

Xolotl: An Intuitive and Approachable Neuron & Network Simulator in MATLAB

Alec Hoyland^{1†}, Srinivas Gorur-Shandilya^{1†}, and Eve Marder^{1*}

¹ *Volen Center and Biology Department
Brandeis University
Waltham, MA 02454
USA*

† These authors have equally contributed to this article.

Correspondence*:
Srinivas Gorur-Shandilya
Marder Lab
srinivas@brandeis.edu

An Intuitive Neuronal Simulator

ABSTRACT

Information processing by neurons relies on the transmission and interaction of electrical signals that arise from the biophysics of ion channels and synapses. Electrophysiological characterization of these low-level mechanisms have allowed for the construction of conductance-based models that can reproduce many features of neuronal and circuit behavior. However, working with conductance-based models continues to be a challenge due to their high dimensionality, hindering intuition of their dynamical features. Here, we present a neuron and circuit simulator using a novel automatic type system that binds class templates written in C++ to object-oriented code in MATLAB. This approach combines the speed of C++ code with the ease-of-use of scientific programming languages like MATLAB. Neuron models are hierarchical, named and searchable, permitting high-level programmatic control over all parameters. The simulator's architecture allows for the live manipulation of any parameter in any model, and for visualizing the effects of changing that parameter on model behavior. The simulator is fully featured with hundreds of ion channel models from the electrophysiological literature, and can be easily extended to include arbitrary mechanisms and dynamics. Finally, the simulator is written in a modular fashion and has been released under a permissive free software license, enabling it to be integrated easily in third party applications.

Keywords: simulator, MATLAB, C++, conductance-based, neuron, network, pedagogy

1 INTRODUCTION

Nervous systems process and transmit information through electrically-excitable membranes. Conductance-based models are the simplest biophysical representation of an electrically-excitable cell (Hodgkin and Huxley 1952a). Studied based on the Hodgkin-Huxley formalism now contribute significantly to mainstream research in small-circuit networks (E. Marder and Abbott 1995; Astrid A Prinz 2006; Astrid A. Prinz 2010). Additionally, these models provide an approachable framework for understanding salient principles of neuroscience. However, challenges remain in simulating biophysically-realistic neuron models. Conductance-based models are typically high-dimensional with many coupled nonlinear

27 differential equations. Conductances are coupled through the membrane potential, and in multicompart-
28 ment models, all membrane potentials are coupled. Simulators written in languages like C, C++, or hoc
29 integrate equations quickly, but these simulators often lack the ease-of-use and interoperability of those
30 written in scientific programming languages (e.g. Python, Julia, MATLAB).

31 Two major approaches have dominated the design of neuron simulators. Some simulators, such as
32 NEURON (M L Hines and Carnevale 1997) are specified in a compiled language with pre-specified net-
33 work components. These simulators tend to perform very fast computations with little overhead, but suffer
34 from a steep learning curve. Implementations in more approachable languages and use of graphical in-
35 terfaces mitigate these drawbacks, but obfuscate the underlying algorithms and parameters (Brette et al.
36 2007; M Hines, Davison, and Muller 2009). In contrast, many simulators have been designed in popu-
37 lar scientific programming languages to emphasize ease-of-use and flexibility. DynaSim (Sherfey et al.
38 2018), ANNarchy (Vitay, Dinkelbach, and Hamker 2015), BRIAN (Stimberg, D F Goodman, et al. 2013)
39 parse strings of equations that are specified in the scientific programming language. The equations can be
40 translated into a fast implementation language (Stimberg, D F M Goodman, et al. 2014). This approach
41 permits tremendous flexibility for simulating systems of differential equations, but the syntax tends to
42 be verbose and the hierarchical nature of conductance-based models is not generally reflected in the user-
43 generated code. There is no simulator that combines the advantages of both approaches without sacrificing
44 efficiency, ease-of-use, or clarity.

45 We have developed a novel neuronal simulator, `xolotl`, written in C++ with a MATLAB interface. De-
46 signed with an emphasis on ease-of-use, `xolotl` can simulate single-compartment conductance-based
47 models, networks of these, and detailed multi-compartment models. `xolotl` exploits a novel automatic
48 type system, `cppplab`, which binds MATLAB code to C++ header files, creating objects in MATLAB
49 which reflect the underlying object-oriented code. `xolotl` implements `cppplab` to represent the nested
50 structure of conductance-based models, and exploits the computational efficiency of the C++ to quickly
51 integrate models. For this reason, models can be implemented entirely in MATLAB with a few lines of
52 code. The software has been implemented in MATLAB due to its ease-of-use and popularity among neu-
53 roscientists while `cppplab` provides a powerful backend for specifying and integrating models without
54 relying on the significantly slower and limiting MATLAB `codegen`. While automated C++ transpiling
55 using the proprietary `codegen` can drastically improve performance over loops through strong typing
56 and memory pre-allocation, supervenience of MATLAB over C++ prevents efficient use of features not
57 accessible in MATLAB, such as passing by reference and strongly-typed object-oriented programming.
58 Minimal experience with MATLAB is required to use `xolotl`, and all equations and integration methods
59 are provided transparently to the end user. No string parsing of equations is required (Sherfey et al. 2018;
60 Stimberg, D F M Goodman, et al. 2014; Stimberg, D F Goodman, et al. 2013).

