## Contents

Summary / Zusammenfassung	ii		
Abbreviations	$\mathbf{vi}$		
Introduction	1		
Multiple Myeloma and Other Monoclonal Gammopathies	2		
Dissemination of Myeloma Cells	3		
Retention of Myeloma Cells in the Bone Marrow	4		
Release of Myeloma Cells from the Bone Marrow	5		
MSCs: Mesenchymal Stromal (Stem) Cells	6		
Molecular Interactions between MSCs Myeloma Cells	8		
Multidimensional Data in Biomedical Research	11		
Nontransparencies in Biomedical Data Analyses	12		
Semi-Big Data: Big Enough to Cause Problems	13		
The Shortcomings of Common Biomedical Analysis Tools	14		
Modern Standards of Software Development	16		
What makes Python an "Easy" Programming Language?	18		
The Potential of Python Data Science Packages for Biomedicine	23		
Aims	26		
Chapter 1: Modelling Myeloma Dissemination in vitro	<b>27</b>		
Introduction	28		
Materials and Methods	30		
Results	34		
Discussion	49		
Chapter 2: Semi-Automating Data Analysis with plotastic	<b>53</b>		
Introduction	54		
Statement of Need	56		
Example	57		
Overview	57		
Discussion	61		
Summarising Discussion			
Semi-Automation was Critical for Establishing in vitro Methods	67		
plotastic Exceled in Re-Doing Statistical Analyses and Plots			
Conclusion 1: Demonstrating the Advantages of Semi-Automation in Biomedical Data Analysis	71		
How Exploratory Live-Cell Imaging Transformed the Research Focus	73		
Potential and Challenges of Image Cytometry	75		
Technical Considerations for Automated Microscopy	77		
Conclusion 2: Automating Microscopy, an Emerging Trend for Exploring Unknown Cell Phenomena?	79		
Isolating & Quantifying Subpopulations within Cells in Direct Contact with MSCs	80		
Integrating Evidence and Hypotheses for a Mechanistic Understanding of Dissemination	84		
Hypothesis 1: Cell Attachment and Detachment Dynamics (CADD) is Adapted during Dissemination	86		
Hypothesis 2: High Adaptability of CADD is a Hallmark of Aggressive Myeloma	89		
Hypothesis 3: CADD is Highly Diverse Within both Patients and Cell Lines	91		
Hypothesis 4: Detachment is Caused by Multiple Cues of Varying Nature	92		
Outlook: High-Value Research Topics for Myeloma Research Arising from this Work	93		
Conclusion 3: The Dynamic Adhesion Hypothetical Framework for Myeloma Dissemination	94		
$Overall\ Conclusion\ .$	95		

### Contents

$\mathbf{Re}$	feren	ces	96
Appendices			
$\mathbf{A}$	Supp	plementary Data & Methods	118
	A.1	Figures	. 118
	A.2	Tables	. 135
	A.3	Materials & Methods	. 143
В	Doc	umentation of plotastic	159
	B.1	Class Diagram	. 160
	B.2	Readme	. 162
	B.3	Example Analysis "qpcr"	. 175
$\mathbf{C}$	Subr	mission Forms & Documents	182
	C.1	Author Contributions	. 182
	C.2	Affidavit	. 189
	C.3	Usage of Generative AI and Other Software	. 191
	C.4	Curriculum Vitae	. 195

## Abbreviations

## Introduction

#### Aims

This PhD thesis is designed to bridge significant gaps in the understanding and analysis of myeloma cell behavior and the handling of complex biomedical datasets. The specific aims are as follows:

- Develop an *in vitro* model to elucidate the mechanisms of myeloma cell dissemination in interaction with mesenchymal stromal cells (hMSCs), focusing particularly on:
  - Observing and quantifying cell proliferation, attachment, and detachment dynamics using time-lapse microscopy.
  - Isolating and characterizing distinct myeloma subpopulations interacting with hMSCs to understand differential gene expression related to cell adhesion and patient survival.
- Design and implement a Python-based software tool, plotastic, to facilitate the analysis of multidimensional datasets generated in biomedical research. This tool will aim to:
  - Streamline the data analysis process, making it more efficient and reproducible.
  - Integrate visualization and statistical analysis capabilities to ensure that data analysis
    protocols are aligned with the ways in which data is visualized.
  - Provide a case study demonstrating the application of plotastic in the analysis of in vitro dissemination experiments, emphasizing the tool's ability to handle semi-big data and enhance reproducibility.
- Synthesize the findings from the experimental and software development components to advance
  the understanding of myeloma dissemination and improve research practices in biomedical data
  analysis.

These aims are crafted to address both the biological and technical challenges in current cancer research methodologies and data science applications in biomedicine, fostering advancements that could lead to novel therapeutic strategies and more robust scientific inquiries.

### **Summarising Discussion**

The subsequent sections will discuss the chapters presented earlier, focusing on how they fit within current scientific fields and the technical and academic challenges encountered during this project. Given the extensive scope of the topics covered, this discussion is divided into three main sections: Microscopy, Molecular Biology, and Data Analysis. Each section will detail key experiments that led to shifts in understanding and present intermediary conclusions to ensure clarity on broad topics.

# Isolating & Quantifying Subpopulations within Cells in Direct Contact with MSCs

This project aimed to develop methodologies for isolating cells after direct contact with human Mesenchymal Stromal Cells (hMSCs). The primary challenge was the scarcity of *in vitro* methods that could effectively separate and isolate adhering cell subpopulations for subsequent molecular analysis. Most available techniques predominantly focus on the quantification of cell adhesion (Khalili & Ahmad, 2015; Kashef & Franz, 2015), and often employ indirect contact setups, complex micromanipulation, or are unsuitable for using live hMSCs as the immobilizing surface. To address the limitations of current adhesion assays, we developed and enhanced innovative methodologies, specifically the Well Plate Sandwich Centrifugation (WPSC) and V-Well adhesion assays.

Variability of Washing Steps: Given the complexity of the requirements, this project first attempts relied on simple and traditional adhesion assays that rely on manual washing steps (Humphries, 2009). Washing involves aspirating the medium, dispensing washing buffer, and potentially repeating these steps multiple times. This introduces variability due to differences in pipetting techniques, which affect the accuracy of volume transfer (Guan et al., 2023; Pushparaj, 2020). However, adhesion assays don't rely on precise volume transfer, but accurate detachment of cells adhering at the well bottom. This introduces a new set of considerations for the pipetting technique, especially since cells are highly sensitive to shear forces applied by fluid flow. From the author's experience with washing experiments and subsequent microscopic evaluations (data not shown), several factors could contribute to the variability of washing steps:

- 1. The distance of the pipette tip from the well bottom, which decreases during aspiration.
- 2. The position of the pipette tip relative to the well bottom (center or edge).
- 3. The angle of the pipette tip.
- 4. The speed of aspiration.
- 5. Accidental or intended contact between the pipette tip and the cell layer.
- 6. The residual volume left after aspiration.
- 7. The same considerations apply when dispensing the washing buffer.

In addition to user-dependent factors, other variables such as the cells' position on the well bottom can significantly impact the outcome. To the author's experience, cells located at the edge of the well don't detach as easily as those in the center, while cells touching the edge are almost impossible to remove. This phenomenon is likely related to the *boundary layer effect*, where fluids slow down near the edges of the well (Weyburne, 2014).

Together, since both user-dependent and independent factors can affect the outcome of washing steps, adhesive assays that replace washing are highly desirable. Still, since washing is straightforward

and some variability is alleviated by the disciplined execution of washing protocols, it remains a common method for adhesion assays.

Directly Interacting Cells Contain Unexplored Interaction Scenarios: It is evident that direct and indirect contact to Mesenchymal Stromal Cells (MSCs) have varying effects on myeloma cells. That difference is crucial for understanding changes in the Bone Marrow Microenvironment (BMME) during MM progression (Fairfield et al., 2020; Dziadowicz et al., 2022). These studies utilize well-inserts to co-culture myeloma cells in close—indirect—contact with MSCs. However, such comparison of indirect vs. direct co-culturing methods might not fully represent the complexity of intercellular interactions scenarios found in the BMME. This is exemplified by this project, as it relied on the complex growth behavior: INA-6 cells aggregated homotypically in direct proximity to those adhering heterotypically to hMSCs, and detached through cell division. Furthermore, such methods fail to capture the subtle variations in paracrine signaling concentrations, where even a few micrometers of distance could significantly alter cellular responses.

Such knowledge shifted this project's point of view as well: Initially, our hypothesis focused on direct heterotypic interactions, not expecting a nMA-INA6 population, but rather subpopulations within MA-INA6 cells that are separable by varying adhesion strengths. Hence, our assay employed strict conditions favoring one growth scenario—heterotypic interactions—, with co-cultures providing unlimited hMSC-surface availability causing predominantly heterotypic adhesion, while the short incubation time prevented the formation of aggregates. Despite these measures, our assay still captured cells emerging from recent cell divisions rather than from weak heterotypic adherence as initially hypothesized. This demonstrates the robustness of our method in separating subpopulations that arising from unexpected intercellular interaction scenarios. This can be a major an advantage over methods that summarize direct interactions as one population. Analysing the non-adhering subpopulation within directly interacting cells could provide valuable insights not just in multiple myeloma, but also metastasis of other cancer types.

Minimizing Variability: There are innovative adhesion assays that both support the isolation of nonadherent subpopulations from directly interacting cells, and avoid variability introduced by washing steps.

One simple method involves flipping over a 96-well plate, with surface tension preventing medium spills as non-adhering cells fall to the surface for collection (Zepeda-Moreno et al., 2011). However, we found that the medium in fact did spill occasionally (not shown). Other approaches involve sealing the plate, such as with PCR plate seals, and using centrifugation to separate cells (Reyes & García, 2003; Y. Chen et al., 2021). Despite our efforts, we could not consistently avoid air bubbles, which, after flipping, would contact the cell layer and create dry regions during centrifugation.

The V-Well adhesion assay does not flip, but collects non-adhering cells into the nadir of V-shaped wells during centrifugation (Weetall et al., 2001). This work profited greatly from this method,

while—to our knowledge—being the first to use cell monolayers as the immobilizing surface. We value this method for its precision, as centrifugation applies a uniform and configurable force, while the readout remains straightforward, relying on the total fluorescent brightness rather than individual cell counting.

Specializing in Quantifying Adhesion or Isolating Subpopulations: Most adhesion assays primarily focus on quantification rather than isolation. The author attempted to combine both quantification and isolation, but found that the two goals can be mutually exclusive. The author summarizes the key differences between quantification and isolation approaches as such:

- Cell Manipulation for Harvest vs. Readout:
  - Isolation methods are designed to manipulate cells for easy harvest. For instance, the WPSC method uses a catching plate to collect non-adherent cells for subsequent analysis.
  - Quantification methods, on the other hand, manipulate cells to simplify the readout process. For example, the V-Well assay, which pellets cells into a single location, allowing for a pooled fluorescence measurement without the need for extensive cell handling.
- Optimization for Subsequent Analysis vs. Sample Throughput:
  - Isolation methods are optimized for detailed subsequent analyses, such as RNA or protein analysis. For example, WPSC minimizes the introduction of biases such as those from fluorescent staining, making it suitable for downstream molecular assays.
  - Quantification methods are optimized for high sample throughput. The V-Well assay, as an end-point assay, is designed to efficiently handle multiple treatments simultaneously, providing quick and comparative results with lower cell numbers.
- Handling of Cell Numbers:
  - Isolation methods, such as WPSC, require multiple wells (e.g., 96 wells) to gather a sufficient amount of cells per subpopulation, which is crucial for robust downstream analyses.
  - Quantification methods, exemplified by the V-Well assay, are highly efficient even with low cell numbers.

Thus, this adopted two distinct techniques for isolating and quantifying directly interacting subpopulations, each optimizing for different outcomes, but also supporting the separation of subpopulations within direct intercellular interactions.

Still, it is theoretically possible to insert microscopy steps into the WPSC method to scan the well bottom for later cell counting. Also, this work effectively isolated cell pellets from the V-well plate for subsequent fixation and cell cycle profiling. The process was tedious and required multiple technical replicates to achieve sufficient cell numbers for analysis. It also required removing hMSC from the V-well nadir to prevent contamination during pellet aspiration.

Together, while both methods can combine quantification and isolation, they are optimized to-

wards either of them. Knowing these strengths and weaknesses could help to advance these methods in future studies.

Rationales of the Well Plate Sandwich Centrifugation: Inspired by the principles of both flipping and V-Well adhesion assays, we developed the Well Plate Sandwich Centrifugation (Well Plate Sandwich Centrifugation (WPSC)) method to address the challenges of isolating cell populations. This method innovatively combines elements from both techniques to provide a more reliable approach to cell isolation. One of the key advantages of WPSC is its ability to reduce the variability commonly introduced by manual pipetting. Instead of relying on aspiration, which introduce variability in cell collection and requires touching the well bottom for complete removal of medium, WPSC employs centrifugation to remove non-adhering cells. Medium is then returned by pipetting to repeat the process and maximize non-adhering cell collection, as the number of detachable cells plateau after few rounds of centrifugation. Hence, this approach compromises between minimizing washing variability and isolating larger quantities of cells.

The 96 well plate format has advantages, reducing spilling when flipping the sandwich, as surface tension kept fluids in place. The 96 well plate format also reduces per-well variability by performing the same washing procedure up to 96 times.

The slow centrifugation speeds used during WPSC are also decided after thorough consideration. For this, one has to discuss how exactly non-adhering cells detach during centrifugation. While centrifugal force is an obvious factor, the properties of cell adhesion are unclear under dry conditions during centrifugation. The author assumed that the cells are being pulled along by the medium as it is centrifuged into the catching plate. Hence, the centrifugation speed was chosen as fast enough to transfer the medium, without completely drying the co-culture plate and minimizing overall cell stress.

A significant challenge in WPSC is the dissociation of MA-INA6 from the hMSC monolayer. WPSC employs two distinct techniques to achieve this dissociation. The first technique involves repeated treatment with the gentle digestive enzyme Accutase followed by Magnetic-Activated Cell Sortings (MACSs). MACS, despite being effective, is costly, time-consuming, reduces overall cell yield, and potentially introduces biases due to CD45 antibody selection and the requirement for cold-treatment. The second technique utilizes strong pipetting to physically detach non-adhering cells (termed 'Wash'). It is important to note that these techniques did not affect the protocol on detaching nMA-INA6 from the co-culture, hence providing for a consistent ratio of isolated MA-INA6 to nMA-INA6 across all experiments. Ultimately, we preferred Wash, as MACS had to be performed on all samples to ensure comparability, reducing overall cell yield which became limiting for downstream applications, especially for nMA-INA6 cells. Both methods achieved comparable purity of MA-INA6 cells, with few hMSCs per 10e4 MA-INA6 cells (purity assessment not shown). Wash probably posited from the highly durable nature of primary hMSC monolayers, whereas MACS required dissociation

of the co-culture.

Together, WPSC offers a versatile solution for isolating hMSC-interacting myeloma cells. It successfully balances the need for precision with the ability to handle larger cell quantities. WPSC could be adapted to other cell types that combines monolayer forming and suspension cells.

**Key Points:** Ultimately, this work established two methodologies that could represent a significant advancement in the field of adhesion assays, providing cost-effective, precise, reliable, and reproducible techniques for both isolating and quantifying subpopulations within co-cultures of directly interacting cell types. They offered valuable insights into the mechanisms of MM detachment and are potentially applicable to other research questions that focus on multicellular interactions and complex growth scenarios.

# Integrating Evidence and Hypotheses for a Mechanistic Understanding of Dissemination

The results outlined in Chapter 1 encompass various aspects of multiple myeloma research, including colonization of the BMME, myeloma-MSC interactions, and the association of adhesion factor expression with patient survival and disease stages. Such a broad scope invites the formulation of generalized conclusions, potentially compromising scientific rigor. The following sections aim to clearly separate hypotheses from evidence to guide further research on dissemination.

Integrating Observations of INA-6 in the Multistep Dissemination Model: The results gained in this work fit well into the multistep model proposed by Zeissig et al. (2020). For most steps, observations were made that could inspire further hypotheses and research:

#### 1. Retention:

- Observation: INA-6 cells attach quickly and strongly to hMSCs, forming stable aggregates.
- Hypothesis: Myeloma cells are retained in the bone marrow microenvironment (BMME) through strong adhesion to hMSCs and stable homotypic aggregation.
- Experiment: Inject INA-6 cells into mice and examine bone lesions. Compare the growth patterns in mice co-injected with an ICAM-1 or LFA-1 $\alpha$  antibody, which dissolve homotypic aggregates in vitro and prevent INA-6 growth in vivo (Kawano et al., 1991; Klausz et al., 2017). If disrupting aggregation leads to diffuse bone colonization rather than focal lesions, it supports the hypothesis that strong adhesion and aggregation are crucial for retention in the BMME.

#### 2. Release:

• Observation: INA-6 cells detach from hMSCs through cell division, and external forces can detach single cells from INA-6 aggregates.

- *Hypothesis*: Myeloma cells detach from the BMME through cell division and external forces after reaching a minimal aggregate size.
- Experiment: Inject INA-6 cells into mice and compare the cell cycle profiles of circulating cells versus those in the bone marrow. Enrichment of G1/G0 cells among circulating cells would support the hypothesis that detachment is more likely shortly after cell division.

#### 3. Intra-/Extravasation:

This study did not make experiments to study for intra-/extravasation, but these phenomena could be explored with similar methods, if MSCs were replaced by endothelial cells.

