

# Algal Growth Survival Analysis & Event Modeling: A Detailed Handout

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## 1 Goal and Justification

The purpose of this handout is to compare the time it takes for algal cultures under three different treatments to reach a specific growth milestone (an absorbance threshold). Survival analysis is used because it correctly

handles cultures that never reach the threshold (right-censoring) and provides robust, interpretable comparisons of growth rates over time.

### 1.1 Why Survival Analysis is an important way to go!

In simple terms, survival analysis focuses on speed. Instead of just looking at OD at a single time point, we look at the time-to-event.

- The Event: Our chosen growth milestone (e.g.,  $OD \geq 0.6$ ).
- The Time: The hours it took to hit that milestone.
- The Magic (Censoring): If a culture is too slow and never reaches the threshold by the end of the 72-hour experiment, we don't throw it out. We simply record that the event did not occur ( $status = 0$ ) by the time of the last observation. This avoids bias, giving us an honest assessment of growth rates.

Survival Term	Algae Equivalent	Statistical Role
Event	Reaching OD threshold (e.g., $OD = 0.6$ )	Defines the failure point we are tracking
Time	Hours until $OD \geq$ threshold	The variable we are modeling
Censoring	Not reaching threshold by experiment end	Allows us to use incomplete data
Hazard Ratio	Relative rate of reaching threshold	$HR > 1$ means faster growth (speed)

## 2 Packages and Data Setup

We first load the required packages. These provide tools for data manipulation (**tidyverse**), survival analysis (**survival**), visualization (**survminer**), and tidying model output (**broom**).

```
library(tidyverse)

## -- Attaching core tidyverse packages -----
tidyverse 2.0.0 --
## v dplyr     1.1.4     v readr     2.1.5
## v forcats   1.0.0     v stringr   1.5.1
```

```

## v ggplot2     3.4.4      v tibble      3.2.1
## v lubridate   1.9.3      v tidyverse   1.3.1
## v purrr       1.0.2
## -- Conflicts ----- tidyverse_conflicts()
-- 
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>)
# to force all conflicts to become errors

library(survival)
library(survminer)

## Loading required package: ggpubr
##
## Attaching package: 'survminer'
##
## The following object is masked from 'package:survival':
## 
##     myeloma

library(broom)

```

## 2.1 Data Simulation

We simulate growth data for three treatments. The function `make_growth` generates noisy logistic growth curves.

```

set.seed(2025)
n_rep <- 10
times <- seq(0, 72, by = 8)

make_growth <- function(n, treatment_label, mu_time_to_mid = 30, sd_time = 6, maxOD =
  tibble(sample = paste0(treatment_label, "_", seq_len(n))) %>%
  rowwise() %>%
  mutate(
    t_mid = rnorm(1, mu_time_to_mid, sd_time) |> pmax(6),
    slope = (maxOD) / (t_mid + 0.1),
    never = runif(1) < prop_no_reach
  ) %>%

```

```

ungroup() %>%
  expand_grid(time = times) %>%
  rowwise() %>%
  mutate(
    mu = plogis((time - rnorm(1, mu_time_to_mid, sd_time))/7) * maxOD,
    absorbance = mu + rnorm(1, 0, 0.03)
  ) %>%
  ungroup() %>%
  mutate(treatment = treatment_label)
}

# --- ENSURE THESE ADJUSTED VALUES ARE USED ---
# Slower growth for A and B to create failures
df_A <- make_growth(n_rep, "A", mu_time_to_mid = 45, sd_time = 6, maxOD = 1.1, prop_n...
df_B <- make_growth(n_rep, "B", mu_time_to_mid = 40, sd_time = 7, maxOD = 1.15, prop_n...
df_C <- make_growth(n_rep, "C", mu_time_to_mid = 22, sd_time = 5, maxOD = 1.0, prop_n...

df_raw <- bind_rows(df_A, df_B, df_C) %>%
  mutate(treatment = factor(treatment))

