OpenStats: A Robust and Scalable Software Package for Reproducible Analysis of High-Throughput Phenotypic Data

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OpenStats is a freely available R package that presents statistical methods and detailed analyses to promote the hard process of identification of abnormal phenotypes. The package incorporates several checks and cleaning on the input data prior to the statistical analysis. For continuous data, Linear Mixed Model with an optional model selection routine is implemented, whilst for categorical data, Fisher's Exact Test is implemented. For cases where the linear mixed model fails, Reference Range Plus method has been employed for a quick, simple analysis of the continuous data. User can perform inspections and diagnostics of the final fitted model by the visualisation tools that come with the software. Furthermore, the user can export/report the outputs in the form of either standard R list or JavaScript Object Notation (JSON). OpenStats has been tested and demonstrated with an application of 2.5M+ analyses from the Internationa Mouse Phenotyping Consortium (IMPC).

The User's Guide with more details about the statistical analysis is available as part of the online documentation from https://rpubs.com/hamedhm/openstats. Project Github repository including dev version of the package is available on https://git.io/JeOVN.

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1 Building block of the software

OpenStats consists of one input layer and three operational layers:

- 1. (Input layer) Input data and specifying model: this includes the input data and an initial model in the form of standard R formula, e.g. $y \sim x + 1$.
- 2. (**Operational layer 1**) Dataset preprocessing: this includes checking, cleaning and terminology unification procedures and is completed by the function *OpenStatsList* which creates an *OpenStatsList* object.
- 3. (**Operational layer 2**) Data analysis: this is managed by the function *OpenStatsAnalysis* and consists of Linear Mixed Model, Fisher's Exact test and Reference Range plus framework implementations. The results are stored in an *OpenStatsMM/FE/RR* object.
- 4. (**Operational layer 3**) Report/Export: the exports/reports are managed by the function *OpenStat-sReport*. *OpenStats* reports the outputs in the form of either List or JSON objects.

2 Data preprocessing

OpenStatsList function performs data processing and creates an *OpenStatsList* object. As input, *OpenStatsList* function requires dataset of phenotypic data that can be presented as data frame. For instance, it can be dataset stored in csv, tsv or txt file. Data is organised with rows and columns for samples and features respectively. Following shows an example of the input data where rows and columns represent mice and features (mouse id, treatment group, gender, age of animals in days):

```
> library (OpenStats)
> ####################
> # Data preparation
> ##################
> fileCon = system.file("extdata", "test_continuous.csv",
                    package = "OpenStats")
> read.csv(fileCon, as.is = TRUE)[60:75, c('external_sample_id'
                                      'biological_sample_group',
                                      'sex'
                                      'age_in_days')]
##
     external_sample_id biological_sample_group sex age_in_days
## 60
      C10058 control female 53
## 61
               C10059
                                    control female
## 62
              C10060
                                   control female
                                                         53
## 63
              C10061
                                   control female
                                                         53
## 64
              C10062
                                   control female
                                                         53
                                                          53
## 65
              C10063
                                   control male
## 66
                                                          53
              C10064
                                   control male
## 67
                                                          53
              C10065
                                   control male
## 68
               C10066
                                                          53
                                   control male
                                    control male
## 69
               C10067
                                                          53
                                            male
## 70
               C10192
                               experimental
                                                          46
                                            male
## 71
               C10193
                               experimental
                                                          46
## 72
                                                          46
                C10194
                                experimental
                                             male
## 73
                C10195
                                                          46
                                experimental
                                             male
## 74
                C10197
                                experimental
                                             male
                                                          46
                                experimental
## 75
                C10199
                                             male
                                                          46
```

The main preprocessing tasks performed by the *OpenStatsList* function are:

- · terminology unification,
- filtering out undesirable records (when the argument dataset.clean is set to TRUE),
- imputing missings such as blanks, spaces or user-specified terms with NA,

• and checking whether the dataset can be used for the statistical analysis.

We define "terminology unification" as the terminology used to describe data (variables) that are essential for the analysis. *OpenStats* package uses the following nomenclature for the names of columns: "Genotype", the only mandatory variable, "Sex", "Batch" "LifeStage" and "Weight". In addition, expected (default) Sex, LifeStage values are "Male/Female" and "Early/Late" respectively. However, the user can define the custome levels by setting *dataset.values.male*, *dataset.values.female*, *dataset.values.early* and *dataset.values.late* in the OpenStatsList function. Missing value is specified by *dataset.values.missingValue* argument and set to *NA*.

The statistical analysis requires exactly two "Genotype" groups for comparison (e.g. wild-type versus knockout). Thus the function *OpenStatsList* requires users to define the reference genotype (mandatory argument *refGenotype* with default value "control") and test genotype (mandatory argument *testGenotype*), defaulted to "experimental"). If the *OpenStatsList* function argument *dataset.clean* is set to TRUE then all records with genotype values others than reference or test genotype are filtered out.

All tasks in OpenStats are accompanied by step-by-step reports, error messages, warnings and/or other useful information about the progress of the function. If messages are not desirable, *OpenStatsList* function's argument *debug* can be set to FALSE meaning there will be no messages.

