Statistics for Linguists 08 July 2022

10:00	Workshop introduction
10:15	Loading and exploring datasets
10:45	Data transformation and coding
11:15	Practical exercise
12:15	Review of practical
12:30 - 13:30	LUNCH BREAK
13:30	lmer and glmer
14:30	Post-hoc analysis and model visualization
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16:00	Review of practical
16:15	Model building
17:00	End of workshop

Statistics for Linguists

Learning objectives

- You will learn to load/import data
- Explore a dataset and create descriptive statistics
- Transform a dataset (if needed)
- Code your factors
- Build a mixed model
- Perform post-hoc statistics
- Visualize your data and your model

Various approaches being used (keeping it maximal, minimal)

- Build up models one factor at a time
 - Imer(ReadingTime ~ capitalization)
 - Interactions + main effects: determiner * capitalization
 - Interaction only (no main effects): determiner : capitalization
 - intercept: (1 | randEf)
 - Intercept + slope: (1 + fixedEf| randEf)
- Compare models based on the AIC

One common approach:

Fit maximal model

```
> mm <- lmer(DV ~ Factor + (Factor | Subj) + (Factor | Item), data=data, REML=FALSE)
```

> print(summary(lmm), corr=FALSE)

- Check random effects structure
- > summary(rePCA(lmm))

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```
$participant
Importance of components:

[,1] [,2]
Standard deviation 0.298 0
Proportion of Variance 1.000 0
Cumulative Proportion 1.000 1
```

\$participant

Importance of components:

```
• Check random effects structure

> summary(rePCA(lmm))

Standard deviation 0.298 0

Proportion of Variance 1.000 0

Cumulative Proportion 1.000 1
```

> VarCorr(lmm)

```
Random effects:
Groups Name Variance Std.Dev. Corr
participant (Intercept) 0.031141 0.1765
capitalization1 0.000625 0.0250 -1.00
Residual 0.357613 0.5980
Number of obs: 2039, groups: participant, 30
```

- Check random effects structure
- > summary(rePCA(lmm))

- If proportion of variance explained is non-zero for all principal components (PCs), both for subject-related and for item-related PCs, you can likely keep them.
- If some are zero, remove them
- Convergence problems mean that the model is not supported by the data

- In this case, I would remove the correlation as the first step
- > mm1 <- lmer(ReadingTime ~ capitalization + (1 + capitalization | | participant), psycholinguistics_data=data)

```
Random effects:

Groups Name Variance Std.Dev. Corr
participant (Intercept) 0.031141 0.1765
capitalization1 0.000625 0.0250 -1.00
Residual 0.357613 0.5980
Number of obs: 2039, groups: participant, 30
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- Convergence problems mean that the model is not supported by the data
- When you obtain a singular fit, this is often indicating that the model is overfitted

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- When you obtain a singular fit, this is often indicating that the model is overfitted

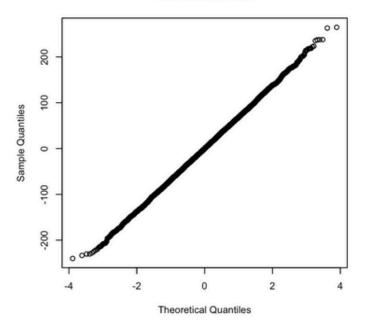
```
> m3.lmer <- lmer(log(ReadingTime) ~ capitalization * determiner + (1 + capitalization | participant), data = psycholinguistics_data)
```

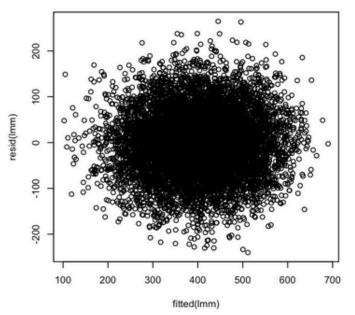
```
boundary (singular) fit: see ?isSingular
Warning message:
Model failed to converge with 1 negative eigenvalue: -8.4e+01
```

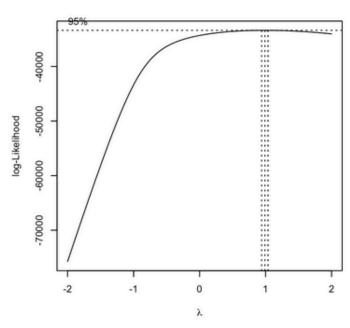
Check residuals

- > qqnorm(resid(lmm))
- > plot(fitted(lmm), resid(lmm))
- > boxcox(DV ~ Factor, data=data)

