

# Group 2: Bokashi/Compost Bioremediation

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2025-04-27

## Introduction

This document performs a Before-After-Control-Impact (BACI) analysis using simulated data. The goal is to detect whether an environmental impact causes a change relative to a control site.

BACI analysis, which stands for Before-After, Control-Impact, is a research design used to assess the effects of a treatment or intervention on a population or environment. In R, it can be performed using various statistical models and packages. The core principle is to compare a “control” group (impact group) with an “impact” group (intervention group) both before and after the treatment.

Here’s a more detailed explanation of BACI analysis and how it can be implemented in R:

### Understanding BACI Design:

1. **Before-After:** This refers to the time periods before and after the intervention or treatment. The “before” period serves as a baseline for comparison, while the “after” period allows researchers to observe any changes that may have occurred due to the intervention.
2. **Control-Impact:** The “control” group is a reference group that is not subjected to the intervention, while the “impact” group is exposed to the treatment or intervention. This allows researchers to assess whether any observed changes in the impact group can be attributed to the intervention rather than other external factors.

### Statistical Modeling in R:

- **t.test** We can test the difference between the means of two groups (e.g., before and after) using a t-test. This is a simple approach but may not be suitable for all data types or distributions – but we will be testing the “delta” or change between treatments. Probably the simplest to run!
- **Simple Linear Models** (See Hypothesis w/ Fake Data): For simple BACI analyses, a linear model can be used to assess the differences between groups and time periods. The model can include interaction terms to test for differences in slopes or intercepts between the control and impact groups.
- **Generalized Linear Models (GLMs)**: For many BACI analyses, a GLM is a suitable model to fit the data. The model can incorporate factors like “before/after” and “control/impact” as independent variables, and the dependent variable will be the measured outcome of interest.
- **Mixed-Effects Models**: If there are hierarchical structures in your data (e.g., multiple sites, replicates, etc.), a mixed-effects model may be more appropriate. These models account for random effects, such as site-specific variability, that can affect the outcome.

## Some Complex Approaches

```
# Load necessary packages
library(lme4) # For mixed models
```

### Create experimental data of random values

```
## Loading required package: Matrix
```

```
# Example data (replace with your own data)
```

```
data <- data.frame(
  site = factor(rep(c("Control", "Impact"), each = 20)),
  time = c(factor(rep(c("Before", "After"), each = 10)), factor(rep(c("Before", "After"), each = 10))),
  outcome = c(rnorm(10, mean = 6, sd = 2), rnorm(10, mean = 4, sd = 2),
              rnorm(10, mean = 5, sd = 2), rnorm(10, mean = 3, sd = 2)) # Example outcome variable
)
```

```
# Fit a GLM (can be adjusted based on your data)
model <- glm(outcome ~ site * time, data = data)
summary(model)
```

### Using GLM

```
##
## Call:
## glm(formula = outcome ~ site * time, data = data)
##
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## -4.3505  -1.2596   0.0505   0.9889   3.3567
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      4.4172     0.5596   7.893 2.29e-09 ***
## siteImpact       -0.7732     0.7914  -0.977  0.3351
## timeBefore        1.7320     0.7914   2.188  0.0352 *
## siteImpact:timeBefore -1.2252     1.1192  -1.095  0.2809
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for gaussian family taken to be 3.131754)
##
##      Null deviance: 148.23  on 39  degrees of freedom
## Residual deviance: 112.74  on 36  degrees of freedom
## AIC: 164.96
##
## Number of Fisher Scoring iterations: 2
```

```
# Fit a mixed model (if you have nested data)
# Example data with site as a random effect
data$conc <- factor(rep(rep(c("Low", "High"), each = 5), 4))
model_mixed <- lmer(outcome ~ site * time + (1 | conc), data = data)
```