#### 4. Colonization:

- Observation: INA-6 cells exhibit quick attachment to hMSCs within one hour and rapidly upregulate numerous adhesion factors, including Extracellular Matrix (ECM) factors.
- *Hypothesis*: Quick attachment and fast expression of adhesion factors enhance the potential to colonize new niches. This is particularly relevant as INA-6 cells were isolated from the pleura, indicating an ability to colonize extramedullary sites (Burger, Günther, et al., 2001).
- Experiment: Inject INA-6 cells into mice and observe if they colonize extramedullary sites. Compare this to INA-6 cells with reduced adaptability to test the hypothesis. Research is required to find techniques to reduce such putative adaptability, one potential option is using XRK3F2 to inhibit p62, an upstream activator of NF-κB (Adamik et al., 2018). In fact, NF-κB signaling seems a robust target, given that it plays a role both in MM patients (Sarin et al., 2020), and inducing adhesion factor expression in INA-6 (this work).

These hypotheses—based on observations from INA-6 cells—provide a starting point for understanding myeloma dissemination. While these insights are specialized for the INA-6 cell line, they inspire the development of a more generalized framework applicable to a broader range of myeloma cases.

Constructing a Generalizable Hypothetical Framework of Dissemination: A mechanistic understanding of myeloma dissemination remains elusive. Although Zeissig et al. (2020) described dissemination as a multistep process, evidence is largely collected for individual steps, leaving the connections between these steps unproven. As a result, the process of dissemination is a patchwork of evidence fragments. The following sections aim to integrate such fragments, especially those derived from the INA-6 cell line in this work, to construct a more coherent understanding of myeloma dissemination.

In this context, the author introduces the *Dynamic Adhesion Hypothetical Framework for Myeloma Dissemination*, which leverages direct observations of Cell Attachment and Detachment Dynam-

ics (CADD)<sup>13</sup>. CADD characterizes the time-dependent changes in cell adhesion and detachment, associating these phases with molecular signatures like CAM expression or cell signaling mediated by CAMs and the microenvironment. By adding a temporal component, CADD aims to predict attachment and detachment events.

**Key Hypotheses:** The Dynamic Adhesion Hypothetical Framework is structured around four key hypotheses, each addressing fundamental aspects of myeloma cell dissemination based on both literature and the results of this work. These hypotheses are as follows:

- 1. CADD is adapted in response to different microenvironments faced during dissemination
- 2. High adaptability of CADD is a hallmark of aggressive myeloma
- 3. CADD is highly diverse within both patients and cell lines
- 4. Detachment is caused by multiple cues of varying nature, including external mechanical forces, cell division, loss of CAM expression, or even pure chance.

This framework sets the stage for a detailed exploration of each hypothesis, linking empirical data with hypothetical constructs to provide a comprehensive framework that can help to identify commonalities in myeloma dissemination, but also inform the development of targeted therapies.

## Hypothesis 1: Cell Attachment and Detachment Dynamics (CADD) is Adapted during Dissemination

As presented in Chapter 1, MA-INA6 cells exhibit rapid upregulation of both adhesion factors and chemoattractants, adapting their *in vitro* CADD from homotypic aggregation to colonizing MSCs. This dynamic behavior includes the loss of adhesion factor expression after cell division, suggesting that myeloma cells can rapidly change their adhesion factor expression in a highly dynamic manner. Given that INA-6 cells were isolated from an extramedullary site—the pleura—, such changes likely facilitate colonization of new microenvironments. This section explores the hypothesis that MM cells adapt their CADD during each step of dissemination.

CADD Adaptation Assumes Distinguishable Niches: The multistep model proposed by Zeissig et al. (2020) posits that myeloma cells acquire regulatory mechanisms specialized for each step of dissemination. The author hypothesizes that the different niches involved in these steps are unique enough to trigger distinct CADD adaptations. This requires thorough knowledge of separate niches. Granata et al. (2022) categorizes the Bone Marrow (BM) into sinusoidal, arteriolar, and endosteal

<sup>&</sup>lt;sup>13</sup> Cell Attachment and Detachment Dynamics (CADD) (defined in this work): The observation and measurement of time-dependent changes in cell adhesion and detachment events. CADD characterizes the time cells spend attached, migrating or detached and associates these phases with molecular signatures, such as Cell Adhesion Molecule (CAM) expression or cell signaling mediated by CAMs or the microenvironment. CADD expands traditional cell adhesion by a time component and implies an intention to predict attachment and detachment events. A focus on dynamics is especially relevant for suspension cells that exhibit intricate attachment and detachment behavior.

niches, each spatially and molecularly distinguishable. The endosteal niche is home to MSC and a majority of plasma cells<sup>14</sup>, and the vascular niches—sinusoidal and arteriolar—include endothelial cells (Zehentmeier et al., 2014; Wilmore & Allman, 2017). Other niches encountered during dissemination include peripheral blood, lymph nodes, and extramedullary sites. Comprehensive mapping and characterization of these niches, including their adhesion molecules and soluble factors, is necessary to understand the adhesion requirements for each niche. This is a highly complex task, yet summarizing available information per niche could provide a powerful basis.

Distinct Adhesion Phenotypes Transitioning between Niches: Adhesion processes are well-documented in MM progression, particularly within the BMME (Bou Zerdan et al., 2022). However, the dynamism of these processes remains unclear. In other cancers, different adhesive phenotypes and transitions, such as those seen in epithelial-mesenchymal transition (EMT), are common (Geng et al., 2014). For myeloma, EMT-like phenotypes have been described, but a clear association with distinct adhesion behaviors is hindered by the cells maintaining their suspension state (Roccaro et al., 2015; Qian et al., 2023). This work might be the first to identify adhesive subtypes through functional separation of detachable myeloma cells. As presented earlier, expanding these findings could reveal transitions in adhesive phenotypes during MM dissemination, such as overcoming retention, initiating release, and establishing colonization.

Extramedullary Niche: CADD adaptation predicts a specialized set of adhesion factors for extramedullary niches. A distinct phenotype has been proposed for extramedullary myeloma<sup>15</sup>, characterized by changes in expression of CD44, CD56, VLA-4, and CXCR4 (S. Gupta et al., 2022). The role of CXCR4 in mediating adhesion factor expression is well established, particularly in extramedullary MM cells (Roccaro et al., 2015; S. Gupta et al., 2022): Extramedullary myeloma cells overexpress CXCR4, making them more responsive to cues that induce adhesion factor expression, such as CD44/H-CAM.

Vascular Niche: CADD adaptation predicts a specialized set of adhesion factors for endothelial interaction, supporting intravasation and extravasation. Although not assessed in this thesis, the vascular niche is a popular therapeutic target for preventing dissemination (Neri & J. Bahlis, 2012). Key adhesion factors like JAM-A and N-Cadherin have been highlighted as potential targets (Solimando et al., 2020; Mrozik et al., 2015). These factors were not differentially expressed between subpopulations isolated in Chapter 1, suggesting distinct regulatory mechanisms for vascular versus MSC interactions.

<sup>&</sup>lt;sup>14</sup>Wilmore & Allman (2017): "We suggest that it is reasonable to approach the notion of physical plasma cell survival niches with some skepticism. It is clear that most BM plasma cells rely heavily on access to APRIL or BLyS (66, 70), and it appears that mature plasma cells are relatively stationary (59). However to us, that plasma cells must remain indefinitely in physical survival niches to survive is less obvious."

<sup>&</sup>lt;sup>15</sup>S. Gupta et al. (2022): "Our analysis concluded that the gain of CD44, loss of CD56, loss of very late antigen-4 (VLA-4), imbalance of the chemokine receptor-4-chemokine ligand-12 (CXCR4-CXCL12) axis, [...] show an increased propensity [...] to leave the bone marrow and hone in extramedullary sites giving rise to more aggressive extramedullary diseases."

Circulating MM: An adaptive CADD would predict that circulating MM cells lose adhesion factors. Studies confirm that—compared to BM-resident cells—circulating Multiple Myeloma (MM) cells exhibit reduced expression of multiple adhesion factors, including  $\alpha 4\beta 1$  and CD138/Syndecan-1 (Paiva et al., 2013, 2011; Akhmetzyanova et al., 2020). Evidence suggests that a dynamic loss of CD138/Syndecan-1 and gain of JAM-C causes intravasation, circulation, and dissemination of MM cells (Akhmetzyanova et al., 2020; Brandl et al., 2022). This thesis also shows that nMA-INA6 cells, after emerging as daughter cells from MA-INA6, not only lose adhesion factor expression but also exhibit increased survival during IL-6 deprivation, potentially aiding survival in circulation.

Intermediary Conclusion: Evidence for Adhesion Phenotypes Lacks Functional Characterization and Proof of Phenotypic Transitions: The concept of CADD adaptation is supported by the existence of distinct BM niches and the identification of separable adhesion phenotypes. However, most transitions between these phenotypes during dissemination are unexplored. Functional characterization of adhesive phenotypes and their transitions could provide a robust framework for understanding dissemination as a multistep process, reinforcing the dynamic adhesion hypothetical framework. Mapping adhesive properties for each involved niche could aid this endeavor.

Implications for Therapy: Adhesion molecules have been targeted for therapy for over a decade (Nair et al., 2012; Neri & J. Bahlis, 2012). Especially inhibiting adhesion molecules involved in interaction with the endothelium effectively reduces tumor burden in mouse models (Asosingh et al., 2001; Mrozik et al., 2015). A deeper understanding of how myeloma cells regulate CADD could be key to predicting and preventing dissemination. CADD adaptation suggests that different adhesion factors should be either antagonized or agonized depending on their role. For instance, adhesion factors involved in intravasation and extravasation should be antagonized, while those facilitating BM retention should be agonized—Tab. 1 provides a list of potential retention targets.

Considerations for Research on Myeloma Cell Adhesion: Studying adhesion factors in MM in vitro requires considering the specific microenvironmental context. Some adhesion factors are not present in MM cells but can be rapidly expressed with appropriate signals. Also, further studies should differentiate between initial adhesion and upregulated adhesion factors. For example, performing a WPSC assay after 30 minutes of adhesion could separate INA-6-6 cells based on initial adhesion capability, with RNAseq of nMA-INA6 vs MA-INA6 identifying initial adhesion factors. This differentiation could be crucial for predicting colonization potential across niches, as initial adhesion is likely to be essential for subsequent growth in BM or extramedullary environments.

Concluding Remarks: The exploration of CADD adaptation across various niches reveals a complex interplay between myeloma cells and their environments, characterized by a dynamic regulation of adhesion factors. The evidence presented supports the hypothesis that myeloma cells modify their adhesion phenotype in response to the unique demands of each microenvironment they encounter during dissemination. This adaptive capability suggests that targeting these specific adhesion mechanisms.

anisms could offer a promising strategy for therapeutic intervention, particularly in preventing the colonization of new niches. The distinctions between the adhesion phenotypes among the niches—vascular, bone marrow, and extramedullary—underscore the necessity for a targeted approach in therapy, which could involve modulation of specific adhesion factors to either promote retention or prevent dissemination.

Despite these insights, the current understanding of the functional roles and transitions of these adhesion phenotypes during myeloma progression remains incomplete. Future research should focus on delineating these roles more clearly by functional assays and real-time imaging to capture the dynamic changes in adhesion factor expression during cell transition between niches. Such studies will be crucial for validating the CADD adaptation hypothesis and for identifying potential therapeutic targets that could disrupt the dissemination process at various stages.

Future Directions: It is imperative to further characterize these adhesion factors in a controlled in vitro environment, where specific microenvironmental contexts are simulated. This approach will allow for a more nuanced understanding of how adhesion factors are upregulated and their role in niche-specific colonization. By integrating detailed molecular and cellular analyses, such as single-cell RNA sequencing and proteomics, researchers can identify critical adhesion factors that facilitate the initial colonization processes. This knowledge could then inform the development of interventions aimed at either enhancing or inhibiting these factors, thereby potentially providing a more strategic approach to the management and treatment of multiple myeloma.

# Hypothesis 2: High Adaptability of CADD is a Hallmark of Aggressive Myeloma

Chapter 1 demonstrates one peculiar paradox of multiple myeloma (MM) progression: The expression of adhesion factors is decreased as the disease advances, but is swiftly increased in direct contact with MSCs in INA-6 cells—a cell line isolated from highly advanced Plasma Cell Leukemia (PCL)—

This assumption dictates that aggressive myeloma cells gain the ability to dynamically express adhesion factors. It could be that INA-6 has gained the capability to express adhesion factors fast in order to colonize new niches, such as pleura from which they were isolated.

#### Disease Stage:

- Higher disease stages imply changes in adhesion factors that favor aggressiveness. In fact, ECM from myeloma patients shows tumor-promoting properties compared to ECM from healthy donors (Ibraheem et al., 2019)
  - Aggressiveness includes:

- Better Colonization of new niches, including extramedullary ones such as the pleura
- This implies a more diverse set of available adhesion factors
- Faster regulation to adapt to new niches
- Better survival in circulation

Overall, it is known that plasma cells express different adhesion factors compared to healthy plasma cells, implying (Cook et al., 1997; Bou Zerdan et al., 2022).

indeed, 3 temporal subtypes have been identified, associating higher risk with faster changes over time (Keats et al., 2012).

Is Disease stage a proxy for tumor aggressiveness?

yes, adhesion has prognostic value: A recent study by Q. Hu et al. (2024) developed a cell adhesion-based prognostic model for MM, calculating an adhesion-related risk score (ARRS) based on expression of only twelve adhesion related genes.

Supporting Literature:

#### 1. Disease Stage

- THIS WORK: Expression decreases during progression from Monoclonal Gammopathy of Undetermined Significance (MGUS) to Multiple Myeloma Relapse (MMR) of adhesion factors involved in hMSC adhesion.
- The idea that MM pathogenesis involves transformative processes has been around for decades (Hallek et al., 1998), but a detailed understanding of changing adhesive properties is still lacking, especially during the progression of MM.
- It is discussed that myeloma cell lines derived from advanced stages show different expression than newly diagnosed patients, discussing that they come from multiply relapsed patients (Sarin et al., 2020). This work also shows that Myeloma cell lines have the lowest expression of adhesion factors compared to all stages of MM and MGUS.
- For B-Cell Chronic Lymphocytic Leukemia, adhesion molecule expression patterns define distinct phenotypes in disease subsets (De Rossi et al., 1993).
- Terpos et al. (2016) reported an increase in adhesion molecule expression of ICAM-1 and VCAM-1 in patients with MM compared to those with MGUS and asymptomatic Multiple Myeloma (aMM).
- However, Pérez-Andrés et al. (2005) reported that CD40 is downregulated in PCL patients. Hence, different CAMs could serve ambiguous roles in MM progression.

How could this be studied?

Databases of expression from Myeloma cells gathered from bone marrow MGUS, aMM, MM,

MMR already exist Akhmetzyanova et al. (2020); Seckinger et al. (2018). Going through such databases gives a good overview. One could categorize genelists using curated databases, get lists associated with extravasation, intravasation, Bone marrow adhesion. For every gene of these genelists, they could be filtered for significant differences between the stages. Further categorizations of pairwise comparisons of stages are required. but overall, these genelists could be a starting point for This approach is similar to the genelists published in chapter 1, with the difference that the genelist was furthere filtered by the RNAseq results of *in vitro* experiments.

What new implications do these dimensions have on targeting adhesion factors for therapy?

- Specialized treatment for each stage?
- Aggressive MM cells have potantial improved control over adhesion factor expression, regulating a more diverse set of adhesion factors faster. This poses further challenges to targeting. It could be smarter to not target effector-molecules, but rather upstream regulators of adhesion. This work shows that NF- $\kappa$ B signaling, which by itself is not treatable, but regulators downstream of NF- $\kappa$ B were shown to be effective (Adamik et al., 2017, 2018)

# Hypothesis 3: CADD is Highly Diverse Within both Patients and Cell Lines

- Describe different cell lines: MM1.S being plastic adhering moderately MSC-adhering non-aggregating, INA-6 being non adhering aggregate forming and MSC-adhering, U266 being plastic adhering, non MSC-adhering and non-aggregating.
  - Results from this work: CXCL12 expresion varies from QM between QM

One important dimension that is missing here is the genetic background of the myeloma cells. These are based on recurrent patterns of chromosomal aberrations or mutational signatures, defining structural and single nucleotide variants (Kumar & Rajkumar, 2018; Hoang et al., 2019). The prognostic value of genetic variants in MM is well established (Sharma et al., 2021), and their identification is becoming precise and cost-effective using *optical genome mapping*, making progress towards personalized therapies (Zou et al., 2024; Budurlean et al., 2024). The prognostic value of adhesion factor expression is nowhere nearly as advanced, with establishing cell adhesion as a reliable prognostic factor only recently (Q. Hu et al., 2024).

What markers can be used to categorize these differences? - Maybe IL-6 dependency/independency (Sprynski et al., 2009)? - *in vitro* growth characteristics: Plastic adherence, MSC ahderence, aggregation

### Hypothesis 4: Detachment is Caused by Multiple Cues of Varying Nature

biological implications: - Different cues could trigger different adhesional changes - Soluble signals? - Loss of CD138 (Akhmetzyanova et al., 2020) - Detachment through intercellular effects: cell division, Saturation of hMSC adhesion surface - Detachment with mechanical influence: External forces and instability after aggregate size -

why is this important? The cues that trigger the detachment of MM cells are not well understood. It could be that MM cells detach due to a combination of factors, such as loss of adhesion factors, changes in the BM microenvironment, or cell division or even completely random. Knowing specific dissemination signals helps preventing dissemination.

Papers like Akhmetzyanova et al. (2020) make it seem as if there is one molecule that decides if a myeloma cell is circulating or not.

It's less about one clear (molecular) mechanism that decides that a myeloma cell decides to become a disseminating cell, but rather a indirect consequence of a combination of many processes. These processes are: - Loss of adhesion factors or dynamic expression of adhesion factors - Loss of dependency from bone marrow microenvironment - asdf

Our thesis postulates that there is no big switch that decides if a myeloma cell detaches from the bone marrow, but rather a prolonged process of continuisly downregulating adhesion factors, a dynamic upregulation of adhesion factors when they're needed, but the ultimate event that triggers release is better explained by external mechanical forces intercellular effects (cell division, saturation of adhesive surface and rising instability of aggregates after reaching a minimum size).

