

Sensitivity/Specificity Analyses - Canine Leishmaniosis

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Exploratory Analyses

##		DPP	
##	PCR	Negative	Positive
##	Negative	743	16
##	Positive	1	11

Models

Angela's paper (Toepp et al., 2019, <https://doi.org/10.1371/journal.pntd.0007058>) uses logistic regression, with age, sex, and variables that have to do with diagnostic tests as explanatory variables. They are something like this:

Model A 1: $\text{logit}(\pi_k) = \beta_0 + \beta_1 \text{Age}_k + \beta_2 \text{Sex}_k + \beta_3 Y_k$, where Y_k is diagnostically positive (as defined in Model 1 below), but for the mom and π_k is the probability of disease for individual k

Model A 2: $\text{logit}(\pi_k) = \beta_0 + \beta_1 \text{Age}_k + \beta_2 \text{Sex}_k + \beta_3 T_{1k} + \beta_4 T_{2k}$, where T_{jk} is the result for Test j (as defined in Model 1 below), but for the mom and π_k is the probability of disease for individual k

Note, these models were fit with a log link function, presumably so that relative risks could be recovered?

We plan to evaluate similar models for our data and to then incorporate sensitivity and specificity of the tests into these models. Then we will compare the model performance to that of other methods. Hopefully we will see an improvement/some details that we miss when we do not include the sensitivity and specificity for the tests.

In all these models, we will assume that the observations are independent.

Model 1:

Data Model

$$T_{1k}|D_k \sim \text{Bernoulli}(\pi_{1k})$$

$$\text{where } \pi_{1k} = P(T_{1k} = 1|D_k) = D_k \times \underbrace{P(T_{1k} = 1|D_k = 1)}_{\text{sensitivity}} + (1 - D_k) \times \underbrace{(1 - P(T_{1k} = 0|D_k = 0))}_{1 - \text{specificity}}.$$

$$T_{2k}|D_k \sim \text{Bernoulli}(\pi_{2k})$$

$$\text{where } \pi_{2k} = P(T_{2k} = 1|D_k) = D_k \times \underbrace{P(T_{2k} = 1|D_k = 1)}_{\text{sensitivity}} + (1 - D_k) \times \underbrace{(1 - P(T_{2k} = 0|D_k = 0))}_{1 - \text{specificity}}.$$

Process Model

$$D_k \sim \text{Bernoulli}(\delta_k)$$

$$\text{where } \delta_k = \text{logit}^{-1}(\text{logit}(\rho) + \mathbf{x}_k^T \boldsymbol{\beta}).$$

Prior Model

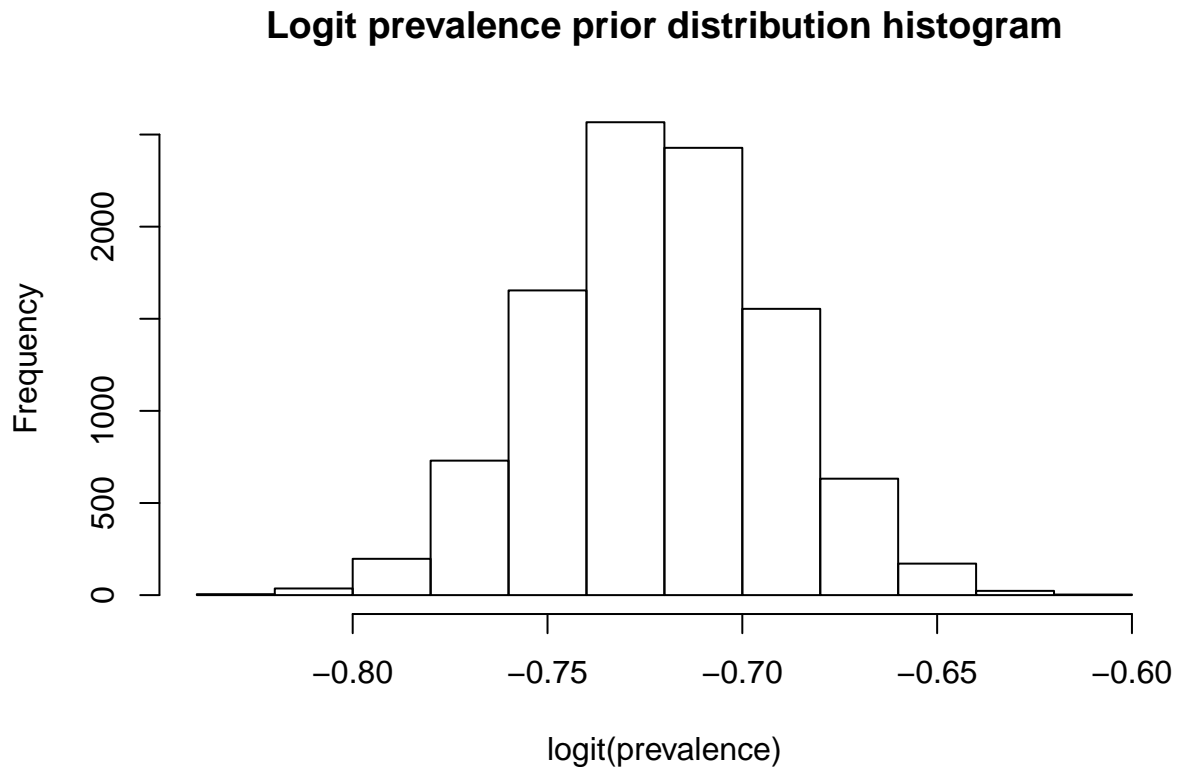
Prevalence

$$\text{logit}(\rho) \sim \text{Normal}(\mu_\rho, \tau^2)$$

where $\mu_\rho = \text{logit}^{-1}(\rho^*)$ and $\tau^2 = \dots$

We have a range for the prevalence of (0.05, 0.10). This corresponds to a range of (-0.7497, -0.6972) on the logit scale.

```
hist(rnorm(10000, mean = log(0.075)/(1-log(0.075)), sd = 0.03),  
     main="Logit prevalence prior distribution histogram",  
     xlab="logit(prevalence)")
```



Note, the prevalence in this population may be higher - these are exposed dogs in the United States in our hunting hound population. We may want to change this, but we also want to generalize this to the larger canine population - to Brazil, if possible.

