

Supplementary information: For Climate drives the high survival synchrony among temperate songbird.

Among-species heterogeneity in sample size does not bias the estimation of synchrony.

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1 Goal and rationale

The primary goal of this appendix is to test the idea that among-species heterogeneity in sample size might bias the estimation of synchrony among species. The verbal argument behind this concern is that the time-dynamic of the most common species might “spill over” the estimation of the time-dynamic common to all species. A suggested solution to this problem has been not to model synchrony explicitly, so that the model does not contain an across-species time-dynamic, but instead to estimate species-specific time dynamics independently, and reconstruct the synchrony a posteriori. Technically, this would be done by fitting a species-by-time random interaction without fitting the main random effect of time (but fitting the main random effect of species).

However, theory suggest that mixed models are generally able to handle unbalanced data, and that the estimation of synchrony should not be biased by unequal sample sizes (Pinheiro & Bates, 2000). Moreover, theory also suggests that the “solution” of not modeling synchrony explicitly and reconstructed it a posteriori from model prediction is flawed, and will generally lead to under-estimation (Hadfield et al., 2010; Houslay & Wilson, 2017).

Here, we test that these two expectations hold for the special case of mixed mark-recapture models used in the main text, and that the results presented in the main text do not suffer from a bias due to unequal sample size among species. To this end, we first simulate multi-species mark-recapture data with synchrony among species or with no synchrony among species. We use the same sample size per year and per species as in the real data, but halved, in order to speed up modeling. We then analyze the simulated data either with a model that explicitly contain synchrony, or with a model that does not contain explicit synchrony but only allow for its reconstruction from species-specific estimates.

We first show that when synchrony is present in simulated data, a model explicitly modeling synchrony can recover it. We then show that on the same data, a model that does not explicitly model synchrony, but instead makes it estimation possible a posteriori, fails to estimate synchrony. Finally, we show that the first model (explicitly modeling synchrony) does not detect synchrony if none is simulated in the data.

2 Load packages and data

We need R2jags to communicate with JAGS. We use `reshape` to convert a matrix into a data frame. We use `lme4` for variance partitioning. We also load the real sample sizes per species per year.

```
setwd(dir = "~/Documents/CanberraPostDoc/BirdSnynchro/penguinteam/SimulsForPubli/")
library(R2jags)
library(reshape)
library(lme4)

SpNumbers <- read.table(file = "SpNumbers.txt", header=TRUE)
SpNumbers <- SpNumbers[, -1]
```

Graphic control:

```
szgr <- 2
szax <- 1.3
marr <- c(4, 4, 1, 1) + 0.1
setPar<-function(){
  par(las=1,mar=marr, cex=szgr, cex.lab=szax , cex.axis=szax, lwd=2 ,pch=1, las=1)
}
```

```
setPar()
prospecies <- rowMeans(SpNumbers)

barplot(prospecies[order(prospecies)],
        xlab="species", ylab="Sample size")
```

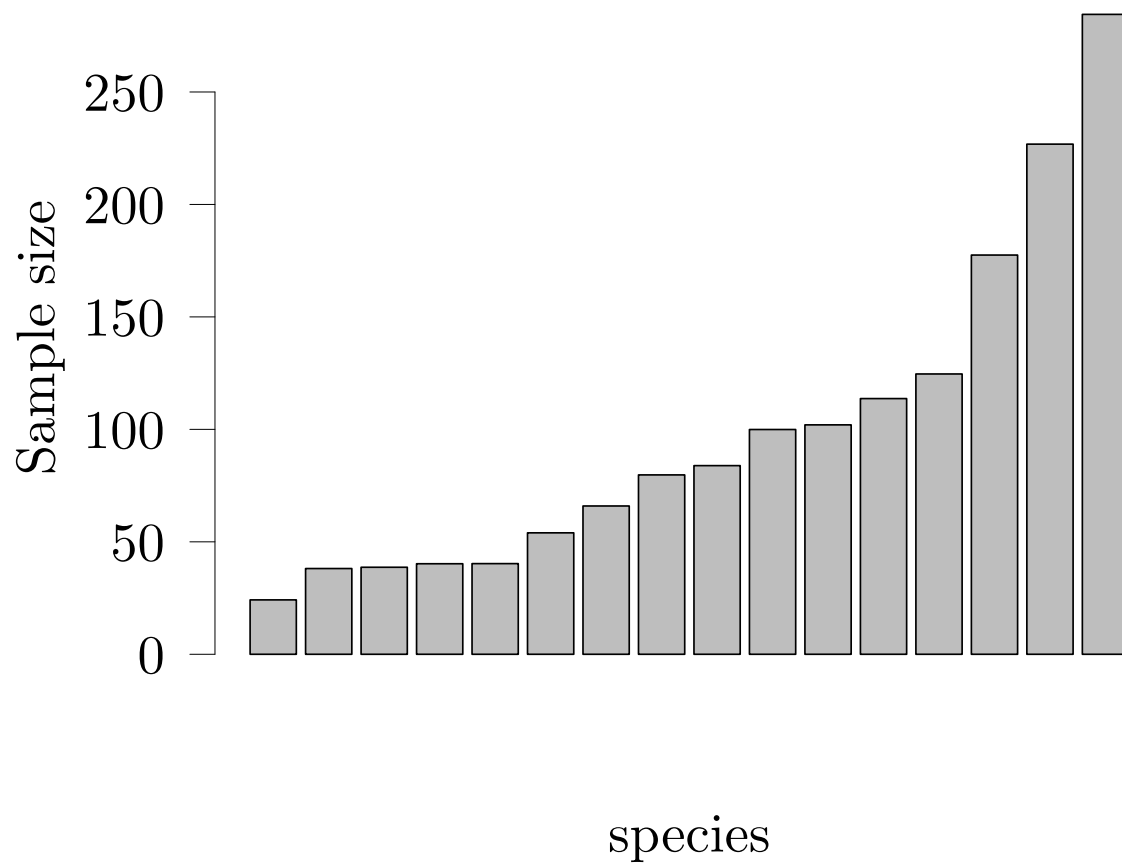


Figure 1: Distribution of the sample size per species.