61 `xolotl` comes packaged with visualization functions and a graphical user interface (GUI) for real-
62 time manipulation of model parameters. Plotting of voltage, intracellular calcium, conductance gating
63 functions, and time constants is provided by built-in `xolotl` methods. The GUI permits real-time tuning
64 of any network parameters using numerical sliders in a graphical interface which displays the resultant
65 membrane potential and intracellular calcium traces. The ease-of-use of these tools lends them to peda-
66 gogical applications and rapid exploration of toy models. This tool aims to simplify the investigation of
67 dynamics of complex neural network models, facilitate collaborative modeling, and complement other
68 tools being developed in the neuroinformatics community.

2 DESIGN GOALS

69 `xolotl` is designed to be easy-to-use without sacrificing speed.

70 The software has been designed in MATLAB due to its popularity among neuroscientists for pedagogy
71 and research. `xolotl` capitalizes on MATLAB's straightforward structure array syntax to permit rapid
72 prototyping and experimentation, especially for small neuronal networks of complex models. Parameters

of conductances, neuronal compartments, and simulations may all be edited in the structure before any calls to integration functions. The underlying code is written in C++ for speed and memory optimization, and while models can indeed be integrated using the compiled binary, symbolic manipulations can be readily performed in MATLAB without ever touching the foundational code.

2.1 FEATURES

MATLAB provides a high level programmatic and graphical interface for implementing, manipulating, and visualizing models without sacrificing the enhancements of the underlying C++ code.

Modular structure. Models are specified by adding compartments and synapses to the `xolotl` object. Conductances are added to compartments and controllers can be added to conductances. This modular structure recapitulates the biophysics of the Hodgkin-Huxley formalism and obviates the need to explicitly write out equations, which in `xolotl` are contained within the conductance header files.

Interface between C++ and MATLAB. `xolotl` relies on `cpplab` constructions, which allow the user to exploit the efficiency of low-level C++ code. MATLAB treats `cpplab` objects as fully-typed variables allowing for symbolic manipulation using only the high-level programming language and graphical interfaces. `xolotl` is fast because all time-intensive code is written in C++. While automated C++ transpiling from MATLAB using the proprietary `codegen` can drastically improve performance over loops through strong typing and memory pre-allocation, supervenience of MATLAB over C++ prevents efficient use of low-level features, such as passing by reference and object-oriented programming. C++ provides speed improvements beyond the benefits of translating MATLAB features into low-level code. For this reason, `cpplab` has been designed to provide an interface for constructing, transpiling, and compiling C++ code to be called from within MATLAB. `xolotl` simulations are run entirely from C++ executables.

Automatic and efficient compiling. `xolotl` automatically handles transpiling and compiling MATLAB code into C++. The MD5 algorithm hashes the network to compile a new binary and MEX bridge file only if needed and to confirm that the correct binary fetched during execution.

2.2 SYNTAX

MATLAB can easily control the `cpplab` objects using the standard, flexible data structure notation popular in high-level scripting languages.

Adding features. The `add` function creates a compartment, conductance, or controller and affixes it as a field in the `xolotl` network structure. This function generates a MATLAB `struct` that faithfully represents the underlying C++ code. Compartments add to the `xolotl` object and conductances add to compartments. Specific properties can be specified using key-value pair arguments (e.g. Figure 1A).

Finding features. `cpplab` comes with several features which simplify the handling of complex, nested models. The `find` function acquires a cell array of all properties of the network which satisfy a search condition. For example, one can find all paths to maximal conductances within the 'HH' compartment by:

```
x.find('HH*gbar');
```

To extract a vector of the maximal conductances:

```
gbars = x.get('HH*gbar');
```

To set the maximal conductances all at once:

```
x.set('HH*gbar', gbars)
```

111 *Compartments.* A model neuron consists of one or more compartments, each representing a section of
112 membrane with capacitance and surface area. Isopotential models require one compartment, whereas
113 models with multiple neurons, units, or non-trivial morphology require multiple compartments.

114 *Synapses.* `xolotl` provides some features for generating complex models. Synapses can be added with
115 the `connect` function. Synapses possess identifiers to presynaptic and postsynaptic compartments and
116 default to electrical synapses. All specifiable properties of synapses are shown in Table ???. To create
117 axons or transport chains, the `slice` function splits a compartment into n discrete segments and adds
118 these compartments to the network connected by electrical synapses.

119 *Conductances and controllers.* All conductances contain fields for maximal conductance and reversal
120 potential. Conductances with activation and inactivation variables include them as m and h respectively.
121 Gating functions and their respective time constants are contained within the conductance header file.
122 `xolotl` comes packaged with conductances from several dozen papers (Dethier et al. 2015).

123 *Creating custom `cpplab` objects.* `xolotl` contains template header files for producing custom con-
124 ductances. The template contains instructions on how to design novel conductances with arbitrary
125 specifications.

126 *Simulation.* Models are simulated in `xolotl` with the `integrate` function which outputs as time series
127 the membrane potentials, intracellular calcium concentrations, controller states, intrinsic currents, and
128 synaptic currents. The `integrate` function also accepts an argument which specifies injected current or
129 clamped voltage.

