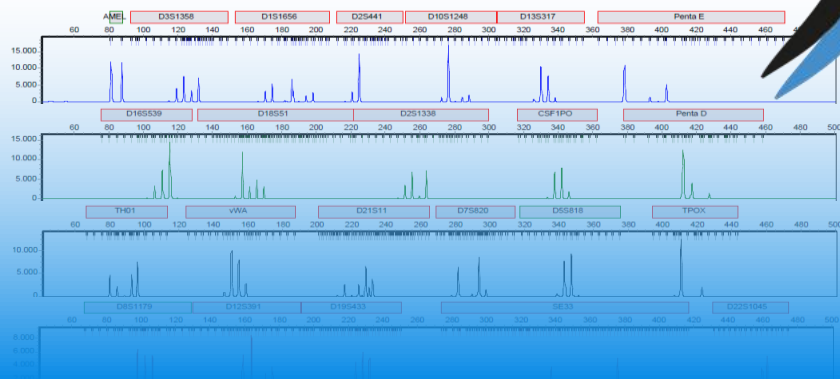
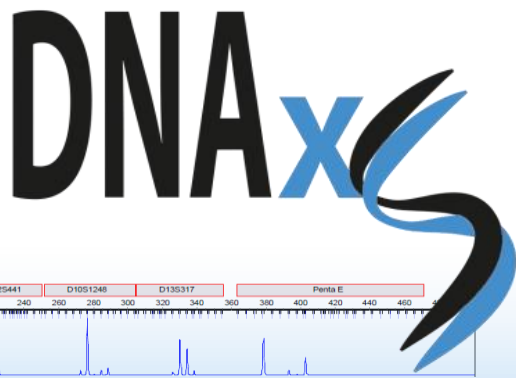




Netherlands Forensic Institute  
Ministry of Justice and Security

# Interpretation of (mixed) DNA profiles within the NFI



*Corina Benschop*

*13/11/2020*

# DNA profile interpretation QOL-00711

**Step 1:**

Define whether the profile is single source or a mixture

**Step 2:**

Identify the number of contributors to the mixture

**Step 3:**

Estimate the relative ratio of the individuals contributing to the mixture, and, if possible deconvolve the mixture

**Step 4:**

Determine whether the profile is suitable for comparison to a reference profile/  
suitable for storage or search in the DNA database

**Step 5A:**

Compare the evidence profile to the reference profile(s)