3 Strong synchrony in the data

3.1 Data simulation

We use an additive model to simulate data, so that the effect of time is the same for all species, leading to strong (virtually perfect) synchrony.

```
n.occasions <- ncol(SpNumbers) # Number of capture occasions
meanphi <- 0 #logit intercept survival (i.e 0.5 on survival scale)
effect_Occasions<-c(0,rnorm(n = n.occasions-2,mean = 0,sd = 0.5))#time effect

n.sp <- nrow(SpNumbers)
effect_Sp<-c(0,rnorm(n = n.sp-1,mean=0,sd=1))

#marked <- round(100*(1:n.sp)^-2) # Annual number of newly marked individuals
p <- rep(0.4, n.occasions-1)#constant recapture probability

# Define matrices with survival and recapture probabilities
PHI <- matrix( data = NA, ncol = n.occasions-1, nrow = sum( SpNumbers) )
SpTots <- rowSums(SpNumbers)
CumulTots <- c(0, cumsum(SpTots))
sp_ind <- vector(length = sum(SpTots))
for (sp in 1:n.sp)
{
  spe <- effect_Sp[sp]
  sp_ind[(CumulTots[sp]+1):CumulTots[sp+1]] <- sp
  #nind <- sum(SpNumbers[sp,])
  for (occ in 1:(n.occasions-1))
  {
    oce <- effect_Occasions[occ]
    PHI[(CumulTots[sp]+1):CumulTots[sp+1],occ] <- 1/(1+exp(-( meanphi + oce + spe) ) )
  }
}

P <- matrix(p, ncol = n.occasions-1, nrow = sum(SpTots))
```

```
# Define function to simulate a capture-history (CH) matrix
simul.cjs <- function(PHI, P, SpTots){
  n.occasions <- dim(PHI)[2] + 1
  CH <- matrix(0, ncol = n.occasions, nrow = sum(SpTots))
  # Define a vector with the occasion of marking
  mark.occ <- rep(1:length(SpTots), SpTots[1:length(SpTots)])

  for (i in 1:sum(SpTots)){# Fill the CH matrix
    CH[i, mark.occ[i]] <- 1 # Write an 1 at the release occasion
    if (mark.occ[i]==n.occasions) next
    for (t in (mark.occ[i]+1):n.occasions){
      # Bernoulli trial: does individual survive occasion?
      sur <- rbinom(1, 1, PHI[i,t-1])
      if (sur==0) break # If dead, move to next individual

      rp <- rbinom(1, 1, P[i,t-1])# Bernoulli trial: is individual recaptured?
      if (rp==1) CH[i,t] <- 1
    } #t
  } #i
  return(CH)
```

```
}
```

```
# Execute function
CH <- simul.cjs(PHI, P, SpTots)

# Create vector with occasion of marking
get.first <- function(x) min(which(x!=0))
f <- apply(CH, 1, get.first)
```

Functions to help fit MR models:

```
known.state.cjs <- function(ch){
  state <- ch
  for (i in 1:dim(ch)[1]){
    n1 <- min(which(ch[i,]==1))
    n2 <- max(which(ch[i,]==1))
    state[i,n1:n2] <- 1
    state[i,n1] <- NA
  }
  state[state==0] <- NA
  return(state)
}

cjs.init.z <- function(ch,f){
  for (i in 1:dim(ch)[1]){
    if (sum(ch[i,])==1) next
    n2 <- max(which(ch[i,]==1))
    ch[i,f[i]:n2] <- NA
  }
  for (i in 1:dim(ch)[1]){
    ch[i,1:f[i]] <- NA
  }
  return(ch)
}
```

3.2 Interaction model with explicit synchrony

We now fit a model that try to estimate the synchrony on the data containing strong synchrony.

```
sink("cjs-phi(spXt)p(.).jags")
cat([1183 chars quoted with '"'],fill = TRUE)
sink()
```

```
# Bundle data
jags.data <- list(y = CH, f = f, nind = dim(CH)[1],
                 n.occasions = dim(CH)[2],
                 z = known.state.cjs(CH),
                 sp_ind=sp_ind, n.sp=n.sp)

#### Initial values####
#interaction init

beta.tXsp_ini<-function()
{beta.tXsp<-matrix(NA,nrow = n.occasions-1,ncol = n.sp)
```

```

for (i in 1:(n.occasions-1))
{
  for (j in 1:n.sp)
  {
    beta.tXsp[i,j]<-rnorm(n = 1,mean = 0,sd = 1)
  }
}
return(beta.tXsp)
}

inits <- function(){list(z = cjs.init.z(CH, f),
                        phi.mean = runif(1, 0, 1), p.mean = runif(1, 0, 1),
                        beta.t=c(rnorm(n = n.occasions-1,mean = 0,sd=1 ) ),
                        beta.sp=c(rnorm(n = n.sp,mean = 0,sd=1 ) ),
                        beta.tXsp= beta.tXsp_ini() )}

# Parameters monitored
parameters <- c("phi.mean", "p.mean",
                "beta.t","beta.sp",
                "beta.tXsp", "sigma.sp",
                "sigma.t", "sigma.tXsp")

# MCMC settings
ni <- 11000
nt <- 10
nb <- 1000
nc <- 3

cjs.interactionS <- jags(jags.data, inits, parameters,
                        "cjs-phi(spXt)p(.).jags",
                        n.chains = nc, n.thin = nt,
                        n.iter = ni, n.burnin = nb,
                        working.directory = getwd())

print(cjs.interactionS, digits = 3)

```

```
load(file = "ForCluster/cjs.interactionS_dataSynchro")
```

The synchrony is well recovered graphically (Figure 2)

```

# Summarize posteriors

predictions <- cjs.interactionS$BUGSoutput$mean$beta.tXsp +
  matrix(cjs.interactionS$BUGSoutput$mean$beta.t,
        nrow = n.occasions-1, ncol = n.sp, byrow = FALSE) +
  matrix(cjs.interactionS$BUGSoutput$mean$beta.sp,
        nrow = n.occasions-1, ncol=n.sp, byrow = TRUE)

```

The intra-Class correlation is close to one, as revealed by the point estimate:

```

(cjs.interactionS$BUGSoutput$mean$sigma.t^2)/
  (cjs.interactionS$BUGSoutput$mean$sigma.tXsp^2+
   cjs.interactionS$BUGSoutput$mean$sigma.t^2)
## [1] 0.9717394

```

As well as the posterior distribution (Figure 3).

```

setPar()
plot(predictions[,1], type="l", ylim=c(-3,3),
      xlab="year", main="", ylab="Latent survival probability")
for(i in 1:n.sp)
{
  lines(predictions[,i])
}

