaK ATPase in PCT (24, 44)	NaK ATPase in PCT acts on basolateral membrane(26)	Blocks ENaC, ↑ GFR, blocks NaK ATPase All in PCT (30, 45, 46) ↓ H <sub>2</sub> O absorption in CT, ↓ urine concentration (47, 48)	Similar to ANP	Direct vasodilator and simulator to ANP (47)	Blocks ENaC acts from basolateral surface (17)
Natriuresis	Variable from no natriuresis to mild natriuresis (21, 25, 33, 36, 38, 39) No effect on α1-NaK ATPase	Variable natriuresis (24, 44)	Moderate natriuresis (26)	Moderate through CGMP (28, 30, 32, 46)	Same as ANP(28, 30, 32, 46, 49)	None (28, 47)	Eliminated by blocking ENaC (17)
RBF	↓ (40)	?	?	↑ RBF → ↑ GFR (30, 45, 50)	Variable (50)	No	No effect (17)
GFR	↓ Or no change (40)	?	?	↑ GFR dilates afferent arteriole and constricts efferent (49)	Same as ANP (49)	No	No effect (17)
K excretion	↓	↓	↓	↑	↑	↑	Minimal (11)
Vasoactivity	↑ BP; vasoconstriction (21, 22, 33, 34, 36, 40–43) ↑ Ca influx and Na influx into vessel wall (22, 34, 42–44)	↑ BP; vasoconstriction (24, 44)	↑ BP (27) ↑ Ca and Na influx into vessel wall (25, 27)	↓ BP (32, 47)	↓ BP (32, 47)	?	None (17)

10. Presence of natriuretic activity in urine of normal subjects during water immersion – an experience known to produce central hypervolemia.
11. Increased natriuretic activity in normal dogs on a high salt diet and superimposed mineralocorticoid hormone.
12. No natriuretic activity in normal dogs on a low salt diet and superimposed mineralocorticoid hormone.

### OTHER "NATRIURETIC" FACTORS

Four other categories of putative natriuretic hormones are considered in separate papers in this symposium. These compounds include: (1) ouabain [or ouabain-like substances (OLS)]; (2) marinobufogenin (MFG); (3) vanadium diascorbate (VD); and atrial natriuretic peptides (ANP).

A summary of key properties of each is shown in **Table 2** and a brief description of some relevant characteristics follows.

### OUABAIN (OLS)

A small molecule (MW 584.6) isolated from plasma (18), urine, adrenal cortex, and hypothalamus (19–23). The primary and relevant effects are the inhibition of NaK ATPase and the cross reaction with ouabain antibodies (21).

But they are inconsistently natriuretic (21, 22, 33, 34), presumably because ouabain has little effect on the  $\alpha 1$  subunit of NaK ATPase (22, 35). Indeed, there also recent evidence that OLS may actually cause Na retention (26).

### MARINOBUFogenin

Also a small molecule (MW between 387 and 600) (31), which is isolated from plasma, urine, and the adrenal cortex (24, 44). No studies have shown that administration of MFG causes natriuresis in assay animals. However, administration of anti-MBG antibodies reduces Na excretion (24, 44). Exogenous administration of bufalin, a very closely related compound, injected into the renal artery of sheep has been shown to produce natriuresis. No similar studies have as yet been published using MFG (51).

### VANADIUM DIASCORBATE

A small molecule (MW 403) isolated from plasma and urine (26). Inhibits NaK ATPase in the proximal convoluted tubule. Produces moderate natriuresis, may produce influx of  $\text{Ca}^{++}$  into vessel walls and thereby increase BP.

### ATRIAL NATRIURETIC PEPTIDES

Molecular weight 2,000–3,000 $\pm$  (32). The natriuretic peptides, while producing natriuresis, also increase GFR and increase glomerular pressure by dilating the afferent arteriole, and constricting the efferent arteriole (30, 45, 50). It may cause progressive renal failure in animal models.

If ANP is the final modulator of Na balance, there is a paradox. In advanced cirrhosis, there is an increase in preload and a decrease in central volume and EAV. ANP levels are elevated (50). Likewise in CHF there is a decrease in EAV and an increase in preload and again ANP levels are elevated. Physiologically both situations should invoke Na retention due to the decrease in the EAV, which in turn should shut off natriuresis, but in both of these situations ANP is elevated.

The arterial natriuretic peptides are primarily vasoactive and act on multiple sites of the nephron including the proximal tubule.

### CONCLUSION

The cumulative data presented in this paper lend support to the view that xanthurenic acid 8-O- $\beta$ -D-glucoside and xanthurenic acid 8-O-sulfate could be the long sought after and elusive natriuretic hormone. But to validate, or refute this thesis, additional experimental evidence is required. A partial list of key areas of future study includes:

1. A sensitive assay system for quantifying the levels of the putative hormone in body fluids.
2. Establishing the site of production.
3. The character of the signaling process (including the nature of the receptors), involved in initiating and controlling the induced natriuresis.
4. The enzymes involved in the synthesis.

### ACKNOWLEDGMENTS

Thanks to Nancy Price for her untiring patience and expertise.

### REFERENCES

1. de Wardener HE, Mills IH, Clapham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intravenous saline in the dog. *Clin Sci* (1961) **21**:249–58.
2. Mills IH, de Wardener HE, Hayter CJ, Clapham WF. Studies on the afferent mechanisms of the sodium diuresis which follows the administration of saline in the dog. *Clin Sci* (1961) **21**:259–64.
3. Nutbourne DM, de Wardener HE. The effect of a water diuresis on the urinary excretion hydrogen ions in man. *Clin Sci* (1961) **20**:63–73.
4. Bricker NS. The control of sodium excretion with normal and reduced nephron populations. The pre-eminence of third factor. *Am J Med* (1967) **43**:313–21. doi:10.1016/0002-9343(67)90188-X
5. Bricker NS, Zeal L, Shapiro M, Sanclemente E, Shankel S. Biologic and physical characteristics of the non-peptidic, non-digitalis-like natriuretic hormone. *Kidney Int* (1993) **44**:937–47. doi:10.1038/ki.1993.335
6. Bourgoignie J, Klahr S, Bricker NS. Inhibition of transepithelial sodium transport in the frog skin by a low molecular weight fraction of uremic serum. *J Clin Invest* (1971) **50**:303–11. doi:10.1172/JCI106495
7. Buckalew VM, Martinez FJ, Green WE. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**:926–35. doi:10.1172/JCI106312
8. Fine LG, Bourgoignie JJ, Hwang KH, Bricker NS. On the influence of the natriuretic factor from patients with chronic uremia on the bioelectric properties and sodium transport of the isolated mammalian collecting tubule. *J Clin Invest* (1976) **58**:590–7. doi:10.1172/JCI108505
9. Fine LG, Bourgoignie JJ, Weber H, Bricker NS. Enhanced end-organ responsiveness of the uremic kidney to the natriuretic factor. *Kidney Int* (1976) **10**:364–72. doi:10.1038/ki.1976.122
10. Bricker NS, Klahr S, Purkerson M, Schultze RG, Avioli LV, Birge SJ. *In vitro* assay for a humoral substance present during volume expansion and uraemia. *Nature* (1968) **219**:1059–1059. doi:10.1038/2191058a0
11. Cain CD, Schroeder FC, Shankel SW, Mitchnick M, Schmertzler M, Bricker NS. Identification of xanthurenic acid 8-O- $\beta$ -D-glucoside and xanthurenic acid 8-O-sulfate as human natriuretic hormones. *Proc Natl Acad Sci U S A* (2007) **104**:17873–8. doi:10.1073/pnas.0705553104
12. Bourgoignie JJ, Hwang KH, Ipakchi E, Bricker NS. The presence of a natriuretic factor in urine of patients with chronic uremia. The absence of the factor in nephrotic uremic patients. *J Clin Invest* (1974) **53**:1559–67. doi:10.1172/JCI107706
13. Danovitch GM, Bourgoignie JJ, Bricker NS. Reversibility of the “salt-losing” tendency of chronic renal failure. *N Engl J Med* (1977) **296**:14–9. doi:10.1056/NEJM197701062960104

14. Epstein M. Renal effects of head-out water immersion in man: implications for an understanding of volume homeostasis. *Physiol Rev* (1978) **58**:529–81.
15. Epstein M, Bricker NS, Bourgoignie JJ. Presence of a natriuretic factor in urine of normal men undergoing water immersion. *Kidney Int* (1978) **13**:152–8. doi:10.1038/ki.1978.22
16. Favre H, Hwang KH, Schmidt RW, Bricker NS, Bourgoignie JJ. An inhibitor of sodium transport in the urine of dogs with normal renal function. *J Clin Invest* (1975) **56**:1302–11. doi:10.1172/JCI108206
17. Hoffman A, Ovcharenko E, Karram T, Okun-Gurevich M, Goltzman I, Cain C, et al. Renal effects of a novel endogenous natriuretic agent xanthurenic acid 8-O-β-D-glucoside in rats. *Physiol Rep* (2013) **1**:e00155. doi:10.1002/phy2.155
18. Hamlyn JM, Blaustein MP, Bova S, DuCharme DW, Harris DW, Mandel F, et al. Identification and characterization of a ouabain-like compound from human plasma. *Proc Natl Acad Sci U S A* (1991) **88**:6259–62. doi:10.1073/pnas.88.21.9907-d
19. Dvela M, Rosen H, Ben-Ami HC, Lichtstein D. Endogenous ouabain regulates cell viability. *Am J Physiol Cell Physiol* (2012) **302**:C442–52. doi:10.1152/ajpcell.00336.2011
20. Tamura M, Lam TT, Inagami T. Specific endogenous  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase inhibitor purified from bovine adrenal. *Biochem Biophys Res Commun* (1987) **149**:468–74. doi:10.1016/0006-291X(87)90391-3
21. Nesher M, Dvela M, Igboekwe U, Rosen H, Lichtstein D. Physiological roles of endogenous ouabain in normal rats. *Am J Physiol* (2009) **297**:H2026–34. doi:10.1152/ajpheart.00734.2009
22. Blaustein MP, Leenen FHH, Chen L, Golovina VA, Hamlyn JM, Pallone TL, et al. How NaCl raises blood pressure: a new paradigm for the pathogenesis of salt-dependent hypertension. *Am J Physiol* (2011) **302**:H1031–49. doi:10.1152/ajpheart.00899.2011
23. Tamura M, Konishi F, Sakakibara M, Inagami T. Large scale purification of an endogenous  $\text{Na}^+/\text{K}^+$ -pump inhibitor from bovine adrenal glands. In: Bamberg E, Schoner W, editors. *Sodium Pump: Structure, Mechanism, Hormonal Control and its Role in Disease*. New York, NY: Springer (1994). p. 763–6.
24. Haller ST, Kennedy DJ, Shidyak A, Budny GV, Malhotra D, Fedorova OV, et al. Monoclonal antibody against marinobufagenin reverses cardiac fibrosis in rats with chronic renal failure. *Am J Hypertens* (2012) **25**:690–6. doi:10.1038/ajh.2012.17
25. Nowicki S, Enero MA, de Lores Arnaiz G. Diuretic and natriuretic effect of a brain soluble fraction that inhibits neuronal  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase. *Life Sci* (1990) **47**:1091–8. doi:10.1016/0024-3205(90)90167-P
26. Kramer HJ, Krampitz A, Bäcker A, Meyer-Lehnert H. Ouabain-like factors in human urine: identification of a Na-K-ATPase inhibitor as vanadium-diascorbate adduct. *Clin Exp Hypertens* (1998) **20**:557–71. doi:10.3109/10641969809053234
27. Beltowski J, Wójcicka G. Regulation of renal tubular sodium transport by cardiac natriuretic peptides: two decades of research. *Med Sci Monit* (2002) **8**:RA39–52.
28. Stingo AJ, Clavell AL, Aarhus LL, Burnett JC Jr. Cardiovascular and renal actions of C-type natriuretic peptide. *Am J Physiol* (1992) **262**:H308–12.
29. Bourgoignie JJ, Hwang KH, Espinel C, Klahr S, Bricker NS. A natriuretic factor in the serum of patients with chronic uremia. *J Clin Invest* (1972) **51**:1514–27. doi:10.1172/JCI106948
30. Brenner BM, Ballerman BJ, Gunning ME, Zeidel ML. Diverse biological actions of atrial natriuretic peptide. *Physiol Rev* (1990) **70**:665–99.
31. Puschett JB, Agunanne E, Uddin MH. Emerging role of bufadienolides in cardiovascular and kidney diseases. *Am J Kidney Dis* (2010) **56**:359–70. doi:10.1053/j.ajkd.2010.01.023
32. Lee CYW, Burnett JC. Natriuretic peptides and therapeutic applications. *Heart Fail Rev* (2007) **12**:131–42. doi:10.1007/s10741-007-9016-3
33. Manunta P, Messaggio E, Ballabeni C, Sciarrone MT, Lanzani C, Ferrandi M, et al. Plasma ouabain-like factor during acute and chronic changes in sodium balance in essential hypertension. *Hypertension* (2001) **38**:198–203. doi:10.1161/01.HYP.38.2.198
34. Anderson DE, Fedorova OV, Morrell CH, Longo DL, Kashkin VA, Metzler JD, et al. Endogenous sodium pump inhibitors and age-associated increases in salt sensitivity of blood pressure in normotensives. *Am J Physiol* (2008) **294**:R1248–54. doi:10.1152/ajpregu.00782.2007
35. Katz A, Lifshitz Y, Bab-Dinitz E, Kapri-Pardes E, Goldschleger R, Tal DM, et al. Sensitivity of digitalis glycosides for isoforms of human Na,K-ATPase. *J Biol Chem* (2010) **285**:19582–92. doi:10.1074/jbc.M110.119248
36. Liu J, Xie Z-J. The sodium pump and cardiotonic steroids-induced signal transduction protein kinases and calcium-signaling microdomain in regulation of transporter trafficking. *Biochim Biophys Acta* (2010) **1802**:1237–45. doi:10.1016/j.bbadi.2010.01.013
37. Khundmiri SJ. Advances in understanding the role of cardiac glycosides in control of sodium transport in renal tubules. *J Endocrinol* (2014) **222**(1):R11–24. doi:10.1530/JOE-13-0613
38. Gupta S, Yan Y, Malhotra D, Liu J, Xie Z, Najjar SM, et al. Ouabain and insulin induce sodium pump endocytosis in renal epithelium. *Hypertension* (2012) **59**:665–72. doi:10.1161/HYPERTENSIONAHA.111.176727
39. Periyasamy SM, Liu J, Tanta F, Kabak B, Wakefield B, Malhotra D, et al. Salt loading induces redistribution of the plasmalemmal Na/K-ATPase in proximal tubule walls. *Kidney Int* (2005) **67**:1868–77. doi:10.1111/j.1523-1755.2005.00285.x
40. Nechay BR, Chinoy DA. Effect of ouabain on renal transport of P-aminohippuric acid (PAH) and blood flow in the dog. *Eur J Pharmacol* (1968) **3**:322–9. doi:10.1016/0014-2999(68)90115-5
41. Liu J, Yan Y, Liu L, Xie Z, Malhotra D, Joe B, et al. Impairment of Na/K-ATPase signaling in renal proximal tubule contributes to Dahl salt-sensitive hypertension. *J Biol Chem* (2011) **286**:22806–13. doi:10.1074/jbc.M111.246249
42. Zhang J, Lee MY, Cavalli M, Chen L, Berra-Romani R, Balke CW, et al. Sodium pump  $\alpha 2$  subunits control myogenic tone and blood pressure in mice. *J Physiol* (2005) **569**:243–56. doi:10.1113/jphysiol.2005.091801
43. Blaustein MP, Zhang J, Chen L, Song H, Raina H, Kinsey SP, et al. The pump, the exchanger, and endogenous ouabain. Signaling mechanisms that link salt retention to hypertension. *Hypertension* (2008) **53**:291–8. doi:10.1161/HYPERTENSIONAHA.108.119974
44. Fedorova OV, Talan MI, Agalakova NI, Lakatta EG, Bagrov AY. Endogenous ligand of  $\alpha_1$  sodium pump, marinobufagenin, is a novel mediator of sodium chloride-dependent hypertension. *Circulation* (2002) **105**:1122–7. doi:10.1161/hc0902.104710
45. Light DB, Corbin JD, Stanton BA. Dual ion channel regulation by cyclic GMP and cyclic-dependent protein kinase. *Nature* (1990) **344**:336–9. doi:10.1038/344336a0
46. Valdivieso A. The kidney in chronic liver disease: circulatory abnormalities, renal sodium handling and role of natriuretic peptides. *Biol Res* (1998) **31**:291–304.
47. Weidmann P, Hasler L, Gnadinger MP, Lang RE, Uehlinger DE, Shaw S, et al. Blood levels and renal effects of atrial natriuretic peptide in normal man. *J Clin Invest* (1986) **77**:734–42. doi:10.1172/JCI112368
48. Pandey KN. Biology of natriuretic peptides and their receptors. *Peptides* (2005) **26**:901–32. doi:10.1016/j.peptides.2005.03.055
49. Eiskjaer H, Pedersen EB. Dose-response study of atrial natriuretic peptide bolus injection in healthy man. *Eur J Clin Invest* (1993) **23**:37–45. doi:10.1111/j.1365-2362.1993.tb00715.x
50. Gunning ME, Brenner BM. Natriuretic peptides and the kidney: current concepts. *Kidney Int* (1992) **42**:S127–33.
51. Yates NA, McDougall IG. Effect of direct arterial infusion of bufalin and ouabain in conscious sheep. *Br J Pharmacol* (1993) **108**:627–30. doi:10.1111/j.1476-5381.1993.tb12852.x

**Conflict of Interest Statement:** During the long period of study covered by this review, major support was provided by Program Project Grants from the NIH and from Naturon Pharmaceutical Company. The authors do not have any conflict of interest in relation to this review.

Received: 02 October 2014; accepted: 24 November 2014; published online: 11 December 2014.

Citation: Bricker NS, Cain CD and Shankel S (2014) Natriuretic hormone: the ultimate determinant of the preservation of external sodium balance. *Front. Endocrinol.* **5**:212. doi: 10.3389/fendo.2014.00212

This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Bricker, Cain and Shankel. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Natriuretic hormones, endogenous ouabain, and related sodium transport inhibitors

John M. Hamlyn\*

Department of Physiology, University of Maryland School of Medicine, Baltimore, MD, USA

**Edited by:**

Vardaman Buckalew, Wake Forest School of Medicine, USA

**Reviewed by:**

Vardaman Buckalew, Wake Forest School of Medicine, USA

Frans H. H. Leenen, University of Ottawa Heart Institute, Canada

**\*Correspondence:**

John M. Hamlyn, Department of Physiology, University of Maryland School of Medicine, 655 West Baltimore Street, Baltimore, MD 21201, USA

e-mail: jhamlyn@umaryland.edu

The work of deWardener and colleagues stimulated longstanding interest in natriuretic hormones (NHs). In addition to the atrial peptides (APs), the circulation contains unidentified physiologically relevant NHs. One NH is controlled by the central nervous system (CNS) and likely secreted by the pituitary. Its circulating activity is modulated by salt intake and the prevailing sodium concentration of the blood and intracerebroventricular fluid, and contributes to postprandial and dehydration natriuresis. The other NH, mobilized by atrial stretch, promotes natriuresis by increasing the production of intrarenal dopamine and/or nitric oxide (NO). Both NHs have short (<35 min) circulating half lives, depress renotubular sodium transport, and neither requires the renal nerves. The search for NHs led to endogenous cardiotonic steroids (CTS) including ouabain-, digoxin-, and bufadienolide-like materials. These CTS, given acutely in high nanomole to micromole amounts into the general or renal circulations, inhibit sodium pumps and are natriuretic. Among these CTS, only bufalin is cleared sufficiently rapidly to qualify for an NH-like role. Ouabain-like CTS are cleared slowly, and when given chronically in low daily nanomole amounts, promote sodium retention, augment arterial myogenic tone, reduce renal blood flow and glomerular filtration, suppress NO in the renal vasa recta, and increase sympathetic nerve activity and blood pressure. Moreover, lowering total body sodium raises circulating endogenous ouabain. Thus, ouabain-like CTS have physiological actions that, like aldosterone, support renal sodium retention and blood pressure. In conclusion, the mammalian circulation contains two non-AP NHs. Identification of the CNS NH should be a priority.

**Keywords:** salt, sodium, urine, excretion, sodium pump, ouabain, hormone

## INTRODUCTION

Natriuretic hormones (NHs) can be defined as substances whose circulating levels and effects fluctuate in a parallel manner with dietary sodium intake (1). NHs have long been implicated in sodium balance and are likely to be of the most significance in western acculturated societies where sodium intake typically is >100 meq/day (2). Indeed, ingestion of high salt meals raises the osmolarity of the circulation, stimulates secretion of antidiuretic hormone (ADH), and raises the natriuretic activity of the blood. In principle, the mode of action of NHs includes suppression of primary active sodium transport in the kidney and/or damping of secondary active transport systems involving sodium (1) or even potassium (3), effects on renal vascular tone and glomerular filtration rate (GFR), and activation of intrarenal natriuretic factors, such as prostaglandins, nitric oxide (NO), or dopamine. This article presents a personal and condensed overview of known and unknown non-atrial NHs and addresses the role of endogenous sodium pump inhibitors as NHs.

## SEARCHING FOR NATRIURETIC HORMONES

It is well accepted that sodium balance is not fully explained by the up and downregulation of glomerular filtration and mineralocorticoid-stimulated reabsorption (4, 5). The first clear evidence for a “third factor” arose from the pioneering

experiments of deWardener in which dogs that received excess mineralocorticoid and vasopressin increased their urinary sodium excretion in response to blood volume expansion with saline at a time when glomerular filtration was being lowered experimentally (6). Thus, the increase in sodium excretion was mediated by diminished tubular reabsorption of sodium and water. Cross-circulation studies, as well as work using isolated kidney studies in dogs and rats (6–10) excluded significant alterations in the composition of the blood, changes in renal nerve activity, glomerular filtration, renal blood flow, or renal perfusion pressure as mediators. A humoral “NH” was required.

The discovery of the atrial peptides (APs) and their natriuretic activity initially promised to explain some of the outstanding functions of an NH (11–13). APs augment sodium excretion (14–16) and saline infusions raise plasma AP (17, 18). However, in dogs, the effects of physiological changes in plasma APs and low dose infusions on sodium excretion were less obvious and, under certain experimental conditions, circulating APs and sodium excretion changed diametrically or, were temporally unconnected (19–21). Thus, some other NH was required.

The search for humoral agents that trigger salt excretion has relied on a variety of assays that range from isolated enzymes all the way to whole kidneys and animals (22). **Table 1** lists some tissues and fluids from which a variety of natriuretic factors

**Table 1 | An overview of sources and characteristics of natriuretic factors.**

Source for isolation	Characteristics	References
Adrenal	No short acting factors described Ouabain, <sup>a</sup> prosclilaridin A-like compound <sup>b</sup>	(8, 28, 29) (30)
Blood	Rapid onset, chymotrypsin-sensitive Rapid sustained natriuresis, MW < 500–700 Trypsin sensitive, slow onset Precursor? slow onset Leucine aminopeptidase-sensitive, chymotrypsin-resistant Ouabain <sup>a</sup>	(31–34) (35) (36) (37) (38–41) (42)
Hypothalamus/ pituitary	ADH, Oxytocin, MSH Ouabain <sup>a</sup>	See text (43)
Intestine	Guanylin (small heat stable peptide)	(26, 44)
Kidney	High MW, release PGE <sub>2</sub> dependent Urodilatin <sup>c</sup> (ANP 95–126) Small peptide	(45–48) (24, 49, 50)
Liver	Long acting, high MW (bound?), hepatic blood > portal blood	(51–59)
Urine	Low MW, Chymotrypsin-sensitive peptide Low MW, non-peptidic, acidic, Sephadex post salt fraction LLU- $\alpha$ <sup>d</sup> High MW, slow onset  Marinobufagenin <sup>e</sup> Prolidase-sensitive peptide Urodilatin <sup>c</sup> (small peptide) Uroguanylin (small heat stable peptide) Xanthurenic acid $\beta$ -glucoside and xanthurenic acid sulfate	(33) (60–62) (3) (36, 46, 63–65) (66) (61) See kidney (27, 44) (25, 67)

MW, molecular weight.

All materials listed with high MW are likely proteins.

<sup>a</sup>Natriuretic at supraphysiological and pharmacological doses.

<sup>b</sup>Expected to have similar natriuretic activity as the bufadienolides (68, 69).

<sup>c</sup>Not likely to circulate in significant amounts.

<sup>d</sup>LLU-alpha; 2,7,8-trimethyl-2-(pcarboxyethyl)-6-hydroxychroman.

<sup>e</sup>Immunoreactivity present in the circulation (70) but not isolated from blood. The natriuretic effect of MBG per se has not been reported but is inferred from studies with bufalin and closely related steroids (68).

were obtained. It is a significant accomplishment that numerous factors with natriuretic activity including guanylin, uroguanylin, urodilatin, LLU- $\alpha$ , xanthurenic acid, and a number of steroid sodium pump inhibitors have been isolated and identified (14, 22–27). These materials likely account for some of the bioactivity in some, but not all, studies where natriuretic activity has been demonstrated. It is less clear that any of these materials fits the physiological profile expected for a NH as will be apparent from the discussion that follows.

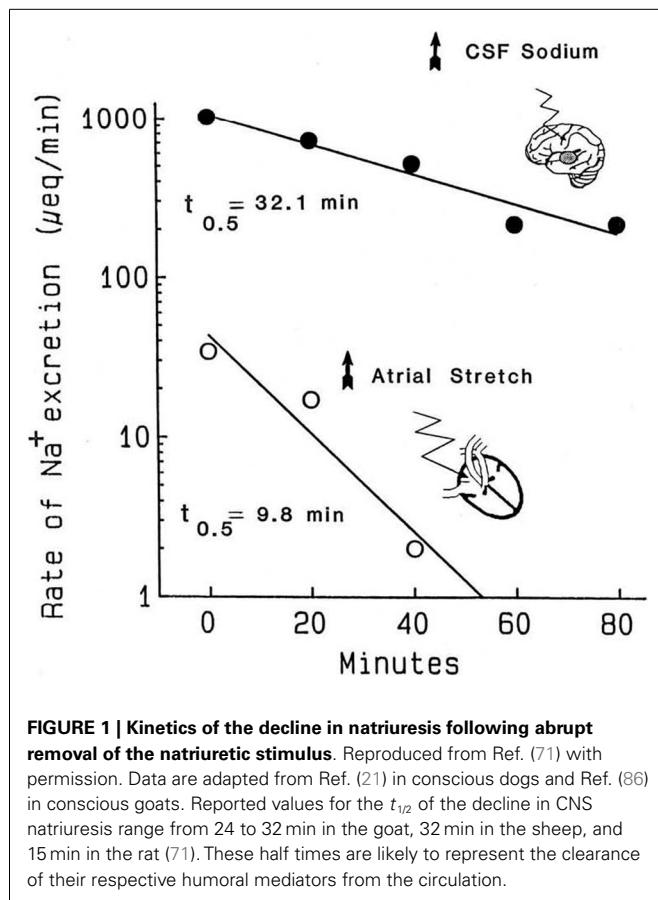
## NATRIURETIC HORMONES: HOW MANY?

Other than the APs, there are numerous hormones and endogenous materials that are known natriuretic agents. These include melanocyte stimulating hormone, dopamine, certain phospholipids, prostaglandins, kinins, and parathyroid hormone (71). These are not discussed here.

Evidence based upon pharmacological interventions, as well as an analysis of the kinetics of salt excretion mentioned below, suggests there are at least two major NH mechanisms unrelated to the APs. One mechanism is activated by the central nervous system (CNS) and the other involves maneuvers that increase atrial stretch. Pharmacological inhibition of renal NO blunts the magnitude of saline natriuresis (72) and both specific and non-selective dopamine antagonists attenuate volume expansion and water immersion (i.e., atrial stretch mediated) natriuresis but not that activated by CNS sodium (73–77). Yet another key factor that distinguishes these two NH systems is their kinetics; the rates of the decline in sodium excretion when the natriuretic stimuli are abruptly removed differ markedly for CNS- and atrial distention natriuresis. The kinetic features are potentially diagnostic; they can be used to evaluate candidate NHs.

The atrial distention arising from balloon inflation requires intact cardiac but not renal nerves, the stretch can be reversed in seconds, and the evoked natriuresis declines rapidly (21). Critically, the kinetics of the decline in natriuresis are uncontaminated by residual volume that typically would remain following a saline load (78). The second experimental paradigm is the natriuresis evoked by infusion of hypertonic saline into the brain. As the flow rates in the cerebral ventricles are much higher than the rates at which hypertonic stimuli are typically infused, simply stopping the infusion exposes the kinetics of the decline in salt excretion. Accordingly, **Figure 1** compares the decline in renal sodium excretion evoked by either atrial distension or CNS sodium. Three points are apparent: (1) the decay kinetics in both instances are first order; for CNS natriuresis, they remain linear for well over 1 h. The kinetics demonstrate that a single reaction likely is the dominant rate limiting step for the natriuresis evoked by each stimulus. (2) The CNS natriuresis, when activated by hypertonic saline (79–83), dehydration (84), or norepinephrine (85), produces similar rate constants with no major species differences. (3) The rate constants for the decline in CNS natriuresis are ~2–3-fold less (slower) than that evoked by atrial distension. Thus, the combined evidence derived from the sensitivity to pharmacological agents and the kinetic observations indicate that CNS- and atrial distension natriuresis must be mediated by different mechanisms.

Compensatory mechanisms might conceivably alter the kinetics in **Figure 1**, especially if significant salt and water loss were to occur along with declining blood pressures. During the 40 min atrial distension in **Figure 1**, blood pressure increased modestly. Plasma renin was suppressed in one set of experiments but not another. Following the distension, in one set of experiments, blood pressure remained elevated even though the natriuresis declined rapidly and aldosterone was unchanged or increased. Nevertheless, changes in aldosterone would have been too slow to have had impact. Under the conditions used, and among the measured hormonal and hemodynamic variables, the only changes



**FIGURE 1 | Kinetics of the decline in natriuresis following abrupt removal of the natriuretic stimulus.** Reproduced from Ref. (71) with permission. Data are adapted from Ref. (21) in conscious dogs and Ref. (86) in conscious goats. Reported values for the  $t_{1/2}$  of the decline in CNS natriuresis range from 24 to 32 min in the goat, 32 min in the sheep, and 15 min in the rat (71). These half times are likely to represent the clearance of their respective humoral mediators from the circulation.

convincingly associated with the decline in natriuresis following atrial distension were the return of left, right, and pulmonary pressures (i.e., cardiac nerve activity) to normal. With regard to CNS natriuresis, the decline in natriuresis is an extended first-order process; the absence of curvature over the time course implies no major influence by a compensatory process.

Among the candidate NHs in **Table 1**, there is, unfortunately, no readily interpretable information regarding the halftimes for the decline in their natriuretic effects. Most of the unidentified materials were impure, with variable onset times, and, reminiscent of urodilatin (49), some produced a natriuresis that lasted many hours following infusion. The absence of kinetic information is understandable; the primary experimental emphasis was the demonstration of natriuresis *per se*. And for decay kinetics to be informative, a near steady-state natriuresis would ideally be desirable prior to stimulus removal. This is not always an easy condition to meet. Regarding the recently identified materials in **Table 1**, no kinetic information is available. However, among all the materials, urodilatin shows a most interesting physiological correlate; in human beings, urinary urodilatin excretion closely paralleled the circadian rhythm for sodium excretion over many days (49). As urodilatin itself is not found in the circulation, it is not, by definition, an NH; although the unknown substance (?) that presumably links sodium intake with urinary urodilatin and sodium excretion could be. Thus, for all listed materials in **Table 1**, there is currently no compelling evidence that their behaviors

fit the definition of a physiologically relevant NH given in the introduction.

Hereafter, I focus primarily on CNS natriuresis and consider the potential role of sodium pump inhibitors as NHs.

### CNS Natriuresis

The brain, via an unknown humoral NH, mediates the natriuresis evoked by increased plasma sodium concentration, intracerebroventricular (icv) sodium, and dehydration (79, 81, 87). The natriuresis may be damped but is not eliminated by renal denervation (88), is activated by small increases of plasma sodium (1–2 mM). The CNS NH may have a dominant influence in post-prandial natriuresis (89) and is a blood-bourne factor distinct from APs (90, 91), ADH (92), or dopamine (74).

Central nervous system natriuresis can be activated by the elevation of either blood-bourne or cerebrospinal fluid sodium; both dehydration and postprandial natriuresis are blocked or reversed by hyponatremic CSF (93–95) or rehydration (96). Push–pull perfusion techniques suggest a discrete area of the third ventricle is near the sodium sensing apparatus (97). Further, the ablation of central structures, including the anteroventral and posterior hypothalamus in a variety of species, or decapitation, profoundly influence the ability to regulate osmotic balance, tolerate hyperosmotic challenge, and excrete sodium (80, 83, 98–106). The lesioned areas have included the median eminence, medial preoptic nucleus, organum vasculosum of the lamina terminalis, and the periventricular preoptic area. The consequences of these lesions are impaired thirst and ADH secretion, reduced renal natriuretic response, and hypernatremia. In contrast, this same system, when overactivated, can lead to profound hyponatremia. This phenomenon, sometimes termed “cerebral salt wasting,” and resembling some of the features of the syndrome of inappropriate ADH secretion, has been noted in some CNS disorders (107–111).

The observation that dehydration results in hypernatremia and provokes a compensatory natriuresis in the face of reduced extracellular fluid volumes, and that the natriuresis subsides with rehydration, suggests that the tendency to hypernatremia during dehydration and following a high salt meal is actively opposed by an unknown osmotically sensitive mechanism [see in Ref. (84)]. In each instance, the natriuretic response to these stimuli is present in animals with denervated kidneys but absent in animals with hypothalamic lesions (84, 112). Further, CNS natriuresis is not explained by blood pressure changes and persists when renal artery pressures are servo controlled (92, 113).

In each of the aforementioned situations, changes in circulating ADH have been implicated as the efferent mediator of CNS natriuresis (114, 115). Indeed, CNS natriuresis is either absent or slowed in rats congenitally deficient in AVP (116, 117), is absent in hypophysectomized rats but reappears in rats pretreated with large amounts of ADH and in rats given a dD-AVP analog (88). ADH certainly contributes to the control of sodium excretion in rats, dogs, and man (84, 89, 118–123). ADH infusions are natriuretic, and specifically implicated in CNS (114, 115) but not saline natriuresis. However, ADH is not sufficient to account for CNS natriuresis (92, 117), although it may be permissive (124, 125). For example, AV3V-lesioned sheep and dehydrated normal sheep both lost similar amounts of body water, although the hypernatremia

was much worse in the lesioned animals (112). Thus, something other than ADH was lacking in the lesioned animals to explain the greater hypernatremia with the same overall water loss.

Little is known about the chemical nature of the CNS NH other than it appears to be heat stable (126). Its actions have an interesting temporal association with ADH and/or oxytocin (92, 127). For example, plasma ADH rises during the prehypertensive period associated with mineralocorticoid escape; a period when increased CNS NH would be expected (128). Consistent with the latter supposition, urinary sodium excretion in sheep given 3–4 day infusions of aldosterone was almost entirely blocked by the acute CNS administration of a low sodium cerebrospinal fluid during mineralocorticoid escape (129). Further, mineralocorticoids also augment the osmotic sensitivity of ADH secretion (130).

Oxytocin also has a role in renal sodium excretion (131–134) and restores the ability of hypophysectomized dogs and rats to excrete sodium at a brisk rate during saline expansion (20, 135). Yet other humoral factors implicated during CNS and ADH natriuresis include an inhibitor of prostacyclin synthesis (136) and a humoral substance that inhibits active sodium transport in toad bladder (137). Hemorrhage, paradoxically, also evokes a natriuresis that depends on the CNS (118). The natriuresis is blocked when intrarenal prostaglandin synthesis is inhibited (119). The simplest interpretation is that activation of intrarenal V1 receptors stimulates prostaglandin synthesis and the resultant products influence sodium reabsorption at distal tubular sites (138). Overall, the phenomenon ascribed to CNS natriuresis has complex interdependencies and is associated with the diminution of renal tubular sodium transport.

Of significant relevance, CNS-mediated natriuresis depends upon the prevailing level of dietary sodium intake. In sodium depleted dogs, infusion of hypertonic saline into the carotid artery is not natriuretic (139). Moreover, the phenomenon of postprandial natriuresis in the sheep is activated only when dietary sodium intakes reach a threshold of 50–75 mmol of sodium/24 h (140), i.e., when plasma renin and aldosterone are largely suppressed. Thus, the CNS NH system is likely of great physiological relevance; it is appropriately integrated with other key factors that govern long-term sodium balance.

The CNS also has a permissive role in the response to saline expansion of blood volume (79, 102, 103). Hypophysectomy reduces saline natriuresis; the deficit is reversed partially by administration of oxytocin and ADH (141). Furthermore, the application of a constricting vice to the neck of anesthetized dogs so as to exclude the brain and pituitary factors from the circulation impairs saline natriuresis (102). In view of the abovementioned role of the CNS, it is surprising that remarkably little attention has been focused on the natriuretic activity associated with extracts from brain and pituitary (Table 1). The little that is known is that the bioactivity of natriuretic extracts from hypothalamus persists following treatment with thioglycollate (to exclude oxytocin or ADH), and that an unidentified tridecapeptide was found in bioactive fractions from the posterior pituitary (142, 143).

## ARE SODIUM PUMP INHIBITORS NATRIURETIC HORMONES?

There is much evidence linking sodium pump inhibitors with salt balance and cardiovascular and renal disease (144, 145). The

Na,K-ATPase inhibitory activity of plasma from normal individuals on a high sodium diet was 25 times greater than that when the individuals were on a low sodium intake (146). Further, the plasma from individuals on high sodium diets, purified natriuretic material from urine, and ouabain, all stimulated glucose-6-phosphate dehydrogenase (G6PD) activity. G6PD activity is claimed to be inversely related to Na,K-ATPase activity (147) and related to inhibition of proximal tubular Na,K-ATPase (148), although the G6PD assay is not considered a surrogate method for the Na,K-ATPase.

Nevertheless, increased blood levels of sodium pump inhibitors, as measured by traditional well-accepted means, have been repeatedly associated with acute volume expansion, high dietary salt, mineralocorticoid excess, chronic renal failure, and CNS natriuresis (31, 32, 35, 38, 149–155). Haddy and coworkers using animal models of low renin hypertension observed that sodium pump inhibition could be reproduced in normal animals given a rapid volume expansion and that this effect could be transferred to the arteries of another animal via the plasma (156). Further, in acutely saline-expanded dogs, the plasma levels of a polar Na,K-ATPase inhibitor and a digoxin immunoreactive material were elevated at a time when endogenous ouabain (EO) was unchanged (37, 157). Moreover, the plasma of dogs undergoing atrial distension strongly inhibited the ouabain-sensitive <sup>86</sup>Rb uptake into human red cells. Notably, the bioactivity of the plasma declined substantially when retested a few days later, and was undetectable after 10 days (Hamlyn and Goetz, unpublished observations). This indicates that the inhibitor is unstable in plasma and is reminiscent of the labile digoxin-like material described by Graves et al. (158). Other work implicated the CNS in the control of humoral sodium pump inhibitors; Buckalew et al. (159) found that the jugular effluent inhibited active sodium transport to a greater extent than the blood from the femoral vein. Further, increased levels of circulating sodium pump inhibitors depend upon the integrity of hypothalamic structures within the AV3V area (103, 160). Moreover, the lesion sites overlap those whose integrity is required for CNS natriuresis. Thus, the interrelationship between increased circulating sodium pump inhibitors and natriuresis continues to be of interest. When taken together, there is no doubt that the circulation contains inhibitors of sodium transport, but what are these materials, do their levels change appropriately with salt, and are they natriuretic? Below we focus on sodium pump inhibitors that have been isolated and that have been previously linked with the aforementioned criteria.

## IDENTIFICATION OF SODIUM PUMP INHIBITORS

Starting from either human plasma or urine (brain, adrenal, and the eye are not discussed here), four groups isolated sodium pump inhibitors and identified them as ouabain- (42, 161, 162), digoxin- (163), marinobufagenin- [MBG, (66)], and telocinobufagin-like steroids (162, 164), respectively. There are altered levels of these materials in numerous experimental and clinical studies (70, 164–169). All these steroids inhibit the sodium pump and, when bound, at least one evokes biased signaling in a manner strikingly reminiscent of the β-adrenergic receptor (168, 170–172). These cardiotonic steroids (CTS) typically are natriuretic and variably kaliuretic when infused acutely at pharmacological (micromolar) doses into anesthetized animals or the renal artery and, in the

case of ouabain, selectively inhibit sodium transport in the distal tubules (68, 69, 173, 174). The natriuretic response is linearly related to the inhibition of Na pumps in the dog (175). But are they physiologically relevant NHs?