**Step 5B:**

DDB storage/search

**Step 6:**

Statistical analyses to define the weight of evidence

Today

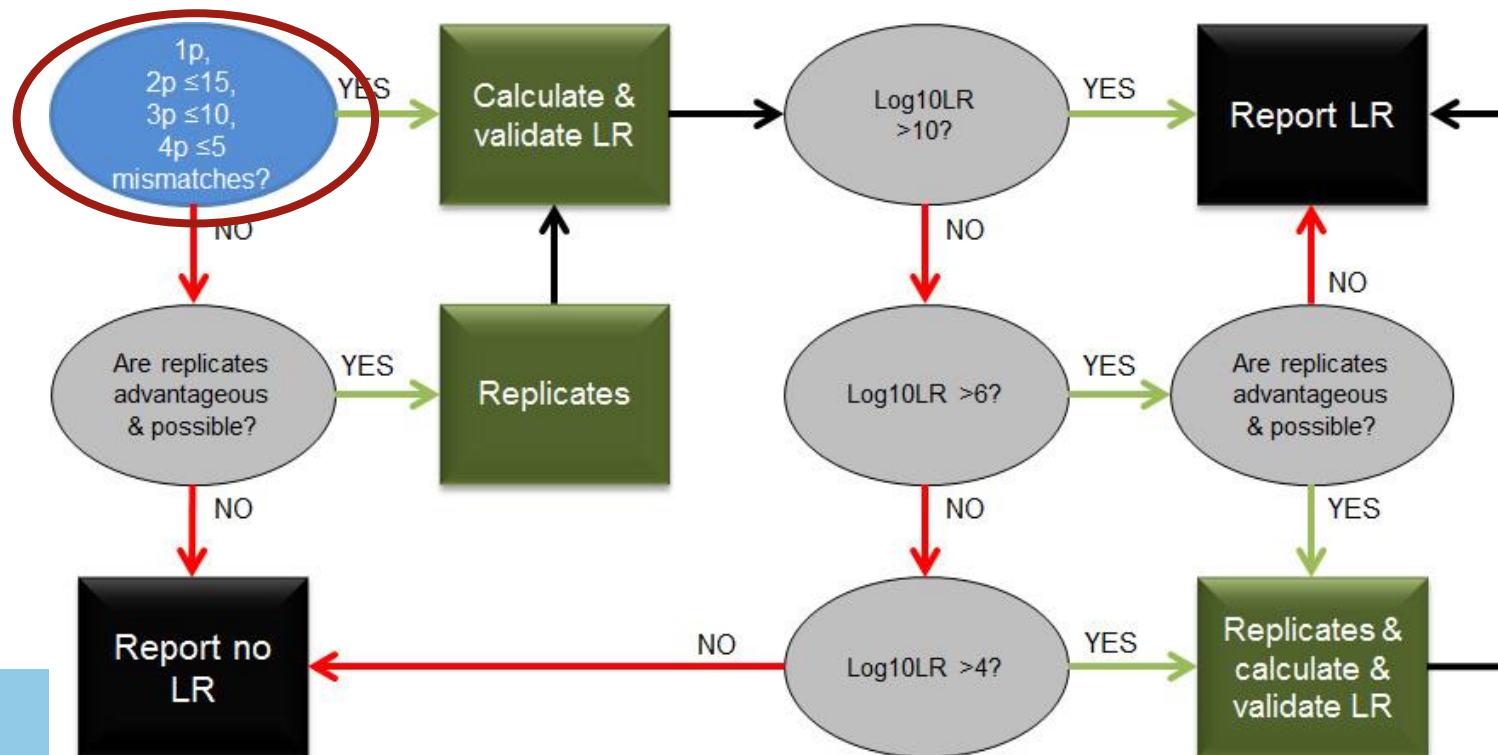




# Guidelines for use in forensic casework

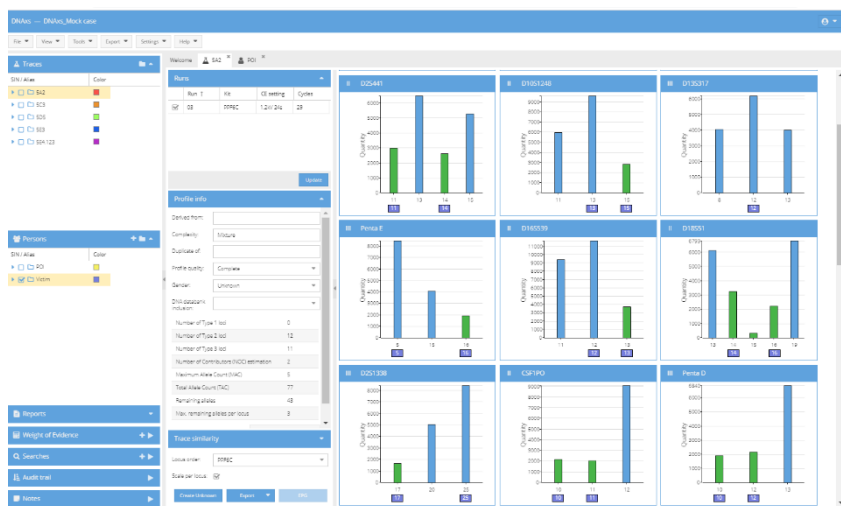
- Based on results from research, casework experience, including efficiency and usefulness of calculations.
- Clearly, the case circumstances, availability of other stains and profiles within the case are factors considered in the process.

Summarized guidelines to be used as helpful tool in the decision making process are as follows:



# Mismatches?

- Alleles of, for example, a person of interest (POI) that are not observed in the trace profile. I.e. unseen alleles, and in case of true donors, drop-outs.

