

Class 9: Structural Bioinformatics pt.1

Marriane Allahwerdi (A16802759)

The main database for structural data is called the PDB (protein Data Bank). Let's see what it contains:

Data From: <https://www.rcsb.org/stats/summary>

```
pdb <- read.csv("Data Export Summary.csv", row.names = 1)
head(pdb)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	167,192	15,572	12,529	208	77	32
Protein/Oligosaccharide	9,639	2,635	34	8	2	0
Protein/NA	8,730	4,697	286	7	0	0
Nucleic acid (only)	2,869	137	1,507	14	3	1
Other	170	10	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	195,610					
Protein/Oligosaccharide	12,318					
Protein/NA	13,720					
Nucleic acid (only)	4,531					
Other	213					
Oligosaccharide (only)	22					

```
pdb$Total
```

```
[1] "195,610" "12,318" "13,720" "4,531" "213" "22"
```

I need to remove the comma and convert to numeric to do math:

```
as.numeric(sub(",", "", pdb$Total))
```

```
[1] 195610 12318 13720 4531 213 22
```

```
#as.numeric(pdb$Total)
```

I could turn this into a function to fix the whole table or any future table I read like this:

```
x <- pdb$Total  
as.numeric(sub(",", "", x))
```

```
[1] 195610 12318 13720 4531 213 22
```

```
comma2numeric <- function(x) {  
  as.numeric( sub(",", "", x) )  
}
```

```
comma2numeric(pdb$X.ray)
```

```
[1] 167192 9639 8730 2869 170 11
```

```
apply(pdb, 2, comma2numeric)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other	Total
[1,]	167192	15572	12529	208	77	32	195610
[2,]	9639	2635	34	8	2	0	12318
[3,]	8730	4697	286	7	0	0	13720
[4,]	2869	137	1507	14	3	1	4531
[5,]	170	10	33	0	0	0	213
[6,]	11	0	6	1	0	4	22

```
# install.packages("readr")
```

```
library(readr)  
pdbn <- read_csv("Data Export Summary.csv")
```

Rows: 6 Columns: 8

-- Column specification -----

Delimiter: ","

chr (1): Molecular Type

dbl (3): Multiple methods, Neutron, Other

num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.

i Specify the column types or set `show_col_types = FALSE` to quiet this message.

```
pdbn$Total
```

```
[1] 195610 12318 13720 4531 213 22
```

```
sum(pdbn$Total)
```

```
[1] 226414
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

83.3% are X-Ray and 10.2% are EM so about 93% are of structures are solved by both X-Ray and EM.

```
sum(pdbn$`X-ray`)/sum(pdbn$Total) * 100
```

```
[1] 83.30359
```

```
sum(pdbn$`EM`)/sum(pdbn$Total) * 100
```

```
[1] 10.18091
```

Q2: What proportion of structures in the PDB are protein?

86.4% of the structures in the pdb are protein.

```
pdbn$Total[1]/sum(pdbn$`Total`) * 100
```

```
[1] 86.39483
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are currently 226,414 HIV-1 protease structures found in the PDB.

Mol

Mol (“molstar”)

We will use the pdb code: 1HSG



Figure 1: A first image from molstar

some more custom images:

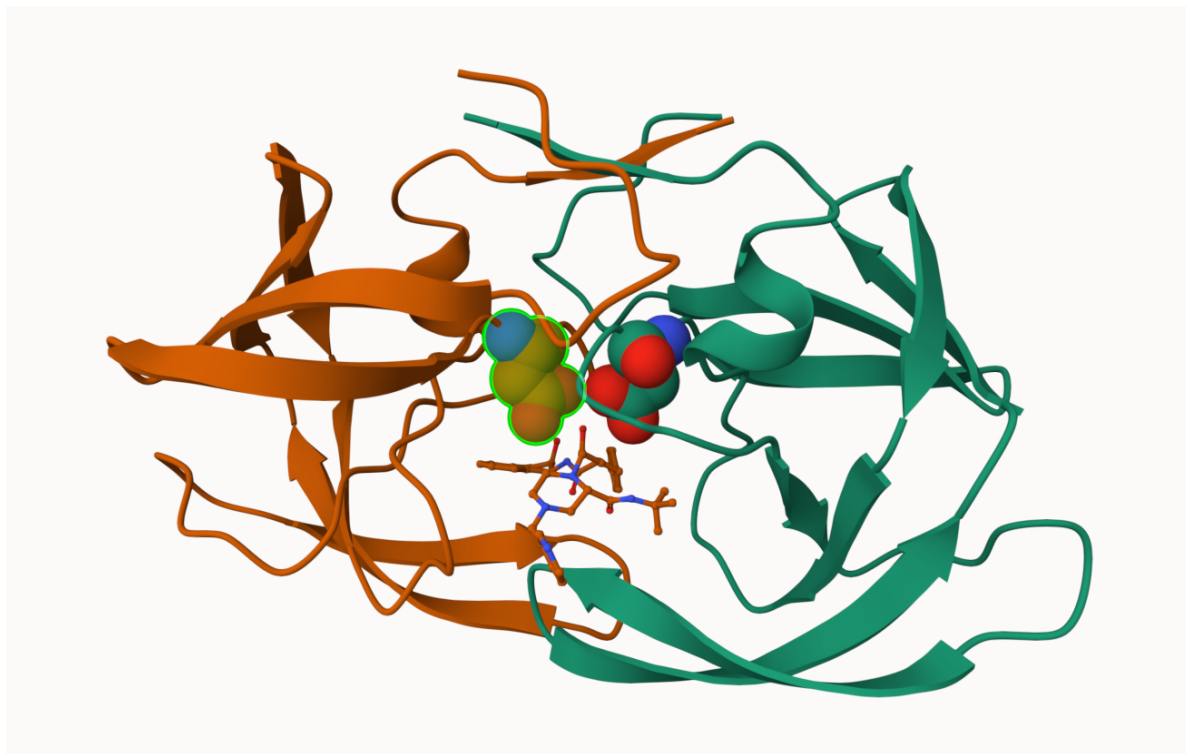


Figure 2: The catalytic ASP25 amino acids

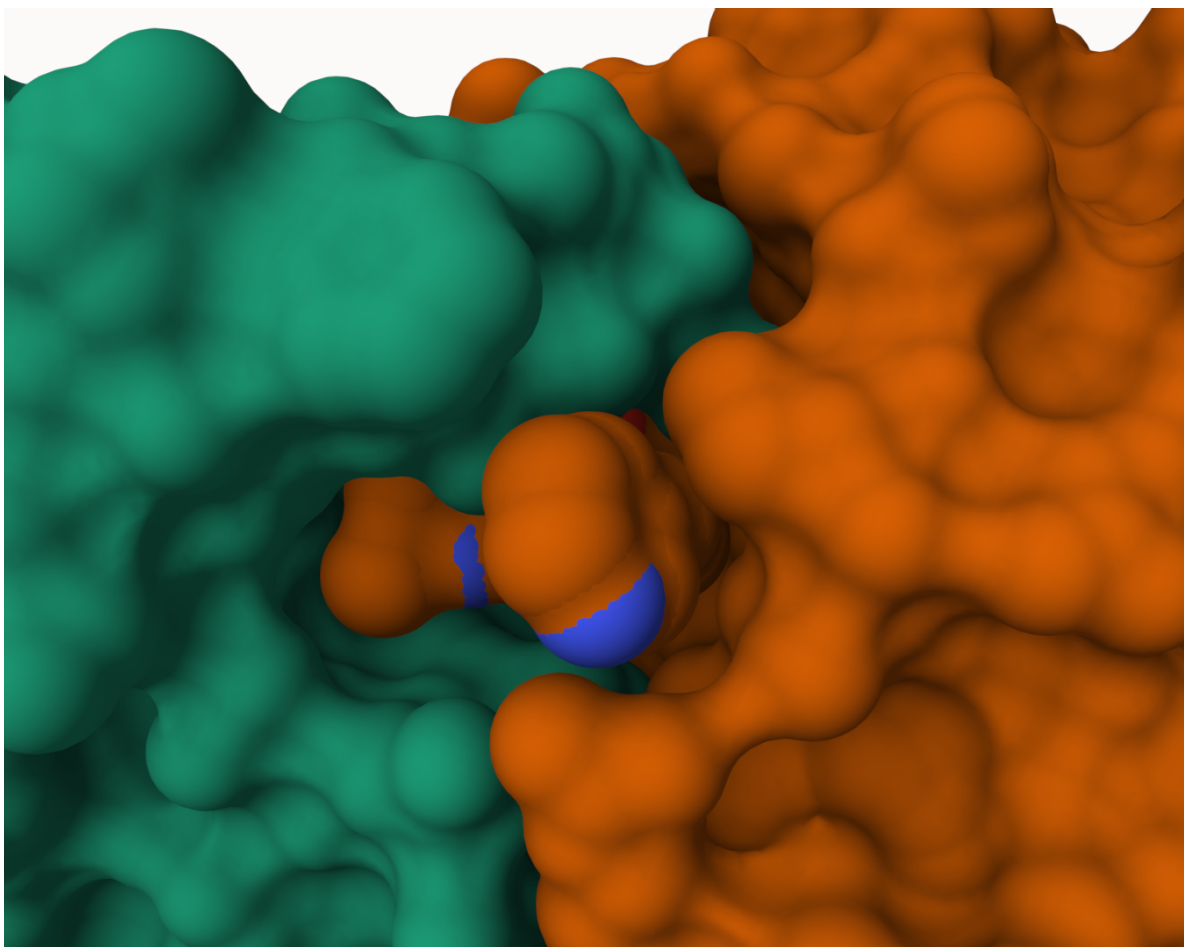


Figure 3: A third image from molstar

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

This is because it primarily detects the electron density of heavier atoms like oxygen, but not hydrogen atoms. Hydrogen atoms have much fewer electrons and typically don't show up well in electron density maps. Thus, water molecules are depicted with only their oxygen atoms.

Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

This water molecule has a residue of 308.



Figure 4: Water molecule

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

The Bio3D package

The bio3d package allows us to do all sorts of structural bioinformatics work in R.

Let's start with how it can read these pdb files:


```
library(bio3d)
```

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
```

```
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD  
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE  
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP  
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

`pdbseq(pdb)`

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20
"P"	"Q"	"I"	"T"	"L"	"W"	"Q"	"R"	"P"	"L"	"V"	"T"	"I"	"K"	"I"	"G"	"G"	"Q"	"L"	"K"
21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40
"E"	"A"	"L"	"L"	"D"	"T"	"G"	"A"	"D"	"D"	"T"	"V"	"L"	"E"	"E"	"M"	"S"	"L"	"P"	"G"