## Mixed-Effects Model

```
## boundary (singular) fit: see help('isSingular')
null_model <- lmer(outcome ~ (1 | conc), data = data)

## boundary (singular) fit: see help('isSingular')
summary(model_mixed)

## Linear mixed model fit by REML ['lmerMod']
## Formula: outcome ~ site * time + (1 | conc)
## Data: data
##
## REML criterion at convergence: 152.5
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -2.45835 -0.71175  0.02856  0.55879  1.89681
##
## Random effects:
## Groups Name Variance Std.Dev.
## conc (Intercept) 0.000 0.00
## Residual 3.132 1.77
## Number of obs: 40, groups: conc, 2
##
## Fixed effects:
##              Estimate Std. Error t value
## (Intercept)      4.4172    0.5596  7.893
## siteImpact       -0.7732    0.7914 -0.977
## timeBefore        1.7320    0.7914  2.188
## siteImpact:timeBefore -1.2252    1.1192 -1.095
##
## Correlation of Fixed Effects:
##              (Intr) stImpc timBfr
## siteImpact  -0.707
## timeBefore  -0.707  0.500
## stImpct:tmB  0.500 -0.707 -0.707
## optimizer (nloptwrap) convergence code: 0 (OK)
## boundary (singular) fit: see help('isSingular')
anova(model_mixed, null_model)

## refitting model(s) with ML (instead of REML)

## Warning in optwrap(optimizer, devfun, x@theta, lower = x@lower, calc.derivs =
## TRUE, : convergence code 3 from bobyqa: bobyqa -- a trust region step failed to
## reduce q

## Data: data
## Models:
## null_model: outcome ~ (1 | conc)
## model_mixed: outcome ~ site * time + (1 | conc)
##
##      npar    AIC    BIC logLik deviance Chisq Df Pr(>Chisq)
## null_model      3 171.91 176.98 -82.955   165.91
## model_mixed      6 166.96 177.10 -77.482   154.96 10.946  3 0.01202 *
```

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## Group Hypotheses

Bokashi will be a more efficient method to reduce pesticide concentrations in decomposed green waste.

-The bokashi samples will see a greater decrease in absorption level compared to the control group and traditional composting group samples after 2 weeks of breakdown

NOTE: No one cares about absorbance, that's a proxy for concentration; please revise to test concentration. . .

## Data Set

```
# Load the data
fakedata.csv = "/home/mwl04747/RTricks/00_Project_Group_Demos/Group2_FakeData.csv"
group1data = read.csv(fakedata.csv, header = TRUE)
str(group1data)

## 'data.frame':   54 obs. of  5 variables:
## $ Sample.ID      : chr  "B1ZB" "B2ZB" "B3ZB" "B1FB" ...
## $ Treatment      : chr  "Bokashi" "Bokashi" "Bokashi" "Bokashi" ...
## $ Before.After   : chr  "Before" "Before" "Before" "Before" ...
## $ Concentraion..ppm.: int  0 0 0 5 5 5 10 10 10 0 ...
## $ Absorption     : num  0.045 0.021 0.034 0.241 0.223 0.234 0.461 0.452 0.443 0.056 ...

names(group1data)

## [1] "Sample.ID"      "Treatment"      "Before.After"
## [4] "Concentraion..ppm." "Absorption"

unique(group1data$Treatment)

## [1] "Bokashi"      "Compost"      "Water (control)"

unique(group1data$Before.After)

## [1] "Before" "After"
```

NOTE: Misspelled concentration. . . suggest you come up with one word column names and factors () (## Removing Water – since that is a correction, not a factor. I suggest this is Concentration\_Initial and then you can have Concentration\_Final as a column.

Really, intial will be use to “correct” final values; perhaps we want to test the difference between the two, but I think we want to test the difference between the bokashi and compost treatments.

```
# Remove water
group1data <- group1data %>%
  filter(Treatment != "Water (control)")
# Remove the "Water" treatment

# Check the data
unique(group1data$Treatment)

## [1] "Bokashi" "Compost"
```

