

Cellpose + Trackmate

Wednesday, August 21, 2024 11:23 AM

Setting up cellpose -----

- 1.
2. Install Anaconda or Miniconda
3. Install Visual Studio <https://visualstudio.microsoft.com>
4. Install Microsoft C++ Build Tools <https://visualstudio.microsoft.com/visual-cpp-build-tools/>
 - a. The package you want to install is this the one on the workloads tab called: "Desktop Development with C++"
5. Update your graphics driver. We all have NVIDIA -> <https://www.nvidia.com/en-us/drivers/>
6. Install CUDA Toolkit 12.6 : <https://developer.nvidia.com/cuda-downloads>
7. Make a new anaconda environment (name it as you please), BUT MAKE SURE IT'S PYTHON VERSION 3.8.XX

In the command prompt for your new python environment where you want to install cellpose AND cellprofiler, copy and paste **ONE** of these sets of commands (depending if you have a GPU to use with cellpose):

conda activate urenvironment

CPU VERSION: If running cellpose on the CPU use this chunk of code:

```
python -m pip install cellpose[gui]
python -m pip install cellprofiler
```

OR

GPU VERSION: If using a GPU to run cellpose run these lines:

```
python -m pip install cellprofiler
python -m pip install cellpose[gui]
python -m pip install cuda-python
python -m pip uninstall torch -y
python -m pip install torch torchvision torchaudio --index-url https://download.pytorch.org/whl/cu121
```

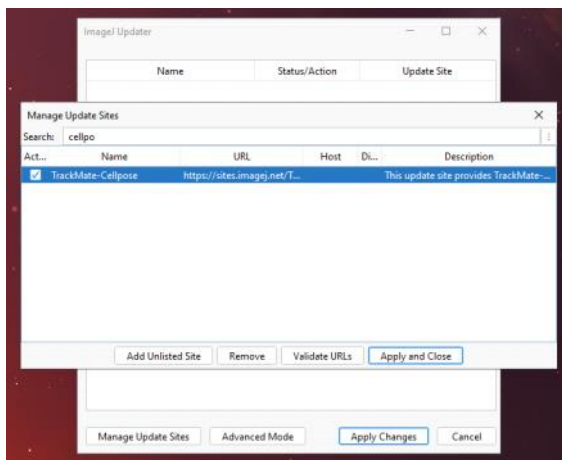
Adding cellpose to TrackMate in Fiji -----

Install Fiji

Open Fiji and go to Help > Update

Once Fiji has updated, click on the bottom left of the new update window : Manage Update Sites

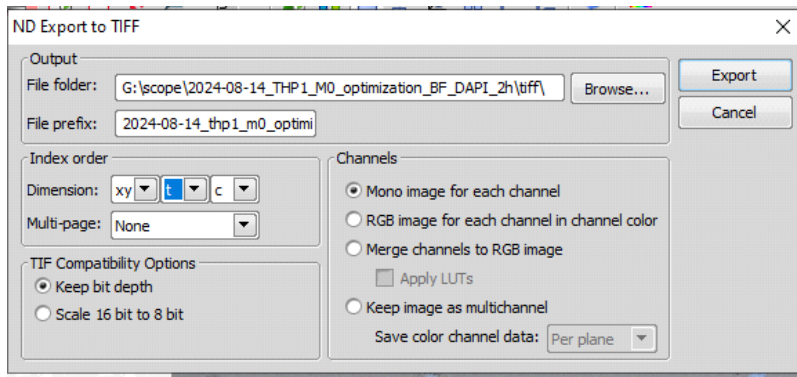
Check the TrackMate-Cellpose box:



Using TrackMate -----

From NIS:

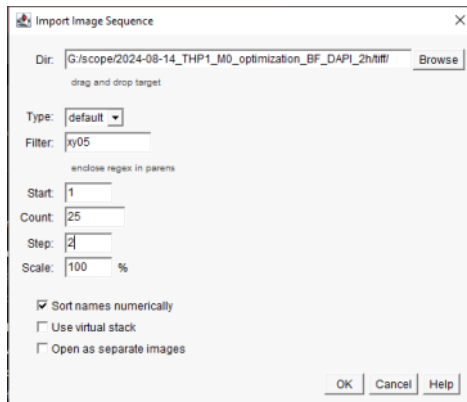
File > Import/Export > ND export to TIFF



I have 2 channels, the brightfield and the DAPI, they're gonna be saved as _c1 and _c2

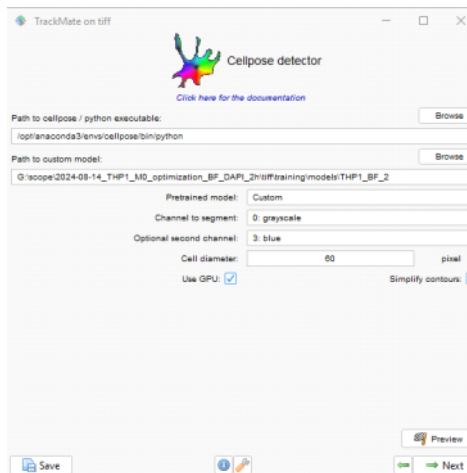
From Fiji:

File > Import > Image Sequence



Your images should open as a video, frame by frame.

Plugins > Track > TrackMate



Path to the cellpose/python executable if installed with anaconda: C:/Users/mhardy/AppData/Local/anaconda3/envs/cellpose/python.exe

```

Configured detector Cellpose detector with settings:
- target channel: 0
- cellpose model: Custom
- cellpose model filepath: G:\scope\2024-08-
14_THP1_M0_optimization_BF_DAPI_2h\tiff\training\models\THP1_BF_2
- simplify contours: true
- use gpu: true
- optional channel 2: 3
- cell diameter: 60.0
- logger: LogPanelLogger
- cellpose python filepath: C:/Users/mhardy/AppData/Local/anaconda3/envs/cellpose/python.exe

Starting detection process using 20 threads.
Saving single time-points.
Running Cellpose with args:
C:/Users/mhardy/AppData/Local/anaconda3/envs/cellpose/python.exe -m cellpose --dir C:
\Users/mhardy/AppData/Local/Temp/TrackMate-Cellpose_5421205060791547441 --chan 0 --chan2 3 --use_gpu
--diameter 60.0 --pretrained_model G:\scope\2024-08-
14_THP1_M0_optimization_BF_DAPI_2h\tiff\training\models\THP1_BF_2 --save_png --no_npy
Reading Cellpose masks.
Converting masks to spots.
Found 2307 spots.
Detection done in 473.7 s.

```

Then explore the options you want to use

- Coloring of edges and tracks
- Filtering of objects and tracks

To export data:

- Click Tracks or Spots
- The new window appears
- Export each to csv

Spots	Label	Spot ID	Track ID	Quality (quality)	X (pixel)	Y (pixel)
Edges	ID5344	5344	0	5,959	1,476.881	143.819
Tracks	ID4977	4977	0	4,592	1,474.478	131.807
	ID6019	6019	0	5,483	1,485.459	131.69
	ID5205	5205	0	5,253	1,464.657	136.893
	ID4967	4967	0	5,011	1,475.054	138.711
	ID6915	6915	1	4,182	1,335.675	846.85
	ID6533	6533	1	5,107	1,352.063	860.301
	ID5768	5768	1	4,768	1,348.846	851.913
	ID5384	5384	1	4,992	1,335.238	847.152
	ID5322	5322	1	4,914	1,337.891	846.465
	ID5976	5976	1	4,765	1,333.808	845.809
	ID5464	5464	1	5,576	1,334.67	844.234
	ID5401	5401	1	4,770	1,339.59	850.238
	ID5981	5981	1	4,911	1,339.951	849.783
	ID5598	5598	1	4,956	1,344.621	848.787

Spots = the cells periphery (in pink)

Edges = the line of the tracks, frame by frame

Tracks = the tracks over time