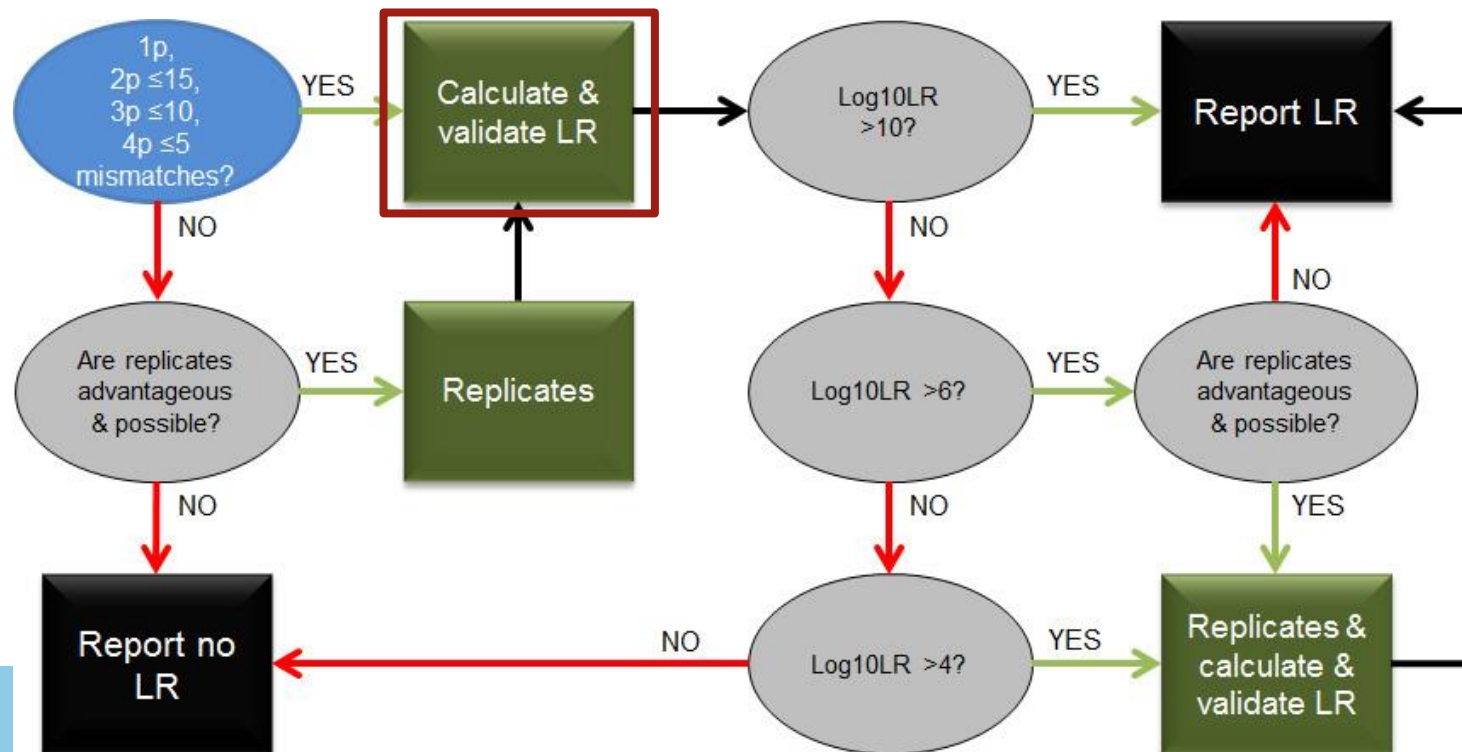




# Guidelines for use in forensic casework

- Based on results from research, casework experience, including efficiency and usefulness of calculations.
- Clearly, the case circumstances, availability of other stains and profiles within the case are factors considered in the process.

Summarized guidelines to be used as helpful tool in the decision making process are as follows:



# Weight of Evidence

- The likelihood ratio (LR) is a measure for weight of evidence.
- Using LR calculations we weigh hypotheses  $H_1$  (or  $H_p$ ) and  $H_2$  (or  $H_d$ ), given observed data.
- LR gives a weight of whether the data supports  $H_1$  or  $H_2$ .

LR value	
$LR = 1$	Neutral evidence, the data support none of the hypotheses
$LR > 1$	The data supports that $H_1$ is true
$LR < 1$	The data supports that $H_2$ is true



## Binary | qualitative | quantitative

- Binary (classical) likelihood ratio calculation
  - Considers genotypic probabilities
- Qualitative/ semi-continuous model:
  - Considers genotypic probabilities
  - Considers drop-out and drop-in
- Quantitative/ continuous model:
  - Considers genotypic probabilities
  - Considers peak height information
  - Drop-out and drop-in based on peak height models





# Underlying model

- LR is a function of the genotypic frequencies
  - Hardy Weinberg Equilibrium, assume independent association of the alleles within loci:
    - Genotype 'aa' has frequency  $p_a^2$
    - Genotype 'ab' has frequency  $2p_ap_b$
  - Population Sub-structure model
    - Introduces "Fst/Theta" correction
    - Genotype probabilities increase (reduction in LR)
    - Balding-Nichols sampling formula
- Linkage equilibrium
  - Markers bear independent information
  - The product rule: Multiply between loci





# Steps in computing likelihood ratios

User: Define propositions (number of contributors, person of interest etc.)

