MarineSDMs

Marine Species Distribution Models

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# 1. Introduction

## 1.1 Background

The best available global distributions are presently AquaMaps (Kaschner et al. 2006; Ready et al. 2010) with supplementation by IUCN RedList range maps[[1]](#footnote-20). These have been used to calculate the biodiversity within national waters (Halpern et al. 2012) as well as beyond in the high seas (Visalli et al. 2020).

## 1.2 Goals

This book aims to capture the overview and details of modeling species distributions in the marine environment for the purposes of advancing the status quo of global and U.S. national species distributions along the following dimensions:

1. **Space**  
   The current AquaMaps distributions are º (~55 km at equator), whereas the best available global bathymetry is º (< 0.5 km).
2. **Time**  
   The current AquaMaps distributions are based on static climatic averages over all seasons, which does not capture temporal dynamics: seasonally within a year, nor long-term climate change trends. This will necessitate sampling the environment contemporaneously with species observations before fitting the model and predicting to different environmental snapshots.
3. **Environment**  
   Other environmental variables besides the initial physiographic (depth) and oceanographic (temperature, chlorophyll, primary productivity and ice) may elicit an improved statistical fit, related to species’ environmental niche. Some candidates include: temperature fronts, eddy kinetic energy, distance from shore, distance from shelf.
4. **Biology**  
   Where sufficient observations exist, additional models should be developed highlighting differences between:
   * **Life stage**, e.g. larval vs adult.
   * **Gender** where varies, such as male sperm whales being more cosmopolitan.
   * **Subpopulations** for understanding metapopulation dynamics
   * **Behavior**, such as migrating, feeding or breeding.

By definition MBONMarine Biodiversity Observation Network; see MarineBON.org is a network, so this is inclusive of and meant for all participants.

## 1.3 Motivations

* [AquaMaps.org](https://aquamaps.org/)  
  AquaMaps (Kaschner et al. 2006; Ready et al. 2010) represents a massive amount of work to gather parameters for >33.5K marine species, including areas to mask out.
* <OBIS.org>  
  The Ocean Biogeographic Information System (Klein et al. 2019; Grassle 2000) is the central portal for continuously added observations with extra flags for quality control, all of which makes marine SDMs possible.
* **Modeling** methods have dramatically improved over time and are ripe for fresh application. The R package dismo originally came came out with an [SDM vignette](https://cran.r-project.org/web/packages/dismo/vignettes/sdm.pdf) as a practical supplement to their excellent review of SDMs (Elith and Leathwick 2009) and using the Maxent algorithm (Elith et al. 2011). The raster package furthered that (raster [sdm](https://rspatial.org/raster/sdm/)) and now there’s [terra sdm](https://rspatial.org/sdm/). Alongside these developments has been a boon of cloud-computing, particularly Google Earth Engine (Gorelick et al. 2017; Campos et al. 2023), allowing for dense global raster processing.
* The world is quickly moving towards a future trying to conserve 30% of the oceans by 2030, so called “[**30 by 30**](https://en.wikipedia.org/wiki/30_by_30)”. In the U.S., this is [America the Beautiful](https://www.noaa.gov/america-the-beautiful) initiative (Carroll, Noss, and Stein 2022) for which MBONMarine Biodiversity Observation Network; see MarineBON.org is well poised to inform (Fautin et al. 2010; Muller-Karger et al. 2018). We need biodiversity indicators to track progress. This push for conservation is driven by increasing impacts of **climate change**, as evidenced by marine heatwaves and shifts in population distributions.

## 1.4 Process

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| Figure 1.1: Diagram of SDM data preparation and model fitting. |

## 1.5 Contribute

We very much welcome your feedback, contributions and collaboration. As soon as you contribute, we will add you to to the authors list. Here are a few ways to contribute from least to most involved:

1. Email Ben (ben@ecoquants.com) with any suggestions, including suggested revisions of this online book.

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| * Note |
| * Note that you can download this entire book as:   + Adobe Acrobat [pdf](https://marinebon.github.io/MarineSDMs/MarineSDMs.pdf) to add annotations; or   + Microsoft Word [docx](https://marinebon.github.io/MarineSDMs/MarineSDMs.docx) to edit with Track Changes on. * These are available in the upper left navigation menu by clicking the download icon . |

1. Submit a [New Issue](https://github.com/marinebon/MarineSDMs/issues/new/choose) on Github.
2. Click on “ Edit this Page” in the upper right. If you have a Github account, then you can fork this repository from owner “marinebon” to your username, edit the page(s) and submit a pull request. See [Hello World - GitHub Docs](https://docs.github.com/en/get-started/quickstart/hello-world).
3. If you are a regular contributor, you can be added to the collaborators of this repository to push changes directly (without needing a pull request).