## WHAT DO THE KINETICS OF THE DECLINE IN NATRIURESES TELL US ABOUT THE ROLE OF KNOWN SODIUM PUMP INHIBITORS?

By comparing the circulating half lives of any putative NH with the half times in **Figure 1**, it is possible to determine whether it is a plausible mediator of natriuresis. Here, I examine the circulatory half lives of a number of well-known sodium pump inhibitors and compare them with the information in **Figure 1**. For example, in the dog, the plasma half lives for intravenous ouabain, digoxin, resibufagenin, and bufalin were ~18 h, ~30 h, 21 min, and 25 min, respectively [Ref. (176–178)]. In the rat, the circulating half lives for intravenous cinobufagin, resibufagenin, and bufalin were 44, 42, and 25 min, respectively (179). Therefore, it is apparent that, among these known steroid sodium pump inhibitors all, with the exception of bufalin, are simply cleared too slowly from the circulation to be kinetically plausible humoral mediators of CNS natriuresis. In the case of atrial distention natriuresis, the kinetic analysis reveals that none of the abovementioned sodium pump inhibitors are likely primary humoral mediators. With regard to CNS natriuresis, only the clearance of bufalin is sufficiently fast in both dogs and rats to warrant further investigation. The kinetic analysis does not prove bufalin as the humoral mediator in CNS natriuresis, but simply suggests that this steroid (or those that are closely related but for which no clearance data are available, e.g., MBG) cannot, as yet, be excluded. A lingering concern with bufalin, or any CTS sodium pump inhibitor, as a NH is the potentially serious conceptual problem that their acute vasoconstrictive action within the renal vasculature will oppose their tubular effects (180).

## RENAL SODIUM PUMP ISOFORMS: IS THEIR OUABAIN SENSITIVITY IMPORTANT?

Nearly all mammalian tissues express the  $\alpha$ -1 catalytic subunit of the sodium pump; muscle and muscle and nerve also express sodium pump isoforms with  $\alpha$ -2 and  $\alpha$ -3 subunits (181). In the rat kidney, sodium pumps with the  $\alpha$ -1 catalytic subunit are insensitive to micromolar ouabain but are somewhat sensitive to bufalin and marinobufagenin; the acute natriuretic effect of bufalin is greater than that of ouabain (69). For many years, it was believed that the kidney expressed only the  $\alpha$ 1 isoform even though the ouabain sensitivity of the renal Na pump increases progressively along the nephron (182); the distal tubules are believed to be ~50–100-fold more sensitive than their proximal tubule counterparts. More recently, small numbers of highly ouabain-sensitive  $\alpha$ -2 sodium pumps have been detected in rat kidney and they are functionally significant. For example, in response to acute low doses of ouabain, the  $\alpha$ 2 sodium pumps trigger enhanced  $\text{Ca}^{2+}$  signaling and NO generation in the descending vasa recta (183). It is not known if these signaling effects extend to the renal epithelia, but if they do then the acute natriuretic effects of ouabain could involve short-term NO-mediated events. In contrast, the acute natriuretic effects of bufalin and

other bufadienolides are thought to be mediated by inhibition of  $\alpha$ 1 sodium pumps (184).

In the kidney, the renal ouabain-insensitive  $\alpha$ -1 sodium pumps far outnumber their ouabain-sensitive  $\alpha$ -2 cousins. Interestingly, saline natriuresis was augmented when rodent  $\alpha$ -1 sodium pumps were made highly ouabain-sensitive (185). Further, the augmented component of the natriuresis was blocked by digoxin antibody fragments (Fab). However, the kinetic analysis in **Figure 1** makes it clear that neither ouabain nor digoxin are viable mediators of atrial distention (saline) natriuresis; the digoxin Fab fragments must, therefore, have interacted with an unknown material that preferred ouabain-sensitive sodium pumps. Thus, occupation of the ouabain binding site by this material can contribute to, but does not fully account for, the phenomenon of saline natriuresis.

## OUABAIN AS A SALT RETAINING STEROID

In contrast to the well-accepted acute natriuretic effects of high doses of sodium pump inhibitors, the chronic effects of low concentrations can be diametrically opposite. In the case of ouabain, the prolonged daily administration of low nanomole amounts in the rat suppresses  $\text{Ca}^{2+}$  signaling and NO generation in the endothelium of the descending vasa recta, reduces renal blood flow and glomerular filtration, raises sympathetic nerve activity, directly augments vascular myogenic tone and contractility, and raises blood pressure (186–193). Further, chronically reduced total body sodium in human beings is associated with elevated circulating levels of EO (194, 195), i.e., the chronic relationship between plasma EO and salt intake is, like aldosterone and renin, roughly “L”-shaped (196). In addition, and as might be anticipated from the above noted chronic observations, clinical studies have shown that among salt-loaded EH patients, renal tubular sodium reabsorption was highest in the group with elevated circulating EO (197). Thus, the behavior of circulating EO under physiological circumstances, as well as its long-term vascular and renal tubular actions, all appear to favor sodium retention.

Dramatic increases in circulating EO have been reported during exercise, a state associated with increased sympathetic activity and a decline in renal blood flow (198). The circulating levels of EO rise acutely in response to the stress of cardiac surgery (199) and the preoperative plasma levels of EO enhance the identification of those patients who will develop acute kidney injury postsurgery (200). Once again, the behavior and actions of EO in these stressful situations is associated directly or indirectly with salt and water retention, rather than salt excretion. When taken together, the current evidence strongly favors the view that EO is a physiologically relevant hormone with a variety of interesting actions that augment vascular tone and promote renal sodium retention.

In summary, the hunt for NHs has led recently to the complete identification of numerous natriuretic materials. In spite of these notable successes, none of the materials seems to fit the anticipated physiological profile for a mammalian NH. Much evidence indicates there are two major non-AP NHs that remain to be isolated and identified. It may be argued that identification of the CNS NH should be a priority in view of its broad physiological relevance, relationship to dietary sodium intake, and the implication

of a profound role in salt balance in a number of pathological disorders.

## ACKNOWLEDGMENTS

I thank Mordecai P. Blaustein for helpful comments. Supported in part by the US National Institutes of Health HL107555.

## REFERENCES

1. de Wardener HE, Clarkson EM. Concept of natriuretic hormone. *Physiol Rev* (1985) **65**:658–759.
2. Brown IJ, Tzoulaki I, Candeias V, Elliott P. Salt intakes around the world: implications for public health. *Int J Epidemiol* (2009) **38**(3):791–813. doi:10.1093/ije/dyp139
3. Wechter WJ, Kantoci D, Murray ED Jr, D'Amico DC, Jung ME, Wang WH. A new endogenous natriuretic factor: LLU-alpha. *Proc Natl Acad Sci USA* (1996) **93**(12):6002–7. doi:10.1073/pnas.93.12.6002
4. Selkert EE. Sodium excretion by the mammalian kidney. *Physiol Rev* (1954) **34**:287–333.
5. Slatopolsky E, Elkan IO, Weerts C, Bricker NS. Studies on the characteristics of the control system governing sodium excretion in uremic man. *J Clin Invest* (1968) **47**:521–30. doi:10.1172/JCI105877
6. de Wardener HE, Mills IH, Clapham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intravenous saline in the dog. *Clin Sci* (1961) **21**:249–58.
7. Bahlman JS, McDonald J, Ventoni MG, de Wardener HE. The effect on urinary sodium excretion of blood volume expansion without changing the composition of the blood. *Clin Sci* (1967) **32**:403–13.
8. Tobian L, Coffee K, McCrea P. Evidence for a humoral factor of non-renal and non-adrenal origin which influences sodium excretion. *Trans Assoc Am Physicians* (1967) **80**:200–6.
9. Kaloyanides GJ, Azer M. Evidence of a humoral mechanism in volume expansion natriuresis. *J Clin Invest* (1971) **50**:1603–12. doi:10.1172/JCI106648
10. Knock CA. Further evidence in vivo for a circulating natriuretic substance after expanding the blood volume in rats. *Clin Sci* (1980) **59**:423–33.
11. Goetz KL. Physiology and pathophysiology of atrial peptides. *Am J Physiol* (1988) **254**:E1–15.
12. Goetz KL. Evidence that atriopeptin is not a physiological regulator of sodium excretion. *Hypertension* (1990) **15**:9–19. doi:10.1161/01.HYP.15.1.9
13. Rubattu S, Sciarretta S, Valenti V, Stanzione R, Volpe M. Natriuretic peptides: an update on bioactivity, potential therapeutic use, and implication in cardiovascular diseases. *Am J Hypertens* (2008) **21**(7):733–41. doi:10.1038/ajh.2008.174
14. De Bold AJ, Borenstein HB, Veress AT, Sonnenberg H. A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats. *Life Sci* (1981) **28**:89–94. doi:10.1016/0024-3205(81)90370-2
15. Richards AM, McDonald D, Fitzpatrick MA, Nicholls MG, Espiner EA, Ikram H, et al. Atrial natriuretic hormone has biological effects in man at physiological plasma concentrations. *J Clin Endocrinol Metab* (1988) **67**(6):1134–9. doi:10.1210/jcem-67-6-1134
16. Anderson JV, Struthers AD, Christofides ND, Bloom SR. Atrial natriuretic peptides: an endogenous factor enhancing sodium excretion in man. *Clin Sci* (1986) **70**:327–31.
17. Lang RE, Tholken H, Ganter D, Luft FC, Ruskoaho H, Unger T. Atrial natriuretic factor – a circulating hormone stimulated by volume loading. *Nature* (1985) **314**:264–6. doi:10.1038/314264a0
18. Richards AM, Tonolo G, Polonia J, Montorsi P. Contrasting plasma atrial natriuretic factor concentrations during comparable natriuresis with infusions of atrial natriuretic factor and saline in normal man. *Clin Sci (Lond)* (1988) **75**(5):455–62.
19. Goetz KL, Wang BC, Geer PG, Leadley RJ Jr, Reinhardt HW. Atrial stretch increases sodium excretion independently of release of atrial peptides. *Am J Physiol* (1986) **250**:R946–50.
20. Goetz KL, Wang BC, Geer PG, Sundet WD, Needelman P. Effects of atriopeptin infusion versus effects of left atrial stretch in awake dogs. *Am J Physiol* (1986) **250**:R221–6.
21. Goetz KL, Wang BC, Bie P, Leadley RJ, Geer PG. Natriuresis during atrial distension and a concurrent decline in plasma atriopeptin. *Am J Physiol* (1988) **255**:R259–67.
22. Wechter WJ, Benaksas EJ. Natriuretic hormones. *Prog Drug Res* (1990) **34**:231–60.
23. De Bold AJ. On the shoulders of giants: the discovery of atrial natriuretic factor. *Can J Physiol Pharmacol* (1987) **65**(10):2007–12. doi:10.1139/y87-314
24. Schulz-Knappe P, Forssmann K, Herbst F, Hock D, Pipkorn R, Forssmann WG. Isolation and structural analysis of “urodilatin”, a new peptide of the cardiodilatin-(ANP)-family, extracted from human urine. *Klin Wochenschr* (1988) **66**(17):752–9. doi:10.1007/BF01726570
25. Cain CD, Schroeder FC, Shankel SW, Mitchnick M, Schmertzler M, Bricker NS. Identification of xanthurenic acid 8-O-beta-D-glucoside and xanthurenic acid 8-O-sulfate as human natriuretic hormones. *Proc Natl Acad Sci U S A* (2007) **104**(45):17873–8. doi:10.1073/pnas.0705553104
26. Currie MG, Fok KF, Kato J, Moore RJ, Hamra FK, Duffin KL, et al. Guanylin: an endogenous activator of intestinal guanylate cyclase. *Proc Natl Acad Sci U S A* (1992) **89**:947–51. doi:10.1073/pnas.89.3.947
27. Hamra FK, Forte LR, Eber SL, Pidhorodeckyj NV, Krause WJ, Freeman RH, et al. Uroguanylin: structure and activity of a second endogenous peptide that stimulates intestinal guanylate cyclase. *Proc Natl Acad Sci USA* (1993) **90**:10464–8. doi:10.1073/pnas.90.22.10464
28. Veress AT, Pearce JW, Solomon S. The effect of acute adrenalectomy on volume natriuresis in the rat. *Proc Soc Exp Biol Med* (1974) **145**(2):533–6. doi:10.3181/00379727-145-37846
29. Kaloyanides GJ, Cohen L, DiBona GF. Failure of selected end organ ablation to modify the natriuresis of blood volume expansion in the dog. *Clin Sci Mol Med* (1977) **52**:351–6.
30. Schneider R, Wray V, Niemtz M, Lehmann WD, Kirch U, Antolovic R, et al. Bovine adrenals contain, in addition to ouabain, a second inhibitor of the sodium pump. *J Biol Chem* (1998) **273**(2):784–92. doi:10.1074/jbc.273.2.784
31. Kramer HJ, Gonick HC. Effect of extracellular volume expansion on renal Na,K-ATPase and cell metabolism. *Nephron* (1974) **12**:281–96. doi:10.1159/000180341
32. Kramer HJ. Natriuretic activity in plasma following extracellular volume expansion. In: Kauffman W, Krause DK, editors. *Central Nervous Control of Na<sup>+</sup> Balance—Relations to the Renin-Angiotensin System*. Stuttgart: Georg Thieme Verlag (1976). p. 126–35.
33. Sedlakova E, Lichardus B, Cort JH. Plasma saluretic activity: its nature and relation to oxytocin analogues. *Science* (1969) **164**:580–92. doi:10.1126/science.164.3879.580
34. Kramer HJ, Rietzel C, Klingmuller D, Dusing R. Further studies on the isolation and purification of a small molecular weight natriuretic hormone. In: Lichardus B, Schrier RW, Ponec J, editors. *Hormonal Regulation of Sodium Excretion*. North Holland: Elsevier (1980). p. 313–22.
35. Buckalew VM Jr, Nelson DB. Natriuretic and sodium transport inhibitory activity in plasma of volume-expanded dogs. *Kidney Int* (1974) **5**:12–22. doi:10.1038/ki.1974.2
36. Sealey JE, Kirshman JD, Laragh JH. Natriuretic activity in plasma and urine of salt-loaded man and sheep. *J Clin Invest* (1969) **48**:2210–24. doi:10.1172/JCI106187
37. Gruber KA, Whitaker JM, Buckalew VM Jr. Endogenous digitalis-like substance in plasma of volume expanded dogs. *Nature* (1980) **287**:743–5. doi:10.1038/287743a0
38. Bourgoignie JJ, Hwang KH, Espinel C, Klahr S, Bricker NS. A natriuretic factor in the serum of patients with chronic uremia. *J Clin Invest* (1972) **51**:1514–27. doi:10.1172/JCI106948
39. Bourgoignie JJ, Weisser F, Rolf D, Klahr S, Bricker NS. Demonstration of a low molecular weight natriuretic factor in uremic serum. *Trans Assoc Am Physicians* (1970) **83**:277–87.
40. Bourgoignie JJ, Klahr S, Bricker NS. Inhibition of transepithelial sodium transport in the frog skin by a low molecular weight fraction of uremic serum. *J Clin Invest* (1971) **50**:303–11. doi:10.1172/JCI106495
41. Bourgoignie JJ, Hwang KH, Ipakchi EJ, Bricker NS. The presence of a natriuretic factor in the urine of patients with chronic uremia. *J Clin Invest* (1974) **53**:1559–67. doi:10.1172/JCI107706
42. Hamlyn JM, Blaustein MP, Bova S, DuCharme DW, Harris DW, Mandel F, et al. Identification and characterization of a ouabain-like compound from human plasma. *Proc Natl Acad Sci USA* (1991) **88**(14):6259–63. doi:10.1073/pnas.88.21.9907-d
43. Kawamura A, Guo J, Itagaki Y, Bell C, Wang Y, Haupert GT Jr, et al. On the structure of endogenous ouabain. *Proc Natl Acad Sci U S A* (1999) **96**(12):6654–9. doi:10.1073/pnas.96.12.6654

44. Forte LR, Fan X, Hamra FK. Salt and water homeostasis: uroguanylin is a circulating peptide hormone with natriuretic activity. *Am J Kidney Dis* (1996) **28**(2):296–304. doi:10.1016/S0272-6386(96)90318-2
45. Cambier P, Godon JP. Role of prostaglandins in the production of natriuretic factor by the isolated rat kidney. *Ren Physiol* (1984) **7**(3):163–75.
46. Godon JP, Nizet A. Release by isolated dog kidney of a natriuretic material following saline loading. *Arch Int Physiol Biochim* (1974) **82**:309–11. doi:10.3109/13813457409070478
47. Gonick HJ, Saldanha LF. A natriuretic principle derived from kidney tissue of volume expanded rats. *J Clin Invest* (1975) **56**:247–55. doi:10.1172/JCI108087
48. Louis F, Favre H. Basal activity of the natriuretic factor extracted from the rat kidney as a function of the diet and its role in the regulation of the acute sodium balance. *Clin Sci* (1980) **58**:385–91.
49. Drummer C, Fiedler F, König A, Gerzer R. Urodilatin, a kidney-derived natriuretic factor, is excreted with a circadian rhythm and is stimulated by saline infusion in man. *J Am Soc Nephrol* (1991) **1**(9):1109–13.
50. Feller SM, Gagelmann M, Forssmann WG. Urodilatin: a newly described member of the ANP family. *Trends Pharmacol Sci* (1989) **10**(3):93–4. doi:10.1016/0165-6147(89)90199-5
51. Ivanov YI. The evidence of production or activation of a natriuretic factor in the liver. *Endocrinol Exp* (1979) **13**:195–200.
52. Milies E. A new diuretic factor of hepatic origin. *Acta Physiol Lat Am* (1959) **10**:178–93.
53. Zubiaur M, Fernandez Munoz MD, Hernando L, Lopez-Novoa JM. Role of the natriuretic hormone in the specific natriuresis induced by intraportal infusion of hypertonic saline in dogs. *Miner Electrolyte Metab* (1987) **13**:13–8.
54. Sealey JE, Laragh JH. Further studies of a natriuretic substance occurring in human urine and plasma. *Circ Res* (1971) **28**(Suppl II):II32–43. doi:10.1161/01.RES.28.5.II-32
55. Passo SS, Thornborough JR, Rothballer AB. Hepatic receptors in the control of sodium excretion in anesthetized cats. *Am J Physiol* (1973) **224**:373–5.
56. Perlmutt JH, Aziz O, Haberich FJ. A comparison of sodium excretion in response to infusion of isotonic saline into the vena porta and vena cava of conscious dogs. *Pflugers Arch* (1975) **357**:1–14. doi:10.1007/BF00584540
57. Daly JJ, Roe JW, Horrocks P. A comparison of sodium excretion following the infusion of saline into systemic and portal veins in the dog: evidence for a hepatic role in the control of sodium excretion. *Clin Sci* (1967) **33**:481–7.
58. Potkay S, Gilmore JP. Renal response to vena cava and portal venous infusions of sodium chloride in unanesthetized dogs. *Clin Sci* (1970) **39**:13–20.
59. Strandhoy JW, Williamson HE. Evidence for an hepatic role in the control of sodium excretion. *Proc Soc Exp Biol Med* (1970) **133**:419–22. doi:10.3181/00379727-133-34487
60. Clarkson EM, Raw SM, de Wardener HE. Further observations on a low-molecular weight natriuretic substance in the urine of normal man. *Kidney Int* (1979) **16**:710–21. doi:10.1038/ki.1979.187
61. Clarkson EM, Young DR, Raw SM, de Wardener HE. Chemical properties, physiological action and further separation of a low molecular weight natriuretic substance in the urine of normal man. In: Richardus B, Schrier RW, Poncet J, editors. *Hormonal Regulation of Sodium Excretion*. North Holland: Elsevier (1980). p. 333–40.
62. Fine LG, Bourgoignie JJ, Hwang KH, Bricker NS. On the influence of the natriuretic factor from patients with chronic uremia on the bioelectric properties and sodium transport of the isolated mammalian collecting tubule. *J Clin Invest* (1976) **58**(3):590–7. doi:10.1172/JCI108505
63. Clarkson EM, Raw SM, de Wardener HE. Two natriuretic substances in extracts of urine from normal man when salt-depleted and salt-loaded. *Kidney Int* (1976) **10**:381–94. doi:10.1038/ki.1976.124
64. Kruck F. Influence of humoral factors on renal tubular sodium handling. *Nephron* (1969) **6**:205–16. doi:10.1159/000179729
65. Viskoper JR, Czaczkeo JW, Schwartz N, Ullmann TD. Natriuretic activity of a substance isolated from human urine during the excretion of a salt load. *Nephron* (1971) **8**:540–8. doi:10.1159/000179959
66. Bagrov AY, Fedorova OV, Dmitrieva RI, Howald WN, Hunter AP, Kuznetsova EA, et al. Characterization of a urinary bufadienolide Na<sup>+</sup>, K<sup>+</sup>-ATPase inhibitor in patients after acute myocardial infarction. *Hypertension* (1998) **31**:1097–103. doi:10.1161/01.HYP.31.5.1097
67. Hoffman A, Okun-Gurevich M, Ovcharenko E, Goltsman I, Karram T, Cain C, et al. Renal effects of a novel endogenous natriuretic agent xanthurenic acid 8- $\alpha$ - $\beta$ -D-glucoside in rats. *Physiol Rep* (2013) **1**(6):e00155. doi:10.1002/phy2.155
68. Eliades D, Pamnani MB, Swindall BT, Haddy FJ. Effects of bufalin on renal venous outflow, urine flow and natriuresis in the anesthetized dog. *Adv Exp Med Biol* (1991) **308**:205–10. doi:10.1007/978-1-4684-6015-5\_17
69. Pamnani MB, Chen S, Bryant HJ, Schooley JF Jr, Eliades DC, Yuan CM, et al. Effects of three sodium-potassium adenosine triphosphatase inhibitors. *Hypertension* (1991) **18**(3):316–24. doi:10.1161/01.HYP.18.3.316
70. Bagrov AY, Shapiro JI, Fedorova OV. Endogenous cardiotonic steroids: physiology, pharmacology, and novel therapeutic targets. *Pharmacol Rev* (2009) **61**(1):9–38. doi:10.1124/pr.108.000711
71. Hamlyn JM, Ludens JH. Nonatrial natriuretic hormones. In: Seldin DW, Giebisch G, editors. *The Kidney, Physiology and Pathophysiology*. New York, NY: Raven Press (1992). p. 1885–924.
72. Alberola A, Pinilla JM, Quesada T, Romero JC, Salom MG, Salazar FJ. Role of nitric oxide in mediating renal response to volume expansion. *Hypertension* (1992) **19**(Pt 2):780–4. doi:10.1161/01.HYP.19.6.780
73. Coruzzi P, Novarini A, Musiari L, Ravanetti C, Ghelmi S, Rodella A, et al. The antinatriuretic effect of dopaminergic blockade during volume expansion is independent of circulating atrial natriuretic factor. *Clin Sci (Lond)* (1989) **77**(5):479–84.
74. Hansell P, Sjoquist M, Fasching A, Isaksson B, Karlsson M, Ulfendahl HR. CNS-induced natriuresis during dopamine receptor blockade. Further support for the existence of, at least, two separate natriuretic hormonal systems. *Acta Physiol Scand* (1988) **133**:373–80. doi:10.1111/j.1748-1716.1988.tb08419.x
75. Jose PA, Holloway RR, Campbell TW, Eisner GM. Dopamine blockade attenuates the natriuresis of saline loading in the adrenalectomized rat. *Nephron* (1988) **48**:54–7. doi:10.1159/000184869
76. McClanahan M, Sowers JR, Beck FWJ, Mohanty PK, McKenzie T. Dopaminergic regulation of natriuretic response to acute volume expansion in dogs. *Clin Sci* (1985) **68**:263–9.
77. Hegde S, Jadhav AL, Lokhandwala MF. Role of kidney dopamine in the natriuretic response to volume expansion in rats. *Hypertension* (1989) **13**:828–34. doi:10.1161/01.HYP.13.6.828
78. Buckalew VM Jr, Adkins TG. Effects of interrupting intravenous saline infusion on sodium excretion in the dog. *Ren Physiol* (1978) **1**:11–8.
79. Andersson B, Jobin M, Olsson K. Stimulation of urinary salt excretion following injections of hypertonic NaCl solution into the 3rd brain ventricle. *Acta Physiol Scand* (1966) **67**:127–8. doi:10.1111/j.1748-1716.1966.tb03293.x
80. Andersson B, Dallman MF, Olsson K. Evidence for a hypothalamic control of renal sodium excretion. *Acta Physiol Scand* (1969) **75**:496–510. doi:10.1111/j.1748-1716.1969.tb04403.x
81. Andersson B, Westbye O. Synergistic action of sodium and angiotension on brain mechanisms controlling fluid balance. *Life Sci* (1970) **9**:601–8. doi:10.1016/0024-3205(70)90090-1
82. Blaine E, Denton DA, McKinley MJ, Weller S. A central osmosensitive receptor for renal sodium excretion. *J Physiol* (1975) **244**:497–509.
83. Morris M, McCann SM, Orias R. Evidence for hormonal participation in the natriuretic and kaliuretic responses to intraventricular hypertonic saline and norepinephrine. *Proc Soc Exp Biol Med* (1976) **152**:95–8. doi:10.3181/00379727-152-39336
84. McKinley MJ, Denton DA, Coghlan JP, Harvey RB, McDougall JG, Rundgren M, et al. Cerebral osmoregulation of renal sodium excretion—a response analogous to thirst and vasopressin release. *Can J Physiol Pharmacol* (1987) **65**:1724–9. doi:10.1139/y87-271
85. Thornborough JR, Passo SS, Rothballer AB. Receptors in cerebral circulation affecting sodium excretion in the cat. *Am J Physiol* (1973) **225**(1):138–41.
86. Andersson B, Jobin M, Olsson K. A study of thirst and other effects of an increased sodium concentration in the 3rd brain ventricle. *Acta Physiol Scand* (1967) **69**(1):29–36. doi:10.1111/j.1748-1716.1967.tb03488.x
87. Mouw DR, Vander AJ. Evidence for brain Na receptors controlling renal Na excretion and plasma renin activity. *Am J Physiol* (1970) **219**:822–32.
88. Ulfendahl HR, Goransson A, Hansell P, Karlsson M, Sjoquist M. Natriuresis obtained by stimulation of the cerebroventricular system with sodium ions indicates a blood borne natriuretic factor. *Acta Physiol Scand* (1986) **127**:269–71. doi:10.1111/j.1748-1716.1986.tb07904.x
89. Saville MA, Geer PG, Wang BC, Leadley RJ, Goetz KL. A high-salt meal produces natriuresis in humans without elevating plasma atriopeptin. *Proc Soc Exp Biol Med* (1988) **188**:387–93. doi:10.3181/00379727-188-3-RC2

90. Hansell P, Goransson A, Leppaluoto J, Arjamaa O, Vakkurri O, Ulfendahl HR. CNS-induced natriuresis is not mediated by the atrial natriuretic factor. *Acta Physiol Scand* (1987) **129**:221–7. doi:10.1111/j.1748-1716.1987.tb08062.x
91. Hansell P. Natriuretic factors. Influence on glomerular filtration, renal blood flow and electrolyte excretion in the rat. *Acta Univ Ups* (1988) **154**:1–29.
92. Ulfendahl HR, Ahlsson A, Hansell P, Hoglund U, Jacobsson E, Lee SL, et al. Studies on the mechanisms underlying CNS-induced natriuresis. *Acta Physiol Scand* (1989) **136**(Suppl 583):75–8.
93. Leksell LG, Congiu M, Denton DA, Fei DT, McKinley MJ, Tarjan E, et al. Influence of mannitol-induced reduction in CSF Na on nervous and endocrine mechanisms involved in the control of fluid balance. *Acta Physiol Scand* (1981) **112**(1):33–40. doi:10.1111/j.1748-1716.1981.tb06779.x
94. Leksell LG, Denton DA, Fei DT, McKinley MJ, Müller AF, Weisinger RS, et al. On the importance of CSF Na in the regulation of renal sodium excretion and renin release. *Acta Physiol Scand* (1982) **115**(1):141–6. doi:10.1111/j.1748-1716.1982.tb07056.x
95. McKinley MJ, Denton DA, Fryday HW, Weisinger RS. Cerebral mechanisms influencing renal sodium excretion in dehydrated sheep. *Clin Exp Pharmacol Physiol* (1983) **10**:521–6. doi:10.1111/j.1440-1681.1983.tb00220.x
96. McKinley MJ, Evered MD, Mathai ML. Renal Na excretion in dehydrated and rehydrated adrenalectomized sheep maintained with aldosterone. *Am J Physiol Regul Integr Comp Physiol* (2000) **279**(1):R17–24.
97. Cox PS, Denton DA, Mouw DR, Tarjan E. Natriuresis induced by localized perfusion within the third cerebral ventricle of sheep. *Am J Physiol* (1987) **252**:R1–6.
98. Cort JH, Lichardus B. The natriuretic activity of jugular vein blood during carotid occlusion. *Physiol Bohemoslov* (1963) **12**:497–501.
99. Andersson B, Lishajko F, Leskell LG. Perturbations in fluid balance induced by medially placed forebrain lesions. *Brain Res* (1975) **99**:261–75. doi:10.1016/0006-8993(75)90028-1
100. Keeler R. Effect of chronic preoptic lesions on the renal excretion of sodium in rats. *Am J Physiol* (1975) **228**:1725–8.
101. Johnson AK, Buggy J. Periventricular preoptic-hypothalamus is vital for thirst and normal water economy. *Am J Physiol* (1978) **234**:R122–9.
102. Kaloyanides GJ, Balabanian MB, Bowman RL. Evidence that the brain participates in the humoral natriuretic mechanism of blood volume expansion in the dog. *J Clin Invest* (1978) **62**:1288–95. doi:10.1172/JCI109249
103. Bealer SL, Haywoood JR, Gruber KA, Buckalew VM Jr, Fink GD, Brody MJ. Preoptic-hypothalamic periventricular lesions reduce natriuresis to volume expansion. *Am J Physiol* (1983) **244**:R51–7.
104. Mangiapane ML, Thrasher TN, Keil LC, Simpson JB, Ganong WF. Deficits in drinking and vasopressin secretion after lesions of the nucleus medianus. *Neuroendocrinology* (1983) **37**:73–7. doi:10.1159/000123518
105. McKinley MJ, Congiu M, Denton DA, Park RG, Penschow J, Simpson JB, et al. The anterior wall of the third cerebral ventricle and homeostatic responses to dehydration. *J Physiol* (1984) **79**:421–7.
106. McKinley MJ, Lichardus B, McDougall JG, Weisinger RS. Periventricular lesions block natriuresis to hypertonic but not isotonic NaCl loads. *Am J Physiol* (1992) **262**(1 Pt 2):F98–107.
107. Peters JP, Welt LG, Simms EAH. A salt wasting syndrome associated with cerebral disease. *Trans Assoc Am Physicians* (1950) **63**:57–64.
108. Cort JH. Cerebral salt wasting. *Lancet* (1954) **1**:1752–4. doi:10.1016/S0140-6736(54)92715-4
109. Nelson PB, Seif SM, Maroon JC, Robinson AG. Hyponatremia in intracranial disease: perhaps not the syndrome of inappropriate secretion of antidiuretic hormone (SIADH). *J Neurosurg* (1981) **55**:938–41. doi:10.3171/jns.1981.55.6.0938
110. Nelson PB, Seif SM, Gutal J, Robinson AG. Hyponatremia and natriuresis following subarachnoid hemorrhage in a monkey model. *J Neurosurg* (1984) **60**:233–7. doi:10.3171/jns.1984.60.2.0233
111. Diringer M, Ladenson PW, Borel C, Hart GK, Kirsch JR, Hanley DF. Sodium and water regulation in a patient with cerebral salt wasting. *Arch Neurol* (1989) **46**:928–30. doi:10.1001/archneur.1989.00520440124031
112. McKinley MJ, Denton DA, Park RG, Weisinger RS. Cerebral involvement in dehydration-induced natriuresis. *Brain Res* (1983) **263**:340–3. doi:10.1016/0006-8993(83)90326-8
113. Mouw DR, Vander AJ, Bourgoignie JJ, Kutschinski SS, Mathias NP. Nonpressor mechanisms in CNS-induced natriuresis. *Am J Physiol* (1979) **237**:F157–66.
114. Luke RG. Natriuresis and chloruresis during hydropenia in the rat. *Am J Physiol* (1973) **224**:13–20.
115. Pierce ET, Grekin RJ, Mouw DR. Efferent role of ADH in CNS-induced natriuresis. *Am J Physiol* (1984) **246**:F32–8.
116. Mouw DR, Vander AJ, Landis C, Kutschinski S, Mathias N, Zimmerman D. Dose-response relation of CSF sodium and renal sodium excretion, and its absence in homozygous Brattleboro rats. *Neuroendocrinology* (1980) **30**(4):206–12. doi:10.1159/000123002
117. Beasley D, Malvin RL, Mouw DR. CNS-induced natriuresis and renal hemodynamics in conscious rats. *Am J Physiol* (1983) **245**:F763–71.
118. Humphreys MH, Friedler RM, Earley LE. Natriuresis produced by vasopressin or hemorrhage during water diuresis in the dog. *Am J Physiol* (1970) **219**:658–64.
119. Fejes-Toth G, Magyar A, Walter J. Renal response to vasopressin after inhibition of prostaglandin synthesis. *Am J Physiol* (1977) **232**:F416–23.
120. Balment RJ, Brimble MJ, Forsling ML, Musabayane CJ. Natriuretic response of the rat to plasma concentrations of arginine vasopressin within the physiological range. *J Physiol* (1984) **352**:517–26.
121. Thrasher TN, Wade CE, Keil LC, Ramsay DJ. Sodium balance and aldosterone during dehydration and rehydration in the dog. *Am J Physiol* (1984) **247**:R76–83.
122. Lote CJ, McVicar AJ, Smyth DG. Effects of vasopressin-glycine and vasopressin-glycine-lysine-arginine on renal function in the rat. *J Endocrinol* (1986) **108**:255–60. doi:10.1677/joe.0.1080255
123. Lote CJ, Thewles A, Wood JA. Vasopressin-induced natriuresis in the conscious rat: role of blood pressure, renal prostaglandin synthesis and the peptide. *Am J Physiol* (1989) **411**:481–91.
124. Park RG, Congiu M, Denton DA, McKinley MJ. Natriuresis induced by arginine vasopressin infusion in sheep. *Am J Physiol* (1985) **249**:F799–805.
125. Merrill DC, Skelton MM, Cowley AW. Humoral control of water and electrolyte excretion during water restriction in sodium-deprived dogs. *Kidney Int* (1986) **29**:1152–61. doi:10.1038/ki.1986.121
126. Beasley D, Malvin RL. Role of natriuretic factor in central nervous system (CNS)-induced natriuresis. *Proc Soc Exp Biol Med* (1985) **178**:575–9. doi:10.3181/00379727-178-42044
127. Balment RJ, Brimble MJ, Forsling ML, Kelly LP, Musabayane CT. A synergistic effect of oxytocin and vasopressin on sodium excretion in the neurohypophysectomized rat. *J Physiol* (1986) **381**:453–64.
128. Haller H, Bähr V, Bock A, Distler A, Philipp T. Vasopressin is increased in mineralocorticoid-induced blood pressure increase in man. *J Hypertens* (1987) **5**(5):111–3.
129. Pennington GL, McKinley MJ. Reduction of cerebral NaCl concentration can abolish mineralocorticoid escape. *Am J Physiol* (1990) **259**(5 Pt 2):F839–46.
130. Brooks DP, Share L, Crofton JT, Guthe C, Ling WD, Bohr DF. Increased sensitivity of the osmotic control of vasopressin in sheep with deoxycorticosterone acetate-induced hypertension. *J Endocrinol* (1985) **107**(3):309–15. doi:10.1677/joe.0.1070309
131. Brooks FP, Pickford M. The effect of posterior pituitary hormones on the excretion of electrolytes in dogs. *J Physiol* (1958) **142**:468–93.
132. Sawyer WH. Posterior pituitary extracts and excretion of electrolytes by the rat. *Am J Physiol* (1952) **169**:583–7.
133. Chan WY, Sawyer WH. Saluretic action of neurohypophyseal peptides in conscious dogs. *Am J Physiol* (1961) **201**:799–803.
134. Barter FC, Mills IH. The effect of oxytocin on sodium excretion. *J Endocrinol* (1969) **45**(1):v–vi.
135. Kleinman LI, Banks RO. Natriuretic effect of oxytocin in saline-expanded neonatal dogs. *Am J Physiol* (1980) **239**:F589–94.
136. Diez J, Colina I, Guarner F, Quiroga J, Corzo J, Purroy A, et al. Intracerebroventricular infusion of sodium chloride-rich artificial cerebrospinal fluid in rats induces natriuresis and releases an inhibitor of prostaglandin synthesis. *Clin Sci* (1984) **66**:621–4.
137. Buckalew VM Jr, Dimond KA. Effect of vasopressin on sodium excretion and plasma antinatriferic activity in the dog. *Am J Physiol* (1976) **231**:28–33.
138. Schwartzman M, Ferreri NR, Carroll MA, Songu-Mize E, McGiff JC. Renal cytochrome P450-related arachidonate metabolite inhibits Na,K-ATPase. *Nature* (1985) **314**:620–2. doi:10.1038/314620a0

139. Gilmore JP, Nemeh MN. Salt depletion inhibits cerebral-induced natriuresis in the dog. *Am J Physiol* (1984) **247**:F725–8.
140. Grim CE, Scoggins BA. The rapid adjustment of renal sodium excretion to changes in dietary sodium intake in sheep. *Life Sci* (1986) **39**:215–22. doi:10.1016/0024-3205(86)90533-3
141. Lichardus B, Ponec J. On the role of the hypophysis in the mechanism of body fluid volume regulation in acutely hypophysectomized rats. *Endocrinologie* (1973) **61**:403–12.
142. Clarkson EM, Koutsaimanis KG, Davidman M, DuBois M, Penn WP, deWardener HE. The effects of brain extracts on urinary sodium excretion of the rat and the intracellular sodium concentration of renal tubule fragments. *Clin Sci Mol Med* (1974) **41**:210–3.
143. Sedlakova E, Prusik Z, Skopkova J, Barth T, Kluh I, Cort JH. Isolation of a tridecapeptide from natriuretic fractions of bovine posterior pituitary. *Eur J Clin Invest* (1974) **4**:285–92. doi:10.1111/j.1365-2362.1974.tb00405.x
144. Goto A, Yamada K, Yagi N, Yoshioka M, Sugimoto T. Physiology and pharmacology of endogenous digitalis-like factors. *Pharmacol Rev* (1992) **44**(3):377–99.
145. Schoner W. Endogenous digitalis-like factors. *Clin Exp Hypertens A* (1992) **14**(5):767–814. doi:10.3109/10641969209036220
146. de Wardener HE, MacGregor GA, Clarkson EM, Alaghband-Zadeh J, Bitensky L, Chayen J. Effect of sodium intake on ability of human plasma to inhibit renal Na,K-ATPase. *Lancet* (1981) **i**:411–2. doi:10.1016/S0140-6736(81)91792-X
147. Dikstein S. Stimulability, adenosine triphosphatases, and their control by cellular redox processes. *Naturwissenschaften* (1971) **58**:439–43. doi:10.1007/BF00624617
148. Fenton S, Clarkson E, MacGregor G, Alaghband-Zadeh J, de Wardener HE. An assay of the capacity of biological fluids to stimulate renal glucose-6-phosphate dehydrogenase activity in vitro as a marker of their ability to inhibit sodium potassium-dependent adenosine triphosphatase activity. *J Endocrinol* (1982) **94**(1):99–110. doi:10.1677/joe.0.0940099
149. Lichardus B, Ponec J. Conditions for biological evidence of a natriuretic hormone in experiments with rat cross circulation. *Physiol Bohemoslov* (1970) **19**:330.
150. Nutbourne DM, Howse JD, Schrier RW, Talner LB, Ventom MG, Verroust PJ, et al. The effect of expanding the blood volume of a dog on the short circuit current across an isolated frog skin incorporated in the dogs circulation. *Clin Sci* (1970) **38**:629–48.
151. Clarkson EM, Talner LB, de Wardener HE. The effect of plasma from blood volume expanded dogs on sodium, potassium and PAH transport of renal tubule fragments. *Clin Sci* (1970) **38**:617–27.
152. Buckalew VM Jr, Lancaster CD Jr. Studies of a humoral sodium transport in normal dogs and dogs with ligation of the inferior vena cava. *Circ Res* (1971) **28–29**(Suppl II):II-44–II-52.
153. Epstein M. Cardiovascular and renal effects of head out water immersion in man. *Circ Res* (1976) **39**:619–28. doi:10.1161/01.RES.39.5.619
154. Poston L, Wilkinson SP, Sewell R, Williams R. Sodium transport during natriuresis of volume expansion. A study using peripheral leucocytes. *Clin Sci* (1982) **63**:243–5.
155. Jandhyala BS, Ansari AF. Elevation of sodium levels in the cerebral ventricles of anesthetized dogs triggers the release of an inhibitor of ouabain-sensitive sodium-potassium-ATPase into the circulation. *Clin Sci* (1986) **70**:103–10.
156. Haddy FJ. Natriuretic hormone – the missing link in low renin hypertension? *Biochem Pharmacol* (1982) **31**:3159–61. doi:10.1016/0006-2952(82)90544-5
157. Ludens JH, Clark MA, Kolbasa KP, Hamlyn JM. Digitalis-like factor and ouabain-like compound in plasma of volume-expanded dogs. *J Cardiovasc Pharmacol* (1993) **22**(Suppl 2):S38–41. doi:10.1097/00005344-199322002-00014
158. Graves SW, Markides KE, Hollenberg NK. Application of supercritical fluid chromatography to characterize a labile digitalis-like factor. *Hypertension* (2000) **36**(6):1059–64. doi:10.1161/01.HYP.36.6.1059
159. Buckalew VM Jr, Martinez FJ, Green WE. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**:926–35. doi:10.1172/JCI106312
160. Buggy J, Huot S, Pamnani M, Haddy F. Periventricular forebrain mechanisms for blood pressure regulation. *Fed Proc* (1984) **43**(1):25–31.
161. Mathews WR, DuCharme DW, Hamlyn JM, Harris DW, Mandel F, Clark MA, et al. Mass spectral characterization of an endogenous digitalislike factor from human plasma. *Hypertension* (1991) **17**(6 Pt 2):930–5. doi:10.1161/01.HYP.17.6.930
162. Komiyama Y, Nishimura N, Dong XH, Hirose S, Kosaka C, Masaki H, et al. Liquid chromatography mass spectrometric analysis of ouabainlike factor in biological fluid. *Hypertens Res* (2000) **23**(Suppl):S21–7. doi:10.1291/hypres.23. Supplement\_S21
163. Goto A, Yamada K, Yagi N, Hui C, Nagoshi H, Sasabe M, et al. Digitalis-like factors from human urine. *J Cardiovasc Pharmacol* (1993) **22**(Suppl 2):S58–9. doi:10.1097/00005344-199322002-00019
164. Komiyama Y, Dong XH, Nishimura N, Masaki H, Yoshika M, Masuda M, et al. A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin Biochem* (2005) **38**(1):36–45. doi:10.1016/j.clinbiochem.2004.08.005
165. Komiyama Y, Nishimura N, Munakata M, Okuda K, Nishino N, Kosaka C, et al. Increases in plasma ouabainlike immunoreactivity during surgical extirpation of pheochromocytoma. *Hypertens Res* (1999) **22**(2):135–9. doi:10.1291/hypres.22.135
166. Ferrandi M, Manunta P, Ferrari P, Bianchi G. The endogenous ouabain: molecular basis of its role in hypertension and cardiovascular complications. *Front Biosci* (2005) **10**:2472–7. doi:10.2741/1711
167. Huang BS, Amin MS, Leenen FH. The central role of the brain in salt-sensitive hypertension. *Curr Opin Cardiol* (2006) **21**(4):295–304. doi:10.1097/0000231398.64362.94
168. Schoner W, Scheiner-Bobis G. Endogenous and exogenous cardiac glycosides: their roles in hypertension, salt metabolism, and cell growth. *Am J Physiol Cell Physiol* (2007) **293**(2):C509–36. doi:10.1152/ajpcell.00098.2007
169. Manunta P, Ferrandi M, Bianchi G, Hamlyn JM. Endogenous ouabain in cardiovascular function and disease. *J Hypertens* (2009) **27**(1):9–18. doi:10.1097/HJH.0b013e32831cf2c6
170. Xie Z, Askari A. Na(+)/K(+) -ATPase as a signal transducer. *Eur J Biochem* (2002) **269**(10):2434–9. doi:10.1046/j.1432-1033.2002.02910.x
171. Drake MT, Violin JD, Whalen EJ, Wisler JW, Shenoy SK, Lefkowitz RJ. Beta-arrestin-biased agonism at the beta2-adrenergic receptor. *J Biol Chem* (2008) **283**(9):5669–76. doi:10.1074/jbc.M708118200
172. Zulian A, Linde CI, Pulina MV, Baryshnikov SG, Papparella I, Hamlyn JM, et al. Activation of c-SRC underlies the differential effects of ouabain and digoxin on Ca(2+) signaling in arterial smooth muscle cells. *Am J Physiol Cell Physiol* (2013) **304**(4):C324–33. doi:10.1152/ajpcell.00337.2012
173. Wilde WS, Howard PJ. Renal tubular action of ouabain on Na and K transport during stop-flow and slow-flow technique. *J Pharmacol Exp Ther* (1960) **130**:232–8.
174. Yates NA, McDougall JG. Effects of direct renal arterial infusion of bufalin and ouabain in conscious sheep. *Br J Pharmacol* (1993) **108**(3):627–30. doi:10.1111/j.1476-5381.1993.tb12852.x
175. Hook JB. A positive correlation between natriuresis and inhibition of renal Na, K-adenosine triphosphatase by ouabain. *Proc Soc Exp Biol Med* (1969) **13**:731–4. doi:10.3181/00379727-131-33963
176. Selden R, Smith TW. Ouabain pharmacokinetics in dog and man: determination by radioimmunoassay. *Circulation* (1972) **45**:1176–82. doi:10.1161/01.CIR.45.6.1176
177. Butler VP Jr, Schmidt DH, Smith TW, Haber E, Raynor BD, Demartini P. Effects of sheep digoxin-specific antibodies and their Fab fragments on digoxin pharmacokinetics in dogs. *J Clin Invest* (1977) **59**(2):345–59. doi:10.1172/JCI108647
178. Cao Y, Zhao L, Liang Q, Bi K, Wang Y, Luo G. Study of the determination and pharmacokinetics of bufadienolides in dog's plasma after administration of Liu-Shen-Wan by high performance liquid chromatography time-of-flight mass spectrometry. *J Chromatogr B Analys Technol Biomed Life Sci* (2007) **853**(1–2):227–33. doi:10.1016/j.jchromb.2007.03.018
179. Zhang Y, Tang X, Liu X, Li F, Lin X. Simultaneous determination of three bufadienolides in rat plasma after intravenous administration of bufadienolides extract by ultra performance liquid chromatography electrospray ionization tandem mass spectrometry. *Anal Chim Acta* (2008) **610**(2):224–31. doi:10.1016/j.aca.2008.01.029
180. Patel AR, Kurashina T, Granger JP, Kirchner KA. Acute Na+, K+-ATPase inhibition with bufalin impairs pressure natriuresis in the rat. *Hypertension* (1996) **27**(3 Pt 2):668–71. doi:10.1161/01.HYP.27.3.668
181. Blanco G, Mercer RW. Isozymes of the Na-K-ATPase: heterogeneity in structure, diversity in function. *Am J Physiol* (1998) **275**(5 Pt 2):F633–50.
182. Doucet A, Barlet C. Evidence for differences in the sensitivity to ouabain of Na, K-ATPase along the nephrons of rabbit kidney. *J Biol Chem* (1986) **261**:993–5.