The chunk of code below demonstrates an example of using *OpenStatsList* when the user sets outmessages to TRUE/FALSE:

```
# Default behaviour with messages
    >
   library(OpenStats)
>
   fileCon = system.file("extdata", "test_continuous.csv",
                         package = "OpenStats")
   test_Cont = OpenStatsList(
     dataset = read.csv(fileCon),
      testGenotype = 'experimental',
      refGenotype = 'control',
     dataset.colname.genotype = 'biological_sample_group',
dataset.colname.batch = 'date_of_experiment',
      dataset.colname.lifestage = NULL,
     dataset.colname.weight = 'weight',
dataset.colname.sex = 'sex'
## 2020-03-30 17:01:29. Input data of the dimensions, rows = 410, columns = 75
\#\# 2020-03-30 17:01:29. Checking the input data in progress ...
## 2020-03-30 17:01:29. Checking the specified missing values [x2] (' ', ") ...
## 2020-03-30 17:01:29. 1/2. Checking (' ') ...
## 2020-03-30 17:01:29. 2/2. Checking (") ...
## 2020-03-30 17:01:29. Checking whether variable 'biological_sample_group' exists
in the data ...
## 2020-03-30 17:01:29. Result = TRUE
## 2020-03-30 17:01:29. Levels (Total levels = 2, missings = 0%):
## 2020-03-30 17:01:29. 1. control
## 2020-03-30 17:01:29. 2. experimental
## 2020-03-30 17:01:29. Checking whether variable 'sex' exists in the data ...
## 2020-03-30 17:01:29. Result = TRUE
## 2020-03-30 17:01:29. Levels (Total levels = 2, missings = 0%):
## 2020-03-30 17:01:29.
                          1. female
## 2020-03-30 17:01:29. 2. male
## 2020-03-30 17:01:29. Checking whether variable 'date_of_experiment' exists in
the data ...
## 2020-03-30 17:01:29. Result = TRUE
```

```
## 2020-03-30 17:01:29. Levels (Total levels = 43, missings = 0%):
## 2020-03-30 17:01:29. 1. 2012-07-23T00:00:00Z
                        2. 2012-07-30T00:00:002
3. 2012-08-06T00:00:002
4. 2012-08-13T00:00:002
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
                          5. 2012-08-20T00:00:00Z
6. 2012-11-26T00:00:00Z
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
                        6. 2012-11 201
7. 2012-12-24T00:00:00Z
## 2020-03-30 17:01:29.
                         8. 2013-01-02T00:00:00Z
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
                         9. 2013-01-15T00:00:00Z
                         10. 201 ...
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29. Checking whether variable 'weight' exists in the data ...
## 2020-03-30 17:01:29. Result = TRUE
## 2020-03-30 17:01:29. Summary:
                                    = 20.0036585365854
## 2020-03-30 17:01:29. mean
## 2020-03-30 17:01:29.
                         sd
                                    = 2.63972182732584
## 2020-03-30 17:01:29. Missings = 0%
## 2020-03-30 17:01:29. Variable 'biological_sample_group' renamed to 'Genotype'
## 2020-03-30 17:01:29. Variable 'sex' renamed to 'Sex'
## 2020-03-30 17:01:29. Variable 'date_of_experiment' renamed to 'Batch'
## 2020-03-30 17:01:29. Variable 'weight' renamed to 'Weight'
## 2020-03-30 17:01:29. Total samples in Genotype:Sex interactions:
## 2020-03-30 17:01:29. Level(frequency):
## 2020-03-30 17:01:29. 1. control.Female(196)

    experimental.Female(5)
    control.Male(201)
    experimental.Male(8)

## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29. Total 'Weight' data points for Genotype:Sex interactions:
## 2020-03-30 17:01:29. Level(frequency):
## 2020-03-30 17:01:29.
                          1. control.Female(196),

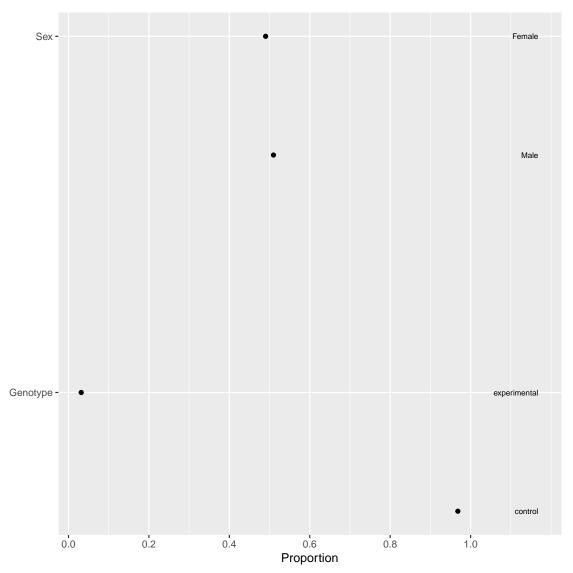
    experimental.Female(5),

## 2020-03-30 17:01:29.
                          3. control.Male(201),
## 2020-03-30 17:01:29.
                         4. experimental.Male(8)
## 2020-03-30 17:01:29.
## 2020-03-30 17:01:29. Successfully performed checks in 0.09 second(s).
    # Default behaviour without messages
    fileCon = system.file("extdata", "test_continuous.csv",
                          package = "OpenStats")
    test_Cont = OpenStatsList(
     dataset = read.csv(fileCon),
     testGenotype = 'experimental',
     refGenotype = 'control',
     dataset.colname.genotype = 'biological_sample_group',
     dataset.colname.batch = 'date of experiment',
     dataset.colname.lifestage = NULL,
     dataset.colname.weight = 'weight',
     dataset.colname.sex
                               = 'sex',
+
     debug
                                = FALSE
> # No output printed
```

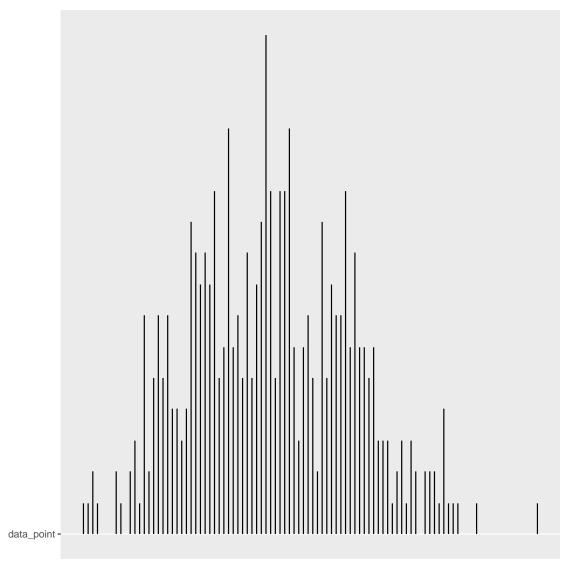
2.1 OpenStatsList Object

The output of the *OpenStatsList* function is the *OpenStatsList* object that contains a cleaned dataset as well as a copy of the original dataset. *OpenStats* allows **plot** and **summary** of the OpenStatList object. Below

is an example of the OpenStatsList function accompanied by the plot and summary:



```
##
## $Continuous
```



OpenStatsList object stores many characteristics of the data, for instance, reference genotype, test genotype, original column names, factor levels etc.