Linear predictors

The regression parameters are $\beta = (\beta_{age}, \beta_{sex}, \beta_{age*sex})^T$;

$$\beta \sim \text{Normal}(\mu_\beta, \Sigma_\beta)$$

where $\mu_\beta = \mathbf{0}$ and $\Sigma_\beta = \mathbf{I}$ in our code.

```

## ranges of sensitivities and specificities
sens.pcr.range <- c(0.839, 0.990)
sens.dpp.range <- c(0.832, 0.930)
spec.pcr.range <- c(0.871, 0.970)
spec.dpp.range <- c(0.682, 0.951)

sens.pcr <- mean(sens.pcr.range)
sens.dpp <- mean(sens.dpp.range)
spec.pcr <- mean(spec.pcr.range)
spec.dpp <- mean(spec.dpp.range)

## range of prevalence for visceral leishmaniasis
prev.range <- c(0.05,0.10)
prev <- mean(prev.range)

```

OpenBUGS Model 1 Implementation

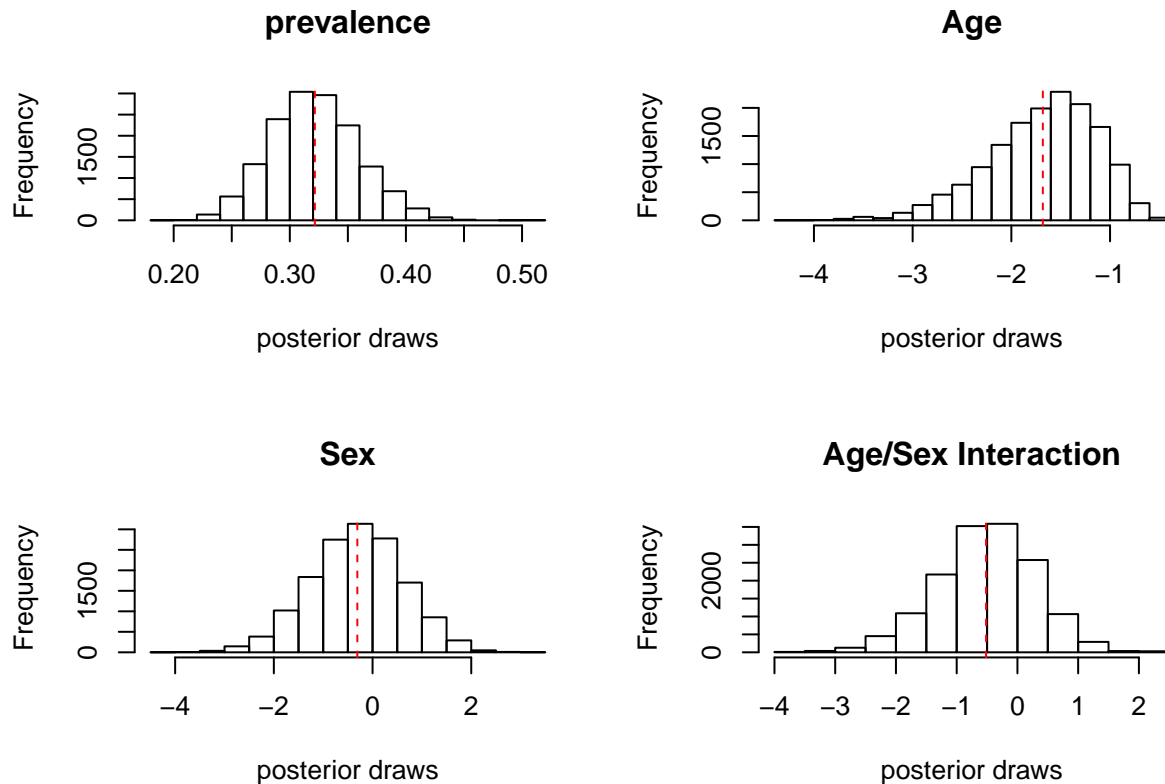
OpenBUGS Model 1 Posterior Distributions

```

par(mfrow=c(2,2))
## Graphical summaries of posterior distributions
hist(exp(model1_df$lpi)/(1+exp(model1_df$lpi)), main="prevalence", xlab="posterior draws")
abline(v=mean(exp(model1_df$lpi)/(1+exp(model1_df$lpi))), lty="dashed", col="red")

hist(model1_df$b1, main="Age", xlab="posterior draws")
abline(v=mean(model1_df$b1), lty="dashed", col="red")
hist(model1_df$b2, main="Sex", xlab="posterior draws")
abline(v=mean(model1_df$b2), lty="dashed", col="red")
hist(model1_df$b3, main="Age/Sex Interaction", xlab="posterior draws")
abline(v=mean(model1_df$b3), lty="dashed", col="red")

```



OpenBUGS Model 1 Disease State Classification

```
## Set up storage for model results
pred_df_m1 <- data.frame(obs=1:nind,
  D=rep(NA,nind), ## average estimate
  SD=rep(NA,nind),
  LB=rep(NA,nind), ## 2.5th percentile
  UB=rep(NA,nind), ## 97.5th percentile
  model_assignment=rep(NA,nind),
  Clinical_status=ss_data2$ClinicalStatus,
  Diagnostic_status=ss_data2$Diagnostically_positive)

## Calculate probabilities of compartment membership for each posterior draw
pred_df_m1$D <- apply(model1_df[,grep("D", names(model1_df))], 2, mean)
pred_df_m1$SD <- apply(model1_df[,grep("D", names(model1_df))], 2, sd)
pred_df_m1$LB <- apply(model1_df[,grep("D", names(model1_df))], 2,
  quantile, probs=0.025)
pred_df_m1$UB <- apply(model1_df[,grep("D", names(model1_df))], 2,
  quantile, probs=0.975)

summary(pred_df_m1)
```