```

### 3 Visualization and Event Definition

#### 3.1 Raw Growth Curves and Threshold

We plot individual growth curves and treatment means. This shows variability and average trends. The dashed red line marks our target OD = 0.6 threshold.

```

df_means <- df_raw %>%
  group_by(treatment, time) %>%
  summarise(mean_abs = mean(absorbance), .groups = "drop")

threshold <- 0.6

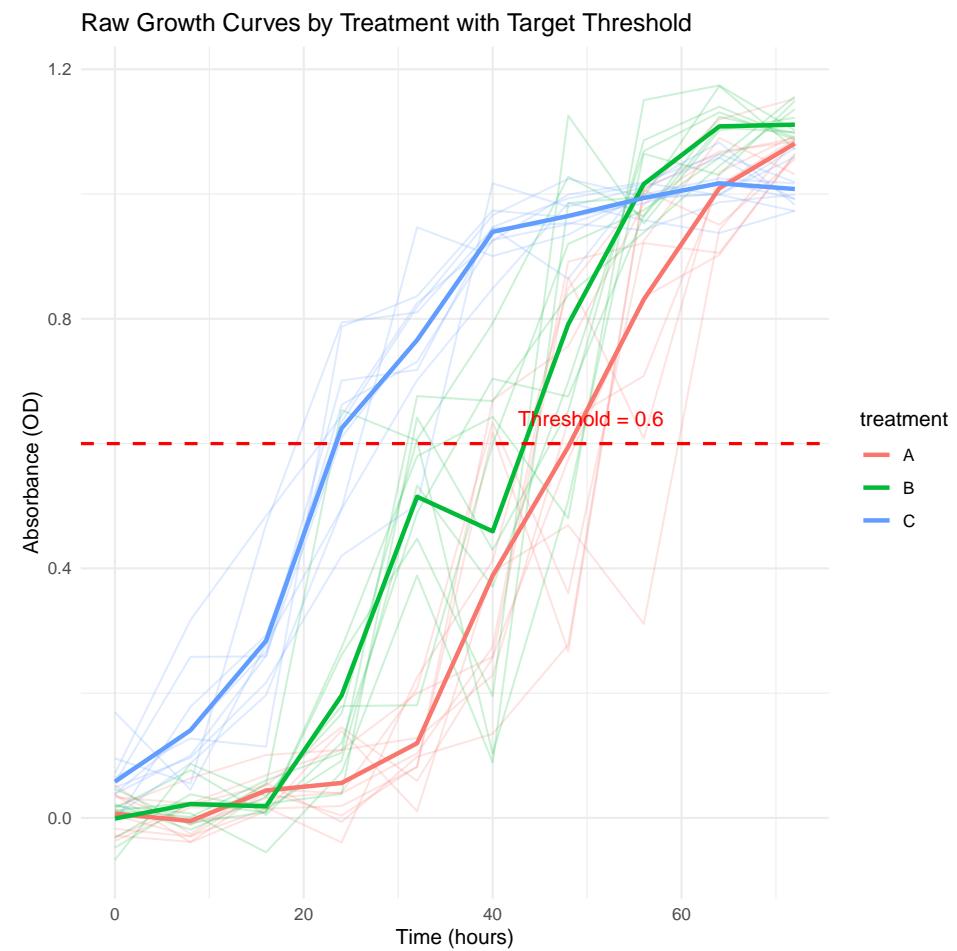
p_raw <- ggplot() +
  geom_line(data = df_raw,
            aes(time, absorbance, group = sample, color = treatment),
            alpha = 0.2, show.legend = FALSE) +

```

```

geom_line(data = df_means,
          aes(time, mean_abs, color = treatment),
          linewidth = 1.1) +
geom_hline(yintercept = threshold, linetype = "dashed", color = "red", linewidth = 1) +
annotate("text", x = max(df_raw$time)*0.7, y = threshold + 0.04, label = paste0("Threshold = ", threshold),
         color = "red", fontface = "bold") +
labs(x = "Time (hours)",
      y = "Absorbance (OD)",
      title = "Raw Growth Curves by Treatment with Target Threshold") +
theme_minimal()

