

MIIN Part 4: Meta-dataset overview

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Filename: MIIN_4_datasetOverview.Rmd

This markdown file does the following tasks: 1. Article selection statistics

2. Number of papers and observations
3. Types of observations
4. Plant species statistics
5. Cover data statistics
6. Trait data statistics
7. Soil measurement statistics
8. Effect size statistics
9. CWM trait value statistics

```
knitr::opts_chunk$set(cache=TRUE)
```

```
citation()
```

```
##
## To cite R in publications use:
##
## R Core Team (2015). R: A language and environment for
## statistical computing. R Foundation for Statistical Computing,
## Vienna, Austria. URL http://www.R-project.org/.
##
## A BibTeX entry for LaTeX users is
##
## @Manual{,
##   title = {R: A Language and Environment for Statistical Computing},
##   author = {{R Core Team}},
##   organization = {R Foundation for Statistical Computing},
##   address = {Vienna, Austria},
##   year = {2015},
##   url = {http://www.R-project.org/},
## }
##
## We have invested a lot of time and effort in creating R, please
## cite it when using it for data analysis. See also
## 'citation("pkgname")' for citing R packages.
```

```
library(plyr)
if(nchar(system.file(package="plyr")) > 0) citation("plyr")
```

```
##
## To cite plyr in publications use:
##
##   Hadley Wickham (2011). The Split-Apply-Combine Strategy for Data
##   Analysis. Journal of Statistical Software, 40(1), 1-29. URL
##   http://www.jstatsoft.org/v40/i01/.
##
## A BibTeX entry for LaTeX users is
##
##   @Article{,
##     title = {The Split-Apply-Combine Strategy for Data Analysis},
##     author = {Hadley Wickham},
##     journal = {Journal of Statistical Software},
##     year = {2011},
##     volume = {40},
##     number = {1},
##     pages = {1--29},
##     url = {http://www.jstatsoft.org/v40/i01/},
##   }
```

```
library(doby)
```

```
## Loading required package: survival
```

```
library(ggplot2)
if(nchar(system.file(package="ggplot2")) > 0) citation("ggplot2")
```

```
##
## To cite ggplot2 in publications, please use:
##
##   H. Wickham. ggplot2: elegant graphics for data analysis.
##   Springer New York, 2009.
##
## A BibTeX entry for LaTeX users is
##
##   @Book{,
##     author = {Hadley Wickham},
##     title = {ggplot2: elegant graphics for data analysis},
##     publisher = {Springer New York},
##     year = {2009},
##     isbn = {978-0-387-98140-6},
##     url = {http://had.co.nz/ggplot2/book},
##   }
```

```
library(reshape2)
library(gridExtra)
library(metafor)
```

```
## Loading required package: Matrix
## Loading 'metafor' package (version 1.9-7). For an overview
## and introduction to the package please type: help(metafor).
```

```
if(nchar(system.file(package="metafor")) > 0) citation("metafor")
```

```
##
## To cite the metafor package in publications, please use:
##
##   Wolfgang Viechtbauer (2010). Conducting meta-analyses in R with
##   the metafor package. Journal of Statistical Software, 36(3),
##   1-48. URL http://www.jstatsoft.org/v36/i03/.
##
## A BibTeX entry for LaTeX users is
##
##   @Article{,
##     title = {Conducting meta-analyses in {R} with the {metafor} package},
##     author = {Wolfgang Viechtbauer},
##     journal = {Journal of Statistical Software},
##     year = {2010},
##     volume = {36},
##     number = {3},
##     pages = {1--48},
##     url = {http://www.jstatsoft.org/v36/i03/},
##   }
```

```
source('CODE/mytheme.R')
```

```
## Loading required package: grid
```

```
figuresPath<-file.path(getwd()[1], "FIGURES_TABLES", "overview") #where to put the saved plots
fig.height<-2.5 #inches
fig.width<- 2.5 #inches
fig.res<-300

#from MIIN_3_calcEffectSizes.Rmd
papers<-read.table("DATA/DATA_SYNTHESIZED/calcES/papers.txt", sep="\t")
observations<-read.table("DATA/DATA_SYNTHESIZED/calcES/observations.txt", header=TRUE, sep="\t")
cover<-read.table("DATA/DATA_SYNTHESIZED/calcES/cover.txt", header=TRUE, sep="\t")
species<-read.table("DATA/DATA_SYNTHESIZED/calcES/species.txt", header=TRUE, sep="\t")
traits<-read.table("DATA/DATA_SYNTHESIZED/calcES/traits.txt", header=TRUE, sep="\t")
measures<-read.table("DATA/DATA_SYNTHESIZED/calcES/measures.txt", header=TRUE, sep="\t")
cwm<-read.table("DATA/DATA_SYNTHESIZED/calcES/cwm.txt", header=TRUE, sep="\t")
spIDcover<-read.table("DATA/DATA_SYNTHESIZED/calcES/spIDcover.txt", header=TRUE, sep="\t")
spIDtraits<-read.table("DATA/DATA_SYNTHESIZED/calcES/spIDtraits.txt", header=TRUE, sep="\t")
metaDataset<-read.table("DATA/DATA_SYNTHESIZED/calcES/metaDataset.txt", header=TRUE, sep="\t")
```

1. Article selection statistics

```
### Number of papers detected by source ###
summ.papers <- ddply(papers,~source,summarise,
                      numPapers=length(read),
                      numAcceptedPapers=sum(reject=='No'))
summ.papers<-orderBy(~numPapers, summ.papers)
summ.papers
```

```
##
##          source numPapers numAcceptedPapers
## 19      search2_111714      388             36
## 18      search1_111714      219             46
## 12          Liao2007        94             47
## 11 independent search for plant traits        3             0
## 3          cited by 249        2             2
## 8          cited by 368        2             2
## 10         cited by 626        2             1
## 1          cited by 155        1             1
## 2          cited by 229        1             0
## 4          cited by 25         1             1
## 5          cited by 256        1             0
## 6          cited by 29         1             1
## 7          cited by 317        1             1
## 9          cited by 455        1             1
## 13         ReferencedBy        1             1
## 14         related record 181    1             1
## 15         related record 188    1             1
## 16         related record 4      1             0
## 17         related record 570    1             1
```

```
### Number of unique number of papers detected ###
summ.papers2 <- ddply(papers,~source+rejectRationale,summarise,
                      numPapers=length(read),
                      numAcceptedPapers=sum(reject=='No'))
summ.papers2<-orderBy(~numPapers, summ.papers2)
totalNumReturned<-sum(summ.papers$numPapers) #total number of papers detected
numAlreadyFound<-sum(summ.papers2[summ.papers2$rejectRationale == 'alreadyFound' & !is.na(summ.papers2$
numUnique<-totalNumReturned - numAlreadyFound #total number of unique papers detected
paste(numUnique, 'unique papers identified by search criteria and their references')
```

```
## [1] "483 unique papers identified by search criteria and their references"
```

```
paste(sum(summ.papers$numAcceptedPapers), 'papers were accepted')
```

```
## [1] "143 papers were accepted"
```

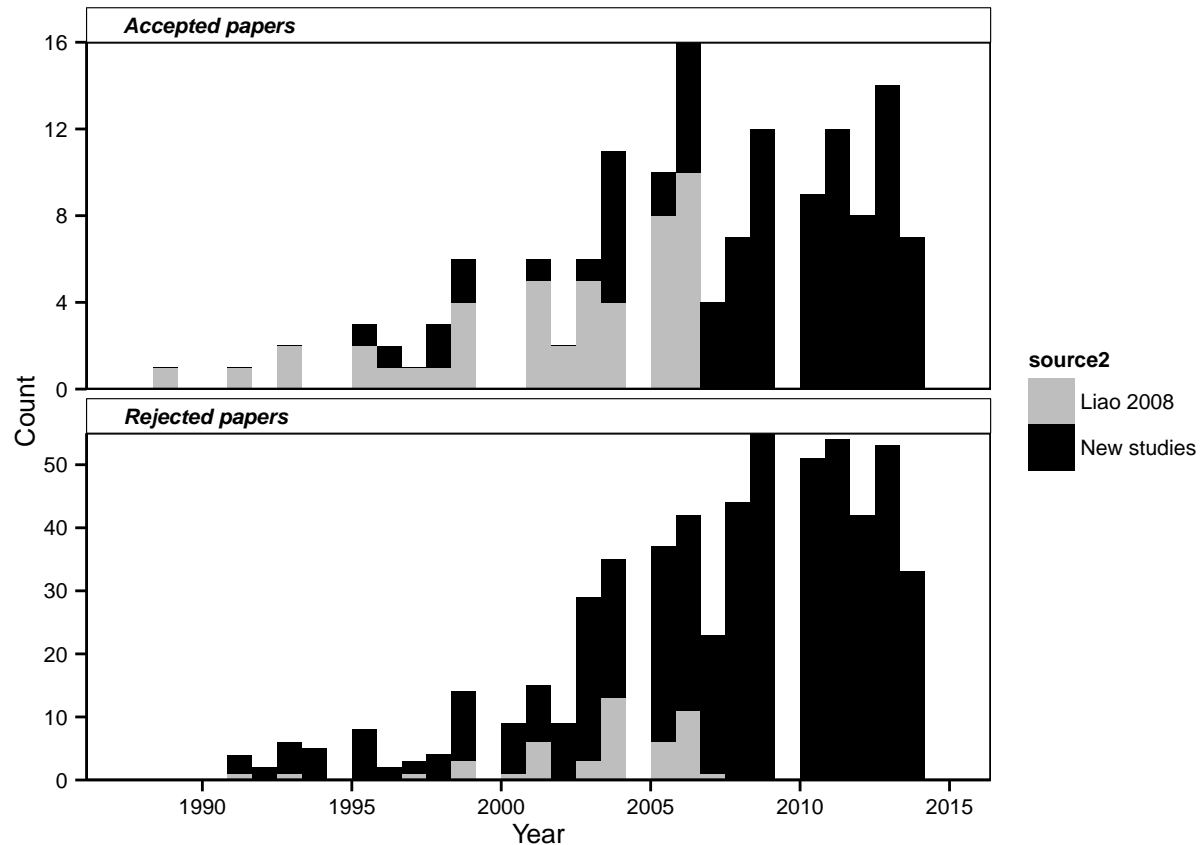
```
length(unique(metaDataset$paperID)) #this should be the same number
```

```
## [1] 143
```

```
### Subset papers detected by the previous meta-analysis, Liao2007 ###
papers$source2<-'New studies'
```

```
papers[papers$source=='Liao2007','source2']<-'Liao 2008'
papers$reject<-revalue(papers$reject, c("No"="Accepted papers", "Yes"="Rejected papers"))

#plot number of papers that were accepted/rejected from Liao 2008 and this search
pHist_papers<-ggplot(papers, aes(x=year, fill=source2)) + mytheme +
  facet_wrap(~reject, scales="free_y", ncol=1) +
  geom_histogram() + scale_y_continuous(expand = c(0,0)) +
  ylab('Count') + xlab('Year') +
  scale_fill_manual(values=c('gray','black'))
pHist_papers
```



```
newfilename<-"pHist_papers.png"
png(paste(figuresPath,newfilename, sep='/'),
  units='in', width = fig.width*1.5, height = fig.height*2, res=fig.res)
pHist_papers
dev.off()
```

```
## pdf
## 2
```

```
#what was the year of the most recent data included in Liao 2008?
maxLiaoyr<-max(papers[papers$source == 'Liao2007','year'])
paste(maxLiaoyr, 'was the most recent year that data was included in the Liao 2008 meta-analysis')
```

```
## [1] "2007 was the most recent year that data was included in the Liao 2008 meta-analysis"
```

```
#how many accepted papers were published after the most recent Liao 2008 reference?
accepted.after<-subset(papers, source != 'Liao2007' & reject == 'Accepted papers' & year > maxLiaoYr)
paste(dim(accepted.after)[1], 'papers were accepted after the most recent reference included in Liao 2008')
```

```
## [1] "69 papers were accepted after the most recent reference included in Liao 2008"
```

```
#how many papers were rejected that were referenced in Liao 2008? Remember that Liao 2008 also addressed rejected papers
rejected.Liao<-subset(papers, source == 'Liao2007' & reject == 'Rejected papers')
numLiaoRej<-dim(rejected.Liao)[1]
all.Liao<-subset(papers, source == 'Liao2007')
numLiaoAll<-dim(all.Liao)[1]
paste(dim(rejected.Liao)[1], 'papers that were used in Liao 2008 were rejected from this study, or', round(numLiaoRej/numLiaoAll, 2), '% of Liao 2008 references were rejected')
```

```
## [1] "47 papers that were used in Liao 2008 were rejected from this study, or 50 % of Liao references were rejected"
```

```
#How many papers were accepted that were published before the most recent Liao 2008 reference and were not rejected?
accepted.before<-subset(papers, source != 'Liao2007' & reject == 'Accepted papers' & year < maxLiaoYr)
numAccBef<-dim(accepted.before)[1]
paste(dim(accepted.before)[1], 'papers that were published before the most recent Liao 2008 reference were included in this study, or', round(numAccBef/numLiaoAll, 2), '% of Liao 2008 references were included in this study')
```

```
## [1] "23 papers that were published before the most recent Liao 2008 reference were included in this study, or 25 % of Liao 2008 references were included in this study"
```

```
#attached the 'source2' column to the metaDataset
temp_idx<-papers[,c('paperID', 'source2')]
metaDataset<-merge(metaDataset, temp_idx, by='paperID')

#re-write the metaDataset file so that it has the source2 column
newfilename<-'metaDataset.txt'
synthdataPath<-file.path(getwd()[1], "DATA", "DATA_SYNTHESIZED", "calcES")
write.table(metaDataset, file=paste(synthdataPath, newfilename, sep='/'), sep='\t')
```