183. Pittner J, Rhinehart K, Pallone TL. Ouabain modulation of endothelial calcium signaling in descending vasa recta. *Am J Physiol Renal Physiol* (2006) **291**(4):F761–9. doi:10.1152/ajprenal.00326.2005
184. Bagrov AY, Fedorova OV. Cardenolide and bufadienolide ligands of the sodium pump. How they work together in NaCl sensitive hypertension. *Front Biosci* (2005) **10**:2250–6. doi:10.2741/1694
185. Loreaux EL, Kaul B, Lorenz JN, Lingrel JB. Ouabain-Sensitive alpha1 Na,K-ATPase enhances natriuretic response to saline load. *J Am Soc Nephrol* (2008) **19**(10):1947–54. doi:10.1681/ASN.2008020174
186. Doursout MF, Chelly JE, Liang YY, Buckley JP. The ouabain-dependent Na(+)–K+ pump and the brain renin-angiotensin system. *Clin Exp Hypertens A* (1992) **14**(3):393–411. doi:10.3109/10641969209036197
187. Yuan CM, Manunta P, Hamlyn JM, Chen S, Bohen E, Yeun J, et al. Long-term ouabain administration produces hypertension in rats. *Hypertension* (1993) **22**(2):178–87. doi:10.1161/01.HYP.22.2.178
188. Manunta P, Rogowski AC, Hamilton BP, Hamlyn JM. Ouabain-induced hypertension in the rat: relationships among plasma and tissue ouabain and blood pressure. *J Hypertens* (1994) **12**(5):549–60. doi:10.1097/00004872-199405000-00008
189. Kurashina T, Kirchner KA, Granger JP, Patel AR. Chronic sodium-potassium-ATPase inhibition with ouabain impairs renal haemodynamics and pressure natriuresis in the rat. *Clin Sci (Lond)* (1996) **91**(4):497–502.
190. Huang BS, Huang X, Harmsen E, Leenen FH. Chronic central versus peripheral ouabain, blood pressure, and sympathetic activity in rats. *Hypertension* (1994) **23**(6 Pt 2):1087–90. doi:10.1161/01.HYP.23.6.1087
191. Zhang J, Lee MY, Cavalli M, Chen L, Berra-Romani R, Balke CW, et al. Sodium pump alpha2 subunits control myogenic tone and blood pressure in mice. *J Physiol* (2005) **569**(Pt 1):243–56. doi:10.1113/jphysiol.2005.091801
192. Cao C, Payne K, Lee-Kwon W, Zhang Z, Lim SW, Hamlyn J, et al. Chronic ouabain treatment induces vasa recta endothelial dysfunction in the rat. *Am J Physiol Renal Physiol* (2009) **296**(1):F98–106. doi:10.1152/ajprenal.90429.2008
193. Blaustein MP, Leenen FH, Chen L, Golovina VA, Hamlyn JM, Pallone TL, et al. How NaCl raises blood pressure: a new paradigm for the pathogenesis of salt-dependent hypertension. *Am J Physiol Heart Circ Physiol* (2012) **302**(5):H1031–49. doi:10.1152/ajpheart.00899.2011
194. Manunta P, Hamilton BP, Hamlyn JM. Salt intake and depletion increase circulating levels of endogenous ouabain in normal men. *Am J Physiol Regul Integr Comp Physiol* (2006) **290**(3):R553–9. doi:10.1152/ajpregu.00648.2005
195. Wang JG, Staessen JA, Messaggio E, Nawrot T, Fagard R, Hamlyn JM, et al. Salt, endogenous ouabain and blood pressure interactions in the general population. *J Hypertens* (2003) **21**(8):1475–81. doi:10.1097/00004872-200308000-00010
196. Hamlyn JM, Manunta P. Endogenous ouabain: a link between sodium intake and hypertension. *Curr Hypertens Rep* (2011) **13**(1):14–20. doi:10.1007/s11906-010-0161-z
197. Manunta P, Maillard M, Tantardini C, Simonini M, Lanzani C, Citterio L, et al. Relationships among endogenous ouabain, alpha-adducin polymorphisms and renal sodium handling in primary hypertension. *J Hypertens* (2008) **26**(5):914–20. doi:10.1097/HJH.0b013e3282f5315f
198. Bauer N, Müller-Ehmsen J, Krämer U, Hambarchian N, Zobel C, Schwinger RH, et al. Ouabain-like compound changes rapidly on physical exercise in humans and dogs: effects of beta-blockade and angiotensin-converting enzyme inhibition. *Hypertension* (2005) **45**(5):1024–8. doi:10.1161/01.HYP.0000165024.47728.f7
199. Bignami E, Casamassima N, Frati E, Messaggio E, Corno L, Zangrillo A, et al. Endogenous ouabain changes rapidly during cardiac pulmonary bypass. *J Steroids Horm Sci* (2011) **S3**:002. doi:10.4172/2157-7536.S3-002
200. Bignami E, Casamassima N, Frati E, Lanzani C, Corno L, Alfieri O, et al. Preoperative endogenous ouabain predicts acute kidney injury in cardiac surgery patients. *Crit Care Med* (2013) **41**(3):744–55. doi:10.1097/CCM.0b013e3182741599

**Conflict of Interest Statement:** The Review Editor Frans H. H. Leenen declares that, despite having collaborated with the author John Hamlyn, the review process was handled objectively and no conflict of interest exists. The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Received: 29 August 2014; accepted: 10 November 2014; published online: 03 December 2014.*

*Citation: Hamlyn JM (2014) Natriuretic hormones, endogenous ouabain, and related sodium transport inhibitors. Front. Endocrinol. 5:199. doi: 10.3389/fendo.2014.00199 This article was submitted to Neuroendocrine Science, a section of the journal Frontiers in Endocrinology.*

*Copyright © 2014 Hamlyn. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*



# Natriuretic hormones in brain function

Anastasia Hodes and David Lichtstein\*

Faculty of Medicine, Department of Medical Neurobiology, Institute for Medical Research Israel-Canada, The Hebrew University of Jerusalem, Jerusalem, Israel

**Edited by:**

Harvey Craig Gonick, University of California Berkeley, USA

**Reviewed by:**

Joao Carlos Dos Reis Cardoso, University of Algarve, Portugal  
Kazuhiro Nakamura, Kyoto University, Japan

**\*Correspondence:**

David Lichtstein, Faculty of Medicine, Department of Medical Neurobiology, Institute for Medical Research Israel-Canada, The Hebrew University of Jerusalem, Ein-Kerem, Jerusalem 91120, Israel  
e-mail: davidli@ekmd.huji.ac.il

Natriuretic hormones (NH) include three groups of compounds: the natriuretic peptides (ANP, BNP and CNP), the gastrointestinal peptides (guanylin and uroguanylin), and endogenous cardiac steroids. These substances induce the kidney to excrete sodium and therefore participate in the regulation of sodium and water homeostasis, blood volume, and blood pressure (BP). In addition to their peripheral functions, these hormones act as neurotransmitters or neuromodulators in the brain. In this review, the established information on the biosynthesis, release and function of NH is discussed, with particular focus on their role in brain function. The available literature on the expression patterns of each of the NH and their receptors in the brain is summarized, followed by the evidence for their roles in modulating brain function. Although numerous open questions exist regarding this issue, the available data support the notion that NH participate in the central regulation of BP, neuroprotection, satiety, and various psychiatric conditions, including anxiety, addiction, and depressive disorders. In addition, the interactions between the different NH in the periphery and the brain are discussed.

**Keywords:** atrial natriuretic peptide, cardiac steroids, ouabain, guanylin, brain function

## INTRODUCTION

Natriuretic hormones (NH) are compounds that act in an endocrine or paracrine fashion to regulate extracellular fluid volume and blood pressure (BP) through the stimulation of sodium excretion by the kidney. Three groups of compounds fall into this broad definition: the natriuretic peptides (NP: ANP, BNP, and CNP), the guanylin peptides (GP), and the endogenous cardiac steroids (CS: ouabain, digoxin, and marinobufagenin). A large body of evidence supports the notion that in addition to their natriuretic effects, these hormones participate in numerous brain functions. Our goal is to review the established information on the biosynthesis, release, and physiological roles of NH, with particular focus on the brain. The available literature on the interactions between the different NH families in the periphery and in the brain is also addressed.

## NATRIURETIC PEPTIDES

The first demonstration of an endocrine link between the heart and kidneys came from the pioneering experiments of De Bold, which led to the discovery of atrial NP (ANP), the founding member of the family of NP. De Bold and his colleagues found that injecting rats with an atrial homogenate caused significant natriuresis and diuresis (1). Additional members of this family of peptides were purified over the course of the following years: B-type NP (BNP) (2) and C-type NP (CNP) (3). ANP, BNP, and CNP are expressed as pre-pro-hormones and are proteolytically processed to form the mature peptides. The three peptides share a similar structure consisting of two cysteine residues flanking a 17-residue disulfide-linked ring that is essential for biological activity (3). The main inducer of ANP release is atrial wall stretch (4). BNP is released from the atrium, as is ANP, but its main sources are the ventricles, where BNP is transcriptionally regulated by cardiac wall stretch resulting from volume overload (5). There are

three known NP receptors: NP receptor-A (NPR-A), or guanylyl cyclase A (GC-A), which binds ANP and BNP (6); NPR-B (GC-B), which is highly specific for CNP (7); and NPR-C. NPR-A and NPR-B are membrane-bound receptors consisting of an extracellular ligand binding domain, a single transmembrane region and an intracellular GC domain that rapidly releases cyclic guanosine monophosphate (cGMP) in response to the NP binding (8). The cGMP then acts as a second messenger that activates protein kinase-G (PKG) and subsequent cellular signaling cascades (9). A third receptor, NPR-C, contains only a short intracellular fragment and has no GC activity. The main function of NPR-C is to clear NP through receptor-mediated internalization and degradation (10).

## PHYSIOLOGICAL ROLES OF NP

Atrial natriuretic peptide has a major role in the regulation of BP. In the kidney, ANP induces natriuresis and diuresis by increasing the glomerular filtration rate (GFR) and inhibiting sodium and water reabsorption (11). ANP acts as a functional antagonist of the renin–angiotensin–aldosteron system by inhibiting renin secretion from the kidney and aldosterone production in the adrenal glands (12). It stimulates smooth muscle cell relaxation in blood vessels, causing vasorelaxation (13). It also regulates the intravascular volume by increasing endothelium permeability (14). In accordance with these effects, it was found that ANP knockout mice developed salt-sensitive hypertension (15). ANP also directly affects the heart by inhibiting cardiac hypertrophy (16). BNP activates the same receptor as ANP but its precise functional significance is not well understood. Studies on mice with targeted disruption of BNP (17) and on cultured cardiac fibroblasts (18) established BNP as an antifibrotic factor that plays a role in ventricular remodeling. Indeed, high concentrations of BNP were found in the ventricles following congestive heart failure or myocardial infarction, rendering it an important biomarker for

these conditions (19). Unlike the other two family members, CNP acts in an autocrine/paracrine fashion. Although NPR-B is present in the kidney, CNP has little natriuretic or diuretic effect, and it is a much more potent cardiovascular effector (20). CNP is produced by the endothelium and induces vasorelaxation (21). It also participates in vascular remodeling following injury (22). In addition to their cardiovascular and renal effects, NP show a wide-spread effect throughout the body (8): They act as bronchodilators and vasorelaxants in the lungs (23), elicit anti-inflammatory effects in the immune system (24), and have metabolic effects on the adipose tissue (25) and on long bones (26).

### NP IN THE BRAIN

The ANP, BNP, and CNP and their receptors are expressed in the brain, which implies a possible role for these peptides in brain function. CNP is the most abundantly present NP in the brain (27), suggesting that it acts as a neurotransmitter or neuromodulator rather than a cardiac hormone (28). Accordingly, the CNP-specific receptor – NPR-B is widely spread throughout the brain: NPR-B mRNA was detected in the cerebral cortex, the limbic area, preoptic-hypothalamic regions, motor nuclei, and the brainstem (29). ANP and BNP are also present in the brain and have interesting neuromodulatory functions. ANP expression was first found in the hypothalamus (30), which is the main source of NP in the brain (31, 32). ANP is present in neurons and glia in the cerebral cortex (33) and in the cerebellum (34). ANP was also described in neurons and fibers in the limbic area, olfactory bulb, thalamus, and striatum (31, 35–37). BNP was found in the hypothalamus (38) and cerebral cortex (33). Unlike ANP and CNP, no BNP mRNA was detected in the brain (39), suggesting the peripheral origin of this peptide. Interestingly, ANP and BNP were described in some of the circumventricular organs in the brain – the highly vasculated structures in the hypothalamus that allow endocrine communication between the periphery and CNS (40). Considerable ANP-like immunoreactivity was found in nerve fibers of the vascular organ of the lamina terminalis and the subfornical organ in rat brain (31). Neurons in the subfornical organ were shown to respond to ANP by increased cGMP production (41). Neurons of the circumventricular organs express receptors for the majority of the cardiovascular hormones (42), including NP receptors: NPR-A and NPR-B were found in the vascular organ of lamina terminalis, the subfornical organ, area postrema, and the choroid plexus (43).

### NP in central regulation of BP

The presence of NP and their receptors in the brain, and in the circumventricular organs in particular, led to the postulation that NP, either locally produced in the brain or arriving via the peripheral circulation, might affect neuronal pathways that centrally regulate BP. However, the results are inconsistent. Intracerebroventricular (i.c.v.) administration of ANP was reported to cause a decrease in BP in normal and spontaneously hypertensive conscious rats, but only at concentrations 10 times higher than the physiological level (44). Low concentrations of ANP were shown to have a depressor effect in anesthetized rats with sinoarticular denervation, leading to a decrease in BP and sympathetic outflow (45, 46). A study performed on conscious sheep showed that CNP, but not ANP, decreased BP upon i.c.v. administration (47). Numerous studies

found no change in BP upon central administration of ANP (48–52) or BNP (53). However, there are reports describing a decrease in vasopressin secretion following central ANP infusion, suggesting that ANP and vasopressin may interact to attenuate the central pressor effects of vasopressin (49, 51–54). Pretreatment of rats with i.c.v. BNP was also shown to suppress vasopressin secretion (53). In several studies it was postulated that ANP acts in the brain by partially inhibiting the angiotensin II (ANG II) pathway. ANP injection prevented the pressor effect of centrally administered ANG II (46, 51). On the behavioral level, centrally administered ANP was shown to inhibit water intake induced by ANG II or dehydration in rats (55). It was also found to attenuate salt appetite in spontaneously hypertensive rats (SHR) (48). These results suggest that ANP may not be directly involved in central regulation of BP, but rather act as a secondary modulator of other mechanisms, perhaps, similar to its peripheral effect, by counteracting to the effects of vasopressin and ANG II.

### NP in neuroprotection

Natriuretic peptides were shown to exert a neuroprotective effect in cultured cells and *in vivo*. Cortical spreading depression (CSD) is a wave of depolarization followed by transient suppression of electrical activity in the brain (56). Rats preconditioned with an evoked episode of CSD were protected from neuronal damage following cerebral ischemia (57). Wiggins and his colleagues found that an acute episode of CSD caused an elevation in ANP mRNA and peptide levels in the rat cortex. The elevation was prolonged, overlapped the time window for CSD-induced neuroprotection and accompanied by ANP-dependent activation of cGMP signaling cascades (58). Increased cGMP levels were previously implicated in the neuroprotective mechanism of CSD (59). This notion is supported by studies showing that ANP and BNP caused an elevation in cGMP levels and inhibited apoptosis of PC12 cells (60). However, there is no direct evidence of this effect in the brain. A neuroprotective effect was also demonstrated in rat retinal neurons, where ANP was shown to ameliorate NMDA-induced neurotoxicity, presumably in a dopamine-dependent manner (61). It was postulated that the ANP neuroprotective effect is mediated via the cerebral blood flow. Indeed, an increased number of ANP-immunoreactive astrocytes and other glial cells were found in the white matter surrounding an infarction area in rats (62). This neuroprotective effect may be modulated by cGMP signaling, since cGMP-phosphodiesterase inhibitor was found to have a protective effect in a focal brain injury model in rats (63). In ischemic brain edema induced in rats, intravenous (i.v.) administration of ANP proved to have a beneficial effect. At pharmacological doses, the peptide significantly suppressed the elevation of the brain's water and sodium content and reduced the area of edema, as revealed by magnetic resonance imaging (MRI) (64). ANP was beneficial even after delayed administration, and reduced brain edema when injected i.c.v. 4 h after induction of hemorrhagic brain injury in rats (65). BNP too was implicated in neuroprotection following brain injury. James and colleagues demonstrated that i.v. administration of BNP improved cerebral blood flow and reduced inflammation in brain injury models in mice, as manifested by reduced neurodegeneration and improved functional outcome (66). Although these experiments were performed using

high doses of exogenous human recombinant BNP (nesiritide), endogenous BNP may play a role in recovery from brain injury, as elevated BNP levels have been associated with this condition. Elevated plasma BNP levels were described in patients with traumatic brain injury (67, 68), stroke (69), and other brain injuries (70, 71). Elevated BNP levels were also reported in the cerebrospinal fluid (CSF) of brain trauma patients (67). These changes, however, correlated with a poor clinical outcome in trauma and stroke patients (72, 73). This may indicate an insufficiency of the endogenous neuroprotective mechanism. The mechanism of NP neuroprotection could be mediated through immunomodulation, as was demonstrated in the periphery (74). All these clinical and pre-clinical observations lead to the premise that ANP and BNP are part of an endogenous protective mechanism in the brain against injury or damage.

### **NP in behavior**

Natriuretic peptides modulate the function of the hypothalamic–pituitary–adrenal (HPA) axis and influence anxiety and addictive behavior. NP regulate the HPA-axis at several levels: ANP inhibits the release of corticotrophin (ACTH) and corticotrophin releasing hormone (CRH) (75, 76), which, in turn, stimulate ANP release, acting in a feedback loop (76). ANP also directly inhibits cortisol secretion, whereas CNP exerts the opposite effect (77). Central or peripheral administration of ANP decreased anxiety-associated behavior in rats (78). In humans, lower levels of ANP were described in patients with anxiety-related disorders, including panic disorder (79) and posttraumatic stress disorder (80), and high ANP levels were associated with lower anxiety levels in patients recovering from cardiac failure (81). Experimentally induced panic attacks were followed by an increase in plasma ANP levels, which was faster and more pronounced in panic disorder patients (79, 82). These observations suggest a therapeutic potential for ANP agonists in the treatment of anxiety-related disorders (83). Indeed, pretreatment with i.v. ANP significantly reduced the number of experimentally induced panic attacks in panic disorder patients and in healthy individuals (84, 85). The effects of ANP on anxiety are presumed to be mediated through inhibition of the HPA-axis. In healthy individuals, pretreatment with ANP was able to partially block the sympathetic activation induced by a bolus injection of CRH (86). However, further investigation is needed to fully understand the interplay between ANP and the HPA-axis.

B-type natriuretic peptide, like ANP, was found to have an anxiolytic effect (87). On the other hand, CNP enhances cortisol secretion (77) and has an anxiogenic effect in rodents and humans (88, 89). However, it is worth mentioning that high doses of CNP (up to 5 µg), were used in these experiments; at low doses (100 ng), CNP reduced anxiety-like behavior in rats (87). At doses similar to those used for ANP, CNP increased the levels of anxiety-related behavior when administered i.c.v. in rats (88). This effect was abolished by a CRH antagonist, pointing toward an HPA-axis related mechanism (89). In humans, pretreatment with CNP enhanced the emotional effect of the anxiogenic agent CCK-4, and increased the release of ACTH following this treatment (90). CNP was also shown to stimulate cortisol and prolactin release (91). All these findings indicate that CNP is a potent anxiogenic substance

that acts by stimulating the HPA-axis. It is therefore that CNP antagonists were considered in anti-anxiety therapy (83).

Atrial natriuretic peptide may modulate alcohol withdrawal-related anxiety. In alcohol-dependant patients, abrupt cessation of alcohol consumption is accompanied by an array of symptoms known as alcohol withdrawal (92). ANP is involved in some of the neurobehavioral aspects of alcohol withdrawal, including anxiety and alcohol craving (93). In mice, i.p. ANP administration attenuated anxiety-like behavior following alcohol withdrawal (94). Handling-induced convulsions resulting from withdrawal were reduced by i.c.v. infusion of ANP, whereas anti-ANP antibodies had the opposite effect (95). Consistently, NPR-A knockout mice showed increased stress-related alcohol consumption and aggravated withdrawal symptoms (96). In humans, acute alcohol consumption elevated plasma ANP levels in healthy individuals (97). In patients with alcohol-dependence, plasma ANP levels were lower during detoxification compared with those in non-drinking individuals (93). The lower levels correlated with alterations in promoter DNA methylation, which was significantly reduced as compared with that in healthy controls (98). On the emotional level, lower ANP levels were associated with increased anxiety and alcohol craving during withdrawal (93, 99). It was postulated that the mechanism of ANP involvement in withdrawal-related stress is mediated through the HPA-axis (100). However, although the HPA-axis stress response affects the patient's recovery from alcohol addiction (101) as well as relapse rate (102), cortisol and ACTH levels do not correlate with those factors affected by ANP, such as alcohol craving (102) and perceived stress (99). ANP involvement in alcohol dependence is supported by recent genetic studies. A genome-wide association study (GWAS) revealed alcohol dependence to be associated with a single-nucleotide polymorphism located in gene GATA4, which encodes a transcription factor regulating ANP (103, 104). This finding was confirmed by a candidate association study, which found GATA4 to be linked to alcohol dependence at the gene level (105). This genetic variation in GATA4 was also shown to be associated with an increased relapse rate in patients (106), and greater reactivity in the amygdala to alcohol-related images, as shown by functional MRI (107). These results suggest that NP, possibly by modifying the stress response of the HPA-axis, are involved in the pathological states of anxiety disorders and alcohol dependence.

### **FUTURE CHALLENGES**

As described above, there is evidence for the involvement of NP in several brain functions. These observations open exciting new venues for research and drug development. However, many open questions remain to be clarified. In all the cases described above, it appears that NP do not regulate brain functions directly, but rather modulate other endocrine pathways, such as ANG II in BP regulation, or the HPA-axis in anxiety-related disorders. As for their neuroprotective qualities, those could be mediated via other mechanisms activated by brain injury, such as the immune system. The intricate interactions between NP and other cellular systems need to be studied in depth. On the more basic level, although the control of NP biosynthesis and release in the periphery is well established, not much is known about locally produced NP in non-cardiac tissues, the brain in particular. Information is lacking as to

the specific cell types in the brain that produce NP, the factors regulating NP production and release, the modes of their elimination, and the neuronal signaling pathways that they affect. Electrophysiological studies are necessary to establish the effects of NP on neuronal excitation and channel activation. It is possible that NP do not directly regulate neuronal activity, but rather modulate it via their effect on glial cells. Studies on the NP effect on calcium release and neurotransmitters uptake in glial cells may help elucidate this point. Also, the interactions between NP and known neurotransmitters and their receptors may be of importance, and should be addressed.

### GUANYLIN AND UROGUANYLIN

Dietary sodium leads to increased natriuresis in an aldosterone-independent manner. This observation led to the postulation that an additional NH is released from the intestine in response to salt intake (108). Such a hormone was discovered in 1992 – the previously unknown endogenous ligand of the intestinal receptor GC-C, activated by bacterial enterotoxins (109), and termed guanylin. Guanylin was purified from rat jejunum, and it was shown to activate GC-C in T84 human intestinal cells (109). One year later, a second endogenous ligand of GC-C was purified from the urine and intestinal mucosa of opossums, and named uroguanylin (110). More recently, additional members of the family, such as lymphoguanylin and renoguanylin were identified (111, 112). GP are expressed as pre-pro-hormones and are proteolytically cleaved to produce the biologically active peptides (113). They share a similar structure two pairs of cysteine residues forming disulfide bonds in conserved positions that are essential for their biological function (114, 115). Like NP, guanylin and uroguanylin bind to a single-membrane-spanning receptor GC (116). GC-C has a similar GC-C domain architecture to GC-A and GC-B and elicit an increase in cellular cGMP (8), which may account for the similar function of the two peptide families. GC-C is mainly expressed in the intestine, but GC-C transcripts were also found in the adrenal gland, kidney, lung, reproductive system, lymphatic organs, and brain (117, 118).

### PHYSIOLOGICAL FUNCTION OF GP

Guanylin peptides are produced in the intestine after oral sodium intake and are secreted into the intestinal lumen (119). The resulting increase in enterocyte cGMP stimulates chloride and bicarbonate secretion and inhibits sodium absorption, causing greater secretion of fluids into the intestine (113). Guanylin and uroguanylin also exert long-term effects on the intestine by regulating intestinal cell proliferation (120). In the kidney, these peptides cause increased natriuresis, kaliuresis, and diuresis without changes in GFR or renal blood flow (121). The renal effects of GP are maintained in GC-C null mice (122), suggesting the existence of an additional receptor whose identity is yet to be discovered. Indeed, novel members of the receptor GC family were described in specific cell types in the olfactory system (123, 124).

### GP IN THE BRAIN

There are few reports on the expression of GP in the brain (118). Their main effect in the CNS is likely endocrine: guanylin and uroguanylin are secreted from the gut and enter the circulation,

mainly as prohormones (125, 126), and subsequently affect extra-intestinal tissues such as the kidney (127) and the brain (128). GC-C, the main intestinal receptor for GP, was found in the midbrain (129) and the hypothalamus (128).

### *Uroguanylin in satiety control*

The intestine is an important endocrine organ, secreting hormones that centrally regulate satiety and food intake (130). The intestinal hormones are vigorously studied as possible therapeutic targets for the growing public health concern regarding obesity and metabolic diseases (131). Valentino and colleagues identified uroguanylin as a novel satiety hormone and showed that food intake causes increased intestinal prouroguanylin secretion in fasting individuals and mice (128). Administration of bacterial enterotoxins (a GC-C agonist) i.v. or i.c.v. (but not orally) reduced food intake in fasting mice, and i.v. administration of anti-prouroguanylin antibodies blocked this response (128). The receptor GC-C is expressed in the mouse hypothalamus. However, uroguanylin expression was not found in this region, suggesting an endocrine rather than a paracrine mode of regulation (128). To strengthen this postulation, Valentino showed an increase in cGMP in the hypothalamus in response to treatment with prouroguanylin, suggesting that this prohormone is cleaved in the hypothalamus by an unknown enzyme to produce the active peptide form (128). Mice lacking GC-C exhibited impaired satiety that resulted in increased food intake, obesity, and metabolic syndrome. In these animals, as opposed to the normal controls, i.v. administration of bacterial enterotoxins did not reduce food intake (128). Although further validation of this new endocrine pathway is necessary, the study provides strong evidence that uroguanylin is a central mediator of food intake, and it may provide a new therapeutic target for obesity and metabolic diseases (132).

### *GP in behavior*

Guanylate cyclase C is expressed in dopaminergic neurons in the midbrain, and GC-C knockout was associated with behavioral changes in mice (129). Gong and colleagues showed the expression of the GC-C protein in the ventral tegmental area and substantia nigra compacta in mice (129). Voltage clamp recordings from mouse dopaminergic neurons revealed that guanylin and uroguanylin significantly increased the neuronal activation evoked by metabotropic and muscarinic receptor agonists. This effect was abolished by blocking PKG signaling downstream from GC-C, and it was absent in GC-C knockout mice (129). The knockout mice exhibited increased locomotor activity, high levels of novelty seeking behavior and impulsivity. Such behavior was attenuated by low doses (1 mg/kg) of amphetamine, used to treat attention deficit hyperactivity disorder (ADHD) in humans. It is widely accepted that the dopaminergic system in the midbrain is involved in the etiology of ADHD (133). The GC-C knockout mice were described by the authors as a new animal model for ADHD, as they exhibit some of the symptoms related to this condition (129, 134). However, as the behavioral changes described could mimic several human conditions, further validation of the model is needed, and evidence of the involvement of GC-C in ADHD in humans is required. This pathway can provide new therapeutic targets for diseases involving the dopamine system, such as ADHD and schizophrenia.

## FUTURE CHALLENGES

Unlike NP and CS, it appears that GP are not synthesized in the brain, but rather arrive via the circulation from the intestine. However, the link between the intestine and the brain is not clear. Which brain-derived factors, if any, induce GP release from the intestine, and what signaling pathways they regulate require further investigation. The humoral or neuronal pathways that mediate the differential endocrine and paracrine effects of GP on remote organs such as the brain and kidney need to be established. Additionally, new members of the GC receptor family have been recently described in specific sensory neurons (135). There is a strong possibility that there are additional receptors which mediate GP functions in specific brain areas.

## CARDIAC STEROIDS

Cardiac steroids are a group of compounds that bind to and inhibit  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase. These compounds, originally discovered in plants and toad skin, have been used for centuries in Eastern and Western medicine to treat cardiac failure (136). Investigation into these substances started with the search for a missing “third factor” in the regulation of sodium homeostasis, as described in the classic work by de Wardener et al. (137). Although the interest in endogenous CS as the “third factor” has subsided with the finding of the NH, these studies paved the way for the recognition of CS in mammalian tissues and circulation. Rat brain extracts were shown to inhibit  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity and ouabain binding (138–140). Consequently, ouabain (141, 142), digoxin (143), and several bufadienolides (144–148) were identified in mammalian tissues, urine and plasma. CS are considered to be produced in the adrenal cortex and hypothalamus (149, 150), although their complete synthetic pathway is unknown. CS are subdivided into cardenolides, such as ouabain and digoxin, and bufadienolides, including bufalin and marinobufagenin. All CS have a steroid nucleus with a lactone ring at position C-17, and a hydroxyl group at C-14. The 5-member- and 6-member lactone rings are essential for the biological function of the cardenolides and bufadienolides, respectively (151). The only established receptor for all CS is the catalytic  $\alpha$  subunit of the plasma membrane  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase. In addition to the inhibition of the  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase pumping function (152), the binding of CS results in the activation of signaling transduction cascades, including the Src-kinase, the MAP-kinase, and the PKC signaling pathways (153, 154).

## PHYSIOLOGICAL FUNCTION OF CS

$\text{Na}^+$ ,  $\text{K}^+$ -ATPase is an essential enzyme expressed in all mammalian cells. CS have widespread effects in different types of cells, including cardiac myocytes, smooth muscle cells, epithelial cells, and neurons (153, 154). CS play important roles in many physiological and pathological processes, among them sodium homeostasis (155), cardiac function (156), BP (157), cell growth (158), and behavior (159). CS form the link between dietary sodium intake and salt-sensitive hypertension (155). As described below, CS regulate BP and hypertension by their effects in the periphery and in the CNS. Given their presence in the brain and CSF, these substances were postulated to act as neurotransmitters or neuromodulators, and they were shown to be involved in psychiatric conditions such as depressive disorders (159). On the cellular level,

CS were found to function in cell growth and proliferation (158) as well as in cell migration (160) and they may be associated with the development of cancer (161).

## CS IN THE BRAIN

Based on their ability to inhibit ouabain binding and  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity, CS were identified in bovine hypothalamus (140), rat brain (138), and CSF from humans (162, 163). Immunohistochemical studies of mammalian brains revealed high concentrations of CS in the paraventricular nucleus and the supraoptic nucleus (164). Cultured rat hypothalamic neurons were shown to secrete CS *in vitro* (164, 165), supporting the premise that the hypothalamus is the source of endogenous brain CS. The only established CS receptor,  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase, is expressed throughout the brain. Three isoforms of this enzyme are expressed in the brain:  $\alpha 1$ ,  $\alpha 2$ , and  $\alpha 3$ . They display a complex expression pattern: neurons are the principal source of the  $\alpha 3$  isoform (166) [although some express  $\alpha 2$ , especially in the neonate (167)], whereas glial cells predominantly express  $\alpha 2$  (168). The  $\alpha 1$  isoform is expressed in all cell types, and considered a house keeping protein. The different subunit isoforms vary in their sensitivity to CS and may mediate differential functions of these substances.

## CS in central regulation of BP

It is widely accepted that excess dietary sodium is an extremely important factor in essential hypertension (169), although the mechanism by which sodium elevates BP is not clear. A large body of evidence links endogenous CS to the regulation of BP and hypertension. In patients with essential hypertension, plasma levels of ouabain and marinobufagenin were increased in about 40%, with a high correlation with BP (170–175). The plasma levels of these substances in hypertensive patients and in rats increased with sodium intake (176–178). Several animal models for hypertension showed increased circulating levels of CS (178–180). Furthermore, prolonged infusion of ouabain produced hypertension in animals (181–183), but had no effect in genetically modified ouabain-insensitive mice (183, 184). In transgenic mice, a greater natriuretic response to sodium loading was demonstrated in animals expressing a highly CS-sensitive  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase  $\alpha 1$  subunit (185). Studies on mice carrying mutations in the gene encoding  $\alpha 2$  showed that ouabain-induced elevation of BP in rodents was mediated via this isoform: reduction of the expression level of  $\alpha 2$  was associated with increased BP (186). In contrast, animals overexpressing  $\alpha 2$  were hypotensive (187). Treatment of hypertensive rats with anti-digoxin antibodies (185, 188) or anti-marinobufogenin antibodies (178) administered to rats on a high sodium intake, resulted in a marked reduction in BP. Endogenous ouabain was put forward as a putative target for the treatment of hypertension; the ouabain inhibitor rostafuroxin showed promising results in hypertensive rats (189). Studies by Leenen and colleagues indicated that CS involvement in BP regulation is partially mediated by their effect in the CNS. The first indication of brain involvement came from experiments in SHR, in which adrenalectomy did not prevent the increase in CS levels following high sodium intake (177). Lesions in the most anteroventral part of the third ventricle (AV3V) showed that this region is essential in mediating the pressor effects of increased CSF sodium concentration via endogenous

ouabain (190, 191). The effects of both acute and prolonged ouabain infusion in sodium-loaded rodents were abolished by administration of ANG II type 1 receptor blockers such as losartan (192, 193), as well as in transgenic rats with reduced brain renin-angiotensin pathway activity (194). These results pointed to the involvement of this pathway in the effect of ouabain. All of these findings led to a unifying hypothesis regarding the role of CS in sodium-induced hypertension: sodium loading increases the levels of ouabain in salt-sensitive individuals (195, 196). In addition to induction of vasoconstriction in the periphery, ouabain also acts in the brain, where it activates the renin-angiotensin pathway, causing sympathetic activation, vasoconstriction and consequently, an elevation in BP.