3 Data Analysis

OpenStats package contains three statistical frameworks for the phenodeviants identification:

- 1. Simple Linear/Linear Mixed Models framework that assumes baseline values of the dependent variable are normally distributed but batch (defined as the date of experiment in the IMPC) is the between-group source of variation.
- 2. Reference Range Plus framework identifies the normal variation form a group called *Reference variable* (wild-type animals in the IMPC), classifies dependent variable as low, normal or high and compare proportions. This framework recommended for the sufficient number of controls (more than 60 records) to correctly identify normal variation.
- 3. Fishers Exact Test is a standard framework for categorical data which compares data proportions and calculates the percentage change in classification.

OpenStats's function *OpenStatsAnalysis* works as a hub for the different statistical analysis methods. It checks the dependent variable, the data, missings, not proper terms in the model (such as terms that do not exist in the input data) and runs the selected statistical analysis framework and returns modelling/testing results. All analysis frameworks output a statistical significance measure, effect size measure, model diagnostics, and graphical visualisations.

Here we explain the main bits of the OpenStatsAnalysis function:

- OpenStatsListObject: defines the dataset stored in an OpenStatsList object.
- model: defines the fixed effect model for example, $Response \sim Genotype + Sex$
- **method**: defines which statistical analysis framework to use.

The possible values for the *method* arguments are "MM" which stands for mixed model framework, "FE" to perform Fisher's exact test model and "RR" for Reference Range Plus framework. The semantic naming in the input arguments of the OpenStatsAnalysis function allows natural distinction of the input arguments For example, MM_{-} , RR_{-} and FE_{-} prefix represent the arguments that can be set in the corresponging frameworks. Having said that,

- MM_fixed, MM_random, MM_weight refer to the fixed effect terms, random effect term and between group variation
- FE_formula refers to the model that need to be analysed by Fisher's exacts test(the default $category \sim Genotype + Sex + LifeStage$ in the IMPC)

• RR_formula, RRrefLevel, RR_prop refer to the Reference Range plus model (default data_point ~ Genotype + Sex + LifeStage in the IMPC). Note that the first term on the right hand side of the model (here Genotype) is the Reference Variable and the reference level is defined by "RRrefLevel" (default is set "control" in the IMPC). Finally the natural variation of the reference level to define the so called "NORMAL" category is determined by RR_prop (IMPC 0.95 that is mutants outside the .025 quantile from right/left tails of the distribution are labeles as high/low respectively).

The *OpenStatsAnalysis* function performs basic checks to ensure that the data and model match, the model is feasible for the type of the data and reports step-by-step progress of the function. Some of the checks are listed below:

• Mixed Model (MM) frameworks:

MM_checks: A vector of four 1/0 or TRUE/FALSE values such as c(TRUE, TRUE, TRUE, TRUE)[default]. Performing pre-checks on the input model for some known scenarios. The first element of the vector activates checks on the model terms (in MM_fixed) to have existed in data. The second term removes any single level -factor- from the model (in MM_fixed). The third term removes the single value (such as a column of constants/no variation) from the -continuous- terms in the model (in MM_fixed). The Fourth element checks the interaction term to make sure all interactions have some data attached. Caution is needed for this check as it may take longer than usual if the formula in MM_fixed contains many factors.

Note that OpenStatsAnalysis function always removes duplicated columns in the dataset prior to applying the linear mixed model.

Regardless of the check settings, the OpenStatsAnalysis function always checks for the existence of the "MM_random" terms (provided "MM_random" is set) in the input data

• Reference Range Plus (RR) and Fisher's exact test (FE) framework's:

FERR_FullComparisions Only applies to the "RR" or "FE" frameworks. A vector of two logical flags, default c(TRUE, FALSE). Setting the first value to TRUE, then all combinations of the effects (all levels of factors in the input model - for example Male_LifeStage, Male_Genotype, Male_Mutant, Male_control, Female_control, Female_Mutant, Female_LifeStage and so on) will be tested. Otherwise only main effects (no sub-levels - for example Sex_LifeStage [not for instance Male_LifeStage]) will be tested. Setting the second element of the vector to TRUE (default FALSE) will force the Fisher's Exact test to do all comparisons between different levels of the RESPONSE variable. For example, if the response has three levels such as 1) positive, 2) negative and 3) neutral then the comparison will be between 1&2, 1&3, 2&3 and 1&2&3 (obviously the latter is the full table).

· All frameworks

OpenStatsAnalysis allows confidence intervals for all estimates in three frameworks. One can set the confidence level by setting MMFERR_conf.level to a value in (0,1) interval (default 0.95).

All frameworks are equipped with the step-by-step report of the progress of the function. Warnings, errors and messages are reported to the user. In the situation where the function encounters a critical failure, then the output object contains a slot called *messages* that reports back the cause of the failure.

4 Examples

In this section, we show some examples of the functionalities in *OpenStats* for the continuous and categorical data. Each section contains the code and different possible scenarios.