```



## 4 Analysis 1: Threshold Crossing (Speed Analysis)

### 4.1 Defining the Event (Survival Data Setup)

We define the event as the first time a culture reaches  $OD \geq 0.6$ . If a culture never reaches this threshold by the end of the growth experiment (72 hours), it is censored at  $t = 72$ .

```
time_to_event <- df_raw %>%
  group_by(sample, treatment) %>%
  arrange(time) %>%
  summarise(
    event_time = {
      hit_rows <- which(absorbance >= threshold)
      if(length(hit_rows) == 0) NA_real_ else time[min(hit_rows)]
    },
    last_time = max(time),
    .groups = "drop"
  ) %>%
  mutate(
    status = if_else(is.na(event_time), 0L, 1L), # 1=Event Occurred, 0=Censored
    time = if_else(is.na(event_time), last_time, event_time)
  )

cat("Censoring Status (0 = Censored, 1 = Event Occurred):\n")

## Censoring Status (0 = Censored, 1 = Event Occurred):

print(time_to_event %>% count(treatment, status))

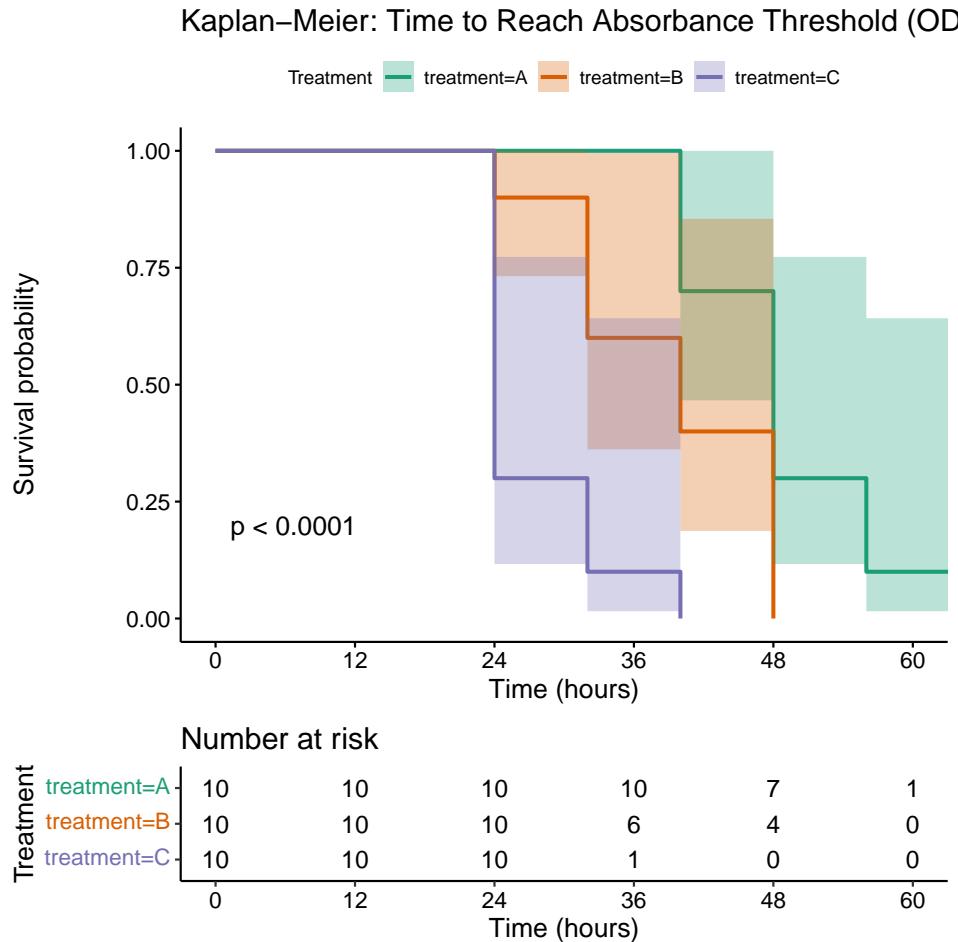
## # A tibble: 3 x 3
##   treatment status     n
##   <fct>     <int> <int>
## 1 A          1     10
## 2 B          1     10
## 3 C          1     10
```

## 4.2 Kaplan-Meier Curves: Visualizing Speed

Kaplan-Meier curves estimate the probability of not yet reaching the threshold over time. Steeper, earlier drops mean faster growth. We use ‘surv.median.line’ = “none” to avoid the interpolation error encountered earlier.

```
km_fit <- survfit(Surv(time, status) ~ treatment, data = time_to_event)

ggsurvplot(km_fit, data = time_to_event, risk.table = TRUE, pval = TRUE,
            conf.int = TRUE, palette = "Dark2",
            surv.median.line = "none",
            title = "Kaplan-Meier: Time to Reach Absorbance Threshold (OD 0.6)",
            xlab = "Time (hours)", legend.title = "Treatment",
            break.time.by = 12)
```



### 4.3 Cox Proportional Hazards Model: Quantifying Speed

The Cox model estimates Hazard Ratios (HR), which quantify the relative speed of hitting the threshold. We use Treatment A as the reference group.

