

# RWorksheet\_5.Rmd

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1. Create a data frame for the table below. Show your solution.

```
student <- c(1,2,3,4,5,6,7,8,9,10)
pre_test <- c(55,54,47,57,51,61,57,54,63,58)
post_test <-c(61,60,56,63,56,63,59,56,62,61)
```

```
Student <- data.frame(
  Student = student,
  Pre_Test = pre_test,
  Post_Test = post_test
)
```

Student

##	Student	Pre_Test	Post_Test
## 1	1	55	61
## 2	2	54	60
## 3	3	47	56
## 4	4	57	63
## 5	5	51	56
## 6	6	61	63
## 7	7	57	59
## 8	8	54	56
## 9	9	63	62
## 10	10	58	61

- a. Compute the descriptive statistics using different packages (Hmisc and pastecs). Write the codes and its result.

```
library(Hmisc)
```

```
## Loading required package: coda
```

```
summary(Student)
```

##	Student	Pre_Test	Post_Test
## Min.	: 1.00	Min. :47.00	Min. :56.00
## 1st Qu.:	: 3.25	1st Qu.:54.00	1st Qu.:56.75
## Median :	: 5.50	Median :56.00	Median :60.50
## Mean :	: 5.50	Mean :55.70	Mean :59.70
## 3rd Qu.:	: 7.75	3rd Qu.:57.75	3rd Qu.:61.75
## Max.	:10.00	Max. :63.00	Max. :63.00

```
library(pastecs)
```

```
stat.desc(Student)
```

```
##           Student      Pre_Test      Post_Test
## nbr.val    10.0000000    10.0000000    10.0000000
## nbr.null    0.0000000    0.0000000    0.0000000
## nbr.na      0.0000000    0.0000000    0.0000000
## min         1.0000000    47.0000000    56.0000000
## max        10.0000000    63.0000000    63.0000000
## range       9.0000000    16.0000000     7.0000000
## sum        55.0000000   557.0000000   597.0000000
## median      5.5000000    56.0000000    60.5000000
## mean       5.5000000    55.7000000    59.7000000
## SE.mean     0.9574271     1.46855938    0.89504811
## CI.mean.0.95 2.1658506     3.32211213    2.02473948
## var         9.1666667    21.56666667     8.01111111
## std.dev      3.0276504     4.64399254     2.83039063
## coef.var     0.5504819     0.08337509     0.04741023
```

2. The Department of Agriculture was studying the effects of several levels of a fertilizer on the growth of a plant. For some analyses, it might be useful to convert the fertilizer levels to an ordered factor.

```
fertilizer <- c(10, 10, 10, 20, 20, 50, 10, 20, 10, 50, 20, 50, 20, 10)
ordered <- ordered(fertilizer)
ordered
```

```
## [1] 10 10 10 20 20 50 10 20 10 50 20 50 20 10
## Levels: 10 < 20 < 50
```

The specified order will be reflected in the levels of the ordered\_factor as 10, 20, 50.

3. Abdul Hassan, president of Floor Coverings Unlimited, has asked you to study the exercise levels undertaken by 10 subjects were “l”, “n”, “n”, “i”, “l”, “l”, “n”, “n”, “i”, “l”,

“n”, “i”, “l” ; n=none, l=light, i=intense a. What is the best way to represent this in R?

```
subjects <- c("l", "n", "n", "i", "l", "l", "n", "n", "i", "l")
factor <- factor(subjects, levels=c("n", "l", "i"))
factor
```

```
## [1] l n n i l l n n i l
## Levels: n l i
```

4. Sample of 30 tax accountants from all the states and territories of Australia and their individual state of origin is specified by a character vector of state mnemonics

```
state <- c("tas", "sa", "qld", "nsw", "nsw", "nt", "wa", "wa", "qld",
          "vic", "nsw", "vic", "qld", "qld", "sa", "tas", "sa", "nt",
          "wa", "vic", "qld", "nsw", "nsw", "wa", "sa", "act", "nsw",
          "vic", "vic", "act")
```

- a. Apply the factor function and factor level. Describe the results.

```
state_factor <- factor(state)
summary(state_factor)
```

```
## act nsw nt qld sa tas vic wa
##    2  6  2  5  4  2  5  4
```

*#In the given sample, we can determine the frequency of each state.*

```
state_levels <- c("nsw", "vic", "qld", "wa", "sa", "tas", "nt", "act")

state <- factor(state, levels = state_levels)

summary(state)
```

```
## nsw vic qld wa sa tas nt act
## 6 5 5 4 4 2 2 2
```

*#Offer a straightforward tally of occurrences for each level in the specified order.*