4.1 Linear mixed model framework

The linear mixed model framework applies to continuous data. In this example, data is extracted from the sample data that accompany the software. Here, "Genotype" is the effect of interest. The response is stored in the variable "data_point" and genotype (Genotype) and body weight (Weight) are covariates. The model selection is left to the default, stepwise, and between-group covariance structure are assumes proportional to the genotype levels (different variation for controls than mutants):

```
library(nlme)
>
    library(OpenStats)
          ##################
>
>
          # Data preparation
          ##################
          ##################
          # Continuous data - Creating OpenStatsList object
          #################
          fileCon = system.file("extdata", "test_continuous.csv",
                                package = "OpenStats")
          test_Cont = OpenStatsList(
                 dataset = read.csv(fileCon),
                  testGenotype = 'experimental',
                  refGenotype = 'control',
                  dataset.colname.genotype = 'biological_sample_group',
                  dataset.colname.batch = 'date_of_experiment',
                   dataset.colname.lifestage = NULL,
                   dataset.colname.weight = 'weight',
                   dataset.colname.sex
                                              = 'sex',
                                              = FALSE
                   debug
          ###################
          # LinearMixed model (MM) framework
          ##################
          MM_result = OpenStatsAnalysis(
                 OpenStatsList = test_Cont,
                  method = 'MM',
                  MM_fixed = data_point ~ Genotype + Weight
## 2020-03-30 17:01:30. OpenStats loaded.
## 2020-03-30 17:01:30. Checking the input model for including functions ...
## 2020-03-30 17:01:30. Linear Mixed Model (MM framework) in progress ...
## 2020-03-30 17:01:30. Removing possible leading/trailing whitespace from the
variables in the formula ...
## 2020-03-30 17:01:30. Checking duplications in the data model:
## 2020-03-30 17:01:30. Genotype, data_point, Weight
## 2020-03-30 17:01:30. No duplicate found.
## 2020-03-30 17:01:30. Checking for the feasibility of terms and interactions
\#\# 2020-03-30 17:01:30. Formula: data_point ~ Genotype + Weight \#\# 2020-03-30 17:01:30. 1 of 1. Checking for the feasibility of terms and interactions
## 2020-03-30 17:01:30. Checking Genotype ...
## 2020-03-30 17:01:30. Checked model: data_point ~ Genotype + Weight
## 2020-03-30 17:01:30. Check missings in progress ...
## 2020-03-30 17:01:30. Missings in variable 'data_point': 0%
## 2020-03-30 17:01:30. Missings in variable 'Genotype': 0%
## 2020-03-30 17:01:30. Missings in variable 'Weight': 0%
## 2020-03-30 17:01:30. Checking the random effect term ...
## 2020-03-30 17:01:30. Formula: ~1 | Batch
```

```
## 2020-03-30 17:01:30. Lme: Fitting the full model ...
## 2020-03-30 17:01:30. Applied model: lme
\#\# 2020-03-30 17:01:31. The full model successfully applied.
\#\# 2020-03-30 17:01:31. Computing the confidence intervals at the level of 0.95
## 2020-03-30 17:01:31. CI for 'all' term(s) successfully estimated
## 2020-03-30 17:01:31. The specified "lower" model:
## 2020-03-30 17:01:31. ~Genotype + 1
## 2020-03-30 17:01:31. The model optimisation is in progress ...
## 2020-03-30 17:01:31. The direction of the optimisation (backward, forward, both):
hoth
## 2020-03-30 17:01:31. Optimising the model ...
## 2020-03-30 17:01:31. Optimised model: data_point ~ Genotype + Weight
\#\# 2020-03-30 17:01:31. Computing the confidence intervals at the level of 0.95
## 2020-03-30 17:01:31. CI for 'all' term(s) successfully estimated
## 2020-03-30 17:01:31. Testing varHom ...
\#\# 2020-03-30 17:01:31. Computing the confidence intervals at the level of 0.95
## 2020-03-30 17:01:31. CI for 'all' term(s) successfully estimated
## 2020-03-30 17:01:31. VarHom checked out ...
## 2020-03-30 17:01:31. Testing Batch ...
\#\# 2020-03-30 17:01:31. Computing the confidence intervals at the level of 0.95
## 2020-03-30 17:01:31. CI for 'all' term(s) successfully estimated
## 2020-03-30 17:01:31. Estimating effect sizes ...
## 2020-03-30 17:01:31. Total effect sizes estimated:
## 2020-03-30 17:01:31. Quality tests in progress ...
## 2020-03-30 17:01:31. MM framework executed in 0.71 second(s).
```

4.2 Sub-model estimation

OpenStats llows stimateing submodels from an input model. This is called Split model effects in the outputs and it is mainly useful for reporting sex/age-specific etc. effects. This is performed by creating submodels of a full model. For instance, for the input fixed effect (MM_fixed) model $response \sim Genotype + Sex + Weight$ a possible submodel is $response \sim Sex + Sex : Genotype + Weight$ that can be used to estimate sex-specific effects for genotype. This model is then estimated under the configuration of the optimal model. One can turn off Split model effects by setting the fourth element of $MM_optimise$ to FALSE.

An alternative to the analytically estimating the sub-models is to break the input data into splits and run the model on the subset of the data. This can be performed by passing the output of OpenStatsAnalysis function, *OpenStatsMM*, to the function, *OpenStatsComplementarySplit*. This function allows the OpenStatsMM object as input and a set of variable names that split happens on. The example below shows the split on "Sex":

```
dataset = read.csv(fileCon),
                 testGenotype = 'experimental',
                 refGenotype = 'control',
                 dataset.colname.genotype = 'biological_sample_group',
                 dataset.colname.batch = 'date_of_experiment',
                 dataset.colname.lifestage = NULL,
                 dataset.colname.weight = 'weight',
dataset.colname.sex = 'sex',
                 dataset.colname.sex
                                          = FALSE
+
         ##################
         # LinearMixed model (MM) framework
         ##################
         MM_result = OpenStatsAnalysis(
                 OpenStatsList = test_Cont,
                 method = 'MM',
                 MM_fixed = data_point ~ Genotype + Weight,
                 debug
                              = FALSE
         # SplitEffect estimation with respect to the Sex levels
         Spliteffect = OpenStatsComplementarySplit(object = MM_result,
                                                  variables = 'Sex')
## 2020-03-30 17:01:32. Split effects in progress ...
## 2020-03-30 17:01:32. Variable(s) to split:
## 2020-03-30 17:01:32. Sex
## 2020-03-30 17:01:32. Splitting in progress ...
## 2020-03-30 17:01:32. Spliting on Sex ...
## 2020-03-30 17:01:32. Processing the levels: Female
## 2020-03-30 17:01:33. [Successful]
## 2020-03-30 17:01:33. Processing the levels: Male
## 2020-03-30 17:01:33. [Successful]
```