Detachment is triggered by external mechanical forces on cell conglomerates previously sensitized by changes in cell adhesion behaviour

Supporting Literature:

#### 1. Cues or Processes

- This work showed that detachment happened mostly mechanically and cell biologically through cell division. - Detachment through intercellular effects: cell division, Saturation of hMSC adhesion surface - Detachment with mechanical influence: External forces and instability after aggregate size.
- Soluble signals within the BM microenvironment, such as cytokines and chemokines, play significant roles in modulating adhesion factor expression in MM cells (Aggarwal et al., 2006; Alsayed et al., 2007).

• CD138 was proposed as a switch between adhesion and migration in MM cells, its blockage triggering migration and intravasation (Akhmetzyanova et al., 2020).

How can this be studied?

Identifying such signals might be challenging without having understood the other two hypotheses about adaptability first.

What new implications do these dimensions have on targeting adhesion factors for therapy?

- It could represent a valid strategy to strengthen myeloma adhesion, provided that targeted adhesion molecule is proven to not be involved in other steps of dissemination, such as extravasation. Stimulating adhesion factor expression or activity is harder than inhibition, yet not impossible. For instance, the short polypeptide SP16 can activate the receptor LRP1—its high expression being associated with improved survival of MM patients in this work—, showing promising results during phase I clinical trial (Wohlford et al., 2021), but could potentially increase survival of MM through PI3K/Akt signaling (Potere et al., 2019; Heinemann et al., 2022) -
- One could also accept that many cues are simply not controllable, and hope for systemic therapies like CAR- T Cells

### Outlook: High-Value Research Topics for Myeloma Research Arising from this Work

As an Outlook, the Author lists research topics arising from this work that have great potential for breakthroughs in myeloma research.

Anti tumor effects of MSCs: This thesis has discussed the pro-tumor effects of MSCs. However, MSCs have also been shown to have anti-tumor effects (Galderisi et al., 2015). This work has also shown that primary hMSCs can induce apoptosis in INA-6 6 cells initially—probably through the action of death domain receptors—, but inhibit apoptosis during prolonged culturing.

This shows that hMSCs could be leveraged as a therapeutic target that could prevent myloma progression.

Cell Division as a Mechanism for Dissemination Initiation: The author describes how the detachment of daughter cells from the mother cell after a cycle of hMSC-(re)attachment and proliferation could be a key mechanism in myeloma dissemination. This mechanism was shown in other studies of intra-/extravasation. The author sees great potential in this mechanism as a target for future research. It is probably under-researched due to requirement of sophisticated time-lapse equipment, yet the simplicity of detachment through cell division is intriguing through its simplicity. It implies asymmetric cell division. Cancer cells are known to divide asymmetrically, e.g. moving

miRNAs to one daughter cell.

Lists of Adhesion Gene Associated With Prolonged Patient Survival: The author lists adhesion genes that are associated with prolonged patient survival. These genes are highly expressed in myeloma samples from patients with longer overall

At this time we could be on the verge of a new era of myeloma therapy, including bi-specific antibodies and cell based approaches (Morè et al., 2023; Engelhardt et al., 2024). Currently, available CAR-T Cell therapies (ide-cel, cilta-cel) are extremely expensive, but show complete remission rates of up to 80 % and a 18-month progression free survival rate of 66 % (Bobin & Leleu, 2022). An affordable "off-the-shelf" CAR-T Cell product could become reality since the problem of deadly graft-versus-host disease during allogeneic transplantation seems to be solvable (Qasim et al., 2017), hence, research groups and biotech companies are racing towards developing a safe allogeneic CAR-T Cell technology (Depil et al., 2020).

the list of genes could be good targets because the BM niche is highly hypoxic, car t cells are not well, but directing them to the BM niche could increase efficacy.

Find MSC and Myeloma crosstalk: Do another GSEA analysis using the list from factors upregulated in Dotterweich et al. (2016), since there, INA-6 and primary hMSC were used as well. Redoing an analysis with the background of the associated processes gained here could reveal insights on the communication between hMSC and INA-6 cells.

## Conclusion 3: The Dynamic Adhesion Hypothetical Framework for Myeloma Dissemination

How does limited understanding of one dimension prevent the understanding of the other dimensions?

Location & Progression: If we don't know the expression profile of an MM cell depending on their source, results become incomparable.

Location & Cues: If we don't know the cues that trigger detachment, we can't predict where the MM cells will detach.

### References

- Abadi, M., Agarwal, A., Barham, P., Brevdo, E., Chen, Z., Citro, C., ... Zheng, X. (2016, March). *TensorFlow: Large-Scale Machine Learning on Heterogeneous Distributed Systems* (No. arXiv:1603.04467). arXiv. Retrieved 2024-03-07, from http://arxiv.org/abs/1603.04467 doi: 10.48550/arXiv.1603.04467
- Abdallah, N. H., Lakshman, A., Kumar, S. K., Cook, J., Binder, M., Kapoor, P., ... Rajkumar, S. V. (2024, January). Mode of progression in smoldering multiple myeloma: A study of 406 patients. *Blood Cancer Journal*, 14(1), 1–7. Retrieved 2024-05-22, from https://www.nature.com/articles/s41408-024-00980-5 doi: 10.1038/s41408-024-00980-5
- Abdelrazik, H. (2023, August). Mesenchymal Stem Cells: A Hope or a Hype? International Journal of Molecular Sciences, 24(17), 13218. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10487858/doi: 10.3390/ijms241713218
- Adamik, J., Jin, S., Sun, Q., Zhang, P., Weiss, K. R., Anderson, J. L., ... Galson, D. L. (2017, April). EZH2 or HDAC1 Inhibition Reverses Multiple Myeloma-Induced Epigenetic Suppression of Osteoblast Differentiation. *Molecular cancer research: MCR*, 15(4), 405–417. doi: 10.1158/1541-7786.MCR-16-0242-T
- Adamik, J., Silbermann, R., Marino, S., Sun, Q., Anderson, J. L., Zhou, D., ... Galson, D. L. (2018). XRK3F2 Inhibition of p62-ZZ Domain Signaling Rescues Myeloma-Induced GFI1-Driven Epigenetic Repression of the Runx2 Gene in Pre-osteoblasts to Overcome Differentiation Suppression. Frontiers in Endocrinology, 9, 344. doi: 10.3389/fendo.2018.00344
- Aggarwal, R., Ghobrial, I. M., & Roodman, G. D. (2006, October). Chemokines in multiple myeloma. *Experimental hematology*, 34(10), 1289–1295. Retrieved 2023-04-02, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3134145/doi: 10.1016/j.exphem.2006.06.017
- Akhmetzyanova, I., McCarron, M. J., Parekh, S., Chesi, M., Bergsagel, P. L., & Fooksman, D. R. (2020). Dynamic CD138 surface expression regulates switch between myeloma growth and dissemination. *Leukemia*, 34(1), 245–256. Retrieved 2023-04-04, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6923614/ doi: 10.1038/s41375-019-0519-4
- Allegra, A., Casciaro, M., Barone, P., Musolino, C., & Gangemi, S. (2022, May). Epigenetic Crosstalk between Malignant Plasma Cells and the Tumour Microenvironment in Multiple Myeloma. *Cancers*, 14(11), 2597. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9179362/ doi: 10.3390/cancers14112597
- Alsayed, Y., Ngo, H., Runnels, J., Leleu, X., Singha, U. K., Pitsillides, C. M., ... Ghobrial, I. M. (2007, April). Mechanisms of regulation of CXCR4/SDF-1 (CXCL12)-dependent migration and homing in multiple myeloma. *Blood*, 109(7), 2708–2717. doi: 10.1182/blood-2006-07-035857
- Anders, S., Pyl, P. T., & Huber, W. (2015, January). HTSeq—a Python framework to work with high-throughput sequencing data. *Bioinformatics (Oxford, England)*, 31(2), 166–169. doi: 10.1093/bioinformatics/btu638
- Andrews, S. (2010). FASTQC. A quality control tool for high throughput sequence data.
- Arefin, S. E., Heya, T. A., Al-Qudah, H., Ineza, Y., & Serwadda, A. (2023, July). Unmasking the giant: A comprehensive evaluation of ChatGPT's proficiency in coding algorithms and data structures (No. arXiv:2307.05360). arXiv. Retrieved 2024-05-03, from http://arxiv.org/abs/2307.05360 doi: 10.48550/arXiv.2307.05360
- Armstrong, R. A. (2014, September). When to use the Bonferroni correction. Ophthalmic & Physiological Optics: The Journal of the British College of Ophthalmic Opticians (Optometrists), 34(5), 502–508. doi: 10.1111/opo.12131
- Asosingh, K., Günthert, U., De Raeve, H., Van Riet, I., Van Camp, B., & Vanderkerken, K. (2001). A unique pathway in the homing of murine multiple myeloma cells: CD44v10 mediates binding to bone marrow endothelium. *Cancer Research*, 61(7), 2862–2865.

- Baker, M. (2016, May). 1,500 scientists lift the lid on reproducibility. *Nature*, 533(7604), 452–454. Retrieved 2024-04-22, from https://www.nature.com/articles/533452a doi: 10.1038/533452a
- Bao, L., Lai, Y., Liu, Y., Qin, Y., Zhao, X., Lu, X., ... Huang, X. (2013, September). CXCR4 is a good survival prognostic indicator in multiple myeloma patients. *Leukemia Research*, 37(9), 1083–1088. doi: 10.1016/j.leukres .2013.06.002
- Barnes, D. G., Vidiassov, M., Ruthensteiner, B., Fluke, C. J., Quayle, M. R., & McHenry, C. R. (2013, September). Embedding and Publishing Interactive, 3-Dimensional, Scientific Figures in Portable Document Format (PDF) Files. *PLOS ONE*, 8(9), e69446. Retrieved 2024-06-13, from https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0069446 doi: 10.1371/journal.pone.0069446
- Barzilay, R., Ben-Zur, T., Bulvik, S., Melamed, E., & Offen, D. (2009, May). Lentiviral delivery of LMX1a enhances dopaminergic phenotype in differentiated human bone marrow mesenchymal stem cells. *Stem cells and development*, 18(4), 591–601. doi: 10.1089/scd.2008.0138
- Begley, C. G., & Ioannidis, J. P. A. (2015, January). Reproducibility in science: Improving the standard for basic and preclinical research. *Circulation Research*, 116(1), 116–126. doi: 10.1161/CIRCRESAHA.114.303819
- Berg, S., Kutra, D., Kroeger, T., Straehle, C. N., Kausler, B. X., Haubold, C., ... Kreshuk, A. (2019, December). Ilastik: Interactive machine learning for (bio)image analysis. *Nature Methods*, 16(12), 1226–1232. Retrieved 2024-06-16, from https://www.nature.com/articles/s41592-019-0582-9 doi: 10.1038/s41592-019-0582-9
- Bianco, P. (2014). "Mesenchymal" stem cells. Annual review of cell and developmental biology, 30, 677–704. doi: 10.1146/annurev-cellbio-100913-013132
- BioStudies. (n.d.). BioStudies < The European Bioinformatics Institute < EMBL-EBI. Retrieved 2024-06-12, from https://www.ebi.ac.uk/biostudies/bioimages/studies/S-BIAD1092?key=69bafe9c-74ff-492b-9e68-bd42655c4d1b
- Bladé, J., Beksac, M., Caers, J., Jurczyszyn, A., von Lilienfeld-Toal, M., Moreau, P., ... Richardson, P. (2022, March). Extramedullary disease in multiple myeloma: A systematic literature review. *Blood Cancer Journal*, 12(3), 1–10. Retrieved 2023-03-24, from https://www.nature.com/articles/s41408-022-00643-3 doi: 10.1038/s41408-022-00643-3
- Blonska, M., Zhu, Y., Chuang, H. H., You, M. J., Kunkalla, K., Vega, F., & Lin, X. (2015, February). Junregulated genes promote interaction of diffuse large B-cell lymphoma with the microenvironment. *Blood*, 125(6), 981–991. Retrieved 2023-03-01, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4319238/ doi: 10.1182/blood-2014-04-568188
- Bobin, A., & Leleu, X. (2022, September). Recent advances in the treatment of multiple myeloma: A brief review. Faculty Reviews, 11, 28. Retrieved 2024-03-27, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9523543/doi: 10.12703/r/11-28
- Bokeh Development Team. (2018). Bokeh: Python library for interactive visualization [Manual]. Retrieved from https://bokeh.pydata.org/en/latest/
- Bolado-Carrancio, A., Rukhlenko, O. S., Nikonova, E., Tsyganov, M. A., Wheeler, A., Garcia-Munoz, A., ... Kholodenko, B. N. (2020, July). Periodic propagating waves coordinate RhoGTPase network dynamics at the leading and trailing edges during cell migration. *eLife*, 9, e58165. Retrieved 2024-04-25, from https://elifesciences.org/articles/58165 doi: 10.7554/eLife.58165
- Bondi, A. B. (2000, September). Characteristics of scalability and their impact on performance. In *Proceedings of the 2nd international workshop on Software and performance* (pp. 195–203). New York, NY, USA: Association for Computing Machinery. Retrieved 2024-03-07, from https://dl.acm.org/doi/10.1145/350391.350432 doi: 10.1145/350391.350432
- Bosch-Queralt, M., Tiwari, V., Damkou, A., Vaculčiaková, L., Alexopoulos, I., & Simons, M. (2022, March). A fluorescence microscopy-based protocol for volumetric measurement of lysolecithin lesion-associated de- and re-

- myelination in mouse brain. STAR protocols, 3(1), 101141. doi: 10.1016/j.xpro.2022.101141
- Boswell, D., & Foucher, T. (2011). The Art of Readable Code: Simple and Practical Techniques for Writing Better Code. "O'Reilly Media, Inc.".
- Bou Zerdan, M., Nasr, L., Kassab, J., Saba, L., Ghossein, M., Yaghi, M., ... Chaulagain, C. P. (2022). Adhesion molecules in multiple myeloma oncogenesis and targeted therapy. *International Journal of Hematologic Oncology*, 11(2), IJH39. Retrieved 2023-02-01, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9136637/ doi: 10.2217/ijh-2021-0017
- Brandl, A., Solimando, A. G., Mokhtari, Z., Tabares, P., Medler, J., Manz, H., ... Beilhack, A. (2022, March). Junctional adhesion molecule C expression specifies a CD138low/neg multiple myeloma cell population in mice and humans. *Blood Advances*, 6(7), 2195–2206. Retrieved 2023-04-04, from https://doi.org/10.1182/bloodadvances.2021004354 doi: 10.1182/bloodadvances.2021004354
- Brankatschk, R., Bodenhausen, N., Zeyer, J., & Bürgmann, H. (2012, June). Simple Absolute Quantification Method Correcting for Quantitative PCR Efficiency Variations for Microbial Community Samples. *Applied and Environmental Microbiology*, 78(12), 4481–4489. Retrieved 2023-05-27, from https://journals.asm.org/doi/10.1128/AEM.07878-11 doi: 10.1128/AEM.07878-11
- Breiman, L. (2001, October). Random Forests. *Machine Learning*, 45(1), 5–32. Retrieved 2024-06-14, from https://doi.org/10.1023/A:1010933404324 doi: 10.1023/A:1010933404324
- Brooke, J. (1996, January). SUS a quick and dirty usability scale. In (pp. 189–194).
- Bubendorf, L. (2001, August). High-throughput microarray technologies: From genomics to clinics. European Urology, 40(2), 231–238. doi: 10.1159/000049777
- Budurlean, L., Tukaramrao, D. B., Zhang, L., Dovat, S., & Broach, J. (2024, March). Integrating Optical Genome Mapping and Whole Genome Sequencing in Somatic Structural Variant Detection. *Journal of Personalized Medicine*, 14(3), 291. Retrieved 2024-06-23, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10971281/ doi: 10.3390/jpm14030291
- Burger, R., Guenther, A., Bakker, F., Schmalzing, M., Bernand, S., Baum, W., ... Gramatzki, M. (2001). Gp130 and ras mediated signaling in human plasma cell line INA-6: A cytokine-regulated tumor model for plasmacytoma. *The Hematology Journal: The Official Journal of the European Haematology Association*, 2(1), 42–53. doi: 10.1038/sj.thj.6200075
- Burger, R., Günther, A., Bakker, F., Schmalzing, M., Bernand, S., Baum, W., ... Gramatzki, M. (2001, January). Gp130 and ras mediated signaling in human plasma cell line INA6: A cytokine-regulated tumor model for plasmacytoma. Hematology Journal - HEMATOL J, 2, 42–53. doi: 10.1038/sj.thj.6200075
- Bustin, S. A. (2014, December). The reproducibility of biomedical research: Sleepers awake! *Biomolecular Detection and Quantification*, 2, 35-42. Retrieved 2024-03-18, from https://www.sciencedirect.com/science/article/pii/S2214753515000030 doi: 10.1016/j.bdq.2015.01.002
- Bustin, S. A., Benes, V., Garson, J., Hellemans, J., Huggett, J., Kubista, M., ... Vandesompele, J. (2013, November). The need for transparency and good practices in the qPCR literature. *Nature Methods*, 10(11), 1063–1067. Retrieved 2024-05-16, from https://www.nature.com/articles/nmeth.2697 doi: 10.1038/nmeth.2697
- Caplan, A. (1991). Mesenchymal stem cells. Journal of orthopaedic research: official publication of the Orthopaedic Research Society, 9(5), 641–50. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/1870029 doi: 10.1002/jor.1100090504
- Caplan, A. I. (1994, July). The mesengenic process. Clinics in plastic surgery, 21(3), 429–435.
- Carlson, M. (2016). Org.Hs.eg.db. *Bioconductor*. Retrieved 2023-06-09, from http://bioconductor.org/packages/org.Hs.eg.db/doi: 10.18129/B9.bioc.org.Hs.eg.db
- Chacon, S., & Straub, B. (2024, March). Git Book. Retrieved 2024-03-07, from https://git-scm.com/book/de/v2 Charlier, F., Weber, M., Izak, D., Harkin, E., Magnus, M., Lalli, J., ... Repplinger, S. (2022, October). Tree