```

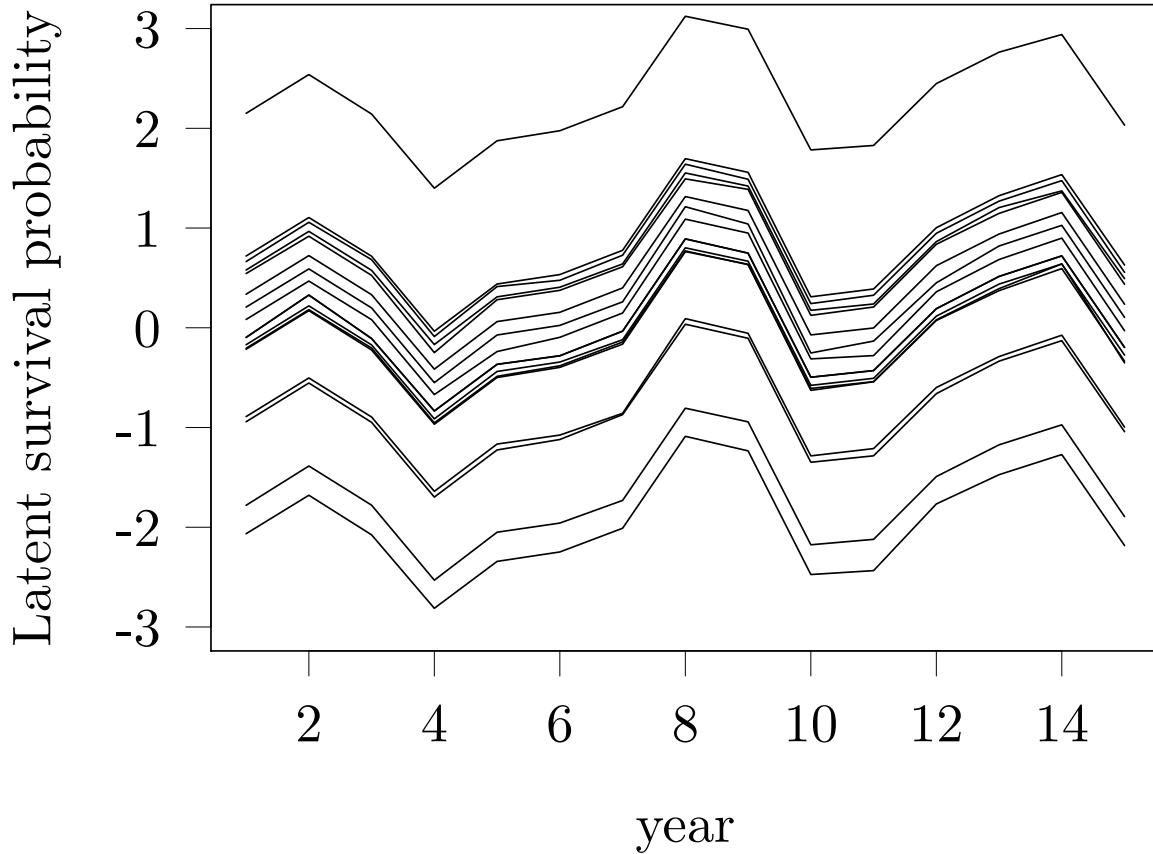


Figure 2: Predicted latent values for survival for all species and years. The model used for predictions explicitly modeled synchrony, and the data were simulated with a strong synchrony. Synchrony is well recovered by the model, as evidenced by the parallel time-dynamics among species.

```

iccpst <- vector(length=1000)
for (ch in 1:1)
{
  for (itt in 1:1000)
  {
    iccpst[itt + 1000*(ch-1)] <-
      cjs.interactionS$BUGSoutput$sims.array[itt, ch,"sigma.t"]^2 /
      ( cjs.interactionS$BUGSoutput$sims.array[itt, ch,"sigma.t"]^2 +
        cjs.interactionS$BUGSoutput$sims.array[itt, ch,"sigma.tXsp"]^2)
  }
}

sdprior <- runif(n = 10000, min = 0, max = 10)
sdprior2 <- sdprior

```

```
iccprior <- (sdprior^2)/(sdprior^2+sdprior2^2)
```

```
setPar()
plot(density(iccpost), xlim = c(0,1), lwd=5,
     main="",
     xlab="ICC", ylab="Probability density")

lines(density(iccprior), lwd=5, lty=2, col="gray")

legend(x = "topleft", legend=c("Posterior", "Prior"), lty=c(1,2), lwd=5,
      col=c("black", "gray"))
```