## 1.6 Features

This Quarto book has a few cool features:

* Multiple formats  
  From the singe set of source Quarto documents (\*.qmd), several output formats are rendered: html, pdf, docx. This is particularly helpful when suggesting changes. It also lends itself well to being carved into manuscripts.
* Self-rendering  
  Github hosts the web pages (\*.html), which get rendered from the source code (\*.qmd) using a Github Action. So edits can be made simply through the web interface and all outputs get updated (html, pdf, docx). It also ensures the reproducibility of the document with a common setup environment.
* Mermaid diagrams  
  e.g., [Figure 1.1](#fig-process), [Figure 3.1](#fig-prep), [Figure 7.1](#fig-model)
* Quarto document listings
* References
* Glossary
* Search

# 2. Prepare

Prepare observations and environmental data for modeling

# 3. Prepare

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| Figure 3.1: Diagram of SDM data preparation for model fitting. |

* **obs**  
  observations: occurrences from OBIS; masked by FAO regions defined by AquaMaps (Skyttner 2020)
  + **presence**  
    OBIS: species occurrence
  + **absence**  
    OBIS not-species, but same family
* **env**  
  environment
* **tbl**  
  table of observations (presence and absence) with environmental values

## 3.1 Environmental Predictors

### 3.1.1 Physiographic

* depth  
  Bathymetric Depth
* d2coast  
  Distance to Coast
* d2shelf  
  Distance to Shelf

### 3.1.2 Time Varying

* vgpm  
  Vertically integrated primary Productivity model

### 3.1.3 Depth & Time Varying

* temp  
  Temperature, either sea-surface temperature (SST) or some modeled product from HyCOM, ROMS or Copernicus
* salin  
  Salinity

# 4. Occurrences

Fetch presence observations and filter for quality control

To describe:

* robis
* Filter based on quality flags
* Remove outliers
  + [eks](https://cran.r-project.org/web/packages/eks/vignettes/tidysf_kde.html)  
    *Tidy and Geospatial Kernel Smoothing for spatially filtering outlier observations*

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| Figure 4.1: Source: Kernel density estimates for tidy and geospatial data in the eks package |

## 4.1 Fetch OBIS

## 4.2 Filter occurrences

# 5. Pseudo-absences

Generate pseudo-absence or background environmental values to compare with occurrence environment

Describe various strategies for generating pseudo-absences.

* [Pseudo-absences • biomod2](https://biomodhub.github.io/biomod2/articles/vignette_pseudoAbsences.html)
  + (Barbet-Massin et al. 2012)

## 5.1 All background

A common Maxent strategy is to feed all background points into Maxent, and then to use the resulting distribution as a null model. This is the default strategy in Maxent (Phillips et al. 2017; Phillips, Anderson, and Schapire 2006; Phillips and Dudík 2008).

## 5.2 Mask by FAO areas

The FAO areas applicable to species are included in the aquamapsdata, presumably from evaluating OBIS observations and the literature.

## 5.3 Use occurrences from same Family, different species

By using the same family, we can be sure that the pseudo-absences are ecologically similar to the species of interest.

# 6. Environment

Extract environmental predictors (static and/or dynamic) from various sources for observations (presence and pseudo-absence)

Environmental data are used to fit the model and predict distribution onto the seascape, e.g. [Table 6.1](#tbl-Roberts2016-env).