### **CS in depressive disorders**

Mood disorders include major depression and dysthymia, characterized by depressive episodes, and bipolar disorder (BD) marked by both depressive and manic episodes. These conditions pose a growing public health concern in the Western world. The etiology of these diseases is not completely understood. Early reports of the psychiatric effects of CS came from doctors describing a syndrome termed “foxglove frenzy” or “digitalis delirium” in patients with digitalis intoxication (197). More recently, a comprehensive hypothesis was put forward, linking brain CS levels and  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity with BD (198, 199). BD has consistently been associated with abnormalities in  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity in erythrocytes (200, 201). A significant mood-related decrease in the enzyme’s activity was found in manic BD patients (202). Furthermore,  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase density was significantly lower in BD patients than in major depressed and schizophrenic patients (159). The plasma levels of endogenous CS were found to be significantly decreased in manic individuals as compared with those in normal controls (203, 204). Conversely, the levels of these compounds were higher in the parietal cortex of BD patients (159). More recently, it was found that there is a reduction in brain  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase  $\alpha 1$  isoform expression in mice treated with the mood stabilizer lithium (205). An allelic association between BD and a  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase  $\alpha$  subunit gene (ATP1A3) was reported (206). We have demonstrated the prominent linkage to BD of six single-nucleotide polymorphisms (SNPs) in the three genes of the  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase  $\alpha$  isoforms. Haplotype analysis of the  $\alpha 2$  isoform showed the significant association of two loci haplotypes with BD (207). A genetic knockdown of the neuron-specific  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase  $\alpha 3$  isoform induced manic-like behavior in mice (208). Numerous studies have demonstrated that i.c.v. injection of ouabain induces hyperactive behavior in rats (159, 209), which could be ameliorated by administration of mood stabilizing drugs such as lithium (210). Reduction of the endogenous brain CS level by i.c.v. injection of anti-ouabain antibodies had anti-depressive effects in rats (159, 211). This was reflected by significant changes in catecholamine metabolism in the hippocampus and ventral tegmentum, two regions known to be associated with mood disorders (211). The molecular pathway underlying the CS behavioral effect is unknown. Ouabain injected i.c.v. elicited the activation of the ERK and Akt signaling pathways in the brain (212, 213), which are known to be activated via  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase. Other effects of ouabain include a reduction

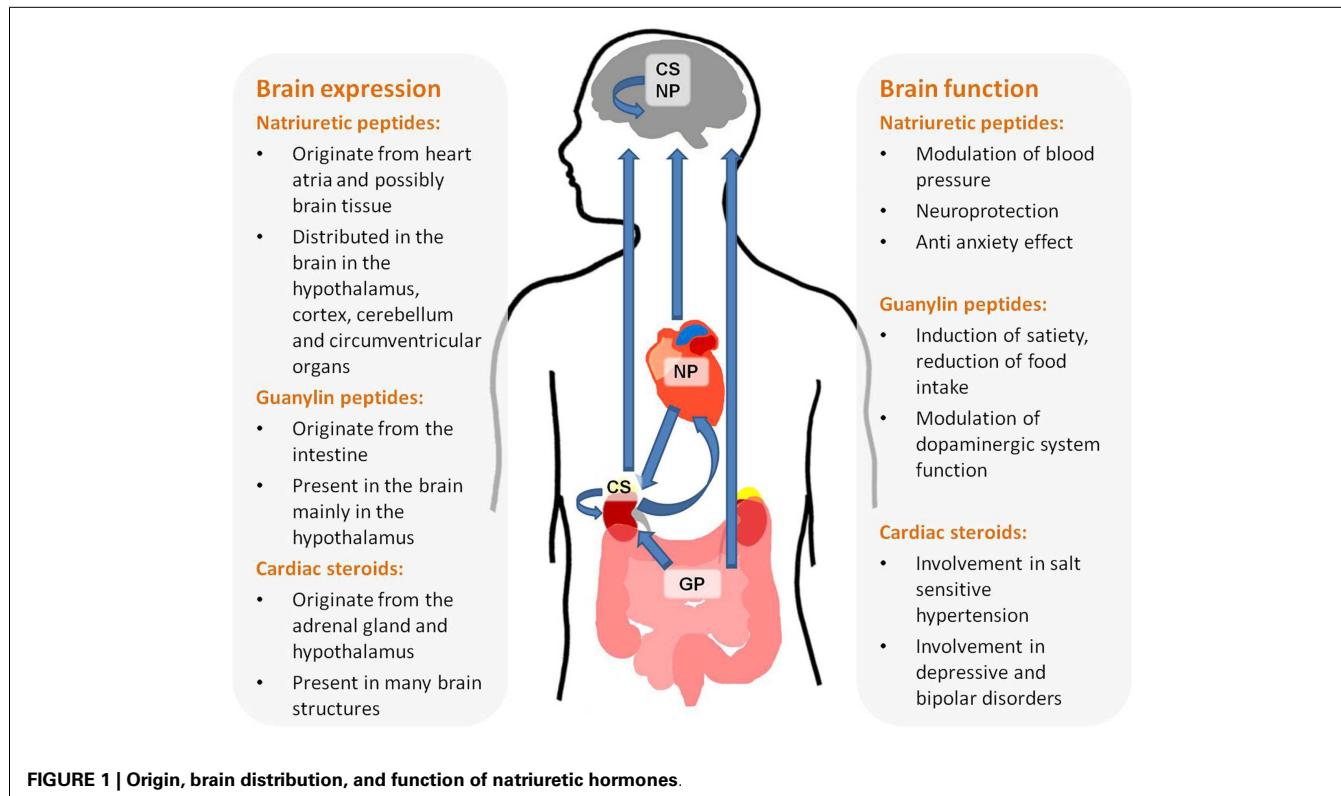
in brain-derived neurotrophic factor (BDNF) (214), activation of mammalian target of rapamycin (mTOR) signaling (213) and an increased level of oxidative stress (215). These findings strongly link the  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase and CS system to the etiology of depressive disorders, and BD in particular, and suggest their potential application in future drug development.

### **FUTURE CHALLENGES**

Despite the identification of cardenolides and bufadienolides in mammalian tissue in many independent studies (see Cardiac steroids), some still question the validity of these findings. Recently, it was claimed that ouabain, the most studied cardenolide, could not be detected in human plasma (216). This issue must be clarified. An additional major missing piece of information for the establishment of CS as neurotransmitters or neuromodulators is the elucidation of their biosynthetic pathway in the adrenal gland and brain. Although the available literature supports the notion that these steroids are synthesized in mammals, the key enzymes involved have not been identified. This issue was recently reviewed in Ref. (217). Several CS were identified in the human body. It was postulated that the different  $\alpha$  isoforms of the  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase serve as receptors for the different CS. Which of the CS are involved in brain functions, and which isoform combinations they activate are topics for future research.

### **INTERACTIONS OF ANP WITH CS AND GP**

Mutual interactions exist between CS and NP in the periphery and in the brain. Ouabain and digoxin were shown to cause increased ANP expression and secretion in rat and rabbit atria (218–221), and in anesthetized dogs (222). In patients with congestive heart failure, i.v. administration of digitalis increased the plasma levels of ANP and BNP (223). Indeed, ANP regulates the secretion of CS in the brain (224–226). ANP decreased the release of CS from rat brain extract when added to the tissue, or when administered i.v. to the animals prior to their sacrifice (224). On the other hand, another study showed that i.c.v. injection of synthetic ANP increased blood CS levels, whereas i.v. administration or incubation with this peptide had no effect (225). The effect of ANP on CS release was abolished by lesions in the AV3V region (226), the area in the hypothalamus that is thought to mediate CS central regulation of BP (191). In addition to secretion regulation, NP and CS interact at the functional level. ANP differentially modulates the effect of marinobufogenin in the rat heart and kidney (227). Ouabain was shown to antagonize the effect of ANP on vasorelaxation in rabbit aorta and in dogs (228, 229), whereas ANP abolished an ouabain-induced increase in aldosterone secretion (230). Administration of anti-ouabain antibodies caused increased sensitivity to ANP-induced vasodilation in rat aorta (231). In rat heart muscle preparations, ANP was shown to attenuate several effects of ouabain, including ouabain-induced increase in contractility,  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase and ERK phosphorylation (232). ANP also interacts with GP. Santos-Neto and colleagues showed synergy between ANP, guanylin, and uroguanylin in the kidney (233). They demonstrated in an isolated perfused rat kidney that pretreatment with ANP significantly enhanced the natriuretic, kaliuretic, and chloruretic responses to low doses of guanylin and uroguanylin (233). Low doses of ANP enhanced GP induced diuresis, and vice versa



**FIGURE 1 |** Origin, brain distribution, and function of natriuretic hormones.

(233). Since GP and NP activate GC receptors, their interaction may be mediated through the shared second messenger, cGMP (6, 116, 234). These initial studies suggest the physiological crosstalk between ANP and CS, particularly in the cardiovascular system and in the brain, and between ANP and GP in the kidney. More studies are needed to deepen our understanding of the nature of these interactions, which may be of significance in the regulation of peripheral and central functions of the NH.

## CONCLUSION

This review summarizes the available data implicating NH in brain function. There is a vast amount of data supporting the assessment that the three families of NH, NP, Guanylin, and endogenous CS are present in the brain and participate in high brain functions (see Figure 1). These include central regulation of BP, satiety, neuroprotection, and behavior. In depth research of these effects will not only increase our thorough understanding of brain function but may also lead to new treatments for brain-related diseases.

## REFERENCES

1. de Bold AJ, Borenstein HB, Veress AT, Sonnenberg H. A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats. *Life Sci* (1981) **28**:89–94. doi:10.1016/0024-3205(81)90370-2
2. Sudoh T, Kangawa K, Minamino N, Matsuo H. A new natriuretic peptide in porcine brain. *Nature* (1988) **332**:78–81. doi:10.1038/332078a0
3. Sudoh T, Minamino N, Kangawa K, Matsuo H. C-type natriuretic peptide (CNP): a new member of natriuretic peptide family identified in porcine brain. *Biochem Biophys Res Commun* (1990) **168**:863–70. doi:10.1016/0006-291X(90)92401-K
4. Edwards BS, Zimmerman RS, Schwab TR, Heublein DM, Burnett JC Jr. Atrial stretch, not pressure, is the principal determinant controlling the acute release of atrial natriuretic factor. *Circ Res* (1988) **62**:191–5. doi:10.1161/01. RES.62.2.191
5. Thuerau DJ, Hanford DS, Glembotski CC. Regulation of rat brain natriuretic peptide transcription. A potential role for GATA-related transcription factors in myocardial cell gene expression. *J Biol Chem* (1994) **269**:17772–5.
6. Suga S, Nakao K, Hosoda K, Mukoyama M, Ogawa Y, Shirakami G, et al. Receptor selectivity of natriuretic peptide family, atrial natriuretic peptide, brain natriuretic peptide, and C-type natriuretic peptide. *Endocrinology* (1992) **130**:229–39. doi:10.1210/en.130.1.229
7. Koller KJ, Lowe DG, Bennett GL, Minamino N, Kangawa K, Matsuo H, et al. Selective activation of the B natriuretic peptide receptor by C-type natriuretic peptide (CNP). *Science* (1991) **252**:120–3. doi:10.1126/science.1672777
8. Potter LR, Yoder AR, Flora DR, Antos LK, Dickey DM. Natriuretic peptides: their structures, receptors, physiologic functions and therapeutic applications. *Handb Exp Pharmacol* (2009) **191**:341–66. doi:10.1007/978-3-540-68964-5\_15
9. Tremblay J, Desjardins R, Hum D, Gutkowska J, Hamet P. Biochemistry and physiology of the natriuretic peptide receptor guanylyl cyclases. *Mol Cell Biochem* (2002) **230**:31–47. doi:10.1023/A:1014260204524
10. Matsukawa N, Grzesik WJ, Takahashi N, Pandey KN, Pang S, Yamauchi M, et al. The natriuretic peptide clearance receptor locally modulates the physiological effects of the natriuretic peptide system. *Proc Natl Acad Sci U S A* (1999) **96**:7403–8. doi:10.1073/pnas.96.13.7403
11. Marin-Grez M, Fleming JT, Steinhausen M. Atrial natriuretic peptide causes pre-glomerular vasodilatation and post-glomerular vasoconstriction in rat kidney. *Nature* (1986) **324**:473–6. doi:10.1038/324473a0
12. Richards AM, McDonald D, Fitzpatrick MA, Nicholls MG, Espiner EA, Ikram H, et al. Atrial natriuretic hormone has biological effects in man at physiological plasma concentrations. *J Clin Endocrinol Metab* (1988) **67**:1134–9. doi:10.1210/jcem-67-6-1134
13. Currie MG, Geller DM, Cole BR, Boylan JG, Yusheng W, Holmberg SW, et al. Bioactive cardiac substances: potent vasorelaxant activity in mammalian atria. *Science* (1983) **221**:71–3. doi:10.1126/science.6857267
14. Baron DA, Lofton CE, Newman WH, Currie MG. Atriopeptin inhibition of thrombin-mediated changes in the morphology and permeability of

- endothelial monolayers. *Proc Natl Acad Sci U S A* (1989) **86**:3394–8. doi:10.1073/pnas.86.9.3394
15. John SW, Kregel JH, Oliver PM, Hagaman JR, Hodgin JB, Pang SC, et al. Genetic decreases in atrial natriuretic peptide and salt-sensitive hypertension. *Science* (1995) **267**:679–81. doi:10.1126/science.7839143
  16. Holtwick R, Van Eickels M, Skryabin BV, Baba HA, Bubikat A, Begrow F, et al. Pressure-independent cardiac hypertrophy in mice with cardiomyocyte-restricted inactivation of the atrial natriuretic peptide receptor guanylyl cyclase-A. *J Clin Invest* (2003) **111**:1399–407. doi:10.1172/JCI17061
  17. Tamura N, Ogawa Y, Chusho H, Nakamura K, Nakao K, Suda M, et al. Cardiac fibrosis in mice lacking brain natriuretic peptide. *Proc Natl Acad Sci U S A* (2000) **97**:4239–44. doi:10.1073/pnas.070371497
  18. Cao L, Gardner DG. Natriuretic peptides inhibit DNA synthesis in cardiac fibroblasts. *Hypertension* (1995) **25**:227–34. doi:10.1161/01.HYP.25.2.227
  19. Nagaya N, Nishikimi T, Uematsu M, Satoh T, Kyotani S, Sakamaki F, et al. Plasma brain natriuretic peptide as a prognostic indicator in patients with primary pulmonary hypertension. *Circulation* (2000) **102**:865–70. doi:10.1161/01.CIR.102.8.865
  20. Schulz S. C-type natriuretic peptide and guanylyl cyclase B receptor. *Peptides* (2005) **26**:1024–34. doi:10.1016/j.peptides.2004.08.027
  21. Drewett JG, Fendly BM, Garbers DL, Lowe DG. Natriuretic peptide receptor-B (guanylyl cyclase-B) mediates C-type natriuretic peptide relaxation of precontracted rat aorta. *J Biol Chem* (1995) **270**:4668–74. doi:10.1074/jbc.270.9.4668
  22. Matsuo H, Furuya M. C-type natriuretic protein inhibits intimal thickening after vascular injury. *Ann N Y Acad Sci* (1997) **811**:45–7. doi:10.1111/j.1749-6632.1997.tb51987.x
  23. Hamad AM, Clayton A, Islam B, Knox AJ. Guanylyl cyclases, nitric oxide, natriuretic peptides, and airway smooth muscle function. *Am J Physiol Lung Cell Mol Physiol* (2003) **285**:L973–83. doi:10.1152/ajplung.00033.2003
  24. Morita R, Ukyo N, Furuya M, Uchiyama T, Hori T. Atrial natriuretic peptide polarizes human dendritic cells toward a Th2-promoting phenotype through its receptor guanylyl cyclase-coupled receptor A. *J Immunol* (2003) **170**:5869–75. doi:10.4049/jimmunol.170.12.5869
  25. Sengenes C, Berlan M, De Glisezinski I, Lafontan M, Galitzky J. Natriuretic peptides: a new lipolytic pathway in human adipocytes. *FASEB J* (2000) **14**:1345–51. doi:10.1096/fj.14.10.1345
  26. Chusho H, Tamura N, Ogawa Y, Yasoda A, Suda M, Miyazawa T, et al. Dwarfism and early death in mice lacking C-type natriuretic peptide. *Proc Natl Acad Sci U S A* (2001) **98**:4016–21. doi:10.1073/pnas.071389098
  27. Kaneko T, Shirakami G, Nakao K, Nagata I, Nakagawa O, Hama N, et al. C-type natriuretic peptide (CNP) is the major natriuretic peptide in human cerebrospinal fluid. *Brain Res* (1993) **612**:104–9. doi:10.1016/0006-8993(93)91649-D
  28. Komatsu Y, Nakao K, Suga S, Ogawa Y, Mukoyama M, Arai H, et al. C-type natriuretic peptide (CNP) in rats and humans. *Endocrinology* (1991) **129**:1104–6. doi:10.1210/endo-129-2-1104
  29. Herman JP, Dolgas CM, Rucker D, Langub MC Jr. Localization of natriuretic peptide-activated guanylate cyclase mRNAs in the rat brain. *J Comp Neurol* (1996) **369**:165–87. doi:10.1002/(SICI)1096-9861(19960527)369:2<165::AID-CNE1>3.0.CO;2-1
  30. Tanaka I, Misono KS, Inagami T. Atrial natriuretic factor in rat hypothalamus, atria and plasma: determination by specific radioimmunoassay. *Biochem Biophys Res Commun* (1984) **124**:663–8. doi:10.1016/0006-291X(84)91606-1
  31. Kawata M, Nakao K, Morii N, Kiso Y, Yamashita H, Imura H, et al. Atrial natriuretic polypeptide: topographical distribution in the rat brain by radioimmunoassay and immunohistochemistry. *Neuroscience* (1985) **16**:521–46. doi:10.1016/0306-4522(85)90190-3
  32. Marei HE. Fine structural and immunohistochemical localization of cardiac hormones (ANP) in the right atrium and hypothalamus of the white rat. *Eur J Morphol* (2002) **40**:37–41. doi:10.1076/ejom.40.1.0037
  33. Mckenzie JC, Berman NE, Thomas CR, Young JK, Compton LY, Cothran LN, et al. Atrial natriuretic peptide-like (ANP-LIR) and ANP prohormone immunoreactive astrocytes and neurons of human cerebral cortex. *Glia* (1994) **12**:228–43. doi:10.1002/glia.440120308
  34. Mckenzie JC, Juan YW, Thomas CR, Berman NE, Klein RM. Atrial natriuretic peptide-like immunoreactivity in neurons and astrocytes of human cerebellum and inferior olfactory complex. *J Histochem Cytochem* (2001) **49**:1453–67. doi:10.1177/002215540104901113
  35. Standaert DG, Needleman P, Saper CB. Organization of atriopeptin-like immunoreactive neurons in the central nervous system of the rat. *J Comp Neurol* (1986) **253**:315–41. doi:10.1002/cne.902530304
  36. Brown J, Czarnecki A. Autoradiographic localization of atrial and brain natriuretic peptide receptors in rat brain. *Am J Physiol* (1990) **258**:R57–63.
  37. Ryan MC, Gundlach AL. Anatomical localisation of preproatrial natriuretic peptide mRNA in the rat brain by *in situ* hybridisation histochemistry: novel identification in olfactory regions. *J Comp Neurol* (1995) **356**:168–82. doi:10.1002/cne.903560204
  38. Abdelalim EM, Takada T, Torii R, Tooyama I. Molecular cloning of BNP from heart and its immunohistochemical localization in the hypothalamus of monkey. *Peptides* (2006) **27**:1886–93. doi:10.1016/j.peptides.2006.01.001
  39. Langub MC Jr, Watson RE Jr, Herman JP. Distribution of natriuretic peptide precursor mRNAs in the rat brain. *J Comp Neurol* (1995) **356**:183–99. doi:10.1002/cne.903560205
  40. Ganong WF. Circumventricular organs: definition and role in the regulation of endocrine and autonomic function. *Clin Exp Pharmacol Physiol* (2000) **27**:422–7. doi:10.1046/j.1440-1681.2000.03259.x
  41. de Vente J, Bol JG, Steinbusch HW. cGMP-producing, atrial natriuretic factor-responding cells in the rat brain. *Eur J Neurosci* (1989) **1**:436–60. doi:10.1111/j.1460-9568.1989.tb00351.x
  42. Smith PM, Ferguson AV. Circulating signals as critical regulators of autonomic state – central roles for the subfornical organ. *Am J Physiol Regul Integr Comp Physiol* (2010) **299**:R405–15. doi:10.1152/ajpregu.00103.2010
  43. Saavedra JM, Kurihara M. Autoradiography of atrial natriuretic peptide (ANP) receptors in the rat brain. *Can J Physiol Pharmacol* (1991) **69**:1567–75. doi:10.1139/y91-233
  44. Levin ER, Weber MA, Mills S. Atrial natriuretic factor-induced vasodepression occurs through central nervous system. *Am J Physiol* (1988) **255**:H616–22.
  45. Schultz HD, Steele MK, Gardner DG. Central administration of atrial peptide decreases sympathetic outflow in rats. *Am J Physiol* (1990) **258**:R1250–6.
  46. Steele MK, Gardner DG, Xie PL, Schultz HD. Interactions between ANP and ANG II in regulating blood pressure and sympathetic outflow. *Am J Physiol* (1991) **260**:R1145–51.
  47. Charles CJ, Richards AM, Espiner EA. Central C-type natriuretic peptide but not atrial natriuretic factor lowers blood pressure and adrenocortical secretion in normal conscious sheep. *Endocrinology* (1992) **131**:1721–6. doi:10.1210/endo.131.4.1396317
  48. Itoh H, Nakao K, Katsuura G, Morii N, Shiono S, Sakamoto M, et al. Centrally infused atrial natriuretic polypeptide attenuates exaggerated salt appetite in spontaneously hypertensive rats. *Circ Res* (1986) **59**:342–7. doi:10.1161/01.RES.59.3.342
  49. Lee J, Malvin RL, Claybaugh JR, Huang BS. Atrial natriuretic factor inhibits vasopressin secretion in conscious sheep. *Proc Soc Exp Biol Med* (1987) **185**:272–6. doi:10.3181/00379727-185-42544
  50. Shoji M, Kimura T, Matsui K, Ota K, Iitake K, Inoue M, et al. Effects of centrally administered atrial natriuretic peptide on renal functions. *Acta Endocrinol (Copenh)* (1987) **115**:433–40.
  51. Al-Barazanj KA, Balment RJ. The renal and vascular effects of central angiotensin II and atrial natriuretic factor in the anaesthetized rat. *J Physiol* (1990) **423**:485–93.
  52. Stepniakowski K, Budzikowski A, Lon S, Szczepanska-Sadowska E. Central ANP attenuates pressor responses to central AVP in WKY and SHR. *Brain Res Bull* (1991) **27**:247–9. doi:10.1016/0361-9230(91)90076-V
  53. Yamada T, Nakao K, Itoh H, Shirakami G, Kangawa K, Minamino N, et al. Intracerebroventricular injection of brain natriuretic peptide inhibits vasopressin secretion in conscious rats. *Neurosci Lett* (1988) **95**:223–8. doi:10.1016/0304-3940(88)90661-1
  54. Samson WK, Aguila MC, Martinovic J, Antunes-Rodrigues J, Norris M. Hypothalamic action of atrial natriuretic factor to inhibit vasopressin secretion. *Peptides* (1987) **8**:449–54. doi:10.1016/0196-9781(87)90008-8
  55. Antunes-Rodrigues J, Mccann SM, Rogers LC, Samson WK. Atrial natriuretic factor inhibits dehydration- and angiotensin II-induced water intake in the conscious, unrestrained rat. *Proc Natl Acad Sci U S A* (1985) **82**:8720–3. doi:10.1073/pnas.82.24.8720
  56. Leao AAP. Spreading depression of activity in the cerebral cortex. *J Neurophysiol* (1944) **7**:359–90.

57. Kawahara N, Ruetzler CA, Klatzo I. Protective effect of spreading depression against neuronal damage following cardiac arrest cerebral ischaemia. *Neurol Res* (1995) **17**:9–16.
58. Wiggins AK, Shen PJ, Gundlach AL. Atrial natriuretic peptide expression is increased in rat cerebral cortex following spreading depression: possible contribution to sd-induced neuroprotection. *Neuroscience* (2003) **118**:715–26. doi:10.1016/S0306-4522(03)00006-X
59. Read SJ, Hirst WD, Upton N, Parsons AA. Cortical spreading depression produces increased cGMP levels in cortex and brain stem that is inhibited by tonabersat (SB-220453) but not sumatriptan. *Brain Res* (2001) **891**:69–77. doi:10.1016/S0006-8993(00)03191-7
60. Fiscus RR, Tu AW, Chew SB. Natriuretic peptides inhibit apoptosis and prolong the survival of serum-deprived PC12 cells. *Neuroreport* (2001) **12**:185–9. doi:10.1097/00001756-200102120-00003
61. Kurabayashi K, Kitaoka Y, Kumai T, Munemasa Y, Isenoumi K, Motoki M, et al. Neuroprotective effect of atrial natriuretic peptide against NMDA-induced neurotoxicity in the rat retina. *Brain Res* (2006) **1071**:34–41. doi:10.1016/j.brainres.2005.11.068
62. Nogami M, Shiga J, Takatsu A, Endo N, Ishiyama I. Immunohistochemistry of atrial natriuretic peptide in brain infarction. *Histochem J* (2001) **33**:87–90. doi:10.1023/A:1017996113871
63. Pifarre P, Prado J, Giralt M, Molinero A, Hidalgo J, Garcia A. Cyclic GMP phosphodiesterase inhibition alters the glial inflammatory response, reduces oxidative stress and cell death and increases angiogenesis following focal brain injury. *J Neurochem* (2010) **112**:807–17. doi:10.1111/j.1471-4159.2009.06518.x
64. Naruse S, Aoki Y, Takei R, Horikawa Y, Ueda S. Effects of atrial natriuretic peptide on ischemic brain edema in rats evaluated by proton magnetic resonance method. *Stroke* (1991) **22**:61–5. doi:10.1161/01.STR.22.1.61
65. Rosenberg GA, Estrada EY. Atrial natriuretic peptide blocks hemorrhagic brain edema after 4-hour delay in rats. *Stroke* (1995) **26**:874–7. doi:10.1161/01.STR.26.5.874
66. James ML, Wang H, Venkatraman T, Song P, Lascola CD, Laskowitz DT. Brain natriuretic peptide improves long-term functional recovery after acute CNS injury in mice. *J Neurotrauma* (2010) **27**:217–28. doi:10.1089/neu.2009.1022
67. Kirchhoff C, Stegmaier J, Bogner V, Buhmann S, Mussack T, Kreimeier U, et al. Intrathecal and systemic concentration of NT-proBNP in patients with severe traumatic brain injury. *J Neurotrauma* (2006) **23**:943–9. doi:10.1089/neu.2006.23.943
68. Powner DJ, Hergenroeder GW, Awili M, Atik MA, Robertson C. Hyponatremia and comparison of NT-pro-BNP concentrations in blood samples from jugular bulb and arterial sites after traumatic brain injury in adults: a pilot study. *Neurocrit Care* (2007) **7**:119–23. doi:10.1007/s12028-007-0079-8
69. Koenig MA, Puttgen HA, Prabhakaran V, Reich D, Stevens RD. B-type natriuretic peptide as a marker for heart failure in patients with acute stroke. *Intensive Care Med* (2007) **33**:1587–93. doi:10.1007/s00134-007-0704-1
70. Mcgirt MJ, Blessing R, Nimjee SM, Friedman AH, Alexander MJ, Laskowitz DT, et al. Correlation of serum brain natriuretic peptide with hyponatremia and delayed ischemic neurological deficits after subarachnoid hemorrhage. *Neurosurgery* (2004) **54**:1369–73. doi:10.1227/01.NEU.0000125016.37332.50
71. James ML, Blessing R, Phillips-Bute BG, Bennett E, Laskowitz DT. S100B and brain natriuretic peptide predict functional neurological outcome after intracerebral haemorrhage. *Biomarkers* (2009) **14**:388–94. doi:10.1080/13547500903015784
72. Sviri GE, Soustiel JE, Zaaroor M. Alteration in brain natriuretic peptide (BNP) plasma concentration following severe traumatic brain injury. *Acta Neurochir (Wien)* (2006) **148**:529–33. doi:10.1007/s00701-005-0666-4
73. Montaner J, Garcia-Berrocose T, Mendioroz M, Palacios M, Perea-Gainza M, Delgado P, et al. Brain natriuretic peptide is associated with worsening and mortality in acute stroke patients but adds no prognostic value to clinical predictors of outcome. *Cerebrovasc Dis* (2012) **34**:240–5. doi:10.1159/000341858
74. De Vito P. Atrial natriuretic peptide: an old hormone or a new cytokine? *Peptides* (2014) **58**:108–16. doi:10.1016/j.peptides.2014.06.011
75. Fink G, Dow RC, Casley D, Johnston CI, Lim AT, Copolov DL, et al. Atrial natriuretic peptide is a physiological inhibitor of ACTH release: evidence from immunoneutralization in vivo. *J Endocrinol* (1991) **131**:R9–12. doi:10.1677/joe.0.131R009
76. Vesely DL, San Miguel GI, Hassan I, Schocken DD. Atrial natriuretic hormone, vessel dilator, long-acting natriuretic hormone, and kaliuretic hormone decrease the circulating concentrations of CRH, corticotropin, and cortisol. *J Clin Endocrinol Metab* (2001) **86**:4244–9. doi:10.1210/jcem.86.9.7829
77. Charles CJ, Espiner EA, Richards AM, Donald RA. Central C-type natriuretic peptide augments the hormone response to hemorrhage in conscious sheep. *Peptides* (1995) **16**:129–32. doi:10.1016/0196-9781(94)00160-8
78. Strohle A, Jahn H, Montkowski A, Liebsch G, Boll E, Landgraf R, et al. Central and peripheral administration of atriopeptin is anxiolytic in rats. *Neuroendocrinology* (1997) **65**:210–5. doi:10.1159/000127274
79. Kellner M, Knaudt K, Jahn H, Holsboer F, Wiedemann K. Atrial natriuretic hormone in lactate-induced panic attacks: mode of release and endocrine and pathophysiological consequences. *J Psychiatr Res* (1998) **32**:37–48. doi:10.1016/S0022-3956(97)00034-4
80. Levinson DF, Zubenko GS, Crowe RR, Depaulo RJ, Scheftner WS, Weissman MM, et al. Genetics of recurrent early-onset depression (GenRED): design and preliminary clinical characteristics of a repository sample for genetic linkage studies. *Am J Med Genet B Neuropsychiatr Genet* (2003) **119B**:118–30. doi:10.1002/ajmg.b.20009
81. Herrmann-Lingen C, Binder L, Klinge M, Sander J, Schenker W, Beyermann B, et al. High plasma levels of N-terminal pro-atrial natriuretic peptide associated with low anxiety in severe heart failure. *Psychosom Med* (2003) **65**:517–22. doi:10.1097/01.PSY.0000073870.93003.C4
82. Kellner M, Herzog L, Yassouridis A, Holsboer F, Wiedemann K. Possible role of atrial natriuretic hormone in pituitary-adrenocortical unresponsiveness in lactate-induced panic. *Am J Psychiatry* (1995) **152**:1365–7.
83. Kellner M, Jahn H, Wiedemann K. Atrial natriuretic peptides and panic disorder: therapeutic prospects. *Expert Rev Neurother* (2003) **3**:381–6. doi:10.1586/14737175.3.3.381
84. Strohle A, Kellner M, Holsboer F, Wiedemann K. Anxiolytic activity of atrial natriuretic peptide in patients with panic disorder. *Am J Psychiatry* (2001) **158**:1514–6. doi:10.1176/appi.ajp.158.9.1514
85. Wiedemann K, Jahn H, Yassouridis A, Kellner M. Anxiolyticlike effects of atrial natriuretic peptide on cholecystokinin tetrapeptide-induced panic attacks: preliminary findings. *Arch Gen Psychiatry* (2001) **58**:371–7. doi:10.1001/archpsyc.58.4.371
86. Arlt J, Jahn H, Kellner M, Strohle A, Yassouridis A, Wiedemann K. Modulation of sympathetic activity by corticotropin-releasing hormone and atrial natriuretic peptide. *Neuropeptides* (2003) **37**:362–8. doi:10.1016/j.nep.2003.09.006
87. Biro E, Toth G, Telegyd G. Effect of receptor blockers on brain natriuretic peptide and C-type natriuretic peptide caused anxiolytic state in rats. *Neuropeptides* (1996) **30**:59–65. doi:10.1016/S0143-4179(96)90056-6
88. Montkowski A, Jahn H, Strohle A, Poettig M, Holsboer F, Wiedemann K. C-type natriuretic peptide exerts effects opposing those of atrial natriuretic peptide on anxiety-related behaviour in rats. *Brain Res* (1998) **792**:358–60. doi:10.1016/S0006-8993(98)00274-1
89. Jahn H, Montkowski A, Knaudt K, Strohle A, Kiefer F, Schick M, et al. Alpha-helical-corticotropin-releasing hormone reverses anxiogenic effects of C-type natriuretic peptide in rats. *Brain Res* (2001) **893**:21–8. doi:10.1016/S0006-8993(00)03275-3
90. Kellner M, Yassouridis A, Hua Y, Wendrich M, Jahn H, Wiedemann K. Intra-venous C-type natriuretic peptide augments behavioral and endocrine effects of cholecystokinin tetrapeptide in healthy men. *J Psychiatr Res* (2002) **36**:1–6. doi:10.1016/S0022-3956(01)00042-5
91. Kellner M, Diehl I, Knaudt K, Schule C, Jahn H, Wiedemann K. C-type natriuretic peptide exerts stimulatory effects on the corticotropin-releasing hormone-induced secretion of hormones in normal man. *Eur J Endocrinol* (1997) **136**:388–93. doi:10.1530/eje.0.1360388
92. Noble JM, Weimer LH. Neurologic complications of alcoholism. *Continuum (Minneapolis Minn)* (2014) **20**:624–41. doi:10.1212/01.CON.0000450970.99322.84
93. Kiefer F, Andersohn F, Jahn H, Wolf K, Raedler TJ, Wiedemann K. Involvement of plasma atrial natriuretic peptide in protracted alcohol withdrawal. *Acta Psychiatr Scand* (2002) **105**:65–70. doi:10.1034/j.1600-0447.2002.0\_011.x
94. von der Goltz C, Jahn H, Mutschler J, Wiedemann K, Kiefer F. Intraperitoneal atrial natriuretic peptide attenuates anxiety-related behaviour during alcohol withdrawal in mice. *Pharmacopsychiatry* (2014) **47**:97–100. doi:10.1055/s-0034-1372645
95. Kovacs GL. Alpha-atrial natriuretic peptide attenuates ethanol withdrawal symptoms. *Eur J Pharmacol* (1993) **238**:417–9. doi:10.1016/0014-2999(93)90878-L

96. Mutschler J, Bilbao A, von der Goltz C, Demiralay C, Jahn H, Wiedemann K, et al. Augmented stress-induced alcohol drinking and withdrawal in mice lacking functional natriuretic peptide-A receptors. *Alcohol Alcohol* (2010) **45**:13–6. doi:10.1093/alcalc/agp065
97. Gianoulakis C, Guillaume P, Thavundayil J, Gutkowska J. Increased plasma atrial natriuretic peptide after ingestion of low doses of ethanol in humans. *Alcohol Clin Exp Res* (1997) **21**:162–70. doi:10.1111/j.1530-0277.1997.tb03744.x
98. Glahn A, Riera Knorrrenschild R, Rhein M, Haschemi Nassab M, Groschl M, Heberlein A, et al. Alcohol-induced changes in methylation status of individual CpG sites, and serum levels of vasopressin and atrial natriuretic peptide in alcohol-dependent patients during detoxification treatment. *Eur Addict Res* (2014) **20**:143–50. doi:10.1159/000357473
99. Koopmann A, Lemenager T, Wolf ND, Reinhard I, Hermann D, Koch J, et al. The impact of atrial natriuretic peptide on anxiety, stress and craving in patients with alcohol dependence. *Alcohol Alcohol* (2014) **49**:282–6. doi:10.1093/alcalc/agt160
100. Kiefer F, Jahn H, Schick M, Wiedemann K. Alcohol self-administration, craving and HPA-axis activity: an intriguing relationship. *Psychopharmacology (Berl)* (2002) **164**:239–40. doi:10.1007/s00213-002-1255-3
101. Adinoff B, Junghanns K, Kiefer F, Krishnan-Sarin S. Suppression of the HPA axis stress-response: implications for relapse. *Alcohol Clin Exp Res* (2005) **29**:1351–5. doi:10.1097/01.ALC.0000176356.97620.84
102. Kiefer F, Jahn H, Tarnaske T, Helwig H, Briken P, Holzbach R, et al. Comparing and combining naltrexone and acamprose in relapse prevention of alcoholism: a double-blind, placebo-controlled study. *Arch Gen Psychiatry* (2003) **60**:92–9. doi:10.1001/archpsyc.60.1.92
103. Treutlein J, Cichon S, Ridinger M, Wodarz N, Soyka M, Zill P, et al. Genome-wide association study of alcohol dependence. *Arch Gen Psychiatry* (2009) **66**:773–84. doi:10.1001/archgenpsychiatry.2009.83
104. Edenberg HJ, Koller DL, Xuei X, Wetherill L, Mcclintick JN, Almasy L, et al. Genome-wide association study of alcohol dependence implicates a region on chromosome 11. *Alcohol Clin Exp Res* (2010) **34**:840–52. doi:10.1111/j.1530-0277.2010.01156.x
105. Karpyak VM, Winham SJ, Biernacka JM, Cunningham JM, Lewis KA, Geske JR, et al. Association of GATA4 sequence variation with alcohol dependence. *Addict Biol* (2014) **19**:312–5. doi:10.1111/j.1369-1600.2012.00482.x
106. Kiefer F, Witt SH, Frank J, Richter A, Treutlein J, Lemenager T, et al. Involvement of the atrial natriuretic peptide transcription factor GATA4 in alcohol dependence, relapse risk and treatment response to acamprose. *Pharmacogenomics J* (2011) **11**:368–74. doi:10.1038/tpj.2010.51
107. Jorde A, Bach P, Witt SH, Becker K, Reinhard I, Vollstadt-Klein S, et al. Genetic variation in the atrial natriuretic peptide transcription factor GATA4 modulates amygdala responsiveness in alcohol dependence. *Biol Psychiatry* (2014) **75**:790–7. doi:10.1016/j.biopsych.2013.10.020
108. Carey RM. Evidence for a splanchnic sodium input monitor regulating renal sodium excretion in man. Lack of dependence upon aldosterone. *Circ Res* (1978) **43**:19–23. doi:10.1161/01.RES.43.1.19
109. Currie MG, Fok KF, Kato J, Moore RJ, Hamra FK, Duffin KL, et al. Guanylin: an endogenous activator of intestinal guanylate cyclase. *Proc Natl Acad Sci U S A* (1992) **89**:947–51. doi:10.1073/pnas.89.3.947
110. Hamra FK, Forte LR, Eber SL, Pidhorodeckyj NV, Krause WJ, Freeman RH, et al. Uroguanylin: structure and activity of a second endogenous peptide that stimulates intestinal guanylate cyclase. *Proc Natl Acad Sci U S A* (1993) **90**:10464–8. doi:10.1073/pnas.90.22.10464
111. Forte LR, Eber SL, Fan X, London RM, Wang Y, Rowland LM, et al. Lym-phoguanylin: cloning and characterization of a unique member of the guanylin peptide family. *Endocrinology* (1999) **140**:1800–6. doi:10.1210/endo.140.4.6630
112. Yuge S, Inoue K, Hyodo S, Takei Y. A novel guanylin family (guanylin, uroguanylin, and renoguanylin) in eels: possible osmoregulatory hormones in intestine and kidney. *J Biol Chem* (2003) **278**:22726–33. doi:10.1074/jbc.M303111200
113. Hamra FK, Fan X, Krause WJ, Freeman RH, Chin DT, Smith CE, et al. Prouruguanylin and proguanylin: purification from colon, structure, and modulation of bioactivity by proteases. *Endocrinology* (1996) **137**:257–65. doi:10.1210/endo.137.1.8536621
114. de Sauvage FJ, Keshav S, Kuang WJ, Gillett N, Henzel W, Goeddel DV. Precursor structure, expression, and tissue distribution of human guanylin. *Proc Natl Acad Sci U S A* (1992) **89**:9089–93. doi:10.1073/pnas.89.19.9089
115. Kita T, Smith CE, Fok KF, Duffin KL, Moore WM, Karabatsos PJ, et al. Characterization of human uroguanylin: a member of the guanylin peptide family. *Am J Physiol* (1994) **266**:F342–8.
116. Forte LR, Eber SL, Turner JT, Freeman RH, Fok KF, Currie MG. Guanylin stimulation of Cl<sup>-</sup>secretion in human intestinal T84 cells via cyclic guanosine monophosphate. *J Clin Invest* (1993) **91**:2423–8. doi:10.1172/JCI116476
117. Schulz S, Chrisman TD, Garbers DL. Cloning and expression of guanylin. Its existence in various mammalian tissues. *J Biol Chem* (1992) **267**:16019–21.
118. Fan X, Wang Y, London RM, Eber SL, Krause WJ, Freeman RH, et al. Signaling pathways for guanylin and uroguanylin in the digestive, renal, central nervous, reproductive, and lymphoid systems. *Endocrinology* (1997) **138**:4636–48. doi:10.1210/endo.138.11.5539
119. Kita T, Kitamura K, Sakata J, Eto T. Marked increase of guanylin secretion in response to salt loading in the rat small intestine. *Am J Physiol* (1999) **277**:G960–6.
120. Pitari GM, Di Guglielmo MD, Park J, Schulz S, Waldman SA. Guanylyl cyclase-C agonists regulate progression through the cell cycle of human colon carcinoma cells. *Proc Natl Acad Sci U S A* (2001) **98**:7846–51. doi:10.1073/pnas.141124698
121. Fontes MC, Greenberg RN, Monteiro HS, Currie MG, Forte LR. Natriuretic and kaliuretic activities of guanylin and uroguanylin in the isolated perfused rat kidney. *Am J Physiol* (1998) **275**:F191–7.
122. Carrithers SL, Ott CE, Hill MJ, Johnson BR, Cai W, Chang JJ, et al. Guanylin and uroguanylin induce natriuresis in mice lacking guanylyl cyclase-C receptor. *Kidney Int* (2004) **65**:40–53. doi:10.1111/j.1523-1755.2004.00375.x
123. Fulle HJ, Vassar R, Foster DC, Yang RB, Axel R, Garbers DL. A receptor guanylyl cyclase expressed specifically in olfactory sensory neurons. *Proc Natl Acad Sci U S A* (1995) **92**:3571–5. doi:10.1073/pnas.92.8.3571
124. Chao YC, Cheng CJ, Hsieh HT, Lin CC, Chen CC, Yang RB. Guanylate cyclase-G, expressed in the Grueneberg ganglion olfactory subsystem, is activated by bicarbonate. *Biochem J* (2010) **432**:267–73. doi:10.1042/BJ20100617
125. Nakazato M, Yamaguchi H, Shiomori K, Date Y, Fujimoto S, Kangawa K, et al. Identification of 10-kDa proguanylin as a major guanylin molecule in human intestine and plasma and its increase in renal insufficiency. *Biochem Biophys Res Commun* (1994) **205**:1966–75. doi:10.1006/bbrc.1994.2901
126. Moss NG, Fellner RC, Qian X, Yu SJ, Li Z, Nakazato M, et al. Uroguanylin, an intestinal natriuretic peptide, is delivered to the kidney as an unprocessed propeptide. *Endocrinology* (2008) **149**:4486–98. doi:10.1210/en.2007-1725
127. Kinoshita H, Fujimoto S, Nakazato M, Yokota N, Date Y, Yamaguchi H, et al. Urine and plasma levels of uroguanylin and its molecular forms in renal diseases. *Kidney Int* (1997) **52**:1028–34. doi:10.1038/ki.1997.424
128. Valentino MA, Lin JE, Snook AE, Li P, Kim GW, Marszalowicz G, et al. A uroguanylin-GUCY2C endocrine axis regulates feeding in mice. *J Clin Invest* (2011) **121**:3578–88. doi:10.1172/JCI57925
129. Gong R, Ding C, Hu J, Lu Y, Liu F, Mann E, et al. Role for the membrane receptor guanylyl cyclase-C in attention deficiency and hyperactive behavior. *Science* (2011) **333**:1642–6. doi:10.1126/science.1207675
130. Strader AD, Woods SC. Gastrointestinal hormones and food intake. *Gastroenterology* (2005) **128**:175–91. doi:10.1053/j.gastro.2004.10.043
131. Badman MK, Flier JS. The gut and energy balance: visceral allies in the obesity wars. *Science* (2005) **307**:1909–14. doi:10.1126/science.1109951
132. Seeley RJ, Tschop MH. Uroguanylin: how the gut got another satiety hormone. *J Clin Invest* (2011) **121**:3384–6. doi:10.1172/JCI58297
133. Volkow ND, Wang GJ, Kollins SH, Wigal TL, Newcorn JH, Telang F, et al. Evaluating dopamine reward pathway in ADHD: clinical implications. *JAMA* (2009) **302**:1084–91. doi:10.1001/jama.2009.1308
134. Sagvolden T, Russell VA, Aase H, Johansen EB, Farshbaf M. Rodent models of attention-deficit/hyperactivity disorder. *Biol Psychiatry* (2005) **57**:1239–47. doi:10.1016/j.biophys.2005.02.002
135. Zufall F, Munger SD. Receptor guanylyl cyclases in mammalian olfactory function. *Mol Cell Biochem* (2010) **334**:191–7. doi:10.1007/s11010-009-0325-9
136. Chen KK, Kovarikova A. Pharmacology and toxicology of toad venom. *J Pharm Sci* (1967) **56**:1535–41. doi:10.1002/jps.2600561202
137. de Wardener HE, Clarkson EM, Nutbourne DM, Schrier RW, Talner LB, Ven-tom MG, et al. Evidence for a hormone other than aldosterone which controls urinary sodium excretion. *Adv Nephrol Necker Hosp* (1971) **1**:97–111.

