

Metadata of the chapter that will be visualized online

Chapter Title	Bacterial Inoculants for Control of Fungal Diseases in <i>Solanum lycopersicum</i> L. (Tomatoes): A Comprehensive Overview	
Copyright Year	2023	
Copyright Holder	The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd.	
Author	Family Name	Shahid
	Particle	
	Given Name	Mohammad
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
Corresponding Author	Family Name	Singh
	Particle	
	Given Name	Udai B.
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
Author	Family Name	Ilyas
	Particle	
	Given Name	Talat
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
Author	Family Name	Malviya
	Particle	
	Given Name	Deepti
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
Author	Family Name	Vishwakarma

	Particle	
	Given Name	Shailesh K.
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
	<hr/>	
Author	Family Name	Shafi
	Particle	
	Given Name	Zaryab
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
	<hr/>	
Author	Family Name	Yadav
	Particle	
	Given Name	Babita
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
	<hr/>	
Author	Family Name	Singh
	Particle	
	Given Name	Harsh V.
	Suffix	
	Division	Plant-Microbe Interaction and Rhizosphere Biology Lab
	Organization	ICAR-National Bureau of Agriculturally Important Microorganisms
	Address	Kushmaur, Maunath Bhanjan, Uttar Pradesh, India
	<hr/>	
Abstract	<p>Globally, <i>Solanum lycopersicum</i> L. (tomatoes) is the second most widely grown vegetable. This crop is sensitive to over 200 diseases caused by a variety of phytopathogenic microorganisms, specifically, soil-borne fungi. The major fungal pathogen causing diseases in tomatoes are <i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>, <i>Botrytis cinerea</i>, <i>Verticillium dahliae</i>, <i>Sclerotium rolfsii</i>, <i>Colletotrichum</i> sp., <i>Alternaria</i> sp. <i>Rhizoctonia solani</i>, etc. Even though a wide range of chemical fungicides is now available to combat fungal diseases, the overuse of these chemicals has been shown to leave negative/adverse influences on the texture, yield and nutritive value of the fruits. In this regard, to manage the fungus-induced tomato diseases, plant</p>	

growth-promoting (PGP) bacteria are one of the most environmentally friendly, effective, safe and economically sound solutions. A variety of beneficial soil microorganisms (BSMs) are currently being employed as soil or plant inoculants in several crop plants, including tomatoes, as biocontrol agents (BCAs). These BCAs also work as growth regulators, in addition to preventing fungal diseases. The current chapter discusses the application of beneficial and antagonistic BCAs, their effectiveness as well as bacterial-mediated mechanisms involved in the management of diseases in tomatoes. The specific mechanisms are antibiosis, competition, production of cellulolytic enzymes, cyanogenic compounds (HCN) and siderophore and induced systemic resistance (ISR). The ability of PGP rhizobacteria to antagonize a pathogen and suppress the disease through multiple pathways has been intensively studied to use them as effective BCAs. As a result, this chapter highlights a full explanation of various bacterial-mediated biocontrol mechanisms used by BCA. As environmental and health issues highlight the need to transition to a more sustainable agriculture system, the use of indigenous PGP rhizobacteria in plant disease prevention is gaining attraction. It's also recommended that using a bacterial consortium guarantees that BCA performs consistently in field settings.

Keywords (separated by ‘-’)	<i>Solanum lycopersicum</i> - Tomatoes - Fungal diseases - Biocontrol agents (BCAs) - Antifungal metabolites
--------------------------------	--

Chapter 15

Bacterial Inoculants for Control of Fungal Diseases in *Solanum lycopersicum* L. (Tomatoes): A Comprehensive Overview

Mohammad Shahid, Udai B. Singh, Talat Ilyas, Deepti Malviya,
Shailesh K. Vishwakarma, Zaryab Shafi, Babita Yadav, and Harsh V. Singh

Abstract Globally, *Solanum lycopersicum* L. (tomatoes) is the second most widely grown vegetable. This crop is sensitive to over 200 diseases caused by a variety of phytopathogenic microorganisms, specifically, soil-borne fungi. The major fungal pathogen causing diseases in tomatoes are *Fusarium oxysporum* f. sp. *lycopersici*, *Botrytis cinerea*, *Verticillium dahliae*, *Sclerotium rolfsii*, *Colletotrichum* sp., *Alternaria* sp. *Rhizoctonia solani*, etc. Even though a wide range of chemical fungicides is now available to combat fungal diseases, the overuse of these chemicals has been shown to leave negative/adverse influences on the texture, yield and nutritive value of the fruits. In this regard, to manage the fungus-induced tomato diseases, plant growth-promoting (PGP) bacteria are one of the most environmentally friendly, effective, safe and economically sound solutions. A variety of beneficial soil microorganisms (BSMs) are currently being employed as soil or plant inoculants in several crop plants, including tomatoes, as biocontrol agents (BCAs). These BCAs also work as growth regulators, in addition to preventing fungal diseases. The current chapter discusses the application of beneficial and antagonistic BCAs, their effectiveness as well as bacterial-mediated mechanisms involved in the management of diseases in tomatoes. The specific mechanisms are antibiosis, competition, production of cellulolytic enzymes, cyanogenic compounds (HCN) and siderophore and induced systemic resistance (ISR). The ability of PGP rhizobacteria to antagonize a pathogen and suppress the disease through multiple pathways has been intensively studied to use them as effective BCAs. As a result, this chapter highlights a full explanation of various bacterial-mediated biocontrol mechanisms used by BCA. As environmental and health issues highlight the need to transition to a more sustainable agriculture system, the use of indigenous PGP rhizobacteria in plant disease prevention is gaining attraction. It's also recommended that using a bacterial consortium guarantees that BCA performs consistently in field settings.

M. Shahid · U. B. Singh (✉) · T. Ilyas · D. Malviya · S. K. Vishwakarma · Z. Shafi · B. Yadav · H. V. Singh

Plant-Microbe Interaction and Rhizosphere Biology Lab, ICAR-National Bureau of Agriculturally Important Microorganisms, Kushmaur, Maunath Bhanjan, Uttar Pradesh, India

35 **Keywords** *Solanum lycopersicum* · Tomatoes · Fungal diseases · Biocontrol agents
36 (BCAs) · Antifungal metabolites

37 15.1 Introduction

38 Tomatoes (*Lycopersicum esculentum* Mill.), commonly known as ‘golden apples’,
39 are the world’s second most widely farmed vegetable crop after potatoes, due to their
40 flavourful fruit and high nutritional content (Costa and Heuvelink 2018). Vitamin A,
41 vitamin C and β -carotene, as well as vital minerals, are abundant in tomatoes (Çolak
42 et al. 2020). It is also high in phosphorus (P), potassium (K), magnesium (Mg) and
43 iron (Fe), all of which are needed to keep nerves and muscles functioning normally
44 (Ali et al. 2020). Tomatoes contain β -carotene (a precursor to vitamin A) and
45 phytosterols, making them the third-best source of vitamin C and the fourth-best
46 source of vitamin A in human diet (Poiroux-Gonord et al. 2010). A strong antiox-
47 idant found in natural lycopene is a phyto-nutrient found in red, ripe and cooked
48 tomatoes that helps prevent heart-related diseases and sarcoma (Kaur and Kaur
49 2015). Other preventive processes, such as anti-inflammatory and antithrombotic
50 functions, are also attributed to this fruit. The short growing season, cheap cost and
51 great economic returns have enticed growers to cultivate the crop throughout the
52 year, even in hotter climates (Villareal 2019). Because of the diverse nature of the
53 fruit, tomatoes are often considered a cash and industrial crop in several countries
54 like India, the USA, China, Turkey and Egypt (Costa and Heuvelink 2018). China is
55 the world’s biggest tomato producer, with yearly production of approximately
56 34 million tonnes, according to international statistics (Nicola et al. 2009). Tomato
57 output in the world is expected to be 145 million tonnes per year, covering 4.36
58 million ha (Chohan et al. 2017).

59 Tomatoes are grown all over the world due to their resilience to a wide range of
60 soils and climates (Morganelli 2007), but their softness makes them vulnerable to
61 insect pests and various abiotic (pesticides, heavy metals, salinity and drought) and
62 biotic stresses (Atkinson et al. 2011). Diseases caused by phytopathogens are the
63 primary limiting factors in overall crop losses around the world, and they are
64 becoming increasingly important as the global population grows. Over 200 patho-
65 gen-caused illnesses have been recorded in tomatoes around the world (Watterson
66 1986). Seed-borne infections, on the other hand, can quickly travel from one location
67 to another and act as the first source of inoculum (Rennie and Cokerell 2006). High-
68 quality seeds, therefore, play an important role in producing long-term, lucrative
69 veggies, and tomatoes are no exception to this. Microorganisms such as fungi,
70 bacteria, viruses and nematodes cause seed-borne illnesses. Among disease-causing
71 parasitic microbes, fungi are the most commonly seen on seeds. As a result, infected
72 seeds have a negative impact on seed health, limiting germination capacity, poor
73 seedling vigour, transmitting fungus pathogen to seedlings, speeding up storage
74 deterioration, transferring pathogens into new areas and expanding the inoculum
75 source in the field. Fungi, a crucial category of microorganisms, are responsible for a
76 number of seed-borne illnesses of tomato, which result in significant yield losses.

Several seed-borne fungi, such as *Aspergillus flavus*, *Rhizopus solani*, *Curvularia* spp. and *Fusarium stolonifer* induce problems in tomato seeds, including wilt, necrosis, rotting and toxification of seeds (Neergard 1997). Late blight, a disease of tomato (caused by *Phytophthora infestans*), is one of the most destructive tomato diseases, resulting in significant financial losses (Fry 2020). *Sclerotinia* rot (caused by *Sclerotinia sclerotiorum*) is another major fungal infection that is threatening tomato crop productivity (Jnr 2000). Several studies have been published on *Fusarium* species-caused wilt, crown and root rot infections in tomatoes (Yezli et al. 2019). The wilt (*Fusarium oxysporum* f. sp. *lycopersici*) and early blight (*Alternaria solani*) diseases are regarded the most damaging fungal diseases that affect tomato plants.

Since the fungal diseases are pandemic, simple cultural cleanliness is ineffective. To protect the fungal diseases, chemical fungicides are often used for better yield and productivity of tomatoes. Even though agrochemicals have long been employed as a reliable method of harvest insurance, their increased use has resulted in several undesirable consequences, including disease resistance and non-target natural effects. Furthermore, the increased use of fungicides to combat tomato diseases has resulted in an increase in health risks due to phytotoxic residues and environmental consequences. As a result, it is strongly recommended that alternate disease management methods be used in place of agrochemical substances.

Managing fungal diseases using biological means is seen to be a viable method for reducing disease severity. In comparison to the previously mentioned use of chemical fungicides, the use of soil beneficial bacteria that colonize the underlying foundations of harvest plants and suppress the soil-borne illnesses is becoming an elective alternative. In this regard, PGP bacteria can be used as soil inoculants to reduce soil-borne illnesses, which is an organic solution. Because they have the potential to colonize the rhizosphere swiftly and spread down the root from a single seed treatment or soak application into the soil, PGPR with the viability of their biocontrol activity typically provide long-term protection from soil-borne diseases at the root surface. A huge number of PGPR have been recovered and characterized in the hopes of developing them as biocontrol agents for tomato illnesses. For example, Kilani-Feki et al. (2016) also employed *B. subtilis* strains to suppress the *Botrytis cinerea*, the pathogen that causes tomato fruit rot. In another study, biocontrol agents (BCAs) were recovered to combat the tomato wilt disease. It was observed that *Ochrobactrum intermedium* and *B. amyloliquefaciens*, among strains, potentially inhibited the disease incidence and increased the seedling growth and vigour indices of tomatoes (Gowtham et al. 2016). Furthermore, the effectiveness of *P. fluorescens* strains against a variety of fungal diseases (leaf blight, damping-off, stem canker, and root rot) in tomatoes (Singh et al. 2017) has been reported. Under in vitro and pothouse settings, BCAs, viz. *P. aeruginosa*, *P. fluorescens*, *B. amyloliquefaciens* and *B. subtilis* were able to successfully suppress the canker and wilt disease of tomato (Abo-Elyousr et al. 2019). Recently, a new technique for inducing systemic resistance (ISR) in plants by employing PGPR has been investigated by Attia et al. (2020). They confirmed that PGPRs were found to be effective in reducing the growth of *A. solani* (causing early blight disease) in tomato plants. In comparison to

122 non-treated plants, they achieved an 84.3% protection rate. Effective selection,
123 screening and safety investigation of promising PGPR strains are required for
124 incorporating soil microbes for disease control/plant growth promotion in cropping
125 systems and eliminating the need for chemicals. Furthermore, information on disease
126 targeting, mass-scale production costs and procedures of registration must all be
127 updated to improve the market standing of these BCAs. The current chapter aims to
128 look into the potentiality of antagonistic PGP bacteria (BCAs) in managing the
129 major fungal diseases of tomatoes. This chapter also discusses the challenges and
130 benefits of commercializing these bacteria in the agriculture sector.