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| Table 6.1: Example of environmental predictors from Roberts et al. (2016).  librarian::shelf(  here, knitr, readr) library(here) library(knitr) library(readr)  d <- read\_csv(  here("data/Roberts-2016\_env-predictors.csv"),  show\_col\_types = F)  options(knitr.kable.NA = '') kable(d, format="pipe")   | **Type** /Covariates | Resolution | Time range | Description | | --- | --- | --- | --- | | **Physiographic** |  |  |  | | Depth, Slope | 30 arc sec |  | Seafloor depth and slope, derived from SRTM30-PLUS global bathymetry20 | | DistToShore, DistTo125m, DistTo300m, DistTo1500m | 30 arc sec |  | Distance to the closest shoreline, excluding Bermuda and Sable Island, and various ecologically-relevant isobaths20 | | DistToCanyon, DistToCanyon OrSeamount | 30 arc sec |  | Distance to the closest submarine canyon, and to the closest canyon or seamount21 | | **SST & Winds** |  |  |  | | SST, DistToFront | 0.2°, daily | 1991-2014 | Foundation sea surface temperature (SST), from GHRSST Level 4 CMC SST22, and distance to the closest SST front identified with the Canny edge detection algorithm23 | | WindSpeed | 0.25°, daily | 1991-2014 | 30-day running mean of NOAA NCDC 1/4° Blended Sea Winds24 | | **Currents** |  |  |  | | TKE, EKE | 0.25°, daily | 1993-2013 | Total kinetic energy (TKE) and eddy kinetic energy (EKE), from Aviso 1/4° DT-MADT geostrophic currents | | DistToEddy, DistToAEddy, DistToCEddy | 0.25°, weekly | 1993-2013 | Distance to the ring of the closest geostrophic eddy having any (DistToEddy), anticyclonic (DistToAEddy), or cyclonic (DistToCEddy) polarity, from Aviso 1/4° DT-MADT using a revision of the Chelton et al. algorithm25; we tested eddies at least 9, 4, and 0 weeks old | | **Biological** |  |  |  | | Chl | 9 km, daily | 1997-2014 | GSM merged SeaWiFS/Aqua/MERIS/VIIRS chlorophyll (Chl) a concentration26, smoothed with a 3D Gaussian smoother to reduce data loss to < 10% | | VGPM, CumVGPM45, CumVGPM90 | 9 km, 8 days | 1997-2014 | Net primary production (mg C m-2 day-1) derived from SeaWiFS and Aqua using the Vertically Generalized Production Model (VPGM)27; we tested the original 8 day estimates as well as 45 and 90 day running accumulations | | PkPP, PkPB | 0.25°, weekly | 1997-2013 | Zooplankton production (PkPP; g m-2 day-1) and biomass (PkPB; g m-2) from the SEAPODYM ocean model28 | | EpiMnkPP, EpiMnkPB | 0.25°, weekly | 1997-2013 | Epipelagic micronekton production (EpiMnkPP; g m-2 day-1) and biomass (EpiMnkPB; g m-2) from the SEAPODYM model(28) | |

### 6.0.1 Physiographic

* depth  
  Bathymetric Depth
* d2coast  
  Distance to Coast
* d2shelf  
  Distance to Shelf

### 6.0.2 Time Varying

* vgpm  
  Vertically integrated primary Productivity model

### 6.0.3 Depth & Time Varying

* temp  
  Temperature, either sea-surface temperature (SST) or some modeled product from HyCOM, ROMS or Copernicus
* salin  
  Salinity

# 7. Model

Model the distribution of a species

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| Figure 7.1: Diagram of SDM Modeling processes. |

# 8. Split

Split data into training (to fit) and test (to evaluate prediction)

Data is often split so that ~20% of the observations (presence and absence) are set aside from the model fitting to be used for model evaluation.

The k-fold function is often used to split the data into k groups, and then the model is fit k times, each time using a different group as the test data and the remaining groups as the training data.

# 9. Fit

Fit environmental relationship distinguishing presence from absence of species

Model fitting in theory is quite complex, but quite simple in practice, with feeding the prepared data into the modeling function.

However there are MANY modeling techniques from which to choose. For instance check out 238 entries in [6 Available Models | The caret Package](https://topepo.github.io/caret/available-models.html).

# 10. Calibrate

Calibrate model fit, i.e., model selection

The process of refining the model to only the most relevant environmental predictor terms is commonly called “Model Selection.” One of the most cited scientific paper of all time (Akaike 1974) is based on taking a most parsimonious approach to this process – the so called Akaike Information Criteria (AIC).

It is important to avoid using environmental predictors that are correlated with each other, since the effect of a predictor on the response could be the ecologically inverse, the result of explaining variance on the residuals of the other correlated predictor.

# 11. Predict

Predict distribution of the species with environmental relationship from fitted model

The prediction step applies the environmental relationships from the fitted model to a new set of data, typically the seascape of interest, and perhaps with some sort of temporal snapshot (e.g., climatic annual or monthly average).