2. Number of papers and observations

```
#how many observations?
paste(length(unique(observations$obsID)), 'observations in the full dataset')
```

```
## [1] "404 observations in the full dataset"
```

```
length(unique(metaDataset$obsID)) #these should be the same
```

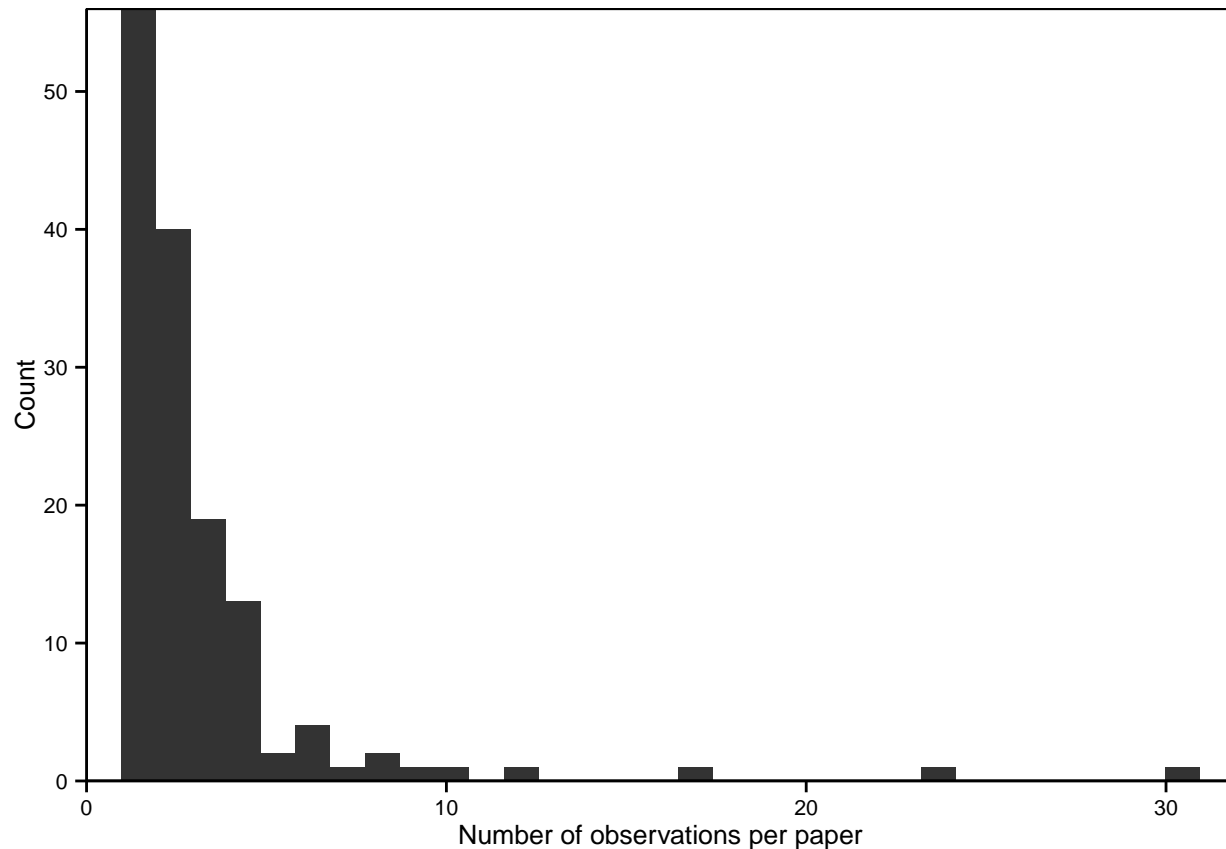
```
## [1] 404
```

```
#how many observations per paper?
summ.obs <- ddply(observations, ~paperID, summarise, numObs=length(paperID))
median(summ.obs$numObs); range(summ.obs$numObs)
```

```
## [1] 2
```

```
## [1] 1 30
```

```
pHist_obs<-ggplot(summ.obs, aes(x=numObs)) +  
  scale_y_continuous(expand=c(0,0)) + scale_x_continuous(expand=c(0,0)) +  
  geom_histogram() + mytheme +  
  ylab('Count') + xlab('Number of observations per paper')  
pHist_obs
```



3. Types of observations

```
summ.obs.eco <- ddply(observations,~ecosystCat,summarise, numObs=length(paperID))  
summ.obs.st <- ddply(observations,~studyType,summarise, numObs=length(paperID))  
summ.obs.nfix <- ddply(observations,~Nfix,summarise, numObs=length(paperID))  
factorlist<-list(summ.obs.eco, summ.obs.st, summ.obs.nfix)  
factortab<-ldply(factorlist)  
factortab$factor<-c(rep('ecosystem',5), rep('studyType', 4), rep('Nfix',4))  
factortab$level<-NA  
factortab[!is.na(factortab$ecosystCat),'level']<-as.character(factortab[!is.na(factortab$ecosystCat),'e
```

```
factortab[!is.na(factortab$studyType), 'level']<-as.character(factortab[!is.na(factortab$studyType), 'studyType'])
factortab[!is.na(factortab$Nfix), 'level']<-as.character(factortab[!is.na(factortab$Nfix), 'Nfix'])
factortab1<-factortab[,c('factor', 'level', 'numObs')]
factortab1
```

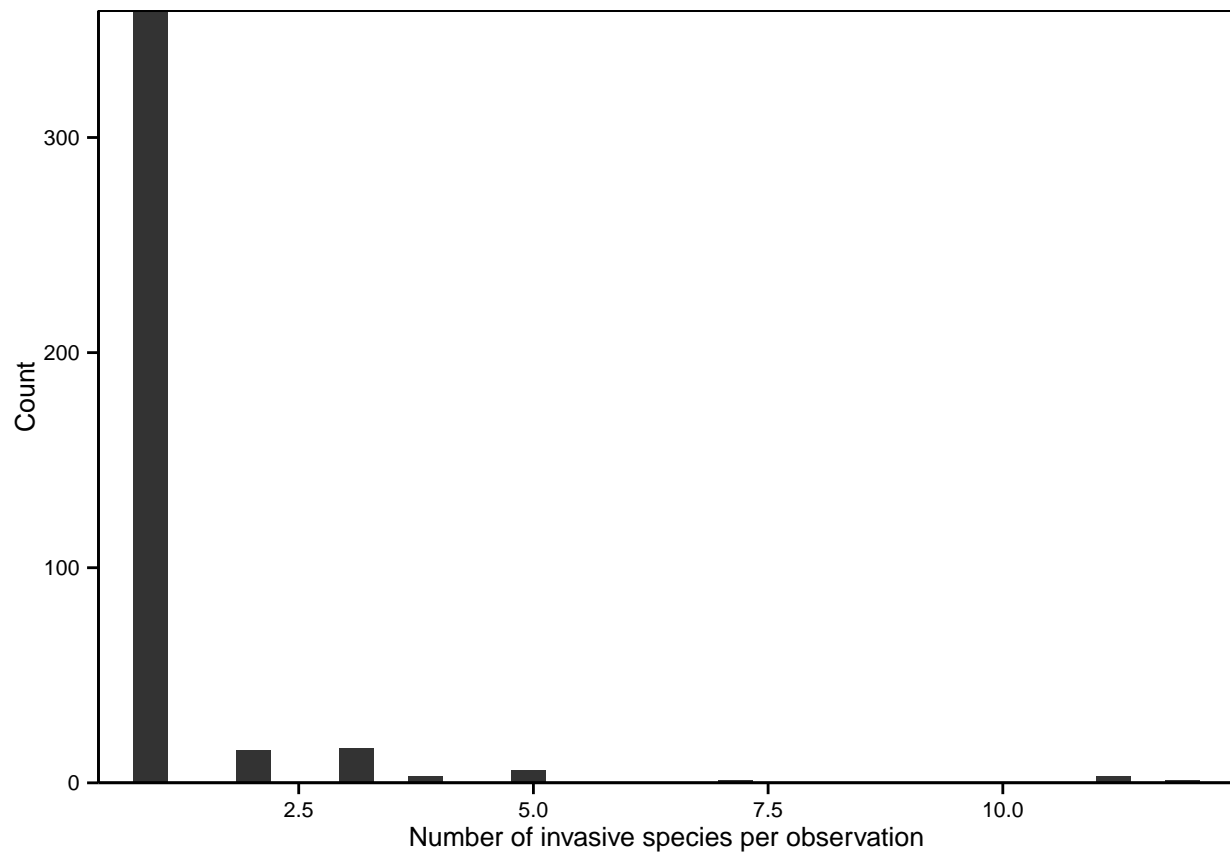
```
##      factor                level numObs
## 1 ecosystem                forest   123
## 2 ecosystem                grassland 176
## 3 ecosystem                other     4
## 4 ecosystem                shrubland  73
## 5 ecosystem                wetland   28
## 6 studyType                field expt addition 45
## 7 studyType                field expt removal 25
## 8 studyType                field study    273
## 9 studyType                greenhouse expt  61
## 10      Nfix Invasive and resident N-fixers 20
## 11      Nfix      Invasive N-fixers only  51
## 12      Nfix      No N-fixers          295
## 13      Nfix      Resident N-fixers only  38
```

```
newfilename<-'numObsTable.txt'
write.table(factortab1, file=paste(figuresPath,newfilename, sep='/'), sep='\t')
```