- $\text{HR} > 1$ : Treatment reaches the threshold faster than A.
- $\text{HR} < 1$ : Treatment reaches the threshold slower than A.

```
time_to_event <- time_to_event %>% mutate(treatment = relevel(treatment, ref = "A"))
cox1 <- coxph(Surv(time, status) ~ treatment, data = time_to_event)
```

```

summary(cox1)

## Call:
## coxph(formula = Surv(time, status) ~ treatment, data = time_to_event)
##
##    n= 30, number of events= 30
##
##          coef exp(coef) se(coef)     z Pr(>|z|)
## treatmentB 1.0199    2.7729   0.5053 2.019   0.0435 *
## treatmentC 2.7386   15.4646   0.6127 4.470 7.83e-06 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
##          exp(coef) exp(-coef) lower .95 upper .95
## treatmentB    2.773    0.36063    1.030     7.465
## treatmentC   15.465    0.06466    4.654    51.387
##
## Concordance= 0.817  (se = 0.032 )
## Likelihood ratio test= 21.02  on 2 df,  p=3e-05
## Wald test          = 20.34  on 2 df,  p=4e-05
## Score (logrank) test = 26.43  on 2 df,  p=2e-06

```

#### 4.4 Hazard Ratio Forest Plot

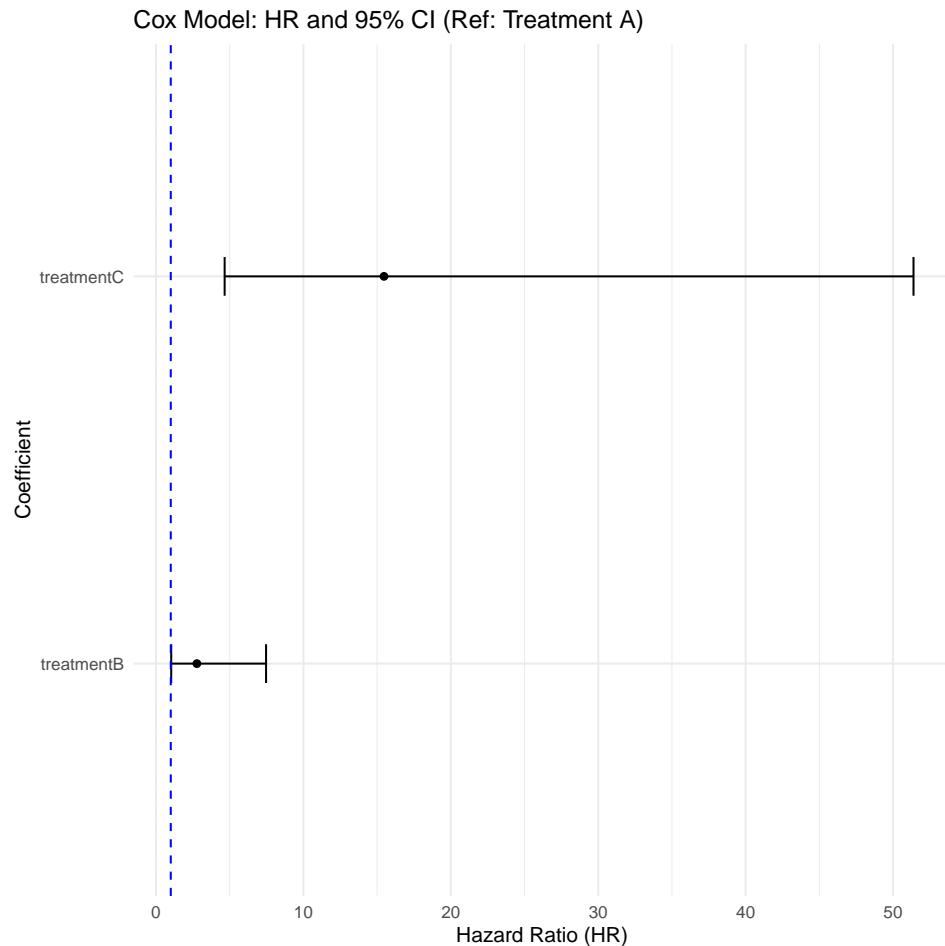
This plot visualizes the HR estimates and confidence intervals (CI). If the CI for an HR does not cross 1 (the blue dashed line), the difference in speed is statistically significant.

```

hr <- tidy(cox1, exponentiate = TRUE, conf.int = TRUE)

ggplot(hr, aes(x = term, y = estimate)) +
  geom_point() +
  geom_errorbar(aes(ymin = conf.low, ymax = conf.high), width = 0.1) +
  geom_hline(yintercept = 1, linetype = "dashed", color = "blue") +
  coord_flip() +
  labs(y = "Hazard Ratio (HR)", x = "Coefficient", title = "Cox Model: HR and 95% CI")