- vismd/statannotations: V0.5. Zenodo. Retrieved 2023-11-16, from https://zenodo.org/record/7213391 doi: 10.5281/ZENODO.7213391
- Chatterjee, M., Hönemann, D., Lentzsch, S., Bommert, K., Sers, C., Herrmann, P., ... Bargou, R. C. (2002, November). In the presence of bone marrow stromal cells human multiple myeloma cells become independent of the IL-6/gp130/STAT3 pathway. *Blood*, 100(9), 3311–3318. doi: 10.1182/blood-2002-01-0102
- Chauhan, D., Uchiyama, H., Akbarali, Y., Urashima, M., Yamamoto, K., Libermann, T., & Anderson, K. (1996, February). Multiple myeloma cell adhesion-induced interleukin-6 expression in bone marrow stromal cells involves activation of NF-??B. *Blood*, 87, 1104–12. doi: 10.1182/blood.V87.3.1104.bloodjournal8731104
- Chen, H., & Zhou, L. (2022, June). Treatment of ischemic stroke with modified mesenchymal stem cells. *International Journal of Medical Sciences*, 19(7), 1155–1162. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9339408/ doi: 10.7150/ijms.74161
- Chen, W.-C., Hu, G., & Hazlehurst, L. A. (2020, October). Contribution of the bone marrow stromal cells in mediating drug resistance in hematopoietic tumors. *Current Opinion in Pharmacology*, 54, 36–43. Retrieved 2022-12-12, from https://www.sciencedirect.com/science/article/pii/S1471489220300576 doi: 10.1016/j.coph.2020.08.006
- Chen, Y., Wang, Q., Mills, C. E., Kann, J. G., Shull, K. R., Tullman-Ercek, D., & Wang, M. (2021, July). High-Throughput Screening Test for Adhesion in Soft Materials Using Centrifugation. *ACS Central Science*, 7(7), 1135–1143. Retrieved 2024-06-18, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8323114/ doi: 10.1021/acscentsci.1c00414
- Cheng, Y., Li, W., Jin, T., Wu, S., & Zhang, L. (2023, February). [Frontiers and development in live-cell super-resolution fluorescence microscopy]. Sheng Wu Yi Xue Gong Cheng Xue Za Zhi = Journal of Biomedical Engineering = Shengwu Yixue Gongchengxue Zazhi, 40(1), 180–184. doi: 10.7507/1001-5515.202210060
- Cippitelli, M., Stabile, H., Kosta, A., Petillo, S., Lucantonio, L., Gismondi, A., ... Fionda, C. (2023, January). Role of NF-κB Signaling in the Interplay between Multiple Myeloma and Mesenchymal Stromal Cells. *International Journal of Molecular Sciences*, 24(3), 1823. Retrieved 2024-06-08, from https://www.mdpi.com/1422-0067/24/3/1823 doi: 10.3390/ijms24031823
- Codecov. (2024). Retrieved 2024-05-02, from https://github.com/codecov
- Cole, R. (2014). Live-cell imaging. Cell Adhesion & Migration, 8(5), 452–459. doi: 10.4161/cam.28348
- Committee on Strategies for Responsible Sharing of Clinical Trial Data, Board on Health Sciences Policy, & Institute of Medicine. (2015). Sharing Clinical Trial Data: Maximizing Benefits, Minimizing Risk. Washington (DC): National Academies Press (US). Retrieved 2024-04-23, from http://www.ncbi.nlm.nih.gov/books/NBK269030/
- Cook, G., Dumbar, M., & Franklin, I. M. (1997). The role of adhesion molecules in multiple myeloma. *Acta Haematologica*, 97(1-2), 81–89. doi: 10.1159/000203663
- Cooper, G. M. (2000). The Cell: A Molecular Approach. 2nd Edition. *Sinauer Associates*, Proliferation in Development and Differentiation. Retrieved from https://www.ncbi.nlm.nih.gov/books/NBK9906/
- da Silva Meirelles, L., Chagastelles, P. C., & Nardi, N. B. (2006, June). Mesenchymal stem cells reside in virtually all post-natal organs and tissues. *Journal of cell science*, 119(Pt 11), 2204–2213. doi: 10.1242/jcs.02932
- Davidson-Pilon, C. (2019, August). Lifelines: Survival analysis in Python. *Journal of Open Source Software*, 4(40), 1317. Retrieved 2024-05-02, from https://joss.theoj.org/papers/10.21105/joss.01317 doi: 10.21105/joss.01317
- Depil, S., Duchateau, P., Grupp, S. A., Mufti, G., & Poirot, L. (2020, March). 'Off-the-shelf' allogeneic CAR T cells: Development and challenges. *Nature Reviews Drug Discovery*, 19(3), 185–199. Retrieved 2024-03-27, from https://www.nature.com/articles/s41573-019-0051-2 doi: 10.1038/s41573-019-0051-2
- De Rossi, G., Zarcone, D., Mauro, F., Cerruti, G., Tenca, C., Puccetti, A., ... Grossi, C. E. (1993, May). Adhesion Molecule Expression on B-Cell Chronic Lymphocytic Leukemia Cells: Malignant Cell Phenotypes Define Distinct Disease Subsets. *Blood*, 81(10), 2679–2687. Retrieved 2024-06-20, from https://www.sciencedirect.com/science/article/pii/S0006497120678636 doi: 10.1182/blood.V81.10.2679.2679

- Ding, W., Goldberg, D., & Zhou, W. (2023, August). PyComplexHeatmap: A Python package to visualize multimodal genomics data. *iMeta*, 2(3), e115. doi: 10.1002/imt2.115
- Dobin, A., Davis, C. A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., ... Gingeras, T. R. (2013, January). STAR: Ultrafast universal RNA-seq aligner. *Bioinformatics*, 29(1), 15–21. Retrieved 2023-05-27, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3530905/ doi: 10.1093/bioinformatics/bts635
- Doddi, S., & Rashid, M. H. (2024). Disparities in Multiple Myeloma Mortality Rate Trends by Demographic Status in the USA. Cancer Diagnosis & Prognosis, 4(3), 288–294. doi: 10.21873/cdp.10322
- Dominici, M., Le Blanc, K., Mueller, I., Slaper-Cortenbach, I., Marini, F., Krause, D., ... Horwitz, E. (2006). Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy*, 8(4), 315–317. doi: 10.1080/14653240600855905
- Dotterweich, J., Schlegelmilch, K., Keller, A., Geyer, B., Schneider, D., Zeck, S., ... Schütze, N. (2016, December). Contact of myeloma cells induces a characteristic transcriptome signature in skeletal precursor cells -Implications for myeloma bone disease. *Bone*, 93, 155–166. doi: 10.1016/j.bone.2016.08.006
- D'souza, N., Rossignoli, F., Golinelli, G., Grisendi, G., Spano, C., Candini, O., ... Dominici, M. (2015, August). Mesenchymal stem/stromal cells as a delivery platform in cell and gene therapies. *BMC medicine*, 13, 186. doi: 10.1186/s12916-015-0426-0
- D'Souza, S., del Prete, D., Jin, S., Sun, Q., Huston, A. J., Kostov, F. E., ... Galson, D. L. (2011, December). Gfi1 expressed in bone marrow stromal cells is a novel osteoblast suppressor in patients with multiple myeloma bone disease. *Blood*, 118(26), 6871–6880. Retrieved 2024-06-08, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3245209/ doi: 10.1182/blood-2011-04-346775
- Dunn, W., Burgun, A., Krebs, M.-O., & Rance, B. (2017, November). Exploring and visualizing multidimensional data in translational research platforms. *Briefings in Bioinformatics*, 18(6), 1044–1056. Retrieved 2024-04-23, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5862238/ doi: 10.1093/bib/bbw080
- Duvall, P., Matyas, S., & Glover, A. (2007). Continuous integration: Improving software quality and reducing risk. Pearson Education. Retrieved from https://books.google.de/books?id=PV9qfEdv9L0C
- Dziadowicz, S. A., Wang, L., Akhter, H., Aesoph, D., Sharma, T., Adjeroh, D. A., ... Hu, G. (2022, January). Bone Marrow Stroma-Induced Transcriptome and Regulome Signatures of Multiple Myeloma. *Cancers*, 14(4), 927. Retrieved 2022-10-25, from https://www.mdpi.com/2072-6694/14/4/927 doi: 10.3390/cancers14040927
- Ekmekci, B., McAnany, C. E., & Mura, C. (2016, July). An Introduction to Programming for Bioscientists: A Python-Based Primer. *PLOS Computational Biology*, 12(6), e1004867. Retrieved 2024-03-10, from https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1004867 doi: 10.1371/journal.pcbi.1004867
- Engelhardt, M., Kortüm, K. M., Goldschmidt, H., & Merz, M. (2024, February). Functional cure and long-term survival in multiple myeloma: How to challenge the previously impossible. *Haematologica*. doi: 10.3324/haematol.2023.283058
- Evers, M., Schreder, M., Stühmer, T., Jundt, F., Ebert, R., Hartmann, T. N., ... Leich, E. (2023, March). Prognostic value of extracellular matrix gene mutations and expression in multiple myeloma. *Blood Cancer Journal*, 13(1), 43. doi: 10.1038/s41408-023-00817-7
- Ewels, P., Magnusson, M., Lundin, S., & Käller, M. (2016, October). MultiQC: Summarize analysis results for multiple tools and samples in a single report. *Bioinformatics*, 32(19), 3047–3048. Retrieved 2023-06-09, from https://doi.org/10.1093/bioinformatics/btw354 doi: 10.1093/bioinformatics/btw354
- Excel, M. (2023, August). Announcing Python in Excel: Combining the power of Python and the flexibility of Excel. Retrieved 2024-03-11, from https://techcommunity.microsoft.com/t5/excel-blog/announcing-python-in-excel-combining-the-power-of-python-and-the/ba-p/3893439
- Fairfield, H., Costa, S., Falank, C., Farrell, M., Murphy, C. S., D'Amico, A., ... Reagan, M. R. (2020). Multiple Myeloma Cells Alter Adipogenesis, Increase Senescence-Related and Inflammatory Gene Transcript Expression, and

- Alter Metabolism in Preadipocytes. Frontiers in Oncology, 10, 584683. doi: 10.3389/fonc.2020.584683
- Federer, L. M., Lu, Y.-L., & Joubert, D. J. (2016, January). Data literacy training needs of biomedical researchers. *Journal of the Medical Library Association : JMLA*, 104(1), 52–57. Retrieved 2024-04-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4722643/ doi: 10.3163/1536-5050.104.1.008
- Fermand, J.-P., Bridoux, F., Dispenzieri, A., Jaccard, A., Kyle, R. A., Leung, N., & Merlini, G. (2018, October). Monoclonal gammopathy of clinical significance: A novel concept with therapeutic implications. *Blood*, 132(14), 1478–1485. doi: 10.1182/blood-2018-04-839480
- Fernandez-Rebollo, E., Mentrup, B., Ebert, R., Franzen, J., Abagnale, G., Sieben, T., ... Wagner, W. (2017, July). Human Platelet Lysate versus Fetal Calf Serum: These Supplements Do Not Select for Different Mesenchymal Stromal Cells. *Scientific Reports*, 7, 5132. Retrieved 2023-05-02, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5506010/doi: 10.1038/s41598-017-05207-1
- Fernando, R. C., Mazzotti, D. R., Azevedo, H., Sandes, A. F., Rizzatti, E. G., de Oliveira, M. B., ... Colleoni, G. W. B. (2019, January). Transcriptome Analysis of Mesenchymal Stem Cells from Multiple Myeloma Patients Reveals Downregulation of Genes Involved in Cell Cycle Progression, Immune Response, and Bone Metabolism. Scientific Reports, 9. Retrieved 2021-01-29, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6355867/doi: 10.1038/s41598-018-38314-8
- Flier, J. S. (2022). The Problem of Irreproducible Bioscience Research. *Perspectives in Biology and Medicine*, 65(3), 373–395. doi: 10.1353/pbm.2022.0032
- Forster, S., & Radpour, R. (2022, July). Molecular Impact of the Tumor Microenvironment on Multiple Myeloma Dissemination and Extramedullary Disease. Frontiers in Oncology, 12. Retrieved 2024-05-23, from https://www.frontiersin.org/journals/oncology/articles/10.3389/fonc.2022.941437/full doi: 10.3389/fonc.2022.941437
- Frassanito, M. A., Cusmai, A., Iodice, G., & Dammacco, F. (2001, January). Autocrine interleukin-6 production and highly malignant multiple myeloma: Relation with resistance to drug-induced apoptosis. *Blood*, 97(2), 483–489. doi: 10.1182/blood.v97.2.483
- Friedenstein, A., & Kuralesova, A. I. (1971, August). Osteogenic precursor cells of bone marrow in radiation chimeras. Transplantation, 12(2), 99–108.
- Friedenstein, A. J., Piatetzky-Shapiro, I. I., & Petrakova, K. V. (1966, December). Osteogenesis in transplants of bone marrow cells. *Journal of embryology and experimental morphology*, 16(3), 381–390.
- Gabr, M. M., Zakaria, M. M., Refaie, A. F., Ismail, A. M., Abou-El-Mahasen, M. A., Ashamallah, S. A., ... Ghoneim, M. A. (2013). Insulin-producing cells from adult human bone marrow mesenchymal stem cells control streptozotocin-induced diabetes in nude mice. *Cell transplantation*, 22(1), 133–145. doi: 10.3727/096368912X647162
- Galbraith, C. G. (2023, January). Pumping up the volume. *Journal of Cell Biology*, 222(2), e202212042. Retrieved 2024-06-14, from https://doi.org/10.1083/jcb.202212042 doi: 10.1083/jcb.202212042
- Galderisi, U., Özcan, S., Alessio, N., Acar, M. B., Toprak, G., Onal, Z. B., & Peluso, G. (2015, October). Myeloma cells can corrupt senescent mesenchymal stromal cells and impair their anti-tumor activity. *Oncotarget*, 6(37), 39482–39492. Retrieved 2024-06-10, from https://www.oncotarget.com/article/5430/text/ doi: 10.18632/oncotarget .5430
- Gao, D., Ji, L., Bai, Z., Ouyang, M., Li, P., Mao, D., ... Shou, M. Z. (2024, January). ASSISTGUI: Task-Oriented Desktop Graphical User Interface Automation (No. arXiv:2312.13108). arXiv. Retrieved 2024-05-16, from http://arxiv.org/abs/2312.13108 doi: 10.48550/arXiv.2312.13108
- Gao, S., Wang, Y.-T., Ma, G.-Y., Lu, M.-Q., Chu, B., Shi, L., ... Bao, L. (2024, April). Solitary bone plasmacytoma: Long-term clinical outcomes in a single center. *Current Problems in Cancer*, 50, 101095. doi: 10.1016/j.currproblcancer.2024.101095
- Garcés, J.-J., Simicek, M., Vicari, M., Brozova, L., Burgos, L., Bezdekova, R., ... Paiva, B. (2020, February). Transcrip-

- tional profiling of circulating tumor cells in multiple myeloma: A new model to understand disease dissemination. Leukemia, 34(2), 589-603. doi: 10.1038/s41375-019-0588-4
- García-Ortiz, A., Rodríguez-García, Y., Encinas, J., Maroto-Martín, E., Castellano, E., Teixidó, J., & Martínez-López, J. (2021, January). The Role of Tumor Microenvironment in Multiple Myeloma Development and Progression. Cancers, 13(2). Retrieved 2021-02-02, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7827690/ doi: 10.3390/cancers13020217
- Gaur, T., Lengner, C. J., Hovhannisyan, H., Bhat, R. A., Bodine, P. V. N., Komm, B. S., ... Lian, J. B. (2005, September). Canonical WNT signaling promotes osteogenesis by directly stimulating Runx2 gene expression. *The Journal of Biological Chemistry*, 280(39), 33132–33140. doi: 10.1074/jbc.M500608200
- Geng, Y., Chandrasekaran, S., Agastin, S., Li, J., & King, M. R. (2014, March). Dynamic Switch Between Two Adhesion Phenotypes in Colorectal Cancer Cells. *Cellular and Molecular Bioengineering*, 7(1), 35–44. Retrieved 2024-06-27, from https://doi.org/10.1007/s12195-013-0313-8 doi: 10.1007/s12195-013-0313-8
- Gentleman. (n.d.). *Bioconductor Bioc Views*. Retrieved 2023-06-09, from https://bioconductor.org/packages/3.17/BiocViews.html
- Ghobrial, I. M. (2012, July). Myeloma as a model for the process of metastasis: Implications for therapy. *Blood*, 120(1), 20–30. Retrieved 2022-10-15, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3390959/ doi: 10.1182/blood-2012-01-379024
- Giorgi, F. M., Ceraolo, C., & Mercatelli, D. (2022, April). The R Language: An Engine for Bioinformatics and Data Science. *Life*, 12(5), 648. Retrieved 2024-04-21, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9148156/doi: 10.3390/life12050648
- Glavey, S. V., Naba, A., Manier, S., Clauser, K., Tahri, S., Park, J., ... Ghobrial, I. M. (2017, November). Proteomic characterization of human multiple myeloma bone marrow extracellular matrix. *Leukemia*, 31(11), 2426–2434. Retrieved 2023-09-05, from https://www.nature.com/articles/leu2017102 doi: 10.1038/leu.2017.102
- Gomez-Cabrero, D., Abugessaisa, I., Maier, D., Teschendorff, A., Merkenschlager, M., Gisel, A., ... Tegnér, J. (2014, March). Data integration in the era of omics: Current and future challenges. *BMC Systems Biology*, 8(2), I1. Retrieved 2024-03-18, from https://doi.org/10.1186/1752-0509-8-S2-I1 doi: 10.1186/1752-0509-8-S2-I1
- Gómez-López, G., Dopazo, J., Cigudosa, J. C., Valencia, A., & Al-Shahrour, F. (2019, May). Precision medicine needs pioneering clinical bioinformaticians. *Briefings in Bioinformatics*, 20(3), 752–766. doi: 10.1093/bib/bbx144
- Goodman, S. N., Fanelli, D., & Ioannidis, J. P. A. (2016, June). What does research reproducibility mean? *Science Translational Medicine*, 8(341), 341ps12-341ps12. Retrieved 2024-03-18, from https://www.science.org/doi/10.1126/scitranslmed.aaf5027 doi: 10.1126/scitranslmed.aaf5027
- Gorelick, M., & Ozsvald, I. (2020). High Performance Python: Practical Performant Programming for Humans. "O'Reilly Media, Inc.".
- Gosselin, R.-D. (2021, February). Insufficient transparency of statistical reporting in preclinical research: A scoping review. *Scientific Reports*, 11(1), 3335. Retrieved 2024-03-11, from https://www.nature.com/articles/s41598-021-83006-5 doi: 10.1038/s41598-021-83006-5
- Gramatzki, M., Burger, R., Trautman, U., Marschalek, R., Lorenz, H., Hansen-Hagge, T., ... Kalden, J. (1994). Two new interleukin-6 dependent plasma cell lines carrying a chromosomal abnormality involving the IL-6 gene locus., 84 Suppl. 1, 173a-173a. Retrieved 2023-03-24, from https://www.cellosaurus.org/cellopub/CLPUB00060
- Granata, V., Crisafulli, L., Nastasi, C., Ficara, F., & Sobacchi, C. (2022, May). Bone Marrow Niches and Tumour Cells: Lights and Shadows of a Mutual Relationship. Frontiers in Immunology, 13. Retrieved 2024-06-28, from https://www.frontiersin.org/journals/immunology/articles/10.3389/fimmu.2022.884024/full doi: 10.3389/fimmu.2022.884024
- $\label{lem:complex} \textit{GraphPad Prism 10 User Guide.} \ \ (2024). \ \ \text{Retrieved 2024-05-14, from https://www.graphpad.com/guides/prism/latest/user-guide/multiple-variable-tables.htm}$