5. From #4 - continuation:

```
incomes <- c(60, 49, 40, 61, 64, 60, 59, 54,
             62, 69, 70, 42, 56, 61, 61, 61, 58, 51, 48,
             65, 49, 49, 41, 48, 52, 46, 59, 46, 58, 43)
state <- c("tas", "sa", "qld", "nsw", "nsw", "nt", "wa", "wa", "qld",
          "vic", "nsw", "vic", "qld", "qld", "sa", "tas", "sa", "nt",
          "wa", "vic", "qld", "nsw", "nsw", "wa", "sa", "act", "nsw",
          "vic", "vic", "act")
incmeans <- tapply(incomes, state, mean)
incmeans
```

```
##      act      nsw      nt      qld      sa      tas      vic      wa
## 44.50000 57.33333 55.50000 53.60000 55.00000 60.50000 56.00000 52.25000
```

b. Copy the results and interpret.

In the ACT, the median income is \$44,500, while in NSW accountants have a median income of \$57,333.33. In the NT, the median income is \$55,500, and in QLD, accountants have a median income of \$53,600. SA reports a median income of \$55,000, while in TAS, accountants have a median income of \$60,500. In VIC, the median income is \$56,000, and accountants in WA earn a median income of \$52,250. These figures offer insights into the central income tendencies for accountants in each region.

6. Calculate the standard errors of the state income means (refer again to number 3)

```
stdError <- function(x) sqrt(var(x)/length(x))

incster <- tapply(incomes, state, stdError)

print(incster)
```

```
##      act      nsw      nt      qld      sa      tas      vic      wa
## 1.500000 4.310195 4.500000 4.106093 2.738613 0.500000 5.244044 2.657536
```

a. What is the standard error? Write the codes.

```
mean_incomes <- tapply(incomes, state, mean)

std_incomes <- tapply(incomes, state, sd)

n_incomes <- tapply(incomes, state, length)

stdError <- function(x) sqrt(var(x)/length(x))
incster <- tapply(incomes, state, stdError)

print(incster)
```

```
##      act      nsw      nt      qld      sa      tas      vic      wa
## 1.500000 4.310195 4.500000 4.106093 2.738613 0.500000 5.244044 2.657536
```

b. Interpret the result.

ACT demonstrates stability at 1.5, with NSW showing resilience at 4.31. Following closely, NT and QLD secure scores of 4.5 and 4.11, signaling robust economic activity. SA posts a score of 2.74, TAS registers 0.5, and VIC takes the lead at 5.24, highlighting economic strength. WA maintains a solid standing at 2.66. These scores offer a concise overview, assisting in targeted interventions and policy considerations.

7. Use the titanic dataset.

```
data("Titanic")
```

a. subset the titanic dataset of those who survived and not survived. Show the codes and its result.

```
data("Titanic")
```

```
no_adult <- as.vector(Titanic[, , "Adult", "No"])
no_child <- as.vector(Titanic[, , "Child", "No"])
yes_adult <- as.vector(Titanic[, , "Adult", "Yes"])
yes_child <- as.vector(Titanic[, , "Child", "Yes"])
```

```
cat("Number of Adults who did not survive:", sum(no_adult), "\n")
```

```
## Number of Adults who did not survive: 1438
```

```
cat("Number of Children who did not survive:", sum(no_child), "\n")
```

```
## Number of Children who did not survive: 52
```

```
cat("Number of Adults who survived:", sum(yes_adult), "\n")
```

```
## Number of Adults who survived: 654
```

```
cat("Number of Children who survived:", sum(yes_child), "\n")
```

```
## Number of Children who survived: 57
```

8. The data sets are about the breast cancer Wisconsin. The samples arrive periodically as Dr. Wolberg reports his clinical cases. The database therefore reflects this

a. describe what is the dataset all about.

The data concentrates on women grappling with breast cancer, employing a survey scale spanning 1 to 10. This scale evaluates diverse attributes of cell nuclei associated with breast cancer, including clump thickness, size uniformity, shape uniformity, marginal adhesion, epithelial size, bare nucleoli, bland chromatin, normal nucleoli, and mitoses. Each score on the scale signifies the severity or abnormality of the corresponding characteristic. The dataset strives to capture and analyze these features to glean insights into the nature of breast cancer among the surveyed women.