131 **15.2 Prevalence of Seed-Borne Mycoflora: A Historical** 132 **Perspective**

133 Seed-borne fungi are microorganisms that can host the seeds both within and
134 externally, causing diseases or contaminating the environment (Amza 2018). Phy-
135 topathogenic fungi may cause post-germination death, rendering them poisonous
136 and lowering their quality for human food and seed production. The conidia,
137 oospores, sclerotia, hyphae and chlamydospores are some examples of the main
138 forms in which they are present (Arora 1986). The tomato, which is employed as a
139 model crop in genetics, is susceptible to a variety of seed-borne fungal diseases.

140 A total of 12 phytopathogenic fungi were recovered from the fruits and seeds of
141 *Solanum lycopersicum*. It was found that the majority of the pathogens were
142 habitants of fruits; however, the species of *Cladosporium* were detected in seeds
143 (Dhekle and Bodke 2013). In reality, a considerable number of fungal isolates
144 belonging to genera *Fusarium*, *Pythium*, *Botrytis*, *Alternaria* and *Rhizoctonia* were
145 found in tomato seeds, causing several seeds-borne diseases. Mycoflora are isolated
146 as surface contaminants, internally seed-borne flora, and are known to cause major
147 field diseases depending on the presence of fungi on the seed coat or in the seed.
148 Various fungal phytopathogens, viz. *F. oxysporum*, *Alternaria solani*, *Aspergillus*
149 *flavus*, *A. fumigatus*, etc., have been detected and identified from infected tomato
150 plants and reported to cause serious seed damage (Fig. 15.1) as reported by several
151 workers. The investigation of grey mould disease in tomato leaves was done using a
152 hyperspectral imaging technique based on competitive adaptive reweighted sam-
153 pling (CARS) and correlation analysis (WANG et al. 2017). Similarly, tomato plants
154 were infected by fungal pathogens like *F. semitectum* (1–3% infection),
155 *F. moniliforme* (0.5% infection), and *Curvularia lunata* (0.5–7.5% infection) and
156 *Bipolaris* spp. (1.5% infection) (Bhatti et al. 2010). The duration of storage has a
157 significant impact on the prevalence of various mycoflora. On a modest scale,
158 *Phytophthora infestans* has caused the full destruction of tomato harvests around
159 the world (Panthee and Chen 2010). *Hormonema* spp., one of the most prevalent
160 genera on tomato seeds, were discovered by Nishikawa et al. (2006) while assessing
161 109 species of seed-borne fungi from three cultivars of tomato. The seed could be

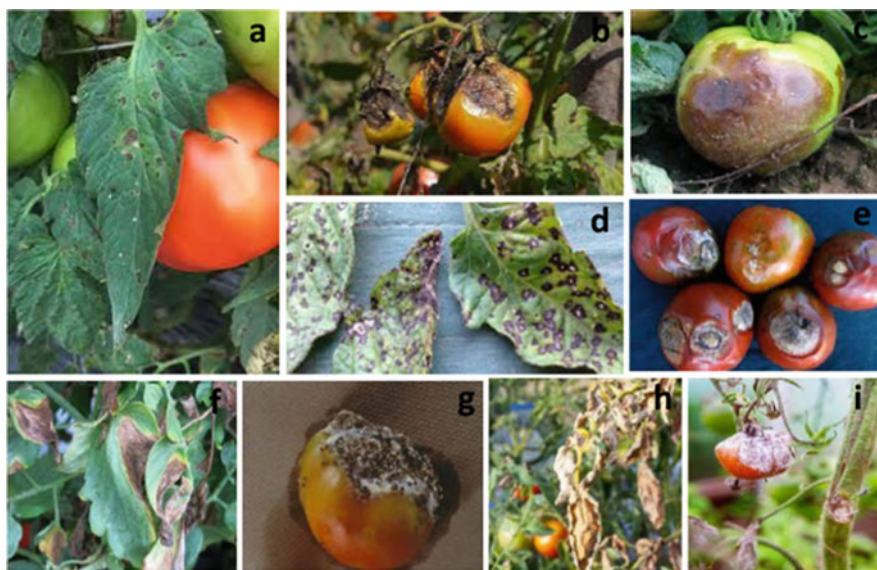


Fig. 15.1 Major tomato diseases caused by fungal pathogens. Early blight (*Alternaria solani*) (a), late blight (*Phytophthora infestans*) (b), buckeye rot (*Phytophthora*) (c), septoria leaf spot (*Septoria lycopersici*) (d), anthracnose (*Colletotrichum*) (e), verticillium wilt (*Verticillium dahliae*) (f), southern blight (*Sclerotium rolfsii*) (g), fusarium wilt (*Fusarium oxysporum* f. sp. *lycopersici*) (h) and grey mould (*Botrytis cinerea*) (i) (source: <https://www.google.com/>)

diseased within or be contaminated on the exterior. The fungus *F. oxysporum* f. sp. *lycopersici* causes fusarium wilts, one of the most destructive diseases of tomatoes. Similarly, previous researchers found a total of 17 fungal phytopathogens linked with tomato var. local seeds, with *Aspergillus niger*, *A. flavus*, *Fusarium moniliform*, *Rhizopus nigricans*, *Curvularia lunata* and *Alternaria alternata* being the most prevalent. In the Indian state of Gujarat, 12 fungi were discovered with four tomato cultivar seeds. *A. alternata*, *A. flavus* and *A. niger* were the most common, whereas *A. amstelodami* and *Cunninghamella echinulata* were discoveries.

15.3 Management of Fungal Diseases in Tomatoes

In the fight against crop diseases, seed health is crucial. The rise of endemic diseases as a result of changing global environment poses a difficulty in maintaining the health of the plant. As a result, timely and precise diagnosis of the problem, as well as pathogen surveillance, gives time for mitigation actions to be implemented. Treatment of seeds to destroy pathogens carried within or on the seed has been demonstrated to be effective in preventing epiphytotic plant diseases. Seed-borne fungi, on the other hand, are easy to control compared to airborne or soil-borne

178 fungi. Farmers are encountering financial difficulties as a result of significant crop
179 losses caused by seed-borne mycoflora on their crops. Controlling seed-borne
180 infections via various approaches is a crucial element in every agricultural crop
181 production and protection programme. To eliminate pathogens from seeds both
182 inside and externally, as well as to protect seeds from soil-borne diseases, a variety
183 of chemical, biological, physical, and mechanical techniques have been applied
184 (Shahid et al. 2017).

185 **15.3.1 Use of Chemical Fungicides (Chemical Method)**

186 Mycotoxins released by seed-borne fungal pathogens cause serious issues in
187 humans. Hence, seeds should be treated with appropriate chemical before planting.
188 Chemical fungicides are used since pre-historic times for the management of phy-
189 topathogenic fungi and for obtaining a better yield of crops. Historically, treating
190 seeds with chemical fungicide using spraying, drenching of soils and dusting has
191 been important (Copping and Duke 2007). These methods could protect the seeds
192 and seedlings from the soil-dwelling fungal pathogens causing damping-off and rot
193 diseases (Divya Rani and Sudini 2013). Amini and Sidovich (2010) conducted
194 in vitro and in vivo experiments where they used six chemical fungicides, viz.
195 fludioxonil (FLN), bromuconazole (BMZL), benomyl (BML), carbendazim
196 (CBZM), azoxystrobin (AZBN) and prochloraz (PCZ), against the fusarium wilt
197 disease in tomato crop. When administered to seedlings at prescribed levels, PCZ
198 and BMZL were found to be the most effective against *Fusarium oxysporum* f. sp.
199 *lycopersici*. Among the fungicides tested, FLN and BMZL showed a phytotoxic
200 impact on tomato seedlings. Similarly, in another investigation, Al-Kassim and
201 Monawar (2000) treated (in vitro) five vegetable seeds including *Solanum*
202 *lycopersicum*, *Solanum melongena*, *Abelmoschus esculentus*, etc., in Gazan prov-
203 ince with 0.2% of chemical fungicides like benomyl, cozib and mancozeb before
204 incubation. The majority of the isolated fungi were inhibited by all of the tested
205 fungicides. Benomyl, on the other hand, was the most effective against all of the
206 fungi found on the seeds of the tomato.

Major fungal diseases of tomatoes, their symptoms and chemical control measures

207	S. no.	Disease	Causal agent	Symptoms	Chemicals used	Effectiveness	Reference
208	1	Early blight	<i>Alternaria solani</i>	<ul style="list-style-type: none">• Brown-black spots (in the form of lesion 1/ 2-inch diameter)• Formation of blotches (irregular)• Defoliation, browning of infected leaves• Appearance of lesion (dark) on stems• Girdling of the stem or collar rot	Mancozeb 80WP, Bavistin 50WP, Indofil M-45, Sulcox 50WP and Tall-25EC, chlorothalonil (0.2%), kasugamycin (0.2%), azoxystrobin (0.1%), propiconazole (0.1%), pyraclostrobin (0.2%), perfekt (0.2%), metalaxyl (0.2%), mancozeb (0.25%)	Suppressed the fungal growth Reduced the disease incidence	Roy et al. (2019), Arunakumara et al. (2010)

(continued)

S. no.	Disease	Causal agent	Symptoms	Chemicals used	Effectiveness	Reference	
2	Septoria leaf spot	<i>Septoria lycopersici</i>	<ul style="list-style-type: none"> • Disease infection on lower leaf • Symptoms also occur on stems and blossom • Symptoms on fruits appear in the form of small-sized water-soaked spots • Development of greyish white centres with dark edges • Yellowing, withering of leaves and ultimately fall off 	Fungicides pyraclostrobin (116.6 ppm), fluxapyroxad (58.5 ppm), mancozeb (4000 ppm), difenoconazole (125 ppm), chlorothalonil (1500 ppm), propineb (2100 ppm), fluazinam + thiophanate-methyl (375 + 375 ppm) and metiram + pyraclostrobin (1100 + 100 ppm)	Suppressed the fungal growth Reduced the disease incidence	Monteiro et al. (2021)	209 210
3	Late blight	<i>Phytophthora infestans</i>	<ul style="list-style-type: none"> • Younger/older leaves infected • Appearance of pale-green water-soaked spots starting at leaf tips that enlarge rapidly, forming irregular, greenish-black blotches • Development of white moulds at the margins of infected areas • Under the favourable condition, whole plants rapidly defoliated • Infection on petioles and stems occurs as brown streaks 	Fungicides oxathiapiprolin, chlorothalonil, azoxystrobin, mandipropamid and mfenoxam were effective against the disease	Chemicals single or in mixture controlled the disease Suppressed the growth of pathogens Improved the growth and yield of tomato	Cohen et al. (2018)	211
4	Fusarium wilt	<i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>	<ul style="list-style-type: none"> • Development of leaves • Affected leaves soon wilt and dry up, but they remain attached to the plant • The wilting continues successively on younger foliage and eventually results in plant death • The stem remains firm and green on the outside but exhibits a narrow band of brown discolouration in the vascular tissue • Brown streaking in the vascular tissue of infected plants becomes plugged during the attack by the fungus, leading to wilting and yellowing of the leaves 	Chemical fungicides difenoconazole (200 mg/L), benomyl, carbendazim, prochloraz, fludioxonil, bromuconazole and azoxystrobin were effective		Amini and Sidovich (2010)	212
5	Verticillium wilt	<i>Verticillium dahliae</i>	<ul style="list-style-type: none"> • Leaf edges and areas between the veins turn yellow and then brown • Infected plants often have a characteristic V-shaped lesion at the edge of the leaf 	Nano-fungicide (leaf extract olive oil-loaded chitosan nanoparticles)	Diminished the disease symptoms	Mazzotta et al. (2022)	213

(continued)