41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60
"R"	"W"	"K"	"P"	"K"	"M"	"I"	"G"	"G"	"I"	"G"	"G"	"F"	"I"	"K"	"V"	"R"	"Q"	"Y"	"D"
61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80
"Q"	"I"	"L"	"I"	"E"	"I"	"C"	"G"	"H"	"K"	"A"	"I"	"G"	"T"	"V"	"L"	"V"	"G"	"P"	"T"
81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	1
"P"	"V"	"N"	"I"	"I"	"G"	"R"	"N"	"L"	"L"	"T"	"Q"	"I"	"G"	"C"	"T"	"L"	"N"	"F"	"P"
2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21
"Q"	"I"	"T"	"L"	"W"	"Q"	"R"	"P"	"L"	"V"	"T"	"I"	"K"	"I"	"G"	"G"	"Q"	"L"	"K"	"E"
22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41
"A"	"L"	"L"	"D"	"T"	"G"	"A"	"D"	"D"	"T"	"V"	"L"	"E"	"E"	"M"	"S"	"L"	"P"	"G"	"R"
42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61
"W"	"K"	"P"	"K"	"M"	"I"	"G"	"G"	"I"	"G"	"G"	"F"	"I"	"K"	"V"	"R"	"Q"	"Y"	"D"	"Q"
62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81
"I"	"L"	"I"	"E"	"I"	"C"	"G"	"H"	"K"	"A"	"I"	"G"	"T"	"V"	"L"	"V"	"G"	"P"	"T"	"P"
82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99		
"V"	"N"	"I"	"I"	"G"	"R"	"N"	"L"	"L"	"T"	"Q"	"I"	"G"	"C"	"T"	"L"	"N"	"F"		

Q7: How many amino acid residues are there in this pdb object?

```
length(pdbseq(pdb))
```

```
[1] 198
```

```
sum(pdb$calpha)
```

```
[1] 198
```

There are 198 amino acid residues in this pdb object.

Q8: Name one of the two non-protein residues?

HOH and MK1

Q9: How many protein chains are in this structure?