138. Lichtstein D, Samuelov S. Endogenous 'ouabain like' activity in rat brain. *Biochem Biophys Res Commun* (1980) **96**:1518–23. doi:10.1016/0006-291X(80)91346-7
139. Fishman MC. Endogenous digitalis-like activity in mammalian brain. *Proc Natl Acad Sci U S A* (1979) **76**:4661–3. doi:10.1073/pnas.76.9.4661
140. Haupert GT Jr, Sancho JM. Sodium transport inhibitor from bovine hypothalamus. *Proc Natl Acad Sci U S A* (1979) **76**:4658–60. doi:10.1073/pnas.76.9.4658
141. Hamlyn JM, Blaustein MP, Bova S, Ducharme DW, Harris DW, Mandel F, et al. Identification and characterization of a ouabain-like compound from human plasma. *Proc Natl Acad Sci U S A* (1991) **88**:6259–63. doi:10.1073/pnas.88.21.9907-d
142. Ludens JH, Clark MA, Ducharme DW, Harris DW, Lutzke BS, Mandel F, et al. Purification of an endogenous digitalis like factor from human plasma for structural analysis. *Hypertension* (1991) **17**:923–9. doi:10.1161/01.HYP.17.6.923
143. Goto A, Ishiguro T, Yamada K, Ishii M, Yoshioka M, Eguchi C, et al. Isolation of a urinary digitalis-like factor indistinguishable from digoxin. *Biochem Biophys Res Commun* (1990) **173**:1093–101. doi:10.1016/S0006-291X(05)80898-8
144. Lichtstein D, Gati I, Samuelov S, Berson D, Rozenman Y, Landau L, et al. Identification of digitalis-like compounds in human cataractous lenses. *Eur J Biochem* (1993) **216**:261–8. doi:10.1111/j.1432-1033.1993.tb18141.x
145. Hilton PJ, White RW, Lord GA, Garner GV, Gordon DB, Hilton MJ, et al. An inhibitor of the sodium pump obtained from human placenta. *Lancet* (1996) **348**:303–5. doi:10.1016/S0140-6736(96)02257-X
146. Bagrov AY, Fedorova OV, Dmitrieva RI, Howald WN, Hunter AP, Kuznetsova EA, et al. Characterization of a urinary bufadienolide Na<sup>+</sup>,K<sup>+</sup>-ATPase inhibitor in patients after acute myocardial infarction. *Hypertension* (1998) **31**:1097–103. doi:10.1161/01.HYP.31.5.1097
147. Schneider R, Antolovic R, Kost H, Sich B, Kirch U, Tepel M, et al. Proscillarinidin A immunoreactivity: its purification, transport in blood by a specific binding protein and its correlation with blood pressure. *Clin Exp Hypertens* (1998) **20**:593–9. doi:10.3109/10641969809053237
148. Komiyama Y, Dong XH, Nishimura N, Masaki H, Yoshika M, Masuda M, et al. A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin Biochem* (2005) **38**:36–45. doi:10.1016/j.clinbiochem.2004.08.005
149. Lichtstein D, Steinitz M, Gati I, Samuelov S, Deutsch J, Orly J. Biosynthesis of digitalis-like compounds in rat adrenal cells: hydroxycholesterol as possible precursor. *Life Sci* (1998) **62**:2109–26. doi:10.1016/S0024-3205(98)00186-6
150. Murrell JR, Randall JD, Rosoff J, Zhao JL, Jensen RV, Gullans SR, et al. Endogenous ouabain: upregulation of steroidogenic genes in hypertensive hypothalamus but not adrenal. *Circulation* (2005) **112**:1301–8. doi:10.1161/CIRCULATIONAHA.105.554071
151. Greeff K, Gänert TW, Linde HHA. *Chemistry and Structure-Activity Relationships of Cardioactive Steroids*, in *Cardiac Glycosides*. Berlin Heidelberg: Springer (1981). p. 13–24.
152. Klein M, Nejad NS, Lown B, Hagemeijer F, Barr I. Correlation of the electrical and mechanical changes in the dog heart during progressive digitalization. *Circ Res* (1971) **29**:635–45.
153. Xie Z, Cai T. Na<sup>+</sup>-K<sup>+</sup> – ATPase-mediated signal transduction: from protein interaction to cellular function. *Mol Interv* (2003) **3**:157–68. doi:10.1124/mi.3.3.157
154. Schonher W, Scheiner-Bobis G. Endogenous and exogenous cardiac glycosides and their mechanisms of action. *Am J Cardiovac Drugs* (2007) **7**:173–89. doi:10.2165/00129784-200707030-00004
155. Huang BS, Amin MS, Leenen FH. The central role of the brain in salt-sensitive hypertension. *Curr Opin Cardiol* (2006) **21**:295–304. doi:10.1097/01.hco.0000231398.64362.94
156. Lichtstein D. Na<sup>+</sup>, K<sup>(+)</sup>-ATPase and heart excitability. *Adv Exp Med Biol* (1995) **382**:23–30. doi:10.1007/978-1-4615-1893-8\_3
157. Blaustein MP, Zhang J, Chen L, Hamilton BP. How does salt retention raise blood pressure? *Am J Physiol Regul Integr Comp Physiol* (2006) **290**:R514–23. doi:10.1152/ajpregu.00819.2005
158. Dvela M, Rosen H, Ben-Ami HC, Lichtstein D. Endogenous ouabain regulates cell viability. *Am J Physiol Cell Physiol* (2012) **302**:C442–52. doi:10.1152/ajpcell.00336.2011
159. Goldstein I, Levy T, Galili D, Ovadia H, Yirmiya R, Rosen H, et al. Involvement of Na<sup>(+)</sup>, K<sup>(+)</sup>-ATPase and endogenous digitalis-like compounds in depressive disorders. *Biol Psychiatry* (2006) **60**:491–9. doi:10.1016/j.biopsych.2005.12.021
160. Pongrakhananon V, Chunhacha P, Chanvorachote P. Ouabain suppresses the migratory behavior of lung cancer cells. *PLoS One* (2013) **8**:e68623. doi:10.1371/journal.pone.0068623
161. Newman RA, Yang P, Pawlus AD, Block KI. Cardiac glycosides as novel cancer therapeutic agents. *Mol Interv* (2008) **8**:36–49. doi:10.1124/mi.8.1.8
162. Halperin J, Schaeffer R, Galvez L, Malave S. Ouabain-like activity in human cerebrospinal fluid. *Proc Natl Acad Sci U S A* (1983) **80**:6101–4. doi:10.1073/pnas.80.19.6101
163. Lichtstein D, Minc D, Bourrit A, Deutsch J, Karlish SJ, Belmaker H, et al. Evidence for the presence of 'ouabain like' compound in human cerebrospinal fluid. *Brain Res* (1985) **325**:13–9. doi:10.1016/0006-8993(85)90297-5
164. Yamada H, Ihara N, Takahashi H, Yoshimura M, Sano Y. Distribution of the endogenous digitalis-like substance (EDLS)-containing neurons labeled by digoxin antibody in hypothalamus and three circumventricular organs of dog and macaque. *Brain Res* (1992) **584**:237–43. doi:10.1016/0006-8993(92)9000-T
165. Morgan K, Lewis MD, Spurlock G, Collins PA, Foord SM, Southgate K, et al. Characterization and partial purification of the sodium-potassium-ATPase inhibitor released from cultured rat hypothalamic cells. *J Biol Chem* (1985) **260**:13595–600.
166. Blanco G, Mercer RW. Isozymes of the Na-K-ATPase: heterogeneity in structure, diversity in function. *Am J Physiol* (1998) **275**:F633–50.
167. Moseley AE, Lieske SP, Wetzel RK, James PF, He S, Shelly DA, et al. The Na,K-ATPase alpha 2 isoform is expressed in neurons, and its absence disrupts neuronal activity in newborn mice. *J Biol Chem* (2003) **278**:5317–24. doi:10.1074/jbc.M211315200
168. Mcgrail KM, Phillips JM, Sweadner KJ. Immunofluorescent localization of three Na,K-ATPase isozymes in the rat central nervous system: both neurons and glia can express more than one Na,K-ATPase. *J Neurosci* (1991) **11**:381–91.
169. Meneton P, Jeunemaitre X, De Wardener HE, Macgregor GA. Links between dietary salt intake, renal salt handling, blood pressure, and cardiovascular diseases. *Physiol Rev* (2005) **85**:679–715. doi:10.1152/physrev.00056.2003
170. Goto A, Yamada K, Ishii M, Sugimoto T. Digitalis-like activity in human plasma: relation to blood pressure and sodium balance. *Am J Med* (1990) **89**:420–6. doi:10.1016/0002-9343(90)90369-O
171. Rossi G, Manunta P, Hamlyn JM, Pavan E, De Toni R, Semplicini A, et al. Immunoreactive endogenous ouabain in primary aldosteronism and essential hypertension: relationship with plasma renin, aldosterone and blood pressure levels. *J Hypertens* (1995) **13**:1181–91. doi:10.1097/00004872-199510000-00013
172. Gonick HC, Ding Y, Vaziri ND, Bagrov AY, Fedorova OV. Simultaneous measurement of marinobufagenin, ouabain, and hypertension-associated protein in various disease states. *Clin Exp Hypertens* (1998) **20**:617–27. doi:10.3109/10641969809053240
173. Manunta P, Messaggio E, Ballabeni C, Sciarrone MT, Lanzani C, Ferrandi M, et al. Plasma ouabain-like factor during acute and chronic changes in sodium balance in essential hypertension. *Hypertension* (2001) **38**:198–203. doi:10.1161/01.HYP.38.2.198
174. Pierdomenico SD, Bucci A, Manunta P, Rivera R, Ferrandi M, Hamlyn JM, et al. Endogenous ouabain and hemodynamic and left ventricular geometric patterns in essential hypertension. *Am J Hypertens* (2001) **14**:44–50. doi:10.1016/S0895-7061(00)01225-5
175. Fridman AI, Matveev SA, Agalakova NI, Fedorova OV, Lakatta EG, Bagrov AY. Marinobufagenin, an endogenous ligand of alpha-1 sodium pump, is a marker of congestive heart failure severity. *J Hypertens* (2002) **20**:1189–94. doi:10.1097/00004872-200206000-00032
176. Hasegawa T, Masugi F, Ogihara T, Kumahara Y. Increase in plasma ouabainlike inhibitor of Na<sup>+</sup>, K<sup>+</sup>-ATPase with high sodium intake in patients with essential hypertension. *J Clin Hypertens* (1987) **3**:419–29.
177. Leenen FH, Harmsen E, Yu H, Yuan B. Dietary sodium stimulates ouabain-like activity in adrenalectomized spontaneously hypertensive rats. *Am J Physiol* (1993) **265**:H421–4.
178. Fedorova OV, Talan MI, Agalakova NI, Lakatta EG, Bagrov AY. Endogenous ligand of alpha(1) sodium pump, marinobufagenin, is a novel mediator of sodium chloride – dependent hypertension. *Circulation* (2002) **105**:1122–7. doi:10.1161/hc0902.104710

179. Manunta P, Rogowski AC, Hamilton BP, Hamlyn JM. Ouabain-induced hypertension in the rat: relationships among plasma and tissue ouabain and blood pressure. *J Hypertens* (1994) **12**:549–60. doi:10.1097/00004872-199405000-00008
180. Yamada K, Goto A, Omata M. Adrenocorticotropin-induced hypertension in rats: role of ouabain-like compound. *Am J Hypertens* (1997) **10**:403–8. doi:10.1016/S0895-7061(97)90523-9
181. Yuan CM, Manunta P, Hamlyn JM, Chen S, Bohen E, Yeun J, et al. Long-term ouabain administration produces hypertension in rats. *Hypertension* (1993) **22**:178–87. doi:10.1161/01.HYP.22.2.178
182. Manunta P, Hamilton J, Rogowski AC, Hamilton BP, Hamlyn JM. Chronic hypertension induced by ouabain but not digoxin in the rat: antihypertensive effect of digoxin and digitoxin. *Hypertens Res* (2000) **23**(Suppl):S77–85. doi:10.1291/hypres.23.Supplement\_S77
183. Dostanic I, Paul RJ, Lorenz JN, Theriault S, Van Huyse JW, Lingrel JB. The alpha2-isoform of Na-K-ATPase mediates ouabain-induced hypertension in mice and increased vascular contractility in vitro. *Am J Physiol Heart Circ Physiol* (2005) **288**:H477–85. doi:10.1152/ajpheart.00083.2004
184. Dostanic-Larson I, Lorenz JN, Van Huyse JW, Neumann JC, Moseley AE, Lingrel JB. Physiological role of the alpha1- and alpha2-isoforms of the Na+-K+-ATPase and biological significance of their cardiac glycoside binding site. *Am J Physiol Regul Integr Comp Physiol* (2006) **290**:R524–8. doi:10.1152/ajpregu.00838.2005
185. Loreaux EL, Kaul B, Lorenz JN, Lingrel JB. Ouabain-sensitive alpha1 Na,K-ATPase enhances natriuretic response to saline load. *J Am Soc Nephrol* (2008) **19**:1947–54. doi:10.1681/ASN.2008020174
186. Zhang J, Lee MY, Cavalli M, Chen L, Berra-Romani R, Balke CW, et al. Sodium pump alpha2 subunits control myogenic tone and blood pressure in mice. *J Physiol* (2005) **569**:243–56. doi:10.1113/jphysiol.2005.091801
187. Poburko D, Liao CH, Lemos VS, Lin E, Maruyama Y, Cole WC, et al. Transient receptor potential channel 6-mediated, localized cytosolic [Na<sup>+</sup>] transients drive Na<sup>+</sup>/Ca<sup>2+</sup> exchanger-mediated Ca<sup>2+</sup> entry in purinergically stimulated aorta smooth muscle cells. *Circ Res* (2007) **101**:1030–8. doi:10.1161/CIRCRESAHA.107.155531
188. Kaide J, Ura N, Torii T, Nakagawa M, Takada T, Shimamoto K. Effects of digoxin-specific antibody Fab fragment (Digibind) on blood pressure and renal water-sodium metabolism in 5/6 reduced renal mass hypertensive rats. *Am J Hypertens* (1999) **12**:611–9. doi:10.1016/S0895-7061(99)00029-1
189. Wenceslau CF, Rossini LV. Rostafuroxin ameliorates endothelial dysfunction and oxidative stress in resistance arteries from deoxycorticosterone acetate-salt hypertensive rats: the role of Na<sup>+</sup>K<sup>+</sup>-ATPase/cSRC pathway. *J Hypertens* (2014) **32**:542–54. doi:10.1097/HJH.0000000000000059
190. Veerasingham SJ, Leenen FH. Excitotoxic lesions of the ventral anteroventral third ventricle and pressor responses to central sodium, ouabain and angiotensin II. *Brain Res* (1997) **749**:157–60. doi:10.1016/S0006-8993(96)01381-9
191. Veerasingham SJ, Leenen FH. Ouabain- and central sodium-induced hypertension depend on the ventral anteroventral third ventricle region. *Am J Physiol* (1999) **276**:H63–70.
192. Huang BS, Leenen FH. Sympathoexcitatory and pressor responses to increased brain sodium and ouabain are mediated via brain ANG II. *Am J Physiol* (1996) **270**:H275–80.
193. Huang BS, Leenen FH. Brain renin-angiotensin system and ouabain-induced sympathetic hyperactivity and hypertension in Wistar rats. *Hypertension* (1999) **34**:107–12. doi:10.1161/01.HYP.34.1.107
194. Huang BS, Ganten D, Leenen FH. Responses to central Na<sup>+</sup> and ouabain are attenuated in transgenic rats deficient in brain angiotensinogen. *Hypertension* (2001) **37**:683–6. doi:10.1161/01.HYP.37.2.683
195. Kawano Y, Yoshida K, Kawamura M, Yoshimi H, Ashida T, Abe H, et al. Sodium and noradrenaline in cerebrospinal fluid and blood in salt-sensitive and non-salt-sensitive essential hypertension. *Clin Exp Pharmacol Physiol* (1992) **19**:235–41. doi:10.1111/j.1440-1681.1992.tb00444.x
196. Huang BS, Van Vliet BN, Leenen FH. Increases in CSF [Na<sup>+</sup>] precede the increases in blood pressure in Dahl S rats and SHR on a high-salt diet. *Am J Physiol Heart Circ Physiol* (2004) **287**:H1160–6. doi:10.1152/ajpheart.00126.2004
197. Bhatia SJ. Digitalis toxicity – turning over a new leaf? *West J Med* (1986) **145**:74–82.
198. el-Mallakh RS, Wyatt RJ. The Na,K-ATPase hypothesis for bipolar illness. *Biol Psychiatry* (1995) **37**:235–44. doi:10.1016/0006-3223(94)00201-D
199. Traub N, Lichtstein D. The mood cycle hypothesis: possible involvement of steroid hormones in mood regulation by means of Na<sup>+</sup>, K<sup>+</sup>-ATPase inhibition. *J Basic Clin Physiol Pharmacol* (2000) **11**:375–94. doi:10.1515/JBCPP.2000.11.4.375
200. Naylor GJ, McNamee HB, Moody JP. Changes in erythrocyte sodium and potassium on recovery from a depressive illness. *Br J Psychiatry* (1971) **118**:219–23. doi:10.1192/bj.p.118.543.219
201. Nurnberger J Jr, Jimerson DC, Allen JR, Simmons S, Gershon E. Red cell ouabain-sensitive Na<sup>+</sup>-K<sup>+</sup>-adenosine triphosphatase: a state marker in affective disorder inversely related to plasma cortisol. *Biol Psychiatry* (1982) **17**:981–92.
202. Looney SW, el-Mallakh RS. Meta-analysis of erythrocyte Na,K-ATPase activity in bipolar illness. *Depress Anxiety* (1997) **5**:53–65. doi:10.1002/(SICI)1520-6394(1997)5:2<53::AID-DA1>3.0.CO;2-6
203. Grider G, El-Mallakh RS, Huff MO, Buss TJ, Miller J, Valdes R Jr. Endogenous digoxin-like immunoreactive factor (DLIF) serum concentrations are decreased in manic bipolar patients compared to normal controls. *J Affect Disord* (1999) **54**:261–7. doi:10.1016/S0165-0327(98)00208-0
204. El-Mallakh RS, Stoddard M, Jortani SA, El-Masri MA, Sephton S, Valdes R Jr. Aberrant regulation of endogenous ouabain-like factor in bipolar subjects. *Psychiatry Res* (2010) **178**:116–20. doi:10.1016/j.psychres.2009.03.032
205. Chetcuti A, Adams LJ, Mitchell PB, Schofield PR. Microarray gene expression profiling of mouse brain mRNA in a model of lithium treatment. *Psychiatr Genet* (2008) **18**:64–72. doi:10.1097/YPG.0b013e3282fb0051
206. Mynett-Johnson L, Murphy V, McCormack J, Shields DC, Claffey E, Manley P, et al. Evidence for an allelic association between bipolar disorder and a Na<sup>+</sup>, K<sup>+</sup> adenosine triphosphatase alpha subunit gene (ATP1A3). *Biol Psychiatry* (1998) **44**:47–51. doi:10.1016/S0006-3223(97)00343-0
207. Goldstein I, Lerer E, Laiba E, Mallet J, Mujahed M, Laurent C, et al. Association between sodium- and potassium-activated adenosine triphosphatase alpha isoforms and bipolar disorders. *Biol Psychiatry* (2009) **65**:985–91. doi:10.1016/j.biopsych.2008.10.033
208. Kirshenbaum GS, Clapcote SJ, Duffy S, Burgess CR, Petersen J, Jarowek KJ, et al. Mania-like behavior induced by genetic dysfunction of the neuron-specific Na<sup>+</sup>,K<sup>+</sup>-ATPase alpha3 sodium pump. *Proc Natl Acad Sci USA* (2011) **108**:18144–9. doi:10.1073/pnas.1108416108
209. El-Mallakh RS, El-Masri MA, Huff MO, Li XP, Decker S, Levy RS. Intracerebroventricular administration of ouabain as a model of mania in rats. *Bipolar Disord* (2003) **5**:362–5. doi:10.1034/j.1399-5618.2003.00053.x
210. Brocardo PS, Budni J, Pavesi E, Franco JL, Ulian-Silva M, Trevisan R, et al. Folic acid administration prevents ouabain-induced hyperlocomotion and alterations in oxidative stress markers in the rat brain. *Bipolar Disord* (2010) **12**:414–24. doi:10.1111/j.1399-5618.2010.00827.x
211. Goldstein I, Lax E, Gispán-Herman I, Ovadia H, Rosen H, Yadid G, et al. Neutralization of endogenous digitalis-like compounds alters catecholamines metabolism in the brain and elicits anti-depressive behavior. *Eur Neuropsychopharmacol* (2012) **22**:72–9. doi:10.1016/j.euroneuro.2011.05.007
212. Yu HS, Kim SH, Park HG, Kim YS, Ahn YM. Activation of Akt signaling in rat brain by intracerebroventricular injection of ouabain: a rat model for mania. *Prog Neuropsychopharmacol Biol Psychiatry* (2010) **34**:888–94. doi:10.1016/j.pnpbp.2010.04.010
213. Kim SH, Yu HS, Park HG, Ha K, Kim YS, Shin SY, et al. Intracerebroventricular administration of ouabain, a Na/K-ATPase inhibitor, activates mTOR signal pathways and protein translation in the rat frontal cortex. *Prog Neuropsychopharmacol Biol Psychiatry* (2013) **45**:73–82. doi:10.1016/j.pnpbp.2013.04.018
214. Freitas TP, Rezin GT, Goncalves CL, Jeremias GC, Gomes LM, Scaini G, et al. Evaluation of citrate synthase activity in brain of rats submitted to an animal model of mania induced by ouabain. *Mol Cell Biochem* (2010) **341**:245–9. doi:10.1007/s11010-010-0455-0
215. Riegel RE, Valvassori SS, Moretti M, Ferreira CL, Steckert AV, De Souza B, et al. Intracerebroventricular ouabain administration induces oxidative stress in the rat brain. *Int J Dev Neurosci* (2010) **28**:233–7. doi:10.1016/j.ijdevneu.2010.02.002
216. Lewis LK, Yandle TG, Hilton PJ, Jensen BP, Begg EJ, Nicholls MG. Endogenous ouabain is not ouabain. *Hypertension* (2014) **64**:680–3. doi:10.1161/HYPERTENSIONAHA.114.03919

217. Lichtstein D, Rosen H, Dvela M. Cardenolides and bufadienolides as hormones: what is missing? *Am J Physiol Renal Physiol* (2012) **302**:F957–8. doi:10.1152/ajprenal.00042.2012
218. Bloch KD, Zamir N, Lichtstein D, Seidman CE, Seidman JG. Ouabain induces secretion of proatrial natriuretic factor by rat atrial cardiocytes. *Am J Physiol* (1988) **255**:E383–7.
219. Morise T, Takeuchi Y, Okamoto S, Takeda R. Stimulation of atrial natriuretic peptide secretion and synthesis by Na-K-ATPase inhibitors. *Biochem Biophys Res Commun* (1991) **176**:875–81. doi:10.1016/S0006-291X(05)80267-0
220. Schiebinger RJ, Cragoe EJ Jr. Ouabain. A stimulator of atrial natriuretic peptide secretion and its mechanism of action. *Circ Res* (1993) **72**:1035–43. doi:10.1161/01.RES.72.5.1035
221. Liu LP, Hong L, Yu L, Li HY, Ding DZ, Jin SJ, et al. Ouabain stimulates atrial natriuretic peptide secretion via the endothelin-1/ET(B) receptor-mediated pathway in beating rabbit atria. *Life Sci* (2012) **90**:793–8. doi:10.1016/j.lfs.2012.04.008
222. Yamamoto A, Shouji T, Kimura S, Aki Y, Nakamura A, Fukui K, et al. Effects of hypercalcemia and ouabain on plasma atrial natriuretic polypeptide in anesthetized dogs. *Am J Physiol* (1988) **255**:E437–41.
223. Tsutamoto T, Wada A, Maeda K, Hisanaga T, Fukai D, Maeda Y, et al. Digitalis increases brain natriuretic peptide in patients with severe congestive heart failure. *Am Heart J* (1997) **134**:910–6. doi:10.1016/S0002-8703(97)80014-2
224. Crabos M, Ausiello DA, Haupert GT Jr, Cantiello HF. Atrial natriuretic peptide regulates release of Na+-K+-ATPase inhibitor from rat brain. *Am J Physiol* (1988) **254**:F912–7.
225. Songu-Mize E, Bealer SL, Hassid AI. Centrally administered ANF promotes appearance of a circulating sodium pump inhibitor. *Am J Physiol* (1990) **258**:H1655–9.
226. Songu-Mize E, Bealer SL. Effect of hypothalamic lesions on interaction of centrally administered ANF and the circulating sodium-pump inhibitor. *J Cardiovasc Pharmacol* (1993) **22**(Suppl 2):S4–6. doi:10.1097/00005344-199322002-00003
227. Fedorova OV, Agalakova NI, Morrell CH, Lakatta EG, Bagrov AY. ANP differentially modulates marinobufagenin-induced sodium pump inhibition in kidney and aorta. *Hypertension* (2006) **48**:1160–8. doi:10.1161/01.HYP.0000248129.20524.d0
228. Sybertz EJ, Desiderio DM. The role of Na+-K+-ATPase in the vasorelaxant actions of synthetic atrial natriuretic factor. *Arch Int Pharmacodyn Ther* (1985) **278**:142–9.
229. Chintala MS, Jandhyala BS. Interaction between atrial natriuretic factor and ouabain: vascular reactivity to noradrenaline in pentobarbital anaesthetized dogs. *Clin Exp Pharmacol Physiol* (1988) **15**:591–9. doi:10.1111/j.1440-1681.1988.tb01118.x
230. Szalay KS, Beck M, Toth M, De Chatel R. Interactions between ouabain, atrial natriuretic peptide, angiotensin-II and potassium: effects on rat zona glomerulosa aldosterone production. *Life Sci* (1998) **62**:1845–52. doi:10.1016/S0024-3205(98)00150-7
231. Nesher M, Dvela M, Igbokwe VU, Rosen H, Lichtstein D. Physiological roles of endogenous ouabain in normal rats. *Am J Physiol Heart Circ Physiol* (2009) **297**:H2026–34. doi:10.1152/ajpheart.00734.2009
232. Nesher M, Bai Y, Li D, Rosen H, Lichtstein D, Liu L. Interaction of atrial natriuretic peptide and ouabain in the myocardium. *Can J Physiol Pharmacol* (2012) **90**:1386–93. doi:10.1139/y2012-112
233. Santos-Neto MS, Carvalho AF, Monteiro HS, Forte LR, Fonteles MC. Interaction of atrial natriuretic peptide, urodilatin, guanylin and uroguanylin in the isolated perfused rat kidney. *Regul Pept* (2006) **136**:14–22. doi:10.1016/j.regpep.2006.04.017
234. Potter LR. Regulation and therapeutic targeting of peptide-activated receptor guanylyl cyclases. *Pharmacol Ther* (2011) **130**:71–82. doi:10.1016/j.pharmthera.2010.12.005

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 09 September 2014; accepted: 12 November 2014; published online: 28 November 2014.

Citation: Hodes A and Lichtstein D (2014) Natriuretic hormones in brain function. *Front. Endocrinol.* 5:201. doi: 10.3389/fendo.2014.00201

This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Hodes and Lichtstein. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Identification of putative natriuretic hormones isolated from human urine

Herbert J. Kramer\*

Center of Internal Medicine, Rheinische-Friedrich-Wilhelms-University, Bonn, Germany,

## OPEN ACCESS

**Edited by:**

Harvey C. Gonick,  
University of California Berkeley, USA

**Reviewed by:**

Harvey C. Gonick,  
University of California Berkeley, USA

Vardaman M. Buckalew,  
Wake Forest School of Medicine,  
USA

**\*Correspondence:**

Herbert J. Kramer,  
Center of Internal Medicine,  
Rheinische-Friedrich-Wilhelms-  
University, Sigmund-Freud-Strasse  
25, Bonn D-53127, Germany  
[hkramer@uni-bonn.de](mailto:hkramer@uni-bonn.de)

**Specialty section:**

This article was submitted to  
Neuroendocrine Science, a section of  
the journal *Frontiers in Endocrinology*

**Received:** 16 January 2015

**Accepted:** 14 April 2015

**Published:** 20 May 2015

**Citation:**

Kramer HJ (2015) Identification of putative natriuretic hormones isolated from human urine.  
*Front. Endocrinol.* 6:66.  
doi: 10.3389/fendo.2015.00066

This brief review describes some representative methodological approaches to the isolation of putative endogenous inhibitors of epithelial sodium transport – i.e., as ouabain-like factors (OLF) that inhibit the sodium transport enzyme Na-K-ATPase or inhibit the epithelial sodium channel (ENaC). Gel chromatography and reverse-phase (RP)-high performance liquid chromatography (HPLC) of lyophilized and reconstituted 24 h-urine from salt-loaded healthy humans led to two active fractions, a hydrophilic OLF-1 and a lipophilic OLF-2, whose mass (Ms)-spectroscopic data indicate a  $M_r$  of 391 (1, 2). Further identification was attempted by Ms-, infrared (IR)-, ultraviolet (UV)-, and  $^1\text{H}$ -NMR-spectroscopy. OLF-1 and OLF-2 may be closely related if not identical to (di)ascorbic acid or its salts such as vanadium (V)-V<sup>IV</sup>-diascorbate with  $M_r$  403 (3) and V<sup>IV</sup>-diascorbate. OLF-1 and V<sup>IV</sup>-diascorbate are about 10-fold stronger inhibitors of Na-K-ATPase than OLF-2 and V<sup>IV</sup>-diascorbate, respectively. In conscious rats, i.v. infusion of OLF-1 and OLF-2 resulted in a strong natriuresis. In a similar study, Cain et al. (4) isolated a sodium transport inhibitor from the urine of uremic patients by gel chromatography and RP-HPLC. In uremic rats, a natriuretic response to the injection of the active material was found. Xanthurenic acid 8-O- $\beta$ -D-glucoside ( $M_r$  368) and xanthurenic acid 8-O-sulfate ( $M_r$  284) were identified as endogenous inhibitors of sodium transport acting, e.g., by ENaC blockade. No definite relation to blood pressure, body fluid volume, or sodium balance has been reported for any of these above factors, and further studies to identify the natriuretic and/or ouabain-like compound(s) or hormone(s) will be needed.

**Keywords:** sodium transport, natriuretic hormone, human urine, endogenous inhibitors, epithelial sodium transport

## Introduction – Background

With this brief review, some methodological aspects of the isolation of putative endogenous membrane transport inhibitor(s) and natriuretic factor(s) will be described, and the results compared with those of similar attempts by other groups of investigators.

In 1969 (5), and in more detail in 1974 (6) and 1977 (7), we demonstrated for the first time that acute extracellular fluid volume (ECFV)-expansion in rats may release a natriuretic factor or “hormone,” which was postulated to act through inhibition of the sodium pump. Thus, ECFV-expansion was accompanied by a decrease in Na-K-ATPase in the renal cortex and the appearance of an inhibitor of Na-K-ATPase in the serum of these rats, respectively. Using gel chromatography, this inhibitory activity was also detected in the post-salt fraction of serum from ECFV-expanded dogs and in the serum and urine of salt-loaded humans. Besides the *in vitro*-assay of the inhibitory activity, we also demonstrated the inhibitory effect of this serum fraction on epithelial sodium transport, i.e., on

short-circuit current (SCC) and potential difference (PD) in the isolated frog skin (8). This fraction of serum or urine was also found to cause natriuresis in a rat bioassay. We concluded that a natriuretic factor emerges in the circulation with excessive salt load whose mechanism of action was to modulate the Na-K-ATPase enzyme in the vasculature as well as in the renal tubule.

Attempts to identify this humoral inhibitor(s), however, remained unsuccessful despite the use of extensive methodologic approaches. Besides an endogenous ouabain (9), several bufadienolides (10) have been identified using the methods described in this paper of which two compounds with  $M_r$  of around 400 daltons will be considered in the present mini-review, namely vanadium (V)-diascorbate(s) and derivatives of xanthurenic acid (4). The compounds were assumed to be present in the circulation, and therefore may be excreted in the urine.

## Ouabain-Like Factor(s) as Endogenous Sodium Transport Inhibitors

As source for isolation and identification of a putative natriuretic hormone and/or endogenous epithelial sodium transport enzyme inhibitor, we pooled large quantities (50–100 L) of urine from salt-loaded healthy humans, which was lyophilized to dryness and reconstituted with 0.01M acetic acid, and then subjected to gel chromatography using Sephadex G-25 and Sephadex G-10 columns.

### Ouabain-Like Factors and Vanadium-Diascorbic Acid: Effects on Na-K-ATPase

To detect the serum and urine fractions with the active compound(s), we employed an *in vitro*-assay of Na-K-ATPase using a purified commercially available hog cerebral enzyme preparation. We also used this Na-K-ATPase membrane fraction as marker to follow-up activity with purification steps during the subsequent chromatographic steps. To detect the potential natriuretic activity, all fractions were screened for their natriuretic effect using a bioassay in conscious rats (11).

The transport enzyme Na-K-ATPase inhibitory and natriuretic activity(ies) eluted from the Sephadex G-25 column in a post-salt fraction. When this fraction was then subjected to gel chromatography on Sephadex G-10, a strongly active enzyme inhibitory material eluted in a late fraction (1). This late fraction also showed a significant natriuretic action (11). This enzyme inhibitory and natriuretic fraction was subjected to high performance liquid chromatography (HPLC), and subsequently to thin layer chromatography (TLC). Characterization of the active material was attempted by mass ( $M_r$ )-, nuclear magnetic resonance ( $^1\text{H-NMR}$ )-, infrared (IR)-spectroscopy (1), and ultraviolet (UV)-fluorescence/absorbance. The natriuretic activity was also studied by bioassay to identify the active compounds after gel filtration, reverse phase (RP)-HPLC, and amino acid analysis for its potential peptidic character (11).

Reverse-phase HPLC of this highly active late fraction from Sephadex G-10 resulted in two subfractions with significant Na-K-ATPase enzyme inhibition. They were named ouabain-like factors (OLF); one eluted in the water phase as the more polar hydrophilic OLF-1; the second eluted in a later phase at 20% acetonitrile as the more apolar lipophilic OLF-2. These fractions also produced a significant natriuresis (see below).

### Analysis of Chemical Structure

Both compounds showed signals for hydroxyl and carboxyl groups as well as criteria for esters or lactones (a precursor of ascorbic acid in plants and animals is L-gulono- $\gamma$ -lactone, and 2,3-diketogulonic acid is an oxidation product of ascorbic acid). No signals for aromatic, aliphatic, heterocyclic, or steroid structures were found. Whereas the IR-spectrum of OLF-1 is different from that of OLF-2 (1), UV-,  $M_r$ -, and  $^1\text{H-NMR}$ -criteria were similar and fluorescence of both compounds when separated by TLC required the presence of a dicarboxylic acid-like conformation; dicarboxylic acid [see also Ref. (11): Asp, Glu as carboxylic acids] is an organic compound containing two carboxyl functional groups ( $-\text{COOH}$ ). IR- and  $^1\text{H-NMR}$  spectra of OLF-1 and OLF-2 suggest a chemical structure resembling a sugar or sugar derivative. However, sugars are not fluorescent as are the OLF recovered from TLC. Therefore, these data suggest the unknown compounds to be identical with ascorbic acid or its salts such as V<sup>+</sup>-diascorbate and V<sup>IV</sup>-diascorbate, respectively, with  $M_r$  403 (3). The superscript roman numbers indicate the oxidative state of vanadium (V): V<sup>IV</sup> oxide ( $\text{V}_2\text{O}_5$ ), the most stable oxygen combination, and V<sup>IV</sup> oxide ( $\text{VO}_2$ ) represent two of the four oxygen states of vanadium. V-diascorbates elute from the RP-HPLC column at similar elution times and acetonitrile gradients as the hydrophilic and lipophilic OLF-1 and OLF-2, respectively. V<sup>IV</sup>-diascorbate also showed the same UV-maximum as we found for OLF. Thus, ascorbic acid seems to be an important cornerstone of the structure of the yet unknown humoral ATPase inhibitor.

It is noteworthy that the water solubility of the individual ascorbic acid salts of metals varies remarkably, and it may be assumed that V<sup>+</sup>- and V<sup>IV</sup>-diascorbates with their different water solubility elute from the RP-HPLC column at similar elution times as the OLF-1 and OLF-2, respectively. V-diascorbates also show the same UV-maximum as the OLF and are strong candidates for the urinary hydrophilic OLF-1 and lipophilic OLF-2, respectively.

### Effects on Enzyme Kinetics

These active subfractions, containing OLF-1 and OLF-2, were further purified by two-dimensional preparative TLC to single compounds, whose mass spectroscopic (MS) data suggested a  $M_r$  of around 400. Actually, OLF-2, which dose-dependently inhibited Na-K-ATPase, was found to have a  $M_r$  of 391 (1). With respect to the effects of OLF-1 and OLF-2 and of V<sup>+</sup>- and V<sup>IV</sup>-diascorbates on Na-K-ATPase enzyme activity and kinetics, *in vitro* studies showed that OLF-1 and OLF-2 inhibited the enzyme in its E2 configuration. In analogy to the polar OLF-1, which revealed an approximately 10-fold stronger enzyme inhibition ( $\text{IC}_{50}$   $1.5 \times 10^{-5}$  M) than the apolar OLF-2 ( $\text{IC}_{50}$   $1.5 \times 10^{-4}$  M), we found that V<sup>+</sup>-diascorbate ( $\text{IC}_{50}$   $2 \times 10^{-6}$  M) is a significantly stronger inhibitor of Na-K-ATPase than V<sup>IV</sup>-diascorbate ( $\text{IC}_{50}$  of  $9 \times 10^{-5}$  M) (3, 5, 12). In this context, I should mention that we found previously that certain trace metals are strong inhibitors of this enzyme (13).

