

	Environmental Analysis Teaching and Research Laboratory	Date: 6/12/2022	Number: 23.02
	Standard Operating Procedure	Title: IDEXX Quanti-Tray DRAFT!	
	Approved By: TBD	Revision Date: June 16, 2022	

1. Scope and Application

1.1 This standard operating procedure describes the test method for the collection and analysis of water samples for the enumeration of *Escherichia coli* (*E. coli*) and Total coliform bacteria.

1.2 The applications of this SOP are for...

2. Summary of Method

2.1 Surface water samples are collected in 120ml shrink-banded, sterile IDEXX bottles. An undiluted water sample will be analyzed from the sample collected. The Colilert reagent is added directly to the 100 ml undiluted sample. Both are mixed thoroughly to dissolve the reagent. The sample is transferred to QuantiTrays/2000 and sealed using the Quanti-Tray sealer. Samples are incubated at 35.0 ± 0.5 C for 24 hours. Results are reported as MPN/100mL.

2.2 The detection limit for this analysis is 1 Most Probable Number (MPN) per 100mL of sample.

Contents

1	Scope and Application	1
2	Summary of Method	1
3	Acknowledgements	3
4	Definitions	3
5	Biases and Interferences	3
6	Health and Safety	3
	Safety and Personnel Protective Equipment	4
7	Personnel & Training Responsibilities	4
8	Required Materials and Apparati	4

9 Reagents and Standards	4
10 Estimated Time	4
11 Sample Collection, Preservation, and Storage	5
12 Procedure	5
QA/QC	8
Disposal	8
13 Data Analysis and Calculations	9
14 QC/QA Criteria	9
Data assessment and acceptable criteria	9
Corrective actions for out-of-control or unacceptable data	10
15 Trouble Shooting	10
16 References	10

3. Acknowledgements

4. Definitions

- 4.1** Analytical batch: The set of samples processed at the same time
- 4.2** Control cultures: For each lot of medium, check analytical procedures by testing with known positive and negative control cultures. For example, E.coli is a positive control for this analysis and Staphylococcus aureus is a negative control.
- 4.3** Field duplicate (FD): Two samples taken at the same time and place under identical circumstances and that are treated identically throughout field and laboratory procedures. Analysis of field duplicates indicates the precision associated with sample collection, preservation, and storage as well as laboratory procedures.
- 4.4** Laboratory reagent blank (LRB): An aliquot of sterilized water treated as a sample in all aspects, except that it is not taken to the sampling site. The purpose is to determine if the analytes or interferences are present in the laboratory environment, the reagents, or the apparatus.
- 4.5** Laboratory duplicate (LD): Two aliquots of the same environmental sample treated identically throughout a laboratory analytical procedure. Analysis of laboratory duplicates indicates precision associated with laboratory procedures but not with sample collection, preservation or storage procedures.

5. Biases and Interferences

- 5.1** Water samples containing humic or other material may be colored. If there is background color, compare inoculated trays to a control tray containing only water (SM, 9223 A.)

6. Health and Safety

- 6.1** The analysis involves handling of freshwater samples that may contain live microorganisms and therefore pose some threat of infection. Laboratory personnel who are routinely exposed to such water samples are encouraged to protect themselves from water borne illnesses by wearing clean disposable gloves and washing their hands frequently.
- 6.2** The Colilert reagent is not hazardous according to the manufacturers material safety data sheet. The manufacturer does recommend wearing gloves and safety glasses while using this reagent and washing hands after use.

Safety and Personnel Protective Equipment

Water quality can range from high quality and even potable to very low with assorted pathogens or even toxicity. Thus, it's important to take every precaution possible to protect yourself.

7. Personnel & Training Responsibilities

7.1 Laboratory and field personnel shall have a working knowledge of this analytical procedure and will have received training from a faculty, staff, or student knowledgeable of the proper sample analysis procedures.

7.2 Researchers training is required before the procedures in this method can be used.

7.3 Researchers using this SOP should be trained for the following SOPs:

- SOP01 Laboratory Safety
- SOP02 Field Safety
- SOP23 IDEXX Quanti-Tray

8. Required Materials and Apparatus

8.1 10.1 Sterile, shrink-wrapped 100ml IDEXX bottles.