214	S. no.	Disease	Causal agent	Symptoms	Chemicals used	Effectiveness	Reference
				occurring in a fan pattern • As the disease progresses, younger leaves begin to wilt and die, until only a few healthy leaves remain at the top of the plant			
214	6	Anthrachnose	<i>Colletotrichum</i>	• Symptoms appear first as small, circular, slightly sunken lesions on surface of ripening fruits • Spots quickly enlarge, become bruise like depressions and develop a water-soaked appearance directly beneath the skin (epidermis) of the fruit • As these spots expand, they develop dark centres or concentric rings of dark specks the rings consist of numerous small spore-producing bodies	Azoxystrobin-based fungicides Onestar 23% SC and Amistar 23% SC	Decreased the incidence of disease (69–71%)	Saxena et al. (2016)
215	7	Buckeye rot	<i>Phytophthora</i>	• Symptoms appear as water-soaked greyish-green/brown spots • Further, spot enlarges and develops into a lesion with a target-like pattern of concentric rings of narrow dark-brown and wide light-brown bands	Application of fungicides metalaxyl, cymoxanil, mancozeb and copper oxychloride	Reduced the incidence of disease	Gupta and Bharat (2008)
216	8	Southern blight	<i>Sclerotium rolfsii</i>	• Formation of wilting on plants • Development of water-soaked lesion on stems • Produced sclerotia are white, later becomes dark-brown (spherically structured) • White mycelium and sclerotia at stem base of infected plants are the main symptoms of disease	Fungicides pyraclostrobin, quintozene and fluxapyroxad were used	Chemicals significantly reduced the disease incidence	Keinath and DuBose (2017)
217	9	Grey mould	<i>Botrytis cinerea</i>	• Appearance of grey-brown velvety mould covering on stems/younger leaves • Grey spores cover dying flowers and the calyx of fruit	Difen Super, 55% WP fungicide concentration of 0.08% with active ingredient, difenoconazole, and fungicide Skor, 250 g/l EC with concentration of 0.05% and 0.07%	Effective against the pathogen	Zuparov et al. (2020)

15.3.2 Biological Management

218

Chemical fungicides have a significant impact on seed-borne fungal infections, but they also have a negative impact on beneficial microbial diversity in soils as well as crop productivity. Furthermore, irregular fungicidal usage is not only harmful to animals and humans but also leads to development of resistance among the pathogens targeted. Because fungicides are harmful to non-target organisms (Marinho et al. 2020), scientists are turning to more environmentally benign and cost-effective means of disease management, such as acid treatments, and the use of antagonistic microbes (Droby et al. 2022), and plant extracts (Shuping and Eloff 2017). These are interesting and viable options for releasing plant growth regulators that influence overall crop development and improve morpho-physiological features. The method of biocontrol has been utilized for over 2000 years, and it has been widely used in managing pests since the end of the eighteenth century. The types of biological control can be classified into natural, conservation, inoculative (classical) and augmentative biocontrol. Natural biocontrol has been used to reduce pests since evolution, whereas conservation biocontrol comprises human measures to stimulate and protect the performance of natural foes. The inoculative mode of biocontrol is the most extensively used type of biocontrol, in which natural enemies are dispersed into new places where the pest has been mistakenly introduced. Natural enemies are mass-produced in bio-factories and sent into the market for fast pest control, making augmented biocontrol more appealing. This type of plant protection is regarded to be both environmentally beneficial and food-safe. Moreover, gaining a better knowledge of biocontrol mechanisms via interactions between BCAs and phytopathogens may aid in the improvement and development of biocontrol systems.

15.4 Plant Growth-Promoting Rhizobacteria: Tapping for BCA

242

243

Rhizobacteria are a varied collection of bacteria that colonize the rhizosphere environment. In the root zone, they are strong microbial competitors. They either directly or indirectly influence plant development. The direct method entails PGPR's positive effects that directly boost plant growth. These systems help plants grow, but the ways they do so differ from species to species and strain to strain (Noori and Saud 2012). Through biogeochemical cycling, the PGPR nourishes the plant by converting the nutrients into the soil. They also help plant growth by facilitating the transfer of these nutrients into the plant. They promote plant growth by synthesizing plant hormones such as indole acetic acid (Ahmed et al. 2021), cytokinins (Liu et al. 2013), fixing nitrogen from the atmosphere (Malik et al. 1997), solubilizing minerals like phosphorus (Gomez-Ramirez and Uribe-Velez 2021) and generating siderophores that can solubilize and sequester iron (Sultana et al. 2021) and provide nutrients to plants (Etesami and Adl 2020) in addition to combating soil-borne plant diseases (Hamid et al. 2021). It plays a significant effect in repelling

phytopathogenic bacteria in addition to promoting plant growth (Hassan et al. 2019). *Bacillus subtilis* has a biocontrol efficiency of more than 50% on tomato plants against the plant disease caused by *Ralstonia solanacearum* in greenhouse conditions (Chen et al. 2013). *Rhizoctonia solani*, which causes damping-off in tomatoes, was controlled by a strong biocontrol agent, *Priestia endophytica* FH5 (Zhou et al. 2021). The biofortified vermicompost prepared from chosen BCAs (like *B. subtilis* and *P. fluorescens*) could be used to successfully manage the wilt diseases in tomatoes caused by *Fusarium oxysporum* (Basco et al. 2017). In a similar study, *S. pratensis* strain LMM15 (having the potential of BCA) was sprayed on tomato leaves 1 day before fungal inoculation. After BCA spraying, it was observed that the occurrence of grey mould disease was reduced by approximately 46%. Furthermore, the applied biocontrol agent significantly increased the plant stressor metabolites (proline and lipid peroxidation) as well as defence enzymes in shoot tissues (Lian et al. 2017). These strains of actinomycetes drastically reduced the pathogen proliferation while also improving tomato growth characteristics (Goudjal et al. 2014).

15.5 Mechanisms Involved in Disease Suppression (Indirect Mechanisms)

The PGPR's antagonistic characteristics against a variety of diseases expand their potential as biocontrol agents (Fatima et al. 2022). Several genera of beneficial soil bacterial strains including *Pseudomonas* (Kabdwal et al. 2019), *Bacillus* (Ni and Punja 2019), *Enterobacter* (Xue et al. 2009), *Serratia* (Youssef et al. 2016), *Klebsiella* (Gaur et al. 2017), *Azotobacter* (Alsudani 2022), etc. are familiar as biological control agents (BCAs) used in reducing tomato diseases. Many reports claim that using the PGP consortium as biological control agents has some advantages over other disease control methods, such as being an environmentally safe and non-toxic indigenous microorganism whose application is both environmentally safe and favourable to human health. Induction of systemic resistance (ISR), antibiotic production, competition, secretion of cellulolytic/hydrolytic enzyme and production of HCN and siderophores are all essential mechanisms involved in BCA's antagonistic effects (Narayanasamy 2013). Furthermore, they can help plants cope with numerous stressors such as salinity, drought, hunger, heavy metal toxicity and so on, allowing them to thrive in such environments. Even though various free-living PGPR are regarded as plant development beneficial microbes, not all the strains within the same species have the same metabolic capacity to boost plant growth. It's crucial to understand the rhizosphere microbiota's capabilities, as well as its mechanisms of action, to ensure long-term crop production (Babalola et al. 2021). The PGPR's antagonistic characteristics against a variety of diseases expand their potential as BCAs (Verma et al. 2019). Various genera of *Bacillus* including *Bacillus megaterium*, *B. subtilis* and *B. polymyxa*, *P. fluorescens* and *T. harzianum* were co-inoculated with *Azospirillum* sp. and *Azotobacter* sp. and were reported to effectively control the disease caused by fungal pathogens (Saad et al. 2016).

Some biocontrol agents (BCAs) involved in the management of major fungal diseases of tomatoes

S. no.	Major fungal disease	Causal agent	Biocontrol agent involved	Mechanism involved	Effectiveness	Reference	
1	Fusarium wilt disease	<i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>	<i>Bacillus</i> sp.	Production of cell wall-degrading enzymes like β -1,3-glucanase, protease, chitinase, ammonia, siderophore, HCN production, bioactive volatile and non-volatile metabolites	<ul style="list-style-type: none"> • Reduced the disease severity up to 55%. Enhanced the growth and physiological traits of plants 	Jangir et al. (2018)	299 300
2	Tomato vascular wilt	<i>Fusarium oxysporum</i>	<i>Bacillus velezensis</i> NKG-2	Volatile organic compounds (VOCs)	<ul style="list-style-type: none"> • Suppressed the disease severity • Increased the growth of plants 	Myo et al. (2019)	301
3	<i>Botrytis cinerea</i>	Grey mould disease	<i>Bacillus subtilis</i> L1-21	Antifungal metabolites	<ul style="list-style-type: none"> • Completely inhibited (100%) the fungal growth • Increased the plant growth and improved the fruit quality 	Bu et al. (2021)	302
4	Alternaria rot disease	<i>Alternaria alternata</i>	<i>Bacillus atrophaeus</i>	Antifungal metabolites O-anisaldehyde And lipopeptides	<ul style="list-style-type: none"> • Reduced the germination of the spore. • Decreased the disease severity of <i>Alternaria</i> in tomato fruit. 	Chacon-Lopez et al. (2021)	303
5	Black scurf disease	<i>Rhizoctonia solani</i>	<i>Bacillus subtilis</i> Hussain T-AMU	Lipopeptide, biosurfactant	<ul style="list-style-type: none"> • Inhibited the growth of the pathogen • Inhibited the incidence and severity of disease 	Hussain et al. (2021)	304
6	Alternaria leaf blight	<i>Alternaria solani</i>	<i>Bacillus velezensis</i> NKMV-3	Production of lipopeptide (antibiotic synthesis genes), <i>iturin C</i> , <i>surfactin A</i> and <i>fengycin B</i> and <i>D</i>	<ul style="list-style-type: none"> • Controlled the growth of the pathogen 	Vignesh et al. (2022)	305
7	Fusarium wilt disease	<i>Fusarium oxysporum</i> sp. <i>lycopersici</i>	<i>Bacillus inaquosorum</i> KR2-7	Production of BGCs fengycin, surfactin and bacillomycin F, bacillaene, macrolactin, skf, subtilisin A, bacilysin and bacillibactin	<ul style="list-style-type: none"> • Reduced the disease severity • Increased defence-related enzyme activities 	Kamali et al. (2022)	306
8	Early blight disease	<i>Alternaria alternata</i>	<i>Bacillus</i> sp.	Antibiosis, production of volatile organic compounds	<ul style="list-style-type: none"> • Decreased the disease severity 	Pane and Zaccardelli (2015)	307

(continued)

308	S. no.	Major fungal disease	Causal agent	Biocontrol agent involved	Mechanism involved	Effectiveness	Reference
		Wilt disease	<i>Verticillium dahliae</i>	<i>Bacillus subtilis</i>	Various antibiotic metabolites and VOCs	<ul style="list-style-type: none"> • Suppressed the fungal infection, increased tomato yield 	Rahman et al. (2021)
309	9	Foliar blight disease	<i>Botrytis cinerea</i>	<i>B. nakamurai</i> , <i>B. pseudomycoides</i> , <i>B. proteolyticus</i> , <i>B. thuringiensis</i> , <i>E. asburiae</i> and <i>E. cloacae</i>	Antifungal VOCs such as 3-methylbutan-1-ol, sulphur-containing compounds, 2-heptanone and dodecanal	<ul style="list-style-type: none"> • Increased the growth and yield of tomato 	Chaouachi et al. (2021)
310	10	Anthrachnose disease	<i>Colletotrichum capsici</i>	<i>Bacillus</i> sp. strain M10	Extracellular enzyme production	<ul style="list-style-type: none"> • Suppressed the fungal growth 	Srikhong et al. (2018)
311	11	Fusarium wilt	<i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>	<i>Bacillus pumilus</i>	Loss of protoplasm in fungal cell wall	<ul style="list-style-type: none"> • Reduced (73%) disease incidence • Increased root (60%) and shoot (84%) of tomato 	Heidarzadeh and Baghaee-Ravari (2015)
312	12	Early blight and anthracnose disease	<i>Alternaria solani</i> , <i>Colletotrichum coccodes</i>	<i>Pseudomonas fluorescens</i> A506	Antifungal antibiotics and VOCs	<ul style="list-style-type: none"> • Suppressed anthracnose on fruit and early blight on detached leaves, controlled the disease in tomato 	Cuppels et al. (2013)
313	13	Early blight disease	<i>Alternaria solani</i>	<i>Pseudomonas fluorescens</i>	Antifungal antibiotics and VOCs	<ul style="list-style-type: none"> • Inhibited (45.55%) the fungal growth 	Koley et al. (2015)
314	14	Grey mould disease	<i>Botrytis cinerea</i>	<i>Bacillus licheniformis</i>	Antifungal compounds	<ul style="list-style-type: none"> • Inhibited the fungal growth • Controlled the disease incidence (90%) in tomato 	Lee et al. (2006)
315	15	Foot rot disease	<i>Fusarium oxysporum</i> f. sp. <i>radicis-lycopersici</i>	<i>Collimonas fungivorans</i>	Antifungal compounds, cell wall-degrading enzymes	<ul style="list-style-type: none"> • Suppressed the fungal growth 	Kamilova et al. (2007)
316	16	Black scurf and wilt disease	<i>Rhizoctonia solani</i> , <i>Fusarium oxysporum</i>	<i>Bacillus subtilis</i> (TD11), <i>Bacillus cereus</i> (TD15)	Secretion of antifungal metabolites Induced systemic resistance (ISR)	<ul style="list-style-type: none"> • Inhibited the growth of <i>Rhizoctonia solani</i> (40%) and <i>Fusarium oxysporum</i> (80%) • Controlled (50%) the disease incidence in tomato 	Malik et al. (2022)
317	17	Root rot disease	<i>Sclerotium rolfsii</i>	<i>Bacillus</i> sp.	Production of hydrogen cyanide (HCN) and extracellular enzymes	<ul style="list-style-type: none"> • Suppressed the fungal growth • Reduced the disease incidence 	Kumar et al. (2012)