- Greenstein, S., Krett, N. L., Kurosawa, Y., Ma, C., Chauhan, D., Hideshima, T., ... Rosen, S. T. (2003, April). Characterization of the MM.1 human multiple myeloma (MM) cell lines: A model system to elucidate the characteristics, behavior, and signaling of steroid-sensitive and -resistant MM cells. *Experimental Hematology*, 31(4), 271–282. doi: 10.1016/s0301-472x(03)00023-7
- Gronthos, S., Graves, S. E., Ohta, S., & Simmons, P. J. (1994, December). The STRO-1+ fraction of adult human bone marrow contains the osteogenic precursors. *Blood*, 84(12), 4164–4173.
- Guan, X. L., Chang, D. P. S., Mok, Z. X., & Lee, B. (2023, November). Assessing variations in manual pipetting: An under-investigated requirement of good laboratory practice. *Journal of Mass Spectrometry and Advances in the Clinical Lab*, 30, 25–29. doi: 10.1016/j.jmsacl.2023.09.001
- Gupta, A., Harrison, P. J., Wieslander, H., Pielawski, N., Kartasalo, K., Partel, G., ... Wählby, C. (2019). Deep Learning in Image Cytometry: A Review. *Cytometry Part A*, 95(4), 366–380. Retrieved 2022-04-08, from https://onlinelibrary.wiley.com/doi/abs/10.1002/cyto.a.23701 doi: 10.1002/cyto.a.23701
- Gupta, D., Attal, K., & Demner-Fushman, D. (2023, March). A dataset for medical instructional video classification and question answering. *Scientific Data*, 10(1), 158. doi: 10.1038/s41597-023-02036-y
- Gupta, S., Master, S., & Graham, C. (2022, October). Extramedullary Multiple Myeloma: A Patient-Focused Review of the Pathogenesis of Bone Marrow Escape. World Journal of Oncology, 13(5), 311–319. Retrieved 2024-06-28, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9635794/ doi: 10.14740/wjon1521
- Hallek, M., Bergsagel, P. L., & Anderson, K. C. (1998, January). Multiple Myeloma: Increasing Evidence for a Multistep Transformation Process. Blood, 91(1), 3-21. Retrieved 2024-06-20, from https://www.ncbi.nlm.nih.gov/ pmc/articles/PMC3901996/
- Hannun, A., Digani, J., Katharopoulos, A., & Collobert, R. (2023). MLX: Efficient and flexible machine learning on Apple silicon. Retrieved from https://github.com/ml-explore
- Harada, T., Hiasa, M., Teramachi, J., & Abe, M. (2021, September). Myeloma-Bone Interaction: A Vicious Cycle via TAK1-PIM2 Signaling. *Cancers*, 13(17). Retrieved 2024-06-05, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8431187/ doi: 10.3390/cancers13174441
- Harrington, D. P., & Fleming, T. R. (1982). A Class of Rank Test Procedures for Censored Survival Data. *Biometrika*, 69(3), 553–566. Retrieved 2023-08-07, from https://www.jstor.org/stable/2335991 doi: 10.2307/2335991
- Harris, C. R., Millman, K. J., van der Walt, S. J., Gommers, R., Virtanen, P., Cournapeau, D., ... Oliphant, T. E. (2020, September). Array programming with NumPy. *Nature*, 585(7825), 357–362. Retrieved 2023-08-09, from https://www.nature.com/articles/s41586-020-2649-2 doi: 10.1038/s41586-020-2649-2
- Heinemann, L., Möllers, K. M., Ahmed, H. M. M., Wei, L., Sun, K., Nimmagadda, S. C., ... Khandanpour, C. (2022, June). Inhibiting PI3K-AKT-mTOR Signaling in Multiple Myeloma-Associated Mesenchymal Stem Cells Impedes the Proliferation of Multiple Myeloma Cells. *Frontiers in Oncology*, 12, 874325. Retrieved 2024-06-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9251191/ doi: 10.3389/fonc.2022.874325
- Hideshima, T., Mitsiades, C., Tonon, G., Richardson, P. G., & Anderson, K. C. (2007, August). Understanding multiple myeloma pathogenesis in the bone marrow to identify new therapeutic targets. *Nature Reviews Cancer*, 7(8), 585–598. Retrieved 2023-02-07, from https://www.nature.com/articles/nrc2189 doi: 10.1038/nrc2189
- Hoang, P. H., Cornish, A. J., Dobbins, S. E., Kaiser, M., & Houlston, R. S. (2019, August). Mutational processes contributing to the development of multiple myeloma. *Blood Cancer Journal*, 9(8), 1–11. Retrieved 2024-06-23, from https://www.nature.com/articles/s41408-019-0221-9 doi: 10.1038/s41408-019-0221-9
- Hose, D., Rème, T., Hielscher, T., Moreaux, J., Messner, T., Seckinger, A., ... Goldschmidt, H. (2011, January). Proliferation is a central independent prognostic factor and target for personalized and risk-adapted treatment in multiple myeloma. *Haematologica*, 96(1), 87–95. doi: 10.3324/haematol.2010.030296
- Hothorn, T., & Lausen, B. (n.d.). *Maximally Selected Rank Statistics in R.* Retrieved from http://cran.r-project.org/web/packages/maxstat/index.html.

- Howe, A., & Chain, P. S. G. (2015). Challenges and opportunities in understanding microbial communities with metagenome assembly (accompanied by IPython Notebook tutorial). Frontiers in Microbiology, 6, 678. doi: 10.3389/fmicb.2015.00678
- Hu, Q., Wang, M., Wang, J., Tao, Y., & Niu, T. (2024). Development of a cell adhesion-based prognostic model for multiple myeloma: Insights into chemotherapy response and potential reversal of adhesion effects. Oncology Research, 32(4), 753-768. Retrieved 2024-06-23, from https://www.techscience.com/or/v32n4/55760 doi: 10 .32604/or.2023.043647
- Hu, X., Villodre, E. S., Larson, R., Rahal, O. M., Wang, X., Gong, Y., ... Debeb, B. G. (2021, January). Decorinmediated suppression of tumorigenesis, invasion, and metastasis in inflammatory breast cancer. *Communications Biology*, 4(1), 72. doi: 10.1038/s42003-020-01590-0
- Huang, S.-Y., Lin, H.-H., Yao, M., Tang, J.-L., Wu, S.-J., Hou, H.-A., ... Tien, H.-F. (2015). Higher Decorin Levels in Bone Marrow Plasma Are Associated with Superior Treatment Response to Novel Agent-Based Induction in Patients with Newly Diagnosed Myeloma A Retrospective Study. PloS One, 10(9), e0137552. doi: 10.1371/journal.pone.0137552
- Humphries, M. J. (2009). Cell adhesion assays. Methods in Molecular Biology (Clifton, N.J.), 522, 203-210. doi: 10.1007/978-1-59745-413-1 14
- Hunter, J. D. (2007, May). Matplotlib: A 2D Graphics Environment. Computing in Science & Engineering, 9(3), 90–95. Retrieved 2023-11-15, from https://ieeexplore.ieee.org/document/4160265 doi: 10.1109/MCSE.2007.55
- Huth, J., Buchholz, M., Kraus, J. M., Schmucker, M., von Wichert, G., Krndija, D., ... Kestler, H. A. (2010, April). Significantly improved precision of cell migration analysis in time-lapse video microscopy through use of a fully automated tracking system. *BMC Cell Biology*, 11(1), 24. Retrieved 2024-06-12, from https://doi.org/10.1186/1471-2121-11-24 doi: 10.1186/1471-2121-11-24
- Ibraheem, A., Attar-Schneider, O., Dabbah, M., Dolberg Jarchowsky, O., Tartakover Matalon, S., Lishner, M., & Drucker, L. (2019, May). BM-MSCs-derived ECM modifies multiple myeloma phenotype and drug response in a source-dependent manner. *Translational Research: The Journal of Laboratory and Clinical Medicine*, 207, 83–95. doi: 10.1016/j.trsl.2019.01.003
- Inc., P. T. (2015). Collaborative data science. Montreal, QC: Plotly Technologies Inc. Retrieved from https://plot.ly Incerti, D., Thom, H., Baio, G., & Jansen, J. P. (2019, May). R You Still Using Excel? The Advantages of Modern
  - Software Tools for Health Technology Assessment. Value in Health, 22(5), 575-579. Retrieved 2024-03-11, from https://www.sciencedirect.com/science/article/pii/S1098301519300506 doi: 10.1016/j.jval.2019.01.003
- Ioannidis, J. P. A. (2005, August). Why Most Published Research Findings Are False. PLOS Medicine, 2(8), e124. Retrieved 2024-04-22, from https://journals.plos.org/plosmedicine/article?id=10.1371/journal.pmed.0020124 doi: 10.1371/journal.pmed.0020124
- Ito, S., Sato, T., & Maeta, T. (2021, April). Role and Therapeutic Targeting of SDF-1α/CXCR4 Axis in Multiple Myeloma. Cancers, 13(8), 1793. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8069569/ doi: 10.3390/cancers13081793
- Jansen, B. J. H., Gilissen, C., Roelofs, H., Schaap-Oziemlak, A., Veltman, J. A., Raymakers, R. A. P., ... Adema, G. J. (2010, April). Functional differences between mesenchymal stem cell populations are reflected by their transcriptome. Stem cells and development, 19(4), 481–490. doi: 10.1089/scd.2009.0288
- Jung, S.-H., & Lee, J.-J. (2022, April). Update on primary plasma cell leukemia. *Blood Research*, 57(S1), 62–66. doi: 10.5045/br.2022.2022033
- Kaplan, E. L., & Meier, P. (1958, June). Nonparametric Estimation from Incomplete Observations. *Journal of the American Statistical Association*, 53(282), 457–481. Retrieved 2023-08-07, from http://www.tandfonline.com/doi/abs/10.1080/01621459.1958.10501452 doi: 10.1080/01621459.1958.10501452
- Kashef, J., & Franz, C. M. (2015, May). Quantitative methods for analyzing cell-cell adhesion in development.

- $Developmental\ Biology,\ 401(1),\ 165-174.\ \ Retrieved\ 2024-06-18,\ from\ https://www.sciencedirect.com/science/article/pii/S001216061400579X\ doi:\ 10.1016/j.ydbio.2014.11.002$
- Kastritis, E., Moulopoulos, L. A., Terpos, E., Koutoulidis, V., & Dimopoulos, M. A. (2014, December). The prognostic importance of the presence of more than one focal lesion in spine MRI of patients with asymptomatic (smoldering) multiple myeloma. *Leukemia*, 28(12), 2402–2403. Retrieved 2024-05-23, from https://www.nature.com/articles/leu2014230 doi: 10.1038/leu.2014.230
- Katz, B.-Z. (2010, June). Adhesion molecules—The lifelines of multiple myeloma cells. Seminars in Cancer Biology, 20(3), 186-195. Retrieved 2021-07-04, from https://linkinghub.elsevier.com/retrieve/pii/S1044579X10000246 doi: 10.1016/j.semcancer.2010.04.003
- Kawano, M. M., Huang, N., Tanaka, H., Ishikawa, H., Sakai, A., Tanabe, O., ... Kuramoto, A. (1991). Homotypic cell aggregations of human myeloma cells with ICAM-1 and LFA-1 molecules. *British Journal of Haematology*, 79(4), 583–588. Retrieved 2023-03-29, from https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2141.1991.tb08085.x doi: 10.1111/j.1365-2141.1991.tb08085.x
- Kazman, R., Bianco, P., Ivers, J., & Klein, J. (2020, December). *Maintainability* (Report). Carnegie Mellon University. Retrieved 2024-03-07, from https://kilthub.cmu.edu/articles/report/Maintainability/12954908/1 doi: 10.1184/R1/12954908.v1
- Keats, J. J., Chesi, M., Egan, J. B., Garbitt, V. M., Palmer, S. E., Braggio, E., ... Bergsagel, P. L. (2012, August). Clonal competition with alternating dominance in multiple myeloma. *Blood*, 120(5), 1067–1076. doi: 10.1182/blood-2012-01-405985
- Kelleher, R. (2024, January). NVIDIA CEO: 'This Year, Every Industry Will Become a Technology Industry'. Retrieved 2024-05-03, from https://blogs.nvidia.com/blog/nvidia-ceo-ai-drug-discovery-jp-morgan-healthcare-2024/
- Kelly, B. S., Kirwan, A., Quinn, M. S., Kelly, A. M., Mathur, P., Lawlor, A., & Killeen, R. P. (2023, May). The ethical matrix as a method for involving people living with disease and the wider public (PPI) in near-term artificial intelligence research. *Radiography (London, England: 1995)*, 29 Suppl 1, S103-S111. doi: 10.1016/j.radi.2023.03.009
- Khalili, A. A., & Ahmad, M. R. (2015, August). A Review of Cell Adhesion Studies for Biomedical and Biological Applications. *International Journal of Molecular Sciences*, 16(8), 18149. Retrieved 2024-06-18, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4581240/ doi: 10.3390/ijms160818149
- Kibler, C., Schermutzki, F., Waller, H. D., Timpl, R., Müller, C. A., & Klein, G. (1998, June). Adhesive interactions of human multiple myeloma cell lines with different extracellular matrix molecules. *Cell Adhesion and Communication*, 5(4), 307–323. doi: 10.3109/15419069809040300
- Kim, D., Langmead, B., & Salzberg, S. L. (2015, April). HISAT: A fast spliced aligner with low memory requirements. *Nature methods*, 12(4), 357–360. Retrieved 2024-04-26, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4655817/ doi: 10.1038/nmeth.3317
- Klausz, K., Cieker, M., Kellner, C., Oberg, H.-H., Kabelitz, D., Valerius, T., ... Peipp, M. (2017, September). A novel Fc-engineered human ICAM-1/CD54 antibody with potent anti-myeloma activity developed by cellular panning of phage display libraries. *Oncotarget*, 8(44), 77552–77566. doi: 10.18632/oncotarget.20641
- Kluyver, T., Ragan-Kelley, B., Pérez, F., Granger, B., Bussonnier, M., Frederic, J., ... Jupyter Development Team (2016). *Jupyter Notebooks—a publishing format for reproducible computational workflows*. Retrieved 2024-04-20, from https://ui.adsabs.harvard.edu/abs/2016ppap.book...87K doi: 10.3233/978-1-61499-649-1-87
- Krekel, H., Oliveira, B., Pfannschmidt, R., Bruynooghe, F., Laugher, B., & Bruhin, F. (2004). *Pytest.* Retrieved from https://github.com/pytest-dev/pytest
- Krzywinski, M., & Savig, E. (2013, July). Multidimensional data. *Nature methods*, 10(7), 595. Retrieved 2024-04-22, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6092027/
- Kumar, S. K., & Rajkumar, S. V. (2018, July). The multiple myelomas current concepts in cytogenetic classification