### Renal and Vascular Mechanisms of Action of OLF

Regarding the potential mechanism of the physiological and pathological effects of OLF-1 and OLF-2 on vascular smooth muscle cells (VSMCs) and inner medullary collecting duct cells (IMCD cells), we found in an *in vitro*-assay that OLF-1 and OLF-2 enhanced

VSMC contractility by increasing intracellular  $\text{Ca}^{2+}$  similar to the effect of ouabain (14, 15). Similar effects were found with OLF-1 and OLF-2 on intracellular  $\text{Ca}^{2+}$  in IMCD cells, suggesting inhibition of tubular Na-reabsorption and thus regulating renal excretion, i.e., to enhance Na-excretion (16).

### Ouabain-Like Factors and V-Diascorbates: Natriuretic Effects

For demonstration of the natriuretic activity, we used a bioassay in conscious rats (12). As mentioned above, in our assay system, the post-salt fraction IV from Sephadex G-25 was applied to Sephadex-G-10 and resulted in a late fraction, which was applied to RP-HPLC. When administered i.v., OLF-1 resulted in an immediate, eightfold rise in natriuresis from approximately 1 to 8  $\mu\text{Eq}/\text{min}/\text{mg}$ , whereas the apolar OLF-2 caused a natriuresis of slower onset reaching its maximum after 60 min and lasting for more than 180 min. This was confirmed also by injection of the active fractions obtained by quantitative TLC.

### Natriuretic Factor Unrelated to OLF

Finally, I should mention that we described previously a natriuretic compound, which we suggested to be a peptide. Thus, when the pooled post-salt natriuretic urine fraction obtained by gel chromatography (see above) was subjected to repetitive RP-HPLC, a late eluting fraction showed strong natriuretic activity in the bioassay and was associated with a fluorescence peak when treated with o-phthalodialdehyde as a marker for primary amines (11). Amino acid analysis before and after total acid hydrolysis suggested a peptide tentatively containing the amino acids (AA) Asp, Glu, Gly, Phe, and Ser (1, 11). The natriuretic activity was lost after incubation with chymotrypsin, which splits bonds with aromatic AA (2). We found, in addition, that several synthetic (mono-) peptides of di- and tri-AA are significantly natriuretic when injected i.v. (unpublished data).

### Xanthurenic Acid 8-O- $\beta$ -D-Glucoside and Xanthurenic Acid 8-O-Sulfate as Endogenous Sodium Transport Inhibitors

Cain et al. (4) followed a protocol very similar to that of Kramer et al. for isolation of the natriuretic activity except that they used the urine of uremic patients as source of the inhibitor and a bioassay in (conscious?) uremic rats. As marker for the active material, Cain et al. used changes of the SCC of the isolated frog skin – as we described in 1977 (8) – for monitoring transepithelial sodium transport inhibitory activity. For monitoring its natriuretic effect, the above mentioned bioassay in uremic rats was used. A direct *in vitro*-assay for inhibition of the Na-K-ATPase enzyme by the natriuretic factor or “hormone” was not employed. The authors rather speculate that the natriuretic hormone may act via other sodium pumps in the kidney, e.g., the epithelial sodium channel (ENaC) in the distal tubule.

### Xanthurenic Acid Derivatives: Effects on Epithelial Sodium Transport

Epithelial sodium transport was measured as changes of SCC and PD in the isolated frog skin. For isolation and identification

of the transport inhibitor, one gel chromatographic step and three consecutive HPLC steps were applied. Final identification was achieved by mass (Ms)-, IR-, UV-, and NMR-spectroscopy. Purification of the activity was estimated from UV peak with a characteristic spectrum at 338 nm. Xanthurenic acid 8-O- $\beta$ -D-glucoside ( $M_r$  368) and xanthurenic acid 8-O-sulfate ( $M_r$  284) were identified as the endogenous sodium transport (ENaC, Na-K-ATPase) inhibitors.

### Xanthurenic Acid Derivatives: Natriuretic Effects

The material ( $M_r$  368) obtained from two HPLC runs was tested for natriuretic effect in their uremic rat bioassay. Urinary sodium excretion rose immediately and reached its maximum approximately 40 min after intra-arterial infusion (5). Urinary volume increased slightly and then decreased to below baseline, i.e., a decrease in urine volume with a rise in urinary osmolality.

A pathophysiological role of xanthurenic acid, a tryptophane derivative, is difficult to envisage as this uremic toxin may inhibit transmembranous sodium transport independent of a potential role as specific circulating natriuretic or sodium transport inhibiting “hormone.” Thus, although Bricker et al. showed that the natriuretic action of the isolated inhibitor paralleled the changes in renal function, as an alternative explanation, it may be reasonable to assume that with the progressive decrease in renal function and the accumulation of toxic metabolites, the rise in fractional sodium excretion may parallel the urinary concentration of the xanthurenic derivatives.

### Summary

Although there is no doubt that an as yet unidentified natriuretic compound can be isolated from human urine by gel filtration and RP-HPLC, whose activity changes in parallel with salt (sodium chloride) intake, i.e., it correlates with salt-balance (low or high salt intake). Therefore, the activity may be related to an as yet unidentified “natriuretic hormone” that is assumed to play a crucial role in the fine-tuning of renal tubular sodium handling and may thus be involved in the long-term body fluid and blood pressure regulation.

We found two Na-K-ATPase inhibitors, the hydrophilic OLF-1 and the lipophilic OLF-2 (1). The hydrophilic form was more potent than the lipophilic one. The lipophilic compound was moderately natriuretic but strong ATPase inhibitor. Both compounds showed UV fluorescence/absorbance of lower intensity in the hydrophilic (hydrated) form. Both enzyme inhibitors showed UV-absorbance, which requires the presence of a dicarboxylic acid-like arrangement (1). In addition, from our data the unknown compound(s) most likely fulfill(s) the criteria for lactones.

Unfortunately, for none of the three classes of endogenous sodium transport inhibitors, a physiologic or pathophysiologic role was demonstrated, i.e., no correlation to body fluid and sodium balance or blood pressure was documented. Thus, further studies are required to confirm the structures of the various endogenous factors; final identification of their physiological and pathophysiological significance must await urgent results of additional well-designed studies.

## References

- Kramer HJ, Krampitz G, Bäcker A, Michel H, Krampitz G Jr. Meyer-Lehnert H. Endogenous sodium pump inhibitors in human urine. Further identification of inhibitors of Na-K-ATPase. *Am J Hypertens* (1995) **8**:753–60. doi:10.1016/0895-7061(95)00125-9
- Kramer HJ, Meyer-Lehnert H, Michel H, Predel HG. Endogenous natriuretic and ouabain-like factors. Their roles in body fluid and blood pressure regulation. *Am J Hypertens* (1991) **4**:81–9.
- Kramer HJ, Krampitz G, Bäcker A, Meyer-Lehnert H. Ouabain-like factors in human urine: identification of a Na-K-ATPase inhibitor as vanadium-diascorbate adduct. *Clin Exp Hypertens* (1998) **20**(5&6):557–71. doi:10.3109/10641969809053234
- Cain CD, Schroeder FC, Shankel SW, Mitchnick M, Schmertzler Bricker NS. Identification of xanthurenic acid 8-O-β-D-glucoside and xanthurenic acid 8-O-sulfate as human natriuretic hormones. *Proc Natl Acad Sci U S A* (2007) **104**:17873–8. doi:10.1073/pnas.0705553104
- Kramer HJ, Gonick HC, Paul W, Lu E. Third factor: inhibitor of Na-K-ATPase? Abstracts free comm. *IVth International Congress of Nephrology*. Stockholm, Sweden (1969). p. 373
- Kramer HJ, Gonick HC. Effects of extracellular volume expansion on renal Na-K-ATPase and cell metabolism. *Nephron* (1974) **12**:281–96. doi:10.1159/000180341
- Gonick HC, Kramer HJ, Paul W, Lu E. Circulating inhibitor of sodium-potassium-activated triphosphatase after expansion of extracellular fluid volume in rats. *Clin Sci Mol Med* (1977) **53**:320–34.
- Kramer HJ, Bäcker A, Krück F. Antinatriferic activity in human plasma following acute and chronic salt-loading. *Kidney Int* (1977) **12**:214–22. doi:10.1038/ki.1977.103
- Hamlyn JM, Blaustein MP. Salt sensitivity, endogenous ouabain and hypertension. *Curr Opin Nephrol Hypertens* (2013) **22**:51–8. doi:10.1097/MNH.0b013e32835b36ec
- Bagrov AY, Shapiro JI, Federova OV. Endogenous cardiotonic steroids: physiology, pharmacology, and novel therapeutic targets. *Pharmacol Rev* (2009) **61**:9–38. doi:10.1124/pr.108.000711
- Kramer HJ, Heppe M, Weiler E, Bäcker A, Liddiard C, Klingmüller D. Further characterization of the endogenous natriuretic and digoxin-like immunoreacting activities in human urine. Effects of changes in sodium intake. *Ren Physiol* (1985) **8**:80–9.
- Kramer HJ, Krampitz G, Bäcker A, Meyer-Lehnert H. Vanadium diascorbates are strong candidates for endogenous ouabain-like factors in human urine: effects on Na-K-ATPase enzyme kinetics. *Biochem Biophys Res Commun* (1995) **213**:289–94. doi:10.1006/bbrc.1995.2128
- Kramer HJ, Gonick HC, Lu E. In vitro-inhibition of Na-K-ATPase by trace metals. Relation to cardiovascular and renal damage. *Nephron* (1986) **44**:329–36. doi:10.1159/000184015
- Meyer-Lehnert H, Wanning C, Michel H, Bäcker A, Kramer HJ. Cellular mechanisms of action of a ouabain-like factor in vascular smooth muscle cells. *J Cardiovasc Pharmacol* (1993) **22**(Suppl 2):S16–9. doi:10.1097/00005344-199312000-00035
- Meyer-Lehnert H, Bäcker A, Kramer HJ. Inhibitors of Na-K-ATPase in human urine: effects of ouabain-like factors and of vanadium-diascorbate on calcium mobilization in rat vascular smooth muscle cells. Comparison with the effects of ouabain, angiotensin II, and arginine-vasopressin. *Am J Hypertens* (2000) **13**:364–9. doi:10.1016/S0895-7061(99)00197-1
- Kramer HJ, Bäcker A, Meyer-Lehnert H. Effects of urinary ouabain-like factor (OLF) and vanadium-diascorbate on calcium mobilization in inner medullary collecting duct (IMCD) cells. *Am J Hypertens* (1998) **11**:1208–13. doi:10.1016/S0895-7061(98)00134-4

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Kramer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Evidence for a 12 kDa “carrier protein” for natriuretic hormone

Harvey C. Gonick\*

Division of Nephrology, Department of Medicine, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA, USA

**Edited by:**

Erik Maronde, Goethe University Frankfurt am Main, Germany

**Reviewed by:**

Erik Maronde, Goethe University Frankfurt am Main, Germany

Eric W. Roubos, Radboud University Nijmegen, Netherlands

**\*Correspondence:**

Harvey C. Gonick, 201 Tavistock Avenue, Los Angeles, CA 90049, USA  
e-mail: hgonick@ucla.edu

The search for the elusive Na-K-ATPase-inhibiting natriuretic hormone continues. In this review, evidence is presented that isolating the carrier protein for natriuretic hormone from hypertensive plasma is a necessary first step before splitting off the final hormone. The carrier protein has a molecular weight of 12 kDa while the final hormone has a molecular weight of 408 Da. Both compounds inhibit Na-K-ATPase but the compound containing the carrier protein predominates. The question has been raised as to whether the carrier protein is in actuality proANF, a 17 kDa protein that can be split between a 14 kDa protein (the presumptive proANF) and the 3 kDa ANF.

**Keywords:** natriuretic hormone, carrier proteins, hormones, hypertension, review, Anf, proANF

## INTRODUCTION

Circulating inhibitors of sodium-potassium adenosine triphosphatase (Na-K-ATPase) have been shown to be of possible pathogenetic importance in the mechanism of essential hypertension (1–3). Although previous studies have demonstrated the presence of both high-molecular weight (HMW), ranging from 11 to 70 kDa (4–8) and low-molecular weight (LMW) either natriuretic or Na-K-ATPase inhibitors, no previous attempts had been made to ascertain whether HMW or LMW forms predominate in hypertension. This review summarizes the steps taken by our laboratory to first identify the HMW form, and then split off the final LMW form of the hormone. We have in the process determined the approximate molecular weight of the HMW form and the precise molecular weight of the LMW form. Unfortunately, while awaiting the identification of the latter compound, it was lost due to freezer failures in two different laboratories a continent apart. This review is presented in intricate detail in the hopes of encouraging subsequent investigators to pursue the final identification of the LMW natriuretic hormone, as well as the identity of the “carrier protein.”

## PREDOMINANCE OF HMW PLASMA Na-K-ATPase INHIBITOR IN HYPERTENSION

In an initial study (9), plasma samples obtained from 26 patients with essential hypertension, 12 normotensive controls, and 6 normotensives with a family history of hypertension were separated into HMW and LMW moieties by passage through a 1 kDa Amicon membrane. The LMW moiety was separated on C-18 Sep-Pak cartridges, applying a 10% stepwise acetonitrile trifluoroacetic acid gradient. The HMW moiety was further separated on Sephadex G-75. Sodium dodecyl sulfate polyacrylamide gel (SDS-PAGE) electrophoresis revealed that the fraction with inhibitory activity contained a distinct 12 kDa protein band, with staining intensity depending on the presence or absence of hypertension (Figure 1). Na-K-ATPase inhibitory activity was found in several LMW fractions, but differences between hypertensives and normotensives were observed in only the 50% acetonitrile fraction ( $0.29 \pm 0.12$

SD versus  $0.11 \pm 0.12 \mu\text{mol/L}$  ouabain equivalents,  $p < 0.01$ ). Na-K-ATPase inhibitory activity in the HMW fraction was 38 times the inhibitory activity in the LMW fraction and was significantly increased in hypertensives as compared to normotensive controls ( $10.9 \pm 8.9$  versus  $1.3 \pm 0.8 \mu\text{mol/L}$  ouabain equivalents,  $p < 0.01$ ). Inhibitory activity in both HMW and LMW fractions correlated positively with mean blood pressure ( $r = 0.42$ ,  $p < 0.05$  and  $r = 0.35$ ,  $p < 0.05$ ). The inhibitory activity in the HMW fraction, but not the LMW fraction, also correlated positively with diastolic blood pressure and inversely with the natural log of plasma renin activity ( $r = 0.40$ ,  $p < 0.01$ ). These results indicate that the HMW moiety is the predominant circulating form of the Na-K-ATPase inhibitor in hypertension.

## DISSOCIATION OF THE LMW Na-K-ATPase INHIBITOR FROM THE HMW PROTEIN INHIBITOR

Pooled blood samples from 10 patients with well-documented essential hypertension, not taking any medications for at least 3 weeks, were collected into chilled vacutainers containing sodium ethylenediamine tetraacetic acid (EDTA) and Trasylol (10). Individual samples were also collected from patients with primary aldosteronism, congestive heart failure (CHF), before and after treatment, and normal controls. The treatment of congestive failure employed diuretics and vasorelaxants but avoided digitalis glycosides.

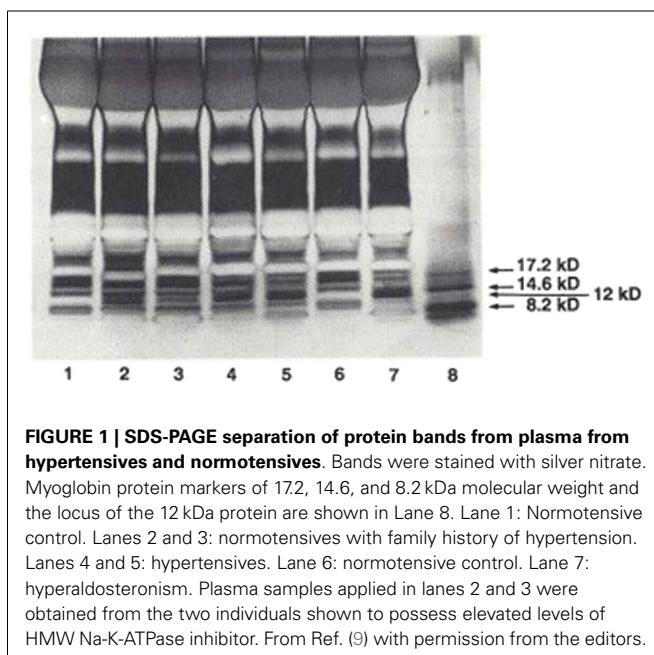
SDS-PAGE was performed according to the procedure described by Laemmli (11).

Plasma samples were also passed through a series of Amicon membranes, the initial ultrafiltration step employing a 1 kDa (YM-2) membrane. The retentate was reconstituted in distilled water and heated for 10 min at 70°C in the presence of 4% beta-mercaptopethanol and 1 mol/L formic acid. The solution was cooled down and subsequently placed on a 30 kDa (YM-30) membrane. The resulting filtrate, containing the dissociated protein, was lyophilized and subjected to further purification.

The dissociated protein was adsorbed onto a SEP-PAK C-18 cartridge. Interfering compounds, e.g., small peptides, hydrophobic

substances, etc., were retained on the SEP-PAK C-18 cartridge. The protein of interest was eluted off the SEP-PAK C-18 cartridge with distilled water. This fraction was lyophilized, reconstituted in 1 mL of distilled water, and subsequently separated on Sephadex G-75. The plasma preparation was eluted off the column with 10 mmol/L ammonium acetate, pH 6.5. Fractions containing the Na-K-ATPase inhibitory material (12 kDa protein), which eluted after the albumin peak, were pooled, lyophilized, and subjected to a series of assays.

Duplicate bioassay procedures for the natriuretic response of the 12 kDa protein were performed according to the method described by Purdy et al. (12). Outcomes were averaged. Results are displayed in **Figure 2**.

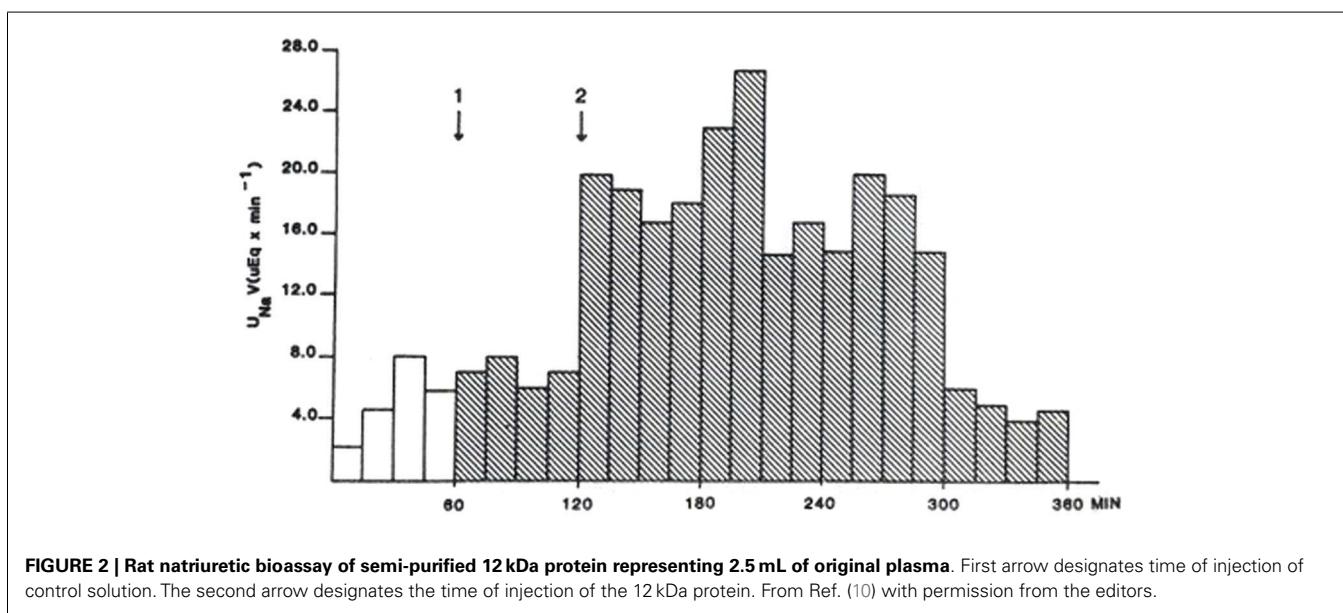


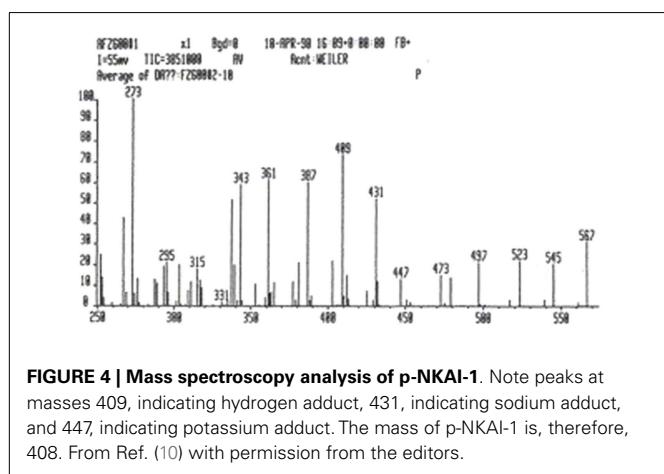
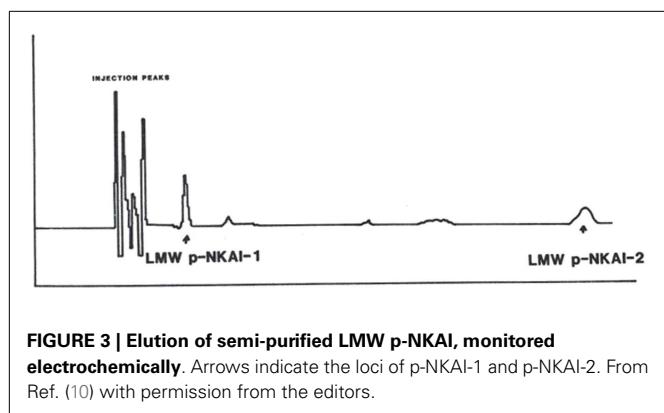
An Econosphere C-18 column was packed with Econosphere C-18 silica, 5  $\mu$ m particle size. The reversed phase C-18 column was equilibrated with triple distilled water. The LMW plasma Na-K-ATPase inhibitor (p-NKAI) was eluted off the column with a linear acetonitrile gradient (0–100% over a period of 30 min). The eluate was continuously monitored at 210 nm. One minute fractions were collected, lyophilized, and subsequently tested for the presence of Na-K-ATPase inhibitory activity.

P-NKAI was further purified by HPLC separation combined with electrochemical detection using a Model 5100A Coulocomb Detection System. On reversed phase C18 chromatography, p-NKAI appeared at 4% acetonitrile, co-eluting with a urinary inhibitor. P-NKAI was ultrafiltrable through an Amicon YM-05 membrane and thus has a presumed molecular weight of less than 500 Da. Rechromatography of active fractions on a 3  $\mu$ m C-18 column monitored electrochemically yielded two active compounds, p-NKAI-1 and p-NKAI-2, both of which were inhibitors of the Na-K-ATPase enzyme system (**Figure 3**). P-NKAI-1 caused 50% inhibition and p-NKAI-2 caused 8% inhibition of Na-K-ATPase in a volume of inhibitor corresponding to 187  $\mu$ L of original plasma. The remaining fractions were without inhibitory activity.

The mass spectrum of p-NKAI-1 showed a fairly intense protonated molecular ion at mass 409 and also the sodium and potassium adduct ions at masses 431 and 447, respectively. This would indicate that the molecular weight of p-NKAI-1 is 408 Da (**Figure 4**).

A purified hog cerebral cortex Na-K-ATPase preparation was employed for Na-K-ATPase and K-pNPPase inhibition assays. The tubes were preincubated with either the 12 kDa protein or the purified LMW plasma factor for 5 min at 37°C. The enzymatic reaction was initiated by adding 0.025 mL enzyme preparation (25 mg/mL). The reaction was stopped by adding 1.0 mL ice cold 10% trichloroacetic acid after an incubation time of 15 min. After centrifugation, 0.5 mL of supernatant was assayed for inorganic phosphate according to the procedure described by Fisk and Subbarow (13). Both the 12 kDa protein and the LMW plasma factor

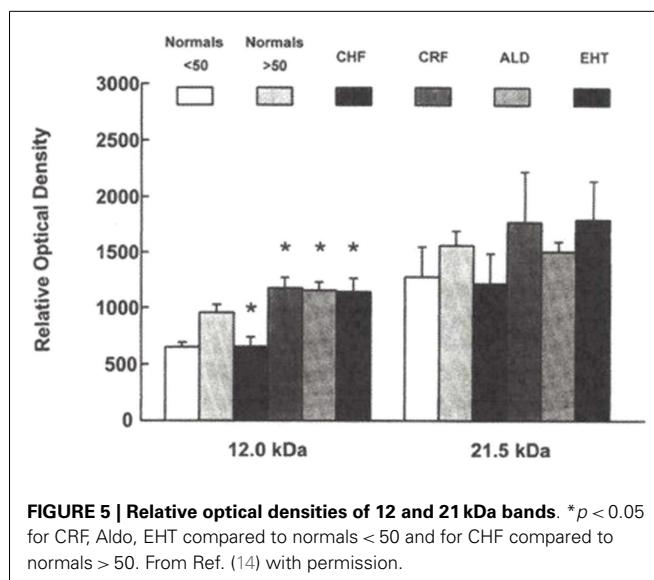




(p-NKAI) were shown to inhibit the Na-K-ATPase and K-pNPPase enzyme systems in a dose-related manner, analogous to ouabain. The IC<sub>50</sub> for inhibition of Na-K-ATPase by p-NKAI corresponds to  $8 \times 10^{-7}$  mol/L ouabain equivalents.

P-NKAI-1 was also tested for its vasoactive properties according to the procedure described by Purdy and Weber (14). Isolated femoral artery segments from New Zealand White rabbits were sectioned into 3 mm segments, then mounted in a 30 mL tissue bath containing Krebs-bicarbonate solution aerated continuously with 95% O<sub>2</sub>/5% CO<sub>2</sub> at 37°C. Subsequently, p-NKAI-1 was assayed for its vasoactive behavior in the presence and absence of norepinephrine. A dose-response curve was established for p-NKAI-1; the concentration of p-NKAI-1 yielding 1% contractile response was selected for the studies of synergy with norepinephrine. One hundred microliters of p-NKAI-1 produced a 1% contractile response, 300 μL produced a 5% contractile response and 600 μL of p-NKAI-1 produced an 18% contractile response. Similarly, the addition of 100 μL of p-NKAI-1 to a bath containing  $10^{-8}$  mol/L norepinephrine increased the contractile response from 60 to 86%.

The dose-response curve for Na-K-ATPase inhibition of the semi-purified 12 kDa protein paralleled the dose-response curve for ouabain; 50% inhibition of Na-K-ATPase, corresponding to  $5 \times 10^{-6}$  mol/L ouabain, was produced by the 12 kDa inhibitor in a fraction containing 2.7 mg/mL Lowry protein.



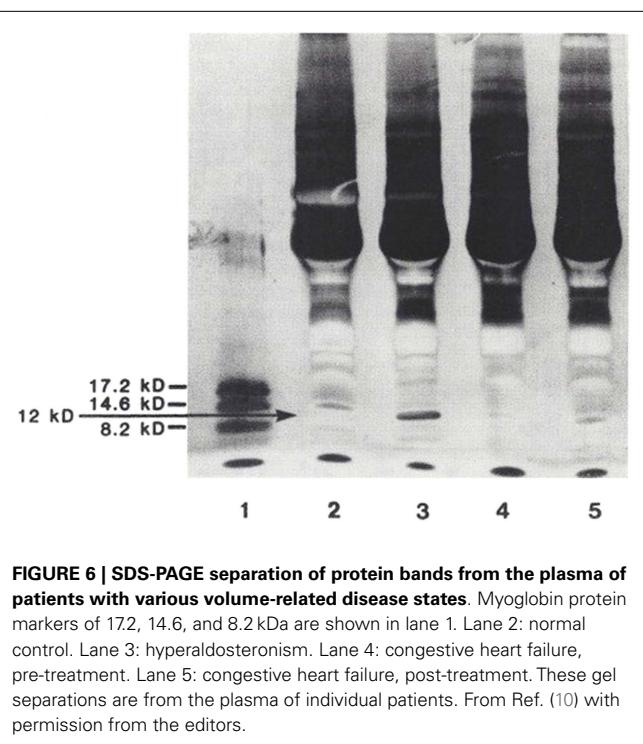
The <sup>3</sup>H ouabain displacement assay revealed that the 12 kDa protein fraction displaces <sup>3</sup>H-ouabain from its receptor in a dose-related manner, similar to ouabain. There was no cross-reactivity with digoxin antibody.

### COMPARISON OF 12 kDa PROTEIN, MARINOBUFAGENIN, AND OUABAIN IN VARIOUS DISEASE STATES

In a third study (15), plasma from 101 patients were examined [25 normals (N) < age 50, 13N > age 50, 7 with acute CHF, 24 with chronic renal failure (CRF), on dialysis, 5 with idiopathic hyperaldosteronism (PA), and 27 with essential hypertension, untreated (EHT)]. Plasma was extracted with 32% acetonitrile, and analyzed by fluoroimmunoassay (DELFIA) for marinobufagenin and ouabain. In addition, from 32 patients (6N < 50, 6N > 50, 5 CHF, 5 CRF, 6 EHT, and 4 PA), SDS gradient gels were obtained. The 12 kDa bands were extracted, analyzed for Na-K-ATPase inhibition, marinobufagenin, and ouabain, and compared to 14 and 21 kDa bands. Marinobufagenin was found to be elevated in CRF, EHT, PA, and CHF. Ouabain was increased only in PA. When the relative optical densities of 12 and 21 kDa bands were contrasted, CRF, PA, and HT were found to be increased and CHF to be decreased in the 12 kDa band, with no discernible changes in the 21 kDa bands (Figure 5). Following extraction of the bands, Na-K-ATPase inhibitory activity measured 38% in 16 pooled 12 kDa bands, with essentially no activity found in the 14 kDa or 21 kDa bands. SDS-PAGE separation of plasma proteins confirmed that the 12 kDa band was elevated in primary aldosteronism, diminished in CHF, with return toward normal after treatment (Figure 6). Thus, only the 12 kDa band possessed all of the attributes of natriuretic hormone.

### DISCUSSION

Following an initial flurry of activity, which utilized natriuretic as an index of hormone activity, most subsequent studies of natriuretic hormone have utilized Na-K-ATPase inhibition as a more rapidly obtained index (1–3). Digitalis-like (EDLF) or



**FIGURE 6 | SDS-PAGE separation of protein bands from the plasma of patients with various volume-related disease states.** Myoglobin protein markers of 17.2, 14.6, and 8.2 kDa are shown in lane 1. Lane 2: normal control. Lane 3: hyperaldosteronism. Lane 4: congestive heart failure, pre-treatment. Lane 5: congestive heart failure, post-treatment. These gel separations are from the plasma of individual patients. From Ref. (10) with permission from the editors.

ouabain-like activities (OLF), measured by radioimmunoassay, were also initially employed as measures of natriuretic hormone. But digoxin-like immunoreactivity was found to be non-specific (16), while the radioimmunoassay for OLF did not prove reliable when measured by HPLC followed by ELISA (17), or by ultrasensitive UPLC-MS/MS (18). The lower limit of quantification by the latter method was 1.7 pmol/L, while ouabain was non-detectable. The suggestion that the presence of endogenous ouabain in human beings is non-detectable has been vigorously debated by Blaustein (19). For the moment, therefore, we must consider this an unresolved matter. Thus, we are left with the Na-K-ATPase inhibition assay as presumably the most reliable as well as the most rapid assay of EDLF activity.

The remaining ouabain-like hormone, which has been suspected to be the putative natriuretic hormone, is marinobufagenin (20), for which studies of activity in several diseases have appeared, including volume-expanded normals (20), CHF (21), CRF (22), essential hypertension (23), primary aldosteronism (15), and pre-eclampsia (24). The one noteworthy discrepancy between natriuretic hormone determined by marinobufagenin radioimmunoassay and natriuretic hormone, as determined by the Na-K-ATPase assay, are the findings in CHF [see above and Ref. (25)]. Urinary sodium values are low in CHF, LMW urinary Na-K-ATPase inhibitors are also lower than normal (25), and arterial central volume is diminished rather than increased. Kramer and Kruck (26) found that a natriuretic substance present in an ultrafiltrate of normal urine from volume-expanded individuals was absent in the urine of patients with edema related to cirrhosis with ascites or with nephrotic syndrome, edematous states physiologically similar to CHF. Furthermore, they also demonstrated

that plasma and urine fractions of normal individuals following Sephadex G-25 separation consistently reduced short-circuit current when applied to the serosal surface of frog skin (anti-natriuretic effect) (26), whereas plasma and urine fractions from patients with edema lacked this effect. In addition, we have shown previously that although the LMW Na-K-ATPase inhibitor in human urine has less activity than normal in CHF, the activity reverts toward normal as CHF improves (25).

The radioimmunoassay for marinobufagenin has been recently validated by high resolution mass spectrometry but has been measured only in CRF, where it is elevated (27). Until the CHF results are similarly verified, it is not possible to be sure that radioimmunoassay results for marinobufagenin in disease states other than CRF also reflect the true status.

We suspect that the HMW Na-K-ATPase inhibitor may be a carrier protein for the LMW inhibitor since the latter can be split off by use of beta-mercaptoethanol, an agent known to cleave S-S bonds, plus heat and formic acid, properties employed by Lindner et al. (28) to dissociate oxytocin and vasopressin from their neurophysin carrier. It is also pertinent that Morich and Garthoff (5) found that both salt-sensitive (DS) and salt-resistant (DR) rats displayed two protein bands in their plasma on SDS-PAGE, in the molecular weight range of 14–15 kDa. When DS rats were given salt and developed hypertension, the upper band diminished but the lower band became more intense. The difference in molecular weight between the two bands was estimated to be between 300 and 400 Da. Mass spectrometry of the first of the LMW inhibitors in the present study (NKAI-1) revealed a molecular weight of 408 Da, as shown by the hydrogen adduct of 409 Da, the sodium adduct of 431 Da, and the potassium adduct of 447 Da. The molecular weight of 408 is identical to that described by Kerek (29), a biochemist, for an initially identified macrocyclic derivative of inorganic carbon suboxide, which is a natriuretic, Na-K-ATPase inhibiting compound derived from plant tissue. We look forward with interest to the comparison between Kerek's 408 Da compound and the 408 Da compound discussed in this review.

The HMW compound of the present dissertation was previously referred to as “hypertension-associated protein” by Van de Voorde et al. (7). These authors claimed an approximate molecular weight of 15 kDa for the compound they isolated by chromatography after reduction of the disulfide bridges of the precursor 105 kDa protein molecule with beta-mercaptoethanol. In a prior study of plasma proteins in essential hypertension, utilizing SDS-PAGE to separate the plasma proteins, Nardi et al. (4) had earlier reported a 14 kDa protein present in such patients but not in patients with hypertension secondary to renovascular hypertension or renal parenchymal disease. Cloix et al. (6) had reported a 13 kDa protein in the plasma of hypertensive human beings and rats. Thus, we are left with four studies that purport to show either 12, 13, 14, or 15 kDa proteins in the plasma of human beings with essential hypertension but not in normal controls or possibly in renovascular hypertension or hypertension with CRF. What could this protein be? In the present study, we have referred to the 12 kDa protein as a “carrier protein” because the Na-K-ATPase inhibitor can be split off by heat and formic acid. But are there alternatives?

To explore this question in all of its ramifications, it is first necessary to review what has been learned about the “other” natriuretic system, namely the natriuretic peptides. Following the initial description of natriuretic peptides by deBold and associates in 1961 (30), it has been found that there are at least three natriuretic peptides released from the hypothalamus and cardiac tissue – atrial natriuretic factor (ANF), B-type natriuretic factor (BNF), and C-type natriuretic factor (CNF). All occur initially as pre-prohormones, which are degraded to prohormones and then finally to the active peptides (31). The molecular weight of the proANF, a circulating compound (32), has been described as 14 kDa (33). Is it possible that pro-ANF is identical to the hypertension-associated protein described by Van de Voorde et al. (7), Nardi et al. (4), Cloix et al. (6), and the present study? A suggestion that this may be the case comes from Melander et al. (34) who described in offspring of hypertensive human beings a strong correlation between salt sensitivity, as defined by the difference in sodium excretion while on a low salt diet and then on a high salt diet, and plasma proANP levels.

Initially, it was thought that EDLF, endogenous digitalis-like factor, or OLF, ouabain-like factor, as the Na-K-ATPase inhibitor became known, could be distinguished physiologically from ANF by its Na-K-ATPase inhibiting property as well as its tendency to increase, rather than decrease, vasoconstriction when applied to isolated blood vessels (10). However, it was recognized by Górný et al. (35) that ANF does inhibit Na-K-ATPase in the rat renal medulla, but not in the rat renal cortex, where the proximal tubule is located. In contrast, Chiou and Vesely (36) reported that kaliuretic peptide, a fraction split off from ANF prohormone, inhibits both renal cortical and medullary Na-K-ATPase. However, these experiments employed rat renal tissue rather than hog cerebral cortex for assay of Na-K-ATPase and the inhibition in the two studies quoted resulted from indirect inhibition of Na-K-ATPase through effects of second messengers, namely, dopamine in the first study (35) and prostaglandin E<sub>2</sub> in the second study (36). Thus, we may no longer be able to depend exclusively on the Na-K-ATPase assay to distinguish between ANF and EDLF. On the other hand, we can still depend on both the molecular weight and the direct vasoconstrictive (10) or vasodilatory (37) actions on isolated vascular smooth muscle preparations to distinguish between EDLF and ANF. The molecular weights for EDLF have been reported as varying between 360 and 620 Da (Table 1), while the molecular weights for ANF have been described as 3800 Da for rat ANF (38) and varying from 3000 Da (33) to 5499 Da (39) for human ANF.

Haupert (44) in 1988 first posed the question as to whether there is an interrelationship between natriuretic peptides and EDLF or OLF. That the interrelationship exists can no longer be in doubt. It has long been recognized that both ANF and EDLF are released from the hypothalamus (45, 46), and in fact from the AV3V region (47). Lesions produced in the AV3V region prevent the natriuresis following isotonic saline volume expansion in experimental animals. Furthermore blood drawn following expansion failed to show an anti-natriferic effect in the toad bladder in contrast to control animals, implying interference with release of the Na-K-ATPase inhibitor. The perfusate from incubation of fragments of rat brain inhibited the Na, K pump by a

**Table 1 | Comparison of sources and molecular weights of various EDLFs.**

Author	Source	Molecular weight (daltons)	Reference
Bricker et al.	Human uremic urine	360	(40)
McKinnon et al.	Human placenta	370	(41)
Kramer et al.	Na-loaded normal human urine	391	(38)
Cloix et al.	Normal human urine	431	(42)
Weiler et al.	Hypertensive human plasma	408	(10)
Kerek	Plant tissue	408	(29)
Tamura et al.	Pig urine	620	(43)

77% reduction of ouabain-sensitive <sup>86</sup>Rb uptake into human erythrocytes. This did not occur when ANF was given intravenously before sacrifice of the test animals (48). ANF injected into lateral cerebral ventricles releases an Na-K-ATPase inhibitor measured as above in cultured aortic smooth muscle cells (49). Ouabain and digoxin, cardiotonic steroids resembling EDLF and OLF, increase ANF secretion by rat atrial cardiocyte superfusions (50). Liu et al. (51) also employed the perfused beating rabbit atria model to show that ouabain significantly increased ANF secretion in a dose-dependent manner, indicating that the interrelationship between Na-K-ATPase inhibitors and ANF can proceed in both directions.

Finally, in an elegant experiment performed by Morgan et al. (52), using extracts from cultured rat hypothalamic cells separated on Sephadex G-25, a sodium transport inhibitor could be recovered from the post-salt fraction as indicated by three assays: (1) inhibition of transport in human erythrocytes, (2) displacement of <sup>3</sup>H ouabain from its binding site, and (3) direct inhibition of canine Na-K-ATPase. Could pro-ANF and EDLF be co-secreted by the hypothalamus in response to volume expansion or as an indicator of pre-disposition to essential hypertension? A suitable way to settle this question would be to perform immunoassays for pro-ANF and ANF on the 12 kDa protein of the present experiment. For this reason, I would again implore currently active investigators to separate the 12 kDa protein from human hypertensive plasma and test it for pro-ANF and ANF immunoreactivity.