(continued)

S. no.	Major fungal disease	Causal agent	Biocontrol agent involved	Mechanism involved	Effectiveness	Reference	
18	Fusarium wilt disease	<i>Fusarium oxysporum</i>	<i>Pseudomonas</i> sp.	Siderophore production	<ul style="list-style-type: none"> • Suppressed the fungal growth • Reduced the disease incidence 	Arya et al. (2018)	318
19	Fusarium wilt disease	<i>Fusarium oxysporum</i>	<i>Streptomyces</i> SNL2	Synthesis of phenolate siderophore	<ul style="list-style-type: none"> • Reduced the prevalence of wilt disease by 88.5% 	Goudjal et al. (2016)	319
20	Early blight of tomato	<i>Alternaria solani</i>	<i>Lysinibacillus fusiformis</i> L-2, <i>Bacillus subtilis</i> B-1 and <i>Achromobacter xylosoxidans</i> A-3	Production of siderophore and other antifungal compounds	<ul style="list-style-type: none"> • Suppressed the fungal growth • Controlled the disease • Improved the growth features of tomato crop 	Attia et al. (2020)	320
21	Fusarium wilt disease	<i>Fusarium oxysporum</i>	<i>B. pumilus</i>	Siderophore production	<ul style="list-style-type: none"> • Suppressed the fungal growth. Reduced the disease incidence 	Heidarzadeh and Baghaee-Ravari (2015)	321
22	Fusarium wilt	<i>F. oxysporum</i> f. sp. <i>lycopersici</i>	<i>Burkholderia contaminans</i> AY001	Systemic induced resistance (ISR), production of antimicrobial compounds, including di (2-ethylhexyl) phthalate and pyrrolo [1,2-a] pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)	<ul style="list-style-type: none"> • Enhanced the growth of tomato plants • Increase the disease resistance • Improved the yield attributes 	Heo et al. (2022)	322
23	Fusarium wilt	<i>Fusarium oxysporum</i>	<i>Pseudomonas aeruginosa</i>	Production of antifungal metabolites like siderophore, HCN and ammonia	<ul style="list-style-type: none"> • Inhibited the fungal growth (75%) • Increased the growth parameters 	Parasuraman et al. (2022)	323
24	Early blight	<i>Alternaria solani</i>	<i>Pseudomonas aeruginosa</i>	Production of antifungal metabolites like siderophore, HCN and ammonia	<ul style="list-style-type: none"> • Inhibited the fungal growth (75%) • Increased the growth parameters 	Parasuraman et al. (2022)	324
25	Early blight disease	<i>Alternaria alternata</i>	<i>B. atrophaeus</i> and <i>Brevibacterium frigoritolerans</i>	Induced systemic resistance (ISR) Secretion of extracellular enzymes	<ul style="list-style-type: none"> • Suppressed the fungal growth • Reduced the disease incidence • Increased the plant growth and biomass 	Chacon-Lopez et al. (2021)	325
26	Verticillium wilt disease	<i>Verticillium dahliae</i>	<i>Pseudomonas stutzeri</i>	Synthesize anti-fungal metabolites like HCN, siderophore and other extracellular enzymes	<ul style="list-style-type: none"> • Enhanced the growth characteristics (stem length, number of leaflets, leaf area and root 	Essalimi et al. (2022)	326

(continued)

S. no.	Major fungal disease	Causal agent	Biocontrol agent involved	Mechanism involved	Effectiveness	Reference
					weight) and biochemical parameters (nitrate reductase activity, proline and chlorophyll content) of tomato.	
27	Fusarium wilt disease	<i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i>	<i>Pseudomonas fluorescens</i> Pf1	Secretion of essential plant growth-regulating substances, synthesize antifungal metabolites	<ul style="list-style-type: none"> • Decreased the disease incidence • Improved the germination, vigour indices, growth and biomass of tomato 	Johnson et al. (2022)
28	Collar rot disease	<i>Sclerotium rolfsii</i>	<i>Pseudomonas fluorescens</i> Pf1	Synthesize antifungal compounds	<ul style="list-style-type: none"> • Suppressed the growth of pathogen • Reduced the disease incidence 	Johnson et al. (2022)

15.5.1 Production of Antibiotics

The chemical compounds which are produced by a species of microbes and are used to kill other species of microbes are known as antibiotics. To survive predation, competition and other threats, microorganisms interact with each other and produce these chemical molecules (Kaya and Koppenhöfer 1996). Bacterial antagonists decrease phytopathogens by secreting inhibitory compounds into the extracellular environment (Freitas et al. 2022). It is a highly effective and extensively researched aspect of biocontrol. The 2,4-diacetylphloroglucinol (DAPG), pyrrolnitrin, phenazine, tensin, pyoluteorin, tropolone, oomycin A, cyclic lipopeptides and hydrogen cyanide (HCN) are some of the well-known antibiotics for biological control (Saeed et al. 2021). The majority of antibiotics produced by *Bacillus* sp. are effective against plant harmful fungi *Fusarium oxysporum* and *Alternaria solani* (Zhang et al. 2020). *B. subtilis* has been reported as a strong biocontrol agent, and it inhibited the fungal growth by producing a wide range of antibiotics, including bacilysin, bacillomycin, fengycin, zwittermicin and difficidin (Kim et al. 2010; Telang 2010). *Bacillus* strain TNAM5 was discovered to be effective in suppressing FOL by producing diffusible and volatile organic (VO) antifungal compounds, ammonia and HCN (Prashar et al. 2013; Kumari et al. 2021). Phichai (2014) discovered that *B. subtilis* could generate antifungal metabolites (bacitracin, subtilin, bacillin and bacillomycin) that inhibited *Alternaria* spp. However, because antibiotic resistance is a problem, having too much reliance on antibiotic-producing bacteria as a BCA could be a disadvantage.

15.5.2 Competition

351

Effective colonization and increased competition are important factors in biocontrol; thus, the biological control agents (BCAs) should be able to tolerate and multiply in a natural environment (Pal and Gardener 2006). Microorganisms must efficiently compete for accessibility of nutrition and niche to establish themselves in the rhizosphere (Lugtenberg 2015). Biocontrol mainly relies heavily on competition between pathogenic and non-pathogenic microorganisms (Daguerre et al. 2014). Plant-associated bacteria are known to provide more protection to the plant by accelerating rhizosphere penetration than pathogens. Biocontrol chemicals exhaust the few available substrates, rendering them inaccessible to infections. At the same time, they produce metabolic chemicals that are harmful to infections (TariqJaveed et al. 2021). As a result, an effective BCA must be able to safeguard the deeper root sections by colonizing well and suppressing the pathogen at the growing tips. In rhizobacterial populations, species of *Pseudomonads* have been identified as very efficient competitors for root exudates (Kuiper et al. 2002). Antimicrobial chemicals are found in some root exudates, providing a favourable ecological niche for PGPR, which may then detoxify them (Badri and Vivanco 2009). This means that PGPR competence is heavily contingent on their capacity to exploit specific environmental conditions or adapt to changing conditions (Amaya-Gómez et al. 2020). Kuiper et al. (2001) found that root colonizing *Pseudomonads*' competitive colonization ability was strongly influenced by their enhanced intake of putrescine, a tomato root exudate.

15.5.3 Induced Systemic Resistance (ISR)

373

Beneficial bacteria living in the rhizosphere region interact with the host plant to help it fight against different phytopathogens (Kumar and Jagadeesh 2016). The enhanced physiological state of defence elicited by broad-spectrum biotic and abiotic stimuli is known as induced resistance. The induction of systemic resistance is characterized by (1) induced systemic resistance (ISR) and (2) systemic acquired resistance (SAR). ISR occurs when plants' intrinsic defence mechanisms are triggered in response to biotic threats (Pineda et al. 2013). Plants in SAR develop greater resistance to uninfected plant sections while dealing with a wide range of diseases (Pieterse et al. 2001). ISR is primarily mediated by the jasmonate or ethylene-sensitive pathway, whereas SAR is mediated by salicylic acid (SA) (Mandal and Ray 2011). Rhizobacteria invading plant roots can induce resistance to a wide range of illnesses. It has been reported that some strains of *Bacillus* (Choudhary and Johri 2009), *Pseudomonas* (Jisha et al. 2019), *Burkholderia* (Ahmad et al. 2022) and *Serratia* (Singh and Jha 2016) cause ISR in response to various infections. By evoking such induced resistance, BCAs display the inhibition of diseases caused by fungal, bacterial, viral and, in some cases, insects and nematodes. The genus

Bacillus is an excellent BCA and the most widely used biopesticide for controlling plant diseases (Miljaković et al. 2020). Some species of *Pseudomonas* also play an imperative role in ISR. For example, *P. fluorescens* was found to provide ISR to tomato plants against the diseases causing fungal pathogens (such as *P. infestans* and *F. oxysporum*) (Santoyo et al. 2012). The use of potential *Pseudomonas fluorescens* VSMKU3054, an efficient BCA, gave clear evidence of ISR-mediated biocontrol of tomato grey mould disease (Suresh et al. 2022). In a study, Chunyu et al. (2017) evaluated the efficiency of PGPR strain *B. amyloliquefaciens* SQRT3 to prevent the tomato crop against pathogens by inducing a systemic resistance mechanism. In a crop-based study, it was observed that by activating the various defence enzymes (PPO, POX, GLU, CHI and PAL), and induction of systemic resistance, antagonistic PGPR strains *S. marcescens*, *Streptomyces cereus* and *Bacillus cereus* increased the resistance of wilt disease in tomato (Ferraz et al. 2015). Reduction in the severity of disease and enhanced upregulation of defensive enzymes triggered by the combined inoculation of *Bacillus atrophaeus*, *B. subtilis*, and *Burkholderia cepacia* exhibited a direct biocontrol and ISR mode of action for suppression of vascular disease in tomato crops (Shanmugam and Kanoujia 2011). Similarly, by activating and upsurging the activities of peroxidase, PPO and phenylalanine ammonia-lyase, the biocontrol agent *Pseudomonas putida* stimulated the systemic responses in tomatoes against early blight disease (Ahmed et al. 2011). In another study, antagonistic bacteria *B. subtilis* OTPB1 suppressed the early blight of tomato (caused by *A. solani*) due to increased systemic response. The bacterial-inoculated tomato seedlings had significantly higher levels of defence-related enzymes (peroxidase, PO; polyphenol oxidase, PPO; and superoxide dismutase, SOD) than uninoculated control seedlings (Chowdappa et al. 2013). In the external environment, bacteria produced exopolysaccharides (EPS). The EPS aids in drought resistance, stress resistance and phytopathogen defence (Fig. 15.2). In various crops, the significant function of bacterial EPS as an elicitor for the generation of systemic resistance has already been established. The exopolysaccharide synthesized by *Bacillus* sp. EPS has been shown to effectively minimize the occurrence of wilt disease in tomatoes caused by *F. oxysporum* (Thenmozhi and Dinakar 2014).