4. Plant species statistics

What is the distribution of invasive species per observation? Native species? Are certain invasive species over-represented?

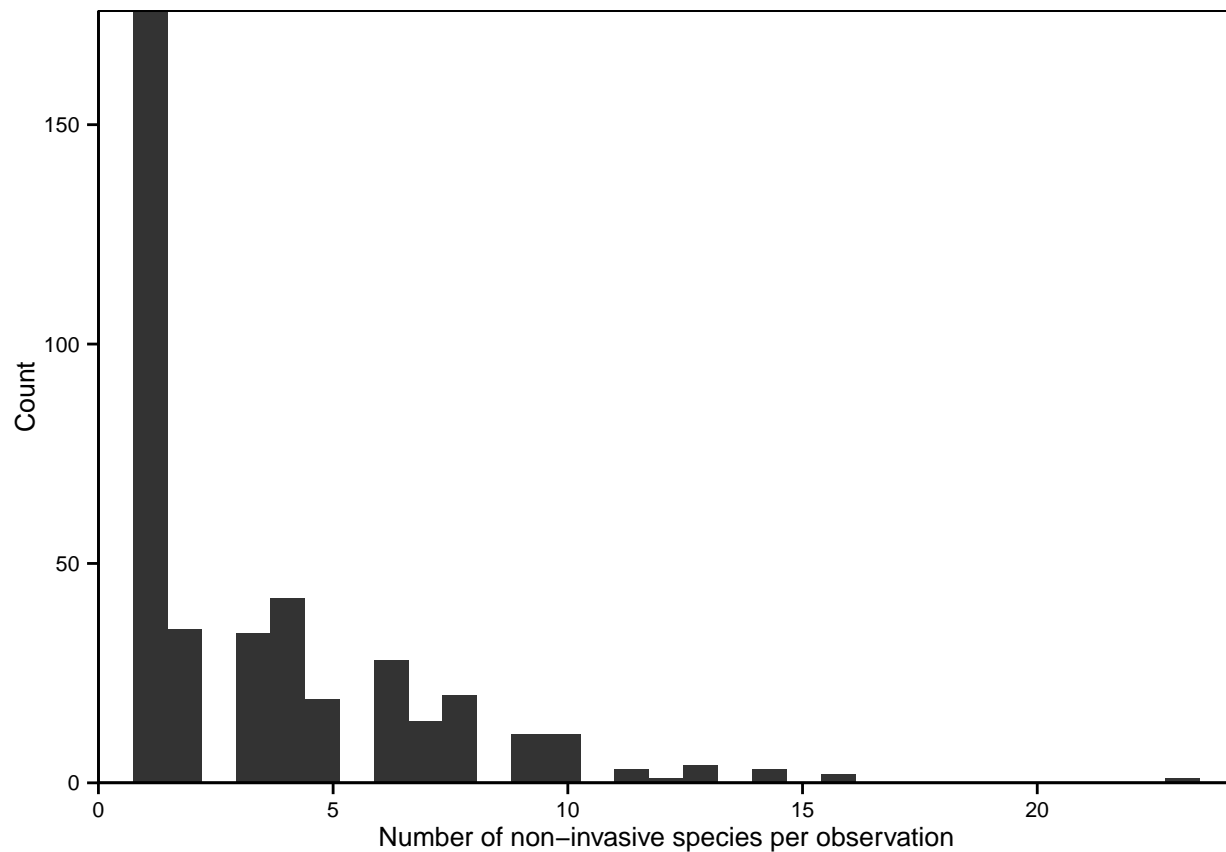
```
#what is the distribution of invasive species per observation?
summ.spp <- ddply(species,~obsID,summarise,
                  numTotalspp=length(obsID),
                  numInv spp=sum(spInvasive=='invasive' & spExotic=='exotic' & spFocal=='focal'),
                  numNonInv spp=sum(spInvasive=='not invasive'),
                  numOthers=numTotalspp-(numInv spp + numNonInv spp))
hist_Inv<-ggplot(summ.spp, aes(x=numInv spp)) + geom_histogram() +
  scale_y_continuous(expand=c(0,0)) + scale_x_continuous(expand=c(0,0)) +
  mytheme +
  ylab('Count') + xlab('Number of invasive species per observation')
hist_Inv; median(summ.spp$numInv spp); range(summ.spp$numInv spp)
```

```
## [1] 1
```

```
## [1] 1 12
```

```
hist_Nat<-ggplot(summ.spp, aes(x=numNonInv spp)) + geom_histogram() +
  scale_y_continuous(expand=c(0,0)) + scale_x_continuous(expand=c(0,0)) +
  mytheme +
  ylab('Count') + xlab('Number of non-invasive species per observation')
hist_Nat; median(summ.spp$numNonInv spp); range(summ.spp$numNonInv spp)
```



```
## [1] 2
```

```
## [1] 1 23
```

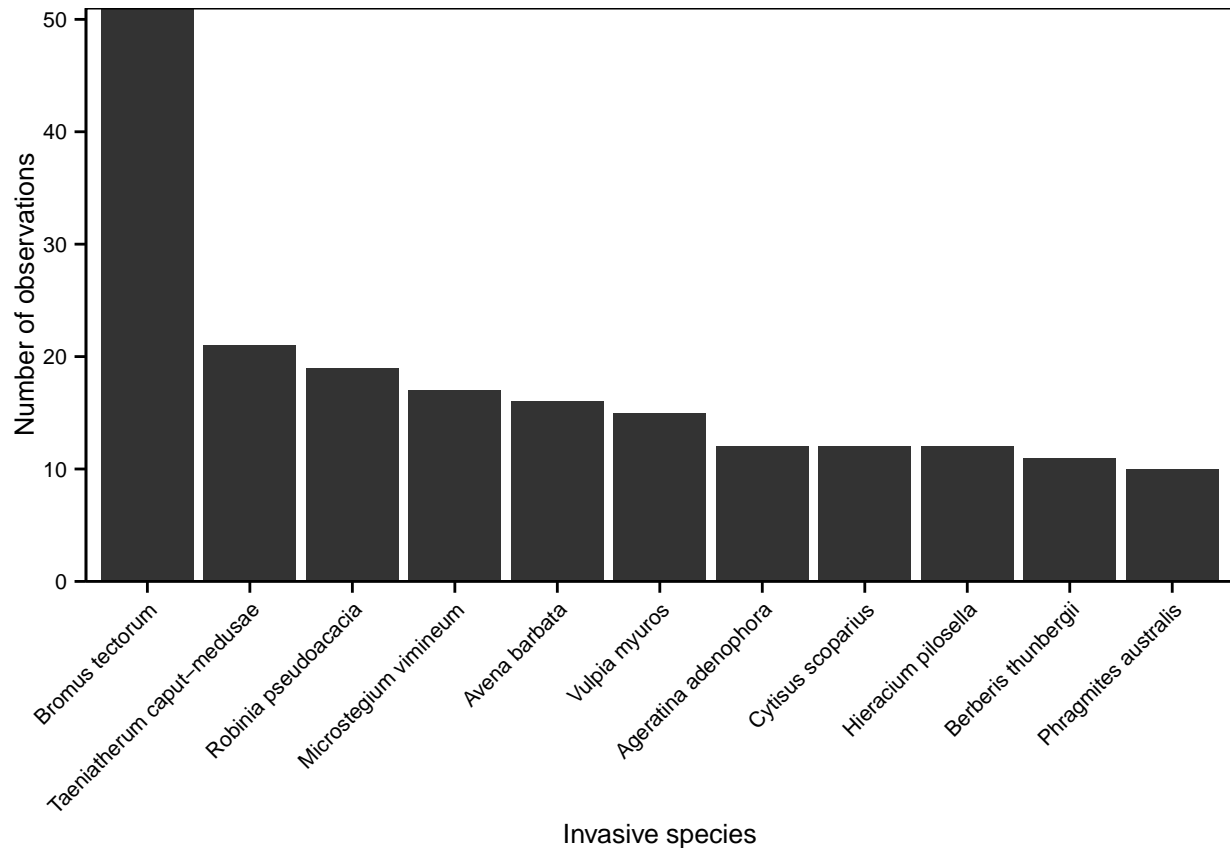
```
#number of observations per species
summ.spp.nam <- ddpby(species,~spName+spFocal+spExotic,summarise,
  numObs=length(obsID),
  numPapers=length(unique(paperID)))
spp.many<-summ.spp.nam[which(summ.spp.nam$numObs > 9 & summ.spp.nam$spFocal == 'focal'),] #more than 9
spp.many.o<-orderBy(~numObs, spp.many)
spp.many.o
```

```
##           spName spFocal spExotic numObs numPapers
## 130      Bromus tectorum focal  exotic    51        19
## 616 Taeniatherum caput-medusae focal  exotic    21         3
## 551      Robinia pseudoacacia focal  exotic    19         7
## 405  Microstegium vimineum focal  exotic    17         7
##  92       Avena barbata focal  exotic    16         5
## 660       Vulpia myuros focal  exotic    15         4
##  29    Ageratina adenophora focal  exotic    12         1
## 216      Cytisus scoparius focal  exotic    12         6
## 314    Hieracium pilosella focal  exotic    12         5
## 103    Berberis thunbergii focal  exotic    11         4
## 463    Phragmites australis focal  exotic    10         5
```

```

positions<-spp.many.o$spName
pHist_spp<-ggplot(spp.many.o, aes(x=spName, y=numObs)) + geom_bar(stat='identity') +
  scale_y_continuous(expand=c(0,0)) + scale_x_discrete(limits = positions) +
  mytheme + theme(axis.text.x=element_text(angle=45, hjust=1)) +
  ylab('Number of observations') + xlab('Invasive species')
pHist_spp