## REFERENCES

1. Gonick HC, Kramer HJ, Paul W, Lu E. Circulating inhibitor of sodium potassium-activated adenosine triphosphatase after expansion of extracellular fluid volume. *Clin Sci Mol Med* (1977) 53:329–34.
2. Kramer HJ, Gonick HC. Effect of extracellular volume expansion on renal Na-K-ATPase and cell metabolism. *Nephron* (1974) 12:281–96. doi:10.1159/000180341
3. Gonick HC. Mechanism of action of natriuretic hormone: inhibitor of Na-K-ATPase. In: Kramer HJ, Kruck F, editors. *Natriuretic Hormone*. Berlin, Heidelberg, New York: Springer (1978). p. 108–21.
4. Nardi R, Sawa H, Carretta R, Bianchi M, Fernandes M. Characteristic variation in the plasma proteins in essential hypertension. *Lancet* (1980) 2:182–3. doi:10.1016/S0140-6736(80)90064-1
5. Morich F, Garthoff B. Characteristic changes of plasma proteins in the Dahl hypertensive rat strain (DS) during the development of hypertension. *J Hypertens* (1985) 3:249–53. doi:10.1097/00004872-198506000-00009
6. Cloix JE, Devynck M-A, Funck Bretano J-L, Meyer P. Plasma protein changes in primary hypertension in humans and rats. *Hypertension* (1983) 5:128–34. doi:10.1161/01.HYP.5.1.128

7. Van de Voorde A, De Broe M, Pollet DE, Rutsaert RJ, Nouwen EJ. Isolation of a plasma protein in patients with essential hypertension. *Biochem Biophys Res Commun* (1982) **111**:1015–21. doi:10.1016/0006-291X(83)91401-8
8. Dey K, Chakraborti T, Roy S, et al. Identification, purification and partial characterization of a 70 kDa inhibitor protein of Na+/K+-ATPase from cytosol of pulmonary artery smooth muscle. *Life Sci* (2010) **86**:473–81. doi:10.1016/j.lfs.2010.02.002
9. Gonick HC, Weiler WJ, Khalil-Manesh F, Weber MA. Predominance of high-molecular weight plasma Na-K-ATPase inhibitor in essential hypertension. *Am J Hypertens* (1993) **6**:680–7.
10. Weiler EW, Khalil-Manesh F, Gonick HC, Prins BA, Purdy RE, Sensharma DK. Na-K-ATPase inhibitor dissociated from hypertension-associated plasma protein. *Am J Hypertens* (1999) **12**:364–73.
11. Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* (1970) **227**:680–5. doi:10.1038/227680a0
12. Purdy RE, Prins BA, Weber MA, Bakhtiarian A, Smith JR, Kim MK, et al. Possible novel action of ouabain: allosteric modulation of vascular serotonergic (5-HT<sub>2</sub>) and angiotensinergic (AT<sub>1</sub>) receptors. *J Pharmacol Exp Ther* (1993) **267**:228–37.
13. Fiske CH, Subbarow Y. The colorimetric determination of phosphorus. *J Biol Chem* (1925) **66**:375–400.
14. Purdy RE, Weber MA. Angiotensin II amplification of alpha adrenergic vasoconstriction: role of receptor reserve. *Circ Res* (1988) **63**:748–57. doi:10.1161/01.RES.63.4.748
15. Gonick HC, Ding Y, Vaziri ND, Bagrov AY, Fedorova OV. Simultaneous measurement of marinobufagenin, ouabain, and hypertension-associated protein in various disease states. *Clin Exp Hypertens* (1998) **20**:617–27. doi:10.3109/10641969809053240
16. Graves SW, Williams GH. Endogenous digitalis-like natriuretic factors. *Ann Rev Med* (1987) **38**:433–44. doi:10.1146/annurev.me.38.020187.002245
17. Lewis LK, Yandle TG, Lewis JG, Richards AM, Pidgeon GB, Kaaja RJ, et al. Ouabain is not detectable in human plasma. *Hypertension* (1994) **24**:549–55. doi:10.1161/01.HYP.24.5.549
18. Baecher S, Kroiss M, Fassnacht M, Vogeser M. No endogenous ouabain is detectable in human plasma by ultra-sensitive UPLC-MS/MS. *Clin Chim Acta* (2014) **431**:87–92. doi:10.1016/j.cca.2014.01.038
19. Blaustein MP. Why isn't endogenous ouabain more widely accepted? *Am J Physiol Heart Circ Physiol* (2014) **307**:H635–9. doi:10.1152/ajpheart.00404.2014
20. Bagrov AY, Fedorova OV, Dmitrieva RI, French AW, Anderson DE. Plasma marinobufagenin-like and ouabain-like immunoreactivity during saline volume expansion in anesthetized dogs. *Cardiovasc Res* (1996) **31**:296–305. doi:10.1016/S0008-6363(95)00208-1
21. Fridman AI, Matveev SA, Agalakova NI, Fedorova OV, Lakatta EG, Bagrov AY. Marinobufagenin, an endogenous ligand of alpha-1 sodium pump is a marker of congestive heart failure severity. *J Hypertens* (2002) **20**:1189–94. doi:10.1097/00004872-200206000-00032
22. Kolmakova EV, Haller ST, Kennedy DJ, Isachkina AN, Budny GV, Frolova EV, et al. Endogenous cardiotonic steroids in chronic renal failure. *Nephrol Dial Transplant* (2011) **26**:2912–9. doi:10.1093/ndt/gfq772
23. Fedorova OV, Talan MI, Agalakova NI, Lakatta EG, Bagrov AY. Endogenous ligand of alpha(1) sodium pump, marinobufagenin, is a novel mediator of sodium chloride-dependent hypertension. *Circulation* (2002) **105**:1122–7. doi:10.1161/hc0902.104710
24. Averina IV, Tapilskaya NI, Reznik VA, Frolova EV, Fedorova OV, Lakatta EG, et al. Endogenous Na/K-ATPase inhibitors in patients with pre-eclampsia. *Cell Mol Biol* (2006) **52**:19–23.
25. Gonick HC, Weiler E, Horn E, et al. Urinary Na-K-ATPase inhibitors and digoxin-like immunoreactive substances in acute congestive heart failure. *Humoral Mechanisms of Heart Failure* (Vol. 3), Tbilisi (1994). p. 23–8.
26. Kramer HJ, Kruck F. Plasma natriuretic activity in oedematous states. *Proc Eur Dial Transplant Assoc* (1976) **12**:321–9.
27. Komiyama Y, Dong XH, Nishimura N, Masaki H, Yoshika M, Masuda M, et al. A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin Biochem* (2005) **38**:36–45. doi:10.1016/j.clinbiochem.2004.08.005
28. Lindner EB, Elmquist A, Porath J. Gel filtration: a method of purification of protein-bound peptides exemplified by oxytocin and vasopressin. *Nature* (1959) **184**:1565–9. doi:10.1038/1841565b0
29. Kerek F. The structure of the digitalislike and natriuretic factors identified as macrocyclic derivatives of the inorganic carbon suboxide. *Hypertens Res* (2000) **23**(Suppl):S33–8. doi:10.1291/hypres.23.Supplement\_S33
30. deBold AJ, Borenstein HB, Veress AT, Sonnenberg H. A rapid and potent natriuretic response to intravenous injection of atrial myocardial extracts in rats. *Life Sci* (1981) **28**:89–94. doi:10.1016/0024-3205(81)90370-2
31. McGrath MF, Kuroski de Bold ML, de Bold AJ. The endocrine function of the heart. *Trends Endocrinol Metab* (2005) **16**(10):469–77. doi:10.1016/j.tem.2005.10.007
32. Buckley MG, Sagnella GA, Markandu ND, Singer DR, MacGregor GA. Concentrations of N-terminal proANP in human plasma: evidence for proANP (1–98) as the circulating form. *Clin Chim Acta* (1990) **191**:1–14. doi:10.1016/0009-8981(90)90052-T
33. Bloch KD, Zisfein JB, Margolies MN, Homcy CJ, Seidman JG, Graham RM. A serum protease cleaves proANF into a 14-kilodalton peptide and ANF. *Am J Physiol* (1987) **252**(1Pt 1):E147–151.
34. Melander O, Frandsen E, Groop L, Hulthén UL. Plasma proANP (1–30) reflects salt sensitivity in subjects with heredity for hypertension. *Hypertension* (2002) **39**:996–9. doi:10.1161/01.HYP.0000017552.91014.2A
35. Górný D, Korzeniowska J, Marcińska A, Pielecki J. Reducing effect of atrial natriuretic factor on Na, K-ATPase activity in rat kidney. *J Physiol Pharmacol* (1994) **45**:173–81.
36. Chiou S, Vesely DL. Kaliuretic peptide: the most potent inhibitor of Na(+)-K+ATPase of the atrial natriuretic peptides. *Endocrinology* (1995) **136**:2033–9. doi:10.1210/endo.136.5.7720651
37. Grammer RT, Fukumi H, Inagami T, Misono KS. Rat atrial natriuretic factor. Purification and vasorelaxant activity. *Biochem Biophys Res Commun* (1983) **116**:696–703. doi:10.1016/0006-291X(83)90581-8
38. Kramer HJ, Krampitz G, Bäcker A, Michel H, Krampitz G, Meyer-Lehnert H. Endogenous sodium pump inhibitors in human urine. Further identification of inhibitors of Na-K-ATPase. *Am J Hypertens* (1995) **8**:753–60. doi:10.1016/0895-7061(95)00125-9
39. deBold AJ, Flynn TJ. Cardionatin I – a novel heart peptide with potent diuretic and natriuretic properties. *Life Sci* (1983) **33**:297–302. doi:10.1016/0024-3205(83)90390-9
40. Bricker NS, Zea L, Shapiro M, San Clemente E, Shankel S. Biologic and physical characteristics of the non-peptidic, non-digitalis-like natriuretic hormone. *Kidney Int* (1993) **44**:937–47. doi:10.1038/ki.1993.335
41. McKinnon W, Lord GA, Forni LG, Hilton PJ. Circulating sodium pump inhibitors in five volume-expanded humans. *J Hypertens* (2003) **21**:2315–21. doi:10.1097/00004872-200312000-00020
42. Cloix JF, Crabos M, Wainer IW, Ruegg U, Seiler M, Meyer P. High yield-purification of a urinary Na+-pump inhibitor. *Biochem Biophys Res Commun* (1985) **131**:1234–40. doi:10.1016/0006-291X(85)90223-2
43. Tamura M, Harris TM, Konishi F, Inagami T. Isolation and characterization of an endogenous Na<sup>+</sup>,K<sup>(+)</sup>-ATPase-specific inhibitor from pig urine. *Eur J Biochem* (1993) **211**:317–27. doi:10.1111/j.1432-1033.1993.tb19901.x
44. Haupert GT Jr. Endogenous Na,K-ATPase inhibitor(s) and atrial natriuretic peptides: are they interrelated? *Prog Biochem Pharmacol* (1988) **23**:71–6.
45. Shibusaki T, Naruse M, Naruse K, Masuda A, Kim YS, Imaki T, et al. Atrial natriuretic factor is released from rat hypothalamus in vitro. *Biochem Biophys Res Commun* (1986) **136**:590–5. doi:10.1016/0006-291X(86)90481-X
46. Songu-Mize E, Bealer SL. Effect of hypothalamic lesions on interaction of centrally administered ANF and the circulating sodium-pump inhibitor. *J Cardiovasc Pharmacol* (1993) **22**(Suppl 2):S4–6. doi:10.1097/00005344-199322002-00003
47. Bealer SL, Haywood JR, Gruber KA, et al. Preoptic-hypothalamic periventricular lesions reduce natriuresis to volume expansion. *Am. J. Physiol.* (1983) **244**:R51–7.
48. Crabos M, Ausiello DA, Haupert GT Jr, Cantiello HF. Atrial natriuretic peptide regulates release of Na<sup>+</sup>-K<sup>+</sup>-ATPase inhibitor from rat brain. *Am J Physiol* (1988) **254**:F912–7.
49. Songu-Mize E, Bealer SL, Hassid AI. Centrally administered ANF promotes appearance of a circulating sodium pump inhibitor. *Am J Physiol* (1990) **258**:H1655–9.
50. Morise T, Takeuchi Y, Okamoto S, Takeda R. Stimulation of atrial natriuretic peptide secretion by Na-K-ATPase inhibitors. *Biochem Biophys Res Commun* (1991) **176**:875–81. doi:10.1016/S0006-291X(05)80267-0

51. Liu LP, Hong L, Yu L, Li HY, Ding DZ, Jin SJ, et al. Ouabain stimulates atrial natriuretic peptide secretion via endothelin-1/ET<sub>B</sub> receptor-mediated pathway in beating rabbit atria. *Life Sci* (2002) **90**:793–8. doi:10.1016/j.lfs.2012.04.008
52. Morgan K, Lewis MD, Spurlock G, Collins PA, Foord SM, Southgate K, et al. Characterization and partial purification of the sodium-potassium-ATPase inhibitor released from cultured rat hypothalamic cells. *J Biol Chem* (1985) **260**:13595–600.

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 15 August 2014; accepted: 30 October 2014; published online: 19 November 2014.

Citation: Gonick HC (2014) Evidence for a 12 kDa “carrier protein” for natriuretic hormone. *Front. Endocrinol.* **5**:196. doi: 10.3389/fendo.2014.00196

This article was submitted to Neuroendocrine Science, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Gonick. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Spherical oligo-silicic acid SOSA disclosed as possible endogenous digitalis-like factor

Franz Kerek<sup>1\*</sup> and Victor A. Voicu<sup>2</sup>

<sup>1</sup> SiNatur GmbH, Martinsried, Germany

<sup>2</sup> Department of Clinical Pharmacology, Toxicology and Psychopharmacology, Carol Davila University of Medicine and Pharmacy, Bucharest, Romania

**Edited by:**

Harvey C. Gonick, University of California Berkeley, USA

**Reviewed by:**

Ricardo Borges, University of La Laguna, Spain

Harvey C. Gonick, University of California Berkeley, USA

**\*Correspondence:**

Franz Kerek, SiNatur GmbH, Am Klopferspitz 19, IZB, 82152 Munich, Germany

e-mail: kerek@sinnatur.net

The  $\text{Na}^+/\text{K}^+$ -ATPase is a membrane ion-transporter protein, specifically inhibited by digitalis glycosides used in cardiac therapy. The existence in mammals of some endogenous digitalis-like factors (EDLFs) as presumed ATPase ligands is generally accepted. But the chemical structure of these factors remained elusive because no weighable amounts of pure EDLFs have been isolated. Recent high-resolution crystal structure data of  $\text{Na}^+/\text{K}^+$ -ATPase have located the hydrophobic binding pocket of the steroid glycoside ouabain. It remained uncertain if the EDLF are targeting this steroid-receptor or another specific binding site(s). Our recently disclosed spherical oligo-silicic acids (SOSA) fulfill the main criteria to be identified with the presumed EDL factors. SOSA was found as a very potent inhibitor of the  $\text{Na}^+/\text{K}^+$ -ATPase,  $\text{Ca}^{2+}$ -ATPase,  $\text{H}^+/\text{K}^+$ -ATPase, and of K-dp-ATPase, with  $\text{IC}_{50}$  values between 0.2 and 0.5  $\mu\text{g}/\text{mL}$ . These findings are even more astonishing while so far, neither monosilicic acid nor its poly-condensed forms have been remarked biologically active. With the diameter  $\phi$  between 1 and 3 nm, SOSA still belong to molecular species definitely smaller than silica nano-particles with  $\phi > 5 \text{ nm}$ . In SOSA molecules, almost all Si-OH bonds are displayed on the external shell, which facilitates the binding to hydrophilic ATPase domains. SOSA is stable for long term in solution but is sensitive to freeze-drying, which could explain the failure of countless attempts to isolate pure EDLF. There is a strong resemblance between SOSA and vanadates, the previously known general inhibitors of P-type ATPases. SOSA may be generated endogenously by spherical oligomerization of the ubiquitously present monosilicic acid in animal fluids. The structure of SOSA is sensitive to the concentration of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ , and other ions suggesting a presumably archaic mechanism for the regulation of the ATPase pumps.

**Keywords:** oligo silicic-acid, ATPase regulation, digitalis-like factor, ouabain

## INTRODUCTION

Investigating the staircase effect on frog ventricular muscle, it was revealed in the early 1950s that human serum contains a factor, which improves the contractile power of the heart, similar to plant-derived digitalis glycosides (1). Besides the serum, this digitalis-like activity was reproduced by a total extract of adrenal but, individual cortisol steroids were, with one exception, inactive. Referring to their finding Szent-Györgyi concluded (2) in more general terms that “such digitalis-like factors are ubiquitously distributed in mammals” and “digitalis glycosides from plants are actually not drugs but only substitutes of the body’s own digitalis-like factors.” This kind of relationship between exogenous drug-substances, which fits accidentally into the receptor of a body’s own factor has been verified by the discovery of endorphins, the endogenous counterparts of the plant-derived morphine (3). Why this broadly accepted rationale was not consequently applied in the differentiation between endogenous digitalis-like factors (EDLFs) and exogenous digitalis glycosides is a subject of the present review.

A further milestone finding of the 1950s was that digitalis glycosides inhibit the transport of  $\text{Na}^+$  and  $\text{K}^+$  ions across the erythrocyte membrane (4) and above all the seminal discovery of

the membrane transport protein of the  $\text{Na}^+$  and  $\text{K}^+$ -ions by Skou (5). This  $\text{Na}^+/\text{K}^+$ -ATPase (adenosine-triphosphatase) abbreviated NKA or sodium pump is present in the membrane of all eukaryotic cells. Every pumping cycle of the NKA moves 3  $\text{Na}^+$  ions outward and 2  $\text{K}^+$  ions inward, powered by the energy of a phosphate bond from ATP.

At the beginning of the 1960s, the existence of a circulating natriuretic factor was postulated by De Wardener et al. (6) showing that the blood, transfused from a saline-loaded, hypertensive dog, produces natriuresis in the recipient normotensive animal. It was further observed that the plasma ultra-filtrate of saline-loaded dogs inhibits the sodium transport in toad bladder and the volume expansion was accompanied by increasing concentrations of a sodium pump-inhibitory factor (7). Ascertaining that both effects were caused by the same factor the name natriuretic hormone was proposed. The natriuretic fraction extracted from the plasma of volume-expanded dogs inhibited the ouabain-insensitive NKA from rat kidney (8).

After the identity of the natriuretic hormone with the sodium pump-inhibitory factor was confirmed, the name EDLF came into use (9). The initial idea that EDLF could be a natriuretic peptide

was rejected after disclosure of the atrial natriuretic peptide (ANP), which has, contrary to EDLF no NKA-inhibitory activity (10). Though the structure of EDLF remained obscure, some of its particular characteristics were established as for instance: its non-peptide nature or its specific interaction with different ATPase isoforms. But the failure of all attempts to obtain weighable amounts of pure EDLF impeded for more than five decades the disclosure of its chemical structure.

$\text{Na}^+/\text{K}^+$ -ATPase controls a broad spectrum of essential cellular functions such as ion homeostasis, membrane potential, pH, temperature, and water osmosis, thereby regulating important physiological processes, e.g., muscle contraction, nervous signal transmission, renal sodium retention, and vascular tone. Study data in animal models and clinical observations in human beings suggest that cardiac insufficiency, essential hypertension, and other diseases may be caused by or connected to malfunction or dysregulation of the sodium pump.

The extensive research work related to the structure, characterization, mechanism of action, and physiological implications of the  $\text{Na}^+/\text{K}^+$ -ATPase was comprehensively reviewed by Gadsby et al. (11), Glynn (12), Kaplan (13), and Jørgensen et al. (14). Similarly, extensive reviews on the whole P-type-ATPase field have been published by Møller et al. (15), Kühlbrandt (16) and by the original contributions of Axelsen and Palmgren (17, 18) with special focus on evolutionary aspects of the P-type ATPases.

### ATPase RECEPTOR SITE

P-type ATPase is the generic designation of several ATP-driven transmembrane ion pumps found in bacteria, archaea, and eukaryotes. The prefix P refers to the ability of these proteins for phosphorylation and de-phosphorylation of their catalytic aspartate residue (15). By the binding and removal of the phosphate group, ATPases interconvert between two conformations, denoted by E1 and E2, each with different affinity to the nucleotide ATP (adenosine triphosphate) and the transported ions (16). A common feature of P-type ATPases is their inhibition by vanadate ions at micro- and sub-micro-molar concentrations.

From about 200 members of the P-type ATPase family, the most prominent pumps are the cell membrane  $\text{Na}^+/\text{K}^+$ -ATPase (NKA); the  $\text{Ca}^{2+}$ -ATPase (SERCA) from sarcoplasmic reticulum (SR), the gastric  $\text{H}^+/\text{K}^+$ -ATPase, and the bacterial K-dp-ATPase. Based on 80–90% similarities of the amino acid (AA) sequences in the conserved regions, it is assumed that P-type ATPases evolved from a common ancestor, probably 3500 million years ago (17, 18).

A seminal breakthrough for the detailed structural understanding of the transmembrane ion pumping was achieved by the first high-resolution (2.6 Å) crystal structure of the SERCA  $\text{Ca}^{2+}$ -ATPase protein from SR solved by Toyoshima et al. (19). This study established the detailed 3D structure and accomplished the functional characterization of the cytoplasmic subunits designated P (phosphorylation), N (nucleotide binding), and A (actuator) domains. It was found that the ion-binding sites are surrounded by the M4–M6 and M8 transmembrane helices where M4 and M6 provide the efficient geometry for the coordination of the  $\text{Ca}^{2+}$  ions. The over 50 Å distance between the membrane site of the  $\text{Ca}^{2+}$  ion translocation and the cytoplasmic phosphorylation site is remarkably long.

In the next high-resolution (3.1 Å) crystal structure of the SERCA pump, the Toyoshima group applied the sesquiterpene lactone thapsigargin, to stabilize the Ca-free E2-(TG) state (20). The comparison of both crystallized forms  $\text{Ca}^{2+}$ E1 and E2-(TG) revealed further details of the ion transport mechanism. In a following contribution (21), the structure of the  $\text{Ca}^{2+}$ -ATPase was solved in the E1 state fixed by the ATP-analog AMPPCP. In the same year, the structure at 2.3 Å resolution of SERCA with phosphate analogs such as  $[\text{MgF}_4]^{2-}$  has been resolved (22). The studies of the Ca-pump fixed with ATP- or phosphate analogs have completed the structural insight into almost all important states of the pump turnover.

Resolving the crystal structure of the SERCA pump with the phosphate mimic  $[\text{BeF}_3]^-$ , Olesen et al. (23) provided support for the presence of an open ion pathway in the pump in which the transmembrane domains form a funnel-shaped geometry. As described later, the crystal structure data confirmed that P-type ATPases share the same architecture regardless of the size, charge, and number of ions that they transport. It seems that the differences are largely confined to the ion-binding pocket (23, 24).

$\text{Na}^+/\text{K}^+$ -ATPase is sensitive to inhibition by digitalis glycosides (e.g., digoxin, ouabain), isolated from medicinal plants used over centuries to treat congestive heart failure. The term “digitalis” designates the entire group of cardiac glycosides and aglycones without regard of their structure and origin (25). Responsible for the cardiac activity is the aglycone, i.e., the steroid nucleus, which results after the sugar moiety from position 3 is removed. Steroid aglycones with five-membered lactone ring in position 17 are named cardenolides while those with six-membered lactone ring bufadienolides. Both terms, cardiotonic (CTS) or cardiac steroids (CSs), are in use but the adjective “tonic” is less rigorous as it refers to a species-dependent physiological response; thus, the designation CS should be preferred over CTS.

Since  $\text{Na}^+/\text{K}^+$ -ATPase is the target of digitalis drugs in heart failure patients, it was of prior importance to establish structure-activity relationships. A general correlation between binding affinity and NKA-inhibitory potency of cardiac glycosides was revealed (26) but some notable exceptions were also identified. By replacement of the five-membered lactone ring in cardenolides with the 2-pyrone ring of bufadienolides, the binding affinity declines but the inhibitory potency increased. Furthermore, while the removal of ouabain’s rhamnose moiety had little effect on inhibitory potency, it caused a decline in ligand binding affinity.

In a recent study, 30 different cardiac glycosides were investigated for their interaction with the  $\alpha 1$ ,  $\alpha 2$ , and  $\alpha 3$  isoforms of the human NKA expressed in *Pichia pastoris* (27). The study revealed significant isoform selectivity by digoxin glycosides but ouabain was found moderately  $\alpha 2$  selective. The observed influence of the sugar moiety on the selectivity was surprising since according to pharmacological data this part influences rather the bioavailability and metabolism of the digitalis drugs.

Biophysical methods provided further insight into the charge transfer processes during the pump cycle. The binding of ouabain is associated with movement of electrical charges; thus, it can be followed by the charge-sensitive fluorescence indicator RH421. These data revealed that the binding of ouabain or generally of

a cardiac glycoside to the  $\text{Na}^+/\text{K}^+$ -ATPase protein will stabilize under physiological conditions the E2P- $2\text{Na}^+$  stage (28).

$\text{Na}^+/\text{K}^+$ -ATPase is expressed in all animal cells and shows highly conserved AA sequences in the main  $\alpha$  subunit (~1000 AA residues) responsible for the catalytic function, similar to that found in SERCA. This catalytic subunit of NKA has 10 transmembrane TM helices numbered M1–M10 from the amino-terminal. Experiments with punctual mutations of AA residues evidenced CS binding sequences in the extracellular loops L1/2, L5/6, and L7/L8. The cytosolic loop L2/3 contains the nucleotide (ATP) binding N site and the phosphorylation site P, while the  $\text{NH}_2$  terminal and the L2/3 loop are responsible for the de-phosphorylation step. NKA contains further the heavily glycosylated  $\beta$ -subunit with ca. ~300 AA and the tissue specific auxiliary  $\gamma$  subunits FXYD of ca. 70–180 residues. The multiple regulatory potential of the NKA is explained by the existence of tissue-specific assemblies of different structural subunits or isoforms (29).

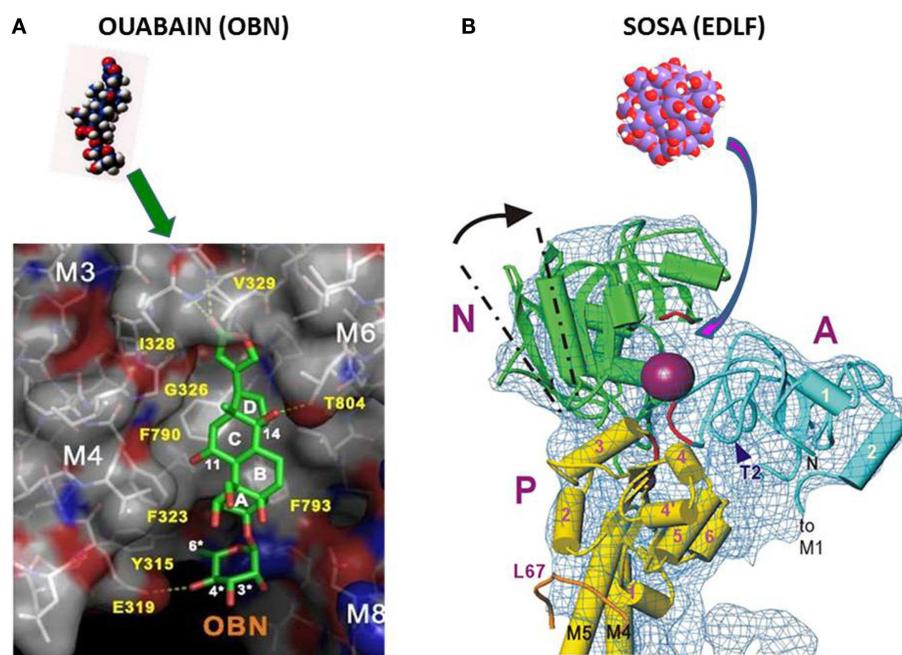
In comparison with more than 20 X-ray structures for  $\text{Ca}^{2+}$ -ATPase, for NKA only 5 crystal structures at better than 5 Å resolution have been published. Some difficulties come from the source of NKA protein, which is limited to rabbit kidney and shark rectal gland, expressing selectively the  $\alpha 1$  isoform. The lack of very high-affinity inhibitors able to fix the enzyme in one particular conformation posed further limitations to NKA crystallization (29).

The crystal structure of the pig-renal  $\text{Na}^+/\text{K}^+$ -ATPase with 2 Rubidium ions as K ion congeners was solved at 3.6 Å resolution by the PUMPKIN group in Denmark (30). The structure shows that the conformation of the  $\alpha$  unit (in Rb/K occlusion) closely matches the conformation of the  $\text{Ca}^{2+}$  ion bound state of SERCA.

The crystal structure of the  $\text{Na}^+/\text{K}^+$ -ATPase from shark enzyme was resolved (31) at 2.4 Å resolution in the: E2- $2\text{K}^+\sim\text{Pi}$  state in which the pump has a high affinity to  $\text{K}^+$  ions. The coordination of  $\text{K}^+$  ions in the transmembrane sites and the critical role of the  $\beta$  subunit in binding of the  $\text{K}^+$  ions were remarked. Despite identical coordinating residues, small differences with the  $\text{Ca}^{2+}$ -ATPase pump were noted.

Solving with 2.8 Å resolution, the crystal structure of  $\text{Na}^+/\text{K}^+$ -ATPase, co-crystallized with ouabain in the E2- $2\text{K}^+\sim\text{Pi}$  state, the authors concluded (32) that ouabain is deeply inserted into the transmembrane domain, with lactone ring near to the  $\text{K}^+$  binding site with partial unwinding of the M4E helix (see **Figure 1A** below). This unwinding should explain why ouabain binding is so slow. The data suggest reconsideration of previous data that CSs bind to the extracellular surface of the ATPase  $\alpha$ -subunit. Since ouabain interacts with transmembrane segments M3, M4, and M6 involved in ion transport this steroid can influence or block these processes.

Based on the crystal structure at 4.6 Å resolution of the pig kidney  $\text{Na}^+/\text{K}^+$ -ATPase with ouabain bound in the E2P state, it was suggested that the high-affinity binding of ouabain stabilizes the phosphorylated state (33). The steroid binds to a site formed between  $\alpha\text{M}1$  and  $\alpha\text{M}6$  domains, plugging the ion pathway from the extracellular side. A high-affinity interaction is formed between the steroid and the  $\alpha\text{M}1\text{--}2$  part from domain  $\alpha$ , which is rotated following the phosphorylation. Solving the crystal structure at 3.4 Å resolution of the phosphorylated pig kidney NKA in complex with ouabain has revealed that the steroid binds with intensive hydrogen bonding network to the  $\alpha\text{M}1$ ,  $\alpha\text{M}2$ , and  $\alpha\text{M}6$  transmembrane segments (34). It was concluded that the binding pocket in the  $[\text{Mg}^{2+}]$  E2P state allows deep ouabain binding with possible long-range interactions with  $\text{Mg}^{2+}$  and  $\text{K}^+$  ions.



**FIGURE 1 | (A)** Binding of ouabain to the hydrophobic site in NKA (32). **(B)** Binding of SOSA to the hydrophilic receptor site of decavanadate in SERCA (19).

Applying fluorescein-labeled ouabain and a lanthanide binding tag in the NKA derived from *Xenopus oocytes* the data of spectroscopic measurements (35) suggested two different type binding sites for ouabain: one external, low-affinity site on the extracellular end of the ion pathway as previously assumed; the second high-affinity site is slightly deeper toward the intracellular end of the ion pathway, as indicated by recent X-ray diffraction studies.

In conclusion, the high-resolution crystal structure data have identified the location of ouabain in the NKA together with the protein domains involved in the interaction with this CS. The ouabain-binding site described in Ref. (32) has a flat hydrophobic surface suitable for the interaction with the steroid frame as shown by **Figure 1A**.

For some hydrophilic ATPase ligands such as the decavanadate or the here disclosed spherical oligo-silicic acid (SOSA) (see below), it is improbable to target the hydrophobic receptor site of CSs and thus to compete directly with ouabain or digoxin. A more suitable target for hydrophilic inhibitors in P-type ATPases is the phosphorylation site in the cytoplasmic region between the N, A, and P domains (**Figure 1B**), as it was suggested for the decavanadate ion in SERCA (19).

The high-structural analogy between the hydrophilic poly-anionic decavanadate and the here disclosed SOSA suggests that SOSA should target ATPase pumps at the phosphorylation site in a similar manner as decavanadate. This assumption is illustrated by **Figure 1B**) with the high-resolution structure of the cytoplasmic domain of SERCA with the decavanadate ion bound to the hydrophilic phosphorylation site (19).

### DILEMMA: EDLF OR CARDIAC STEROIDS

The physiological role of the CS receptors and the existence of endogenous non-steroidal ligands of NKA have been investigated by the group of Lingrel (36, 37). The experiments were conducted among others on some ouabain-resistant and ouabain-sensitive isoforms (cc2, cc3, and cc4) genetically engineered in mice and rat. The biological function and significance of the CS binding site was evidenced with rigorous distinction between cardiotonic steroids (CTS) and the yet undisclosed endogenous ATPase ligands. Reviewing the whole set of his experimental results and their significance for the physiological role of the NKA receptors, Lingrel concluded three main points: (1) the ouabain-binding site of the Na/K-ATPase plays a physiological role, (2) an endogenous ligand for the Na<sup>+</sup>/K<sup>+</sup>-ATPase must exist, and finally, (3) whether the endogenous ligand acts through a change in intracellular Na<sup>+</sup> or through a signaling mechanism is unknown (37).

Despite the convincing arguments of Lingrel, the relation between EDL factors and CSs remained a dilemma, i.e., “a problem offering two possibilities, neither of which is acceptable.” In fact, there are some experimentally identified but physically not isolated (probably labile) EDL factors assumed as ATPase ligands that differ markedly from CSs except for the inhibition of the NKA. Lacking weighable amounts of EDL factors, the studies have been performed only with the commercially available CSs and, therefore, could neither demonstrate nor exclude the identity of EDLF with CSs.

This unsolved dilemma has marked the extensive research work focused to disclose the structure and properties of the putative

endogenous ligands of the ATPases as thoroughly reviewed by Goto et al. (38), Hollenberg and Graves (39), Buckalew (40), Schoner and Scheiner-Bobis (41), Nesher et al. (42), and Bagrov et al. (43).

Numerous attempts to isolate pure EDL factors from various biological sources (organs, glands, plasma, or urine) were listed by these surveys (38–42). Despite applying extraction procedures on several kilogram amounts of starting material and efficient separation techniques, the final yields after multiple purification steps were invariably small: trace, sub-microgram amounts of EDLF, definitely insufficient for structural studies.

The low chemical stability of the EDL factors should also have been considered as possible explanation for the dramatically vanishing EDLF amounts along the purification processes. But this instability was not investigated in detail. Once a labile sodium pump inhibitor was signalized in peritoneal dialyzate (44) with even significantly higher inhibitory potency than ouabain, but the experiment was not reproduced. Of historical interest is the mention published 60 years ago by Szent-Györgyi (2) that, “the cardio-active serum factor if lyophilized and stored, loses its activity, as it also loses it on repeated freezing and thawing.”

Because of these persistent failures to isolate EDL factors, the search for endogenous ATPase ligands and assessment of their putative biological role became increasingly discrepant. Lacking measurable amounts of the pure EDLF on one side and the growing body of evidences that toxic CSs of herbal or amphibian origin are unable to function as endogenous ATPase ligands in mammals became an unsolvable problem. Further controversies were caused by the sugar moieties of the cardiac glycosides comprising desoxy-sugars (e.g., rhamnose, digitoxose) which were never identified in mammals, making it unlikely that such desoxy-glycosides can genuinely exist and act as endogenous ligands in these animals.

Estimated from their evolutionary history CSs identified in flowering plants could not be older than 100 million years and steroids from amphibians must be “younger” than 400 million years. It is very unlikely that such compounds have existed as regulatory ligands of the ATPase pumps 3500 million years ago in the prokaryote membrane.

After the successful evolution of the archaic ATPase 3.5 billion years ago, it was no more evolutionary pressure to improve this perfectly working ion-pumping mechanism or to change its endogenous regulatory ligand. The adaptation on the growing complexity of multicellular organisms was accomplished by diversification of the subunits or by the combination of substructures, without essential changes of the basic pumping mechanism.

Finally, it should be remarked that the search for identification of the EDL factors had considered almost exclusively organic candidates (38). This was not fully justified as the essential chemical reaction of the pumping cycle is the binding (and release) of a simple inorganic phosphate group. The lack of specific 3D structures in solution makes simple inorganic salts rather unable to fit a receptor site and to work as endogenous ligands contrary to organic substances. However, some inorganic poly-oxo-acids derived from metalloids such as Be, Al, Si, Ge, As, V, Cr, or Mn are able to form 3D structures in solution to fit into a receptor site. Actually the predominant part of these poly-oxo-acids and their

salts are toxic for living organisms, which reduces the number of candidate endogenous ligands.

Several criteria of the putative EDLFs have been formulated previously, among others by Goto et al. (38). Completing the earlier list with a few criteria, we consider that the endogenous ATPase ligands should have characteristics as:

- a. inhibitor of the  $\text{Na}^+/\text{K}^+$ -ATPase,
- b. inhibitor of other P-type ATPases,
- c. ubiquitously distributed,
- d. bioavailable and eliminable,
- e. non-toxic for animals,
- f. was present in a very early stage of the evolution,
- g. sensitive to drying and freeze-drying,
- h. specific 3D structure in solution.

These properties can also explain the difficulties met earlier in the isolation and characterization of pure EDLFs. Further efforts are needed to disclose the detailed mechanism of action at cellular and physiological level of the actual EDL factors.

### ATPase INHIBITORY VANADATES

The inhibition of the  $\text{Na}^+/\text{K}^+$ -ATPase by vanadate was discovered in year 1977 with the accidental observation (45, 46) that the reagent grade ATP of Sigma was contaminated with an ATPase inhibitory substance, identified as sodium vanadate ( $\text{Na}_3\text{VO}_4$ ). The similar (isoelectronic) structure of the vanadate and phosphate ions was considered as a probable mechanism of the inhibition by vanadate in competition to the phosphate binding site.

Investigating the interaction of vanadates (47) with fluorescein-labeled SERCA, it was observed that vanadate impeded the high-affinity  $\text{Ca}^{2+}$  binding to the enzyme at 4°C. Vanadate inhibits the phosphorylation reaction by inorganic phosphate but had no effect on the phosphorylation by ATP. It was suggested that vanadate binds to the low-affinity ATP binding site of the ATPases, which is exposed only in the E2 conformation of the enzyme.

Interactions between SERCA and vanadate ions in solution have been investigated by  $^{51}\text{V}$ -NMR spectra indicating that mono- and oligo-vanadates are bound to SR membrane influencing the structure of  $\text{Ca}^{2+}$ -ATPase (48). Actually, the mono and oligovanadate species form some complex equilibria impeding the establishment of rigorous structure–activity correlations.

In the presence of  $\text{Ca}^{2+}$ , it was observed that tetra- and decavanadate  $[\text{V}_{10}\text{O}_{28}]^{6-}$  binds to the SERCA pump, whereas monomeric vanadate binds to the SR only when ATP is present. There are further arguments that decavanadate clearly differs from mono- or small-vanadate oligomers in preventing the accumulation of  $\text{Ca}^{2+}$  ions by SR vesicles, which is coupled to ATP hydrolysis (49).

Biological studies with vanadium often disregarded the formation of decameric vanadate species known to manifest high-affinity interaction with many proteins such as myosin and the SR calcium pump (50). Vanadium is accumulated in mitochondria in particular when decavanadate is administered. These findings point out the contributions of decavanadate to *in vivo* effects induced by vanadium in biological systems.

An increasing volume of data suggests the putative biological importance of decavanadate, a vanadate oligomer that eventually

occurs in the cytoplasm more often than expected (51). Specific interactions of decavanadate have been clearly demonstrated for  $\text{Ca}^{2+}$ -ATPase, myosin, and actin, considered as major proteins in muscle contraction and its regulation. Based on crystal structure data, the binding of the SERCA inhibitory decavanadate was localized (19, 52) to the ATP binding site between the cytoplasmic domains A, N, and P of the thapsigargin-inhibited enzyme in the absence of  $\text{Ca}^{2+}$  as shown by **Figure 1B** of the present paper.

Vanadate compounds show a significant antidiabetic efficacy. Sodium vanadate was applied in diabetes therapy 22 years before the first use of insulin to treat diabetes in human beings (53, 54). Besides its insulin-mimetic action, vanadate inhibits the glucose-6-phosphatase (G6P) enzyme, with a key role in glucose metabolism.

The proposal of Kramer et al. (55) to consider vanadium disorbinate with a molar mass of 403 Da as a candidate EDLF is worth mentioning. However, this hypothesis has not been confirmed since vanadate ions are not ubiquitously distributed in mammals and are toxic in particular by accumulation in some organs.

### ATPase INHIBITORY MCS-FACTORS

Our way to disclose the structure of the assumed EDLFs is a typical example of serendipity. At the end of 1990s, we investigated at the Max-Planck Institute for Biochemistry in Munich an herbal product isolated from the roots of *Helleborus species*. The plant product contained, besides other components, the cardiac glycoside hellebrin with strong NKA-inhibitory potency. The chemical stability of hellebrin was monitored by measuring the inhibition of  $\text{Na}^+/\text{K}^+$ -ATPase, starting therewith a very intensive and prolific collaboration with Hans-Jürgen Apell and Robert Stimac from the University of Konstanz (GER), which led finally to the disclosure of the novel ATPase-inhibitory factor.

The alkaline treatment destroyed hellebrin and annihilated the NKA-inhibitory effect but, as the alkaline boiling was accidentally prolonged for several hours, a very potent novel NKA inhibitor was generated (56). By the HPLC on RP-18 column, the pure inhibitory compound eluted closely after the injection peak, or delayed if it was attached to some lipophilic components. Similar characteristics have been reported by the HPLC analysis of the earlier EDLF preparations from biological samples (38–43).