Pretty sure zero concentration (control) is also not a treatment but will be used as a correction factor... please correct fake data to preprocess that. Or we can use R if that would be something you'd like to do... probably take us an hour or so of working together to do that. But it excel it could take 10 min.

```
# Make sure site and time are factors
group1data <- group1data %>%
  mutate(Treatment = factor(Treatment),
         Before.After = factor(Before.After),
         Concentraion..ppm. = factor(Concentraion..ppm.))

# Check the data
str(group1data)

## 'data.frame': 36 obs. of 5 variables:
## $ Sample.ID : chr "B1ZB" "B2ZB" "B3ZB" "B1FB" ...
## $ Treatment : Factor w/ 2 levels "Bokashi","Compost": 1 1 1 1 1 1 1 1 1 2 ...
## $ Before.After : Factor w/ 2 levels "After","Before": 2 2 2 2 2 2 2 2 2 2 ...
## $ Concentraion..ppm.: Factor w/ 3 levels "0","5","10": 1 1 1 2 2 2 3 3 3 1 ...
## $ Absorption : num 0.045 0.021 0.034 0.241 0.223 0.234 0.461 0.452 0.443 0.056 ...

group1data[sample(1:nrow(group1data), 8), ]

## Sample.ID Treatment Before.After Concentraion..ppm. Absorption
## 30 C3ZA Compost After 0 0.045
## 6 B3FB Bokashi Before 5 0.234
## 8 B2TB Bokashi Before 10 0.452
## 22 B1FA Bokashi After 5 0.153
## 33 C3FA Compost After 5 0.231
## 7 B1TB Bokashi Before 10 0.461
## 16 C1TB Compost Before 10 0.563
## 17 C2TB Compost Before 10 0.554
```

## Summary Stats

Fake data isn't really working – since you don't have any variance with ppm. We are not going reporting absorbance values, but I'll do this now, since it looks like there is some variance there.

```
# Summarize the data
summary_stats <- group1data %>%
  group_by(Treatment, Before.After, Concentraion..ppm.) %>%
  summarise(
    mean = mean(Absorption),
    sd = sd(Absorption),
    n = n()
  ) %>%
  ungroup()

## `summarise()` has grouped output by 'Treatment', 'Before.After'. You can
## override using the `.groups` argument.

summary_stats

## # A tibble: 12 x 6
## Treatment Before.After Concentraion..ppm. mean sd n
## <fct> <fct> <fct> <dbl> <dbl> <int>
## 1 Bokashi After 0 0.04 0.0135 3
```

##	2	Bokashi	After	5	0.142	0.0263	3
##	3	Bokashi	After	10	0.146	0.0108	3
##	4	Bokashi	Before	0	0.0333	0.0120	3
##	5	Bokashi	Before	5	0.233	0.00907	3
##	6	Bokashi	Before	10	0.452	0.00900	3
##	7	Compost	After	0	0.0693	0.0215	3
##	8	Compost	After	5	0.249	0.0157	3
##	9	Compost	After	10	0.329	0.0116	3
##	10	Compost	Before	0	0.056	0.011	3
##	11	Compost	Before	5	0.347	0.0286	3
##	12	Compost	Before	10	0.552	0.0121	3

## Hypothesis Tests w/Fake Data

```
# Fit a linear model
model <- lm(Absorption ~ Treatment + Concentraion..ppm. * Before.After, data = group1data)
# Summarize the model
anova(model)
```

```
## Analysis of Variance Table
##
## Response: Absorption
##
##              Df Sum Sq Mean Sq F value    Pr(>F)
## Treatment      1  0.07719  0.077191   67.876 4.396e-09 ***
## Concentraion..ppm.  2  0.62341  0.311705  274.088 < 2.2e-16 ***
## Before.After      1  0.12192  0.121917  107.204 2.980e-11 ***
## Concentraion..ppm.:Before.After  2  0.11515  0.057574   50.626 3.470e-10 ***
## Residuals      29  0.03298  0.001137
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
model_summary <- tidy(model)
model_summary
```