##	obs	D	SD	LB	UB
----	-----	---	----	----	----

```
## Min. : 1.0 Min. :0.000e+00 Min. :0.000000 Min. :0
## 1st Qu.:193.5 1st Qu.:0.000e+00 1st Qu.:0.000000 1st Qu.:0
## Median :386.0 Median :0.000e+00 Median :0.000000 Median :0
## Mean :386.0 Mean :7.531e-05 Mean :0.001797 Mean :0
## 3rd Qu.:578.5 3rd Qu.:0.000e+00 3rd Qu.:0.000000 3rd Qu.:0
## Max. :771.0 Max. :1.073e-02 Max. :0.103048 Max. :0
## UB model_assignment Clinical_status Diagnostic_status
## Min. :0 Mode:logical A: 0 Negative:743
## 1st Qu.:0 NA's:771 N:736 Positive: 28
## Median :0 S: 35
## Mean :0
## 3rd Qu.:0
## Max. :0
```

```
## Apply a cut off of point estimate of 0.5; if pi.D > 0.5, classify as S (symptomatic), otherwise as N
## Summarize in a table (clinical status versus diagnostic status)
```

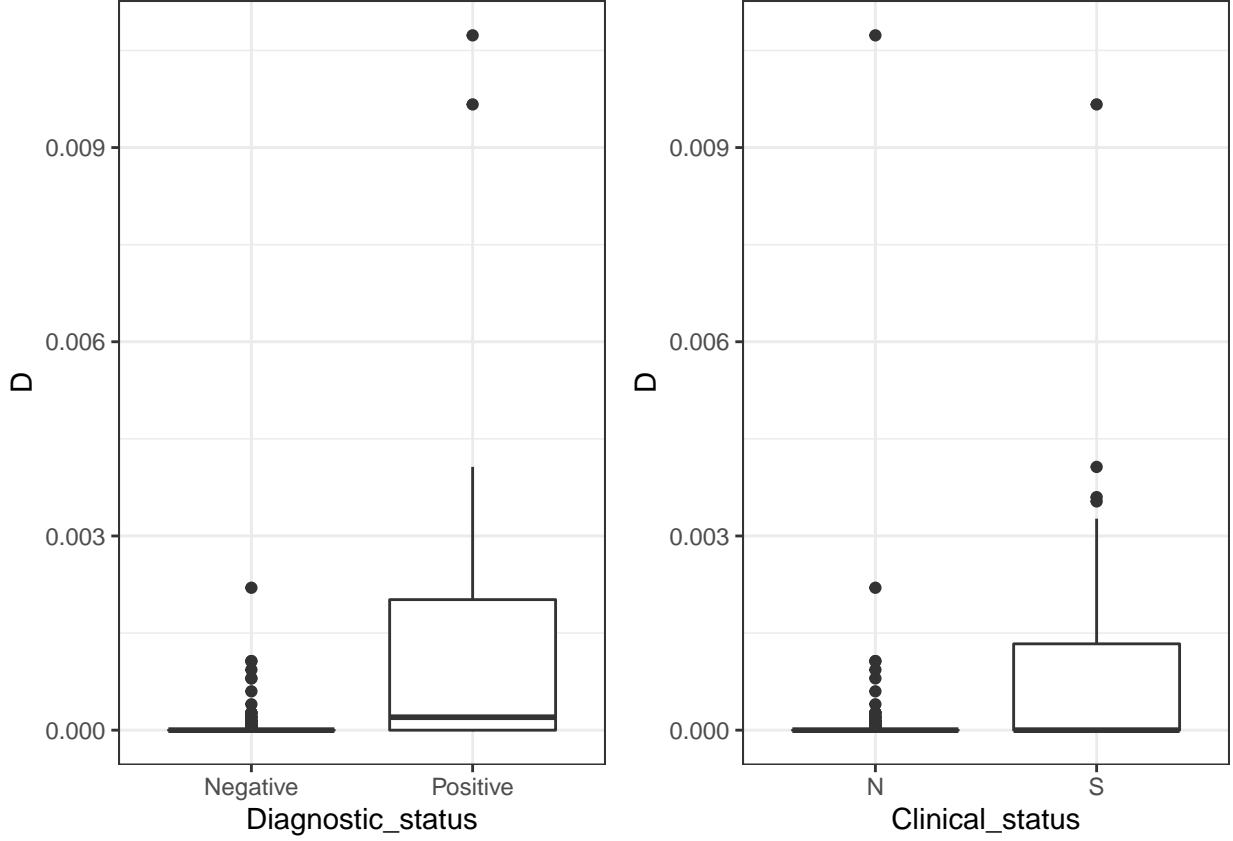
```
table(pred_df_m1[pred_df_m1$D > 0.5,]$Clinical_status,
      pred_df_m1[pred_df_m1$D > 0.5,]$Diagnostic_status)
```

```
##
##      Negative Positive
## A         0         0
## N         0         0
## S         0         0
```

```
## Print summary table of clinical status versus diagnostic status from the original data
table(pred_df_m1$Clinical_status, pred_df_m1$Diagnostic_status)
```

```
##
##      Negative Positive
## A         0         0
## N        728         8
## S         15        20
```

```
## boxplots
p1 <- (ggplot(data=pred_df_m1, aes(x=Diagnostic_status, y=D))
      + geom_boxplot()
      + theme_bw())
p2 <- (ggplot(data=pred_df_m1, aes(x=Clinical_status, y=D))
      + geom_boxplot()
      + theme_bw())
ggarrange(p1,p2, nrow=1)
```



Model 2:

The data outcome we are using is “diagnostically positive”, meaning that an individual tests positive on at least one diagnostic test. This is what we have used in our other papers and seems to be popular in the literature (add some references to this). In this model, we assume that the two diagnostic tests are independent, and that there is some imprecision in the test results, so we include sensitivity and specificity for each test in the model.