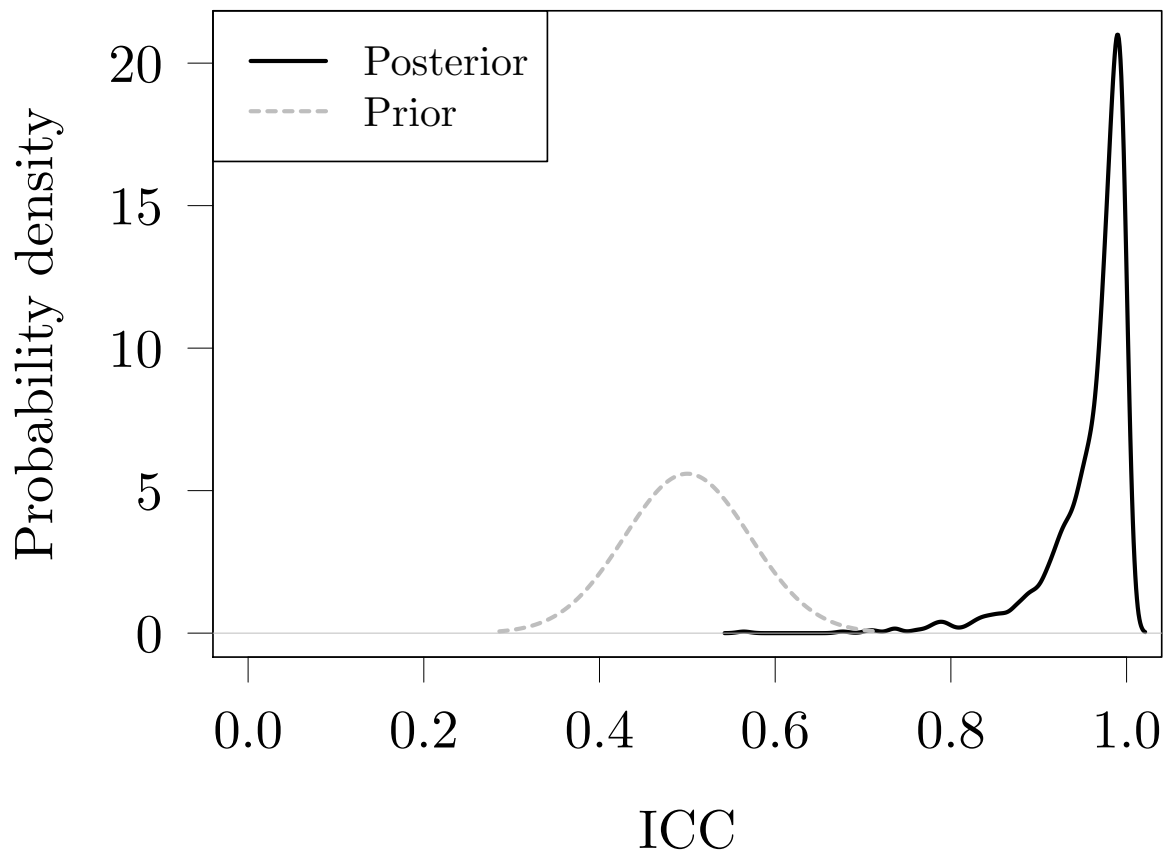


Figure 3: Probability density distributions of the intra-class correlation (ICC) of time, measuring the proportion of time-variation in survival rate that is common to all bird species. Data were simulated with a strong synchrony (ICC close to one). Prior and posterior distributions are shown.

Note that the synchrony manifests itself on the latent scale, but also on the scale of the data (Figure 4).


```

setPar()
plot(1/(1+exp(-predictions))[,1], type="l", ylim=c(0, 1), xlab="year",
     ylab="predicted survival probability")
for(i in 1:n.sp)
{
  lines(1/(1+exp(-predictions))[,i])
}

```

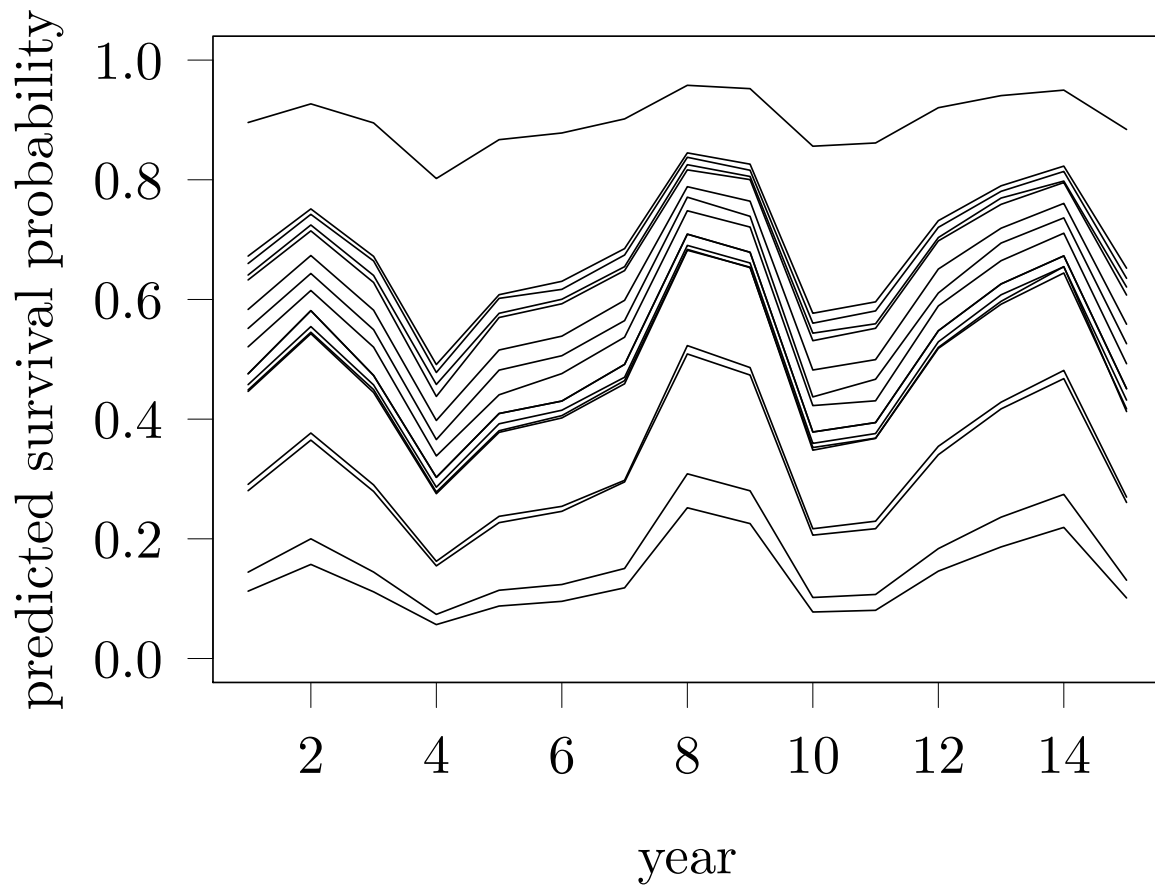


Figure 4: Predicted survival probability by species and by year. Data were simulated with a strong synchrony (ICC close to one). The model used for prediction explicitly modeled synchrony.