4.3 Reference range plus framework

Reference range plus framework applies to continuous data. In this example, data is extracted from the sample data that accompany the software. Here, "Genotype" is the effect of interest. The response is stored in the variable "data_point" and genotype (Genotype) and sex (Sex) are covariate.

```
>
    library(nlme)
    library(OpenStats)
>
         ###################
>
         # Data preparation
          ##################
>
          ##################
>
>
          # Continuous data - Creating OpenStatsList object
>
          #################
          fileCon = system.file("extdata", "test_continuous.csv",
                                package = "OpenStats")
          test_Cont = OpenStatsList(
                  dataset = read.csv(fileCon),
                  testGenotype = 'experimental',
                  refGenotype = 'control',
                  dataset.colname.genotype = 'biological_sample_group',
```

```
dataset.colname.batch = 'date_of_experiment',
                   dataset.colname.lifestage = NULL,
                   dataset.colname.weight = 'weight',
                   dataset.colname.sex
                                               = 'sex',
                                                = FALSE
          )
    ###############
          # Reference range framework
           ###################
          RR_result = OpenStatsAnalysis(
                   OpenStatsList = test_Cont,
                                 = 'RR',
                   method
                   RR_formula = data_point ~ Genotype + Sex
## 2020-03-30 17:01:34. OpenStats loaded.
## 2020-03-30 17:01:34. Checking the input model for including functions ...
## 2020-03-30 17:01:34. Reference Range Plus (RR framework) in progress ...
## 2020-03-30 17:01:34. Optimisation level:
\#\# 2020-03-30 17:01:34. Estimation of all factor combination effects = TRUE
\#\# 2020-03-30 17:01:34. Estimation of inter level factors for the response = FALSE
\#\# 2020-03-30 17:01:34. The input formula: data_point ~ Genotype + Sex
## 2020-03-30 17:01:34. The reformatted formula for the algorithm: \sim data\_point
+ Genotype + Sex
## 2020-03-30 17:01:34. The probability of the middle area in the distribution:
0.95
## 2020-03-30 17:01:34. Tails probability: 0.025
## 2020-03-30 17:01:34. Formula to calculate the tail probabilities: 1-(1-x)/2,
(1-x)/2 where x = 0.95
## 2020-03-30 17:01:34. Discritizing the continuous data into discrete levels.
The quantile = 0.975
## 2020-03-30 17:01:34. Stp 1. Low versus Normal/High
## 2020-03-30 17:01:34. Removing possible leading/trailing whitespace from the
variables in the formula ...
## 2020-03-30 17:01:34. Preparing the reference ranges ...
## 2020-03-30 17:01:34. Preparing the data for the variable: Genotype
## 2020-03-30 17:01:34. Reference level is set to 'control'
## 2020-03-30 17:01:34. Initial quantiles for cutting the data
## 2020-03-30 17:01:34. Probs: 0, 0.025, 1
## 2020-03-30 17:01:34. N.reference: 397
## 2020-03-30 17:01:34. Quantiles: 6.04, 8.08, 17.73
## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): Low = 0.02518892,
NormalHigh = 0.97481108
## 2020-03-30 17:01:34. Spliting on Sex ...
## 2020-03-30 17:01:34. Preparing the data for the combined effect: Female
## 2020-03-30 17:01:34. Reference level is set to 'control'
## 2020-03-30 17:01:34. Initial quantiles for cutting the data
## 2020-03-30 17:01:34. Probs: 0, 0.025, 1
## 2020-03-30 17:01:34. N.reference: 196
## 2020-03-30 17:01:34. Quantiles: 7.579, 9.267, 17.73 ## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): Low = 0.0255102,
NormalHigh = 0.9744898
## 2020-03-30 17:01:34. Preparing the data for the combined effect: Male ## 2020-03-30 17:01:34. Reference level is set to 'control' ## 2020-03-30 17:01:34. Initial quantiles for cutting the data ## 2020-03-30 17:01:34. Probs: 0, 0.025, 1 ## 2020-03-30 17:01:34. N.reference: 201
## 2020-03-30 17:01:34. Quantiles: 6.04, 7.673, 15.645
```

```
## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): Low = 0.02985075,
NormalHigh = 0.97014925
## 2020-03-30 17:01:34. Stp 2. Low/Normal versus High ## 2020-03-30 17:01:34. Removing possible leading/trailing whitespace from the
variables in the formula ...
## 2020-03-30 17:01:34. Preparing the reference ranges ...
## 2020-03-30 17:01:34. Preparing the data for the variable: Genotype
## 2020-03-30 17:01:34. Reference level is set to 'control'
## 2020-03-30 17:01:34. Initial quantiles for cutting the data
## 2020-03-30 17:01:34. Probs: 0, 0.975, 1
## 2020-03-30 17:01:34. N.reference: 397
## 2020-03-30 17:01:34. Quantiles: 6.04, 14.5, 17.73
## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): LowNormal
= 0.97481108, High = 0.02518892
## 2020-03-30 17:01:34. Spliting on Sex ...
## 2020-03-30 17:01:34. Preparing the data for the combined effect: Female
## 2020-03-30 17:01:34. Reference level is set to 'control'
## 2020-03-30 17:01:34. Initial quantiles for cutting the data
## 2020-03-30 17:01:34. Probs: 0, 0.975, 1
## 2020-03-30 17:01:34. N.reference: 196
## 2020-03-30 17:01:34. Quantiles: 7.579, 14.744, 17.73
## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): LowNormal
= 0.9744898, High = 0.0255102
\#\# 2020-03-30 17:01:34. Preparing the data for the combined effect: Male
## 2020-03-30 17:01:34. Reference level is set to 'control' ## 2020-03-30 17:01:34. Initial quantiles for cutting the data ## 2020-03-30 17:01:34. Probs: 0, 0.975, 1
## 2020-03-30 17:01:34. N.reference: 201
## 2020-03-30 17:01:34. Quantiles: 6.04, 13.527, 15.645
## 2020-03-30 17:01:34. Detected percentiles in the data (8 decimals): LowNormal
= 0.97014925, High = 0.02985075
## 2020-03-30 17:01:34. Fisher exact test with 1500 iteration(s) in progress ...
## 2020-03-30 17:01:34. Analysing Low vs NormalHigh ...
## 2020-03-30 17:01:34. Analysing LowNormal vs High ...
## 2020-03-30 17:01:34. RR framework executed in 0.31 second(s).
```