There are 2 protein chains in this structure.

```
unique(pdb$atom$chain)
```

```
[1] "A" "B"
```

Predicting functional motions of a single structure

Let's do a bioinformatics prediction of functional motions - i.e. the movements that one of these molecules needs to make to do its stuff.

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
```

```
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM TAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

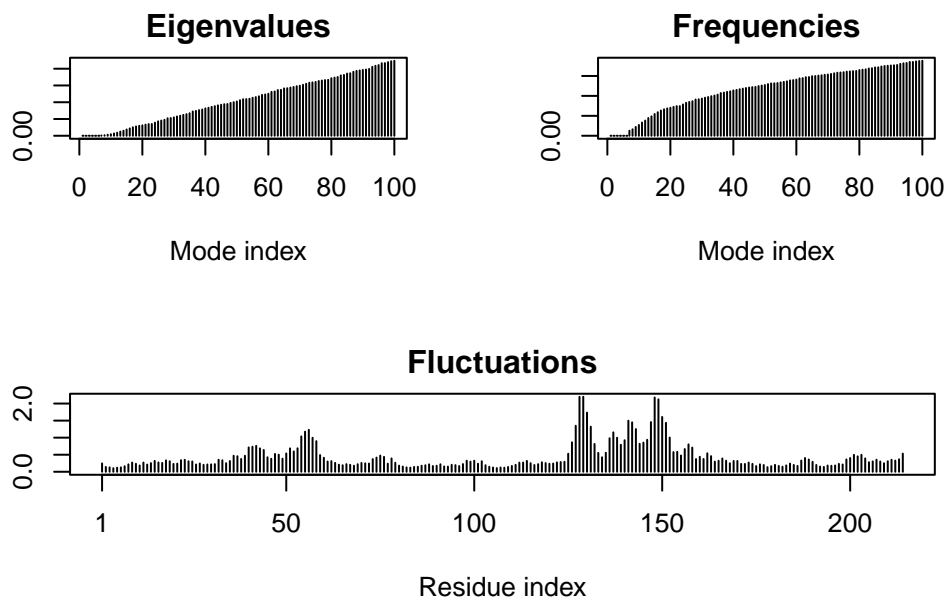
```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

```
# Perform flexibility prediction  
m <- nma(adk)
```

```
Building Hessian... Done in 0.014 seconds.
```

```
Diagonalizing Hessian... Done in 0.282 seconds.
```

```
plot(m)
```



Write out multi-model PDb file (trajectory) that we can use to make an animation of the predicted motions.

```
mktrj(m, file="adk.pdb")
```

I can open this in Mol* to play the trajectory...

Comparative analysis of Protein structure

```
library(bio3d)
```

Here we will find and analyze all ADK structures in the PDB database.

We will start with a single database accession id: "1ake_A"

```
id <-"1ake_A"
aa <- get.seq(id)
```

Warning in get.seq(id): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

aa

```
      1      .      .      .      .      .      .      60
pdb|1AKE|A MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
      1      .      .      .      .      .      .      60

      61      .      .      .      .      .      .      120
pdb|1AKE|A DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      .      120

      121      .      .      .      .      .      .      180
pdb|1AKE|A VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM TAPLIG
      121      .      .      .      .      .      .      180

      181      .      .      .      214
pdb|1AKE|A YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
      181      .      .      .      214
```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

I ran these cmds in the R brain/console

Q10. Which of the packages above is found only on BioConductor and not CRAN?

The `msa` package

Q11. Which of the above packages is not found on BioConductor or CRAN?

N/A

Q12 True or False? Functions from the `devtools` package can be used to install packages from GitHub and BitBucket?

TRUE

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

```
length(aa)
```

```
[1] 3
```

```
attributes(aa)
```

```
$names
```

```
[1] "id"    "ali"   "call"
```

```
$class
```

```
[1] "fasta"
```

```
ncol(aa$ali)
```

```
[1] 214
```

```
#b <- blast.pdb(aa)
```

```
#plot(b)
```

```
#hits <- plot(b)
```

```
#head(hits$pdb.id)
```

Pre-Calculated results

```
hits <- NULL
```

```
hits$pdb.id <- c('1AKE_A', '6S36_A', '6RZE_A', '3HPR_A', '1E4V_A', '5EJE_A', '1E4Y_A', '3X2S_A', '6H
```