# 12. Evaluate

Evaluate performance of the predicted model with the test data

Model evaluation uses the set aside test data from the earlier splitting to evaluate how well the model predicts the response of presence or absence. Since the test response data is binary [0,1] and the prediction from the model is continuous [0-1], a threshold needs to be applied to assign to convert the continuous response to binary. This is often performed through a Receiver Operator Characteristic (**ROC**) curve ([Figure 12.1](#fig-rocr)), which evaluates at each threshold the **confusion matrix** ([Table 12.1](#tbl-confusion-matrix)).

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| Table 12.1: Confusion matrix to understand predicted versus observed.   |  |  |  |  | | --- | --- | --- | --- | |  |  | Predicted |  | |  |  | 0 (absence) | 1 (presence) | | Observed | 0 (absence) | True absence | False presence | |  | 1 (presence) | False absence | True presence | |

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| Figure 12.1: ROC curve generated by showing rates of false positive vs false negative as function of changing the threshold value (rainbow colors). Source: [ROCR: visualizing classifier performance in R](https://cran.rstudio.com/web/packages/ROCR/vignettes/ROCR.html) |

From the ROC curve, the area under the curve (**AUC**) is calculated, which is a measure of the model’s ability to distinguish between presence and absence. AUC values range from 0 to 1, with 0.5 being no better than random, and 1 being perfect.

## 12.1 More Resources

* [Classification: ROC Curve and AUC | Machine Learning | Google for Developers](https://developers.google.com/machine-learning/crash-course/classification/roc-and-auc)

# 13. Combine

Combine SDMs from the same or multiple species

We look at combining SDMs to calculate biodiversity based on addressing questions of interest and relevance.

* See joint species distribution models (jSDMs) per Hartig et al (2023, Box 2) that includes species co-occurrence.

# 14. Ensemble

* [biomod2](https://biomodhub.github.io/biomod2/)  
  *Species distribution modeling, calibration and evaluation, ensemble modeling*  
  ![Graphical overview of `biomod2` functions for data formatting, single SDMs, ensemble methods and visualization. Source: [`biomod2`](https://biomodhub.github.io/biomod2/)](data:image/png;base64,)

# 15. Mosaic

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| Figure 15.1: Hierarchy of preferred model outputs based on response type and age. |

This is illustrated well by [Figure 15.1](#fig-sdm-response-hierarchy).

# 16. Taxa

Group SDMs by taxanomy

Taxonomic groups (Tittensor et al. 2010) in the high seas (Visalli et al. 2020) were packaged with simple query statements in the draft R package [gmbi](https://marinebon.github.io/gmbi/articles/calc.html#assign-taxonomic-groups) (global marine biodiversity indicators).

# 17. Indicators

Calculate indicators of ecological or management interest beyond taxonomic groupings

## 17.1 Diversity

Here are the classic diversity indices from the R package vegan:

where is the proportion of species , and is the number of species so that , and is the base of the logarithm.

## 17.2 Endemism

Endemism could be measured as a function of the presence or average of the species range, given by either a global SDM converted to a binary range or using the existing IUCN range maps.

## 17.3 Extinction Risk

This is provided by IUCN RedList, as well as sometimes at a national level, such as NatureServe’s [Conservation Status Ranks](https://www.natureserve.org/conservation-status-assessment) for the U.S.

## 17.4 Functional Importance

## 17.5 Habitat Forming

Habitat forming species, such as coral, mangrove, seagrasses and kelp are especially important for biodiversity and ecosystem services.

## 17.6 Phylogenetic Uniqueness

## 17.7 Richness

## 17.8 Sensitivity

Sensitivity to specific human activities, such as shipping or fishing. Some activities may have different stages of development, such as construction versus operation of offshore wind energy.

## 17.9 Trophic Index

# 18. Share

Metadata standards and portals to share SDMs

# 19. Metadata

Metadata standards for reproducible and stackable SDMs

What standards (Araújo et al. 2019; Kass et al. 2023; Zurell et al. 2020) are required for models to be hosted?

* input
  + input observations
  + environmental predictors  
    and range of values in original observations
  + model type and object
* outputs
  + model object
  + mean prediction
  + measure(s) of uncertainty  
    standard error, standard deviation (sd), confidence intervals (e.g., 5% and 95%), coefficient of variation (cv)…

# 20. Portal

Portal to host and combine for user-specific needs

Can we supplement an existing portal or create a new one to host different types of model outputs and combine them?