```



```

newfilename<-'pHist_spp.png'
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*2, height = fig.height*2, res=fig.res)
pHist_spp
dev.off()

```

```

## pdf
## 2

```

```

#which species appear both as exotic and native species in the dataset?
summ.spp <- ddply(species,~spName+spExotic, summarise,
  numObs=length(obsID),
  numPapers=length(unique(paperID)))
summ.spp.nam2 <- ddply(summ.spp,~spName,summarise,
  numInvNat=length(spExotic))
summ.spp.nam2[summ.spp.nam2$numInvNat==2,] # if the length of spInvasive col==2, then there is native a

```

```

##          spName numInvNat

```

```
## 5      Acacia longifolia      2
## 10      Acer negundo      2
## 61 Anthoxanthum odoratum      2
## 115     Briza maxima      2
## 118     Bromus hordeaceus      2
## 208     Cytisus scoparius      2
## 209     Dactylis glomerata      2
## 269     Festuca arundinacea      2
## 305     Holcus lanatus      2
## 451 Phragmites australis      2
## 477     Poa pratensis      2
## 498     Prunus serotina      2
## 553 Schedonorus phoenix      2
## 615     Trifolium sp      2
```

5. Cover data statistics

What percent of observations have measured cover data?

```
summ.cov.obs <- ddply(cover,~obsID,summarise,
                      numMeasured= sum(covQuality=='measured'))
numMeasured.obs<-sum(summ.cov.obs$numMeasured > 0) #number of observations with cover measurement value
numtotal.obs<-length(summ.cov.obs$numMeasured > 0) #total number of observations
cov.obs.perc<-round((numMeasured.obs / (numtotal.obs) ) *100, digits=2)
paste(cov.obs.perc, '% of observations have any cover data at all that was measured in the original paper')
```

```
## [1] "27.97 % of observations have any cover data at all that was measured in the original paper"
```

```
#What is the frequency of cover observations for each cover measure type?
summ.cov <- ddply(cover,~covCat,summarise,
                  numMeas = length(obsID),
                  numObs=length(unique(obsID)),
                  numPapers=length(unique(paperID)))
orderBy(~numMeas, summ.cov)
```

```
##      covCat numMeas numObs numPapers
## 3 sp_plantcov   1141    404      143
## 1 sp_biomass    81     32       13
## 2 sp_ind       80     22        6
```

```
#What units are commonly reported for each cover measure type?
summ.covUnit <- ddply(cover,~covCat+covUnit,summarise,
                      numMeas = length(obsID),
                      numObs=length(unique(obsID)),
                      numPapers=length(unique(paperID)))
COVCAT<-unique(summ.covUnit$covCat)
covUnitList<-list()
i<-0
for(i in 1:length(COVCAT)){
```

```

subdf<-summ.covUnit[summ.covUnit$covCat==COVCAT[i],]
covUnitList[[as.character(COVCAT[i])]]<-orderBy(~numMeas, subdf)
}
covUnitList

```

```

## $sp_biomass
##      covCat covUnit numMeas numObs numPapers
## 2 sp_biomass g/m2      38      13         6
## 5 sp_biomass ind/ha     12       6         1
## 7 sp_biomass m2/ha     11       3         1
## 4 sp_biomass g/pot      6       3         1
## 6 sp_biomass kg/m2      6       3         2
## 1 sp_biomass %         4       2         1
## 3 sp_biomass g/m2*y     4       2         1
##
## $sp_ind
##      covCat covUnit numMeas numObs numPapers
## 11 sp_ind notReported    34       8         1
## 9  sp_ind ind/30m2      19       1         1
## 13 sp_ind stems/m2      11       5         1
## 12 sp_ind plants/m2     10       5         1
## 8  sp_ind ind/10m2      4       1         1
## 10 sp_ind ind/ha        2       2         1
##
## $sp_plantcov
##      covCat covUnit numMeas numObs numPapers
## 14 sp_plantcov %      1141     404        143

```

```

#A more detailed look at cover data quality as it contributes to CWM values...
cwm.calc<-subset(cwm, qualityCWMcalc == 'calculated')
summ.cwm <- ddply(cwm.calc,~traitCat+invType,summarise,
  numObs=length(unique(obsID)),
  num1spAll_1=sum(qualityCover=='Measured=All, 1sp=All'),
  num1spAll_2=sum(qualityCover=='Measured=None, 1sp=All'),
  num1spAll_3=sum(qualityCover=='Measured=NA, 1sp=NA'),
  num1spAll_4=sum(qualityCover=='Measured=Mid, 1sp=All'),
  totalspAll=sum(num1spAll_1, num1spAll_2, num1spAll_3, num1spAll_4),
  perc1spAll=(totalspAll/numObs) *100,
  percEqual=100-perc1spAll)
summ.cwm #cover data quality by traitCat and invType

```

```

##      traitCat      invType numObs num1spAll_1 num1spAll_2 num1spAll_3
## 1          cn      InvArea   198          15         138          0
## 2          cn InvSpInvArea   198          18         154          0
## 3          cn      NatArea   198          15          78         57
## 4 littercn      InvArea    40           5          27          0
## 5 littercn InvSpInvArea    40           8          31          0
## 6 littercn      NatArea    40           5          14          6
## 7 litterpercN      InvArea    42           5          24          0
## 8 litterpercN InvSpInvArea    42           6          29          0
## 9 litterpercN      NatArea    42           5          17          7
## 10         percN      InvArea   318          32         189          2
## 11         percN InvSpInvArea   318          52         232          2

```

	percN	NatArea	318	36	144	23
## 12	num1spAll_4	total1spAll	perc1spAll	percEqual		
## 1	2	155	78.28283	21.71717		
## 2	0	172	86.86869	13.13131		
## 3	0	150	75.75758	24.24242		
## 4	1	33	82.50000	17.50000		
## 5	0	39	97.50000	2.50000		
## 6	1	26	65.00000	35.00000		
## 7	0	29	69.04762	30.95238		
## 8	0	35	83.33333	16.66667		
## 9	0	29	69.04762	30.95238		
## 10	6	229	72.01258	27.98742		
## 11	0	286	89.93711	10.06289		
## 12	1	204	64.15094	35.84906		

```
summ.cwm2 <- ddply(summ.cwm, ~invType, summarise,
  mean1sp=mean(perc1spAll),
  meanEqual=mean(percEqual),
  seEqual=sd(percEqual)/sqrt(length(percEqual)))
summ.cwm2 #aggregated across traitCat
```

	invType	mean1sp	meanEqual	seEqual
## 1	InvArea	75.46076	24.53924	3.034981
## 2	InvSpInvArea	89.40978	10.59022	3.015379
## 3	NatArea	68.48903	31.51097	2.647894

```
summ.cwm3 <- ddply(cwm, ~invType+traitCat+obsID, summarise,
  numReported=sum(qualityCWMcalc=='reported'))
summ.cwm4 <- ddply(summ.cwm3, ~traitCat+invType, summarise,
  count=sum(numReported != 0),
  total=length(obsID),
  percCWMReported=(count/total)*100)
summ.cwm4 #percent of CWM data that was reported in the original paper (rather than calculated based on
```

	traitCat	invType	count	total	percCWMReported
## 1	cn	InvArea	14	212	6.603774
## 2	cn	InvSpInvArea	14	212	6.603774
## 3	cn	NatArea	14	212	6.603774
## 4	littercn	InvArea	21	61	34.426230
## 5	littercn	InvSpInvArea	21	61	34.426230
## 6	littercn	NatArea	21	61	34.426230
## 7	litterpercN	InvArea	26	68	38.235294
## 8	litterpercN	InvSpInvArea	26	68	38.235294
## 9	litterpercN	NatArea	26	68	38.235294
## 10	percN	InvArea	53	371	14.285714
## 11	percN	InvSpInvArea	53	371	14.285714
## 12	percN	NatArea	53	371	14.285714