15.5.4 Production of Cellulolytic/Cell Wall-Degrading Enzymes

Cellulolytic/cell wall-degrading enzymes secreted and excreted by certain bacteria can disrupt pathogen development and/or activity. The secretion of these enzymes is a cost-effective way to stop pathogen proliferation through the lysis of pathogenic cell walls. Several bacterial genera produce and release lytic enzymes that are capable of hydrolyzing a wide range of polymeric materials (proteins chitin, cellulose, hemicellulose and DNA). This technique allows the phytopathogen to be directly parasitized. The release and expression of these enzymes by various

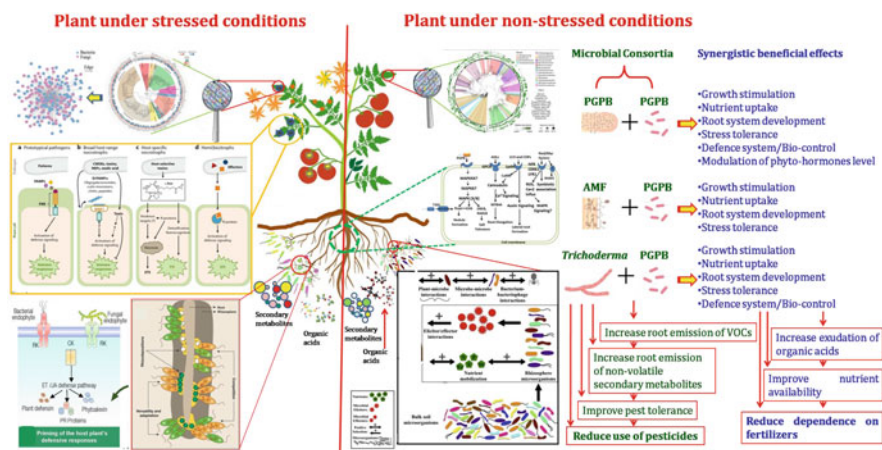


Fig. 15.2 An overview of microbe-mediated modulation of physio-biochemical and molecular mechanisms of biotic stress tolerance in plants. Photograph clearly depicts that plant secreted chemically different biochemicals/secondary metabolites which regulate recruitment of microbial strains under stressed and non-stressed conditions. These microorganisms further regulate the defence mechanisms in plants. Figure showed the multi-trophic interactions in the rhizosphere which define the active rhizosphere effects and how the microbiome served as the first line of defence and maintain defence level and plant growth under stressed conditions

microorganisms can occasionally directly decrease plant pathogen activity. Poly- 430
meric components (such as cellulose, hemicelluloses, chitin, proteins and DNA) 431
found in fungal cell walls can be digested by lytic enzymes including cellulase, 432
chitinase and protease, among others. *Pseudomonas* sp., among soil bacteria, has 433
gotten increased attention since it can produce a variety of cell wall disintegrating 434
enzymes, including chitinase, protease/elastase and 1,3-glucanase (Wang et al. 435
2021). 436

Chitinases and 1,3-glucanases are two of the most important hydrolytic enzymes 437
involved in the breakdown of fungal cell walls. Mycolytic enzymes produced by 438
rhizobia, particularly chitinases, are known to hydrolyze chitin, a significant component 439
of fungal cell walls (Chavan and Deshpande 2013). Production of extracellular 440
cellulolytic enzymes (cellulase, chitinases, protease and β -1,3-glucanase) by 441
Bacillus sp. recovered from *Solanum lycopersicum* rhizosphere inhibited the fungal 442
pathogens and proved to be a viable bioresource for the agricultural business (Kumar 443
et al. 2012). In a study, Ramyabharathi and Raguchander (2014) found that antagonistic 444
PGPR strain *B. subtilis* EPCO16 produced HCN and volatile organic compounds (VOCs) 445
together with proteolytic and extracellular enzymes chitinase and β -1,3-glucanase. This strain 446
retarded the growth of *F. oxysporum* f. sp. *lycopersici* 447
causing fusarium wilt disease in tomatoes and proved to be a successful biocontrol 448
agent in managing the fungal diseases. Further, strains of *Pseudomonas* 449
(*P. fluorescences* NRC1 and *P. fluorescens* NRC3) inhibited the phytopathogens 450

451 *Phytophthora capsica* and *R. solani* (the causal agents of root rot disease in tomato)
452 by secreting different cell wall-degrading enzymes and synthesizing antifungal
453 metabolites (Moataza 2006). Under greenhouse circumstances, the strain *Bacillus*
454 sp., which could synthesize different types of extracellular enzymes, reduced the
455 disease incidence of wilt disease (caused by *F. oxysporum* f. sp. *lycopersici*) by 36%
456 in tomato plants, suggesting that it could be an effective biocontrol agent for tomato
457 wilt (Jangir et al. 2018). *P. fluorescens* has also been identified as a potent BCA
458 against the fungal pathogen, stimulating important enzymes such as peroxidase
459 (POX), polyphenol oxidase (PPO) and superoxide dismutase (SOD), as well as
460 1,3-glucanases (Dorjey et al. 2017). Apart from the extracellular enzymes, the
461 inhibition of phytopathogens could also be aided by other microbial by-products.
462 For instance, at picomolar concentrations, hydrogen cyanide (HCN) efficiently dis-
463 ables the cytochrome oxidase pathway and is extremely hazardous to all aerobic
464 microbes. Hydrolytic enzymes produced by some identified strains of *Bacillus* have
465 been described for the biocontrol of the phytopathogen *F. oxysporum* causing wilt
466 disease in tomatoes (Jadhav and Sayyed 2016). The efficiency of different
467 rhizobacterial strains is dependent on the host plant and soil factors, in addition to
468 the biocontrol methods listed above. Furthermore, their innate capacities and rhizo-
469 sphere competence play a significant role in the manifestation of their biocontrol
470 features (Weert and Bloemberg 2007). In a field trial, Shanmugam and Kanoujia
471 (2011) evaluated the biocontrol potential of antagonistic PGPR strains, *B. cepacia*
472 and *B. subtilis* against *Fusarium oxysporum* f. sp. *lycopersici* causing wilt disease in
473 tomato. They found that the bacterial strains reduced the incidence of disease by
474 secreting extracellular enzymes. Further, they claimed that leaf and root sample
475 analyses had the highest induction of chitinase and 1,3-glucanase.

476 15.5.5 Production of Hydrogen Cyanide (HCN)

477 Hydrogen cyanide (HCN) is an antibacterial and antifungal molecule well known for
478 its ability to prevent disease (Haas and Keel 2003). HCN is likely to disrupt the
479 electron transport chain (ETS) and the cell's energy source, resulting in cell death.
480 Cytochrome oxidase is also known to be inhibited by the action of HCN (Cooper and
481 Brown 2008). This detrimental characteristic gives PGPR a competitive advantage
482 over fungal pathogens, and it can be used in plant disease biocontrol (Shaikh and
483 Sayyed 2015). Among rhizosphere microbes, *Pseudomonas* and *Bacillus* spp., for
484 example, produce cyanogenic compound (HCN) as a secondary metabolite
485 (Sivasakthi et al. 2014). Bacterial cyanogenesis has also been described in the
486 species of *Burkholderia* (Shahid and Khan 2018), *Achromobacter* (Oves et al.
487 2019), *Bacillus* (Hassan et al. 2010), and *Azotobacter* (Shahid et al. 2019). However,
488 fluorescent *Pseudomonas* is the most common HCN produce (Keerthana et al.
489 2022). The majority of research suggests that HCN-producing bacteria are active

against fungal infections and, thus, serve as a biocontrol agent (Sehrawat et al. 2022; Haddoudi et al. 2021; Hernández-León et al. 2015; Pandya and Saraf 2014). In a study, Kumar et al. (2012) found that HCN-synthesizing *Bacillus* sp. suppressed the disease-causing pathogen *Sclerotium rolfsii*. Similarly, PGPR strain *Beijerinckia fluminensis* suppressed the growth of major fungal phytopathogens while assessed under in vitro conditions (Al-Shwaiman et al. 2022).

15.5.6 Production of Siderophore

Iron (Fe) is a necessary micronutrient for all living things and is found in the soil mostly in the form of ferric ion (Fe^{3+}), which is only sparingly soluble. As a result, plants are unable to absorb it. Plants can utilize the microorganism-produced siderophores to absorb iron (Radzki et al. 2013). Beneficial soil microbial diversity synthesizes siderophores (low-molecular-weight Fe-chelating molecules) having high affinity and selectivity for binding and forming a complex Fe complex (III) (Colombo et al. 2014). It functions as a ligand, allowing iron to be sequestered and transported into the cell. In recent times, this characteristic has gotten a lot of attention, and the in vitro assessment of siderophore synthesized by identified PGPR has been reported (Parray et al. 2016). Rather than Gram-positive PGPR transport systems, siderophore-mediated iron transfer mechanisms are best researched in Gram-negative PGPR transport systems. Among different varieties of 500 siderophores, only 270 have been structurally studied. Siderophores have been proposed as an environmentally acceptable alternative to insecticides.

Since siderophores have a higher affinity for iron than fungal pathogens, they have a competitive edge when it comes to effectively reducing the proliferation of phytopathogen (Govindasamy et al. 2008). When there is a lack of iron, fungal pathogens become unable to reproduce and are pushed out of their biological habitat. As a result, siderophore synthesis is an attractive feature of PGPR as a biocontrol agent. Siderophores produced by *Pseudomonas* are reported to have a stronger affinity than other bacterial siderophores (Patel et al. 2018). Several studies have been published on the biocontrol efficacy of PGPR that produce siderophores. Siderophore-producing strains of *B. subtilis* MF497446 and *P. koreensis* MG209738 exhibited strong effectiveness against the fungal pathogens, reduced the disease incidence and improved the crop growth (Ghazy and El-Nahrawy 2021). Under field circumstances, Attia et al. (2020) found that siderophore-producing PGPR strains, viz. *Lysinibacillus fusiformis* L-2, *Bacillus subtilis* B-1 and *Achromobacter xylosoxidans* A-3 suppressed the early blight disease of tomato caused by *Alternaria solani*. Similarly, the indigenously isolated *Pseudomonas* strain reduced the incidence of fusarium wilt disease in tomato crops by producing siderophore (Arya et al. 2018). In addition, siderophore-synthesizing *Streptomyces* SNL2 reduced the prevalence of wilt disease by 88.5% (Goudjal et al. 2016).

529 15.6 Future Perspective

530 Managing sustainable natural reserves can help ensure food security for the world's
531 growing population. PGPB have been shown to play an important function in
532 agricultural management in several studies. Though, there is still a knowledge
533 vacuum underpinning microbe-plant symbiosis under various stress circumstances,
534 especially pathogen stress. Understanding the rhizosphere ecology that governs
535 pathogen and antagonist dispersion could help improve biocontrol efficiency against
536 plant-based pathogens. Future studies will necessitate intense rhizosphere engineer-
537 ing based on the successful discovery and separation of novel metabolites, which
538 could establish a unique environment for plant-microbe interactions. Instead, explor-
539 ing and applying the combined inoculation over a single strain, on the other hand,
540 could be an efficient means for the suppression of fungal diseases. Furthermore,
541 genetic alterations to improve biocontrol values could be a new study area for
542 managing plant diseases. Transforming the strains with higher quantities of antifun-
543 gal and growth-promoting essential metabolites, for example, maybe an excellent
544 option. The use of cutting-edge tools to explore microbiological, biochemical and
545 molecular interactions between plants and interacting microorganisms may provide
546 in-depth knowledge for a better understanding of interactions between plants and
547 microbes. To summarize, the forthcoming challenge will be to improve the effec-
548 tiveness and long-term resilience of biological control in the field. If this issue is
549 handled, biocontrol efficacy could be increased by leveraging expertise to design
550 better screening processes, formulations and application procedures, as well as
551 innovative integrated disease management strategies.

552 15.7 Conclusions

553 Tomato is a highly nutritious vegetable crop grown all over the world and ranks
554 second only to potatoes in terms of consumption. Tomato seed-borne infections, on
555 the other hand, are a major source of concern in the seed industry because they have
556 a negative impact on seedling germination and vigour, resulting in a significant
557 reduction in yield and product quality. The current research shows that PGPR not
558 only have different biological promotional effects on tomato plant development
559 parameters but also operate as biocontrol agents (BCAs) to protect the plant from
560 diseases. It should become increasingly more effective and cost-effective to replace
561 fungicides with a biological pesticide. Before commercialization, molecular analysis
562 can help stabilize the effects of PGPR in biological control and determine potential
563 risks. To use PGPR effectively for disease reduction or crop protection in the future,
564 a logical selection of organisms will be required, as well as technical improvements
565 in upscaling and formulation procedures will be needed. The PGP genomic products
566 may be enhanced through the genetic engineering of PGPR. As a result, a single
567 bacterial strain or a consortium with varying features will reduce pathogen attacks

while also promoting plant growth, which will stimulate producers. Although investigations should focus on the relative contributions of each mechanism responsible for PGPR strains' significant biocontrol activity, it has become obvious that they use numerous pathways to operate as an effective biocontrol agent. An in-depth study of microbial interactions with plants, as well as exploitation of microbial ecology in the soil and rhizosphere, will aid in revealing the many dimensions of disease suppression by these biocontrol agents. Furthermore, for maximal commercial utilization of these strains, cautiously conducted skilful field trials of tomato plants inoculated with BCAs are required. Finally, the success of the microbial inoculant industry, particularly those that use PGPRs, will be determined by factors such as product marketing and substantial research. Furthermore, to incorporate PGPR strains into the agriculture industry, they will need to be optimized for improved fermentation and formulation procedures.