```



#### 4.5 Assumptions Check: Proportional Hazards

The Cox model assumes the HR is constant over time. A non-significant p-value ( $p > 0.05$ ) from the Schoenfeld residuals test indicates the assumption holds.

```
ph_test <- cox.zph(cox1)
ph_test

##          chisq df      p
## treatment  1.74  2 0.42
## GLOBAL    1.74  2 0.42
```

---

## 5 Analysis 2: End of Event (Success Probability Analysis)

In this alternative analysis, we ignore when the event happened and only ask: Did the culture succeed in reaching  $OD \geq 0.6$  by the time the experiment ended (72 hours)?

This shifts our focus from speed to final probability of success. Since the outcome is binary (Success=1 or Failure=0), we use Logistic Regression.

### 5.1 Event Definition (Logistic Data Setup)

We filter the data to the final time point and create a binary success variable.

```
final_data <- df_raw %>%
  filter(time == max(time)) %>%
  mutate(
    success = if_else(absorbance >= threshold, 1, 0),
    treatment = relevel(treatment, ref = "A")
  )

cat("Success/Failure Status at 72 Hours:\n")

## Success/Failure Status at 72 Hours:

print(final_data %>% count(treatment, success))

## # A tibble: 3 x 3
##   treatment success     n
##   <fct>      <dbl> <int>
## 1 A          1     10
## 2 B          1     10
## 3 C          1     10
```

### 5.2 Logistic Regression Model

The model estimates Odds Ratios (OR), which quantify the relative odds of a culture achieving  $OD \geq 0.6$  at the end of the experiment compared to the reference group (Treatment A).

OR > 1: Treatment has higher odds of success than A. OR < 1: Treatment has lower odds of success than A.

```
logis_model <- glm(success ~ treatment, data = final_data, family = "binomial")
summary(logis_model)

##
## Call:
## glm(formula = success ~ treatment, family = "binomial", data = final_data)
##
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## 3.971e-06 3.971e-06 3.971e-06 3.971e-06 3.971e-06
##
## Coefficients:
##             Estimate Std. Error z value Pr(>|z|)
## (Intercept) 2.557e+01 6.831e+04     0      1
## treatmentB -1.759e-09 9.660e+04     0      1
## treatmentC -1.759e-09 9.660e+04     0      1
##
## (Dispersion parameter for binomial family taken to be 1)
##
## Null deviance: 0.000e+00 on 29 degrees of freedom
## Residual deviance: 4.731e-10 on 27 degrees of freedom
## AIC: 6
##
## Number of Fisher Scoring iterations: 24
```

### 5.3 Odds Ratio Plot

We visualize the OR estimates and their confidence intervals. If the CI does not cross 1, the difference in the odds of success is statistically significant.

```
or_results <- tidy(logis_model, exponentiate = TRUE, conf.int = TRUE) %>%
  filter(term != "(Intercept)") # Remove the intercept for the plot

## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
```



```
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning in regularize.values(x, y, ties, missing(ties), na.rm
= na.rm): collapsing to unique 'x' values
## Error in approx(sp$y, sp$x, xout = cutoff): need at least two
non-NA values to interpolate

ggplot(or_results, aes(x = term, y = estimate)) +
  geom_point() +
  geom_errorbar(aes(ymin = conf.low, ymax = conf.high), width = 0.1) +
  geom_hline(yintercept = 1, linetype = "dashed", color = "blue") +
  coord_flip() +
  labs(y = "Odds Ratio (OR)", x = "Coefficient", title = "Logistic Model: Odds of S")
  theme_minimal()

## Error in ggplot(or_results, aes(x = term, y = estimate)): object
'or_results' not found
```