

SubRead Annotations

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Setup

```
knitr::opts_chunk$set(fig.width=12, fig.height=8,
  fig.path=paste0('Figures_subReadAnnotations/'),
  dev='pdf',
  echo=TRUE, warning=FALSE, message=FALSE,
  error=TRUE)
```

```
suppressWarnings(library('tidyverse'))
suppressWarnings(library('magrittr'))
suppressWarnings(library('knitr'))
```

Exons

load table from subRead

```
(exon_anno_tbl <- rtracklayer::import('/Volumes/GenomeDK/ms_tools/subread-2.0.0-Linux-x86_64/annotation.

## # A tibble: 261,752 x 8
##   seqnames start   end width strand GeneID   ExonID score
##   <fct>     <int> <int> <int> <fct>  <chr>    <chr>   <dbl>
## 1 chr1     11874 12227   354 +    100287102 1         0
## 2 chr1     12613 12721   109 +    100287102 2         0
## 3 chr1     13221 14409  1189 +    100287102 3         0
## 4 chr1     14362 14829   468 -     653635   1         0
## 5 chr1     14970 15038    69 -     653635   2         0
## 6 chr1     15796 15947   152 -     653635   3         0
## 7 chr1     16607 16765   159 -     653635   4         0
## 8 chr1     16858 17055   198 -     653635   5         0
## 9 chr1     17233 17368   136 -     653635   6         0
```

```
## 10 chr1      17606 17742   137 -      653635    7      0
## # ... with 261,742 more rows
```

add exon nr and counts

```
(exon_cnts <- exon_anno_tbl %>%
  mutate(GeneID = as.character(GeneID)) %>%
  group_by(GeneID) %>%
  summarize(exon_cnt = n(),
            tr_exons_width = sum(end-start)))
```

```
## # A tibble: 28,395 x 3
##   GeneID exon_cnt tr_exons_width
##   <chr>      <int>      <int>
## 1 1      8      1758
## 2 10     3      1415
## 3 100    13      1851
## 4 1000   20      4874
## 5 10000  23      8411
## 6 100008587 1      155
## 7 100008588 1      1868
## 8 100008589 1      5069
## 9 100009601 2       71
## 10 100009602 2       71
## # ... with 28,385 more rows
```

```
(exon_anno_tbl %<>%
  group_by(GeneID) %>%
  mutate(exon_nr = as.integer(ifelse(strand == '+',
                                     rank(start, ties.method = 'first'),
                                     rank(-start, ties.method = 'first'))),
         exon_width = end-start) %>%
  dplyr::select(GeneID, ExonID, exon_nr, width) %>%
  left_join(., exon_cnts) %>%
  ungroup %>%
  mutate(class = case_when(.$exon_cnt == 1 ~ 'monoexonic',
                           .$exon_nr == 1 ~ 'multiexonic first exon',
                           .$exon_nr == .$exon_cnt ~ 'multiexonic last exon',
                           .$exon_nr < .$exon_cnt ~ 'multiexonic internal')))
```

```
## # A tibble: 261,752 x 7
##   GeneID ExonID exon_nr width exon_cnt tr_exons_width class
##   <chr>   <chr>   <int> <int>   <int>      <int> <chr>
## 1 100287102 1       1   354     3      1649 multiexonic fir~
## 2 100287102 2       2   109     3      1649 multiexonic int~
## 3 100287102 3       3  1189     3      1649 multiexonic las~
## 4 653635   1      11   468    11      1758 multiexonic las~
## 5 653635   2      10    69    11      1758 multiexonic int~
## 6 653635   3       9   152    11      1758 multiexonic int~
## 7 653635   4       8   159    11      1758 multiexonic int~
## 8 653635   5       7   198    11      1758 multiexonic int~
## 9 653635   6       6   136    11      1758 multiexonic int~
## 10 653635  7       5   137    11      1758 multiexonic int~
## # ... with 261,742 more rows
```