d. Compute the descriptive statistics using different packages. Find the values of: d.1 Standard error of the mean for clump thickness.

```
data <- read.csv('breastcancer_wisconsin.csv')
```

```
clump_thickness_column <- data$clump_thickness
stderror <- sd(clump_thickness_column) / sqrt(length(clump_thickness_column))
```

```
print(stderror)
```

```
## [1] 0.1065011
```

d.2 Coefficient of variability for Marginal Adhesion.

```
data <- read.csv('breastcancer_wisconsin.csv')

marginal_adhesion_column <- data$marginal_adhesion
coefficientvar <- sd(marginal_adhesion_column) / mean(marginal_adhesion_column) * 100

print(coefficientvar)
```

```
## [1] 101.7283
```

d.3 Number of null values of Bare Nuclei.

```
data <- read.csv('breastcancer_wisconsin.csv')

bare_nucleoli_column <- data$bare_nucleoli
null <- sum(is.na(bare_nucleoli_column))

print(null)
```

```
## [1] 15
```

d.4 Mean and standard deviation for Bland Chromatin

```
mean <- mean(data$bland_chromatin, )
sd <- sd(data$bland_chromatin, )

print(paste("Mean:", mean))
```

```
## [1] "Mean: 3.43776824034335"
```

```
print(paste("Standard deviation:", sd))
```

```
## [1] "Standard deviation: 2.43836425232425"
```

d.5 Confidence interval of the mean for Uniformity of Cell Shape

```
data <- read.csv('breastcancer_wisconsin.csv')
shape_uniformity <- data$shape_uniformity

anss <- t.test(shape_uniformity)

cat("Mean:", anss$estimate, "\n")
```

```
## Mean: 3.207439
```

```
cat("95% confidence interval:", anss$conf.int[1], anss$conf.int[2], "\n")
```

```
## 95% confidence interval: 2.986741 3.428138
```

d. How many attributes?

```
data <- read.csv('breastcancer_wisconsin.csv')

attributes <- length(names(data))
print(attributes)
```

```
## [1] 11
```

e. Find the percentage of respondents who are malignant. Interpret the results.

```
data <- read.csv('breastcancer_wisconsin.csv')

malignant_count <- sum(data$class == "malignant")
total_count <- nrow(data)

percentage <- (malignant_count / total_count) * 100
print(percentage)
```

```
## [1] 0
```

9. Export the data abalone to the Microsoft excel file. Copy the codes. `install.packages("AppliedPredictiveModeling")`  
`library("AppliedPredictiveModeling")` `view(abalone)` `summary(abalone)`

```
library(openxlsx)
install.packages("AppliedPredictiveModeling")
```

```
## Installing package into '/cloud/lib/x86_64-pc-linux-gnu-library/4.3'
## (as 'lib' is unspecified)
```

```
library("AppliedPredictiveModeling")
```

```
data("abalone")
```

```
head(abalone)
```

```
##   Type LongestShell Diameter Height WholeWeight ShuckedWeight VisceraWeight
## 1    M         0.455   0.365  0.095    0.5140         0.2245         0.1010
## 2    M         0.350   0.265  0.090    0.2255         0.0995         0.0485
## 3    F         0.530   0.420  0.135    0.6770         0.2565         0.1415
## 4    M         0.440   0.365  0.125    0.5160         0.2155         0.1140
## 5    I         0.330   0.255  0.080    0.2050         0.0895         0.0395
## 6    I         0.425   0.300  0.095    0.3515         0.1410         0.0775
##   ShellWeight Rings
## 1         0.150   15
## 2         0.070    7
## 3         0.210    9
## 4         0.155   10
## 5         0.055    7
## 6         0.120    8
```

```
summary(abalone)
```

```
##   Type      LongestShell      Diameter      Height      WholeWeight
## F:1307  Min.   :0.075      Min.   :0.0550  Min.   :0.0000  Min.   :0.0020
## I:1342  1st Qu.:0.450      1st Qu.:0.3500  1st Qu.:0.1150  1st Qu.:0.4415
## M:1528  Median :0.545      Median :0.4250  Median :0.1400  Median :0.7995
##        Mean   :0.524      Mean   :0.4079  Mean   :0.1395  Mean   :0.8287
##        3rd Qu.:0.615      3rd Qu.:0.4800  3rd Qu.:0.1650  3rd Qu.:1.1530
##        Max.   :0.815      Max.   :0.6500  Max.   :1.1300  Max.   :2.8255
## ShuckedWeight VisceraWeight ShellWeight Rings
## Min.   :0.0010  Min.   :0.0005  Min.   :0.0015  Min.   : 1.000
## 1st Qu.:0.1860  1st Qu.:0.0935  1st Qu.:0.1300  1st Qu.: 8.000
## Median :0.3360  Median :0.1710  Median :0.2340  Median : 9.000
## Mean   :0.3594  Mean   :0.1806  Mean   :0.2388  Mean   : 9.934
## 3rd Qu.:0.5020  3rd Qu.:0.2530  3rd Qu.:0.3290  3rd Qu.:11.000
## Max.   :1.4880  Max.   :0.7600  Max.   :1.0050  Max.   :29.000
```

```
write.xlsx(abalone, file = "abalone.xlsx")
```