## Binary

Define possible genotype combinations



Determine genotypic probabilities



Calculate profile likelihoods

Sum up probabilities/locus & multiply between loci



Calculate LR:

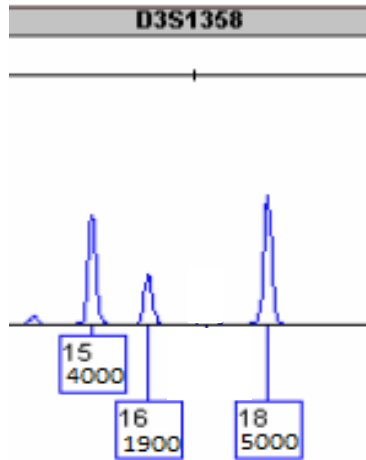
likelihood  $H_p$  / likelihood  $H_d$





# Example of a binary (classical) likelihood ratio calculation

## Example: High-template two-person mixture



### **D3S1358**

Evidence	15, 16, 18
Victim	15, 18
POI	16, 18

$$LR = \frac{Hp: \text{Victim} + POI \text{ contributed to the sample}}{Hd: \text{Victim} + \text{Unknown person} \\ (\text{unrelated to the POI}) \text{ contributed to the sample}}$$



$$LR = \frac{Hp: V + POI}{Hd: V + U}$$



## Under $H_p$

$H_p: V + POI$

**D3S1358**

Evidence	15, 16, 18
----------	------------

Victim	15, 18
--------	--------

POI	16, 18
-----	--------



In this case, there  
are no unknowns

$$Pr(Evidence | H_p) = 1$$



Ne  
Mi

Sum up the probabilities for all plausible genotypes

Calculate profile likelihood

## Under $H_d$

$H_d: V + U$

**D3S1358**

Evidence	15, 16, 18
Victim	15, 18
Unknown	?

$2p_{15}p_{16}$

$2p_{16}p_{18}$

$p_{16}^2$

$$Pr(\text{Evidence} | H_d) = 2p_{15}p_{16} + 2p_{16}p_{18} + p_{16}^2$$

$$(2 \times 0.25 \times 0.04) + (2 \times 0.04 \times 0.15) + (0.04^2) = 0.0336$$

- The victim's profile explains alleles 15 and 18
- The unknown has to have allele **16**: allele **16** is constrained



## Under $H_p$ and $H_d$

$H_p: V + POI$

**D3S1358**

Evidence 15, 16, 18

Victim 15, 18

POI 16, 18

Likelihood under  $H_p$

$$\Rightarrow Pr(\text{Evidence} \mid H_p) = 1$$

$H_d: V + U$

**D3S1358**

Evidence 15, 16, 18

Victim 15, 18

Unknown 15, **16**

**16**, 18

**16**, **16**

Likelihood under  $H_d$

$$\Rightarrow Pr(\text{Evidence} \mid H_d) = 2p_{15}p_{16} + 2p_{16}p_{18} + p_{16}^2$$



## Under $H_p$ and $H_d$ : LR for 1 locus

*$H_p: V + POI$*

**D3S1358**

Evidence 15, 16, 18

Victim 15, 18

POI 16, 18

*$H_d: V + U$*

**D3S1358**

Evidence 15, 16, 18

Victim 15, 18

Unknown 15, **16**

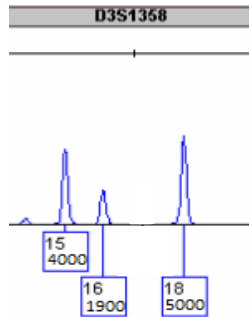
**16**, 18

**16**, **16**

$$LR = \frac{1}{2p_{15}p_{16} + 2p_{16}p_{18} + p_{16}^2}$$

$$LR = \frac{1}{0.0336} = 29.76$$

# Example with more loci: Multiply between loci



## D3S1358

Evidence	15, 16, 18
Victim	15, 18
Unknown	15, <b>16</b>
	<b>16</b> , 18
	<b>16</b> , <b>16</b>