```
# Download related PDB files
```

```
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/1AKE.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6RZE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb.gz exists. Skipping download

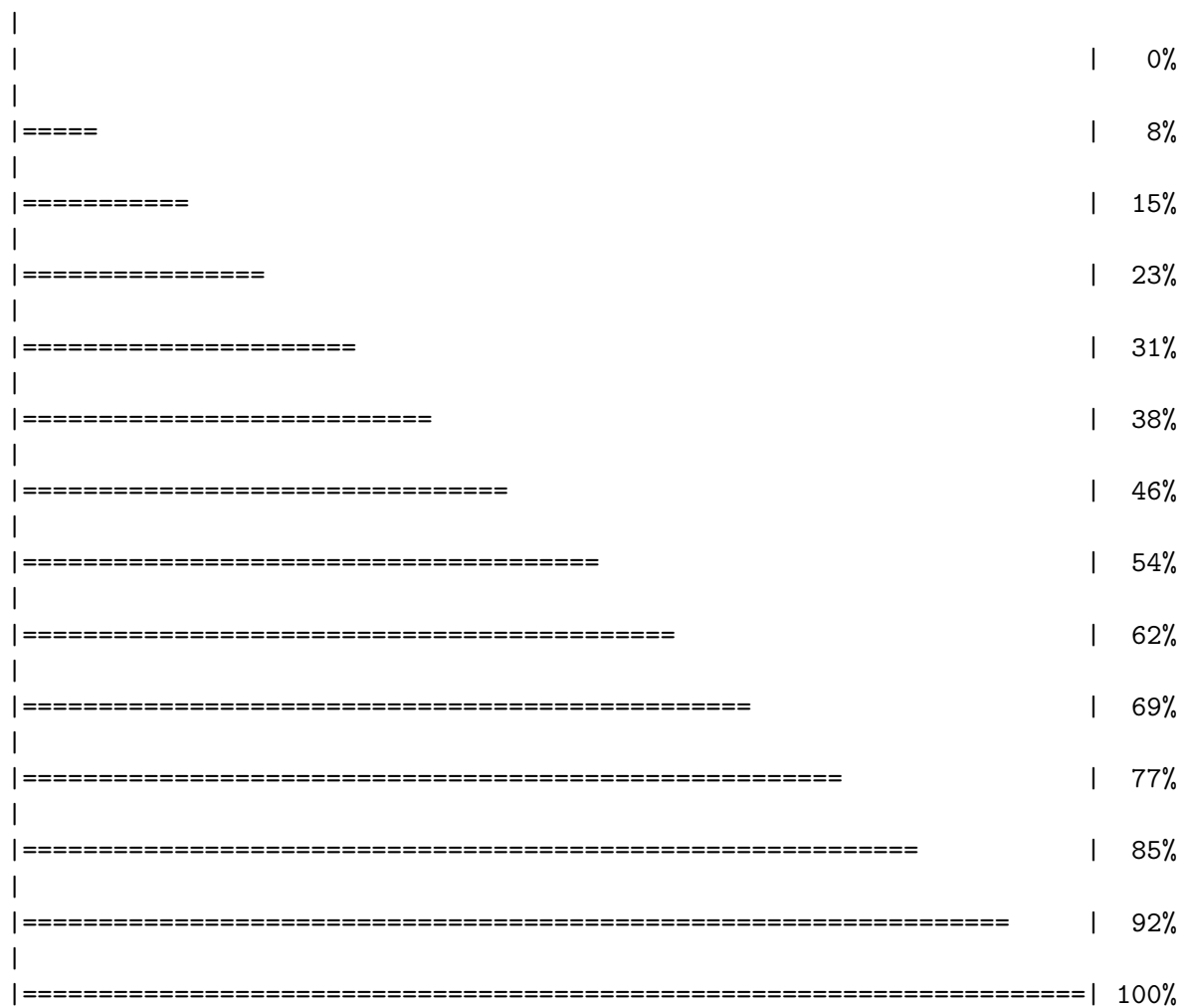
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4PZL.pdb.gz exists. Skipping download



```
# Align releated PDBs
pddb <- pddbain(files, fit = TRUE, exefile="msa")
```

```
Reading PDB files:
pddb/split_chain/1AKE_A.pdb
pddb/split_chain/6S36_A.pdb
pddb/split_chain/6RZE_A.pdb
pddb/split_chain/3HPR_A.pdb
pddb/split_chain/1E4V_A.pdb
pddb/split_chain/5EJE_A.pdb
pddb/split_chain/1E4Y_A.pdb
pddb/split_chain/3X2S_A.pdb
```

```

pdbc/split_chain/6HAP_A.pdb
pdbc/split_chain/6HAM_A.pdb
pdbc/split_chain/4K46_A.pdb
pdbc/split_chain/3GMT_A.pdb
pdbc/split_chain/4PZL_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
....   PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
...