- and therapy. Nature Reviews. Clinical Oncology, 15(7), 409-421. doi: 10.1038/s41571-018-0018-y
- Kundu, S., Jha, S. B., Rivera, A. P., Flores Monar, G. V., Islam, H., Puttagunta, S. M., ... Sange, I. (2022, February). Multiple Myeloma and Renal Failure: Mechanisms, Diagnosis, and Management. *Cureus*, 14(2), e22585. Retrieved 2024-05-23, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8958144/ doi: 10.7759/cureus.22585
- Kuric, M. (2024, April). Markur4/plotastic. Retrieved 2024-05-02, from https://github.com/markur4/plotastic
- Kuric, M., Beck, S., Schneider, D., Rindt, W., Evers, M., Meißner-Weigl, J., ... Ebert, R. (2024, April). Modeling Myeloma Dissemination In Vitro with hMSC-interacting Subpopulations of INA-6 Cells and Their Aggregation/Detachment Dynamics. *Cancer Research Communications*, 4(4), 1150–1164. Retrieved 2024-05-14, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC11057410/ doi: 10.1158/2767-9764.CRC-23-0411
- Kuric, M., & Ebert, R. (2024, March). Plotastic: Bridging Plotting and Statistics in Python. *Journal of Open Source Software*, 9(95), 6304. Retrieved 2024-03-11, from https://joss.theoj.org/papers/10.21105/joss.06304 doi: 10.21105/joss.06304
- Kyle, R. A. (1997, February). Monoclonal gammopathy of undetermined significance and solitary plasmacytoma. Implications for progression to overt multiple myeloma. *Hematology/Oncology Clinics of North America*, 11(1), 71–87. doi: 10.1016/s0889-8588(05)70416-0
- Lai, T.-Y., Cao, J., Ou-Yang, P., Tsai, C.-Y., Lin, C.-W., Chen, C.-C., ... Lee, C.-Y. (2022, April). Different methods of detaching adherent cells and their effects on the cell surface expression of Fas receptor and Fas ligand. *Scientific Reports*, 12(1), 5713. Retrieved 2023-06-01, from https://www.nature.com/articles/s41598-022-09605-y doi: 10.1038/s41598-022-09605-y
- Lakhlifi, C., Lejeune, F.-X., Rouault, M., Khamassi, M., & Rohaut, B. (2023, April). Illusion of knowledge in statistics among clinicians: Evaluating the alignment between objective accuracy and subjective confidence, an online survey. Cognitive Research: Principles and Implications, 8, 23. Retrieved 2024-04-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10118231/ doi: 10.1186/s41235-023-00474-1
- Leek, J. T., & Peng, R. D. (2015, April). Statistics: P values are just the tip of the iceberg. *Nature*, 520(7549), 612-612. Retrieved 2024-04-22, from https://www.nature.com/articles/520612a doi: 10.1038/520612a
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., ... 1000 Genome Project Data Processing Subgroup (2009, August). The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, 25(16), 2078–2079. Retrieved 2023-06-09, from https://doi.org/10.1093/bioinformatics/btp352 doi: 10.1093/bioinformatics/btp352
- Liu, Z., Liu, H., He, J., Lin, P., Tong, Q., & Yang, J. (2020, May). Myeloma cells shift osteoblastogenesis to adipogenesis by inhibiting the ubiquitin ligase MURF1 in mesenchymal stem cells. *Science signaling*, 13(633), eaay8203. Retrieved 2024-06-08, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7376968/ doi: 10.1126/scisignal.aay8203
- Localio, A. R., Goodman, S. N., Meibohm, A., Cornell, J. E., Stack, C. B., Ross, E. A., & Mulrow, C. D. (2018, June). Statistical Code to Support the Scientific Story. *Annals of Internal Medicine*, 168(11), 828–829. Retrieved 2024-04-23, from https://www.acpjournals.org/doi/10.7326/M17-3431 doi: 10.7326/M17-3431
- Love, M. I., Huber, W., & Anders, S. (2014, December). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology*, 15(12), 550. Retrieved 2024-04-26, from https://doi.org/10.1186/s13059-014-0550-8 doi: 10.1186/s13059-014-0550-8
- Mai, E. K., Hielscher, T., Kloth, J. K., Merz, M., Shah, S., Raab, M. S., ... Hillengass, J. (2015, June). A magnetic resonance imaging-based prognostic scoring system to predict outcome in transplant-eligible patients with multiple myeloma. *Haematologica*, 100(6), 818–825. Retrieved 2024-05-23, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4450628/ doi: 10.3324/haematol.2015.124115
- Maichl, D. S., Kirner, J. A., Beck, S., Cheng, W.-H., Krug, M., Kuric, M., ... Jundt, F. (2023, September). Identification of NOTCH-driven matrisome-associated genes as prognostic indicators of multiple myeloma patient survival. *Blood Cancer Journal*, 13(1), 1–6. Retrieved 2023-09-05, from https://www.nature.com/articles/s41408-023-00907-6 doi: 10.1038/s41408-023-00907-6

- Majithia, N., Rajkumar, SV., Lacy, MQ., Buadi, FK., Dispenzieri, A., Gertz, MA., ... Kumar, SK. (2016, November). Early relapse following initial therapy for multiple myeloma predicts poor outcomes in the era of novel agents. *Leukemia*, 30(11), 2208–2213. Retrieved 2022-10-15, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5541860/doi: 10.1038/leu.2016.147
- Mangolini, M., & Ringshausen, I. (2020, February). Bone Marrow Stromal Cells Drive Key Hallmarks of B Cell Malignancies. *International Journal of Molecular Sciences*, 21(4), 1466. Retrieved 2023-05-02, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7073037/ doi: 10.3390/ijms21041466
- Manifesto for Agile Software Development. (2001). Retrieved 2024-05-14, from http://agilemanifesto.org/
- Martin, S. K., Diamond, P., Williams, S. A., To, L. B., Peet, D. J., Fujii, N., ... Zannettino, A. C. W. (2010, May). Hypoxia-inducible factor-2 is a novel regulator of aberrant CXCL12 expression in multiple myeloma plasma cells. *Haematologica*, 95(5), 776–784. doi: 10.3324/haematol.2009.015628
- Mateos María-Victoria, Hernández Miguel-Teodoro, Giraldo Pilar, de la Rubia Javier, de Arriba Felipe, Corral Lucía López, ... San Miguel Jesús-F. (2013). Lenalidomide plus Dexamethasone for High-Risk Smoldering Multiple Myeloma. New England Journal of Medicine, 369(5), 438–447. Retrieved 2024-05-22, from https://www.nejm.org/doi/full/10.1056/NEJMoa1300439 doi: 10.1056/NEJMoa1300439
- Mättig, P. (2022, November). Classifying exploratory experimentation three case studies of exploratory experimentation at the LHC. European Journal for Philosophy of Science, 12(4), 66. Retrieved 2024-06-14, from https://doi.org/10.1007/s13194-022-00496-4 doi: 10.1007/s13194-022-00496-4
- McCall, M. N., McMurray, H. R., Land, H., & Almudevar, A. (2014, August). On non-detects in qPCR data. Bioinformatics, 30(16), 2310–2316. Retrieved 2023-04-25, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4133581/ doi: 10.1093/bioinformatics/btu239
- McKay, B. S., Irving, P. E., Skumatz, C. M., & Burke, J. M. (1997, November). Cell-cell adhesion molecules and the development of an epithelial phenotype in cultured human retinal pigment epithelial cells. *Experimental Eye Research*, 65(5), 661–671. doi: 10.1006/exer.1997.0374
- McKinney, W. (2010, January). Data Structures for Statistical Computing in Python. In (pp. 56–61). doi: 10.25080/Majora-92bf1922-00a
- Mckinney, W. (2011, January). Pandas: A Foundational Python Library for Data Analysis and Statistics. *Python High Performance Science Computer*.
- Mesirov, J. P. (2010, January). Accessible Reproducible Research. *Science*, 327(5964), 415–416. Retrieved 2024-04-22, from https://www.science.org/doi/10.1126/science.1179653 doi: 10.1126/science.1179653
- Moleiro, A. F., Conceição, G., Leite-Moreira, A. F., & Rocha-Sousa, A. (2017). A Critical Analysis of the Available In Vitro and Ex Vivo Methods to Study Retinal Angiogenesis. *Journal of Ophthalmology*, 2017, 3034953. doi: 10.1155/2017/3034953
- Moñivas Gallego, E., & Zurita Castillo, M. (2024, April). Mesenchymal stem cell therapy in ischemic stroke trials. A systematic review. *Regenerative Therapy*, 27, 301–306. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC11021793/ doi: 10.1016/j.reth.2024.03.026
- Moran, M. (2003). Arguments for rejecting the sequential Bonferroni in ecological studies. *Oikos*, 100(2), 403-405. Retrieved 2024-04-24, from https://onlinelibrary.wiley.com/doi/abs/10.1034/j.1600-0706.2003.12010.x doi: 10.1034/j.1600-0706.2003.12010.x
- Morè, S., Corvatta, L., Manieri, V. M., Morsia, E., Poloni, A., & Offidani, M. (2023, November). Novel Immunotherapies and Combinations: The Future Landscape of Multiple Myeloma Treatment. *Pharmaceuticals*, 16(11), 1628. Retrieved 2024-05-22, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10675193/ doi: 10.3390/ph16111628
- Motulsky, H. (2018). Intuitive Biostatistics: A Nonmathematical Guide to Statistical Thinking. Oxford University Press.
- Mrozik, K. M., Cheong, C. M., Hewett, D., Chow, A. W., Blaschuk, O. W., Zannettino, A. C., & Vandyke, K. (2015).

- Therapeutic targeting of N-cadherin is an effective treatment for multiple myeloma. *British Journal of Haematology*, 171(3), 387–399. Retrieved 2024-05-26, from https://onlinelibrary.wiley.com/doi/abs/10.1111/bjh.13596 doi: 10.1111/bjh.13596
- Muruganandan, S., Roman, A. A., & Sinal, C. J. (2009, January). Adipocyte differentiation of bone marrow-derived mesenchymal stem cells: Cross talk with the osteoblastogenic program. *Cellular and molecular life sciences: CMLS*, 66(2), 236–253. doi: 10.1007/s00018-008-8429-z
- Myers, G. J., Sandler, C., & Badgett, T. (2011). *The art of software testing* (3rd ed.). Wiley Publishing. Retrieved from https://malenezi.github.io/malenezi/SE401/Books/114-the-art-of-software-testing-3-edition.pdf
- Nair, R. R., Gebhard, A. W., Emmons, M. F., & Hazlehurst, L. A. (2012, January). Chapter Six Emerging Strategies for Targeting Cell Adhesion in Multiple Myeloma. In K. S. M. Smalley (Ed.), *Advances in Pharmacology* (Vol. 65, pp. 143–189). Academic Press. Retrieved 2024-06-23, from https://www.sciencedirect.com/science/article/pii/B9780123979278000063 doi: 10.1016/B978-0-12-397927-8.00006-3
- Nalbant, P., & Dehmelt, L. (2018, August). Exploratory cell dynamics: A sense of touch for cells? *Biological Chemistry*, 399(8), 809-819. Retrieved 2024-06-12, from https://www.degruyter.com/document/doi/10.1515/hsz-2017-0341/html doi: 10.1515/hsz-2017-0341
- Narzt, W., Pichler, J., Pirklbauer, K., & Zwinz, M. (1998, January). A Reusability Concept for Process Automation Software..
- Nature Video Content. (n.d.). Retrieved 2024-06-13, from https://support.nature.com/en/support/solutions/articles/6000210836-requirements-to-play-video-content-on-the-site
- Neri, P., & J. Bahlis, N. (2012, August). Targeting of Adhesion Molecules as a Therapeutic Strategy in Multiple Myeloma. *Current Cancer Drug Targets*, 12(7), 776-796. Retrieved 2021-07-25, from http://www.eurekaselect.com/openurl/content.php?genre=article&issn=1568-0096&volume=12&issue=7&spage=776 doi: 10.2174/156800912802429337
- Newville, M., Stensitzki, T., Allen, D. B., & Ingargiola, A. (2014, September). LMFIT: Non-Linear Least-Square Minimization and Curve-Fitting for Python. Zenodo. Retrieved 2023-05-30, from https://zenodo.org/record/11813 doi: 10.5281/zenodo.11813
- Niehorster, D. C. (2021, December). Optic Flow: A History. *i-Perception*, 12(6), 20416695211055766. Retrieved 2024-06-14, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8652193/ doi: 10.1177/20416695211055766
- Nilsson, K., Bennich, H., Johansson, S. G., & Pontén, J. (1970, October). Established immunoglobulin producing myeloma (IgE) and lymphoblastoid (IgG) cell lines from an IgE myeloma patient. Clinical and Experimental Immunology, 7(4), 477–489.
- Nitta, C. F., Pierce, M., Elia, J., Ruiz, J., Hipol, A.-D., Fong, N., ... Chan, L. L.-Y. (2023, October). A rapid and high-throughput T cell immunophenotyping assay for cellular therapy bioprocess using the Cellaca® PLX image cytometer. *Journal of Immunological Methods*, 521, 113538. doi: 10.1016/j.jim.2023.113538
- Nowotschin, S., & Hadjantonakis, A.-K. (2010, August). Cellular dynamics in the early mouse embryo: From axis formation to gastrulation. *Current opinion in genetics & development*, 20(4), 420–427. doi: 10.1016/j.gde.2010.05 .008
- Oba, Y., Lee, J. W., Ehrlich, L. A., Chung, H. Y., Jelinek, D. F., Callander, N. S., ... Roodman, G. D. (2005, March). MIP-1alpha utilizes both CCR1 and CCR5 to induce osteoclast formation and increase adhesion of myeloma cells to marrow stromal cells. *Experimental Hematology*, 33(3), 272–278. doi: 10.1016/j.exphem.2004.11.015
- Ocias, L. F., Larsen, T. S., Vestergaard, H., Friis, L. S., Abildgaard, N., Frederiksen, H., & Academy of Geriatric Cancer Research (AgeCare). (2016). Trends in hematological cancer in the elderly in Denmark, 1980-2012. *Acta Oncologica (Stockholm, Sweden)*, 55 Suppl 1, 98–107. doi: 10.3109/0284186X.2015.1115124
- O'Connor, B. P., Raman, V. S., Erickson, L. D., Cook, W. J., Weaver, L. K., Ahonen, C., ... Noelle, R. J. (2004, January). BCMA Is Essential for the Survival of Long-lived Bone Marrow Plasma Cells. *The Journal of Experimental*

- Medicine, 199(1), 91-98. Retrieved 2024-05-26, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1887725/doi: 10.1084/jem.20031330
- Okuno, Y., Takahashi, T., Suzuki, A., Ichiba, S., Nakamura, K., Okada, T., ... Imura, H. (1991, February). In vitro growth pattern of myeloma cells in liquid suspension or semi-solid culture containing interleukin-6. *International Journal of Hematology*, 54(1), 41–47.
- Ordak, M. (2023, September). ChatGPT's Skills in Statistical Analysis Using the Example of Allergology: Do We Have Reason for Concern? *Healthcare (Basel, Switzerland)*, 11(18), 2554. doi: 10.3390/healthcare11182554
- Paiva, B., Paino, T., Sayagues, J.-M., Garayoa, M., San-Segundo, L., Martín, M., ... San Miguel, J. F. (2013, November). Detailed characterization of multiple myeloma circulating tumor cells shows unique phenotypic, cytogenetic, functional, and circadian distribution profile. *Blood*, 122(22), 3591–3598. doi: 10.1182/blood-2013-06-510453
- Paiva, B., Pérez-Andrés, M., Vídriales, M.-B., Almeida, J., de las Heras, N., Mateos, M.-V., ... Myeloma Stem Cell Network (MSCNET) (2011, April). Competition between clonal plasma cells and normal cells for potentially overlapping bone marrow niches is associated with a progressively altered cellular distribution in MGUS vs myeloma. Leukemia, 25(4), 697–706. doi: 10.1038/leu.2010.320
- Paszke, A., Gross, S., Massa, F., Lerer, A., Bradbury, J., Chanan, G., ... Chintala, S. (2019, December). PyTorch: An Imperative Style, High-Performance Deep Learning Library (No. arXiv:1912.01703). arXiv. Retrieved 2024-03-07, from http://arxiv.org/abs/1912.01703 doi: 10.48550/arXiv.1912.01703
- Pattarone, G., Acion, L., Simian, M., Mertelsmann, R., Follo, M., & Iarussi, E. (2021, May). Learning deep features for dead and living breast cancer cell classification without staining. *Scientific Reports*, 11, 10304. Retrieved 2024-06-16, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8119670/ doi: 10.1038/s41598-021-89895-w
- Peng, R. D. (2011, December). Reproducible Research in Computational Science. Science, 334(6060), 1226–1227. Retrieved 2024-03-18, from https://www.science.org/doi/10.1126/science.1213847 doi: 10.1126/science.1213847
- Peras, I., Klemenčič Mirazchiyski, E., Japelj Pavešić, B., & Mekiš Recek, Ž. (2023, September). Digital versus Paper Reading: A Systematic Literature Review on Contemporary Gaps According to Gender, Socioeconomic Status, and Rurality. European Journal of Investigation in Health, Psychology and Education, 13(10), 1986–2005. Retrieved 2024-06-13, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10606230/ doi: 10.3390/ejihpe13100142
- Perez, F., & Granger, B. E. (2007, May). IPython: A System for Interactive Scientific Computing. *Computing in Science & Engineering*, 9(3), 21–29. Retrieved 2024-04-20, from https://ieeexplore.ieee.org/document/4160251 doi: 10.1109/MCSE.2007.53
- Pérez-Andrés, M., Almeida, J., Martín-Ayuso, M., Moro, M. J., Martín-Nuñez, G., Galende, J., ... Spanish Network of Cancer Research Centers (C03/10) (2005, March). Clonal plasma cells from monoclonal gammopathy of undetermined significance, multiple myeloma and plasma cell leukemia show different expression profiles of molecules involved in the interaction with the immunological bone marrow microenvironment. *Leukemia*, 19(3), 449–455. doi: 10.1038/sj.leu.2403647
- Perneger, T. V. (1998, April). What's wrong with Bonferroni adjustments. *BMJ : British Medical Journal*, 316(7139), 1236–1238. Retrieved 2021-11-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1112991/
- Pfaffl, M. W. (2001, May). A new mathematical model for relative quantification in real-time RT–PCR. *Nucleic Acids Research*, 29(9), e45. Retrieved 2024-05-16, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC55695/
- Pittenger, M. F., Mackay, A. M., Beck, S. C., Jaiswal, R. K., Douglas, R., Mosca, J. D., ... Marshak, D. R. (1999). Multilineage Potential of Adult Human Mesenchymal Stem Cells., 284(April), 143–148. doi: 10.1126/science.284.5411.143
- Podar, K., & Leleu, X. (2021, October). Relapsed/Refractory Multiple Myeloma in 2020/2021 and Beyond. *Cancers*, 13(20), 5154. Retrieved 2024-05-22, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8534171/ doi: 10.3390/cancers13205154
- Polager, S., & Ginsberg, D. (2009, October). P53 and E2f: Partners in life and death. Nature Reviews Cancer, 9(10),