## Locus 2

Evidence	10, 12, 14, 15
Victim	14, 15
Unknown	<b>10</b> , <b>12</b>

## Locus 3

Evidence	9, 9.3
Victim	9, 9.3
Unknown	9, 9.3
	9, 9
	9.3, 9.3

$$2p_{15}p_{16} + 2p_{16}p_{18} + p_{16}^2$$

X

$$2p_{10}p_{12}$$

X

$$2p_9p_{9.3} + p_9^2 + p_{9.3}^2$$





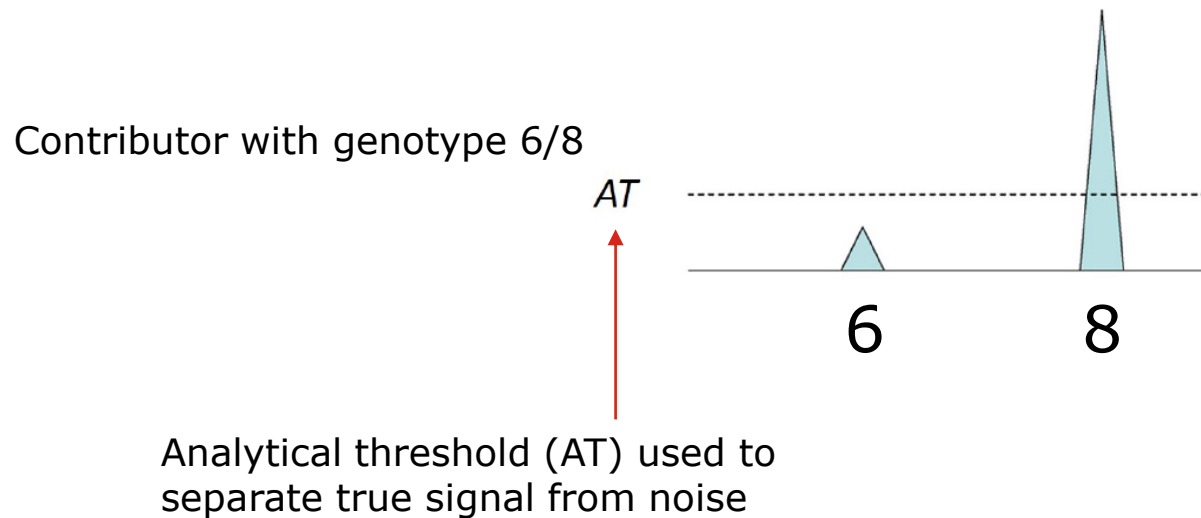
# Steps in computing likelihood ratios

User: Define propositions (number of contributors, person of interest etc.)

	Binary	Qualitative
Define possible genotype combinations	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
Determine genotypic probabilities	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
Estimate drop-out probabilities	<input type="checkbox"/>	<input checked="" type="checkbox"/>
Calculate profile likelihoods Sum up probabilities/locus & multiply between loci	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
Calculate LR: likelihood $H_p$ / likelihood $H_d$	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>