```

Extracting sequences

```

pdb/seq: 1    name: pdbc/split_chain/1AKE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2    name: pdbc/split_chain/6S36_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3    name: pdbc/split_chain/6RZE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 4    name: pdbc/split_chain/3HPR_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5    name: pdbc/split_chain/1E4V_A.pdb
pdb/seq: 6    name: pdbc/split_chain/5EJE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7    name: pdbc/split_chain/1E4Y_A.pdb
pdb/seq: 8    name: pdbc/split_chain/3X2S_A.pdb
pdb/seq: 9    name: pdbc/split_chain/6HAP_A.pdb
pdb/seq: 10   name: pdbc/split_chain/6HAM_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 11   name: pdbc/split_chain/4K46_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12   name: pdbc/split_chain/3GMT_A.pdb
pdb/seq: 13   name: pdbc/split_chain/4PZL_A.pdb

```

pdbc

	1	.	.	.	40
[Truncated_Name:1] 1AKE_A.pdb	-----	MRIILLGAPGAGKGTQAQFIMEKYGIPQIS			
[Truncated_Name:2] 6S36_A.pdb	-----	MRIILLGAPGAGKGTQAQFIMEKYGIPQIS			

[Truncated_Name:12] 3GMT_A.pdb	RLKEADCANGYLFDFGFPRTIAQADAMKEAGVAIDYVLEID	
[Truncated_Name:13] 4PZL_A.pdb	RISKNCNNGFLLDGVPRTIPQAQELDKLGVNIDYIVEVD	
	*~ * *~* ** ***** ** ^ *~ ^***~* *	
	81 . . .	120
	121 . . .	160
[Truncated_Name:1] 1AKE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:2] 6S36_A.pdb	VPDELIVDKIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:3] 6RZE_A.pdb	VPDELIVDAIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:4] 3HPR_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDGTG	
[Truncated_Name:5] 1E4V_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:6] 5EJE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:7] 1E4Y_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:8] 3X2S_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:9] 6HAP_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:10] 6HAM_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG	
[Truncated_Name:11] 4K46_A.pdb	VADSVIVERMAGRAHLASGRTYHNVNPPKVEGKDDVTG	
[Truncated_Name:12] 3GMT_A.pdb	VPFSEIIERMSGRRTHPASGRTYHVKNPPKVEGKDDVTG	
[Truncated_Name:13] 4PZL_A.pdb	VADNLLIERITGRIHPASGRTYHTKFNPPKVADKDDVTG	
	* ^^^ ^ *** * *** * ^***** *** **	
	121 . . .	160
	161 . . .	200
[Truncated_Name:1] 1AKE_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:2] 6S36_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:3] 6RZE_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:4] 3HPR_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:5] 1E4V_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:6] 5EJE_A.pdb	EELTTRKDDQEECVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:7] 1E4Y_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:8] 3X2S_A.pdb	EELTTRKDDQEETVRKRLCEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:9] 6HAP_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:10] 6HAM_A.pdb	EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN	
[Truncated_Name:11] 4K46_A.pdb	EDLVIREDDKEETVLARLGVYHNQTAPLIAYYGKEAEAGN	
[Truncated_Name:12] 3GMT_A.pdb	EPLVQRDDDKKEETVKKRLDVYEAQTKPLITYYGDWARRGA	
[Truncated_Name:13] 4PZL_A.pdb	EPLITRTDDNEDTVKQRLSVYHAQTAKLIDFYRNFSSNT	
	* * * * * ^ * * * * * ^*	
	161 . . .	200
	201 . . .	227
[Truncated_Name:1] 1AKE_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-	
[Truncated_Name:2] 6S36_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-	
[Truncated_Name:3] 6RZE_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-	

```

[Truncated_Name:4] 3HPR_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:5] 1E4V_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:6] 5EJE_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:7] 1E4Y_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:8] 3X2S_A.pdb      T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:9] 6HAP_A.pdb      T--KYAKVDGTPVCEVRADLEKILG-
[Truncated_Name:10] 6HAM_A.pdb      T--KYAKVDGTPVCEVRADLEKILG-
[Truncated_Name:11] 4K46_A.pdb      T--QYLKFDGTKAFAEVSAAELEKALA-
[Truncated_Name:12] 3GMT_A.pdb      E-----NGLKAPA-----YRKISG-
[Truncated_Name:13] 4PZL_A.pdb      KIPKYIKINGDQAVEKVSQDIFDQLNK
                                     *
                                201      .      .      227