Acknowledgements The authors sincerely thank Director Mau, ICAR-NBAIM, for providing scientific and technical support during preparation of the manuscript. The authors gratefully acknowledge the Network Project on Application of Microorganisms in Agriculture and Allied Sectors (AMAAS), ICAR-NBAIM and Indian Council of Agricultural Research, Ministry of Agriculture and Farmers Welfare, Government of India for providing financial support for the study.

Funding This research was supported by Network Project on Application of Microorganisms in Agriculture and Allied Sectors (AMAAS), ICAR-NBAIM and Indian Council of Agricultural Research, New Delhi (India).

Conflicts of Interest The authors declare that they have no known competing financial interest or personal relationship that could have appeared to influence the content reported in this manuscript. The authors declare no conflict of interest.

References

- Abo-Elyousr KA, Bagy HM, Hashem M, Alamri SA, Mostafa YS (2019) Biological control of the tomato wilt caused by *Clavibacter michiganensis* subsp. *michiganensis* using formulated plant growth-promoting bacteria. Egypt J Biological Pest Control 29:1–8
- Ahmad T, Bashir A, Farooq S, Riyaz-Ul-Hassan S (2022) *Burkholderia gladioli* E39CS3, an endophyte of *Crocus sativus* Linn., induces host resistance against corm-rot caused by *Fusarium oxysporum*. J Appl Microbiol 132:495–508
- Ahmed HE, Zienat KM, Mohamed EE, Mohamed GF, Zienat K (2011) Induced systemic protection against tomato leaf spot (early leaf blight) and bacterial speck by rhizobacterial isolates. J Exp Biol 20:49–57
- Ahmed B, Shahid M, Syed A, Rajput VD, Elgorban AM, Minkina T, Bahkali AH, Lee J (2021) Drought tolerant *Enterobacter* sp./*Leclercia adecarboxylata* secretes indole-3-acetic acid and other biomolecules and enhances the biological attributes of *Vigna radiata* (L.) R. Wilczek in water deficit conditions. Biology 10:1149
- Ali MY, Sina AA, Khandker SS, Neesa L, Tanvir EM, Kabir A, Khalil MI, Gan SH (2020) Nutritional composition and bioactive compounds in tomatoes and their impact on human health and disease: a review. Foods 10(1):45

- Al-Kassim MY, Monawar MN (2000) Seed-borne fungi of some vegetable seeds in Jazan province and their chemical control. *Saudi J Biol Sci* 7(2):179–184
- Al-Shwaiman HA, Shahid M, Elgorban AM, Siddique KH, Syed A (2022) *Beijerinckia fluminensis* BFC-33, a novel multi-stress-tolerant soil bacterium: deciphering the stress amelioration, phytopathogenic inhibition and growth promotion in *Triticum aestivum* (L.). *Chemosphere* 295:133843
- Alsudani AA (2022) Biocontrol of *Rhizoctonia solani* (Kühn) and *Fusarium solani* (Marti) causing damping-off disease in tomato with *Azotobacter chroococcum* and *Pseudomonas fluorescens*. *Pak J Biol Sci* 23:1456–1461
- Amaya-Gómez CV, Porcel M, Mesa-Garriga L, Gómez-Álvarez MI (2020) A framework for the selection of plant growth-promoting rhizobacteria based on bacterial competence mechanisms. *Appl Environ Microbiol* 86(14):e00760–e00720
- Amini J, Sidovich D (2010) The effects of fungicides on *Fusarium oxysporum* f. sp. *lycopersici* associated with Fusarium wilt of tomato. *J Plant Protect Res* 50(2):172–178
- Amza J (2018) Seed borne fungi; food spoilage, negative impact and their management: a review. *FSQM* 81:70–79
- Arora DK (1986) Chemotaxis of *Actinoplanes missouriensis* zoospores to fungal conidia, chlamydospores and sclerotia. *Microbiology* 132:1657–1663
- Arunakumara KT, Kulkarni MS, Thammaiah N, Yashoda H (2010) Fungicidal management of early blight of tomato. *Indian Phytopathol* 63:96–97
- Arya N, Rana A, Rajwar A, Sahgal M, Sharma AK (2018) Biocontrol efficacy of siderophore producing indigenous *Pseudomonas* strains against Fusarium wilt in tomato. *Nat Acad Sci Lett* 41:133–136
- Atkinson NJ, Dew TP, Orfila C, Urwin PE (2011) Influence of combined biotic and abiotic stress on nutritional quality parameters in tomato (*Solanum lycopersicum*). *J Agric Food Chem* 59:9673–9682
- Attia MS, El-Sayyad GS, Abd Elkodous M, El-Batal AI (2020) The effective antagonistic potential of plant growth-promoting rhizobacteria against *Alternaria solani*-causing early blight disease in tomato plant. *Sci Hortic* 10(266):109289
- Babalola OO, Emmanuel OC, Adeleke BS, Odelade KA, Nwachukwu BC, Ayiti OE, Adegboyega TT, Igiehon NO (2021) Rhizosphere microbiome cooperations: strategies for sustainable crop production. *Curr Microbiol* 78:1069–1085
- Badri DV, Vivanco JM (2009) Regulation and function of root exudates. *Plant Cell Environ* 32:666–681
- Basco MJ, Bisen K, Keswani C, Singh HB (2017) Biological management of fusarium wilt of tomato using biofortified vermicompost. *Mycosphere* 8:467–483
- Bhatti FJ, Ghazal H, Irshad G, Begum N, Bhutta AR (2010) Study on seed-borne fungi of vegetable seeds. *Pak J Seed Tech* 2:96–106
- Bu S, Munir S, He P, Li Y, Wu Y, Li X, Kong B, He P, He Y (2021) *Bacillus subtilis* L1-21 as a biocontrol agent for postharvest gray mold of tomato caused by *Botrytis cinerea*. *Biol Control* 157:104568
- Chacon-Lopez A, Guardado-Valdivia L, Banuelos-Gonzalez M, Lopez-Garcia U, Montalvo-González E, Arvizu-Gomez J, Stoll A, Aguilera S (2021) Effect of metabolites produced by *Bacillus atrophaeus* and *Brevibacterium frigoritolerans* strains on postharvest biocontrol of *Alternaria alternata* in tomato (*Solanum lycopersicum* L.). *Biocontrol Sci* 26:67–74
- Chaouachi M, Marzouk T, Jallouli S, Elkahoui S, Gentzittel L, Ben C, Djébal N (2021) Activity assessment of tomato endophytic bacteria bioactive compounds for the postharvest biocontrol of *Botrytis cinerea*. *Postharvest Biol Tech* 172:111389
- Chavan SB, Deshpande MV (2013) Chitinolytic enzymes: an appraisal as a product of commercial potential. *Biotechnol Prog* 29:833–846
- Chen Y, Yan F, Chai Y, Liu H, Kolter R, Losick R, Guo JH (2013) Biocontrol of tomato wilt disease by *Bacillus subtilis* isolates from natural environments depends on conserved genes mediating biofilm formation. *Environ Microbiol* 15(3):848–864

- Chohan S, Perveen R, Abid M, Naqvi AH, Naz S (2017) Management of seed borne fungal diseases of tomato: a review. *Pak J Phytopathol* 29:193–200
- Choudhary DK, Johri BN (2009) Interactions of *Bacillus* spp. and plants—with special reference to induced systemic resistance (ISR). *Microbiol Res* 164:493–513
- Chowdappa P, Kumar SM, Lakshmi MJ, Upreti KK (2013) Growth stimulation and induction of systemic resistance in tomato against early and late blight by *Bacillus subtilis* OTPB1 or *Trichoderma harzianum* OTPB3. *Biol Control* 65:109–117
- Chunyu LI, Weicong HU, Bin PA, Yan LI, Saifei YU, Yuanyuan DI, Rong LI, Zheng X, Biao SH, Qirong SH (2017) Rhizobacterium *Bacillus amyloliquefaciens* strain SQRT3-mediated induced systemic resistance controls bacterial wilt of tomato. *Pedosphere* 27:1135–1146
- Cohen Y, Rubin AE, Galperin M (2018) Oxathiapiprolin-based fungicides provide enhanced control of tomato late blight induced by mefenoxam-insensitive *Phytophthora infestans*. *PLoS One* 13:e0204523
- Çolak NG, Eken NT, Ülger M, Frary A, Doğanlar S (2020) Mapping of quantitative trait loci for antioxidant molecules in tomato fruit: carotenoids, vitamins C and E, glutathione and phenolic acids. *Plant Sci* 292:110393
- Colombo C, Palumbo G, He JZ, Pinton R, Cesco S (2014) Review on iron availability in soil: interaction of Fe minerals, plants, and microbes. *J Soils Sediments* 14:538–548
- Cooper CE, Brown GC (2008) The inhibition of mitochondrial cytochrome oxidase by the gases carbon monoxide, nitric oxide, hydrogen cyanide and hydrogen sulfide: chemical mechanism and physiological significance. *J Bioenerg Biomembr* 40:533–539
- Copping LG, Duke SO (2007) Natural products that have been used commercially as crop protection agents. *Pest Manag Sci* 63:524–554
- Costa JM, Heuvelink EP (2018) The global tomato industry. In: *Tomatoes*, vol 1. CABI, Boston, pp 1–26
- Cuppels DA, Higham J, Traquair JA (2013) Efficacy of selected streptomycetes and a streptomycete+ pseudomonad combination in the management of selected bacterial and fungal diseases of field tomatoes. *Biol Control* 67:361–372
- Daguerre Y, Siegel K, Edel-Hermann V, Steinberg C (2014) Fungal proteins and genes associated with biocontrol mechanisms of soil-borne pathogens: a review. *Fungal Biol Rev* 28:97–125
- Dhekke NM, Bodke SS (2013) Studies on fungal diversity associated with cauliflower, tomato and bhendi. *Rev Res J* 2:1–7
- Divya Rani V, Sudini H (2013) Management of soil-borne diseases in crop plants: an overview. *Int J Plant Anim Environ Sci* 3:156–164
- Dorjey S, Dolkar D, Sharma R (2017) Plant growth promoting rhizobacteria *Pseudomonas*: a review. *Int J Curr Microbiol App Sci* 6:1335–1344
- Droby S, Gonzalez-Estrada RR, Avila-Quezada G, Durán P, Manzo-Sánchez G, Hernandez-Montiel LG (2022) Microbial antagonists from different environments used in the biocontrol of plant pathogens. In: *Microbial biocontrol: food security and post harvest management*. Springer, Cham, pp 227–244
- Essalimi B, Esserti S, Rifai LA, Koussa T, Makroum K, Belfaiza M, Rifai S, Venisse JS, Faize L, Albuquerque N, Burgos L (2022) Enhancement of plant growth, acclimatization, salt stress tolerance and verticillium wilt disease resistance using plant growth-promoting rhizobacteria (PGPR) associated with plum trees (*Prunus domestica*). *Sci Hortic* 291:110621
- Etesami H, Adl SM (2020) Plant growth-promoting rhizobacteria (PGPR) and their action mechanisms in availability of nutrients to plants. In: *Phyto-microbiome in stress regulation*, pp 147–203
- Fatima I, Hakim S, Imran A, Ahmad N, Imtiaz M, Ali H, Islam EU, Yousaf S, Mirza MS, Mubeen F (2022) Exploring biocontrol and growth-promoting potential of multifaceted PGPR isolated from natural suppressive soil against the causal agent of chickpea wilt. *Microbiol Res* 260:127015
- Ferraz HG, Resende RS, Moreira PC, Silveira PR, Milagres EA, Oliveira JR, Rodrigues FA (2015) Antagonistic rhizobacteria and jasmonic acid induce resistance against tomato bacterial spot. *Bragantia* 74:417–427