8.2 Quanti-Tray Sealer: catalog number XXXX? . IDEXX Laboratories, Inc., Westbrook, ME (NEED TO ADD SERIAL NUMBER).

8.3 Incubator (add model and serial number)

9. Reagents and Standards

9.1 ... need to add

?? Colilert reagent: for 100 ml samples, catalog number WP200. IDEXX Laboratories, Inc., Westbrook, ME.

9.2 ?? Quanti-Tray/2000: 100 trays containing 97 wells each, part number WQT2K. IDEXX Laboratories, Inc., Westbrook, ME

10. Estimated Time

10.1 This procedure requires XX minutes...

11. Sample Collection, Preservation, and Storage

12. Procedure

NEEDS TO BE UPDATED!!! copied from various methods online... needs to be harmonized.

- a) Good laboratory practices will be used for this procedure.
- b) Pre-heat incubator to bring temperature up to 35C.
- c) Pre-warm rehydration fluid vials to 35-37C. Use blue autoclavable foam vial holder to hold vials.
- d) Discard blue cap from rehydration fluid vial.
- e) Remove organism vial from pouch (vial with colorless cap).
- f) Transfer colorless cap onto pre-warmed rehydration vial and discard vial containing the desiccant.
- g) Place rehydration vials into the foam vial holder. Invert and place in incubator for 10 minutes at 35-37C.
- h) Fill four sterile IDEXX 100 ml. bottles with distilled water to fill line. Label three bottles with each bacteria name and one bottle control. Place in incubator until a temperature of 35 ± 0.5C is reached.
- i) Remove vial from holder. Hold vial upside down and tap cap gently to mix. Remove cap and look at inside surface to ensure that no un-dissolved black particles are present. Inoculate an additional 10 minutes if present.
- j) Add entire contents of each appropriate bacteria vial to pre-warmed 100 ml. labeled bottles.
- k) Add Colilert reagent to sample bottles including control. Place in incubator and follow incubation instructions outlined in section 7. Do not place in QuantiTrays.

The 100ml duplicate water sample is shaken well just prior to preparation for analysis. Samples over the 100 ml mark must not be poured to volume. If there is at least 1 of headspace, the sample may be shaken and excess volume taken out with a sterile pipet. If there is insufficient headspace (1) for proper mixing, do not pour off and discard a portion of the sample. Rather, pour the entire sample into a larger sterile container, mix properly, and proceed with the analysis.

Open a Colilert ampule and pour contents into either the diluted sample or undiluted sample. Repeat for the remaining sample.

Mix thoroughly, making sure the Colilert reagent is completely dissolved.

Follow manufacturers instructions for preparation of QuantiTray/2000 and use of the Quanti-Tray Sealer.

Allow bubbles to settle or dissipate. Failure to do this may result in the wells filling or sealing improperly.

Record the samples site code on the back of the well for identification purposes.

Record the lot number of the reagents and the wells used on the bench sheet in the comments section.

Incubate at 35.0 ± 0.5C for 24 hours.

Count the number of small and large positive wells and refer to the MPN table to find the most probable number for Total coliform.



Figure 1: add caption – pictures from our lab...

E. coli results are obtained by placing the wells under a black light and counting the number of fluorescent wells. Refer to the MPN table to determine the *E. coli* concentration.

Report results on the bench sheet.

The completed bench sheet should be reviewed by the researcher, the laboratory director and the QA manager.

12.1 Arrive at site and record site number, date and time.

12.2 Immerse the thermometer or YSI handheld in the water and leave immersed five minutes before reading temperature. Avoid disturbing the bottom with the thermometer at the sample site.

12.3 Label bottle with location (geographic area name and stream or lake name), date, time, water temperature and sampler's initials. Label bottle before immersion using a black permanent marker or pre-printed labels. QVIR Bacteria Lab, State Certified Lab, purchases only certified sterile, 100 ml, sealed containers from IDEXX.

12.4 Use latex gloves when handling bottles during sampling. Fingers contain contaminants such as nitrates. Bug repellents or sunscreen are particularly troublesome as contaminants. Once the gloves are on, be careful not to touch your face, the ground, or anything but the bottles.

12.5 The sample should be taken from flowing, not stagnant water, facing upstream positioned in the thalweg.

12.6 Be sure to immerse the bottle completely, 10 cm (4 inches) deep, with mouth of bottle pointing upstream, so no water flows over your hand into the bottle. Be sure the bottle does not get near the bottom of the stream where sediments can be disturbed. Water samples should be collected 6-12 inches below the water surface. Fill bottle, to the 100ml line indicated, on first immersion, pour off the excess and



Figure 2: add caption – pictures from our lab...