Data Model

$$Y_k | T_{1k}, T_{2k}, D_k \sim \text{Bernoulli}(\pi_k^{DP})$$

We are assuming that the test outcomes are independent. The probability of a diagnostically positive test for individual k , π_k^{DP} , is

$$\begin{aligned} \pi_k^{DP} &= P(Y_k = 1 | D_k) \\ &= D_k \times P(Y_k = 1 | D_k = 1) + (1 - D_k) \times (1 - P(Y_k = 0 | D_k = 0)), \end{aligned}$$

where $P(Y_k = 1 | D_k = 1) = P(T_{1k} = 1 | D_k = 1) + P(T_{2k} = 1 | D_k = 1) - P(T_{1k} = 1 | D_k = 1) \times P(T_{2k} = 1 | D_k = 1)$ and $P(Y_k = 0 | D_k = 0) = P(T_{1k} = 0 | D_k = 0) \times P(T_{2k} = 0 | D_k = 0)$.

Process Model

$$D_k \sim \text{Bernoulli}(\delta_k)$$

where $\delta_k = \text{logit}^{-1}(\text{logit}(\rho) + \mathbf{x}_k^T \beta)$.

Prior Model

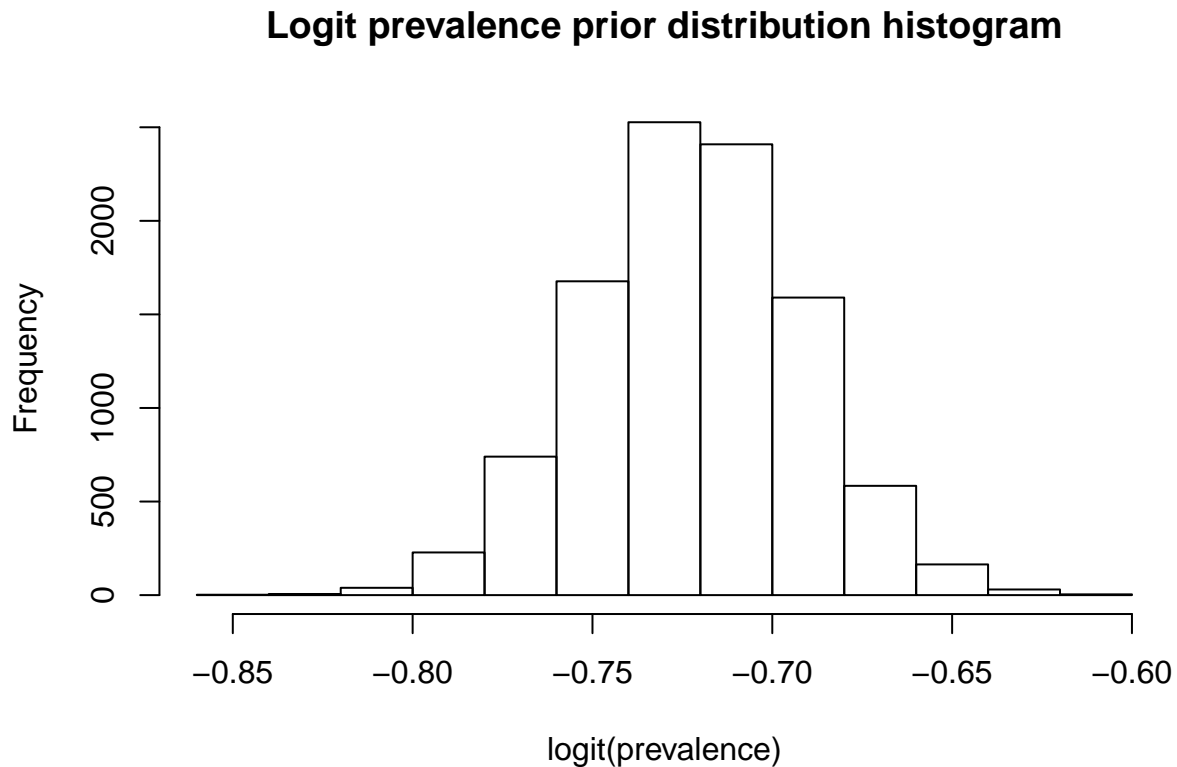
Prevalence

$$\text{logit}(\rho) \sim \text{Normal}(\mu_\rho, \tau^2)$$

where $\mu_\rho = \text{logit}^{-1}(\rho^*)$ and $\tau^2 = \dots$

We have a range for the prevalence of (0.05, 0.10). This corresponds to a range of (-0.7497, -0.6972) on the logit scale.

```
hist(rnorm(10000, mean = log(0.075)/(1-log(0.075)), sd = 0.03),  
     main="Logit prevalence prior distribution histogram",  
     xlab="logit(prevalence)")
```



Note, the prevalence in this population may be higher - these are exposed dogs in the United States in our hunting hound population. We may want to change this, but we also want to generalize this to the larger canine population - to Brazil, if possible.

Linear predictors

The regression parameters are $\beta = (\beta_{age}, \beta_{sex}, \beta_{age*sex})^T$;

$$\beta \sim \text{Normal}(\mu_\beta, \Sigma_\beta)$$

where $\mu_\beta = \mathbf{0}$ and $\Sigma_\beta = \mathbf{I}$ in our code.