```

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdbs, fasta
```

Alignment dimensions:

```
13 sequence rows; 227 position columns (204 non-gap, 23 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

pdbs

```

                                1      .      .      .      40
[Truncated_Name:1] 1AKE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:2] 6S36_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:3] 6RZE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:4] 3HPR_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:5] 1E4V_A.pdb      -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:6] 5EJE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:7] 1E4Y_A.pdb      -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:8] 3X2S_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:9] 6HAP_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:10] 6HAM_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:11] 4K46_A.pdb      -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
[Truncated_Name:12] 3GMT_A.pdb      -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
[Truncated_Name:13] 4PZL_A.pdb      TENLYFQSNAMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
                                **~*****  *****  *  *~ *  **
                                1      .      .      .      40

```

	41	.	.	.	80
[Truncated_Name:1] 1AKE_A.pdb	TG	D	M	L	R
[Truncated_Name:2] 6S36_A.pdb	TG	D	M	L	R
[Truncated_Name:3] 6RZE_A.pdb	TG	D	M	L	R
[Truncated_Name:4] 3HPR_A.pdb	TG	D	M	L	R
[Truncated_Name:5] 1E4V_A.pdb	TG	D	M	L	R
[Truncated_Name:6] 5EJE_A.pdb	TG	D	M	L	R
[Truncated_Name:7] 1E4Y_A.pdb	TG	D	M	L	R
[Truncated_Name:8] 3X2S_A.pdb	TG	D	M	L	R
[Truncated_Name:9] 6HAP_A.pdb	TG	D	M	L	R
[Truncated_Name:10] 6HAM_A.pdb	TG	D	M	L	R
[Truncated_Name:11] 4K46_A.pdb	TG	D	M	L	R
[Truncated_Name:12] 3GMT_A.pdb	TG	D	M	L	R
[Truncated_Name:13] 4PZL_A.pdb	TG	D	M	L	R

****~* ~* *~ ** * ~* ** * ^^ ~*^^

41	.	.	.	80
----	---	---	---	----

	81	.	.	.	120
[Truncated_Name:1] 1AKE_A.pdb	RIA	Q	E	D	C
[Truncated_Name:2] 6S36_A.pdb	RIA	Q	E	D	C
[Truncated_Name:3] 6RZE_A.pdb	RIA	Q	E	D	C
[Truncated_Name:4] 3HPR_A.pdb	RIA	Q	E	D	C
[Truncated_Name:5] 1E4V_A.pdb	RIA	Q	E	D	C
[Truncated_Name:6] 5EJE_A.pdb	RIA	Q	E	D	C
[Truncated_Name:7] 1E4Y_A.pdb	RIA	Q	E	D	C
[Truncated_Name:8] 3X2S_A.pdb	RIA	Q	E	D	S
[Truncated_Name:9] 6HAP_A.pdb	RI	C	Q	E	D
[Truncated_Name:10] 6HAM_A.pdb	RI	C	Q	E	D
[Truncated_Name:11] 4K46_A.pdb	RIA	Q	D	D	C
[Truncated_Name:12] 3GMT_A.pdb	RL	K	E	A	D
[Truncated_Name:13] 4PZL_A.pdb	R	I	S	K	N

*~ * *~* ** ***** ** ^ *~ ^***^* *

81	.	.	.	120
----	---	---	---	-----

	121	.	.	.	160
[Truncated_Name:1] 1AKE_A.pdb	VP	D	E	L	I
[Truncated_Name:2] 6S36_A.pdb	VP	D	E	L	I
[Truncated_Name:3] 6RZE_A.pdb	VP	D	E	L	I
[Truncated_Name:4] 3HPR_A.pdb	VP	D	E	L	I
[Truncated_Name:5] 1E4V_A.pdb	VP	D	E	L	I
[Truncated_Name:6] 5EJE_A.pdb	VP	D	E	L	I
[Truncated_Name:7] 1E4Y_A.pdb	VP	D	E	L	I