- 738-748. Retrieved 2023-02-14, from https://www.nature.com/articles/nrc2718 doi: 10.1038/nrc2718
- Potere, N., Buono, M. G. D., Niccoli, G., Crea, F., Toldo, S., & Abbate, A. (2019, February). Developing LRP1 Agonists into a Therapeutic Strategy in Acute Myocardial Infarction. *International Journal of Molecular Sciences*, 20(3). Retrieved 2024-06-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6387161/ doi: 10.3390/ijms20030544
- Price, A., Schroter, S., Clarke, M., & McAneney, H. (2018, September). Role of supplementary material in biomedical journal articles: Surveys of authors, reviewers and readers. *BMJ Open*, 8(9), e021753. Retrieved 2024-06-13, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6157527/ doi: 10.1136/bmjopen-2018-021753
- Purschke, M., Rubio, N., Held, K. D., & Redmond, R. W. (2010, November). Phototoxicity of Hoechst 33342 in time-lapse fluorescence microscopy. *Photochemical & Photobiological Sciences*, 9(12), 1634–1639. Retrieved 2022-03-03, from https://pubs.rsc.org/en/content/articlelanding/2010/pp/c0pp00234h doi: 10.1039/C0PP00234H
- Pushparaj, P. N. (2020). Revisiting the Micropipetting Techniques in Biomedical Sciences: A Fundamental Prerequisite in Good Laboratory Practice. *Bioinformation*, 16(1), 8. Retrieved 2024-06-18, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6986936/ doi: 10.6026/97320630016008
- Pylvänäinen, J. W., Gómez-de-Mariscal, E., Henriques, R., & Jacquemet, G. (2023, December). Live-cell imaging in the deep learning era. *Current Opinion in Cell Biology*, 85, 102271. Retrieved 2024-06-14, from https://www.sciencedirect.com/science/article/pii/S0955067423001205 doi: 10.1016/j.ceb.2023.102271
- PyMOL. (2024). Retrieved 2024-04-30, from https://pymol.org/
- The Python Language Reference. (2024). Retrieved 2024-03-07, from https://docs.python.org/3/reference/index.html
- Qamar, S., Öberg, R., Malyshev, D., & Andersson, M. (2023, October). A hybrid CNN-Random Forest algorithm for bacterial spore segmentation and classification in TEM images. *Scientific Reports*, 13(1), 18758. Retrieved 2024-06-14, from https://www.nature.com/articles/s41598-023-44212-5 doi: 10.1038/s41598-023-44212-5
- Qasim, W., Zhan, H., Samarasinghe, S., Adams, S., Amrolia, P., Stafford, S., ... Veys, P. (2017, January). Molecular remission of infant B-ALL after infusion of universal TALEN gene-edited CAR T cells. Science Translational Medicine, 9(374), eaaj2013. doi: 10.1126/scitranslmed.aaj2013
- Qian, X., Yang, Y., Deng, Y., Liu, Y., Zhou, Y., Han, F., ... Yuan, H. (2023, April). SETDB1 induces lenalidomide resistance in multiple myeloma cells via epithelial-mesenchymal transition and PI3K/AKT pathway activation. Experimental and Therapeutic Medicine, 25(6), 274. Retrieved 2024-06-28, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10189757/ doi: 10.3892/etm.2023.11973
- Qiang, Y.-W., Barlogie, B., Rudikoff, S., & Shaughnessy, J. D. (2008, April). Dkk1-induced inhibition of Wnt signaling in osteoblast differentiation is an underlying mechanism of bone loss in multiple myeloma. *Bone*, 42(4), 669–680. doi: 10.1016/j.bone.2007.12.006
- Quanbeck, A., Hennessy, R. G., & Park, L. (2022, November). Applying concepts from "rapid" and "agile" implementation to advance implementation research. *Implementation Science Communications*, 3(1), 118. doi: 10.1186/s43058-022-00366-3
- Qureshi, R., Shaughnessy, D., Gill, K. A. R., Robinson, K. A., Li, T., & Agai, E. (2023, April). Are ChatGPT and large language models "the answer" to bringing us closer to systematic review automation? *Systematic Reviews*, 12, 72. Retrieved 2024-05-03, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10148473/ doi: 10.1186/s13643-023-02243-z
- R Core Team. (2018). R: A language and environment for statistical computing [Manual]. Vienna, Austria. Retrieved from https://www.R-project.org/
- Radford, A., Wu, J., Child, R., Luan, D., Amodei, D., & Sutskever, I. (2019). Language Models are Unsupervised Multitask Learners. Retrieved 2024-03-07, from https://www.semanticscholar.org/paper/Language-Models-are -Unsupervised-Multitask-Learners-Radford-Wu/9405cc0d6169988371b2755e573cc28650d14dfe

- Rajkumar, S. V., Dimopoulos, M. A., Palumbo, A., Blade, J., Merlini, G., Mateos, M.-V., ... Miguel, J. F. S. (2014, November). International Myeloma Working Group updated criteria for the diagnosis of multiple myeloma. *The Lancet. Oncology*, 15(12), e538-548. doi: 10.1016/S1470-2045(14)70442-5
- Rajkumar, S. V., & Kumar, S. (2020, September). Multiple myeloma current treatment algorithms. *Blood Cancer Journal*, 10(9), 94. Retrieved 2023-07-03, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7523011/ doi: 10.1038/s41408-020-00359-2
- Ramakers, C., Ruijter, J. M., Deprez, R. H., & Moorman, A. F. (2003, March). Assumption-free analysis of quantitative real-time polymerase chain reaction (PCR) data. *Neuroscience Letters*, 339(1), 62–66. Retrieved 2022-11-27, from https://linkinghub.elsevier.com/retrieve/pii/S0304394002014234 doi: 10.1016/S0304-3940(02)01423-4
- Rayhan, A., & Gross, D. (2023). The Rise of Python: A Survey of Recent Research. doi: 10.13140/RG.2.2.27388.92809 Read the Docs. (2024). Retrieved 2024-05-03, from https://docs.readthedocs.io/en/stable/index.html
- Rebl, H., Finke, B., Schroeder, K., & Nebe, J. B. (2010, October). Time-dependent metabolic activity and adhesion of human osteoblast-like cells on sensor chips with a plasma polymer nanolayer. *The International Journal of Artificial Organs*, 33(10), 738–748.
- Reyes, C. D., & García, A. J. (2003, October). A centrifugation cell adhesion assay for high-throughput screening of biomaterial surfaces. *Journal of Biomedical Materials Research Part A*, 67A(1), 328–333. Retrieved 2024-06-18, from https://onlinelibrary.wiley.com/doi/10.1002/jbm.a.10122 doi: 10.1002/jbm.a.10122
- Ribatti, D., Tamma, R., & Annese, T. (2020, June). Epithelial-Mesenchymal Transition in Cancer: A Historical Overview. *Translational Oncology*, 13(6), 100773. doi: 10.1016/j.tranon.2020.100773
- Richardson, G., Knudby, A., Chen, W., Sawada, M., Lovitt, J., He, L., & Naeni, L. Y. (2023, November). Dense neural network outperforms other machine learning models for scaling-up lichen cover maps in Eastern Canada. *PLOS ONE*, 18(11), e0292839. Retrieved 2024-06-14, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10659193/doi: 10.1371/journal.pone.0292839
- Rigsby, R. E., & Parker, A. B. (2016, September). Using the PyMOL application to reinforce visual understanding of protein structure. *Biochemistry and Molecular Biology Education: A Bimonthly Publication of the International Union of Biochemistry and Molecular Biology*, 44(5), 433–437. doi: 10.1002/bmb.20966
- Robinson, M. D., McCarthy, D. J., & Smyth, G. K. (2010, January). edgeR: A Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics (Oxford, England)*, 26(1), 139–140. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/19910308 doi: 10.1093/bioinformatics/btp616
- Robitaille, M. C., Byers, J. M., Christodoulides, J. A., & Raphael, M. P. (2022, November). Self-supervised machine learning for live cell imagery segmentation. *Communications Biology*, 5(1), 1–8. Retrieved 2024-06-14, from https://www.nature.com/articles/s42003-022-04117-x doi: 10.1038/s42003-022-04117-x
- Roccaro, A. M., Mishima, Y., Sacco, A., Moschetta, M., Tai, Y.-T., Shi, J., ... Ghobrial, I. M. (2015, July). CXCR4 Regulates Extra-Medullary Myeloma through Epithelial-Mesenchymal-Transition-like Transcriptional Activation. *Cell Reports*, 12(4), 622–635. doi: 10.1016/j.celrep.2015.06.059
- Roukos, V., Pegoraro, G., Voss, T. C., & Misteli, T. (2015, February). Cell cycle staging of individual cells by fluorescence microscopy. *Nature protocols*, 10(2), 334–348. Retrieved 2024-06-16, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6318798/ doi: 10.1038/nprot.2015.016
- Roy, P., Mukherjee, T., Chatterjee, B., Vijayaragavan, B., Banoth, B., & Basak, S. (2017, March). Non-canonical NFκB mutations reinforce pro-survival TNF response in multiple myeloma through an autoregulatory RelB:p50 NFκB pathway. *Oncogene*, 36(10), 1417–1429. Retrieved 2024-06-08, from https://www.nature.com/articles/onc2016309 doi: 10.1038/onc.2016.309
- Roy, P., Sarkar, U., & Basak, S. (2018, May). The NF-κB Activating Pathways in Multiple Myeloma. *Biomedicines*, 6(2), 59. Retrieved 2021-07-25, from http://www.mdpi.com/2227-9059/6/2/59 doi: 10.3390/biomedicines6020059 Rueden, C. T., Schindelin, J., Hiner, M. C., DeZonia, B. E., Walter, A. E., Arena, E. T., & Eliceiri, K. W. (2017,

- November). ImageJ2: ImageJ for the next generation of scientific image data. BMC Bioinformatics, 18(1), 529. Retrieved 2024-04-25, from https://doi.org/10.1186/s12859-017-1934-z doi: 10.1186/s12859-017-1934-z
- Ruijter, J. M., Barnewall, R. J., Marsh, I. B., Szentirmay, A. N., Quinn, J. C., van Houdt, R., ... van den Hoff, M. J. B. (2021, June). Efficiency Correction Is Required for Accurate Quantitative PCR Analysis and Reporting. Clinical Chemistry, 67(6), 829–842. Retrieved 2023-05-27, from https://doi.org/10.1093/clinchem/hvab052 doi: 10.1093/clinchem/hvab052
- Ruiz-Villalba, A., Ruijter, J. M., & van den Hoff, M. J. B. (2021, May). Use and Misuse of Cq in qPCR Data Analysis and Reporting. *Life*, 11(6), 496. Retrieved 2023-04-25, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8229287/doi: 10.3390/life11060496
- Ruksakulpiwat, S., Kumar, A., & Ajibade, A. (2023, May). Using ChatGPT in Medical Research: Current Status and Future Directions. *Journal of Multidisciplinary Healthcare*, 16, 1513–1520. Retrieved 2024-05-03, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10239248/ doi: 10.2147/JMDH.S413470
- Sacchetti, B., Funari, A., Remoli, C., Giannicola, G., Kogler, G., Liedtke, S., ... Bianco, P. (2016). No identical "mesenchymal stem cells" at different times and sites: Human committed progenitors of distinct origin and differentiation potential are incorporated as adventitial cells in microvessels. *Stem Cell Reports*, 6(6), 897–913. Retrieved from http://dx.doi.org/10.1016/j.stemcr.2016.05.011 doi: 10.1016/j.stemcr.2016.05.011
- Sandve, G. K., Nekrutenko, A., Taylor, J., & Hovig, E. (2013, October). Ten Simple Rules for Reproducible Computational Research. *PLoS Computational Biology*, 9(10), e1003285. Retrieved 2024-03-07, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3812051/ doi: 10.1371/journal.pcbi.1003285
- Santos, B. S., Silva, I., Ribeiro-Dantas, M. d. C., Alves, G., Endo, P. T., & Lima, L. (2020, October). COVID-19: A scholarly production dataset report for research analysis. *Data in Brief*, 32, 106178. doi: 10.1016/j.dib.2020.106178
- Sanz-Rodríguez, F., Ruiz-Velasco, N., Pascual-Salcedo, D., & Teixidó, J. (1999, December). Characterization of VLA-4-dependent myeloma cell adhesion to fibronectin and VCAM-1: VLA-4-dependent Myeloma Cell Adhesion. British Journal of Haematology, 107(4), 825–834. Retrieved 2023-04-02, from http://doi.wiley.com/10.1046/j.1365-2141.1999.01762.x doi: 10.1046/j.1365-2141.1999.01762.x
- Sarin, V., Yu, K., Ferguson, I. D., Gugliemini, O., Nix, M. A., Hann, B., ... Wiita, A. P. (2020, October). Evaluating the efficacy of multiple myeloma cell lines as models for patient tumors via transcriptomic correlation analysis. *Leukemia*, 34(10), 2754–2765. doi: 10.1038/s41375-020-0785-1
- Seabold, S., & Perktold, J. (2010). Statsmodels: Econometric and Statistical Modeling with Python. In *Python in Science Conference* (pp. 92-96). Austin, Texas. Retrieved 2023-05-29, from https://conference.scipy.org/proceedings/scipy2010/seabold.html doi: 10.25080/Majora-92bf1922-011
- Seckinger, A., Delgado, J. A., Moser, S., Moreno, L., Neuber, B., Grab, A., ... Vu, M. D. (2017, March). Target Expression, Generation, Preclinical Activity, and Pharmacokinetics of the BCMA-T Cell Bispecific Antibody EM801 for Multiple Myeloma Treatment. Cancer Cell, 31(3), 396–410. Retrieved 2023-07-21, from https://www.cell.com/cancer-cell/abstract/S1535-6108(17)30016-8 doi: 10.1016/j.ccell.2017.02.002
- Seckinger, A., Hillengass, J., Emde, M., Beck, S., Kimmich, C., Dittrich, T., ... Hose, D. (2018). CD38 as Immunother-apeutic Target in Light Chain Amyloidosis and Multiple Myeloma-Association With Molecular Entities, Risk, Survival, and Mechanisms of Upfront Resistance. Frontiers in Immunology, 9, 1676. doi: 10.3389/fimmu.2018.01676
- Sharma, N., Smadbeck, J. B., Abdallah, N., Zepeda-Mendoza, C., Binder, M., Pearce, K. E., ... Baughn, L. B. (2021, October). The Prognostic Role of MYC Structural Variants Identified by NGS and FISH in Multiple Myeloma. Clinical Cancer Research, 27(19), 5430. Retrieved 2024-06-24, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8738776/doi: 10.1158/1078-0432.CCR-21-0005
- Shenghui, H., Nakada, D., & Morrison, S. J. (2009). Mechanisms of Stem Cell Self-Renewal. *Annual Review of Cell and Developmental Biology*, 25(1), 377–406. Retrieved from https://doi.org/10.1146/annurev.cellbio.042308.113248 doi: 10.1146/annurev.cellbio.042308.113248

- Sherina, V. (2020). Multiple imputation and direct estimation for qPCR data with non-detects.
- Siclari, V., Guise, T., & Chirgwin, J. (2007, January). Molecular interactions between breast cancer cells and the bone microenvironment drive skeletal metastases. *Cancer metastasis reviews*, 25, 621–33. doi: 10.1007/s10555-006-9023-1
- Siegel, R. L., Giaquinto, A. N., & Jemal, A. (2024). Cancer statistics, 2024. CA: A Cancer Journal for Clinicians, 74(1), 12-49. Retrieved 2024-05-21, from https://onlinelibrary.wiley.com/doi/abs/10.3322/caac.21820 doi: 10.3322/caac.21820
- Simonyan, K., & Zisserman, A. (2015, April). Very Deep Convolutional Networks for Large-Scale Image Recognition. arXiv:1409.1556 [cs]. Retrieved 2022-05-12, from http://arxiv.org/abs/1409.1556
- Smith, A. M., Niemeyer, K. E., Katz, D. S., Barba, L. A., Githinji, G., Gymrek, M., ... Vanderplas, J. T. (2018). Journal of Open Source Software (JOSS): Design and first-year review. *PeerJ Preprints*, 4, e147. doi: 10.7717/peerj-cs.147
- Solimando, A. G., Da Vià, M. C., Leone, P., Borrelli, P., Croci, G. A., Tabares, P., ... Beilhack, A. (2020, April). Halting the vicious cycle within the multiple myeloma ecosystem: Blocking JAM-A on bone marrow endothelial cells restores the angiogenic homeostasis and suppresses tumor progression. *Haematologica*. doi: 10.3324/haematol.2019.239913
- Solimando, A. G., Malerba, E., Leone, P., Prete, M., Terragna, C., Cavo, M., & Racanelli, V. (2022, September). Drug resistance in multiple myeloma: Soldiers and weapons in the bone marrow niche. Frontiers in Oncology, 12, 973836. Retrieved 2022-10-23, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9533079/ doi: 10.3389/fonc.2022.973836
- Spaepen, P., De Boodt, S., Aerts, J.-M., & Sloten, J. V. (2011). Digital image processing of live/dead staining. *Methods in Molecular Biology (Clifton, N.J.)*, 740, 209–230. doi: 10.1007/978-1-61779-108-6 21
- Sphinx.~(2024). Retrieved 2024-05-03, from https://docs.readthedocs.io/en/stable/intro/getting-started-with-sphinx.html
- Sprynski, A. C., Hose, D., Caillot, L., Rème, T., Shaughnessy, J. D., Barlogie, B., ... Klein, B. (2009, May). The role of IGF-1 as a major growth factor for myeloma cell lines and the prognostic relevance of the expression of its receptor. Blood, 113(19), 4614–4626. Retrieved 2023-06-29, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2691749/doi: 10.1182/blood-2008-07-170464
- Standal, T., Seidel, C., Plesner, T., Sanderson, R., Waage, A., Børset, M., & Sundan, A. (2002, November). Osteoprotegerin is bound, internalized, and degraded by multiple myeloma cells. *Blood*, 100, 3002–7. doi: 10.1182/blood-2002-04-1190
- Stock, P., Bruckner, S., Winkler, S., Dollinger, M. M., & Christ, B. (2014, April). Human bone marrow mesenchymal stem cell-derived hepatocytes improve the mouse liver after acute acetaminophen intoxication by preventing progress of injury. *International journal of molecular sciences*, 15(4), 7004–7028. doi: 10.3390/ijms15047004
- Sullivan, G. M., & Feinn, R. S. (2021, August). Facts and Fictions About Handling Multiple Comparisons. *Journal of Graduate Medical Education*, 13(4), 457–460. Retrieved 2024-03-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8370375/ doi: 10.4300/JGME-D-21-00599.1
- Tabolacci, C., De Martino, A., Mischiati, C., Feriotto, G., & Beninati, S. (2019, January). The Role of Tissue Transglutaminase in Cancer Cell Initiation, Survival and Progression. *Medical Sciences*, 7(2), 19. Retrieved 2023-03-17, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6409630/ doi: 10.3390/medsci7020019
- Tai, Y.-T., Li, X.-F., Breitkreutz, I., Song, W., Neri, P., Catley, L., ... Anderson, K. C. (2006, July). Role of B-cell-activating factor in adhesion and growth of human multiple myeloma cells in the bone marrow microenvironment. Cancer Research, 66(13), 6675–6682. doi: 10.1158/0008-5472.CAN-06-0190
- Tam, P. P., & Beddington, R. S. (1987, January). The formation of mesodermal tissues in the mouse embryo during gastrulation and early organogenesis. *Development (Cambridge, England)*, 99(1), 109–126.
- Taskiran, I. I., Spanier, K. I., Dickmänken, H., Kempynck, N., Pančíková, A., Ekşi, E. C., ... Aerts, S. (2024, February). Cell-type-directed design of synthetic enhancers. *Nature*, 626(7997), 212–220. Retrieved 2024-04-21, from https://www.nature.com/articles/s41586-023-06936-2 doi: 10.1038/s41586-023-06936-2