ENTREZ to ENSEMBL

```
library("AnnotationDbi")
library("org.Hs.eg.db")
columns(org.Hs.eg.db)

## [1] "ACCNUM"      "ALIAS"      "ENSEMBL"    "ENSEMBLPROT"
## [5] "ENSEMBLTRANS" "ENTREZID"   "ENZYME"     "EVIDENCE"
## [9] "EVIDENCEALL"  "GENENAME"   "GO"         "GOALL"
## [13] "IPI"         "MAP"        "OMIM"       "ONTOLOGY"
## [17] "ONTOLOGYALL"  "PATH"       "PFAM"       "PMID"
## [21] "PROSITE"     "REFSEQ"     "SYMBOL"     "UCSCKG"
## [25] "UNIGENE"     "UNIPROT"

entrezid_map <- select(org.Hs.eg.db,
  key=unique(exon_anno_tbl$GeneID), columns=c('ENSEMBL', 'SYMBOL', 'REFSEQ'),
  keytype="ENTREZID")

head(entrezid_map)
```

```
##      ENTREZID      ENSEMBL      SYMBOL      REFSEQ
## 1 100287102 ENSG00000223972 DDX11L1 NR_046018
## 2   653635 ENSG00000227232 WASH7P NR_024540
## 3 102466751 ENSG00000278267 MIR6859-1 NR_106918
## 4 100302278 ENSG00000284332 MIR1302-2 NR_036051
## 5   645520 ENSG00000237613 FAM138A NR_026818
## 6   79501 ENSG00000186092 OR4F5 NM_001005484
```

Gencode gene info

```
(gencode_tbl <- rtracklayer::import('/Volumes/GenomeDK/annotations/hg38/Gencode_v28/gencode.v28.genes.g
  dplyr::mutate(ENSEMBL = sub('\\..*', '', gene_id)) %>%
  dplyr::select(ENSEMBL, gene_name, gene_type))

## # A tibble: 58,381 x 3
##      ENSEMBL      gene_name      gene_type
##      <chr>      <chr>      <chr>
## 1 ENSG00000223972 DDX11L1      transcribed_unprocessed_pseudogene
## 2 ENSG00000227232 WASH7P      unprocessed_pseudogene
## 3 ENSG00000278267 MIR6859-1      miRNA
## 4 ENSG00000243485 RP11-34P13.3 lincRNA
## 5 ENSG00000284332 MIR1302-2      miRNA
## 6 ENSG00000237613 FAM138A      lincRNA
## 7 ENSG00000268020 OR4G4P      unprocessed_pseudogene
## 8 ENSG00000240361 OR4G11P      transcribed_unprocessed_pseudogene
## 9 ENSG00000186092 OR4F5      protein_coding
## 10 ENSG00000238009 RP11-34P13.7 lincRNA
## # ... with 58,371 more rows

sort(table(gencode_tbl$gene_type),decreasing = T)

##
##      protein_coding      processed_pseudogene
##      19901      10219
```

##	lincRNA	antisense
##	7490	5501
##	unprocessed_pseudogene	misc_RNA
##	2664	2213
##	snRNA	miRNA
##	1900	1881
##	TEC	snoRNA
##	1067	943
##	sense_intronic	transcribed_unprocessed_pseudogene
##	899	855
##	processed_transcript	rRNA
##	556	544
##	transcribed_processed_pseudogene	IG_V_pseudogene
##	472	188
##	sense_overlapping	IG_V_gene
##	183	144
##	transcribed_unitary_pseudogene	TR_V_gene
##	123	106
##	unitary_pseudogene	TR_J_gene
##	95	79
##	scaRNA	bidirectional_promoter_lncRNA
##	49	47
##	polymorphic_pseudogene	IG_D_gene
##	38	37
##	TR_V_pseudogene	3prime_overlapping_ncRNA
##	33	32
##	Mt_tRNA	IG_J_gene
##	22	18
##	pseudogene	IG_C_gene
##	18	14
##	IG_C_pseudogene	ribozyme
##	9	8
##	TR_C_gene	sRNA
##	6	5
##	TR_D_gene	TR_J_pseudogene
##	4	4
##	IG_J_pseudogene	non_coding
##	3	3
##	Mt_rRNA	translated_processed_pseudogene
##	2	2
##	IG_pseudogene	macro_lncRNA
##	1	1
##	scRNA	vaultRNA
##	1	1