Figure 3: add caption – pictures from our lab...

cap. Do not under fill or over fill, do not redunk. If too much water is poured off, redo sample with new 100 ml container.

12.7 Do not touch bottle mouth or inside of cap. Be careful not to contaminate the sample with surface film, contact with human skin, breathing in/on the bottle or cap, etc. If stream is too shallow to immerse bottle fully, collect as much as possible, being very careful not to touch the bottom. Note depth on field notes.

12.8 Collect one "duplicate" sample every two weeks (sampling frequency). Sample sites chosen for duplicate sampling are selected at random among sites sampled. When a duplicate sample is selected for the site, repeat procedures as with normal stream samples. The duplicate is the second sample when two samples are collected. Duplicates document repeatability of individual sample collections and reproducibility of laboratory results.

12.9 Samples are analyzed in the Biogeochemistry Lab. Keep samples cool while transporting. Store at 4 C until analysis, but do not freeze. The maximum holding time is 6 hours.

12.10 For each sample, the location number, bottle numbers used and time collected will be recorded in the field sample log.

12.11 The samples will be kept in the possession of researcher personnel who both collect and analyze the samples.

QA/QC

12.12 Accuracy: Initial analyst demonstration of capability and for each new lot of Quanti-Tray/2000, analyze the following:

- Check each new lot of Colilert. Shine the ultraviolet lamp on the media snap packs. If the lot is fluorescent it will be discarded.
- Dissolve one packet in 100 ml distilled water. Do not incubate. Check for fluorescents.
- Analyze sterile reagent water blank with each batch of samples to verify that there is a negative result from 24-28 hours
- Gravimetrically check each new lot of sterile, transparent, nonfluorescing 100-ml vessels to ensure the 100-mL fill line is accurately represented on the vessel.

Disposal

needs updating...

12.13 All materials must be autoclaved prior to disposal and workspaces thoroughly disinfected.

Table 1: need to add caption

Organism	Result
Escherichia coli	Yellow wells, fluorescence
Klebsiella pneumoniae	Yellow wells, no fluorescence
Pseudomonas aeruginosa	Clear wells, no fluorescence
Method Blank	Clear wells, no fluorescence

12.14 Dispose of media in accordance with Good Laboratory Practices. ??? need to figure out what this means!

13. Data Analysis and Calculations

13.1 For each sample analyzed, including quality control samples, record the number of small and large positive wells and the MPN in the appropriate places on the bench sheet (see below). Calculate precision for duplicate analyses using equation 1.

13.2 Equation 1.

$$Precision(asRPD) = \frac{(AB) * 100}{(A + B)/2} \quad (1)$$

Where: A = MPN from aliquot A and B = MPN from aliquot B

13.3 The following results should be observed:

14. QC/QA Criteria

Data assessment and acceptable criteria

14.1 The analyst should review all data for correctness (e.g., use of MPN table).

14.2 Precision values are calculated for pairs of duplicate analyses.

14.3 Record the precision values as RPD on the bench sheet.

14.4 The desired precision is 20

14.5 The desired detection limit is 1 MPN/100mL

14.6 The completed bench sheet is reviewed by the analyst's supervisor or the QVIR Lab Director

Corrective actions for out-of-control or unacceptable data

- 14.7** The results for precision and blank data are compared to the acceptable values for this analysis; 20% and 1 MPN/100mL, respectively.
- 14.8** If a precision value exceeds 20% then the analyst should write in the comments section of the bench sheet: These data are associated with an out-of-control duplicate analysis. The UCL = 20%. Note: UCL is the Upper Control Limit (i.e., 20
- 14.9** If a blank value exceeds 1 MPN/100mL then the analyst should write in the comments section of the bench sheet: These data are associated with a blank value that exceeds the detection limit of 1 MPN/100mL.
- 14.10** The samples cannot be reanalyzed because the sample volume will be depleted after the initial analysis.
- 14.11** If data are unacceptable for any reason, the analyst should review their analytical technique prior to conducting this analysis again.

15. Trouble Shooting

TBD!!!

16. References

- 16.1** APHA, AWWA. WEF. (2012) Standard Methods for examination of water and wastewater. 22nd American Public Health Association (Eds.). Washington. 1360 pp. (2014).