3.3 Interactive model without explicit synchrony

We analyze the same data, but this time using a model that does not explicitly contain synchrony.

```
sink("cjs-phi(spXt_no_t)p(.).jags")
cat([1043 chars quoted with ''],fill = TRUE)
sink()

# Bundle data
jags.data <- list(y = CH, f = f, nind = dim(CH)[1],
                 n.occasions = dim(CH)[2],
                 z = known.state.cjs(CH),
                 sp_ind=sp_ind, n.sp=n.sp)

#### Initial values####
#interaction init

beta.tXsp_ini<-function()
{beta.tXsp<-matrix(NA,nrow = n.occasions-1,ncol = n.sp)
for (i in 1:(n.occasions-1))
{
  for (j in 1:n.sp)
  {
    beta.tXsp[i,j]<-rnorm(n = 1,mean = 0,sd = 1)
  }
}
return(beta.tXsp)
}

inits <- function(){list(z = cjs.init.z(CH, f),
                        phi.mean = runif(1, 0, 1), p.mean = runif(1, 0, 1),
                        beta.sp=c(rnorm(n = n.sp,mean = 0,sd=1 ) ),
                        beta.tXsp= beta.tXsp_ini() )}

# Parameters monitored
parameters <- c("phi.mean", "p.mean",
                "beta.sp","beta.tXsp",
                "sigma.sp", "sigma.tXsp")

# MCMC settings
ni <- 11000
nt <- 10
nb <- 1000
nc <- 3

cjs.interactionNot <- jags(jags.data, inits,
                          parameters, "cjs-phi(spXt_no_t)p(.).jags",
                          n.chains = nc, n.thin = nt,
                          n.iter = ni, n.burnin = nb,
                          working.directory = getwd())

print(cjs.interactionNot, digits = 3)

load(file = "ForCluster/cjs.interactionSNot_dataSynchro")
```

```

predictions <- matrix(cjs.interactionNot$BUGSoutput$mean$phi.mean,
                      nrow = n.occasions-1, ncol=n.sp, byrow = TRUE) +
  cjs.interactionNot$BUGSoutput$mean$beta.tXsp +
  matrix(cjs.interactionNot$BUGSoutput$mean$beta.sp,
         nrow = n.occasions-1, ncol=n.sp, byrow = TRUE)

```

Graphically, much less synchrony is apparent (Figure 5)

```

setPar()
plot(predictions[,1], type="l", ylim=c(-3,3), xlab="year", main="",
      ylab="Latent survival probability")
for(i in 1:n.sp)
{
  lines(predictions[,i])
}

```

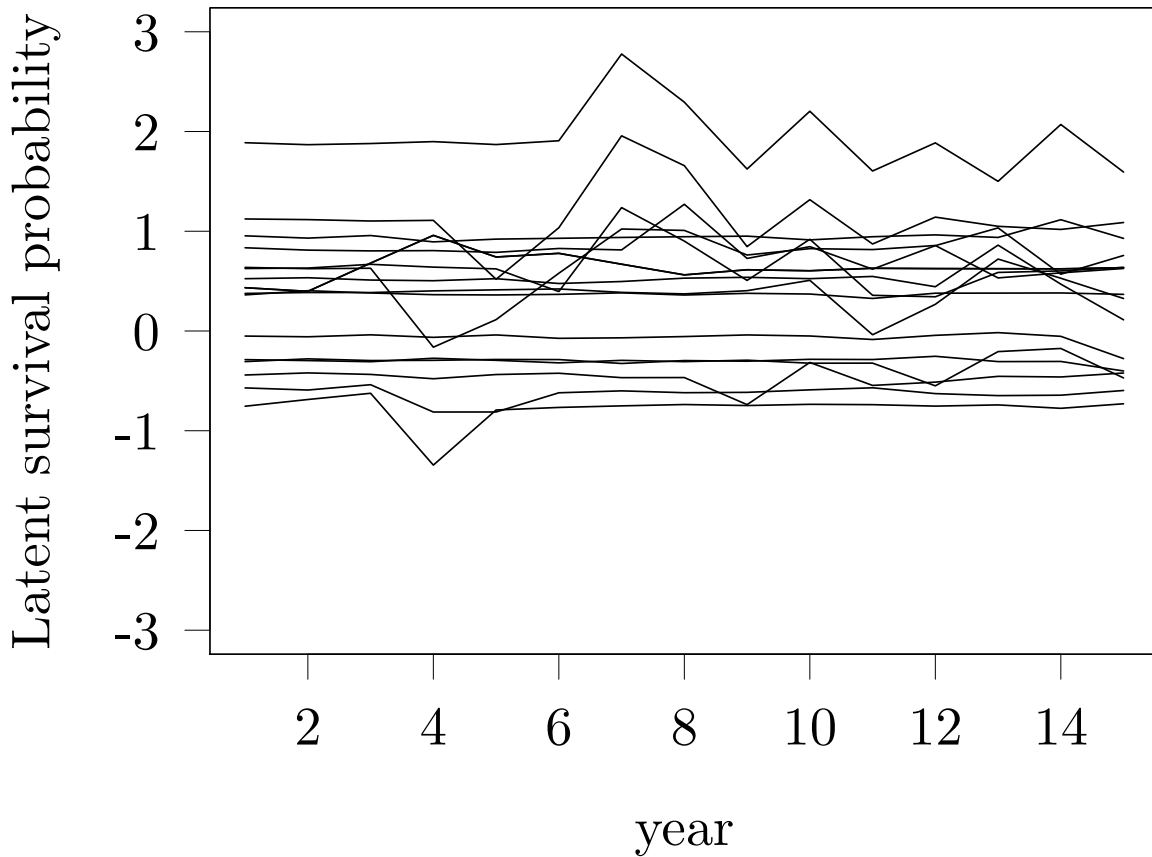


Figure 5: Predicted latent values for survival for all species and years. The model used for predictions did not model explicit synchrony but allow for it with a species-by-time random interaction, and the data were simulated with a strong synchrony. Synchrony is poorly recovered by the model, as evidenced by the lack of parallel time-dynamics among species.