What are publishing workflows for existing portals?

* Existing portals used to share SDMs
  + [AquaMaps](https://aquamaps.org/) *Standardized distribution maps for over 33,500 species of fishes, marine mammals and invertebrates*
  + [DisMAP](https://apps-st.fisheries.noaa.gov/dismap/)  
    *Distribution Mapping and Analysis Portal*
  + [OBIS-SEAMAP Model Repository](https://seamap.env.duke.edu/models/)  
    *World Data Center for Marine Mammal, Seabird, Sea Turtle, Shark & Ray Distributions*
  + [NCEI](https://www.ncei.noaa.gov/)  
    *National Centers for Environmental Information (NOAA)*
  + [DataONE](https://www.dataone.org/)  
    *Data Observation Network for Earth*
  + [ArcGIS Online](https://www.arcgis.com/index.html)
  + *Esri’s commercial data sharing platform*
* Candidate portal  
  <MarineSpeciesMaps.org>  
  BDB registered the domain. Similar to:
  + **MarineRegions.org**  
    *spatial authority*
  + **MarineSpecies.org**  
    *taxonomic authority*

# References

Akaike, H. 1974. “A New Look at the Statistical Model Identification.” *IEEE Transactions on Automatic Control* 19 (6): 716723.

Araújo, Miguel B., Robert P. Anderson, A. Márcia Barbosa, Colin M. Beale, Carsten F. Dormann, Regan Early, Raquel A. Garcia, et al. 2019. “Standards for Distribution Models in Biodiversity Assessments.” *Science Advances* 5 (1): eaat4858. <https://doi.org/10.1126/sciadv.aat4858>.

Barbet-Massin, Morgane, Frédéric Jiguet, Cécile Hélène Albert, and Wilfried Thuiller. 2012. “Selecting Pseudo-Absences for Species Distribution Models: How, Where and How Many?” *Methods in Ecology and Evolution* 3 (2): 327–38. <https://doi.org/10.1111/j.2041-210X.2011.00172.x>.

Campos, João C., Nuno Garcia, João Alírio, Salvador Arenas-Castro, Ana C. Teodoro, and Neftalí Sillero. 2023. “Ecological Niche Models Using MaxEnt in Google Earth Engine: Evaluation, Guidelines and Recommendations.” *Ecological Informatics* 76 (September): 102147. <https://doi.org/10.1016/j.ecoinf.2023.102147>.

Carroll, C., R. F. Noss, and Bruce A. Stein. 2022. “US Conservation Atlas Needs Biodiversity Data.” *Science* 376 (6589): 144–45. <https://doi.org/10.1126/science.abo0526>.

Elith, Jane, and John R. Leathwick. 2009. “Species Distribution Models: Ecological Explanation and Prediction Across Space and Time.” *Annual Review of Ecology, Evolution, and Systematics* 40 (1): 677–97. <https://doi.org/10.1146/annurev.ecolsys.110308.120159>.

Elith, Jane, Steven J. Phillips, Trevor Hastie, Miroslav Dudík, Yung En Chee, and Colin J. Yates. 2011. “A Statistical Explanation of MaxEnt for Ecologists.” *Diversity and Distributions* 17 (1): 43–57. <http://dx.doi.org/10.1111/j.1472-4642.2010.00725.x>.

Fautin, Daphne, Penelope Dalton, Lewis S. Incze, Jo-Ann C. Leong, Clarence Pautzke, Andrew Rosenberg, Paul Sandifer, et al. 2010. “An Overview of Marine Biodiversity in United States Waters.” *PLoS ONE* 5 (8): e11914. <https://doi.org/10.1371/journal.pone.0011914>.

Gorelick, Noel, Matt Hancher, Mike Dixon, Simon Ilyushchenko, David Thau, and Rebecca Moore. 2017. “Google Earth Engine: Planetary-Scale Geospatial Analysis for Everyone.” *Remote Sensing of Environment*, Big remotely sensed data: Tools, applications and experiences, 202 (December): 18–27. <https://doi.org/10.1016/j.rse.2017.06.031>.

Grassle, J. Frederick. 2000. “The Ocean Biogeographic Information System (OBIS): An on-Line, Worldwide Atlas for Accessing, Modeling and Mapping Marine Biological Data in a Multidimensional Geographic Context.” *Oceanography* 13 (3): 5–7. <https://www.jstor.org/stable/43924357>.