- Freitas CS, Maciel LF, Corrêa dos Santos RA, Costa OM, Maia FC, Rabelo RS, Franco HC, Alves E, Consonni SR, Freitas RO, Persinoti GF (2022) Bacterial volatile organic compounds induce adverse ultrastructural changes and DNA damage to the sugarcane pathogenic fungus *Thielaviopsis ethacetica*. *Environ Microbiol* 24(3):1430–1453. <https://doi.org/10.1111/1462-2920.15876>
- Fry WE (2020) Phytophthora infestans: the itinerant invader; “late blight”: the persistent disease. *Phytoparasitica* 48:87–94
- Gaur I, Sharma PD, Paul PK (2017) Effect of *Klebsiella pneumoniae* on speck disease development in *Solanum lycopersicum*. *Indian J Agr Res* 51(5):431–436
- Ghazy N, El-Nahrawy S (2021) Siderophore production by *Bacillus subtilis* MF497446 and *Pseudomonas koreensis* MG209738 and their efficacy in controlling *Cephalosporium maydis* in maize plant. *Arch Microbiol* 203:1195–1209
- Gomez-Ramirez LF, Uribe-Velez D (2021) Phosphorus solubilizing and mineralizing *bacillus* spp. contribute to rice growth promotion using soil amended with rice straw. *Curr Microbiol* 78:932–943
- Goudjal Y, Toumatia O, Yekkour A, Sabaou N, Mathieu F, Zitouni A (2014) Biocontrol of *Rhizoctonia solani* damping-off and promotion of tomato plant growth by endophytic actinomycetes isolated from native plants of Algerian Sahara. *Microbiol Res* 169:59–65
- Goudjal Y, Zamoum M, Sabaou N, Mathieu F, Zitouni A (2016) Potential of endophytic *Streptomyces* spp. for biocontrol of Fusarium root rot disease and growth promotion of tomato seedlings. *Biocontrol Sci Tech* 26:1691–1705
- Govindasamy V, Senthilkumar M, Upendra-Kumar AK (2008) PGPR-biotechnology for management of abiotic and biotic stresses in crop plants. In: *Potential microorganisms for sustainable agriculture*, pp 26–48
- Gowtham HG, Hariprasad P, Nayak SC, Niranjana SR (2016) Application of rhizobacteria antagonistic to *Fusarium oxysporum* f. sp. *lycopersici* for the management of Fusarium wilt in tomato. *Rhizosphere* 2:72–74
- Gupta SK, Bharat NK (2008) Management of buckeye rot and late blight of tomato through combi fungicides. *Pestology* 32:17–19
- Haas D, Keel C (2003) Regulation of antibiotic production in root-colonizing *Pseudomonas* spp. and relevance for biological control of plant disease. *Annu Rev Phytopathol* 41:117–153
- Haddoudi I, Cabrefiga J, Mora I, Mhadhbi H, Montesinos E, Mrabet M (2021) Biological control of Fusarium wilt caused by *Fusarium equiseti* in *Vicia faba* with broad spectrum antifungal plant-associated *Bacillus* spp. *Biol Control* 160:104671
- Hamid S, Lone R, Mohamed HI (2021) Production of antibiotics from PGPR and their role in biocontrol of plant diseases. In: *Plant growth-promoting microbes for sustainable biotic and abiotic stress management*. Springer, Cham, pp 441–461
- Hassan MN, Osborn AM, Hafeez FY (2010) Molecular and biochemical characterization of surfactin producing *Bacillus* species antagonistic to *Colletotrichum falcatum* Went causing sugarcane red rot. *Afr J Microbiol Res* 4:2137–2142
- Hassan MK, McInroy JA, Kloepper JW (2019) The interactions of rhizodeposits with plant growth-promoting rhizobacteria in the rhizosphere: a review. *Agriculture* 9:142
- Heidarzadeh N, Baghaee-Ravari S (2015) Application of *Bacillus pumilus* as a potential biocontrol agent of Fusarium wilt of tomato. *Arch Phytopathol Plant Protect* 48:841–849
- Heo AY, Koo YM, Choi HW (2022) Biological control activity of plant growth promoting rhizobacteria *Burkholderia contaminans* AY001 against tomato Fusarium wilt and bacterial speck diseases. *Biology* 11:619
- Hernández-León R, Rojas-Solís D, Contreras-Pérez M, del Carmen Orozco-Mosqueda M, Macías-Rodríguez LI, Reyes-de la Cruz H, Valencia-Cantero E, Santoyo G (2015) Characterization of the antifungal and plant growth-promoting effects of diffusible and volatile organic compounds produced by *Pseudomonas fluorescens* strains. *Biol Control* 81:83–92
- Hussain T, Khan AA, Khan MA (2021) Biocontrol of soil borne pathogen of potato tuber caused by *Rhizoctonia solani* through Biosurfactant based *Bacillus* strain. *J Nepal Agril Res Council* 7: 54–66

- Jadhav HP, Sayyed RZ (2016) Hydrolytic enzymes of rhizospheric microbes in crop protection. *MOJ Cell Sci Rep* 3:135–136
- Jangir M, Pathak R, Sharma S, Sharma S (2018) Biocontrol mechanisms of *Bacillus* sp., isolated from tomato rhizosphere, against *Fusarium oxysporum* f. sp. *lycopersici*. *Biol Control* 123: 60–70
- Jisha MS, Linu MS, Sreekumar J (2019) Induction of systemic resistance in chilli (*Capsicum annum* L.) by *Pseudomonas aeruginosa* against anthracnose pathogen *Colletotrichum capsici*. *J Tropical Agriculture* 56(2):153–166
- Jnr L (2000) Sclerotinia rot losses in processing tomatoes grown under Centre pivot irrigation in Central Brazil. *Plant Pathol* 49:51–56
- Johnson I, Sreenayana B, Suruthi VP, Manikandan R, Ramjegathesh R, Karthikeyan M (2022) Rhizosphere population dynamics and biocontrol potential of *Pseudomonas fluorescens* Pf1 against Wilt and collar rot pathogens in tomato. *Pharma Innovat* 11(5):1042–1051
- Kabdwal BC, Sharma R, Tewari R, Tewari AK, Singh RP, Dandona JK (2019) Field efficacy of different combinations of *Trichoderma harzianum*, *Pseudomonas fluorescens*, and arbuscular mycorrhiza fungus against the major diseases of tomato in Uttarakhand (India). *Egypt J Biol Pest Control* 29:1. <https://doi.org/10.1186/s41938-018-0103-7>
- Kamali M, Guo D, Naeimi S, Ahmadi J (2022) Perception of biocontrol potential of *Bacillusinaquosorum* KR2-7 against tomato fusarium wilt through merging genome mining with chemical analysis. *Biology* 11:137
- Kamilova F, Leveau JH, Lugtenberg B (2007) *Collimonasfungivorans*, an unpredicted in vitro but efficient in vivo biocontrol agent for the suppression of tomato foot and root rot. *Environ Microbiol* 9:1597–1603
- Kaur P, Kaur J (2015) Potential role of lycopene as antioxidant and implications for human health and disease. In: Bailey JR (ed) *Lycopene food sources, potential role in human health and antioxidant effects*. Nova Science, pp 1–38
- Kaya HK, Koppenhöfer AM (1996) Effects of microbial and other antagonistic organism and competition on entomopathogenic nematodes. *Biocontrol Sci Tech* 1:357–372
- Keerthana U, Prabhukarthikeyan SR, Baite MS, Yadav MK, Kumar RN, Kumar AM, Raghu S, Aravindan S, Rath PC (2022) Fluorescent pseudomonads: a multifaceted biocontrol agent for sustainable agriculture. In: *New and future developments in microbial biotechnology and bioengineering*, pp 83–92
- Keinath AP, DuBose VB (2017) Management of southern blight on tomato with SDHI fungicides. *Crop Prot* 101:29–34
- Kilani-Feki O, Khedher SB, Dammak M, Kamoun A, Jabnoun-Khiareddine H, Daami-Remadi M, Tounsi S (2016) Improvement of antifungal metabolites production by *Bacillus subtilis* V26 for biocontrol of tomato postharvest disease. *Biol Control* 95:73–82
- Kim PI, Ryu JW, Kim YH, Chi YT (2010) Production of biosurfactant lipopeptides iturin A, fengycin, and surfactin A from *Bacillus subtilis* CMB32 for control of *Colletotrichum gloeosporioides*. *J Microbiol Biotechnol* 20:138–145
- Koley S, Mahapatra SS, Koley PC (2015) In vitro efficacy of bio-control agents and botanicals on the growth inhibition of *Alternaria solani* causing early leaf blight of tomato. *Int J Bio-res Env Agril Sci* 1:114–118
- Kuiper I, Bloemberg GV, Noreen S, Thomas-Oates JE, Lugtenberg BJ (2001) Increased uptake of putrescine in the rhizosphere inhibits competitive root colonization by *Pseudomonas fluorescens* strain WCS365. *Mol Plant Microbe Int* 14:1096–1104
- Kuiper I, Kravchenko LV, Bloemberg GV, Lugtenberg BJ (2002) *Pseudomonas putida* strain PCL1444, selected for efficient root colonization and naphthalene degradation, effectively utilizes root exudate components. *Mol Plant Microbe Int* 15:734–741
- Kumar KH, Jagadeesh KS (2016) Microbial consortia-mediated plant defence against phytopathogens and growth benefits. *South Indian J Biol Sci* 2:395–403
- Kumar DP, Anupama PD, Singh RK, Thenmozhi R, Nagasathya A, Thajuddin N, Paneerselvam A (2012) Evaluation of extracellular lytic enzymes from indigenous *Bacillus* isolates. *J Microbiol Biotechnol Res* 2:129–137

- 825 Kumari P, Bishnoi SK, Chandra S (2021) Assessment of antibiosis potential of *Bacillus* sp. against
826 the soil-borne fungal pathogen *Sclerotium rolfsii* Sacc. (*Atheliarolfsii* (Curzi) Tu & Kimbrough).
827 Egyptian J Biol Pest Control 31:1
- 828 Lee JP, Lee SW, Kim CS, Son JH, Song JH, Lee KY, Kim HJ, Jung SJ, Moon BJ (2006) Evaluation
829 of formulations of *Bacillus licheniformis* for the biological control of tomato graymold caused
830 by *Botrytis cinerea*. Biol Control 37:329–337
- 831 Lian Q, Zhang J, Gan L, Ma Q, Zong Z, Wang Y (2017) The biocontrol efficacy of *Streptomyces*
832 *pratensis* LMM15 on *Botrytis cinerea* in tomato. Biomed Res Int 28:2017
- 833 Liu F, Xing S, Ma H, Du Z, Ma B (2013) Cytokinin-producing, plant growth-promoting
834 rhizobacteria that confer resistance to drought stress in *Platycladus orientalis* container seed-
835 lings. Appl Microbiol Biotechnol 97:9155–9164
- 836 Lugtenberg B (2015) Life of microbes in the rhizosphere. In: Principles of plant-microbe interac-
837 tions. Springer, Cham, pp 7–15
- 838 Malik KA, Bilal R, Mehnaz S, Rasul G, Mirza MS, Ali S (1997) Association of nitrogen-fixing,
839 plant-growth-promoting rhizobacteria (PGPR) with kallar grass and rice. In: Opportunities for
840 biological nitrogen fixation in rice and other non-legumes. Springer, Dordrecht, pp 37–44
- 841 Malik MS, Haider S, Rehman A, Rehman SU, Jamil M, Naz I, Anees M (2022) Biological control
842 of fungal pathogens of tomato (*Lycopersicon esculentum*) by chitinolytic bacterial strains. J
843 Basic Microbiol 62:48–62
- 844 Mandal S, Ray RC (2011) Induced systemic resistance in biocontrol of plant diseases. In:
845 Bioaugmentation, bio stimulation and biocontrol. Springer, Berlin, Heidelberg, pp 241–260
- 846 Marinho MD, Diogo BS, Lage OM, Antunes SC (2020) Ecotoxicological evaluation of fungicides
847 used in viticulture in non-target organisms. Environ Sci Pollut Res 27:43958–43969
- 848 Mazzotta E, Muzzalupo R, Chiappetta A, Muzzalupo I (2022) Control of the verticillium wilt on
849 tomato plants by means of olive leaf extracts loaded on chitosan nanoparticles. Microorganism
850 10:136
- 851 Miljaković D, Marinković J, Balešević-Tubić S (2020) The significance of *Bacillus* spp. in disease
852 suppression and growth promotion of field and vegetable crops. Microorganisms 8:1037
- 853 Moataza MS (2006) Destruction of *Rhizoctonia solani* and *Phytophthora capsici* causing tomato
854 root-rot by *Pseudomonas fluorescences* lytic enzymes. Res J Agri Biol Sci 2:274–281
- 855 Monteiro FP, Ogoshi C, Cardoso DA, Perazzoli V, Maindra LC, Pinto FA, Mallmann G (2021)
856 Fungicides in the control of septoriose in tomato plant. Plant Pathol Quarantine 11(1):173–190
- 857 Morganeli A (2007) The biography of tomatoes. Crabtree Publishing Company
- 858 Myo EM, Liu B, Ma J, Shi L, Jiang M, Zhang K, Ge B (2019) Evaluation of *Bacillus velezensis*
859 NKG-2 for bio-control activities against fungal diseases and potential plant growth promotion.
860 Biol Control 134:23–31
- 861 Narayanasamy P (2013) Mechanisms of action of bacterial biological control agents. In: Biological
862 management of diseases of crops. Springer, Dordrecht, pp 295–429
- 863 Neergard P (1997) Seed pathology. The Macmillan Press Limited, Danist Govt. Institute of Seed
864 Pathology for Developing Countries, Copenhagen, p 1
- 865 Ni L, Punja ZK (2019) Management of fungal diseases on cucumber (*Cucumis sativus* L.) and
866 tomato (*Solanum lycopersicum* L.) crops in greenhouses using *Bacillus subtilis*. In: Bacilli and
867 agrobiotechnology: phytostimulation and biocontrol. Springer, Cham, pp 1–28
- 868 Nicola S, Tibaldi G, Fontana E, Crops AV, Plants A (2009) Tomato production systems and their
869 application to the tropics. Acta Hort 821:27–34
- 870 Nishikawa J, Kobayashi T, Shirata K, Chibana T, Natsuaki KT (2006) Seedborne fungi detected on
871 stored solanaceous berry seeds and their biological activities. J General Plant Pathol 72:305–313
- 872 Noori MS, Saud HM (2012) Potential plant growth-promoting activity of *Pseudomonas* sp. isolated
873 from paddy soil in Malaysia as biocontrol agent. J Plant Pathol Microbiol 3:1–4
- 874 Oves M, Khan MS, Qari HA (2019) Chromium-reducing and phosphate-solubilizing
875 *Achromobacter xylosoxidans* bacteria from the heavy metal-contaminated soil of the Brass
876 city, Moradabad, India. Int J Environ Sci Tech 16:6967–6984
- 877 Pal KK, Gardener BM (2006) Biological control of plant pathogens. Plant Health Instructor. <https://doi.org/10.1094/PHI-A-2006-1117-02>
878