The main component of the plant material subjected to alkaline boiling was a resin-like compound, similar to the polymeric carbon suboxide (57). Therefore, it was thought that the obtained potent ATPase inhibitor could be a low molecular weight decomposition product of this polymer. For the structure of the de-polymerization product, we assumed a repeatedly condensed 4-pyrone frame that forms a supplementary cage-type macrocycle with formula  $(\text{C}_3\text{O}_2)_n$  where  $(n = 4, 6, \text{ or } 8)$ . We named the inhibitor MCS, macrocyclic carbon suboxide (56). Tentatively, this MCS was suggested as probable EDLF and natriuretic factor (58).

MCS factors showed a rigorously reproducible potent inhibition of the  $\text{Na}^+/\text{K}^+$ -ATPase,  $\text{Ca}^{2+}$ -ATPase,  $\text{H}^+/\text{K}^+$ -ATPase, and K-dp-ATPase with  $\text{IC}_{50}$  values in the 0.2–0.5  $\mu\text{g}/\text{mL}$  range. The mechanism of  $\text{Na}^+/\text{K}^+$ -ATPase inhibition by the MCS factor was investigated with the fluorescent styryl dye RH421, a dye known to

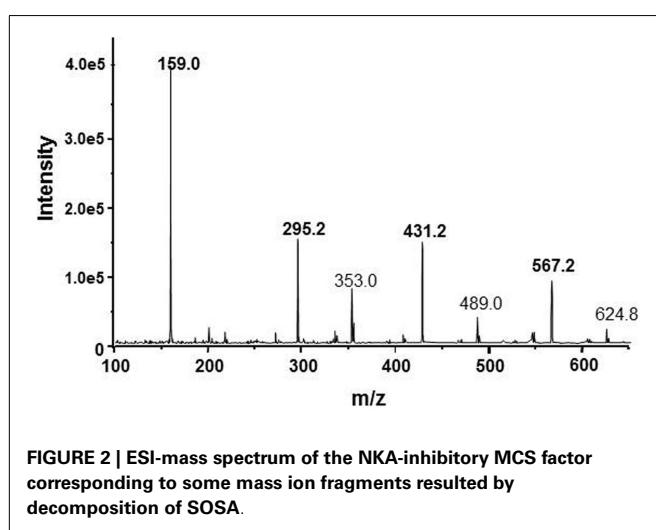
reflect changes of local electric fields in the membrane dielectric. It was found that the binding of the MCS to the  $\text{Na}^+/\text{K}^+$ -ATPase is not competitive with ouabain (59, 60). MCS factors interact with the  $\text{Na}^+/\text{K}^+$ -ATPase in the E1 conformation of the ion pump and induce a structural rearrangement that causes a change of the equilibrium dissociation constant for one of the first 2 intracellular cation binding sites. The MCS-inhibited state was found to have bound one cation ( $\text{H}^+$ ,  $\text{Na}^+$ , or  $\text{K}^+$ ) in one of two non  $\text{Na}^+$  specific binding sites, and the other  $\text{Na}^+$  ion was bound at high  $\text{Na}^+$  concentrations to the highly  $\text{Na}^+$ -selective ion-binding site (60).

The proposal with cage-form condensed macrocyclic carbon suboxide structure was apparently supported by the mass ion peaks ( $m/z$ ) containing multiples of the 68.03 Da unit, the molar mass of the  $\text{C}_3\text{O}_2$ .

The main  $m/z$  peaks in ESI-MS spectra (Figure 2) have been assessed as small multiples of the carbon suboxide unit (68 Da) with 1  $\text{Na}^+$  ion according to the formula  $[(\text{C}_3\text{O}_2)_n \cdot \text{Na}]^+$  thus,  $m/z = 159$  Da correspond to  $n = 2$ ; 295 Da ( $n = 4$ ); 431 Da ( $n = 6$ ), and 567 Da ( $n = 8$ ). The small  $\text{MH}^+$  peaks at 275, 409.2, and 544.2 Da were also perceptible in the mass spectrum (56).

Interestingly, a molar mass ion at  $M = 408$  Da ( $\text{MH}^+ = 409$ ;  $\text{MNa}^+ = 431$  Da) was identified in some earlier EDLF preparations from biological sources, e.g., human plasma (61), placenta (62), or bound to a hypertension-associated plasma protein (63). It can be speculated about the possible identity of these factors and our MCS product but only the same molar mass ion value is not sufficient to prove or disprove this identity. The fine structure of the mass spectra may also differ due to the different ionization techniques, i.e., FAB used by Weiler et al. (63) and ESI-MS applied by us (56).

Although the ATPase-inhibitory effect of the MCS factors on several P-type ATPases and the mechanism of action on NKA were rigorously reproducible, the structure with head-to-tail condensed pyran-4-one rings supplementary bond in a cage-like macro-cycle could not be confirmed by synthesis despite huge experimental efforts with Frank Freudenmann and Luis Moroder at the MPI for biochemistry in Munich.



**FIGURE 2 |** ESI-mass spectrum of the NKA-inhibitory MCS factor corresponding to some mass ion fragments resulted by decomposition of SOSA.

Likewise not confirmed were the specific  $^{13}\text{C}$ -NMR signals and the UV-absorbance peaks expected for the macrocyclic condensed pyran-4-one structure. The assessment of the mass spectrum as multiples of a 68 Da unit was correct but the attribution of this mass to  $\text{C}_3\text{O}_2$  was erroneous. These disagreements required the revision of the proposed macrocyclic cage-structure.

## SPHERICAL OLIGO-SILICIC ACID (SOSA)

The decisive hint to disclose the actual chemical structure of our NKA-inhibitory factor came from revision of the blind probe of the described alkaline preparation. Surprisingly, the several hours boiling of the NaOH solution alone, without any other reagent yielded a similarly potent ATPase inhibitor as that obtained by alkaline boiling of the plant polymer.

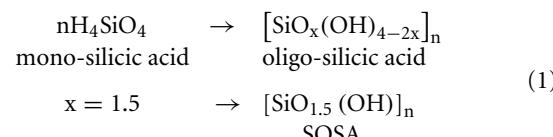
The mystery was explained by identifying small amounts of sodium silicate in the solution leached from the glass flask by its prolonged alkaline heating in oil bath at 120°C. Applying the proper neutralization-activation procedure (56), this silicate was transformed to the highly active ATPase-inhibitory factor identified as spherical oligomers of silicic acid. The generation of biologically active oligomeric condensation products from the inactive monosilicic acid was totally unexpected and thus very surprising.

We considered that this finding could have implications in clearing of some controversial disputes within the following research areas:

Regarding the biological role of silicon, it was generally agreed that Si provides structural support in plants and is beneficial of bones and elasticity of cartilages in animals. But neither a Si containing biologically active substance, nor a protein, which needs Si has been found in animals. Identifying the SOSA as biologically active water-soluble Si compound (64), a decisive argument has been provided in support of the assumed biological role (essentiality) of this element.

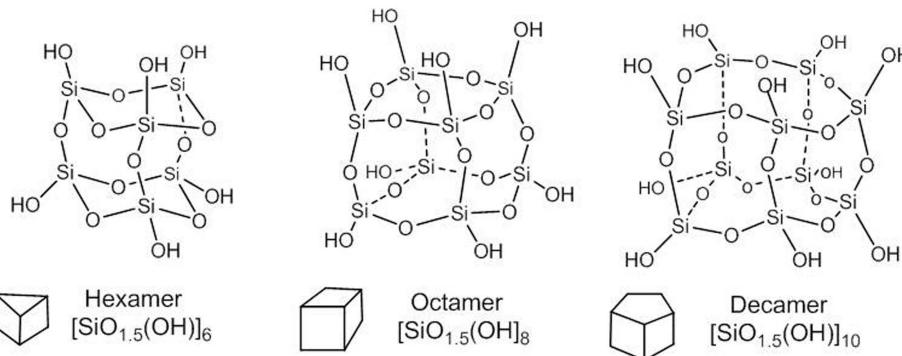
The assumed existence of EDLFs and their putative identity with cardiac glycosides was the subject of debates for several decades. The proposed identity of EDLF (65) with the SOSA was a novel approach suggesting the reconsideration of the divergent opinions.

The structure of the ATPase-inhibitory SOSA is formed by successive condensation of a few (oligos in Greek) molecules of monosilicic acid  $\text{H}_4\text{SiO}_4$  according to the equation (Eq. 1) where “ $n$ ” should be in the range of 16–200.



The value  $x = 1.5$  in the general formula  $[\text{SiO}_x(\text{OH})_{4-2x}]_n$  is congruent with a particular symmetry of the multi-cyclic silicic acid oligomers corresponding to polyhedral symmetry, i.e., prismatic hexamer ( $n = 6$ ), cubic octamer ( $n = 8$ ), and prismatic decamer ( $n = 10$ ) structure, known as silsesquioxanes with general formula  $[\text{SiO}_{1.5}\text{OH}]_n$  and shown by Figure 3.

The spherical form of the oligo-silicic acid SOSA is accomplished for  $x = 1.5$  in the general formula  $[\text{SiO}_x(\text{OH})_{4-2x}]_n$  and



**FIGURE 3 | Structure of polyhedral silsesquioxanes.**

is assumed as the natural continuation of the polyhedral series with formula  $[\text{SiO}_{1.5}(\text{OH})]_n$  for values of  $n > 20$ . There is an interesting formal resemblance with the series of Platonic bodies (tetrahedron, cube, . . . sphere) from geometry.

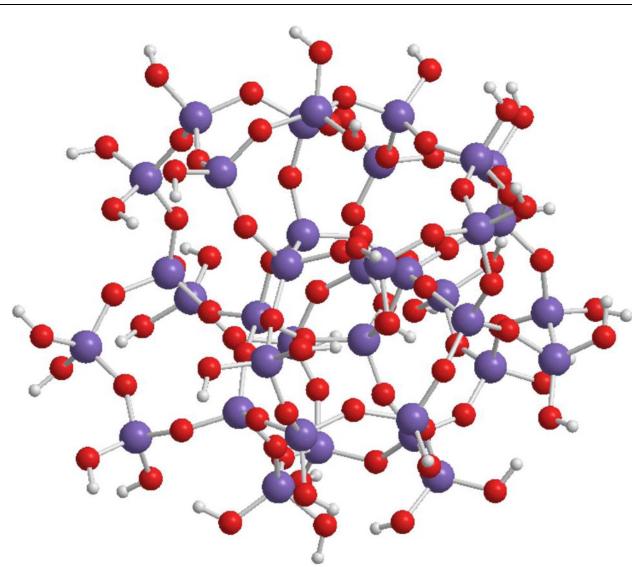
Actually, the cage-type condensed polyhedral silsesquioxanes have all Si-OH groups at the external vertices with nearly axial orientation. A similarly external distribution of the Si-OH groups is accomplished by the here disclosed nearly spherical oligomers of the silicic acid. SOSA molecules as next term in the series of polyhedral structures with the same general formula  $[\text{SiO}_{1.5}(\text{OH})]_n$  have the same ratio between the Si: O: H atom = 1: 2.5: 1.

But, there are significant differences between the chemical structure and properties of the polyhedral and of the spherical silica. The predominant difference is that polyhedral silsesquioxanes inhibit neither  $\text{Na}^+/\text{K}^+$ -ATPase nor other ATPases conversely to the strong ATPase-inhibitory SOSA. Structurally, all polyhedral silsesquioxanes (Figure 3) comprise only Q3 type Si atoms, i.e., each one is involved in three (Si)-O-Si bonds and one (Si)-OH bond.

The spherical shape of the oligo-condensed silicic acid with  $x = 1.5$  is accomplished by some preferred “ $n$ ” values. Figure 4 shows the SOSA molecule with  $n = 36$ , as ball and stick model. This SOSA molecule with formula  $[\text{Si}_{36}\text{O}_{54}(\text{OH})_{36}]$  comprises in its internal shells  $4 + 8 = 12$  Si atoms of type Q<sup>4</sup> (without Si-OH bonds). In the external shell, there are 12 Si atoms of type Q3 (with one Si-OH bond) and 12 Si atoms of type Q<sup>2</sup> (with two Si-OH bonds). It is observed that the 36 external Si-OH bonds are displayed on the external surface strongly facilitating the hydrophilic interactions with proteins.

Applying the gel-permeation chromatography (GPC) method, molar mass values in the range of 1.2–6.0 kDa have been obtained of the spherical condensed silicic acid oligomers. For the SOSA molecule illustrated on Figure 4 there resulted  $M = 3.2$  kDa, corresponding to molecular diameter  $\phi = 2.2$  nm further confirmed by dynamic light scattering (DLS).

The mass ion peaks of SOSA obtained by ESI-MS technique were practically the same as those identified in the spectrum of our MCS factor (Figure 2). Actually these  $m/z$  peaks correspond to small ionic fragments resulted by the split of the large SOSA molecules in the ionization chamber. The main mass ion peaks



**FIGURE 4 | Structure of SOSA with formula:  $[\text{Si}_{36}\text{O}_{54}(\text{OH})_{36}]$  with 36 Si-OH bonds on the external surface of the spherical molecule.**

correspond to the formula  $[(\text{Si}_2\text{O}_5)_q \cdot \text{Na}]^+$  with  $m/z = 159$  Da for  $q = 1$ ; 295 Da for  $q = 2$ ; 431 Da for  $q = 3$ , and 567 Da for  $q = 4$ . Similar mass spectra have been observed by decomposition of condensed silsesquioxanes (66, 67).

In fact, the erroneous assessment of the mass ion peaks as derived from carbon suboxide was caused by the accidental identity of the molar mass of  $\text{C}_3\text{O}_2$   $M = 68.03$  Da and the mass of the  $1/2 \text{Si}_2\text{O}_5$  (di-silicate) ion with  $M = 68.08$  Da, resulted by the decomposition of SOSA. In conclusion, the mass ion peaks on Figure 2 correspond rather to the formula  $[(\text{Si}_2\text{O}_5)_q \cdot \text{Na}]^+$  of SOSA and not to the formula  $[(\text{C}_3\text{O}_2)_n \cdot \text{Na}]^+$  as initially suggested.

### SOSA AS POSSIBLE ENDOGENOUS FACTOR

With the confirmed potent inhibition of several P-type ATPases, SOSA fulfills the main condition as a candidate ATPase ligand. Its classification as an endogenous factor requires further

that SOSA should be produced within the organism, tissue, or cell. This condition is satisfied with the presumed biosynthetic pathway of SOSA by spherical oligomerization of the monosilicic acid ( $H_4SiO_4$ ) ubiquitously distributed in plants and animals (68, 69).

A human body contains approximately 1 g of Si in various combinations with oxygen (generic name: silica). Almost all silica in the body is bound to biomolecules and tissues and only a minor part circulates as dissolved silicic acid in blood plasma and urine (70). The plasma level of silicic acid is 0.7 mg/L corresponding to 0.2 mg/L silicon. Infants have two- to three-fold higher Si plasma level, but in contrast aged persons and pregnant women have significantly lower values of Si. For human beings, the daily ingested amount of Si is estimated to be 30–50 mg and the same amount is eliminated in urine. After meals, the plasma concentration of Si increases 30–50%, and returns after a few hours to the initial level (71). The regulatory mechanisms of Silicon homeostasis require elucidation.

The catalytic action of a biomolecule to accomplish the spherical oligomerization of silicic acid yielding SOSA is presumed. Proteins catalyzing high-grade polymerization of silicic acid have been found (67) in algae (silaffin) or sponges (silicatein). Biomolecules favoring the spherical oligomerization of silicic acid have not yet been identified.

The mechanism of ATPase regulation by the probable endogenous ligand SOSA is not fully understood. According to the unusual physical–chemical properties of SOSA and to its specific interactions with the  $Na^+$ ,  $K^+$ ,  $H^+$ ,  $Mg^{2+}$ , or  $Ca^{2+}$  ions and with the ATPase protein, some challenging proposals for the regulation mechanism may be formulated.

#### ASSUMED REGULATORY MECHANISM OF SOSA

Transmembrane ion pumping may be physically influenced by SOSA located in a receptor cavity along the ion-transport pathway of the ATPase molecule. The SOSA molecule should behave like a multi-anionic gel with selective binding or permeability effects on cationic species depending on their concentration, charge and size.

The interaction of SOSA with the ATPase protein is assumed on the cytoplasmic site of the ATPases in the E1 conformation as revealed by charge transfer investigations with the dye RH421 (60). According to these data, SOSA should inhibit the X1E1 state, manifesting complex interactions with different X ions ( $Na^+$ ,  $K^+$ , or  $H^+$ ) and with  $Na^+$  ions in the  $(Na)NaE1$  state. Structural details of the very complex binding of the sodium ions to the NKA in the state preceding phosphorylation have been disclosed by high-resolution crystal structure (72).

The ion-sensitive nature of our ATPase inhibitory factors MCS (disclosed as SOSA) was remarked in Ref. (60) assuming that the concentration of the  $Na^+$  and of other ions may cause significant structural rearrangements. The *in vitro* ion-sensitive structure of SOSA is also supported by DLS measurements. Tentatively, it could be assumed that the SOSA structure at  $[Na^+] < 5\text{mM}$  inhibits the sodium pump but the structure at  $[Na^+] \geq 5\text{mM}$  should activate it. A possible activation mechanism could be by favoring the differentiate binding of  $Na^+$  ions to the E1 conformation and promoting the phosphorylation step. Accepting this hypothesis, the generally found cytosolic level of  $[Na^+] = 5.0\text{ mM}$  in the

eukaryotic cells could be a consequence of the structural change of the archaic ligand SOSA, which happens accidentally at this  $Na^+$  ion concentration.

The here proposed probably archaic regulatory mechanism of the NKA pump by ion-dependent structural changes of the endogenous ligand could also work for other ATPases. The ion concentration threshold for the activation of other ATPases should depend upon the concentration of the ions to be pumped and their interaction with SOSA. This regulatory mechanism, assuming the ion-sensitive structural variation of the archaic ligand SOSA, can explain the astonishing manifoldness of the same well-conserved ion-pumping mechanism in the hitherto identified more than 200 different P-type ATPase pumps.

#### CONCLUSIONS AND OUTLOOK

With almost all Si-OH bonds disposed on the external surface of SOSA, this substance should bind preferentially to hydrophilic domains of the target proteins. In P-type ATPases, the well-conserved phosphorylation site between the cytoplasmic domains P and N provides an adequate binding site for SOSA molecules similar to decavanadate (Figure 1).

Although SOSA inhibits NKA at sub-micro-molar concentration, its direct competition with ouabain for the hydrophobic steroid binding site in NKA is less probable. The non-competitive binding mechanism of our hydrophilic ATPase inhibitor and ouabain was confirmed by the fluorescence dye measurements of Stimac et al. (59, 60). An apparent competition may appear if the NKA conformation required for SOSA binding and that required for ouabain binding are different (C. Toyoshima, personal communication).

The ion-concentration dependent structural changes of SOSA suggest a probable archaic regulation mechanism of the sodium pump and of other ATPases where the pump ligand is sensing the nature and molarity of the ions to be pumped. It is a challenging idea that the transmembrane ion pumping with fundamental importance for many essential life processes should be regulated by the cation-sensitive structure of an inorganic acid.

The identified chemical properties and enzymatic activities of SOSA are congruent with the predicted characteristics of the EDLFs of the P-type ATPases. Table 1 shows a synoptic presentation of the assumed characteristics of the putative EDLF factors in comparison with that of candidate substances: ouabain, marinobufagenin, vanadate, and the here disclosed SOSA.

Summarizing the characteristics of the SOSA, it may be concluded that these match the predicted criteria of the endogenous ligands of the NKA and probably of further P-type ATPases:

- a. SOSA inhibits with similar potencies ( $IC_{50} \sim 0.2\text{--}0.5\text{ }\mu\text{g/mL}$ ) the ouabain-sensitive  $Na^+/K^+$ -ATPase from rabbit medulla and the ouabain-insensitive enzyme from rat.
- b. SOSA inhibits  $Ca^{2+}$ -ATPase from SR,  $H^+/K^+$ -ATPase from gastric membrane and of K-dp-ATPase from *Escherichia coli* with  $IC_{50}$  values in the range of  $0.2\text{--}0.5\text{ }\mu\text{g/mL}$ .
- c. There are no sensitive methods to differentiate between mono and oligo-silicic acids in cells. But the assay of the NKA-inhibitory factors EDLF in urine and plasma suggest the probable presence of SOSA in these biological fluids.

**Table 1 | Comparison of the endogenous ATPase ligand candidates.**

	<b>EDLF</b>	<b>Ouabain</b>	<b>Marino-bufagin</b>	<b>Vanadate</b>	<b>SOSA</b>
<b>Characteristics</b>					
C-1 Chemical nature	ND	Cardiac glycoside	Bufadienolide steroid	Inorganic, poly-oxo-acid	Inorganic poly-oxo-acid
C-2 Molar mass (kDa)	0.4–5.0	0.58	0.4	0.12–1.5	1.4–6.0
C-3 Stability by drying	Low	Stable	Stable	Limited	Low
C-4 Structural stability	ND	Stable	Stable	pH sensitive equilibra	Cation and pH sensitive
C-5 Nature to ATPase binding site	ND	Hydrophobic	Hydrophobic	Hydrophilic	Hydrophilic
C-6 Distribution	Ubiquitous	Only in a few plant species	Predominantly in amphibians	Limited	Ubiquitous
C-7 Mammalian occurrence	Yes	No	No	Only in traces	Ubiquitous
C-8 Evolutionary age million years Myr	>3500 Myr	<60 Myr	<360 Myr	>3500 Myr	>3500 Myr
<b>Biochemistry</b>					
B-1 Toxicity	Low	High	High	Moderate	Low
B-2 Biosynthesis	Predicted	Only in a few plant species	Predominantly in amphibians	Improbable	From monosilicic-acid
B-3 Na,K-ATPase	Inhibitor	Inhibitor	Inhibitor	Inhibitor	Inhibitor
B-4 SERCA	Inhibitor	No	No	Inhibitor	Inhibitor
B-5 H/K-ATPase	Inhibitor	No	No	Inhibitor	Inhibitor
B-6 K-db-ATPase	Inhibitor	No	No	Inhibitor	Inhibitor

ND: not determined.

- d. Monosilicic acid is present in almost all cells; thus, its adequate transformation into SOSA can occur *in situ*, catalyzed and/or controlled by proteins.
- e. Preliminary data revealed a reduced toxicity by per-oral and a moderate toxicity by intravenous or intramuscular administration of SOSA. Renal elimination is assumed.
- f. There are no reasons to doubt the presence of silicic acid and of SOSA in the early history of the evolution.
- g. SOSA is stable in solution for several years but it loses its activity by freeze-drying probably through the forced intermolecular condensation of water molecules.

It is planned to obtain further structural details of the interaction of SOSA with ATPase proteins with possible implication for the regulation of the pump. One of the very intriguing questions is to investigate the influence of the concentration of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ , and other ions on the structure of SOSA. The further great challenge for ongoing research is to establish the role of SOSA in ATPase related cellular and physiological processes and to explore its possible health-care applications in some connected pathologies (73, 74).

## ACKNOWLEDGMENTS

We are grateful to Chikashi Toyoshima for the critical proofreading of the manuscript and for the suggested clear differentiation between the hydrophobic binding site of CSs and the assumed hydrophilic receptor site of SOSA in  $\text{Na}^+/\text{K}^+$ -ATPase. We thank very much Harvey Gonick for his consistent help in improving the manuscript and for the linguistic corrections.

## REFERENCES

- Hajdu S, Szent-Györgyi A. Action of DOC and serum on the frog heart. *Am J Physiol* (1952) **168**:159–70.
- Szent-Györgyi A. *Chemical Physiology of Contraction in Body and Heart Muscle*. New York, NY: Academic Press (1953). p. 79–91.
- Hughes J, Smith TW, Kosterlitz HW, Fothergill LA, Morgan BA, Morris HR. Identification of two related pentapeptides from brain with potent opiate agonist activity. *Nature* (1975) **258**:577–9. doi:10.1038/258577a0
- Schatzmann HJ. Herzglycoside als Hemmstoffe für den aktiven Kalium und Natrium Transport durch die Erythrozytenmembran. *Helv Physiol Pharmacol Acta* (1953) **11**:346–54.
- Skou JC. The influence of some cations on an adenosine triphosphatase from peripheral nerves. *Biochim Biophys Acta* (1957) **23**:394–401. doi:10.1016/0006-3002(57)90343-8
- De Wardener HE, Mills IH, Clapham WF, Hayter CJ. Studies on the efferent mechanism of the sodium diuresis which follows the administration of intravenous saline in the dog. *Clin Sci* (1961) **21**:249–58.
- Buckalew VM, Martinez FJ, Green WE. The effect of dialysates and ultrafiltrates of plasma of saline-loaded dogs on toad bladder sodium transport. *J Clin Invest* (1970) **49**:926–35. doi:10.1172/JCI106312
- Gonick HC, Kramer HJ, Paul W, Lu E. Circulating inhibitor of sodium-potassium activated adenosine triphosphatase after expansion of extracellular fluid volume in rats. *Clin Sci Mol Med* (1977) **53**:329–34.
- Gruber KA, Whitaker JM, Buckalew VM. Endogenous, digitalis-like substance in plasma of volume-expanded dogs. *Nature* (1980) **287**:743–5. doi:10.1038/287743a0
- DeBold AJ, Borenstein HB, Veress A, Sonnenberg H. Rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats. *Life Sci* (1981) **28**:89–94. doi:10.1016/0024-3205(81)90370-2
- Rakowski RF, Gadsby DC, DeWeer P. Voltage dependence of the Na/K pump. *J Membr Biol* (1997) **155**:105–12. doi:10.1007/s002329900162
- Glynn IM. A hundred years of sodium pumping. *Annu Rev Physiol* (2002) **64**:1–18. doi:10.1146/annurev.physiol.64.081501.130716
- Kaplan JH. Biochemistry of Na/K-ATPase. *Annu Rev Biochem* (2002) **71**:511–35. doi:10.1146/annurev.biochem.71.102201.141218

14. Jørgensen PL, Håkansson KO, Karlsson SJD. Structure and mechanism of Na/K-ATPase: functional sites and their interactions. *Annu Rev Physiol* (2003) **65**:817–49. doi:10.1146/annurev.physiol.65.092101.142558
15. Møller JV, Juul B, le Maire M. Structural organization, ion transport, and energy transduction of P-type ATPases. *Biochim Biophys Acta* (1996) **1286**:1–51. doi:10.1016/0304-4157(95)00017-8
16. Kühlbrandt W. Biology, structure and mechanism of P-type ATPases. *Nat Rev Mol Cell Biol* (2004) **5**:282–95. doi:10.1038/nrm1354
17. Axelsen KB, Palmgren MG. Evolution of substrate specificities in the P-type ATPase superfamily. *J Mol Evol* (1998) **46**:84–101. doi:10.1007/PL00006286
18. Pedersen CNS, Axelsen KB, Harper JE, Palmgren MG. Evolution of plant P-type ATPases. *Front Plant Sci* (2012) **3**:31. doi:10.3389/fpls.2012.00031
19. Toyoshima C, Nakasako M, Nomura H, Ogawa H. Crystal structure of the calcium pump of sarcoplasmic reticulum at 2.6 Å resolution. *Nature* (2000) **405**:647–55. doi:10.1038/35015017
20. Toyoshima C, Nomura H. Structural changes in the calcium pump accompanying the dissociation of calcium. *Nature* (2002) **418**:605–11. doi:10.1038/nature00944
21. Toyoshima C, Mitzutani T. Crystal structure of the calcium pump with a bound ATP analogue. *Nature* (2004) **430**:529–35. doi:10.1038/nature02680
22. Toyoshima C, Nomura H, Tsuda T. Luminal gating mechanism revealed in calcium pump crystal structures with phosphate analogues. *Nature* (2004) **432**:361–8. doi:10.1038/nature02981
23. Olesen C, Picard M, Winther AM, Gyrup C, Morth JP, Oxvig C, et al. The structural basis of calcium transport by the calcium pump. *Nature* (2007) **450**:1036–42. doi:10.1038/nature06418
24. Toyoshima C. Structural aspects of ion pumping by  $\text{Ca}^{2+}$ -ATPase of sarcoplasmic reticulum. *Arch Biochem Biophys* (2008) **476**:3–11. doi:10.1016/j.abb.2008.04.017
25. Hoffman BG, Bigger JT. Digitalis and allied cardiac glycosides. 9th ed. In: Gilman AG, Goodman LS, Gilman A, editors. *The Pharmacologic Basis of Therapeutics*. New York, NY: McGraw-Hill (1992). p. 729–60.
26. Paula S, Tabet MR, Ball WRJ. Interactions between cardiac glycosides and Na/KATP-ase: three-dimensional structure-activity relationship models for ligand binding to the E2-Pi form of the enzyme versus activity inhibition. *Biochemistry* (2005) **44**:498–510. doi:10.1021/bi048680w
27. Katz A, Lifshitz Y, Bab-Dinitz E, Kapri-Pardesh E, Goldshleger Tal D, Karlsson SJD. Selectivity of digitalis glycosides for isoforms of human Na/K-ATPase. *J Biol Chem* (2010) **285**:19582–92. doi:10.1074/jbc.M110.119248
28. Apell HJ. Structure-function relationship in P-type ATPases – a biophysical approach. *Rev Physiol Biochem Pharmacol* (2003) **150**:1–35. doi:10.1007/s10254-003-0018-9
29. Toyoshima C, Kanai R, Cornelius F.  $\text{Na}^+/\text{K}^+$ -ATPase: new light on the oldest ion pump. *Structure* (2011) **19**:1732–8. doi:10.1016/j.str.2011.10.016
30. Morth JP, Pedersen BP, Toustrup-Jensen MS, Sørensen TL, Petersen J, Andersen JP, et al. Crystal structure of the sodium-potassium pump. *Nature* (2007) **450**:1043–9. doi:10.1038/nature06419
31. Shinoda T, Ogawa H, Cornelius F, Toyoshima C. Crystal structure of the sodium-potassium pump at 2.4 Å resolution. *Nature* (2009) **459**:446–50. doi:10.1038/nature07939
32. Ogawa H, Shinoda T, Cornelius F, Toyoshima C. Crystal structure of the sodium-potassium pump ( $\text{Na}^+/\text{K}^+$ -ATPase) with bound potassium and ouabain. *Proc Natl Acad Sci U S A* (2009) **106**:13742–7. doi:10.1073/pnas.0907054106
33. Yatime L, Laursen M, Morth JP, Esmann M, Nissen P, Fedosova N. Structural insights into the high affinity binding of cardiotonic steroids to the  $\text{Na}^+/\text{K}^+$ -ATPase. *J Struct Biol* (2011) **174**:296–306. doi:10.1016/j.jsb.2010.12.004
34. Laursen M, Yatime L, Nissen P, Fedosova NU. Crystal structure of the high affinity Na/K-ATPase ouabain complex with  $\text{Mg}^{2+}$  bound in the cation binding site. *Proc Natl Acad Sci U S A* (2013) **110**:10958–63. doi:10.1073/pnas.1222308110
35. Sandtner W, Egwolff B, Khalili-Araghi F, Sánchez-Rodríguez JE, Roux B, Bezanilla F, et al. Ouabain binding site in a functioning  $\text{Na}^+/\text{K}^+$ -ATPase. *J Biol Chem* (2011) **286**(44):38177–83. doi:10.1074/jbc.M111.267682
36. Dostanic-Larson I, Van Huysse JW, Lorenz JN, Lingrel JB. The highly conserved cardiac glycoside binding site of Na/K-ATPase plays a role in blood pressure regulation. *Proc Natl Acad Sci U S A* (2005) **102**:15845–50. doi:10.1073/pnas.0507358102
37. Lingrel JB. The physiological significance of the cardiotonic steroid/ouabain-binding site of the Na/K-ATPase. *Annu Rev Physiol* (2010) **72**:395–412. doi:10.1146/annurev-physiol-021909-135725
38. Goto A, Yamada K, Yagi N, Yoshioka M, Sugimoto T. Physiology and pharmacology of endogenous digitalis-like factors. *Pharmacol Rev* (1992) **44**:377–99.
39. Hollenberg NK, Graves SW. Endogenous sodium pump inhibition: current status and therapeutic opportunities. *Prog Drug Res* (1996) **46**:9–42.
40. Buckalew VM. Endogenous digitalis-like factors: an historical overview. *Front Biosci* (2005) **10**:2325–34. doi:10.2741/1701
41. Schonher W, Scheiner-Bobis G. Endogenous and exogenous cardiac glycosides and their mechanisms of action. *Am J Cardiovasc Drugs* (2007) **7**:173–89. doi:10.2165/00129784-200707030-00004
42. Nesher M, Shpolansky U, Rosen H, Lichtstein D. The digitalis-like steroid hormones: new mechanisms of action and biological significance. *Life Sci* (2007) **80**:2093–107. doi:10.1016/j.lfs.2007.03.013
43. Bagrov AY, Shapiro JI, Fedorova OV. Endogenous cardiotonic steroids physiology pharmacology, and novel therapeutic targets. *Pharmacol Rev* (2009) **61**:9–38. doi:10.1124/pr.108.000711
44. Graves SW, Tao QF, Markides KE, Williams GH, Hollenberg NK. A labile sodium pump inhibitor from the peritoneal dialysate of hypertensive renal failure patients: estimates of potency. *Clin Exp Hypertens* (1998) **20**:611–6. doi:10.3109/10641969809053239
45. Cantley L, Josephson L, Warner R, Yanagisawa M, Lechene C, Guidotti G. Vanadate is a potent Na/K-ATPase inhibitor found in ATP derived from muscle. *J Biol Chem* (1977) **252**:7421–3.
46. Beaugé LA, Glynn IM. Commercial ATP containing traces of vanadate alters the response of (Na+ + K+) ATPase to external potassium. *Nature* (1978) **272**:551–2. doi:10.1038/272551a0
47. Pick U. The interaction of vanadate ions with the Ca-ATPase from sarcoplasmic reticulum. *J Biol Chem* (1982) **257**:6111–9.
48. Csermely P, Martonosi A, Levy GC, Ejchart AJ. 51V-n.m.r. analysis of the binding of vanadium (V) oligoanions to sarcoplasmic reticulum. *Biochem J* (1985) **230**:807–15.
49. Aureliano M, Madeira VMC. Interactions of vanadate oligomers with sarcoplasmic reticulum Ca-ATPase. *Biochim Biophys Acta* (1994) **1221**:259–71. doi:10.1016/0167-4889(94)90249-6
50. Aureliano M, Gandara RMC. Decavanadate effects in biological system. *J Inorg Biochem* (2005) **99**:979–85. doi:10.1016/j.jinorgbio.2005.02.024
51. Aureliano M. Recent perspectives into biochemistry of decavanadate. *World J Biol Chem* (2011) **2**:215–25. doi:10.4331/wjbc.v2.i10.215
52. Hua S, Inesi G, Toyoshima C. Distinct topologies of mono-and decavanadate binding and photo-oxidative cleavage in the sarcoplasmic reticulum ATPase. *J Biol Chem* (2000) **275**:30546–50. doi:10.1074/jbc.M003218200
53. Goc A. Biological activity of vanadium compounds. *Cent Eur J Biol* (2006) **1**:314–32. doi:10.2478/s11535-006-0029-z
54. Thompson HK, Orwig C. Vanadium in diabetes: 100 year from phase 0 to phase 1. *J Inorg Biochem* (2006) **100**:1925–35. doi:10.1016/j.jinorgbio.2006.08.016
55. Kramer HJ, Krampitz G, Baecker A, Meyer-Lehnert H. Vanadium diascorbates are strong candidates for endogenous ouabain-like factors in human urine: effects on Na/K-ATPase enzyme kinetics. *Biochim Biophys Res Commun* (1995) **213**:289–94. doi:10.1006/bbrc.1995.2128
56. Kerek F, Stimac R, Apell HJ, Freudenmann F, Moroder L. Characterization of the macrocyclic carbon suboxide factors as potent Na/K-ATPase and SR Ca-ATPase inhibitors. *Biochim Biophys Acta* (2002) **1567**:213–20. doi:10.1016/S0005-2736(02)00609-0
57. Kappe T, Ziegler E. Carbon suboxide in preparative organic chemistry. *Angew Chem Int Ed* (1974) **13**:491–558. doi:10.1002/anie.197404911
58. Kerek F. The structure of the digitalis like and natriuretic factors identified as macrocyclic derivatives of the inorganic carbon suboxide. *Hypertens Res* (2000) **23**:S33–8. doi:10.1291/hypres.23.Supplement\_S33
59. Stimac R, Kerek F, Apell HJ. Macrocyclic carbon suboxide oligomers as potent inhibitors of the Na/K-ATPase. *Ann N Y Acad Sci* (2003) **986**:327–9. doi:10.1111/j.1749-6632.2003.tb07204.x
60. Stimac R, Kerek F, Apell HJ. Mechanism of the Na/K-ATPase inhibition by MCS derivatives. *J Membr Biol* (2005) **205**:89–101. doi:10.1007/s00232-005-0767-2
61. Mathews WR, DuCharme DW, Hamlyn JM, Harris DW, Mandel F, Clark MA, et al. Mass spectral characterization of an endogenous digitalislike factor from human plasma. *Hypertension* (1991) **17**:930–5. doi:10.1161/01.HYP.17.6.930
62. Hilton PJ, White RW, Lord GA, Garner GV, Gordon BD, Hilton MJ, et al. An inhibitor of the sodium pump obtained from human placenta. *Lancet* (1996) **48**:303–5. doi:10.1016/S0140-6736(96)02257-X

63. Weiler EW, Khalil-Manesh F, Gonick HC, Prins BA, Purdy RE, Sen-sharma DK. Na/K-ATPase inhibitor dissociated from hypertension associated plasma protein. *Am J Hypertens* (1999) **12**:364–73. doi:10.1016/S0895-7061(00)86962-9
64. Kerek F, Voicu VA. Endogenous digitalis identified as sub-nano silicic acid SNSA. *Therapeutics Pharmacology and Clinical Toxicology* (2010) **XIV**(S3):p. 19.
65. Kerek F, Voicu VA. Sub-nano silicic acid, the putative biologically active form of silica. *Basic Clin Pharmacol Toxicol* (2011) **109**:S26. doi:10.1111/j.1742-7843.2011.00731x
66. Chen H, Tecklenburg RE. Characterization of low- and intermediate molecular weight hydrogen silsesquioxanes by mass spectrometry. *J Am Soc Mass Spectrom* (2006) **17**:1437–41. doi:10.1016/j.jasms.2006.06.010
67. Belton DJ, Deschaume O, Perry CC. An overview of the fundamentals of the chemistry of silica with relevance to biosilicification and technological advances. *FEBS J* (2012) **279**:171–120. doi:10.1111/j.1742-4658.2012.08531.x
68. Currie AH, Perry CC. Silica in plants: biological, biochemical and chemical studies. *Ann Bot* (2007) **100**:1383–9. doi:10.1093/aob/mcm247
69. Epstein E. Silicon. *Annu Rev Plant Physiol Plant Mol Biol* (1999) **50**:641–64. doi:10.1146/annurev.aplant.50.1.641
70. Sripanyakorn S, Jugdaohsingh R, Thompson RPH, Powell JJ. Dietary silicon and bone health. *BNF Nutr Bull* (2005) **30**:222–30. doi:10.1111/j.1467-3010.2005.00507.x
71. Sripanyakorn S, Jugdaohsingh R, Dissayabutr W, Anderson SH, Thompson RP, Powell JJ. The comparative absorption of silicon from different foods and food supplements. *Br J Nutr* (2009) **102**:825–34. doi:10.1017/S0007114509311757
72. Kanai R, Ogawa H, Vilsen B, Cornelius F, Toyoshima C. Crystal structure of a  $\text{Na}^+$ -bound Na/K-ATPase preceding the E1P state. *Nature* (2013) **502**:201–6. doi:10.1038/nature12578
73. Kerek F. *Biologically Active Silicic Acid*. PCT application, WO 2010/012507 A1. (2010).
74. Yatime L, Buch-Pedersen MJ, Musgaard M, Morth JP, Lund Winther AM, Pedersen BP, et al. P-type ATPases as drug targets: tools for medicine and science. *Biochim Biophys Acta* (2011) **1787**:207–20. doi:10.1016/j.bbabi.2008.12.019

**Conflict of Interest Statement:** Dr. Franz Kerek is founder and majority shareholder of the company SiNatur GmbH in Munich, Germany where the R&D activities of the novel factor SOSA were conducted. Prof. Victor A. Voicu has no conflict of interests.

*Received: 03 October 2014; accepted: 16 December 2014; published online: 23 January 2015.*

*Citation:* Kerek F and Voicu VA (2015) Spherical oligo-silicic acid SOSA disclosed as possible endogenous digitalis-like factor. *Front. Endocrinol.* **5**:233. doi:10.3389/fendo.2014.00233

*This article was submitted to Neuroendocrine Science, a section of the journal Frontiers in Endocrinology.*

*Copyright © 2015 Kerek and Voicu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*

## ADVANTAGES OF PUBLISHING IN FRONTIERS



### FAST PUBLICATION

Average 90 days  
from submission  
to publication



### COLLABORATIVE PEER-REVIEW

Designed to be rigorous –  
yet also collaborative, fair and  
constructive



### RESEARCH NETWORK

Our network  
increases readership  
for your article



### OPEN ACCESS

Articles are free to read,  
for greatest visibility



### TRANSPARENT

Editors and reviewers  
acknowledged by name  
on published articles



### GLOBAL SPREAD

Six million monthly  
page views worldwide



### COPYRIGHT TO AUTHORS

No limit to  
article distribution  
and re-use



### IMPACT METRICS

Advanced metrics  
track your  
article's impact



### SUPPORT

By our Swiss-based  
editorial team