Other parameters:

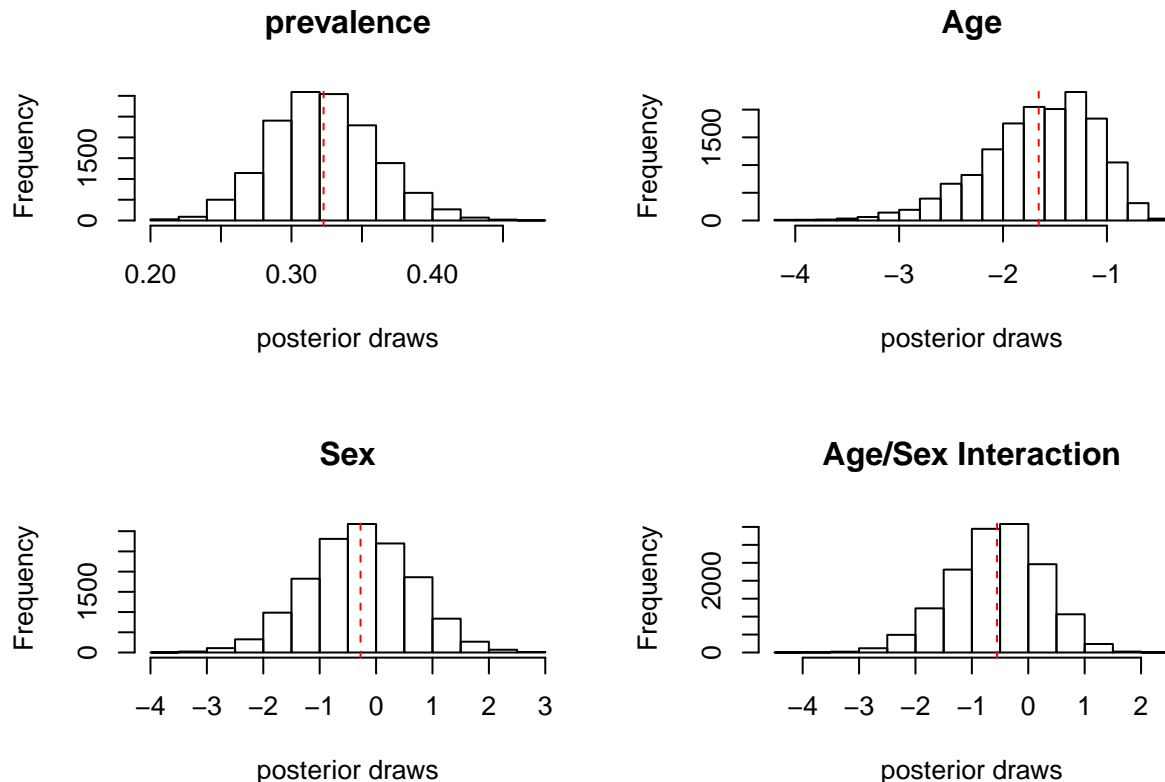
$$\beta \sim \text{Normal}(\mu_\beta, \Sigma_\beta)$$

We will assume that the regression coefficients are independent, so Σ_β is a diagonal matrix.

OpenBUGS Model 2 Implementation

```
par(mfrow=c(2,2))
## Graphical summaries of posterior distributions
hist(exp(model2_df$lpi)/(1+exp(model2_df$lpi)), main="prevalence", xlab="posterior draws")
abline(v=mean(exp(model2_df$lpi)/(1+exp(model2_df$lpi))), lty="dashed", col="red")

hist(model2_df$b1, main="Age", xlab="posterior draws")
abline(v=mean(model2_df$b1), lty="dashed", col="red")
hist(model2_df$b2, main="Sex", xlab="posterior draws")
abline(v=mean(model2_df$b2), lty="dashed", col="red")
hist(model2_df$b3, main="Age/Sex Interaction", xlab="posterior draws")
abline(v=mean(model2_df$b3), lty="dashed", col="red")
```



```
## Numeric summaries of posterior distributions
#boxplot(model2_df[,!(names(model2_df) %in% c("deviance"))])
```

The mean posterior prevalence is 0.3228 and the 95% credible interval is: (0.2526, 0.3991).

OpenBUGS Model 2 Disease State Prediction

```
## Set up storage for model results
pred_df_m2 <- data.frame(obs=1:nind,
                        D=rep(NA,nind), ## average estimate
                        SD=rep(NA,nind),
                        LB=rep(NA,nind), ## 2.5th percentile
                        UB=rep(NA,nind), ## 97.5th percentile
                        model_assignment=rep(NA,nind),
                        Clinical_status=ss_data2$ClinicalStatus,
                        Diagnostic_status=ss_data2$Diagnostically_positive)

## Calculate probabilities of compartment membership for each posterior draw
pred_df_m2$D <- apply(model2_df[,grep("D", names(model2_df))], 2, mean)
pred_df_m2$SD <- apply(model2_df[,grep("D", names(model2_df))], 2, sd)
pred_df_m2$LB <- apply(model2_df[,grep("D", names(model2_df))], 2,
                        quantile, probs=0.025)
pred_df_m2$UB <- apply(model2_df[,grep("D", names(model2_df))], 2,
                        quantile, probs=0.975)

summary(pred_df_m2)
```

```
##      obs          D          SD          LB
## Min.   : 1.0   Min.   :0.0000000   Min.   :0.000000   Min.   :0
## 1st Qu.:193.5 1st Qu.:0.0000000   1st Qu.:0.000000   1st Qu.:0
## Median :386.0 Median :0.0000000   Median :0.000000   Median :0
## Mean   :386.0 Mean   :0.0001646   Mean   :0.001691   Mean   :0
## 3rd Qu.:578.5 3rd Qu.:0.0000000   3rd Qu.:0.000000   3rd Qu.:0
## Max.   :771.0 Max.   :0.0956000   Max.   :0.294052   Max.   :0
##      UB      model_assignment Clinical_status Diagnostic_status
## Min.   :0.000000   Mode:logical   A: 0           Negative:743
## 1st Qu.:0.000000   NA's:771      N:736         Positive: 28
## Median :0.000000           S: 35
## Mean   :0.001297
## 3rd Qu.:0.000000
## Max.   :1.000000
```

```
## Apply a cut off of point estimate of 0.5; if pi.D > 0.5, classify as S (symptomatic), otherwise as N
## Summarize in a table (clinical status versus diagnostic status)
table(pred_df_m2[pred_df_m2$D > 0.5,]$Clinical_status,
      pred_df_m2[pred_df_m2$D > 0.5,]$Diagnostic_status)
```