4.4 Fisher's exact test framework

Fisher's Exact test framework applies to categorical data. In this example, data is extracted from the sample data that accompany the software. Here, Genotype is the effect of interest. The response is stored in the variable *category* and Genotype and Sex are the covariates.

```
library(nlme)
   library(OpenStats)
>
    #################
>
         # Categorical data - Creating OpenStatsList object
>
         #################
         fileCat = system.file("extdata", "test_categorical.csv",
                                 package = "OpenStats")
         test_Cat = OpenStatsList(
                 dataset
                          = read.csv(fileCat, na.strings = '-'),
                 testGenotype = 'Aff3/Aff3',
                 refGenotype = '+/+',
                 dataset.colname.genotype = 'Genotype',
                 dataset.colname.batch = 'Assay.Date',
                 dataset.colname.lifestage = NULL,
                  dataset.colname.weight = 'Weight',
```

```
dataset.colname.sex
                                              = 'Sex'.
                                               = FALSE
          )
    #################
          # Fisher's exact test framework
          ################
          FE_result = OpenStatsAnalysis(
                   OpenStatsList = test_Cat,
                           = "FE",
                   method
                   FE_formula = Thoracic.Processes ~ Genotype + Sex
## 2020-03-30 17:01:34. OpenStats loaded.
## 2020-03-30 17:01:34. Checking the input model for including functions ...
## 2020-03-30 17:01:34. Fisher Exact Test (FE framework) in progress ...
## 2020-03-30 17:01:34. Optimisation level:
## 2020-03-30 17:01:34. Estimation of all factor combination effects = TRUE
## 2020-03-30 17:01:34. Estimation of inter level factors for the response = FALSE
## 2020-03-30 17:01:34. The input formula: Thoracic.Processes ~ Genotype + Sex
## 2020-03-30 17:01:34. The reformatted formula for the algorithm: ~Thoracic.Processes
+ Genotype + Sex
## 2020-03-30 17:01:34.
                          Top framework: FE
## 2020-03-30 17:01:34. Fisher exact test with 1500 iteration(s) in progress ...
## 2020-03-30 17:01:34. Check missings in progress ...
## 2020-03-30 17:01:34. Missings in variable 'Thoracic.Processes': 0.32%
## 2020-03-30 17:01:34. Missings in variable 'Genotype': 0%
## 2020-03-30 17:01:34. Missings in variable 'Sex': 0%
\#\# 2020-03-30 17:01:34. Removing possible leading/trailing whitespace from the
variables in the formula ...
## 2020-03-30 17:01:34. Step 1. Testing "Thoracic.Processes"
## 2020-03-30 17:01:34. The data (variable(s) = Thoracic.Processes) contain 2 missing(s)
## 2020-03-30 17:01:34. Missing data removed
## 2020-03-30 17:01:34. Testing for the main effect:
## 2020-03-30 17:01:34. Testing for the main effect:
## 2020-03-30 17:01:34. Combined effects in progress ...
## 2020-03-30 17:01:34. Splitting in progress ...
## 2020-03-30 17:01:34. Spliting on Genotype ...
## 2020-03-30 17:01:34. Spliting on Sex ...
## 2020-03-30 17:01:34. Shrinking in progress ...
## 2020-03-30 17:01:34. Dichotomising the final tables ...
## 2020-03-30 17:01:34. Finalising the tables ...
## 2020-03-30 17:01:34. Testing for the combined effects ...
## 2020-03-30 17:01:34. Step 2. Testing "Genotype"
## 2020-03-30 17:01:34. Testing for the main effect:
## 2020-03-30 17:01:34. Total tested categories = 2:
                                                         Thoracic.Processes, Genotype
## 2020-03-30 17:01:34. Total tests = 7
## 2020-03-30 17:01:34. FE framework executed in 0.05 second(s).
```

4.5 OpenStatsAnalysis output object

OpenStatsAnalysis output consists of three elements namely, *input*, *output* and *extra*. The *input* object encapsulate the input parameters to the function, *output* hold the analysis results and the *extra* keeps some extra processes on the data/model. Below is an example output from the Reference Rage plus framework:

```
>
   library(nlme)
>
    library(OpenStats)
>
          ##################
>
          # Data preparation
          #################
          #################
          # Continuous data - Creating OpenStatsList object
          ##################
          fileCon = system.file("extdata", "test_continuous.csv",
>
                                package = "OpenStats")
          test_Cont = OpenStatsList(
                 dataset = read.csv(fileCon),
                  testGenotype = 'experimental',
                  refGenotype = 'control',
                 dataset.colname.genotype = 'biological_sample_group',
                 dataset.colname.batch = 'date_of_experiment',
                  dataset.colname.lifestage = NULL,
                  dataset.colname.weight = 'weight',
                                           = 'sex',
                  dataset.colname.sex
                                            = FALSE
+
          )
    ##################
>
>
        # Reference range framework
>
          ###################
         RR_result = OpenStatsAnalysis(
                 OpenStatsList = test_Cont,
                  method = 'RR',
RR_formula = data_point ~ Genotype + Sex,
debug = FALSE
          lapply(RR_result, names)
## $output
## [1] "SplitModels"
##
## $input
                         "data"
## [1] "OpenStatsList"
                            "formula"
                                                "depVariable"
                                                                     "rep"
## [5] "method"
                                                                     "ci_level"
                                                "prop"
## [9] "refLevel"
                            "full_comparisions"
##
## Sextra
## [1] "Cleanedformula" "TransformedRRprop"
          #lapply(RR_result$output,names)
```

5 Summary and export

OpenStats package stores the results of statistical analyses in the *OpenStatsMM/RR/FE* object. The standard *summary* function can be applied to print out a summary table. The function *OpenStatsReport* can be used to create a table of detailed summary in the form of either list or JSON. The following is an example of the summary output of the MM framework. The same function applied to the FE and RR output objects.

```
> summary(MM_result) ## 2020-03-30 17:01:35. Working on the summary table ...
```

```
##
##
##
## Statistic
                          Value
## Applied framework
                          Linear Mixed Model framework, LME, including Weight
## Final model
                          data_point ~ Genotype + Weight
## ......
                          ## Tested Gene
                          experimental
## Reference Gene
                          control
## ......
## Sexual dimorphism detected? FALSE, Genotype-Sex interaction is not part of the input (it
## ......
                          ## Genotype contribution overall 0.343064998063529
## Genotype contribution Females -
## Genotype contribution Males
## ......
                          ## LifeStage contribution
## Genotype contribution Early
## Genotype contribution Late
## .......
## Sex contribution
                          0
## Body weight contribution
```