Halpern, Benjamin S., Catherine Longo, Darren Hardy, Karen L. McLeod, Jameal F. Samhouri, Steven K. Katona, Kristin Kleisner, et al. 2012. “An Index to Assess the Health and Benefits of the Global Ocean.” *Nature*. <https://doi.org/10.1038/nature11397>.

Hartig, Florian, Nerea Abrego, Alex Bush, Jonathan M. Chase, Gurutzeta Guillera-Arroita, Mathew A. Leibold, Otso Ovaskainen, et al. 2023. “Novel Community Data in Ecology-Properties and Prospects.” *Trends in Ecology & Evolution* 0 (0). <https://doi.org/10.1016/j.tree.2023.09.017>.

Kaschner, K., R. Watson, A. W. Trites, and D. Pauly. 2006. “Mapping World-Wide Distributions of Marine Mammal Species Using a Relative Environmental Suitability (RES) Model.” *Marine Ecology Progress Series* 316 (July): 285–310. <https://doi.org/10.3354/meps316285>.

Kass, Jamie M., Gonzalo E. Pinilla-Buitrago, Andrea Paz, Bethany A. Johnson, Valentina Grisales-Betancur, Sarah I. Meenan, Dean Attali, et al. 2023. “Wallace 2: A Shiny App for Modeling Species Niches and Distributions Redesigned to Facilitate Expansion via Module Contributions.” *Ecography* 2023 (3): e06547. <https://doi.org/10.1111/ecog.06547>.

Klein, Eduardo, Ward Appeltans, Pieter Provoost, Hanieh Saeedi, Abigail Benson, Lenore Bajona, Ana Carolina Peralta, and R. Sky Bristol. 2019. “OBIS Infrastructure, Lessons Learned, and Vision for the Future.” *Frontiers in Marine Science* 6. <https://www.frontiersin.org/articles/10.3389/fmars.2019.00588>.

Muller-Karger, Frank E., Patricia Miloslavich, Nicholas J. Bax, Samantha Simmons, Mark J. Costello, Isabel Sousa Pinto, Gabrielle Canonico, et al. 2018. “Advancing Marine Biological Observations and Data Requirements of the Complementary Essential Ocean Variables (EOVs) and Essential Biodiversity Variables (EBVs) Frameworks.” *Frontiers in Marine Science* 5. <https://doi.org/10.3389/fmars.2018.00211>.

Phillips, Steven J., Robert P. Anderson, Miroslav Dudík, Robert E. Schapire, and Mary E. Blair. 2017. “Opening the Black Box: An Open-Source Release of Maxent.” *Ecography* 40 (7): 887–93. <https://doi.org/10.1111/ecog.03049>.

Phillips, Steven J., Robert P. Anderson, and Robert E. Schapire. 2006. “Maximum Entropy Modeling of Species Geographic Distributions.” *Ecological Modelling* 190 (3): 231–59. <https://doi.org/10.1016/j.ecolmodel.2005.03.026>.

Phillips, Steven J., and Miroslav Dudík. 2008. “Modeling of Species Distributions with Maxent: New Extensions and a Comprehensive Evaluation.” *Ecography* 31 (2): 161–75. <https://doi.org/10.1111/j.0906-7590.2008.5203.x>.

Ready, Jonathan, Kristin Kaschner, Andy B. South, Paul D. Eastwood, Tony Rees, Josephine Rius, Eli Agbayani, Sven Kullander, and Rainer Froese. 2010. “Predicting the Distributions of Marine Organisms at the Global Scale.” *Ecological Modelling* 221 (3): 467–78. <https://doi.org/10.1016/j.ecolmodel.2009.10.025>.

Roberts, Jason J., Benjamin D. Best, Laura Mannocci, Ei Fujioka, Patrick N. Halpin, Debra L. Palka, Lance P. Garrison, et al. 2016. “Habitat-Based Cetacean Density Models for the u.s. Atlantic and Gulf of Mexico.” *Scientific Reports* 6 (March): 22615. <https://doi.org/10.1038/srep22615>.

Skyttner, Markus. 2020. “Aquamapsdata: Curated Data from AquaMaps.org.” <https://github.com/raquamaps/aquamapsdata>.