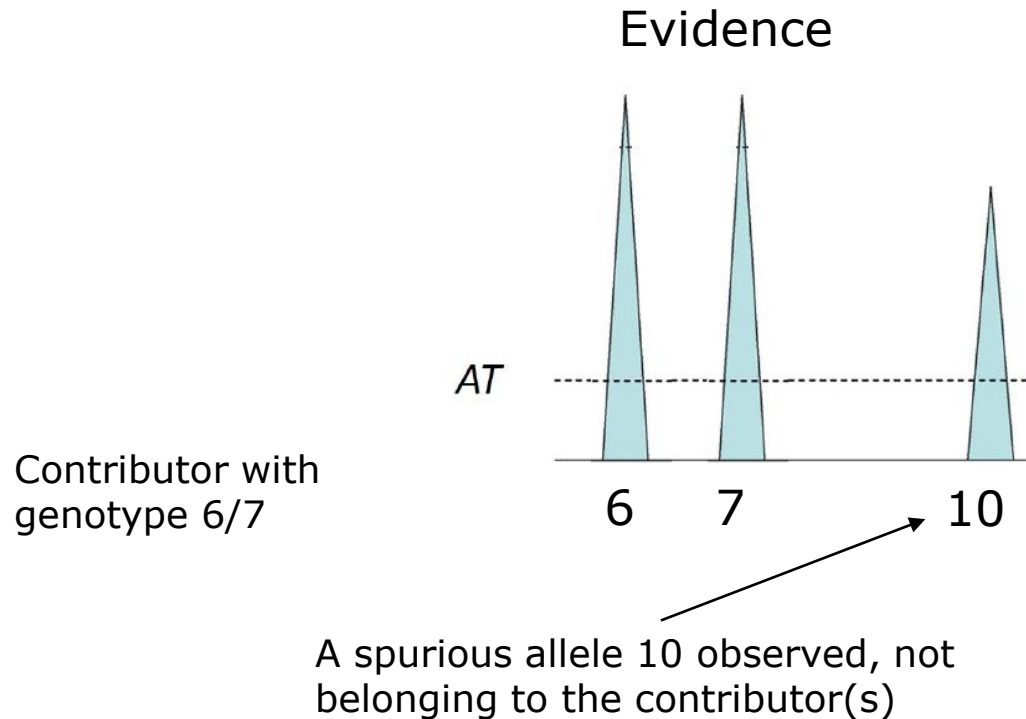
# The drop-out phenomena

## Evidence



Small DNA-amount leads to low peak falling below AT  
Corresponding allele removed from evidence

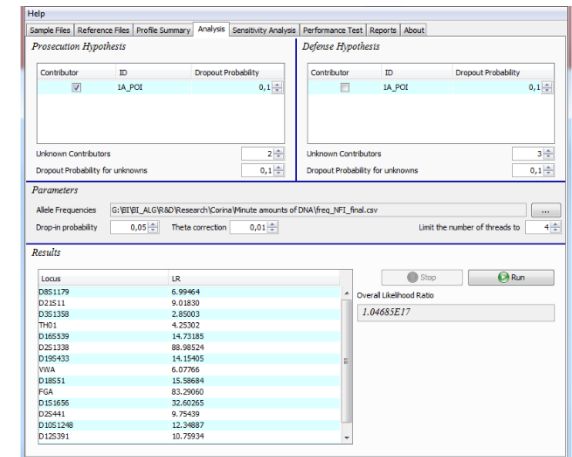
# The drop-in phenomena



Typical more likely with smaller peak heights  
than higher peak heights

# LR with drop-out and drop-in *using the qualitative model*

- Main theory described in:
  - Haned et al, FSIG 2012
  - DNA commission ISFG, FSIG 2012
  - Gill et al, FSI 2007
  - **Curran et al, FSI 2005**
- Two key parameters in the model:
  - Drop-out & drop-in



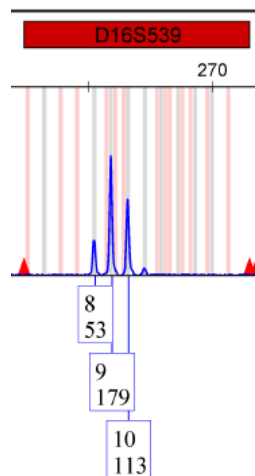
The screenshot shows the LocusZoom software interface. The 'Prosecution Hypothesis' section has a table with columns 'Contributor', 'ID', and 'Dropout Probability'. The 'Defense Hypothesis' section has a similar table. The 'Parameters' section includes 'Allele Frequencies' (G:\WV\AL\GVR\Research\Corne\Minute amounts of DNA\free\_JPT\_final.csv), 'Drop-in probability' (0.05), 'Theta correction' (0.01), and 'Limit the number of threads to' (4). The 'Results' section shows a table of Loci and their LR values. The 'Overall Likelihood Ratio' is displayed as 1.04685E17.

Locus	LR
D8S1179	6.99464
D2S1318	9.01809
D3S1358	2.85003
TH01	4.25302
D16S539	14.73185
D2S1338	89.98324
D19S433	14.15405
VWA	6.07766
D18S51	15.58644
FGA	83.29060
D15S656	32.60285
D2S441	9.79439
D10S1248	12.34887
D12S391	10.75934

## Introducing drop-out parameter $d$

- An allele drops out with a probability of  $d$
- An allele does not drop out with a probability of  $1 - d$
- Allele drop-out from a heterozygote genotype:  $d$
- Allele drop-out from a homozygote genotype:  $d' = d^2$

# The Q-allele: a possible non-observed allele *explained as drop-out*



Q-allele, can be anything except the observed alleles (here 8, 9, 10)

$H_p: POI + U$

D16S539	
Evidence	8, 9, 10
POI	9, 10
Unknown	8, 8 8, 9 8, 10 8, Q

No drop-in →

Assuming drop-in →

Additional possibilities	Allele drop-in
9,9	8
9,10	8
9,Q	8
10,10	8
10,Q	8
Q,Q	8

This increases the number of terms under  $H_d$



# Steps in computing likelihood ratios

User: Define propositions (number of contributors, person of interest etc.)