combine subread exons with gencode those

```
(exon_anno_tbl %<>%
  left_join(., dplyr::rename(entrezid_map, GeneID=ENTREZID)) %>%
  left_join(., gencode_tbl))
```

```
## # A tibble: 4,801,919 x 12
```

```
##   GeneID ExonID exon_nr width exon_cnt tr_exons_width class      ENSEMBL
```

```
##      <chr>  <chr>      <int> <int>      <int>      <int> <chr>      <chr>
## 1 100287~ 1          1   354          3          1649 multiexo~ ENSG000~
## 2 100287~ 2          2   109          3          1649 multiexo~ ENSG000~
## 3 100287~ 3          3  1189          3          1649 multiexo~ ENSG000~
## 4 653635 1          11   468         11          1758 multiexo~ ENSG000~
## 5 653635 2          10    69         11          1758 multiexo~ ENSG000~
## 6 653635 3          9    152         11          1758 multiexo~ ENSG000~
## 7 653635 4          8    159         11          1758 multiexo~ ENSG000~
## 8 653635 5          7    198         11          1758 multiexo~ ENSG000~
## 9 653635 6          6    136         11          1758 multiexo~ ENSG000~
## 10 653635 7          5    137         11          1758 multiexo~ ENSG000~
## # ... with 4,801,909 more rows, and 4 more variables: SYMBOL <chr>,
## #   REFSEQ <chr>, gene_name <chr>, gene_type <chr>
```

```
sort(table(exon_anno_tbl$gene_type),decreasing = T)
```

```
##
##              protein_coding                      lincRNA
##              4355224                          20056
##              antisense transcribed_unprocessed_pseudogene
##              8754                              6492
##              processed_transcript                miRNA
##              4628                              1859
## transcribed_unitary_pseudogene                    polymorphic_pseudogene
##              1361                              1202
##              unprocessed_pseudogene transcribed_processed_pseudogene
##              1010                              551
##              sense_intronic                      snoRNA
##              383                                372
##              bidirectional_promoter_lncRNA        sense_overlapping
##              236                                224
##              TR_C_gene                          processed_pseudogene
##              128                                77
##              snRNA                              TEC
##              33                                29
##              rRNA                              scaRNA
##              18                                17
##              misc_RNA                          3prime_overlapping_ncRNA
##              11                                3
##              non_coding                        ribozyme
##              3                                1
##              scRNA                            unitary_pseudogene
##              1                                1
```

```
save(exon_anno_tbl, file='../data/subRead_exon_annotations.RData')
```