130 *Numerical integration.* `xolotl` uses the exponential Euler method for single compartment models, for-
131 ward Euler for gating variables, and a Crank-Nicholson regime for electrically-coupled compartments
132 (Butcher 2016; Dayan and Abbott 2001; Oh and French 2006). These defaults provide a mix of speed,
133 accuracy, and stability, and are built into the `cpplab` header files. Custom `cpplab` header files can
134 be customized with any iterative integration method. The simulation time-resolution can be specified to
135 target arbitrary precision, and an output time step can be selected to support automatic down-sampling for
136 memory considerations.

137 *‘Closed-loop’ vs. ‘open-loop.’* Simulations can be run in ‘closed-loop’ mode where each simulation be-
138 gins by resetting all dynamical variables to their initial conditions at instantiation, or ‘open-loop’ mode
139 which begins simulation with the current network state.

140 *Using the graphical interface to manipulate parameters.* `xolotl` comes packaged with a graphical user
141 interface for visualizing parameter changes in real-time. The `manipulate` function opens the GUI,
142 which displays a figure plotting the membrane potential and intracellular calcium concentration of all
143 compartments as time series, and a dialog box with customizable sliders for all parameters of the model,
144 much like the `Manipulate` function in Wolfram Mathematica. Moving the sliders integrates the
145 model in ‘open-loop’ mode with the new parameters. The parameters available in the sliders can be cus-
146 tomized by passing a cell array to `manipulate`. For example, to only see sliders for maximal conductances
147 of the HH compartment, call `x.manipulate('HH*gbar')`. Closing the GUI saves the network state
148 of the model to the `xolotl` object. This is particularly helpful for rapid prototyping of models.

149 *Optimizing parameters.* `xolotl` can use the Global Optimization toolbox for MATLAB to optimize any
150 accessible `xolotl` parameters. The toolbox is algorithm-agnostic and accepts any function in MATLAB
151 with a scalar first output as the objective function. Simulations run on multi-core processors or high-
152 performance computing clusters using the Parallel Computing toolbox.

2.3 LIMITATIONS

153 The focus on ease-of-use and speed means some features were intentionally neglected in the streamlining
154 process.

155 *Reliance on compiled C++ code.* While MATLAB comes with robust features for compiling C and C++
156 code, xolotl cannot run without C++ compilation. For users, this necessitates the additional step of
157 setting up the mex compiler which can be problematical, especially for nonstandard (e.g. Arch-based
158 Linux). Secondly, compilation adds a small amount to total processing time. Longer simulations (> 1000
159 time-steps) minimizes this effect. Adding new conductances also requires writing some C++ code. For
160 model conductances in the Hodgkin-Huxley formalism (Dayan and Abbott 2001; Hodgkin, Huxley, and
161 Katz 1952, adjustments consist of changing default values in a template C++ header file. Implementing a
162 new integration scheme requires much more in-depth usage of C++.

163 *Limited to conductance-based models.* xolotl has been developed specifically for conductance-based
164 models. It does not currently support rate- or current-based models.

165 *Limited numerical integration strategies.* While the exponential Euler method performs well in neuronal
166 models (Dayan and Abbott 2001; Oh and French 2006, it may be desirable to use other methods un-
167 der certain conditions. xolotl does not currently support other integration schemes for its built-in
168 conductances, nor does the software support error-sensitive variable step-sizes ‘out-of-the-box.’

169 *Inefficient tools for handling large networks.* While xolotl can integrate large networks (> 1000 com-
170 partments), xolotl uses string-based comprehension for labeling compartments which is suited to
171 descriptive naming, but prohibits vector operations over compartments.

3 USAGE EXAMPLES

172 In MATLAB, users create a xolotl object and populate it with cpplab-generated objects which describe
173 compartments, conductances, synapses, and controllers. The model is integrated with the integrate
174 function where the membrane potential, intracellular calcium concentration, controller states, intrinsic
175 currents, and synaptic currents can be outputs.

176 xolotl comes packaged with a library of pre-existing conductance and synapse objects which greatly
177 simplify the task of constructing model neurons. These objects can be referenced by name and added
178 directly to a compartment. Novel conductance dynamics can be easily written by modifying a template
179 header file contained in the xolotl distribution, or designed entirely from scratch.

3.1 SIMULATING A HODGKIN-HUXLEY MODEL

180 The seminal Hodgkin-Huxley model of action potentials in the squid giant axon (Hodgkin and Huxley
181 1952b; Hodgkin, Huxley, and Katz 1952 contains a fast inactivating sodium conductance (NaV), a non-
182 inactivating delayed rectifier (Kd), and a passive leak current (Figure 1A). A compartment, HH, with
183 membrane capacitance (Cm) and surface area (A) can be specified by Figure 1B. Network properties can
184 be set during construction or afterwards using dot-notation in MATLAB (e.g. x.HH.Cm). Figure 1C shows
185 the MATLAB command prompt after invoking the xolotl object x, displaying the hierarchical structure
186 inherent in conductance-based treatments of neurodynamics.

187 This model was constructed using conductances from Liu et al. 1998 based on electrophysiological
188 recordings from the lobster stomatogastric ganglion (Turrigiano, LeMasson, and E. Marder 1995. In the
189 absence of applied positive current, the model is quiescent. When 0.2 nA is injected, the model ton-
190 ically spikes (Figure 1D). The integrate function takes the applied current as an argument (e.g.