6. Trait data statistics

```
# what percent of observations had trait data reported within the original article?
n.ot<-length(unique(traits$obsID)) # number of observations with trait data
n.o<-length(unique(observations$obsID)) # total number of observations
tr.obs.perc<-round((n.ot/n.o) *100, digits=2) # percent of observations with trait data
paste(tr.obs.perc, '% of observations with species-level trait data from the original paper',collapse=''
```

```
## [1] "34.41 % of observations with species-level trait data from the original paper"
```

```
summ.tr <- ddply(traits,~traitCat,summarise,
                numObs = length(unique(obsID)),
                numPapers = length(unique(paperID)))
summ.tr.o<-orderBy(~numObs, summ.tr)
summ.tr.o
```

```
##      traitCat numObs numPapers
## 4      sp_percN    106         40
## 1          sp_cn     54         21
## 3 sp_litterpercN     48         22
## 2      sp_littercn    32         18
```

```
positions<-summ.tr.o$traitCat
pBar.tr<-ggplot(summ.tr.o, aes(x=traitCat, y=numObs)) + geom_bar(stat='identity') +
  scale_y_continuous(expand=c(0,0)) +
  scale_x_discrete(limits = positions,
                  labels = c("sp_percN" = "Leaf %N",
                             "sp_cn" = "Leaf C:N",
                             "sp_litterpercN" = "Litter %N",
                             "sp_littercn" = "Litter C:N")) +
  mytheme + theme(axis.text.x=element_text(angle=45, hjust=1)) +
  ylab('Number of observations') + xlab('Trait type (species-level)')
```

```
#What units and methods are commonly reported for each measurement?
```

```
summ.traitUnit <- ddply(traits,~traitCat+traitUnit,summarise,
                        numMeas = length(obsID),
                        numObs=length(unique(obsID)),
                        numPapers=length(unique(paperID)))
summ.traitUnit
```

```
##      traitCat traitUnit numMeas numObs numPapers
## 1      sp_cn          %      16      4         2
## 2      sp_cn      %C/%N      11      5         5
## 3      sp_cn molC/molN     128     45        14
## 4      sp_littercn %C/%N       6      2         2
## 5      sp_littercn molC/molN     81     30        16
## 6      sp_litterpercN %      85     29        14
## 7      sp_litterpercN g/kg       6      4         2
## 8      sp_litterpercN mg/g      38     15         6
## 9          sp_percN %     160     58        23
## 10         sp_percN g/kg      50     21         3
```

```
## 11      sp_percN      mg/g      66      23      11
## 12      sp_percN    mmol/kg       4       1       1
## 13      sp_percN      ug/g      14       2       1
## 14      sp_percN      ug/mg       2       1       1
```

```
TRAITCAT<-unique(summ.traitUnit$traitCat)
traitUnitList<-list()
i<-0
for(i in 1:length(TRAITCAT)){
  subdf<-summ.traitUnit[summ.traitUnit$traitCat==TRAITCAT[i],]
  traitUnitList[[as.character(TRAITCAT[i])]]<-orderBy(~numMeas, subdf)
}
traitUnitList
```

```
## $sp_cn
##   traitCat traitUnit numMeas numObs numPapers
## 3   sp_cn molC/molN    128    45      14
## 1   sp_cn      %      16     4       2
## 2   sp_cn   %C/%N     11     5       5
##
## $sp_littercn
##   traitCat traitUnit numMeas numObs numPapers
## 5 sp_littercn molC/molN     81    30      16
## 4 sp_littercn   %C/%N      6     2       2
##
## $sp_litterpercN
##   traitCat traitUnit numMeas numObs numPapers
## 6 sp_litterpercN      %      85    29      14
## 8 sp_litterpercN    mg/g      38    15       6
## 7 sp_litterpercN    g/kg       6     4       2
##
## $sp_percN
##   traitCat traitUnit numMeas numObs numPapers
## 9   sp_percN      %      160    58      23
## 11  sp_percN    mg/g      66    23      11
## 10  sp_percN    g/kg      50    21       3
## 13  sp_percN    ug/g      14     2       1
## 12  sp_percN  mmol/kg       4     1       1
## 14  sp_percN    ug/mg       2     1       1
```

7. Soil measurement statistics

```
summ.meas <- ddply(measures,~measCat,summarise, numObs=length(unique(obsID)))
summ.meas.o<-orderBy(~numObs, summ.meas)
summ.meas.o
```

```
##   measCat numObs
## 9   toti    225
```



```
## 7    soiln    212
## 5      no    177
## 2      nh    162
## 4    nminz    128
## 6   soilcn    126
## 8     som     97
## 3   nitrif     85
## 1 ammonif     54
```

#What units and methods are commonly reported for each measurement?

```
summ.measUnit <- ddply(measures,~measCat+unit,summarise,
                        numMeas = length(obsID),
                        numObs=length(unique(obsID)))
MEASCAT<-unique(summ.measUnit$measCat)
measUnitList<-list()
i<-0
for(i in 1:length(MEASCAT)){
  subdf<-summ.measUnit[summ.measUnit$measCat==MEASCAT[i],]
  measUnitList[[as.character(MEASCAT[i])]]<-orderBy(~numMeas, subdf)
}
measUnitList
```

```
## $ammonif
##   measCat      unit numMeas numObs
## 4  ammonif    mg/kg*d     14     14
## 14 ammonif    ug/g*d      7      7
## 12 ammonif   ug/g*2wks     5      5
## 17 ammonif    ug/g*mo     5      5
## 15 ammonif    ug/g*hr     4      4
## 2  ammonif    mg/kg*10d     2      2
## 8  ammonif   notReported     2      2
## 9  ammonif      ppm      2      2
## 10 ammonif     ug/g      2      2
## 11 ammonif    ug/g*2wk     2      2
## 18 ammonif    umol/g*d     2      2
## 1  ammonif meq per 100g soil     1      1
## 3  ammonif    mg/kg*28d     1      1
## 5  ammonif    mg/kg*mo     1      1
## 6  ammonif    mg/m2*d     1      1
## 7  ammonif    mg/m2*y     1      1
## 13 ammonif    ug/g*30d     1      1
## 16 ammonif    ug/g*IER     1      1
##
## $nh
##   measCat      unit numMeas numObs
## 38      nh      ug/g     59     59
## 28      nh     mg/kg     44     44
## 32      nh    mmol/kg     17     17
## 29      nh      mg/L      7      7
## 35      nh      ppm      5      5
## 26      nh     mg/bag      4      4
## 27      nh      mg/g      4      4
## 21      nh      g/m2      3      3
## 34      nh   notReported     3      3
```

## 37	nh	ug/capsule	3	3
## 22	nh	g/m3	2	2
## 31	nh	mg/m2*y	2	2
## 19	nh	%	1	1
## 20	nh	cmol/kg resin	1	1
## 23	nh	g/mg	1	1
## 24	nh	kg/ha	1	1
## 25	nh	meq per 100g soil	1	1
## 30	nh	mg/m2	1	1
## 33	nh	ng/g*d	1	1
## 36	nh	ug/10cm2*35d	1	1
## 39	nh	ug/kg	1	1

##

\$nitrif

##	measCat	unit	numMeas	numObs
## 56	nitrif	ug/g*d	23	23
## 44	nitrif	mg/kg*d	19	19
## 54	nitrif	ug/g*2wks	5	5
## 57	nitrif	ug/g*hr	5	5
## 59	nitrif	ug/g*mo	5	5
## 40	nitrif	g/m2*6mo	4	4
## 52	nitrif	ug/g*14d	3	3
## 42	nitrif	mg/kg*10d	2	2
## 47	nitrif	mg/m2*mo	2	2
## 49	nitrif	notReported	2	2
## 50	nitrif	ppm/30d	2	2
## 51	nitrif	ug/g	2	2
## 53	nitrif	ug/g*2wk	2	2
## 60	nitrif	umol/g*d	2	2
## 41	nitrif	meq per 100g soil	1	1
## 43	nitrif	mg/kg*28d	1	1
## 45	nitrif	mg/kg*mo	1	1
## 46	nitrif	mg/m2*d	1	1
## 48	nitrif	mg/m2*y	1	1
## 55	nitrif	ug/g*30d	1	1
## 58	nitrif	ug/g*IER	1	1

##

\$nminz

##	measCat	unit	numMeas	numObs
## 80	nminz	ug/g*d	24	24
## 74	nminz	mmol/kg*30d	17	17
## 70	nminz	mg/kg*d	16	16
## 68	nminz	mg/kg	11	11
## 64	nminz	g/m2*s	6	6
## 66	nminz	mg/g*382d	6	6
## 67	nminz	mg/g*wk	6	6
## 78	nminz	ug/g*2wks	5	5
## 62	nminz	g/m2*6mo	4	4
## 81	nminz	ug/g*hr	4	4
## 65	nminz	g/m2*y	3	3
## 75	nminz	ug/g	3	3
## 76	nminz	ug/g*14d	3	3
## 83	nminz	ug/g*mo	3	3
## 61	nminz	g/ha*d	2	2

```

## 69  nminz  mg/kg*60d      2      2
## 72  nminz  mg/m2*mo      2      2
## 77  nminz  ug/g*2wk      2      2
## 84  nminz  ug/g*y        2      2
## 85  nminz  umol/g*d      2      2
## 63  nminz  g/m2*d        1      1
## 71  nminz  mg/m2*d        1      1
## 73  nminz  mg/m2*y        1      1
## 79  nminz  ug/g*30d      1      1
## 82  nminz  ug/g*IER      1      1
##
## $no
##      measCat      unit numMeas numObs
## 104      no      ug/g      59      59
## 95      no      mg/kg      57      57
## 98      no      mmol/kg     17      17
## 100     no      notReported    7      7
## 101     no      ppm          5      5
## 93      no      mg/bag        4      4
## 94      no      mg/g          4      4
## 96      no      mg/L          4      4
## 88      no      g/m2          3      3
## 103     no      ug/capsule     3      3
## 89      no      g/m3          2      2
## 97      no      mg/m2*y        2      2
## 106     no      umol/capsule*d  2      2
## 86      no      %            1      1
## 87      no      cmol/kg resin  1      1
## 90      no      g/mg          1      1
## 91      no      kg/ha          1      1
## 92      no      meq per 100g soil 1      1
## 99      no      ng/g*d        1      1
## 102     no      ug/10cm2*35d    1      1
## 105     no      ug/kg          1      1
##
## $soilcn
##      measCat      unit numMeas numObs
## 109  soilcn  molC/molN      79      79
## 107  soilcn  %C/%N        39      39
## 108  soilcn  gC/gN         8       8
##
## $soiln
##      measCat      unit numMeas numObs
## 110  soiln   %          106     106
## 111  soiln   g/kg        33      33
## 115  soiln   mg/g        29      29
## 112  soiln   g/m2        18      18
## 116  soiln   mg/kg        12      12
## 119  soiln   ug/g         6       6
## 114  soiln   kg/m3         2       2
## 118  soiln   ppm          2       2
## 120  soiln   ug/kg         2       2
## 113  soiln   kg/ha         1       1
## 117  soiln   mmol/kg        1       1

```

```
##
## $som
##      measCat unit numMeas numObs
## 121      som   %       91      91
## 123      som g/kg       4       4
## 122      som   cm       1       1
## 124      som mg/g       1       1
##
## $toti
##      measCat      unit numMeas numObs
## 140      toti      ug/g      79      79
## 130      toti      mg/kg      52      52
## 133      toti      mg/pot      24      24
## 134      toti      mmol/kg      17      17
## 128      toti      mg/bag      10      10
## 141      toti      ug/gIER*d      8       8
## 129      toti      mg/g       7       7
## 131      toti      mg/L       6       6
## 136      toti      ppm       5       5
## 139      toti      ug/bag*d      4       4
## 126      toti      g/m2       3       3
## 135      toti      notReported      3       3
## 142      toti umol/capsule*d      2       2
## 125      toti      %       1       1
## 127      toti      index      1       1
## 132      toti      mg/m2       1       1
## 137      toti      ug/10cm2*35d      1       1
## 138      toti      ug/bag      1       1
```