Transcripts table

```
(transcripts <- rtracklayer::import('/Volumes/GenomeDK/ms_tools/subread-2.0.0-Linux-x86_64/annotation/hg38/'))
```

```
## GRanges object with 28421 ranges and 5 metadata columns:
```

```
##              seqnames      ranges strand |      name      score
##              <Rle>        <IRanges> <Rle> | <character> <numeric>
```

```
##      [1]      chr1    11874-14409    + |    100287102      0
##      [2]      chr1    14362-29370    - |      653635      0
##      [3]      chr1    17369-17436    - |    102466751      0
##      [4]      chr1    30366-30503    + |    100302278      0
##      [5]      chr1    34611-36081    - |      645520      0
##      ...      ...      ...      ...      ...
## [28417] NW_009646208.1    21277-23071    + |    102723722      0
## [28418] NW_009646208.1    25775-30157    - |    101929829      0
## [28419] NW_011332699.1    64666-77517    - |    105379672      0
## [28420] NW_011332699.1    76210-170143    + |      85316      0
## [28421] NW_011332701.1  2741969-2798262 - |    102725021      0
##      itemRgb      thick      blocks
##      <character>      <IRanges>      <IRangesList>
##      [1]      #FF0000    11874-14409      1-354,740-848,1348-2536
##      [2]      #FF0000    14362-29370    1-468,609-677,1435-1586,...
##      [3]      #FF0000    17369-17436      1-68
##      [4]      #FF0000    30366-30503      1-138
##      [5]      #FF0000    34611-36081    1-564,667-871,1111-1471
##      ...      ...      ...
## [28417]      #FF0000    21277-23071    1-250,493-576,966-1008,...
## [28418]      #FF0000    25775-30157    1-254,353-494,949-1136,...
## [28419]      #FF0000    64666-77517    1-220,453-564,3270-3342,...
## [28420]      #FF0000    76210-170143  1-203,1673-1862,56463-56625,...
## [28421]      #FF0000  2741969-2798262 1-731,1522-1659,11728-11842,...
## -----
##      seqinfo: 55 sequences from an unspecified genome; no seqlengths
```

```
(tr_anno_tbl <- exon_anno_tbl %>%
```

```
  distinct(GeneID, exon_cnt, tr_exons_width, ENSEMBL, SYMBOL, REFSEQ, gene_name, gene_type))
```

```
## # A tibble: 287,698 x 8
```

```
##      GeneID exon_cnt tr_exons_width ENSEMBL SYMBOL REFSEQ gene_name
##      <chr>      <int>      <int> <chr>      <chr>      <chr>      <chr>
## 1 100287~      3      1649 ENSG0000~ DDX11L1 NR_0460~ DDX11L1
## 2 653635      11      1758 ENSG0000~ WASH7P NR_0245~ WASH7P
## 3 102466~      1      67 ENSG0000~ MIR6859~ NR_1069~ MIR6859-1
## 4 100302~      1      137 ENSG0000~ MIR1302~ NR_0360~ MIR1302-2
## 5 645520      3      1127 ENSG0000~ FAM138A NR_0268~ FAM138A
## 6 79501      1      917 ENSG0000~ OR4F5 NM_0010~ OR4F5
## 7 79501      1      917 ENSG0000~ OR4F5 NP_0010~ OR4F5
## 8 729737      3      5471 <NA>      LOC7297~ NR_0399~ <NA>
## 9 102725~      4      1169 <NA>      LOC1027~ NR_1483~ <NA>
## 10 102723~     11      2109 ENSG0000~ WASH9P XR_0017~ RP11-34P1~
## # ... with 287,688 more rows, and 1 more variable: gene_type <chr>
```

-> no good fit with RefSeq !?

```
(tr_anno_tbl <- exon_anno_tbl %>%
```

```
  distinct(GeneID, exon_cnt, tr_exons_width, ENSEMBL, SYMBOL, gene_name, gene_type))
```

```
## # A tibble: 31,756 x 7
```

```
##      GeneID exon_cnt tr_exons_width ENSEMBL SYMBOL gene_name gene_type
##      <chr>      <int>      <int> <chr>      <chr>      <chr>      <chr>
## 1 100287102      3      1649 ENSG000~ DDX11L1 DDX11L1 transcrib~
## 2 653635      11      1758 ENSG000~ WASH7P WASH7P unprocess~
## 3 102466751      1      67 ENSG000~ MIR685~ MIR6859-1 miRNA
```

```
## 4 100302278      1      137 ENSG000~ MIR130~ MIR1302-2 miRNA
## 5 645520        3      1127 ENSG000~ FAM138A FAM138A  lincRNA
## 6 79501         1      917 ENSG000~ OR4F5  OR4F5    protein_c~
## 7 729737       3      5471 <NA>    LOC729~ <NA>    <NA>
## 8 102725121    4      1169 <NA>    LOC102~ <NA>    <NA>
## 9 102723897    11     2109 ENSG000~ WASH9P  RP11-34P~ unprocess~
## 10 102465909   1      67 ENSG000~ MIR685~ MIR6859-2 miRNA
## # ... with 31,746 more rows
```

-> much more reasonable list!

```
filter(tr_anno_tbl, gene_type == 'protein_coding')
```

```
## # A tibble: 19,463 x 7
##   GeneID exon_cnt tr_exons_width ENSEMBL SYMBOL gene_name gene_type
##   <chr>   <int>      <int> <chr>   <chr>  <chr>   <chr>
## 1 79501     1        917 ENSG0000001~ OR4F5  OR4F5    protein_co~
## 2 729759    1        938 ENSG0000002~ OR4F29 OR4F29    protein_co~
## 3 81399     1        938 ENSG0000002~ OR4F16 OR4F16    protein_co~
## 4 81399     1        938 ENSG0000002~ OR4F16 OR4F29    protein_co~
## 5 81399     1        938 ENSG0000002~ OR4F16 OR4F3     protein_co~
## 6 148398    14       2540 ENSG0000001~ SAMD11 SAMD11    protein_co~
## 7 26155     19       2781 ENSG0000001~ NOC2L  NOC2L     protein_co~
## 8 339451    12       3438 ENSG0000001~ KLHL17 KLHL17     protein_co~
## 9 84069     15       2545 ENSG0000001~ PLEKH~ PLEKHN1    protein_co~
## 10 84808    4       3900 ENSG0000001~ PERM1  PERM1     protein_co~
## # ... with 19,453 more rows
```

```
save(tr_anno_tbl, file='../data/subRead_tr_annotations.RData')
```