- Pandya U, Saraf M (2014) In vitro evaluation of PGPR strains for their biocontrol potential against fungal pathogens. In: Microbial diversity and biotechnology in food security. Springer, New Delhi, pp 293–305
- Pane C, Zaccardelli M (2015) Evaluation of *Bacillus* strains isolated from solanaceous phylloplane for biocontrol of *Alternaria* early blight of tomato. Biol Control 84:11–18
- Panthee DR, Chen F (2010) Genomics of fungal disease resistance in tomato. Curr Genomics 11: 30–39
- Parasuraman P, Pattnaik SS, Busi S, Marraiki N, Elgorban AM, Syed A (2022) Isolation and characterization of plant growth promoting rhizobacteria and their biocontrol efficacy against phytopathogens of tomato (*Solanum lycopersicum* L.). Plant Biosyst 156:164–170
- Parray JA, Jan S, Kamili AN, Qadri RA, Egamberdieva D, Ahmad P (2016) Current perspectives on plant growth-promoting rhizobacteria. J Plant Growth Regul 35:877–902
- Patel PR, Shaikh SS, Sayyed RZ (2018) Modified chrome azurol S method for detection and estimation of siderophores having affinity for metal ions other than iron. Environ Sustain 1: 81–87
- Phichai K (2014) Biological control of tomato leaf blight disease by high cell density culture of antagonistic *Bacillus subtilis*. Khon Kaen Agric J 42(4):106–112
- Pieterse CM, Van Pelt JA, Van Wees S, Ton J, Léon-Kloosterziel KM, Keurentjes JJ, Verhagen BW, Knoester M, Van der Sluis I, Bakker PA, Van Loon LC (2001) Rhizobacteria-mediated induced systemic resistance: triggering, signalling and expression. Eur J Plant Pathol 107:51–61
- Pineda A, Dicke M, Pieterse CM, Pozo MJ (2013) Beneficial microbes in a changing environment: are they always helping plants to deal with insects? Funct Ecol 27:574–586
- Poiroux-Gonord F, Bidet LP, Fanciullino AL, Gautier H, Lauri-Lopez F, Urban L (2010) Health benefits of vitamins and secondary metabolites of fruits and vegetables and prospects to increase their concentrations by agronomic approaches. J Agric Food Chem 58:12065–12082
- Prashar P, Kapoor N, Sachdeva S (2013) Isolation and characterization of *Bacillus* sp. with *in-vitro* antagonistic activity against *Fusarium oxysporum* from rhizosphere of tomato. J Agric Sci Technol 15:1501–1512
- Radzki W, Gutierrez Mañero FJ, Algar E, Lucas García JA, García-Villarco A, Ramos Solano B (2013) Bacterial siderophores efficiently provide iron to iron-starved tomato plants in hydroponics culture. Antonie Van Leeuwenhoek 104:321–330
- Rahman M, Islam T, Jett L, Kotcon J (2021) Biocontrol agent, bio fumigation, and grafting with resistant rootstock suppress soil-borne disease and improve yield of tomato in West Virginia. Crop Prot 145:105630
- Ramyabharathi SA, Raguchander T (2014) Mode of action of *Bacillus subtilis* EPCO16 against tomato fusarium wilt. Biochem Cell Arch 14:47–50
- Rennie WJ, Cokerell V (2006) Seedborne diseases. In: The epidemiology of plant diseases, pp 357–372
- Roy C, Akter N, Sarkar MK, PK Uddin M, Begum N, Zenat E (2019) Control of early blight of tomato caused by *Alternaria solani* and screening of tomato varieties against the pathogen. Open Microbiol J 13:41–50
- Saad OA, Moharram TM, Aly ME, Muqlad RR (2016) Biological control of fungal wilt of tomato by plant growth promoting rhizobacteria and *Trichoderma harzianum*. J Phytopathol Pest Manag 3(3):1–10
- Saeed Q, Xiukang W, Haider FU, Kučerik J, Mumtaz MZ, Holatko J, Naseem M, Kintl A, Ejaz M, Naveed M, Brtnický M (2021) Rhizosphere bacteria in plant growth promotion, biocontrol, and bioremediation of contaminated sites: a comprehensive review of effects and mechanisms. Int J Molecular Sci 22:10529
- Santoyo G, Orozco-Mosqueda MD, Govindappa M (2012) Mechanisms of biocontrol and plant growth-promoting activity in soil bacterial species of *Bacillus* and *Pseudomonas*: a review. Biocontrol Sci Tech 22:855–872
- Saxena A, Sarma BK, Singh HB (2016) Effect of azoxystrobin based fungicides in management of chilli and tomato diseases. Proc Nat Acad Sci India Sect B Biol Sci 86:283–289

- Sehrawat A, Sindhu SS, Glick BR (2022) Hydrogen cyanide production by soil bacteria: biological control of pests and promotion of plant growth in sustainable agriculture. *Pedosphere* 32:15–38
- Shahid M, Khan M (2018) Glyphosate induced toxicity to chickpea plants and stress alleviation by herbicide tolerant phosphate solubilizing *Burkholderia cepacia* PSBB1 carrying multifarious plant growth promoting activities. *3 Biotech* 8:1–7
- Shahid M, Zaidi A, Khan M, Rizvi A, Saif S, Ahmed B (2017) Recent advances in management strategies of vegetable diseases. In: *Microbial strategies for vegetable production*, pp 197–226
- Shahid M, Zaidi A, Ehtram A, Khan MS (2019) In vitro investigation to explore the toxicity of different groups of pesticides for an agronomically important rhizosphere isolate *Azotobacter vinelandii*. *Pesticide Biochem Physiol* 157:33–44
- Shaikh SS, Sayyed RZ (2015) Role of plant growth-promoting rhizobacteria and their formulation in biocontrol of plant diseases. In: *Plantmicrobes' symbiosis: applied facets*. Springer, New Delhi, pp 337–351
- Shanmugam V, Kanoujia N (2011) Biological management of vascular wilt of tomato caused by *Fusarium oxysporum* f. sp. *lycopersici* by plant growth-promoting rhizobacterial mixture. *Biol Control* 57:85–93
- Shuping DS, Eloff JN (2017) The use of plants to protect plants and food against fungal pathogens: a review. *Afr J Tradit Complement Altern Med* 14:120–127
- Singh RP, Jha PN (2016) The multifarious PGPR *Serratia marcescens* CDP-13 augments induced systemic resistance and enhanced salinity tolerance of wheat (*Triticum aestivum* L.). *PLoS One* 20:e0155026
- Singh VK, Singh AK, Kumar A (2017) Disease management of tomato through PGPB: current trends and future perspective. *3 Biotech* 7:1
- Sivasakthi S, Usharani G, Saranraj P (2014) Biocontrol potentiality of plant growth promoting bacteria (PGPR)-*Pseudomonas fluorescens* and *Bacillus subtilis*: a review. *African J Agric Res* 9:1265–1277
- Srikhong P, Lertmongkonthum K, Sowanpreecha R, Rerngsamran P (2018) *Bacillus* sp. strain M10 as a potential biocontrol agent protecting chili pepper and tomato fruits from anthracnose disease caused by *Colletotrichum capsici*. *BioControl* 63:833–842
- Sultana S, Alam S, Karim MM (2021) Screening of siderophore-producing salt-tolerant rhizobacteria suitable for supporting plant growth in saline soils with iron limitation. *J Agric Food Res* 4:100150
- Suresh P, Shanmugaiah V, Rajagopal R, Muthusamy K, Ramamoorthy V (2022) *Pseudomonas fluorescens* VSMKU3054 mediated induced systemic resistance in tomato against *Ralstonia solanacearum*. *Physiol Mol Plant Pathol* 119:101836
- TariqJaveed M, Farooq T, Al-Hazmi AS, Hussain MD, Rehman AU (2021) Role of *Trichoderma* as a biocontrol agent (BCA) of phytoparasitic nematodes and plant growth inducer. *J Inverteb Pathol* 183:107626
- Telang SM (2010) Effect of extracts of various plant parts on seed mycoflora and seed germination of tomato. *Asian Sci* 5:15–18
- Thenmozhi P, Dinakar S (2014) Exopolysaccharides (EPS) mediated induction of systemic resistance (ISR) in *Bacillus-Fusarium oxysporum* f. sp. *Lycopersici* pathosystem in tomato (var. PKM-1). *Int J Curr Microbiol Appl Sci* 3:839–846
- Verma PP, Shelake RM, Das S, Sharma P, Kim JY (2019) Plant growth-promoting rhizobacteria (PGPR) and fungi (PGPF): potential biological control agents of diseases and pests. In: *Microbial interventions in agriculture and environment*. Springer, Singapore, pp 281–311
- Vignesh M, Shankar SR, MubarakAli D, Hari BN (2022) A novel rhizospheric bacterium: *Bacillus velezensis* NKMV-3 as a biocontrol agent against *Alternaria* leaf blight in tomato. *Appl Biochem Biotechnol* 194:1–7
- Villareal RL (2019) *Tomatoes in the tropics*. CRC Press
- Wang HL, Yang GG, zhang Y, Bao YD, Yong HE (2017) Detection of fungal disease on tomato leaves with competitive adaptive reweighted sampling and correlation analysis methods. *Spectrosc Spectr Anal* 37:2115

- Wang Y, Han X, Chen X, Deng Y (2021) Potential harmful of extracellular proteases secreted by *Pseudomonas fluorescens* W3 on milk quality. J Food Proc Preserv 45(3):e15192
- Watterson JC (1986) Diseases, the tomato crops. Atherton and Rudich. Champan and Hall Ltd, New York, pp 461–462
- Weert SD, Bloemberg GV (2007) Rhizosphere competence and the role of root colonization in biocontrol. In: Plant-associated bacteria. Springer, Dordrecht, pp 317–333
- Xue QY, Chen Y, Li SM, Chen LF, Ding GC, Guo DW, Guo JH (2009) Evaluation of the strains of *Acinetobacter* and *Enterobacter* as potential biocontrol agents against Ralstonia wilt of tomato. Biol Control 48:252–258
- Yezli W, Hamini-Kadar N, Zebboudj N, Blondin L, Tharreau D, Kihal M (2019) First report of crown and root rot of tomato caused by *Fusarium equiseti* in Algeria. J Plant Pathol 101:1249
- Youssef SA, Tartoura KA, Abdelraouf GA (2016) Evaluation of *Trichoderma harzianum* and *Serratia proteamaculans* effect on disease suppression, stimulation of ROS-scavenging enzymes and improving tomato growth infected by *Rhizoctonia solani*. Biol Control 100:79–86
- Zhang D, Yu S, Yang Y, Zhang J, Zhao D, Pan Y, Fan S, Yang Z, Zhu J (2020) Antifungal effects of volatiles produced by *Bacillus subtilis* against *Alternaria solani* in potato. Front Microbiol 11:1196
- Zhou L, de Jong A, Kuipers OP (2021) Characterization of the interaction between *Priestia endophytica* FH5 and *Rhizoctonia solani*: biocontrol potential against tomato damping-off. In: Discovery of natural products from bacilli and pseudomonas for biocontrol of plant diseases, p 159
- Zuparov MA, Khakimov AA, Mamiev MS, Allayarov AN (2020) In vitro efficacy testing of fungicides on *Botrytis cinerea* causing graymold of tomato. Int J Emerging Technol 11:50