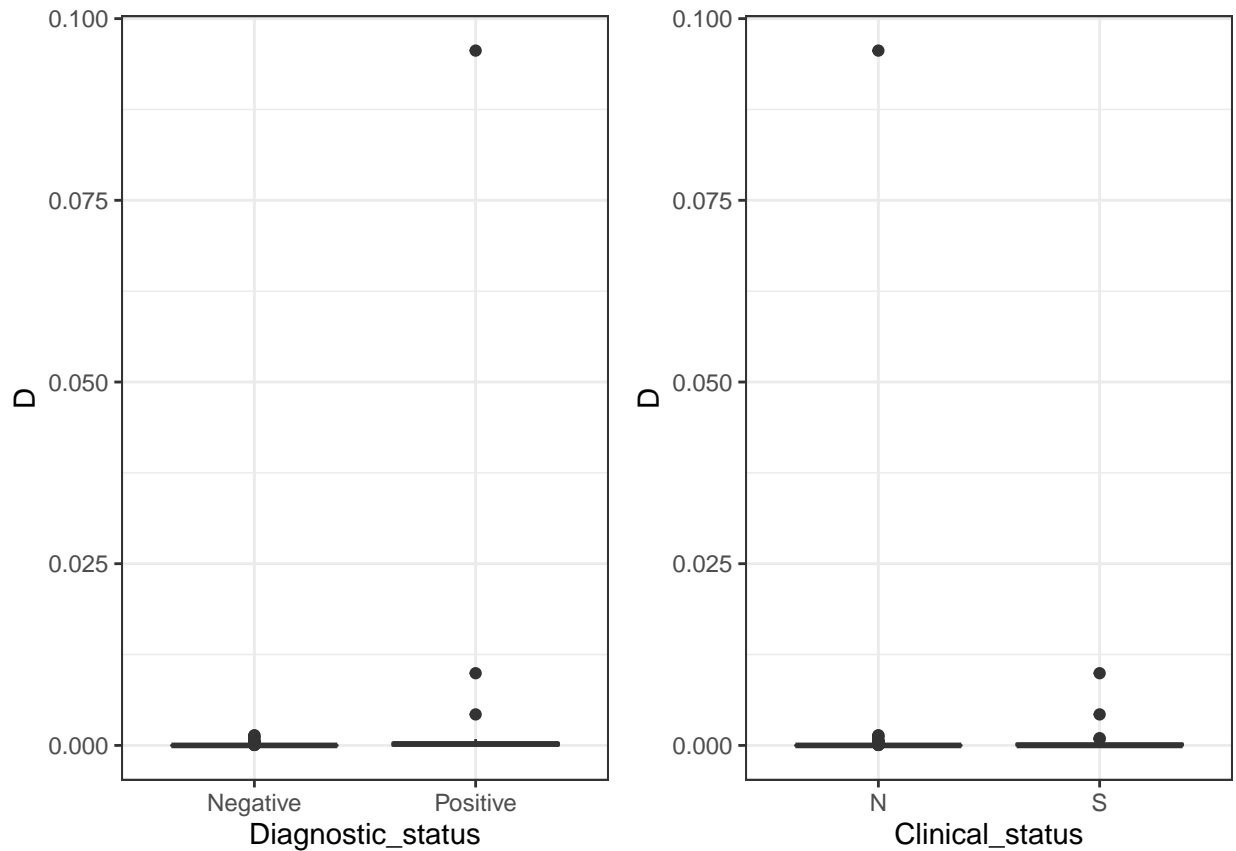
```
##
##      Negative Positive
## A          0         0
## N          0         0
## S          0         0
```

```
## Print summary table of clinical status versus diagnostic status from the original data
table(pred_df_m2$Clinical_status, pred_df_m2$Diagnostic_status)
```

```
##
##      Negative Positive
##      A          0      0
##      N        728      8
##      S          15     20

## boxplots
p1 <- (ggplot(data=pred_df_m2, aes(x=Diagnostic_status, y=D))
      + geom_boxplot()
      + theme_bw())
p2 <- (ggplot(data=pred_df_m2, aes(x=Clinical_status, y=D))
      + geom_boxplot()
      + theme_bw())

ggarrange(p1,p2, nrow=1)
```



Model 3:

The data outcome we are using is “diagnostically positive”, meaning that an individual tests positive on at least one diagnostic test. This is what we have used in our other papers and seems to be popular in the literature (add some references to this). In this model, we assume that the two diagnostic tests are independent, and that there is some imprecision in the test results, so we include sensitivity and specificity for each test in the model. We also include a random intercept term to see how this changes the model.

Data Model

$$Y_k|T_{1k}, T_{2k}, D_k \sim \text{Bernoulli}(\pi_k^{DP})$$

We are assuming that the test outcomes are independent. The probability of a diagnostically positive test for individual k , π_k^{DP} , is

$$\begin{aligned}\pi_k^{DP} &= P(Y_k = 1|D_k) \\ &= D_k \times P(Y_k = 1|D_k = 1) + (1 - D_k) \times (1 - P(Y_k = 0|D_k = 0)),\end{aligned}$$

where $P(Y_k = 1|D_k = 1) = P(T_{1k} = 1|D_k = 1) + P(T_{2k} = 1|D_k = 1) - P(T_{1k} = 1|D_k = 1) \times P(T_{2k} = 1|D_k = 1)$ and $P(Y_k = 0|D_k = 0) = P(T_{1k} = 0|D_k = 0) \times P(T_{2k} = 0|D_k = 0)$.