191 `x.integrate(Iapp)`), so that the `xolotl` object is agnostic to integration-specific perturbations.
 192 The `plot` function generates voltage and intracellular calcium traces, where the voltage trace is colored
 193 by the dominant current. If the membrane potential is increasing, the strongest instantaneous inward cur-
 194 rent colors the trace. Conversely, if the membrane potential is decreasing, the strongest outward current
 195 colors the trace instead. Figure 1F-I display the results of the `show` function. Activation and inactivation
 196 steady-states and the voltage-dependent time constants of these gating variables describe the conductance
 197 dynamics in absence of other channel types.

3.2 PERFORMING A VOLTAGE CLAMP EXPERIMENT *IN-SILICO*

198 `xolotl` can recapitulate the results of voltage clamp experiments (Destexhe and Bal 2009; Swensen and
 199 E. Marder 2000, 2001; Turrigiano, LeMasson, and E. Marder 1995. Figure 2 displays steps in the
 200 procedure to clamp the membrane potential of a cell with delayed rectifier potassium conductance. Dur-
 201 ing an *in-vitro* experiment, confounding currents would be pharmacologically-blocked and two-electrode
 202 voltage clamp used to record tail currents at fixed membrane potential (Connor and Stevens 1971a,b).

203 A single-compartment model with a delayed-rectifier conductance is simulated at stepped membrane
 204 potentials. The model is simulated using the `integrate` function. The second argument determines the
 205 clamped voltage and the fourth output is the current trace.

```
206 [V, Ca, ~, I] = x.integrate([], clamped_voltage)
```

207 Currents under voltage clamp approach the steady-state holding current (Figure 2D-E). The current-
 208 voltage relation is the steady-state current over the clamped voltage, and the effective conductance is the
 209 derivative of that relation (Figure 2F-G). Since the effective conductance is the product of the maximal
 210 conductance and the gating variables (Dayan and Abbott 2001; Turrigiano, LeMasson, and E. Marder
 211 1995 and the tail current is monotonically-increasing with time under voltage clamp, the current can be
 212 represented as non-inactivating. Fitting a sigmoid to various powers yields a model for the current dy-
 213 namics (Figure 2H-I). These figures describe graphically the theoretical underpinnings of current analysis
 214 through voltage clamp and can serve as an effective pedagogical tool for computational and quantitative
 215 neuroscience.

3.3 SIMULATING NETWORK MODELS

216 Network models in `xolotl` consist of compartment objects connected by synapses. Synapses are stored
 217 in a vector array as a field of the `xolotl` object in MATLAB. Presynaptic and postsynaptic labels in-
 218 dicate the connectivity of the synapse. Figure 3 implements a model of the triphasic pyloric
 219 rhythm in the stomatogastric ganglion of crustaceans. The pyloric model contains three compartments
 220 and seven synapses (Figure 3A). This structure is reciprocated in the hierarchy of the `xolotl` object,
 221 where conductances are contained within compartments (Figure 3B).

222 Representing the network in `xolotl` requires constructing three compartments and eight conductances
 223 in each using the `add` function.

```
224 x.add('AB', 'compartment', 'Cm', 10, 'A', 0.628, ...)
225 x.AB.add('prinz/NaV', 'gbar', 1000, 'E', 50)
226 ...
```

227 Synapses are upper-level properties of the network which point between two compartments (Figure
 228 3C). This exploits vectorized operations in MATLAB and does not require each synapse to possess a
 229 unique name. The `connect` function adds synapses to the network. The first two arguments specify
 230 the presynaptic and postsynaptic compartments. The third dictates the type of synapse. All others follow
 231 the 'keyword', value paradigm and preset parameters of the synapse.

```
232 x.connect('AB', 'LP', 'Chol', 'gbar', 30)
```

3.4 SIMULATING INTEGRAL CONTROL

xolotl can implement homeostatic tuning rules as integral control. The controller computes an error signal (typically a function of intracellular calcium concentration), and adjusts the conductance or synapse it controls accordingly (O’Leary et al. 2013). In xolotl, integral controllers are cppLab objects added to the conductance or synapse they regulate.

In a demonstration adapted from O’Leary et al. 2013, integral control changes maximal conductances to bring a neuron from quiescence into a bursting regime. Calcium sensors supervene on maximal conductance density (Figure 4) to change neuronal activity. Each conductance in the xolotl structure contains a calcium-sensitive controller (Figure 4B-C). Maximal conductances increase from random initial conditions to a set which elicits the desired network output by minimizing the error signal (Figure 4D-F).

3.5 USING THE GUI TO MANIPULATE PARAMETERS

The manipulate function opens the GUI which permits visualization of changing parameters in real-time. Moving sliders representing the values of network parameters updates a plot (Figure 5B). By default, the function opens a figure displaying the results of the plot function, which shows the voltage and intracellular calcium traces for each compartment (Figure 5A). manipulate grants slider control over all xolotl parameters by default, but specific ones can be selected by passing them as arguments. For example, to manipulate only the maximal conductances and visualize using the myPlot function

```
x.manipulate('*gbar', @myplot)
```

4 BENCHMARKS

To assess speed and accuracy, xolotl, DynaSim (Sherfey et al. 2018), and NEURON (M L Hines and Carnevale 1997) were compared in simulations over varied time resolution, simulation time, and number of compartments (Figure 6).