Tittensor, Derek P., Camilo Mora, Walter Jetz, Heike K. Lotze, Daniel Ricard, Edward Vanden Berghe, and Boris Worm. 2010. “Global Patterns and Predictors of Marine Biodiversity Across Taxa.” *Nature* 466 (7310): 1098–1101. <https://doi.org/10.1038/nature09329>.

Visalli, Morgan E., Benjamin D. Best, Reniel B. Cabral, William W. L. Cheung, Nichola A. Clark, Cristina Garilao, Kristin Kaschner, et al. 2020. “Data-Driven Approach for Highlighting Priority Areas for Protection in Marine Areas Beyond National Jurisdiction.” *Marine Policy*, March, 103927. <https://doi.org/10.1016/j.marpol.2020.103927>.

Zurell, Damaris, Janet Franklin, Christian König, Phil J. Bouchet, Carsten F. Dormann, Jane Elith, Guillermo Fandos, et al. 2020. “A Standard Protocol for Reporting Species Distribution Models.” *Ecography* 43 (9): 1261–77. <https://doi.org/10.1111/ecog.04960>.

# Glossary

glossary::glossary\_table(as\_kable=F) |>   
 knitr::kable("pipe", escape = F, row.names = F)

| term | definition |
| --- | --- |
| MBON | Marine Biodiversity Observation Network; see <MarineBON.org> |

# AquaMaps Downscaled

Downscale AquaMaps from 1/2º to GEBCO 1/240º using Google Earth Engine and Shiny.

* [website](https://shiny.marinebon.app/am-fine/)
* [code](https://github.com/marinebon/aquamaps-downscaled/tree/main/sp-map)



# AquaMaps Envelope

Extract and plot AquaMaps environmental envelope, ramp rasters, using R.

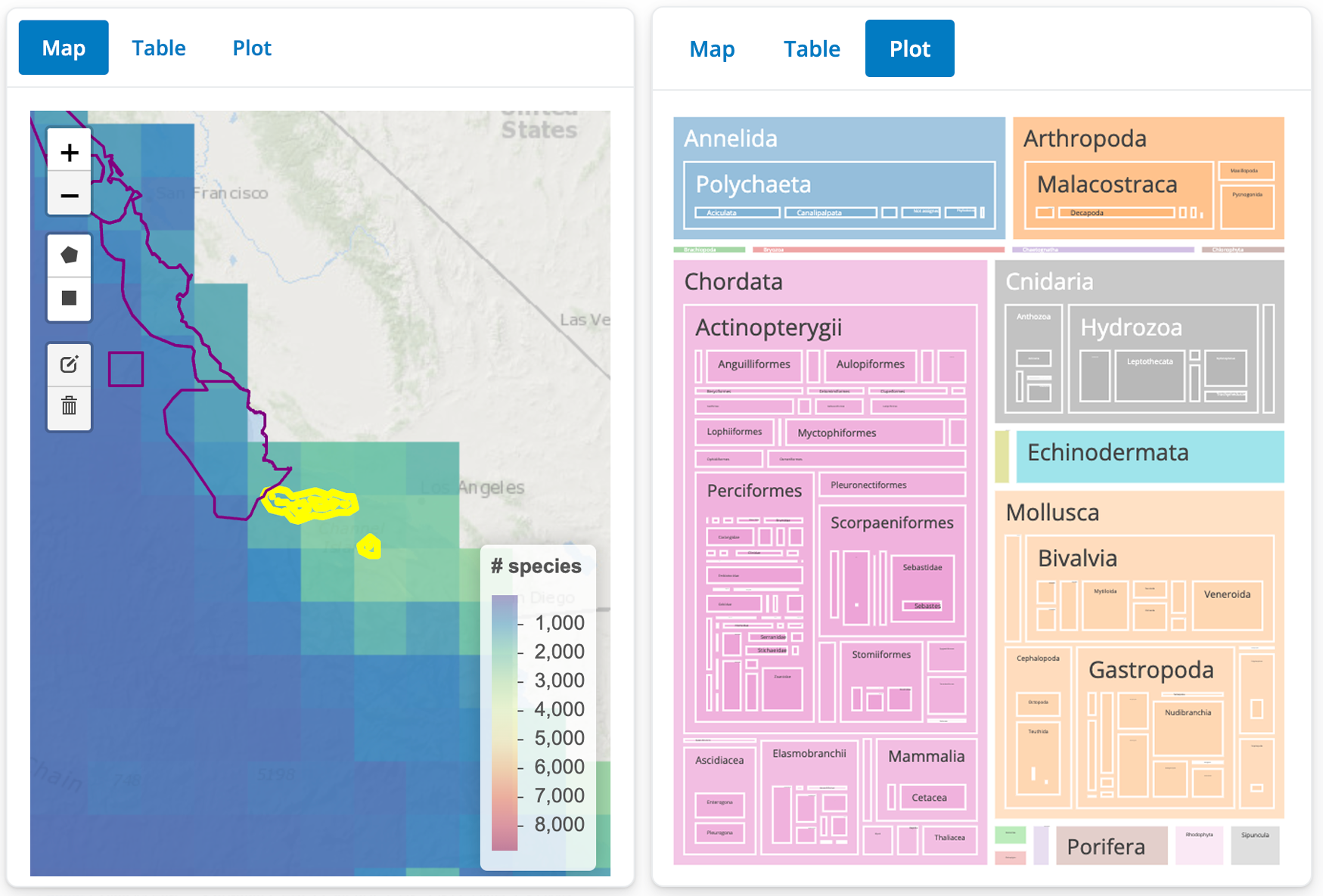
* [website](https://marinebon.github.io/aquamaps-downscaled/)
* [code](https://github.com/marinebon/aquamaps-downscaled/blob/main/index.qmd)