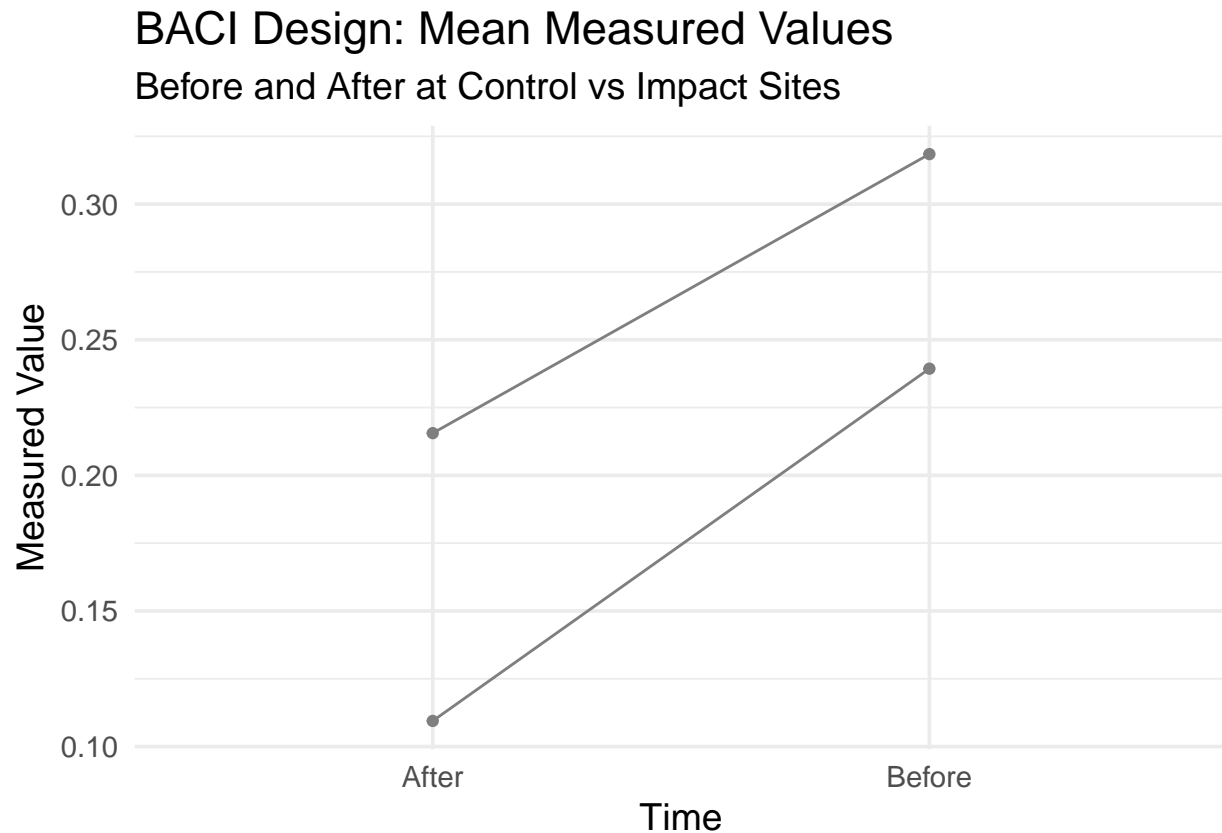
```
## # A tibble: 7 x 5
##   term                                estimate std.error statistic  p.value
##   <chr>                                <dbl>     <dbl>     <dbl>    <dbl>
## 1 (Intercept)                        0.00836    0.0149     0.562 5.78e- 1
## 2 TreatmentCompost                    0.0926    0.0112     8.24  4.40e- 9
## 3 Concentraion..ppm.5                  0.141     0.0195     7.22  5.91e- 8
## 4 Concentraion..ppm.10                 0.183     0.0195     9.39  2.69e-10
## 5 Before.AfterBefore                   -0.0100    0.0195    -0.514 6.11e- 1
## 6 Concentraion..ppm.5:Before.AfterBefore  0.105     0.0275     3.80  6.84e- 4
## 7 Concentraion..ppm.10:Before.AfterBefore 0.274     0.0275     9.97  7.08e-11
```

```
# Key interaction term:
key_interaction <- coef(model)["Treatmentimpact:Before.Afterafter"]
```

## Plots

```
ggplot(group1data, aes(x = Before.After, y = Absorption, color = Treatment, group = Treatment)) +
  stat_summary(fun = mean, geom = "line") +
  stat_summary(fun = mean, geom = "point") +
```

```
labs(
  title = "BACI Design: Mean Measured Values",
  subtitle = "Before and After at Control vs Impact Sites",
  y = "Measured Value",
  x = "Time",
  color = "Treatment"
) +
scale_color_manual(values = c("control" = "#1f77b4", "impact" = "#d62728")) +
theme_minimal(base_size = 14) +
theme(legend.position = "bottom")
```



5. Conclusion The analysis suggests:

There was a decrease in the measured value at the impact site after the event.

The significant site  $\times$  time interaction supports a likely effect of the environmental disturbance.

6. Appendix (Optional) You can improve the model by considering:

Mixed models (lmer) if you have random effects (e.g., multiple sites)

Repeated measures ANOVA

Adding covariates (e.g., weather, seasonality)