Process Model

$$D_k \sim \text{Bernoulli}(\delta_k)$$

where $\delta_k = \text{logit}^{-1}(\text{logit}(\rho) + \mathbf{x}_k^T \boldsymbol{\beta} + \epsilon_k)$.

Prior Model

Prevalence

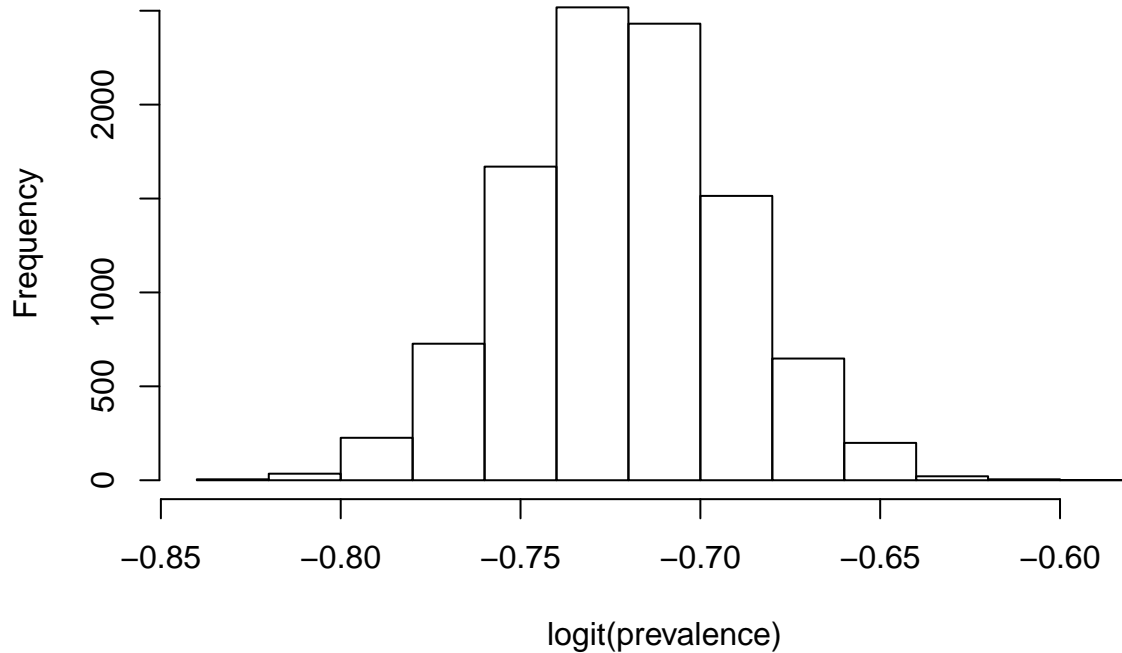
$$\text{logit}(\rho) \sim \text{Normal}(\mu_\rho, \tau^2)$$

where $\mu_\rho = \text{logit}^{-1}(\rho^*)$ and $\tau^2 = \dots$

We have a range for the prevalence of (0.05, 0.10). This corresponds to a range of (-0.7497, -0.6972) on the logit scale.

```
hist(rnorm(10000, mean = log(0.075)/(1-log(0.075)), sd = 0.03),  
     main="Logit prevalence prior distribution histogram",  
     xlab="logit(prevalence)")
```

Logit prevalence prior distribution histogram



Note, the prevalence in this population may be higher - these are exposed dogs in the United States in our hunting hound population. We may want to change this, but we also want to generalize this to the larger canine population - to Brazil, if possible.

Linear predictors

The regression parameters are $\beta = (\beta_{age}, \beta_{sex}, \beta_{age*sex})^T$;

$$\beta \sim Normal(\mu_{\beta}, \Sigma_{\beta})$$

where $\mu_{\beta} = \mathbf{0}$ and $\Sigma_{\beta} = \mathbf{I}$ in our code.

We will assume that the regression coefficients are independent, so Σ_{β} is a diagonal matrix.