The intra class correlation (ratio of temporal variance common to all species over the total temporal variance) is weak and close to zero.

```

meltedpredictions <- melt(predictions, varnames = c("year", "species"))
modvardec <- lmer(value ~ 1 + (1|year) + (1|species), data=meltedpredictions)
summary(modvardec)

```

```

setPar()
plot(1/(1+exp(-predictions))[,1], type="l", ylim=c(0, 1), xlab="year",
     ylab="predicted survival probability")
for(i in 1:n.sp)
{
  lines(1/(1+exp(-predictions))[,i])
}

```

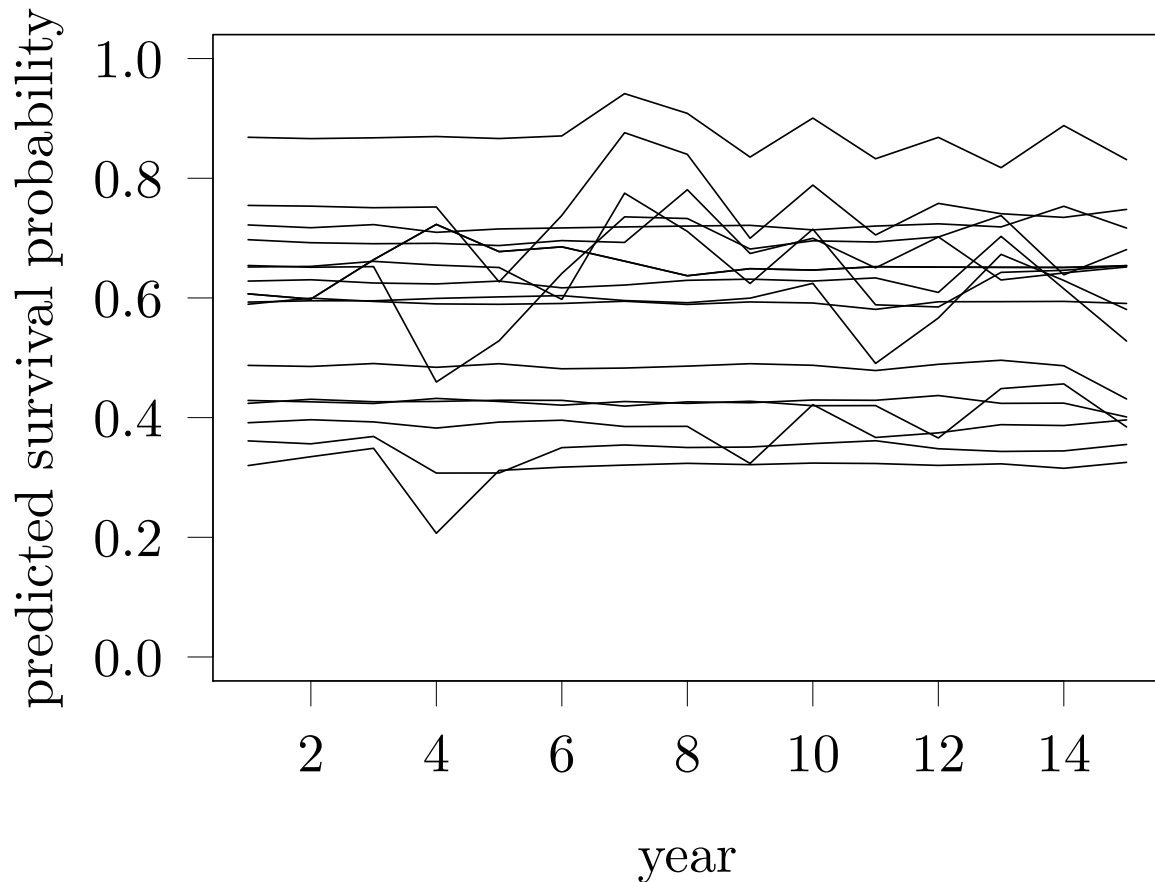


Figure 6: Predicted survival probability by species and by year. Data were simulated with a strong synchrony (ICC close to one). The model used for prediction explicitly did not model synchrony.

```

## Linear mixed model fit by REML ['lmerMod']
## Formula: value ~ 1 + (1 | year) + (1 | species)
## Data: meltedpredictions
##
## REML criterion at convergence: -70.6
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -4.4036 -0.3071  0.0389  0.3131  4.4310
##
## Random effects:
## Groups Name Variance Std.Dev.
## species (Intercept) 0.523956 0.7238
## year (Intercept) 0.004045 0.0636
## Residual 0.027855 0.1669
## Number of obs: 240, groups: species, 16; year, 15
##

```

```
## Fixed effects:
##           Estimate Std. Error t value
## (Intercept)  0.3405    0.1820   1.871

sptimeV <- sigma(modvardec)^2
timeV <- as.numeric(VarCorr(modvardec)$year)

timeV/(timeV+sptimeV)

## [1] 0.1268008
```

4 No synchrony in the data

We now simulate data without any synchrony to check whether a model would detect spurious synchrony because of the over-abundance of some species.

```
effect_OccasionsSP<-matrix(c(rnorm(n = (n.occasions-1)*n.sp,
                                mean = 0,sd = 0.5)),
                            nrow=n.sp,
                            ncol=(n.occasions-1))#time effect

effect_Sp<-c(0,rnorm(n = n.sp-1,mean=0,sd=1))

# Define matrices with survival and recapture probabilities
PHI <- matrix( data = NA, ncol = n.occasions-1, nrow = sum( SpNumbers) )
SpTots <- rowSums(SpNumbers)
CumulTots <- c(0, cumsum(SpTots))
sp_ind <- vector(length = sum(SpTots))
for (sp in 1:n.sp)
{
  spe <- effect_Sp[sp]
  sp_ind[(CumulTots[sp]+1):CumulTots[sp+1]] <- sp
  #nind <- sum(SpNumbers[sp,])
  for (occ in 1:(n.occasions-1))
  {
    oce <- effect_Occasions[occ]
    PHI[(CumulTots[sp]+1):CumulTots[sp+1],occ] <-
      1/(1+exp(-( meanphi + effect_OccasionsSP[sp,occ] + spe) ))
  }
}

# Execute function
CH <- simul.cjs(PHI, P, SpTots)

# Create vector with occasion of marking
get.first <- function(x) min(which(x!=0))
f <- apply(CH, 1, get.first)
```

We fit the model with explicit synchrony.

```
load(file = "ForCluster/cjs.interactionS_dataNoSynchro")
```

There graphically, no synchrony is apparent (Figure 7), which matches the data-generating process that did not have any synchrony.

```
# Summarize posteriors

predictions <- cjs.interactionSdataNoS$BUGSoutput$mean$beta.tXsp +
  matrix(cjs.interactionSdataNoS$BUGSoutput$mean$beta.t,
          nrow = n.occasions-1, ncol = n.sp, byrow = FALSE) +
  matrix(cjs.interactionSdataNoS$BUGSoutput$mean$beta.sp,
          nrow = n.occasions-1, ncol=n.sp, byrow = TRUE)
```