Single-compartment Hodgkin-Huxley-like models were generated using conductance dynamics from Liu et al. 1998 in the simulation environments. xolotl uses the exponential Euler method for integrating membrane potential (Dayan and Abbott 2001). DynaSim was implemented with a 2nd-order Runge-Kutta integration scheme as recommended for high-performance in the documentation. NEURON used an implicit Euler regime (M L Hines and Carnevale 1997).

To compare the integration methods, models were simulated for 5 s at varying time-resolution (Figure 6A). The ratio between ‘simulated’ time and actual runtime was defined as the speed factor. Higher values indicate faster simulations. The coincidence factor determines the correlative overlap between two spike trains (Jolivet et al. 2008). To assess accuracy over decreasing time-resolution for the three simulation environments, spike trains at each resolution were compared to a ‘canonical’ spike train (exponential Euler at a time-step of $dt = 0.001$ ms).

To assess the performance of the simulators in absence of set-up overhead, models were simulated with a time-resolution of 0.1 ms over increasing simulation time (Figure 6B). The speed factor was defined as the ratio between time represented in the simulation and actual runtime (simulation-time). Therefore, the speed factor represents how many times faster the simulation is than a real-time observation.

Many simulators perform well in simulations of many compartments (Brette et al. 2007; Delorme and Thorpe 2003; Sherfey et al. 2018; Vitay, Dinkelbach, and Hamker 2015). To assess how xolotl performs in these conditions, networks of up to 1,000 Hodgkin-Huxley cells were simulated for 5 s at a time-resolution of 0.1 ms (Figure 6C).

272 DESCRIBE BENCHMARK RESULTS

5 DISCUSSION

273 We envision that `xolotl` will be helpful in teaching students how to interpret cellular biophysics. The
274 modular structure of `cpplab` and the graphical interface simplifies the process of manipulating and
275 analyzing the properties of electrical excitability.

5.1 REPRODUCIBILITY

276 `xolotl` fosters reproducibility in science. While the availability of hosting sites with version control (viz.
277 GitHub (<https://github.com>), GitLab (<https://gitlab.com/>), and Open Science Frame-
278 work (<https://osf.io/>)) and the push for reproducibility in computational science (Baker 2016;
279 Eklund, Nichols, and Knutsson 2016; Stodden et al. 2016 has resulted in the availability of source code,
280 much of this code base is bespoke and difficult to implement (Sedano 2016; W Xu, D Xu, and Deng 2017).

281 To this end, `xolotl` provides an environment with readability and reproducibility in mind. Each net-
282 work is hashed to provide a unique alphanumeric identifier. Conductance header files are easily viewed in
283 the `xolotl` source files; conductances in MATLAB contain links to the full path of the generating file.

5.2 CIRCUMVENTING LANGUAGE TRADEOFFS

284 Executing C/C++ code in higher-level languages such as MATLAB or Python often provides speed
285 improvements for iterative code in algorithms.

286 C is statically-typed, with procedural syntax that provides low-level access to memory (Kernighan and
287 Ritchie 1978, providing significant advantages for time-intensive computations. Unfortunately, automatic
288 code-generation is limited by the supervening language. MATLAB, for instance, cannot use pointers or pass
289 by reference, which limits the efficiency of C code automatically generated from MATLAB. Conversely,
290 custom C/C++ code provides significant increases in performance and memory conservation, but lacks
291 the ease-of-use and flexibility of scripting languages.

292 `xolotl` handles this problem through symbolic manipulation of C++ objects in MATLAB. Built from
293 the ground up in C++, `xolotl` maintains all the advantages of custom compiled code, but can run in
294 MATLAB without the user having to touch the C++ code. `xolotl` represents compartment, conductance,
295 synapse, and controllers as `cpplab` objects, which map to underlying C++ header files. In this way,
296 properties of the `xolotl` network can be examined and changed using object-oriented paradigms. The
297 object specifies the `integrate` function, not the other way around.

5.3 APPLICATIONS OF CPPLAB

298 NEED TO WRITE THIS

CONFLICT OF INTEREST STATEMENT

299 The authors declare that the research was conducted in the absence of any commercial or financial
300 relationships that could be construed as a potential conflict of interest.

AUTHOR CONTRIBUTIONS

301 SG-S designed and implemented the core of the `xolotl` toolbox. AH contributed to the code base,
302 created the online user documentation, and wrote the manuscript. EM supervised the project. All authors
303 reviewed the paper.

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SUPPLEMENTAL DATA

308 Tables including all conductances packaged with `xolotl` should be put in the supplementary material.

DATA AVAILABILITY STATEMENT

309 The code to generate all figures is available at ([https://github.com/marderlab/](https://github.com/marderlab/xolotl-paper)
310 `xolotl-paper`). `xolotl` is freely available at (<https://github.com/marderlab/xolotl>).