OpenStatsReport function was developed for large scale application where automatic implementation is require. Following is the JSON output of the function from an *OpenStatsMM* object:

```
OpenStatsReport(
            object = MM_result,
            JSON = TRUE,
            RemoveNullKeys = TRUE,
            pretty = TRUE
##
     "Applied method": "Linear Mixed Model framework, LME, including Weight",
##
     "Dependent variable": "data_point",
##
     "Batch included": true,
##
    "Residual variances homogeneity": false,
##
##
     "Genotype contribution": {
##
      "Overall": 0.343064998063529,
##
      "Sexual dimorphism detected": {
##
         "Criteria": false,
##
         "Note": "Genotype-Sex interaction is not part of the input (it is not part of the fin
##
      }
##
##
     "Genotype estimate": {
      "Value": -0.502512854643372,
##
##
       "Confidence": {
##
         "Genotypeexperimental lower": -1.54340602996328,
##
         "Genotypeexperimental upper": 0.538380320676539
##
##
       "Level": 0.95
##
##
     "Genotype standard error": 0.529316719407687,
     "Genotype p-value": 0.343064998063529,
##
##
     "Genotype percentage change": {
```

```
##
       "control Genotype": -4.66480574285704,
##
       "experimental Genotype": 8.36890228977311
##
    },
##
     "Genotype effect size": {
##
       "Value": 0.572742111514943,
       "Variable": "Genotype",
##
       "Model": "data_point ~ Genotype",
##
       "Type": "Mean differences",
##
       "Percentage change": {
##
##
         "control Genotype": -4.66480574285704,
##
         "experimental Genotype": 8.36890228977311
##
##
     "Weight estimate": {
##
      "Value": -0.390270185772556,
##
##
       "Confidence": {
##
         "Weight lower": -0.432146253204354,
         "Weight upper": -0.348394118340757
##
##
       "Level": 0.95
##
##
##
     "Weight standard error": 0.0212948870837609,
##
     "Weight p-value": 0,
##
     "Weight effect size": {
##
       "Value": -0.598034137811873,
       "Variable": "Weight",
##
##
       "Model": "data_point ~ Weight",
       "Type": "Standardized coefficient",
##
       "Percentage change": {
##
         "Weight": -59.8034137811872
##
##
##
##
     "Gp1 genotype": "control",
##
     "Gp1 Residuals normality test": {
       "P-value": 0.0471145158513991,
##
      "Unique N": 397,
##
##
      "N": 397,
##
       "Unique N/N percent": 100,
##
       "Mean": 4.89284218374938e-015,
##
       "SD": 1.00381856105265,
       "Test": "Shapiro",
##
##
       "Note": "Cautions require when too many duplicates exist in data."
##
##
     "Gp2 genotype": "experimental",
##
     "Gp2 Residuals normality test": {
       "P-value": 0.999989937478505,
##
##
       "Unique N": 13,
##
       "N": 13,
##
       "Unique N/N percent": 100,
       "Mean": 7.37870468518433e-015,
##
##
       "SD": 0.966087192271611,
       "Test": "Shapiro",
##
##
       "Note": "Cautions require when too many duplicates exist in data."
\#\ \#
##
     "Intercept estimate": {
      "Value": 18.7225415243191,
##
       "Confidence": {
##
         "(Intercept) lower": 17.8369200640355,
##
```

```
##
       "(Intercept) upper": 19.6081629846027
##
       },
       "Level": 0.95
##
##
     },
##
     "Intercept standard error": 0.450357690019706,
##
     "Intercept p-value": 0,
##
     "Additional information": {
       "Data": {
##
         "Data signature": "data_point:[n=410,mean=11.0068452195,sd=1.7142008997],Genotype:[cd
##
##
         "Variability": {
##
           "Value": 0.995121951219512,
##
           "Type": "Total unique response divided by total number of response"
##
         },
##
         "Summary statistics": {
           "control": {
##
##
             "Count": 397,
             "Mean": 11.0319801511335,
##
             "SD": 1.70415051295418,
##
             "Normality test": {
##
##
               "P-value": 0.0425147176738847,
##
               "Unique N": 395,
##
               "N": 397,
##
               "Unique N/N percent": 99,
##
               "Mean": 11.0319801511335,
##
               "SD": 1.70415051295418,
##
               "Test": "Shapiro",
               "Note": "Cautions require when too many duplicates exist in data."
##
##
             }
##
           "experimental": {
##
             "Count": 13,
##
             "Mean": 10.2392630769231,
##
             "SD": 1.91256805527448,
##
##
             "Normality test": {
               "P-value": 0.41774224953246,
##
               "Unique N": 13,
##
##
               "N": 13,
##
               "Unique N/N percent": 100,
##
               "Mean": 10.2392630769231,
##
               "SD": 1.91256805527448,
               "Test": "Shapiro",
##
##
               "Note": "Cautions require when too many duplicates exist in data."
##
##
##
         }
##
       },
##
       "Analysis": {
##
         "Model setting": {
##
           "Initial model": {
             "Formula": "data_point ~ Genotype + Weight",
##
             "Random effect": "~1 | Batch"
##
##
           },
##
           "Final model": {
             "Formula": "data_point ~ Genotype + Weight",
##
             "Random effect": "~1 | Batch"
##
##
##
         "Is model optimised": "Fixed term = TRUE, Weight term = TRUE, Random effect term = TR
##
```