The intra-Class correlation is close to zero (Figure 9) as expected.

```
(cjs.interactionSdataNoS$BUGSoutput$mean$sigma.t^2)/
  (cjs.interactionSdataNoS$BUGSoutput$mean$sigma.tXsp^2+
   cjs.interactionSdataNoS$BUGSoutput$mean$sigma.t^2)

## [1] 0.07141729
```

```

iccpst <- vector(length=1000)
for (ch in 1:1)
{
  for (itt in 1:1000)
  {
    iccpst[itt + 1000*(ch-1)] <-
      cjs.interactionSdataNoS$BUGSoutput$sims.array[itt, ch,"sigma.t"]^2 /
      ( cjs.interactionSdataNoS$BUGSoutput$sims.array[itt, ch,"sigma.t"]^2 +
        cjs.interactionSdataNoS$BUGSoutput$sims.array[itt, ch,"sigma.tXsp"]^2)
  }
}

```

```

setPar()
plot(predictions[,1], type="l", ylim=c(-3,3),
      xlab="year", main="", ylab="Latent survival probability")
for(i in 1:n.sp)
{
  lines(predictions[,i])
}

```

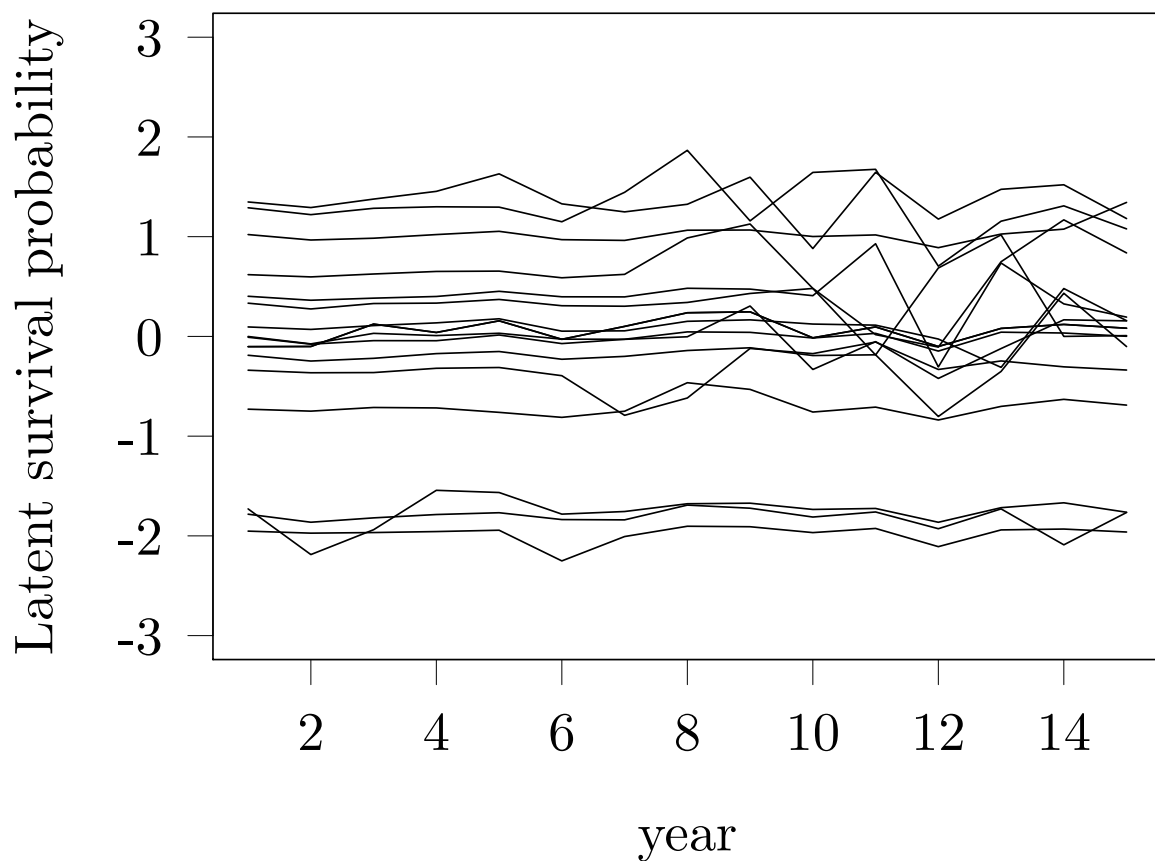


Figure 7: Predicted latent values for survival for all species and years. The model used for predictions explicitly modeled synchrony, and the data were simulated without any synchrony. The model correctly identifies the lack of synchrony, as evidenced by the non-parallel time-dynamics among species.


```

setPar()
plot(1/(1+exp(-predictions))[,1], type="l", ylim=c(0, 1), xlab="year",
     ylab="predicted survival probability")
for(i in 1:n.sp)
{
  lines(1/(1+exp(-predictions))[,i])
}