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FIGURE CAPTIONS

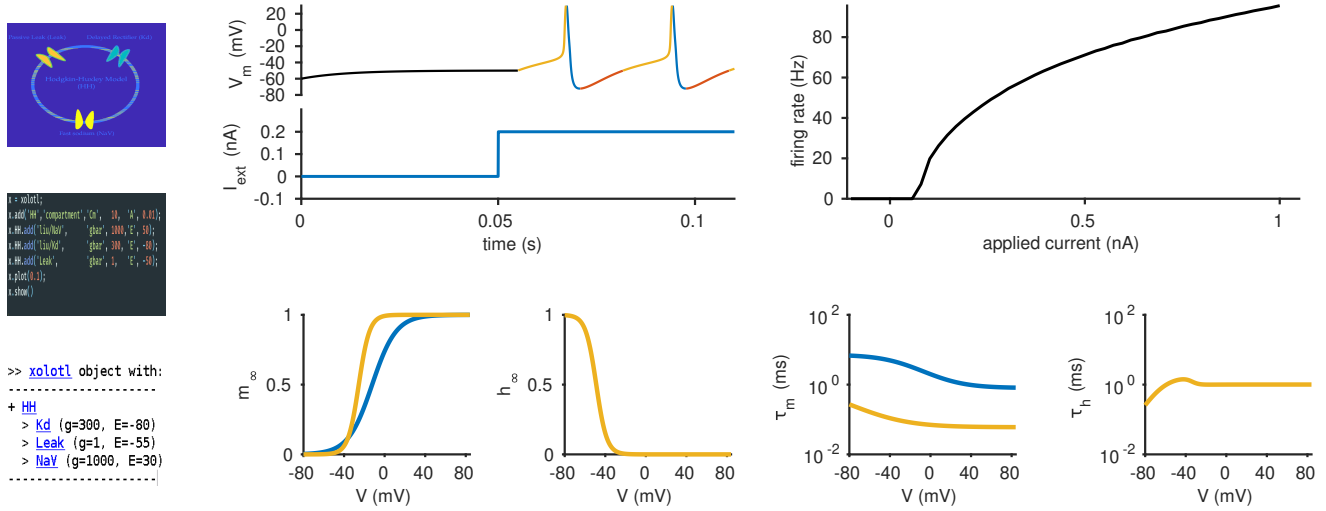


Figure 1: `xolotl` can quickly set up and simulate conductance-based models. (A) Cartoon of a Hodgkin-Huxley single-compartment neuron model with fast sodium, delayed rectifier, and leak currents. (B) Code snippet in MATLAB used to implement D, F-I. (C) `xolotl` schematic displayed in the MATLAB command prompt. (D) Simulated voltage trace of a Hodgkin-Huxley model with three conductances and 0.2 nA of injected current. Colors indicate the dominant current (gold is fast sodium, blue is delayed rectifier, red is leak). (E) Firing rate-input relation displaying firing rate as a function of injected current. (F-G) Steady-state gating functions for activation (m) and inactivation (h) gating variables. (H-I) Voltage-dependence of time constants for activation (m) and inactivation (h) gating variables.

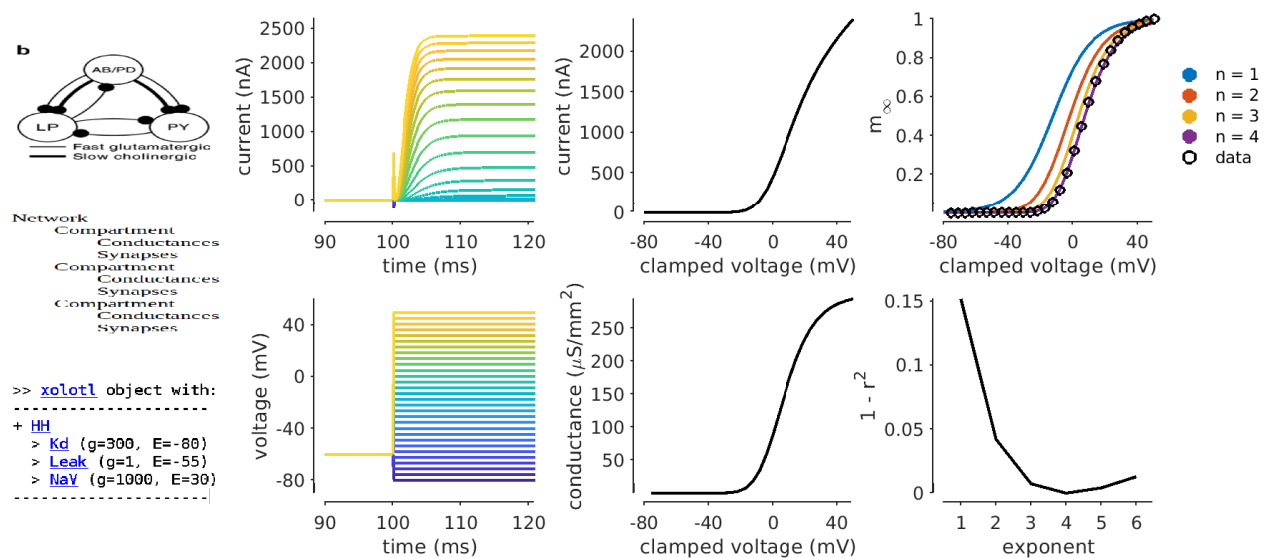


Figure 2: Simulating a voltage-clamp experiment. (A) Cartoon of a cell with delayed rectifier potassium conductance (Liu et al. 1998 with experimentally-fixed voltage). (B) Structure of `xolotl` object in A. (C) Code snippet depicting integration under voltage clamp. (D-E) Current response to steps in voltage from a holding potential of $V_m = -60$ mV. (F) Current-voltage relation of the steady-state current ($t = 400$ ms) indicating a reversal potential of $E = -80$ mV and no inactivation. (G) Conductance-voltage relation at steady-state takes the form of a sigmoid. (H) Sigmoids m fit to the model as m^n data indicating that $n = 4$ is the best fit. (I) Goodness of fit vs. exponent n , suggesting $n = 4$ as the best fit to the data.