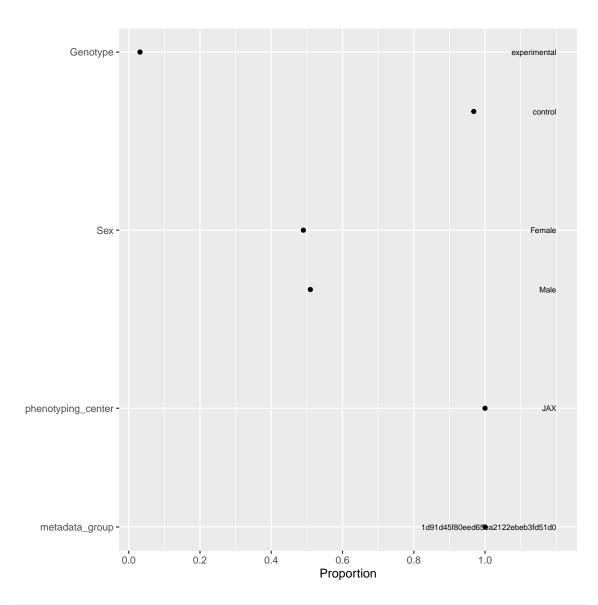
```
"Multibatch in analysis": "Dataset contains multiple batches",
##
##
         "Gender included in analysis": "Both sexes included; Female[201], Male[209]",
         "Effect sizes": {
##
##
           "Genotype": {
##
             "Value": 0.572742111514943,
             "Variable": "Genotype",
##
             "Model": "data_point ~ Genotype",
##
             "Type": "Mean differences",
##
             "Percentage change": {
##
               "control Genotype": -4.66480574285704,
##
##
               "experimental Genotype": 8.36890228977311
##
           },
##
           "Weight": {
##
             "Value": -0.598034137811873,
##
##
             "Variable": "Weight",
##
             "Model": "data_point ~ Weight",
             "Type": "Standardized coefficient",
##
##
             "Percentage change": {
               "Weight": -59.8034137811872
##
##
             }
##
          }
##
         },
##
         "Other residual normality tests": {
##
           "Genotype": {
##
             "control": {
##
               "P-value": 0.0471145158513991,
               "Unique N": 397,
##
               "N": 397,
##
               "Unique N/N percent": 100,
##
               "Mean": 4.89284218374938e-015,
##
               "SD": 1.00381856105265,
##
               "Test": "Shapiro",
##
               "Note": "Cautions require when too many duplicates exist in data."
##
##
             },
             "experimental": {
##
##
               "P-value": 0.999989937478505,
##
               "Unique N": 13,
##
               "N": 13,
##
               "Unique N/N percent": 100,
               "Mean": 7.37870468518433e-015,
##
               "SD": 0.966087192271611,
##
               "Test": "Shapiro",
##
##
               "Note": "Cautions require when too many duplicates exist in data."
##
##
           },
##
           "Overall": {
             "P-value": 0.0502552737582412,
##
##
             "Unique N": 410,
             "N": 410,
##
             "Unique N/N percent": 100,
##
             "Mean": 4.97155881841697e-015,
##
             "SD": 1.00150249979038,
##
             "Test": "Shapiro",
##
             "Note": "Cautions require when too many duplicates exist in data."
##
##
##
         }
##
```

```
## }
## }
```

6 Graphics

Graphics in *OpenStats* are as easy as calling the **plot**() function on a OpenStatsList or the OpenStatsMM/FE/RR object. Calling the plot function on the OpenStatsList object is shown below:

```
> library(OpenStats)
   ####################
         file = system.file("extdata", "test_continuous.csv",
>
                            package = "OpenStats")
+
   ######################
>
   # OpenStatsList object
   #####################
   testGenotype = 'experimental',
        refGenotype = 'control',
dataset.colname.batch = 'date_of_experiment',
        dataset.colname.genotype = 'biological_sample_group',
        dataset.colname.sex = 'sex',
        dataset.colname.weight = 'weight',
         debug = FALSE
  )
        plot (OpenStatsList)
## 2020-03-30 17:01:35. Working on the plot ...
```

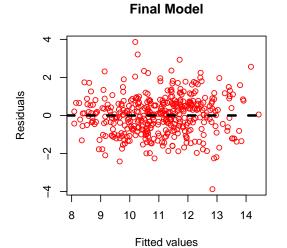


```
summary(
            OpenStatsList,
            OpenStatishist,
style = 'grid',
varnumbers = FALSE,  # See more options ?summarytools::dfSummary
graph.col = FALSE,  # Do not show the graph column
             valid.col = FALSE
## 2020-03-30 17:01:35. Working on the summary table ...
## Data Frame Summary
##
## Dimensions: 410 \times 6
## Duplicates: 320
##
## +---
## | Variable | Stats / Values | Freqs (% of Valid) | Missing |
## +=======
## | Genotype
                      | 1. control
                                                              | 397 (96.8%) | 0
```

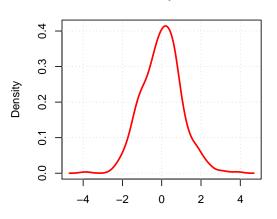
##	[factor]	2. experimental	13 (3.2%)	(0%)
## ## ## -	Sex [factor]	1. Female 2. Male	201 (49.0%)	0
## ## ## ## ## ## ##	Batch [factor] 	1. 2012-07-23T00:00:00Z 2. 2012-07-30T00:00:00Z 3. 2012-08-06T00:00:00Z 4. 2012-08-13T00:00:00Z 5. 2012-08-20T00:00Z 6. 2012-11-26T00:00:00Z 7. 2012-12-24T00:00:00Z 8. 2013-01-02T00:00Z 9. 2013-01-15T00:00:00Z 10. 2013-01-21T00:00:00Z	10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 10 (2.4%) 9 (2.2%) 4 (1.0%) 318 (77.6%)	0 (0%)
## ## ## ##	age_in_weeks [integer] 		6 : 8 (1.9%)	0 (0%)
##	phenotyping_center [factor]	1. JAX 	410 (100.0%)	0
## + # + # + # + # + # + # + # + # + #	[factor]	1. 1d91d45f80eed65ea2122ebeb	410 (100.0%)	0
# # -	T			

There are also graphics for the OpenStatsMM/FE/RR. Below shows an example for the OpenStatsMM output:

```
> plot(MM_result, col = 2)
## 2020-03-30 17:01:36. Working on the plot ...
```

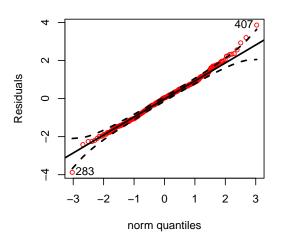


Final Model: Density of the residuals

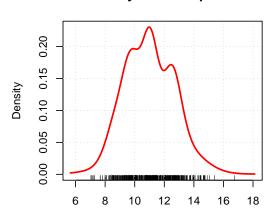


Residuals - [Shapiro] p-value = 0.05025527

Final Model: Normal Q-Q of the residuals



Density of the response



data_point (n = 410) - [Shapiro] p-value = 0.0398488