Normal Q-Q Plot

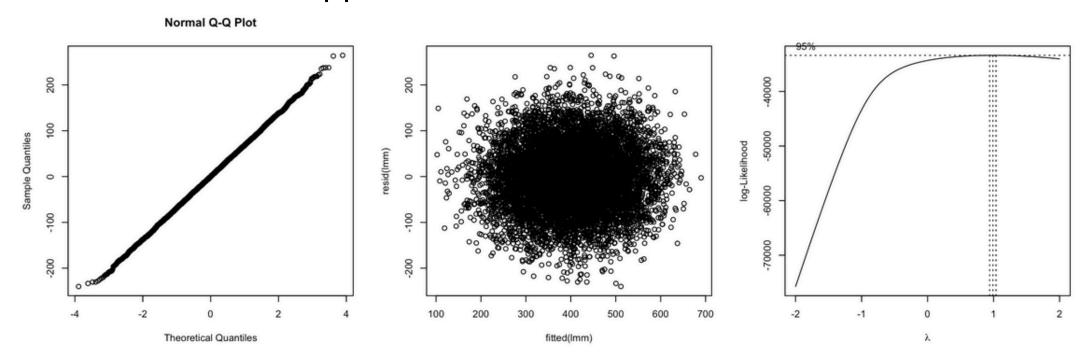






Check residuals

• The figures show that the residuals do not strongly deviate from a normal distribution (left panel) and that the variance of the residuals is similar across different fitted values (homoscedasticity, middle panel). Power transformations (right panel) further show no transformation is needed to better approximate a normal distribution.



- When you've determined the maximal random effects structure, you can start building your fixed effects, step by step
 - You can build up: start with an empty model, add factors and test if they contribute
 - Or build down: start with a full model and remove factors that don't contribute
 - And/Or keep some fixed effects based on your hypotheses

- Your model building choices may depend on your hypothesis and the amount of variables you have
- Example 1: we want to know the effect of *condition*. We can add it and see if it reaches significance. If not, we leave it in and report it
 - Advantage: there is a p-value to report, as the factor is still in the model
 - Disadvantage: leaving factors that don't contribute increases chances of overfitting

- Your model building choices may depend on your hypothesis and the amount of variables you have
- Example 2: we want to know which of 10 cognitive abilities, socio economic background factors, and language history measures relate to reading time
 - Adding all factors and interactions is too much for the model; it does not converge
 - We could add factors stepwise. If they don't contribute, remove them again
 - Advantage: probably the only feasible approach, leads to a converging model
 - Disadvantage: no estimates to report for factors that didn't make the cut
 - Disadvantage: interactions between background factors may be missed

- How do we determine whether a factor 'contributes'?
- This is not based on significance p-value
- Should be based on model comparison. An easy-ish way is with anova()

```
> m8.lmer <- lmer(log(ReadingTime) ~ capitalization + (1 | participant),
      data = psycholinguistics data2)
> m9.lmer <- lmer(log(ReadingTime) ~ capitalization + determiner + (1 |
      participant), data = psycholinguistics data2)
> anova(m8.lmer, m9.lmer)
     Models:
     m8.lmer: log(ReadingTime) ~ capitalization + (1 | participant)
     m9.lmer: log(ReadingTime) ~ capitalization + determiner + (1 | participant)
                   AIC BIC logLik deviance Chisq Df Pr(>Chisq)
     m8.lmer 4 3757.1 3779.6 -1874.5 3749.1
     m9.lmer 5 3531.3 3559.4 -1760.6 3521.3 227.82 1 < 2.2e-16 ***
```

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

- Be careful to only add one factor/ take on step at a time
- Look at the AIC or BIC values
 - Lower is better
 - Rule of thumb: a difference of 2 points indicates a better model
 - BIC corrects more strictly for the number of parameters than AIC

- You can add as many models as you like to anova()
- > anova(m8.lmer, m9.lmer, m10.lmer, m11.lmer, ...)

```
Models:
m8.lmer: log(ReadingTime) ~ capitalization + (1 | participant)
m9.lmer: log(ReadingTime) ~ capitalization + determiner + (1 | participant)
npar AIC BIC logLik deviance Chisq Df Pr(>Chisq)
m8.lmer 4 3757.1 3779.6 -1874.5 3749.1
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---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Strategy for determining a parsimonious model

Here is sketch of recommendation about how to proceed to determine a parsimonious model. Note that theoretical expectations about parameters may lead to different sequences

- 1. Perform a PCA on the maximal model: summary(rePCA(max_lmm)). If the maximal model doesn't converge, then proceed to step (3).
- 2. If all principle component(s) explain non-zero variance, use the maximal model
- If one (or more) PC explain zero variance, fit a zero correlation parameter model
- 4. Remove predictor with smallest variance of random slopes. E.g.: $lmm < -lmer(DV \sim 1 + c1 + c3 + (1 + c1 + c3 | lid), data=dat)$
- 5. Add correlation parameter to this reduced model. E.g., $lmm <- lmer(DV \sim 1 + c1 + c3 + (1 + c1 + c3 | id)$, data=dat)
- 6. Perform a PCA to check whether all PC explain non-zero variance. If not, repeat steps (3-6). Otherwise this is the parsimonious model.

Alternative / additional criterion: Use model comparison based on AIC or BIC

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