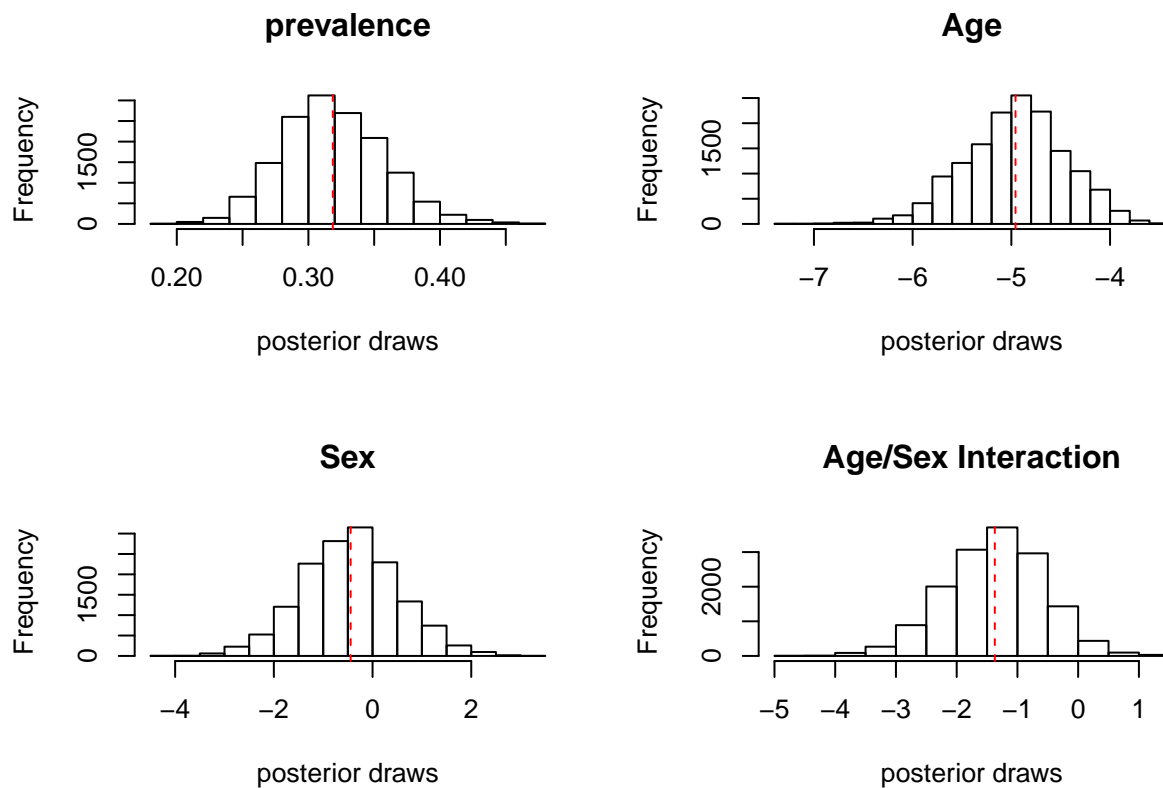
Individual random effect - parameterized using precision.

$$\epsilon \sim Normal(0, 5 \times 10^{-3})$$

OpenBUGS Model 3 Implementation

```
par(mfrow=c(2,2))
## Graphical summaries of posterior distributions
hist(exp(model3_df$lpi)/(1+exp(model3_df$lpi)), main="prevalence", xlab="posterior draws")
abline(v=mean(exp(model3_df$lpi)/(1+exp(model3_df$lpi))), lty="dashed", col="red")
```

```
hist(model3_df$b1, main="Age", xlab="posterior draws")
abline(v=mean(model3_df$b1), lty="dashed", col="red")
hist(model3_df$b2, main="Sex", xlab="posterior draws")
abline(v=mean(model3_df$b2), lty="dashed", col="red")
hist(model3_df$b3, main="Age/Sex Interaction", xlab="posterior draws")
abline(v=mean(model3_df$b3), lty="dashed", col="red")
```



```
## Numeric summaries of posterior distributions
#boxplot(model3_df[,!(names(model3_df) %in% c("deviance"))])
```

The mean posterior prevalence is 0.3185 and the 95% credible interval is: (0.2477, 0.399).

OpenBUGS Model 3 Disease State Prediction

```
## Set up storage for model results
pred_df_m3 <- data.frame(obs=1:nind,
  D=rep(NA,nind), ## average estimate
  SD=rep(NA,nind),
  LB=rep(NA,nind), ## 2.5th percentile
  UB=rep(NA,nind), ## 97.5th percentile
  model_assignment=rep(NA,nind),
  Clinical_status=ss_data2$ClinicalStatus,
  Diagnostic_status=ss_data2$Diagnostically_positive)
```

```
## Calculate probabilities of compartment membership for each posterior draw
pred_df_m3$D <- apply(model3_df[,grep("D", names(model3_df))], 2, mean)
pred_df_m3$SD <- apply(model3_df[,grep("D", names(model3_df))], 2, sd)
pred_df_m3$LB <- apply(model3_df[,grep("D", names(model3_df))], 2,
  quantile, probs=0.025)
pred_df_m3$UB <- apply(model3_df[,grep("D", names(model3_df))], 2,
  quantile, probs=0.975)

summary(pred_df_m3)
```

```
##      obs      D      SD      LB
## Min.   : 1.0   Min.   :0.000000   Min.   :0.000000   Min.   :0
## 1st Qu.:193.5 1st Qu.:0.000000   1st Qu.:0.000000   1st Qu.:0
## Median :386.0 Median :0.000000   Median :0.000000   Median :0
## Mean   :386.0 Mean   :0.001572   Mean   :0.009204   Mean   :0
## 3rd Qu.:578.5 3rd Qu.:0.000000   3rd Qu.:0.000000   3rd Qu.:0
## Max.   :771.0 Max.   :0.502000   Max.   :0.500013   Max.   :0
##      UB      model_assignment Clinical_status Diagnostic_status
## Min.   :0.000000   Mode:logical      A: 0           Negative:743
## 1st Qu.:0.000000   NA's:771          N:736          Positive: 28
## Median :0.000000                      S: 35
## Mean   :0.006485
## 3rd Qu.:0.000000
## Max.   :1.000000
```

```
## Apply a cut off of point estimate of 0.5; if pi.D > 0.5, classify as S (symptomatic), otherwise as N
## Summarize in a table (clinical status versus diagnostic status)
table(pred_df_m3[pred_df_m3$D > 0.5,]$Clinical_status,
  pred_df_m3[pred_df_m3$D > 0.5,]$Diagnostic_status)
```

```
##
##      Negative Positive
## A      0      0
## N      0      1
## S      0      0
```

```
## Print summary table of clinical status versus diagnostic status from the original data
table(pred_df_m3$Clinical_status, pred_df_m3$Diagnostic_status)
```

```
##
##      Negative Positive
## A      0      0
## N     728      8
## S     15     20
```

```
## boxplots
p1 <- (ggplot(data=pred_df_m3, aes(x=Diagnostic_status, y=D))
  + geom_boxplot()
  + theme_bw())
p2 <- (ggplot(data=pred_df_m3, aes(x=Clinical_status, y=D))
```

```
+ geom_boxplot()  
+ theme_bw()
```

```
ggarrange(p1,p2, nrow=1)
```