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdb, fasta
```

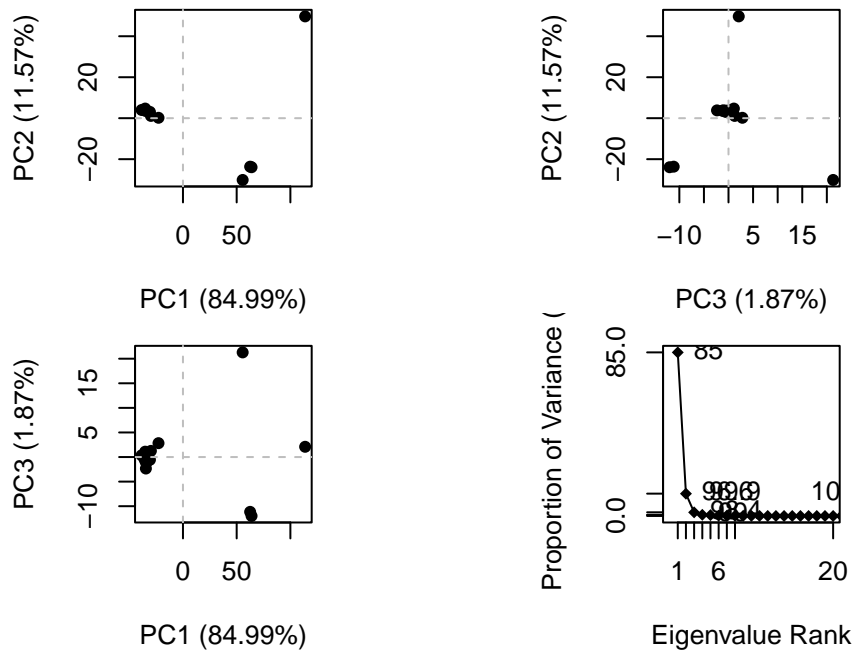
Alignment dimensions:

```
13 sequence rows; 227 position columns (204 non-gap, 23 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

Principle Component Analysis

```
# Perform PCA  
pc.xray <- pca(pdb)  
plot(pc.xray)
```

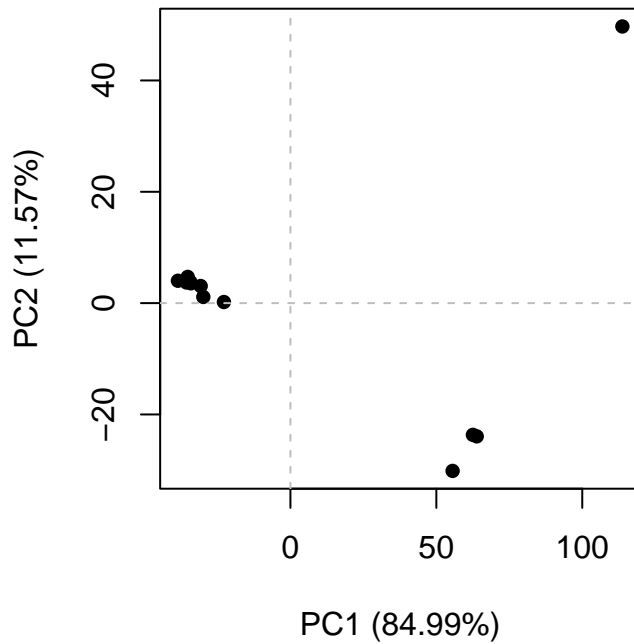


To visualize the major structural variations in the ensemble the function `mktrj()` can be used to generate a trajectory PDB file by interpolating along a given PC (eigenvector):


```
# Visualize first principal component  
pc1 <- mktrj(pc.xray, pc=1, file="pc_1.pdb")
```

```
pc.xray <-pca(pdb)
```

```
plot(pc.xray, pc.axes = c(1,2))
```



```
uniprot <- 24883887  
pdb <- 195610  
pdb/uniprot *100
```

```
[1] 0.0786091
```