```

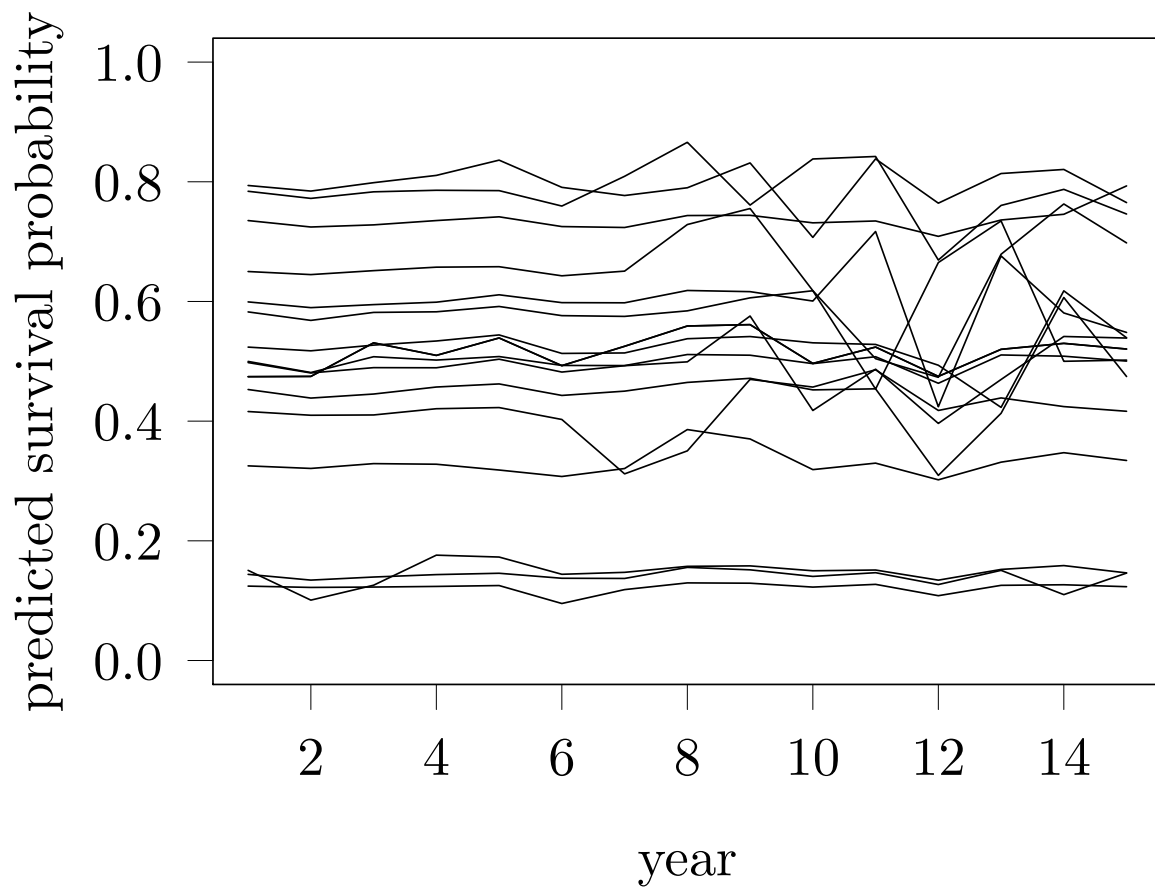


Figure 8: Predicted survival probability by species and by year. Data were simulated without any synchrony (ICC close to zero). The model used for prediction explicitly modeled synchrony.

```

setPar()
plot(density(iccpost), xlim = c(0,1), lwd=5,
     main="",
     xlab="ICC", ylab="Probability density")

lines(density(iccprior), lwd=5, lty=2, col="gray")

legend(x = "topright", legend=c("Posterior", "Prior"), lty=c(1,2), lwd=5,
      col=c("black", "gray"))

```

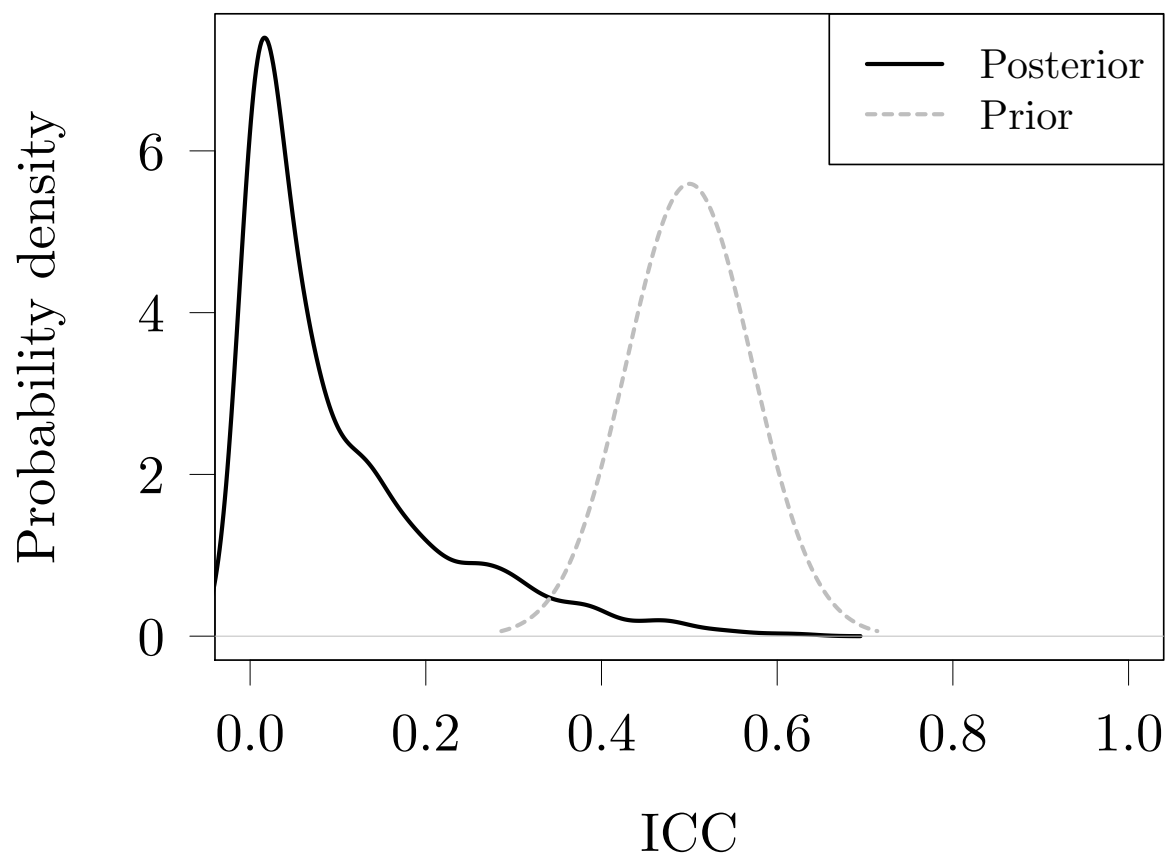


Figure 9: Probability density distributions of the intra-class correlation (ICC) of time, measuring the proportion of time-variation in survival rate that is common to all bird species. Data were simulated without any synchrony (ICC close to zero). Prior and posterior distributions are shown.

References

- Hadfield, J.D., Wilson, A.J., Garant, D., Sheldon, B.C. & Kruuk, L.E.B. 2010. The misuse of BLUP in ecology and evolution. *Am. Nat.* **175**: 116–25.
- Houslay, T.M. & Wilson, A.J. 2017. Avoiding the misuse of BLUP in behavioural ecology. *Behav. Ecol.* **28**: 948–952.
- Pinheiro, J.C. & Bates, D.M. 2000. *Mixed-Effects Models in S and S-PLUS*. Statistics and Computing Series. Springer-Verlag, New York, NY.