# AquaMaps Treemap

Select a Sanctuary or Draw a polygon to filter to 1/2º AquaMaps species and view as Table or treemap Plot for interactive taxonomic composition.

* [website](https://shiny.marinebon.app/am-sanct/)
* [code](https://github.com/marinebon/aquamapsduckdb/tree/main/inst/app)



This app uses [aquamapsdata](https://raquamaps.github.io/aquamapsdata/articles/intro.html) after translating from sqlite (slow) and raster (deprecated) R functions to duckdb (fast) and terra (superceding) R functions. We can use these distributional data to calculate place-based indicators with anticipation of the distributional data getting improved upon in 2024 by AquaX and others.

# SDM predicts

Predict species distribution of N Atlantic right whale using OBIS occcurrences and predicts package in R.

* [website](https://marinebon.github.io/sdm-explore/sdm_1.html)
* [code](https://github.com/marinebon/sdm-explore/blob/main/sdm_1.qmd)



# OBIS Top Classes

Extract the species with the most numerous observations by unique Class from the OBIS parquet archive in R.

* [website](https://marinebon.github.io/sdm-explore/explore_obis.html)
* [code](https://github.com/marinebon/sdm-explore/blob/main/explore_obis.qmd)



# Software

## R

Most packages have not yet migrated from using the deprecated raster R package to the new terra package, except for [biomod2](https://biomodhub.github.io/biomod2/) (ref?).

* [biomod2](https://biomodhub.github.io/biomod2/)  
  *Species distribution modeling, calibration and evaluation, ensemble modeling*  
  ![Graphical overview of `biomod2` functions for data formatting, single SDMs, ensemble methods and visualization. Source: [`biomod2`](https://biomodhub.github.io/biomod2/)](data:image/png;base64,)
* [eks](https://cran.r-project.org/web/packages/eks/vignettes/tidysf_kde.html)  
  *Tidy and Geospatial Kernel Smoothing for spatially filtering outlier observations*

|  |
| --- |
| Figure 20.1: Source: [Kernel density estimates for tidy and geospatial data in the eks package](https://cran.r-project.org/web/packages/eks/vignettes/tidysf_kde.html) |

* [predicts](Species%20distribution%20modeling,%20calibration%20and%20evaluation,%20ensemble%20modeling)  
  *New R library using terra for predicting from fitted model*

## Python

* [Xarray](https://docs.xarray.dev/en/stable/)  
  Xarray makes working with labelled multi-dimensional arrays in Python simple, efficient, and fun!

## Google Earth Engine

* [XEE](https://github.com/google/Xee)  
  XEE is a new Python package for Earth Engine that provides a set of functions to facilitate the use of Earth Engine API. It is designed to be used in Jupyter notebooks and Google Colab. [Documentation](https://google.github.io/Xee/)

1. IUCN RedList range maps: <https://www.iucnredlist.org/resources/spatial-data-download> [↑](#footnote-ref-20)