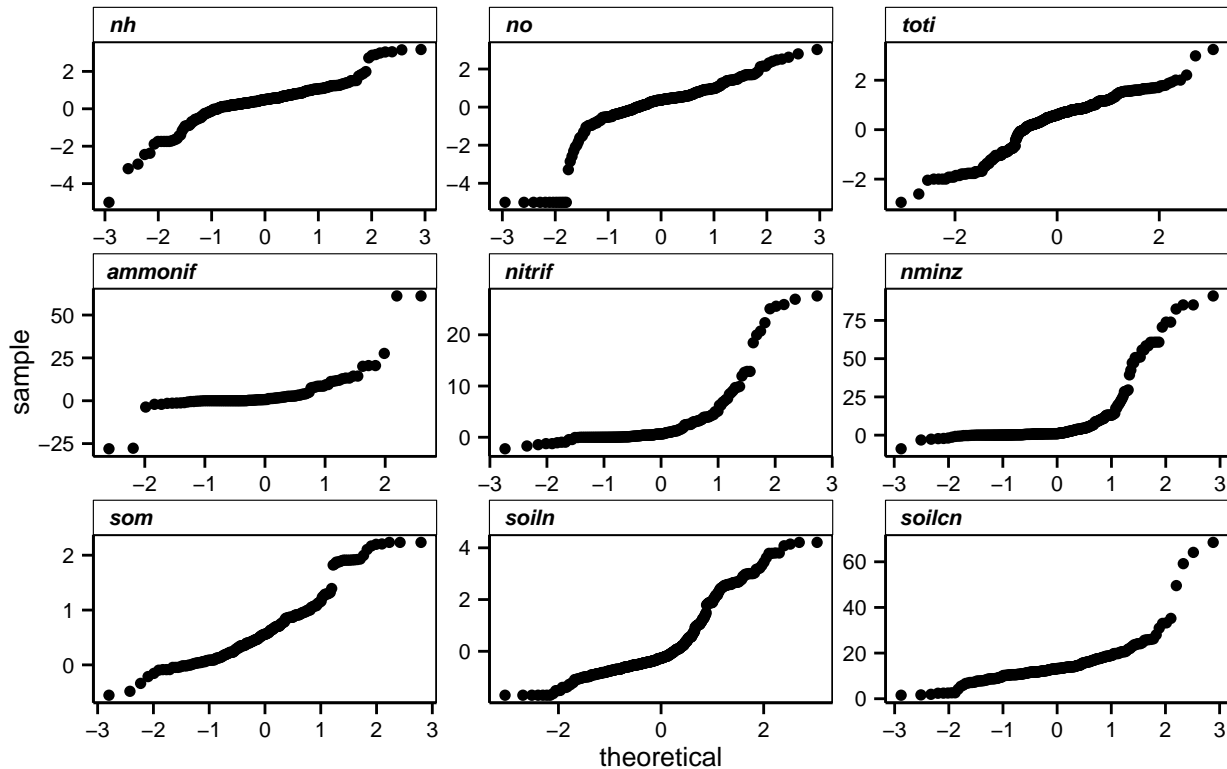
```
#re-order measCat levels
metaDataset$measCat <- factor(metaDataset$measCat, levels = measCat_order)

#re-shape measures so that inv and nat are in the same column temporarily
tmp<-ddply(metaDataset, ~obsID+measCat, summarize,
  m1i_logt = unique(m1i_logt),
  m2i_logt = unique(m2i_logt),
  measQuality = unique(measQuality))
tmp$obsID<-as.factor(tmp$obsID)
m.tmp<-melt(tmp, idcols=c('obsID','measCat','measQuality'))
m.tmp$invType<-rep(NA,length(dim(m.tmp)[1]))
m.tmp[m.tmp$variable == 'm1i_logt','invType']<-'inv'
m.tmp[m.tmp$variable == 'm2i_logt','invType']<-'ref'

#Shapiro Test
# ddply(measures, ~measCat, summarise,
#       shapTest=shapiro.test(value)$p.value,
#       shapTest.Ln=shapiro.test(log(value+1))$p.value)
#none are normal according to Shapiro test

# Q-Q plots
qq<-ggplot(m.tmp, aes(sample=value)) +
  facet_wrap(~measCat, scales='free', ncol=3) +
  stat_qq() + mytheme + ggtitle('QQ Plots of \nstd. measurement values')
qq
```

QQ Plots of std. measurement values



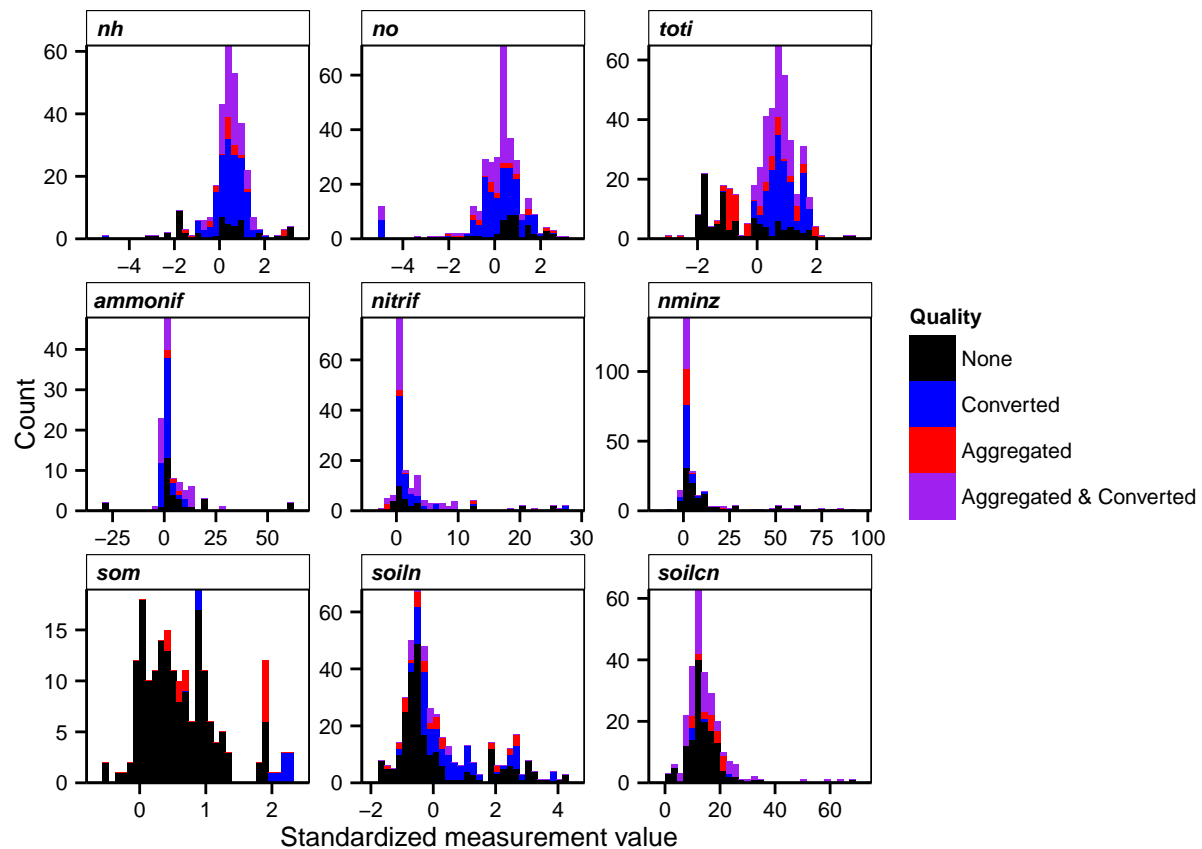
```
newfilename<-"qq_meas.png"
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*3, height = fig.height*6, res=fig.res)
qq
dev.off()
```

```
## pdf
## 2
```

```
# Plot Quality Histograms
#re-order measQuality levels
```

```
m.tmp$measQuality <- factor(m.tmp$measQuality, levels = c('NoAgg.NoConv', 'NoAgg.Conv', 'Agg.NoConv', 'Agg.NoConv'))
```

```
pHist_measQual<-ggplot(data=m.tmp, aes(x=value,fill=measQuality)) + mytheme +
  facet_wrap(~measCat, scales='free', ncol=3) + geom_histogram() +
  scale_y_continuous(expand = c(0,0)) +
  scale_fill_manual(name = "Quality",
                    labels = c("Agg.Conv"="Aggregated & Converted",
                              "Agg.NoConv"="Aggregated",
                              "NoAgg.Conv"="Converted",
                              "NoAgg.NoConv"="None"),
                    values=c("Agg.Conv" = "purple",
                              "Agg.NoConv" = "red",
                              "NoAgg.Conv" = "blue",
                              "NoAgg.NoConv" = "black")) +
  ylab('Count') + xlab('Standardized measurement value')
pHist_measQual
```



```
newfilename<-"pHist_measQual.png"
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*3.5, height = fig.height*6, res=fig.res)
pHist_measQual
dev.off()
```

```
## pdf
## 2
```