- Team, T. P. D. (2020, February). *Pandas-dev/pandas: Pandas.* Zenodo. Retrieved from https://doi.org/10.5281/zenodo.3509134 doi: 10.5281/zenodo.3509134
- Teoh, G., & Anderson, K. C. (1997, February). INTERACTION OF TUMOR AND HOST CELLS WITH ADHESION AND EXTRACELLULAR MATRIX MOLECULES IN THE DEVELOPMENT OF MULTIPLE MYELOMA. *Hematology/Oncology Clinics of North America*, 11(1), 27–42. Retrieved 2021-01-29, from http://www.sciencedirect.com/science/article/pii/S0889858805704135 doi: 10.1016/S0889-8588(05)70413-5
- Teramachi, J., Silbermann, R., Yang, P., Zhao, W., Mohammad, K. S., Guo, J., ... Kurihara, N. (2016, February). Blocking the ZZ Domain of Sequestosome1/p62 Suppresses Myeloma Growth and Osteoclast Formation In Vitro and Induces Dramatic Bone Formation in Myeloma-Bearing Bones In Vivo. *Leukemia*, 30(2), 390–398. Retrieved 2024-06-08, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4740189/ doi: 10.1038/leu.2015.229
- Terpos, E., Migkou, M., Christoulas, D., Gavriatopoulou, M., Eleutherakis-Papaiakovou, E., Kanellias, N., ... Dimopoulos, M. A. (2016, May). Increased circulating VCAM-1 correlates with advanced disease and poor survival in patients with multiple myeloma: Reduction by post-bortezomib and lenalidomide treatment. *Blood Cancer Journal*, 6(5), e428. Retrieved 2021-02-03, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4916305/doi: 10.1038/bcj.2016.37
- Terpos, E., Ntanasis-Stathopoulos, I., Gavriatopoulou, M., & Dimopoulos, M. A. (2018, January). Pathogenesis of bone disease in multiple myeloma: From bench to bedside. *Blood Cancer Journal*, 8(1), 7. doi: 10.1038/s41408-017-0037-4
- Thompson, S., Dowrick, T., Ahmad, M., Xiao, G., Koo, B., Bonmati, E., ... Clarkson, M. J. (2020, July). SciKit-Surgery: Compact libraries for surgical navigation. *International Journal of Computer Assisted Radiology and Surgery*, 15(7), 1075–1084. doi: 10.1007/s11548-020-02180-5
- Thumallapally, N., Meshref, A., Mousa, M., & Terjanian, T. (2017, January). Solitary plasmacytoma: Population-based analysis of survival trends and effect of various treatment modalities in the USA. *BMC Cancer*, 17, 13. Retrieved 2024-05-21, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5216567/ doi: 10.1186/s12885-016-3015-5
- Trapnell, C., Roberts, A., Goff, L., Pertea, G., Kim, D., Kelley, D. R., ... Pachter, L. (2012, March). Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. *Nature Protocols*, 7(3), 562–578. doi: 10.1038/nprot.2012.016
- Tsubaki, M., Seki, S., Takeda, T., Chihara, A., Arai, Y., Morii, Y., ... Nishida, S. (2020, October). The HGF/Met/NFκB Pathway Regulates RANKL Expression in Osteoblasts and Bone Marrow Stromal Cells. *International Journal*of Molecular Sciences, 21(21), 7905. Retrieved 2024-06-08, from https://www.ncbi.nlm.nih.gov/pmc/articles/
  PMC7663721/ doi: 10.3390/ijms21217905
- Turesson, I., Bjorkholm, M., Blimark, C. H., Kristinsson, S., Velez, R., & Landgren, O. (2018, April). Rapidly changing myeloma epidemiology in the general population: Increased incidence, older patients, and longer survival. *European journal of haematology*, 10.1111/ejh.13083. Retrieved 2024-05-22, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6195866/ doi: 10.1111/ejh.13083
- Ullah, I., Subbarao, R. B., & Rho, G. J. (2015). Human mesenchymal stem cells current trends and future prospective Bioscience Reports.

  doi: 10.1042/BSR20150025
- Ullah, T. R. (2019, August). The role of CXCR4 in multiple myeloma: Cells' journey from bone marrow to beyond. Journal of Bone Oncology, 17, 100253. doi: 10.1016/j.jbo.2019.100253
- Urashima, M., Chauhan, D., Uchiyama, H., Freeman, G., & Anderson, K. (1995, April). CD40 ligand triggered interleukin-6 secretion in multiple myeloma. Blood, 85(7), 1903-1912. Retrieved 2021-02-01, from https://ashpublications.org/blood/article/85/7/1903/123565/CD40-ligand-triggered-interleukin6-secretion-in doi: 10.1182/blood.V85.7.1903.bloodjournal8571903
- Väänänen, H. K. (1993, August). Mechanism of bone turnover. Annals of Medicine, 25(4), 353–359. doi: 10.3109/07853899309147297

- Vallat, R. (2018, November). Pingouin: Statistics in Python. Journal of Open Source Software, 3(31), 1026. Retrieved 2023-05-29, from https://joss.theoj.org/papers/10.21105/joss.01026 doi: 10.21105/joss.01026
- van Rossum, G., Lehtosalo, J., & Langa, L. (2014). PEP 484 Type Hints | peps.python.org. Retrieved 2024-03-08, from https://peps.python.org/pep-0484/
- Vande Broek, I., Vanderkerken, K., Van Camp, B., & Van Riet, I. (2008). Extravasation and homing mechanisms in multiple myeloma. Clinical & Experimental Metastasis, 25(4), 325–334. doi: 10.1007/s10585-007-9108-4
- Van Valckenborgh, E., Croucher, P. I., De Raeve, H., Carron, C., De Leenheer, E., Blacher, S., ... Vanderkerken, K. (2004, September). Multifunctional role of matrix metalloproteinases in multiple myeloma: A study in the 5T2MM mouse model. The American Journal of Pathology, 165(3), 869–878. doi: 10.1016/S0002-9440(10)63349-4
- Verzella, D., Cornice, J., Arboretto, P., Vecchiotti, D., Di Vito Nolfi, M., Capece, D., ... Franzoso, G. (2022, September). The NF-κB Pharmacopeia: Novel Strategies to Subdue an Intractable Target. *Biomedicines*, 10(9), 2233. Retrieved 2024-06-10, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9496094/ doi: 10.3390/biomedicines10092233
- Wadgaonkar, R., Phelps, K. M., Haque, Z., Williams, A. J., Silverman, E. S., & Collins, T. (1999, January). CREB-binding protein is a nuclear integrator of nuclear factor-kappaB and p53 signaling. The Journal of Biological Chemistry, 274(4), 1879–1882. doi: 10.1074/jbc.274.4.1879
- Wang, W., Yang, X., Dai, J., Lu, Y., Zhang, J., & Keller, E. T. (2019, June). Prostate cancer promotes a vicious cycle of bone metastasis progression through inducing osteocytes to secrete GDF15 that stimulates prostate cancer growth and invasion. *Oncogene*, 38(23), 4540–4559. doi: 10.1038/s41388-019-0736-3
- Waskom, M. L. (2021, April). Seaborn: Statistical data visualization. *Journal of Open Source Software*, 6(60), 3021. Retrieved 2023-03-26, from https://joss.theoj.org/papers/10.21105/joss.03021 doi: 10.21105/joss.03021
- Webster, G. A., & Perkins, N. D. (1999, May). Transcriptional Cross Talk between NF-κB and p53. *Molecular and Cellular Biology*, 19(5), 3485–3495. Retrieved 2023-07-04, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC84141/
- Weetall, M., Hugo, R., Maida, S., West, S., Wattanasin, S., Bouhel, R., ... Friedman, C. (2001, June). A Homogeneous Fluorometric Assay for Measuring Cell Adhesion to Immobilized Ligand Using V-Well Microtiter Plates. *Analytical Biochemistry*, 293(2), 277–287. Retrieved 2022-09-25, from https://linkinghub.elsevier.com/retrieve/pii/S0003269701951401 doi: 10.1006/abio.2001.5140
- Weiss, C. J. (2022, September). Visualizing protein big data using Python and Jupyter notebooks. *Biochemistry and Molecular Biology Education: A Bimonthly Publication of the International Union of Biochemistry and Molecular Biology*, 50(5), 431–436. doi: 10.1002/bmb.21621
- West, K. (2018, July). Reinventing Research: Agile in the Academic Laboratory / Agile Alliance. Retrieved 2024-05-14, from https://www.agilealliance.org/resources/experience-reports/reinventing-research-agile-in-the-academic-laboratory/
- Weyburne, D. W. (2014, April). New thickness and shape parameters for the boundary layer velocity profile. Experimental Thermal and Fluid Science, 54, 22-28. Retrieved 2024-06-18, from https://www.sciencedirect.com/science/article/pii/S089417771400017X doi: 10.1016/j.expthermflusci.2014.01.008
- Wickham, H. (2014, September). Tidy Data. Journal of Statistical Software, 59, 1–23. Retrieved 2023-11-15, from https://doi.org/10.18637/jss.v059.i10 doi: 10.18637/jss.v059.i10
- Wilkins, A., Kemp, K., Ginty, M., Hares, K., Mallam, E., & Scolding, N. (2009, July). Human bone marrow-derived mesenchymal stem cells secrete brain-derived neurotrophic factor which promotes neuronal survival in vitro. Stem cell research, 3(1), 63-70. doi: 10.1016/j.scr.2009.02.006
- Wilkinson, M. D., Dumontier, M., Aalbersberg, I. J., Appleton, G., Axton, M., Baak, A., ... Mons, B. (2016, March). The FAIR Guiding Principles for scientific data management and stewardship. *Scientific Data*, 3(1), 160018. Retrieved 2024-03-18, from https://www.nature.com/articles/sdata201618 doi: 10.1038/sdata.2016.18
- Wilmore, J. R., & Allman, D. (2017, August). Here, there, and anywhere? Arguments for and against the physical

- plasma cell survival niche. *Journal of immunology (Baltimore, Md. : 1950), 199*(3), 839–845. Retrieved 2024-06-28, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5651088/ doi: 10.4049/jimmunol.1700461
- Witwer, K. W. (2013, February). Data submission and quality in microarray-based microRNA profiling. *Clinical chemistry*, 59(2), 392–400. Retrieved 2024-04-22, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4037921/doi: 10.1373/clinchem.2012.193813
- Wohlford, G. F., Buckley, L. F., Kadariya, D., Park, T., Chiabrando, J. G., Carbone, S., ... Van Tassell, B. (2021, May). A phase 1 clinical trial of SP16, a first-in-class anti-inflammatory LRP1 agonist, in healthy volunteers. PLoS ONE, 16(5), e0247357. Retrieved 2023-03-30, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8101931/doi: 10.1371/journal.pone.0247357
- Wong, A. D., & Searson, P. C. (2017, November). Mitosis-mediated intravasation in a tissue-engineered tumor-microvessel platform. *Cancer research*, 77(22), 6453-6461. Retrieved 2023-07-14, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5690825/ doi: 10.1158/0008-5472.CAN-16-3279
- Xu, W., Zhang, X., Qian, H., Zhu, W., Sun, X., Hu, J., ... Chen, Y. (2004, July). Mesenchymal stem cells from adult human bone marrow differentiate into a cardiomyocyte phenotype in vitro. *Experimental biology and medicine* (Maywood, N.J.), 229(7), 623–631.
- Yang, A., Troup, M., & Ho, J. W. (2017, July). Scalability and Validation of Big Data Bioinformatics Software. Computational and Structural Biotechnology Journal, 15, 379–386. Retrieved 2024-03-07, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5537105/ doi: 10.1016/j.csbj.2017.07.002
- Yang, P., Qu, Y., Wang, M., Chu, B., Chen, W., Zheng, Y., ... Qian, Z. (2022, June). Pathogenesis and treatment of multiple myeloma. *MedComm*, 3(2), e146. Retrieved 2024-05-21, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9162151/ doi: 10.1002/mco2.146
- Yang, Y., Macleod, V., Bendre, M., Huang, Y., Theus, A. M., Miao, H.-Q., ... Sanderson, R. D. (2005, February). Heparanase promotes the spontaneous metastasis of myeloma cells to bone. *Blood*, 105(3), 1303–1309. doi: 10.1182/blood-2004-06-2141
- Zehentmeier, S., Roth, K., Cseresnyes, Z., Sercan, Ö., Horn, K., Niesner, R. A., ... Hauser, A. E. (2014, August). Static and dynamic components synergize to form a stable survival niche for bone marrow plasma cells. *European Journal of Immunology*, 44(8), 2306–2317. doi: 10.1002/eji.201344313
- Zeissig, M. N., Zannettino, A. C. W., & Vandyke, K. (2020, December). Tumour Dissemination in Multiple Myeloma Disease Progression and Relapse: A Potential Therapeutic Target in High-Risk Myeloma. *Cancers*, 12(12). Retrieved 2021-02-03, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7761917/ doi: 10.3390/cancers12123643
- Zeiss OAD Feature Extractors. (n.d.). Retrieved 2024-06-14, from https://github.com/zeiss-microscopy/OAD/blob/master/Machine\_Learning/Feature\_Extractors/feature\_extractors.md
- Zepeda-Moreno, A., Taubert, I., Hellwig, I., Hoang, V., Pietsch, L., Lakshmanan, V. K., ... Ho, A. D. (2011). Innovative method for quantification of cell-cell adhesion in 96-well plates. *Cell Adhesion & Migration*, 5(3), 215–219. Retrieved 2024-06-18, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3210204/ doi: 10.4161/cam.5.3.14648
- Zerbino, D. R., Achuthan, P., Akanni, W., Amode, M. R., Barrell, D., Bhai, J., ... Flicek, P. (2018, January). Ensembl 2018. Nucleic Acids Research, 46(D1), D754-D761. Retrieved 2023-05-27, from https://doi.org/10.1093/nar/gkx1098 doi: 10.1093/nar/gkx1098
- Zhou, F., Meng, S., Song, H., & Claret, F. X. (2013, November). Dickkopf-1 is a key regulator of myeloma bone disease: Opportunities and challenges for therapeutic intervention. *Blood reviews*, 27(6), 261–267. Retrieved 2023-02-18, from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4133945/ doi: 10.1016/j.blre.2013.08.002
- Zhou, Y., Zhou, B., Pache, L., Chang, M., Khodabakhshi, A. H., Tanaseichuk, O., ... Chanda, S. K. (2019, April). Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nature Communications*, 10(1), 1523. Retrieved 2023-02-09, from https://www.nature.com/articles/s41467-019-09234-6 doi: 10.1038/s41467-019-09234-6

- Ziemann, M., Eren, Y., & El-Osta, A. (2016, August). Gene name errors are widespread in the scientific literature. Genome Biology, 17(1), 177. Retrieved 2024-04-30, from https://doi.org/10.1186/s13059-016-1044-7 doi: 10.1186/s13059-016-1044-7
- Zou, Y. S., Klausner, M., Ghabrial, J., Stinnett, V., Long, P., Morsberger, L., ... Tang, G. (2024, May). A comprehensive approach to evaluate genetic abnormalities in multiple myeloma using optical genome mapping. *Blood Cancer Journal*, 14(1), 1–5. Retrieved 2024-06-23, from https://www.nature.com/articles/s41408-024-01059-x doi: 10.1038/s41408-024-01059-x

# Appendices

# A Supplementary Data & Methods

A.1 Figures

## A.2 Tables

## A.3 Materials & Methods

## B Documentation of plotastic