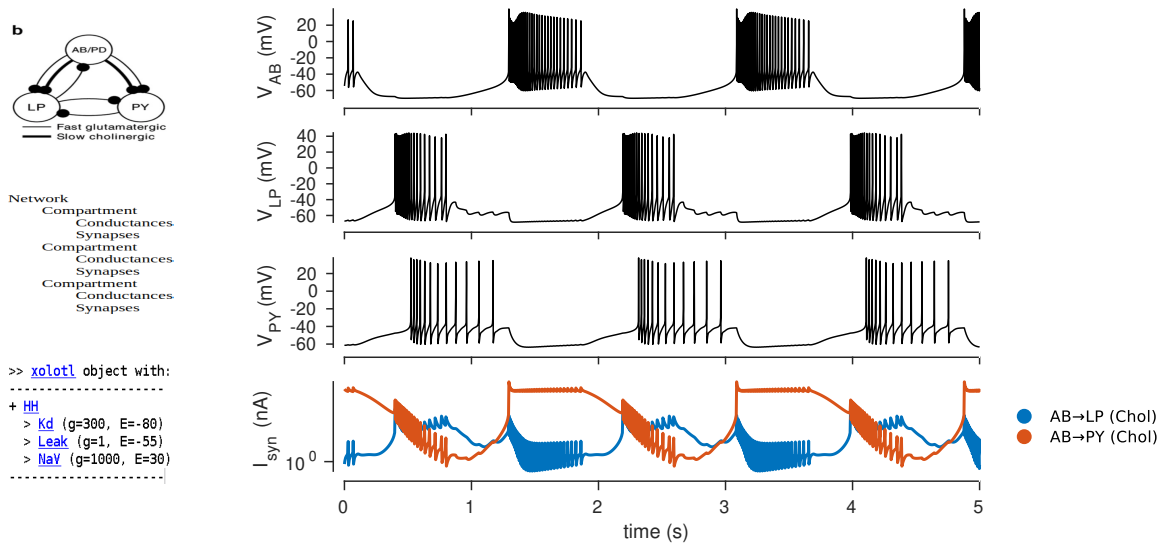


Figure 3: Simulating a network of conductance-based model neurons. (A) Diagram of a network model of the pyloric rhythm in the crustacean stomatogastric ganglion (Prinz *et al.* 2004). (B) Each neuron is modeled as a single compartment with 7-8 intrinsic conductances and 1-3 post-synaptic conductances. (C) *xolotl* implements conductances as fields of compartments and synapses as connections between compartments. (D-F) Simulated voltage trace of a model network for the three compartments. (G) Time series of synaptic currents in the simulated network can be obtained from the integration.

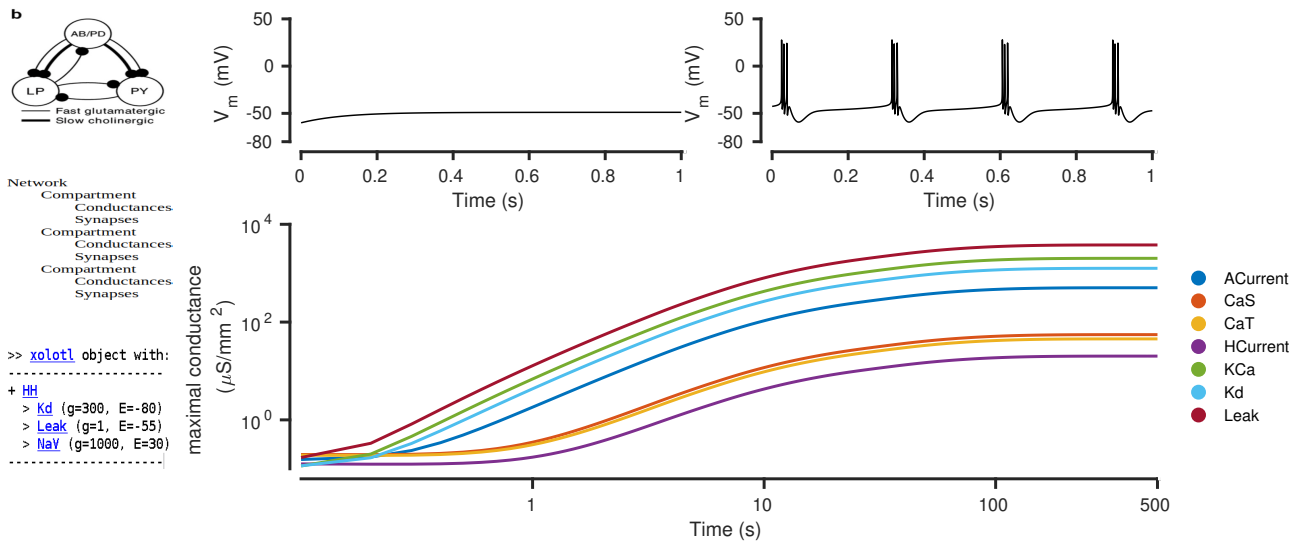


Figure 4: Simulating neurons under homeostatic regulation. (A) Cartoon of a model neuron (Liu et al. 1998) with integral control (O'Leary et al. 2013). (B) Hierarchical structure of a neuronal network considers controllers as components of compartments which act on conductances. (C) *xolotl* implements controllers as properties of conductances and synapses. (D) Calcium sensors change maximal conductances to move a neuron from quiescence to a bursting state. (E) Voltage trace shows regular bursting activity after integral control.