8. Effect size statistics

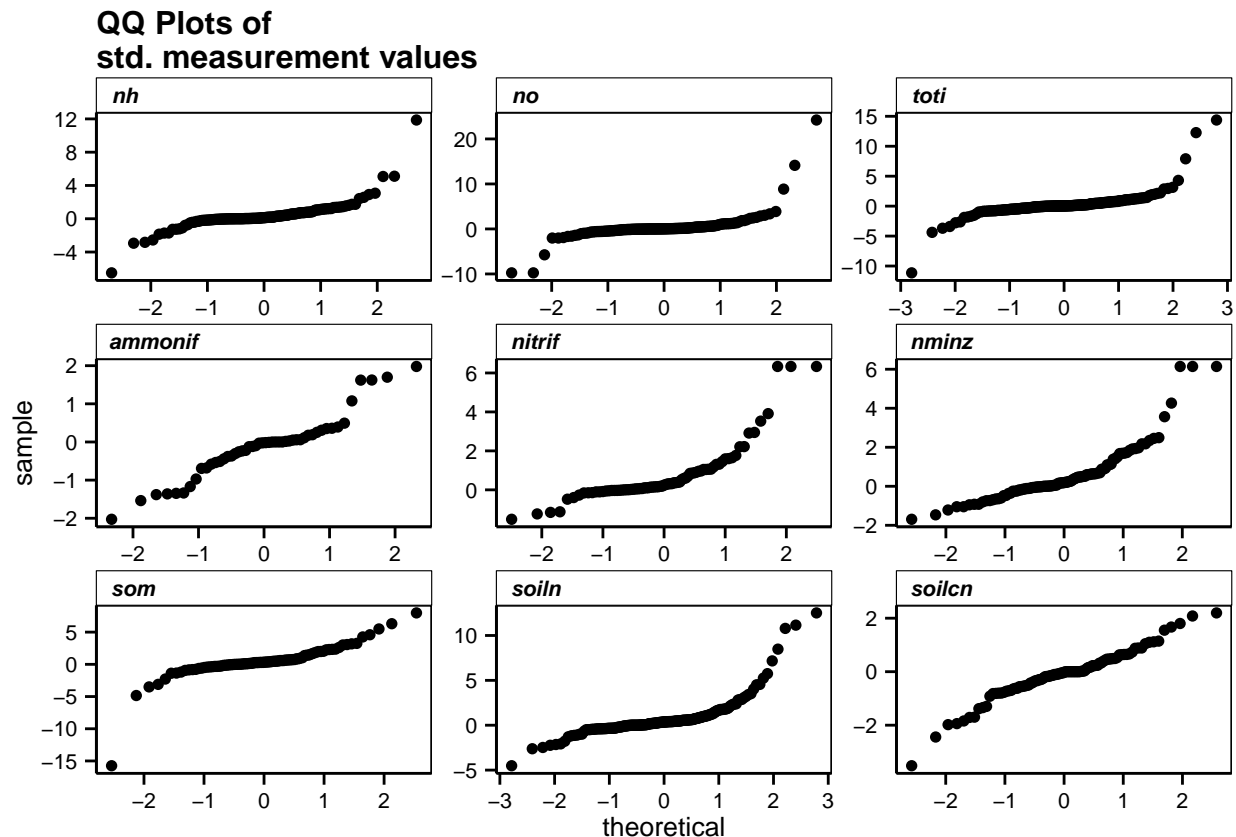
```
#re-shape measures so that inv and nat are in the same column temporarily
tmp<-ddply(metaDataset, ~obsID+measCat, summarize,
    yi = unique(yi),
    measQuality = unique(measQuality))
tmp$obsID<-as.factor(tmp$obsID)

#Shapiro Test
# ddply(measures, ~measCat, summarise,
#     shapTest=shapiro.test(value)$p.value,
#     shapTest.Ln=shapiro.test(log(value+1))$p.value)
```

```
#none are normal according to Shapiro test
```

```
# Q-Q plots
```

```
qq<-ggplot(tmp, aes(sample=yi)) +  
  facet_wrap(~measCat, scales='free', ncol=3) +  
  stat_qq() + mytheme + ggtitle('QQ Plots of \nstd. measurement values')  
qq
```



```
newfilename<-"qq_ESmeas.png"  
png(paste(figuresPath,newfilename, sep='/'),  
    units='in', width = fig.width*3, height = fig.height*6, res=fig.res)  
qq  
dev.off()
```

```
## pdf  
## 2
```

```
#Plot Quality Histograms
```

```
#re-order measQuality levels
```

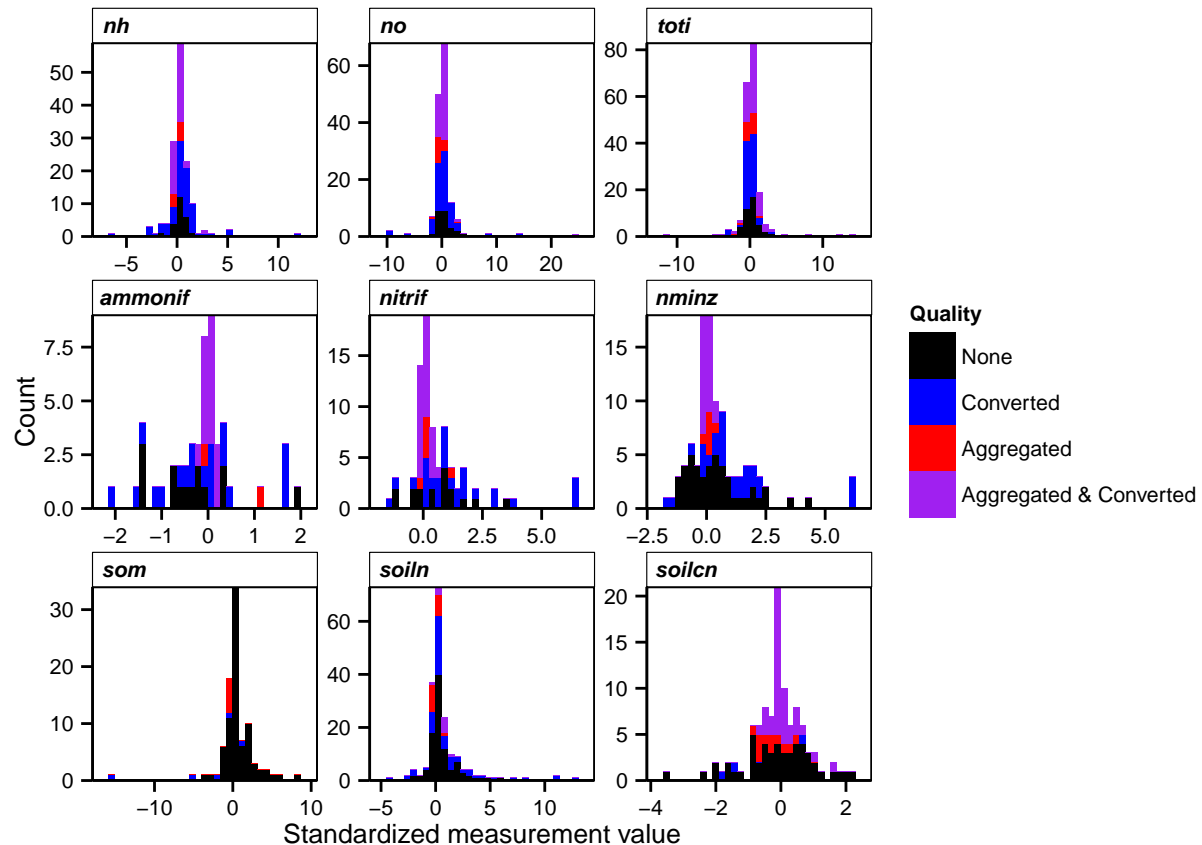
```
tmp$measQuality <- factor(tmp$measQuality, levels = c('NoAgg.NoConv', 'NoAgg.Conv', 'Agg.NoConv', 'Agg.Conv'))
```

```
pHist_ESmeasQual<-ggplot(data=tmp, aes(x=yi,fill=measQuality)) + mytheme +  
  facet_wrap(~measCat, scales='free', ncol=3) + geom_histogram() +  
  scale_y_continuous(expand = c(0,0)) +  
  scale_fill_manual(name = "Quality",
```

```

labels = c("Agg.Conv"="Aggregated & Converted",
           "Agg.NoConv"="Aggregated",
           "NoAgg.Conv"="Converted",
           "NoAgg.NoConv"="None"),
values=c("Agg.Conv" = "purple",
          "Agg.NoConv" = "red",
          "NoAgg.Conv" = "blue",
          "NoAgg.NoConv" = "black")) +
ylab('Count') + xlab('Standardized measurement value')
pHist_ESmeasQual

```



```

newfilename<-"pHist_ESmeasQual.png"
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*3.5, height = fig.height*6, res=fig.res)
pHist_measQual
dev.off()

```

```

## pdf
## 2

```


9. CWM trait value statistics

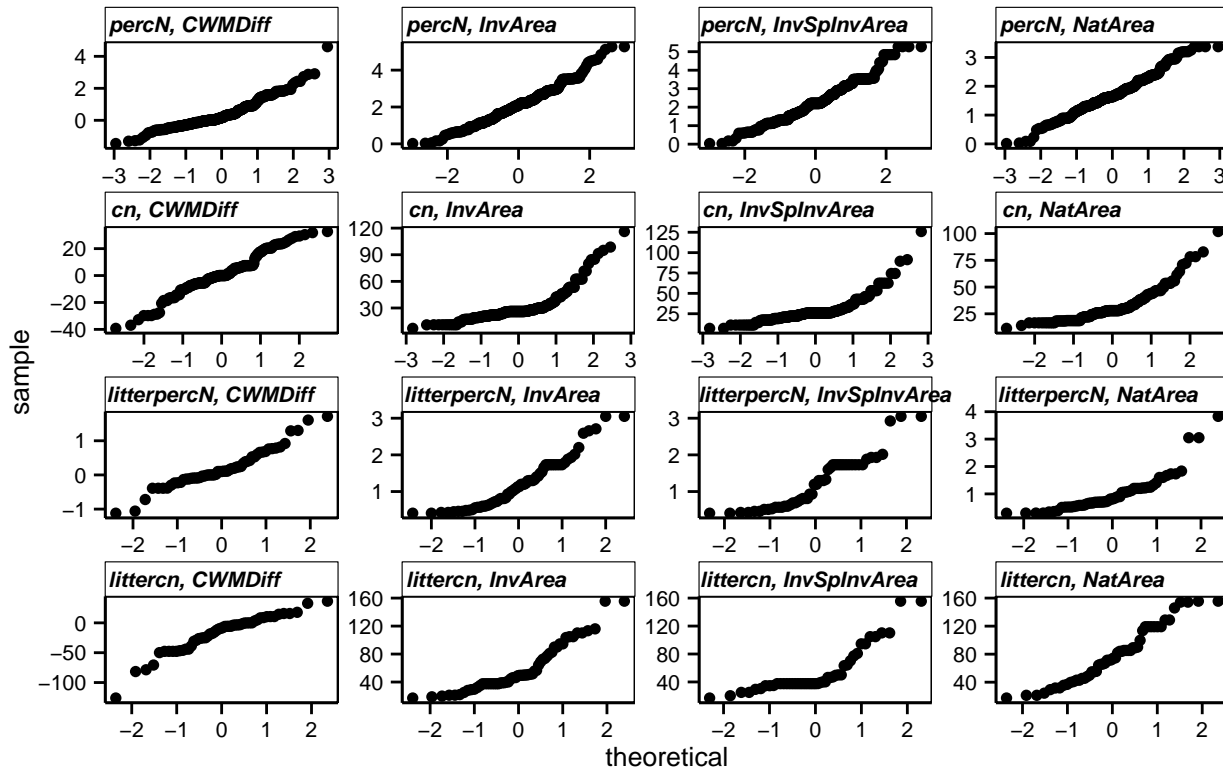
```
#re-order measCat levels
metaDataset$traitCat <- factor(metaDataset$traitCat, levels = traitCat_order)
#re-shape measures so that inv and nat are in the same column temporarily
tmp<-ddply(metaDataset, ~obsID+traitCat, summarize,
  InvArea_cwm = unique(InvArea_cwm),
  InvSpInvArea_cwm = unique(InvSpInvArea_cwm),
  NatArea_cwm = unique(NatArea_cwm),
  CWMDiff_cwm = unique(CWMDiff_cwm),
  InvArea_qualRank = unique(InvArea_qualRank),
  InvSpInvArea_qualRank = unique(InvSpInvArea_qualRank),
  NatArea_qualRank = unique(NatArea_qualRank),
  CWMDiff_qualRank = unique(InvArea_qualRank)+ unique(NatArea_qualRank))

tmp$obsID<-as.factor(tmp$obsID)
m.tmp<-melt(tmp, idcols=c('obsID','traitCat'))
m.tmp$dataType<-rep(NA,length(dim(m.tmp)[1])) #dataType
m.tmp[grepl('_qualRank', m.tmp$variable),'dataType']<-'qualRank'
m.tmp[grepl('_cwm', m.tmp$variable),'dataType']<-'cwm'
m.tmp$invType<-rep(NA,length(dim(m.tmp)[1])) #invType
m.tmp[grepl('InvArea', m.tmp$variable),'invType']<-'InvArea'
m.tmp[grepl('InvSpInvArea', m.tmp$variable),'invType']<-'InvSpInvArea'
m.tmp[grepl('NatArea', m.tmp$variable),'invType']<-'NatArea'
m.tmp[grepl('CWMDiff', m.tmp$variable),'invType']<-'CWMDiff'
c.tmp<-dcast(m.tmp, obsID+traitCat+invType~dataType)
c.tmp<-c.tmp[!is.na(c.tmp$cwm),]

# #Shapiro Test
# ddply(cwm, ~traitCat, summarise,
#   shapTest=shapiro.test(cwm)$p.value,
#   shapTestLn=shapiro.test(log10(cwm))$p.value)
# #none are normal according to Shapiro test

# Q-Q plots
qq<-ggplot(c.tmp, aes(sample=cwm)) +
  facet_wrap(~traitCat+invType, scales='free', ncol=4) +
  stat_qq() + mytheme + ggtitle('QQ Plots of \n cwm trait values')
qq
```