sessionInfo

```
sessionInfo()
```

```
## R version 3.5.0 (2018-04-23)
## Platform: x86_64-apple-darwin15.6.0 (64-bit)
## Running under: macOS 10.14.6
##
## Matrix products: default
## BLAS: /Library/Frameworks/R.framework/Versions/3.5/Resources/lib/libRblas.0.dylib
## LAPACK: /Library/Frameworks/R.framework/Versions/3.5/Resources/lib/libRlapack.dylib
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## attached base packages:
## [1] parallel stats4      stats      graphics  grDevices  utils      datasets
## [8] methods    base
##
## other attached packages:
## [1] org.Hs.eg.db_3.7.0  AnnotationDbi_1.44.0 IRanges_2.14.10
## [4] S4Vectors_0.18.3    Biobase_2.40.0      BiocGenerics_0.26.0
## [7] bindrcpp_0.2.2      knitr_1.20           magrittr_1.5
## [10] forcats_0.3.0       stringr_1.3.1        dplyr_0.7.5
```

```

## [13] purrr_0.2.5          readr_1.1.1          tidyr_0.8.1
## [16] tibble_1.4.2         ggplot2_3.1.0        tidyverse_1.2.1
##
## loaded via a namespace (and not attached):
## [1] httr_1.3.1           bit64_0.9-7
## [3] jsonlite_1.5         modelr_0.1.2
## [5] assertthat_0.2.0     blob_1.1.1
## [7] GenomeInfoDbData_1.1.0 cellranger_1.1.0
## [9] Rsamtools_1.32.0     yaml_2.1.19
## [11] RSQLite_2.1.1        pillar_1.2.3
## [13] backports_1.1.2      lattice_0.20-35
## [15] glue_1.2.0           digest_0.6.15
## [17] GenomicRanges_1.32.3 XVector_0.20.0
## [19] rvest_0.3.2          colorspace_1.3-2
## [21] htmltools_0.3.6      Matrix_1.2-14
## [23] plyr_1.8.4           psych_1.8.4
## [25] XML_3.98-1.11        pkgconfig_2.0.1
## [27] broom_0.4.4          haven_1.1.1
## [29] zlibbioc_1.26.0      scales_0.5.0
## [31] BiocParallel_1.14.1  withr_2.1.2
## [33] SummarizedExperiment_1.10.1 lazyeval_0.2.1
## [35] cli_1.0.0            mnormt_1.5-5
## [37] crayon_1.3.4         readxl_1.1.0
## [39] memoise_1.1.0        evaluate_0.10.1
## [41] nlme_3.1-137         xml2_1.2.0
## [43] foreign_0.8-70       tools_3.5.0
## [45] hms_0.4.2            matrixStats_0.53.1
## [47] munsell_0.5.0        DelayedArray_0.6.0
## [49] Biostrings_2.48.0    compiler_3.5.0
## [51] GenomeInfoDb_1.16.0  rlang_0.2.1
## [53] grid_3.5.0           RCurl_1.95-4.10
## [55] rstudioapi_0.7       bitops_1.0-6
## [57] rmarkdown_1.10       gtable_0.2.0
## [59] DBI_1.0.0            reshape2_1.4.3
## [61] R6_2.2.2             GenomicAlignments_1.16.0
## [63] lubridate_1.7.4      rtracklayer_1.40.3
## [65] bit_1.1-14           utf8_1.1.4
## [67] bindr_0.1.1          rprojroot_1.3-2
## [69] stringi_1.2.3        Rcpp_0.12.17
## [71] tidyselect_0.2.4

```