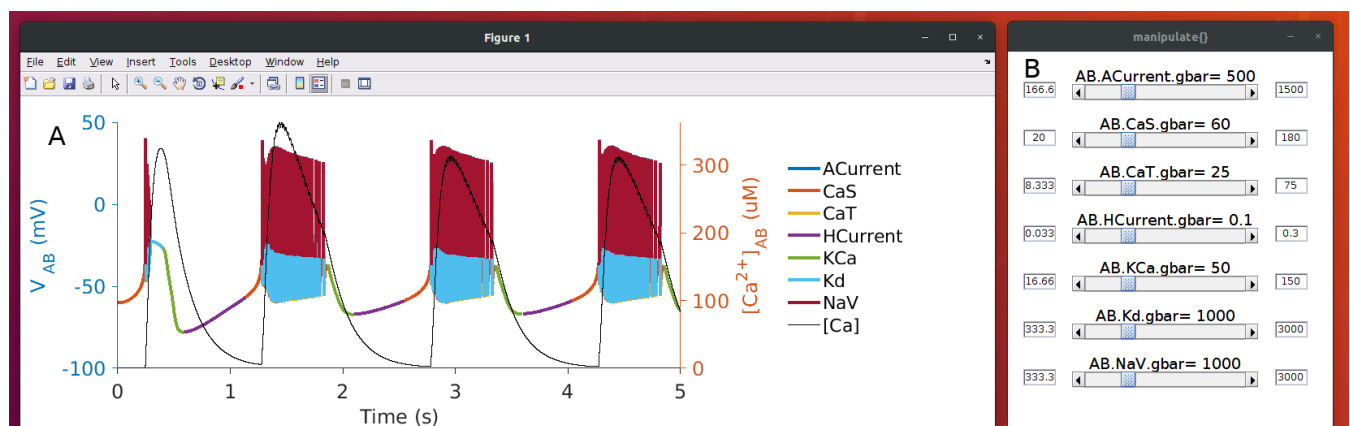


Figure 5: Using the GUI to manipulate neuron parameters. (A) Real-time output of the `plot` function displaying voltage (colored) and intracellular calcium (black) traces of a bursting neuron model (Astrid A. Prinz, Billimoria, and Eve Marder 2003; Astrid A. Prinz, Bucher, and Eve Marder 2004). Colors indicate the dominant current. (B) Sliders control the maximal conductances, which updates on the figure.

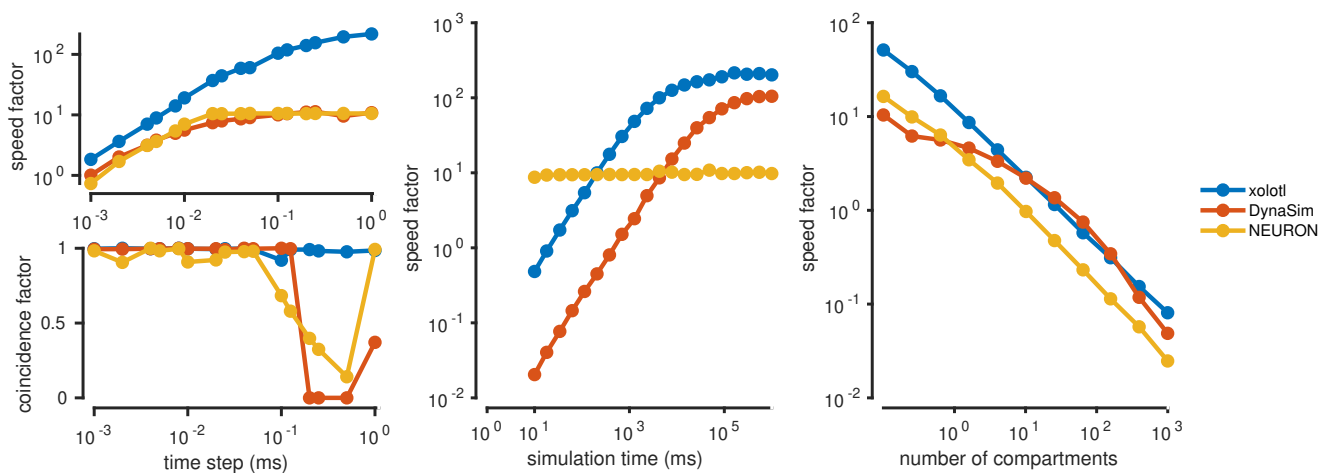


Figure 6: `xolotl` benchmarked against `DynaSim` and `NEURON`. (A) Ratio of 'simulated' time to runtime (speed factor) and accuracy, measured by spike train coincidence plotted against decreasing time-resolution. (B) Speed factor for models at increasing simulation times. (C) Speed factor over number of compartments.