QQ Plots of cwm trait values



```
newfilename<-"qq_cwm.png"
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*4, height = fig.height*3, res=fig.res)
qq
dev.off()
```

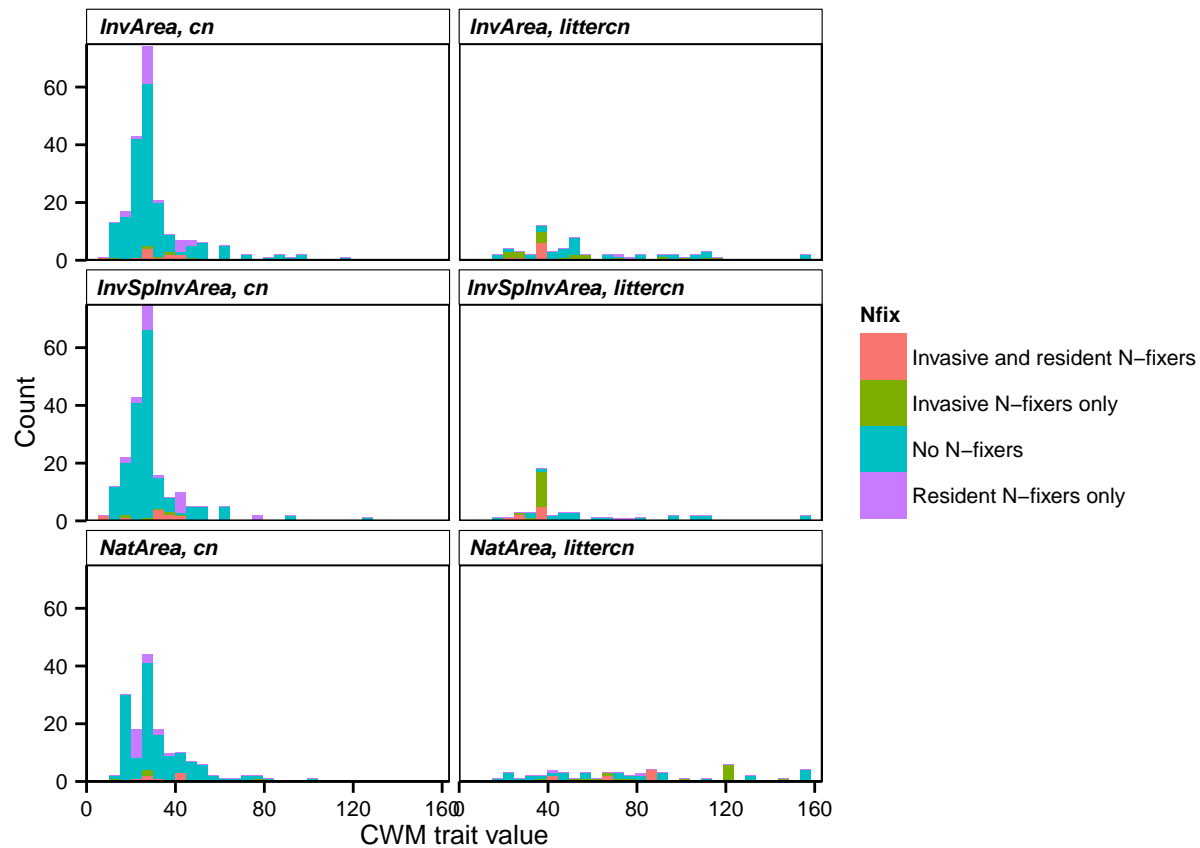
```
## pdf
## 2
```

```
#Plot Factor Histograms
#InvType
cwm$obsID<-as.factor(cwm$obsID)
cwm$n_invSp_invArea<-as.factor(cwm$n_invSp_invArea)
cwm$n_invSp_natArea<-as.factor(cwm$n_invSp_natArea)
cwm$n_natSp_invArea<-as.factor(cwm$n_natSp_invArea)
cwm$n_natSp_natArea<-as.factor(cwm$n_natSp_natArea)

#Nfix
cwm.tmp<-merge(cwm, observations, by='obsID')

#plot
cwm.tmp.cn<-subset(cwm.tmp, traitCat %in% c('cn','littercn'))
pHist_cwm_cn<-ggplot(data=cwm.tmp.cn, aes(x=cwm,fill=Nfix)) +
  facet_wrap(~invType+traitCat, scales='fixed',ncol=2) +
  scale_y_continuous(expand=c(0,0)) + scale_x_continuous(expand=c(0,0)) +
  geom_histogram() + mytheme +
```

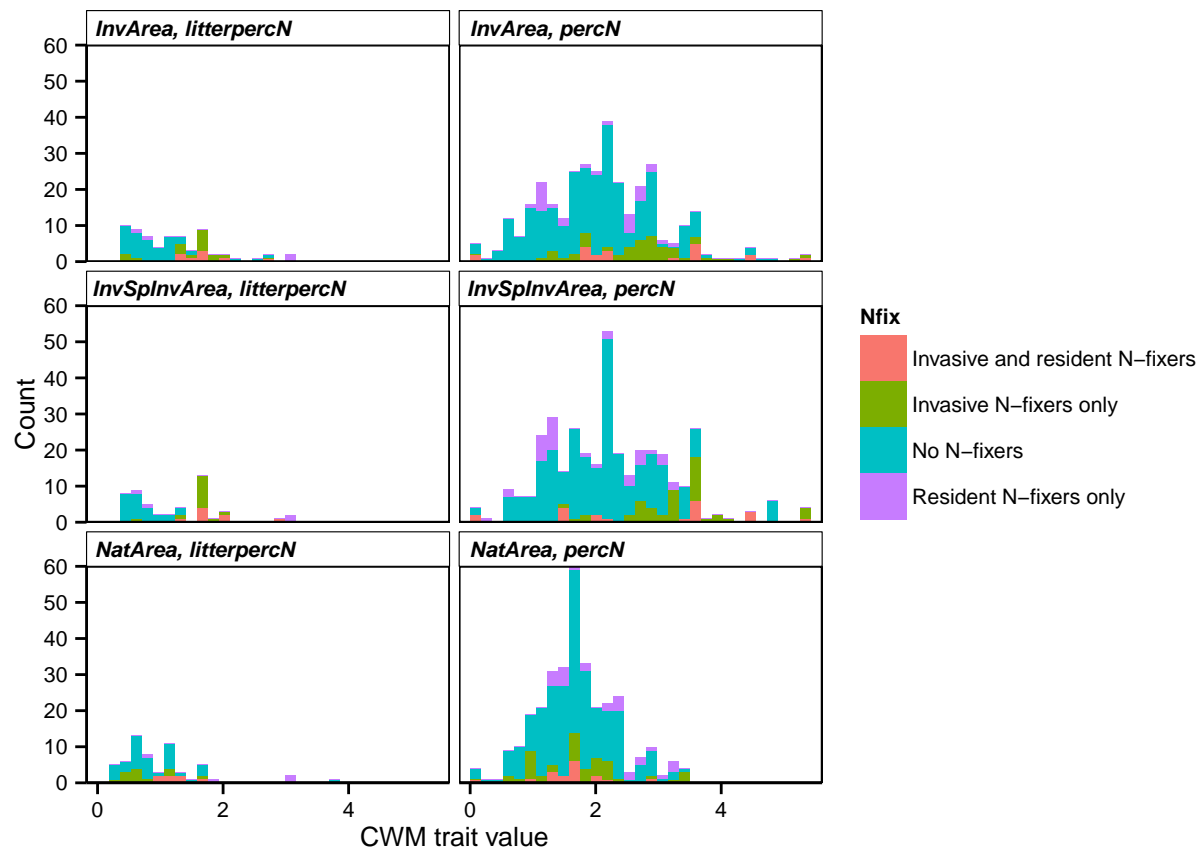
```
ylab('Count') + xlab('CWM trait value')
pHist_cwm_cn
```



```
newfilename<-"pHist_cwm_cn.png"
png(paste(figuresPath,newfilename, sep='/'),
     units='in', width = fig.width*2.5, height = fig.height*3, res=fig.res)
pHist_cwm_cn
dev.off()
```

```
## pdf
## 2
```

```
cwm.tmp.percn<-subset(cwm.tmp, traitCat %in% c('percN','litterpercN'))
pHist_cwm_percn<-ggplot(data=cwm.tmp.percn, aes(x=cwm,fill=Nfix)) +
  facet_wrap(~invType+traitCat, scales='fixed',ncol=2) +
  scale_y_continuous(expand=c(0,0)) + scale_x_continuous(expand=c(0,0)) +
  geom_histogram() + mytheme +
  ylab('Count') + xlab('CWM trait value')
pHist_cwm_percn
```



```

newfilename<-"pHist_cwm_percn.png"
png(paste(figuresPath,newfilename, sep='/'),
    units='in', width = fig.width*2.5, height = fig.height*3, res=fig.res)
pHist_cwm_percn
dev.off()

```

```

## pdf
## 2

```