	Binary	Qualitative	Quantitative
Estimate parameters (by optimizer) Mixture proportion, peak height (PH) expectation, PH variance, degradation slope, stutter			
Define possible genotype combinations			
Determine genotypic probabilities			
Estimate drop-out probabilities			
Compute PH weights			
Calculate profile likelihoods Sum up probabilities/locus & multiply between loci			
Calculate LR: likelihood $H_p$ / likelihood $H_d$			



## Various continuous models available

➤ EuroForMix Open source, freely available

Bulletproof

➤ DNASTatistX

➤ Kongoh

➤ LikeLTD

➤ CeesIT Commercial or proprietary

➤ LiRaHT

➤ TrueAllele

➤ STRmix

➤ DNAmixtures

➤ DNA View Mixture Solution

➤ Genoproof Mixture

➤ maSTR

➤ ...



# DNASTatistX is based on EuroForMix

➤ [euroformix.com](http://euroformix.com)



Forensic Science International: Genetics 21 (2016) 35–44



Contents lists available at ScienceDirect

Forensic Science International: Genetics

journal homepage: [www.elsevier.com/locate/fsig](http://www.elsevier.com/locate/fsig)



Research paper

**EuroForMix: An open source software based on a continuous model to evaluate STR DNA profiles from a mixture of contributors with artefacts**

Øyvind Bleka<sup>a,b,\*</sup>, Geir Storvik<sup>b,1</sup>, Peter Gill<sup>a,c,1</sup>

<sup>a</sup> Department of Forensic Biology, Norwegian Institute of Public Health, Oslo, Norway

<sup>b</sup> Department of Mathematics, University of Oslo, Oslo, Norway

<sup>c</sup> Department of Forensic Medicine, University of Oslo, Oslo, Norway



*Appl. Statist.* (2015)  
**64**, Part 1, pp. 1–32

## Analysis of forensic DNA mixtures with artefacts

R. G. Cowell,  
*City University London, UK*

T. Graversen and S. L. Lauritzen  
*University of Oxford, UK*

and J. Mortera  
*Università Roma Tre, Italy*

- EuroForMix uses the gamma distribution to model peak heights
- Alike DNA·VIEW Mixture Solution, LiRaHT, DNA-mixtures, LikeLTD, Kongoh

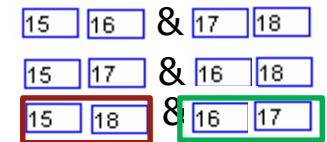
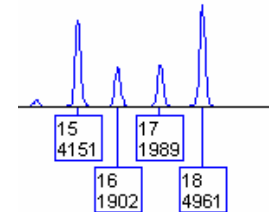


# What to expect from a quantitative model?

- Benefit from using peak heights in case
  - Contributors have distinctive peak heights and LR < highest reporting threshold
  - DNA-profiles show degradation

Trend: Larger weight of evidence for true contributors

Non-contributors that cannot be excluded based on allele matching  
might be excluded based on peak heights



- No/not much difference compared to semi-continuous model when peak heights are not very informative
  - In case donors contributed equally
  - In case all peaks are within the stochastic range
  - In case one can condition on known contributors

Forensic Science International: Genetics 25 (2016) 85–96



Contents lists available at ScienceDirect

Forensic Science International: Genetics

journal homepage: [www.elsevier.com/locate/fsig](http://www.elsevier.com/locate/fsig)



Research paper

A comparative study of qualitative and quantitative models used to interpret complex STR DNA profiles

Øyvind Bleka<sup>a,b,\*</sup>, Corina C.G. Benschop<sup>c</sup>, Geir Storvik<sup>b</sup>, Peter Gill<sup>a,d</sup>





Example of an LR calculation performed  
using a quantitative/ continuous model

*Software: EuroForMix (EFM) or DNASTatistX*

## Estimate parameters

The continuous model uses parameters:

- Mixture proportions
- Peak height expectation
- Peak height variance
- Degradation slope
- Backward stutter (n-1) proportion
- Forward stutter (n+1) proportion

$\beta$

To be **estimated** under  
 $H = H_p$  and  $H = H_d$   
separately

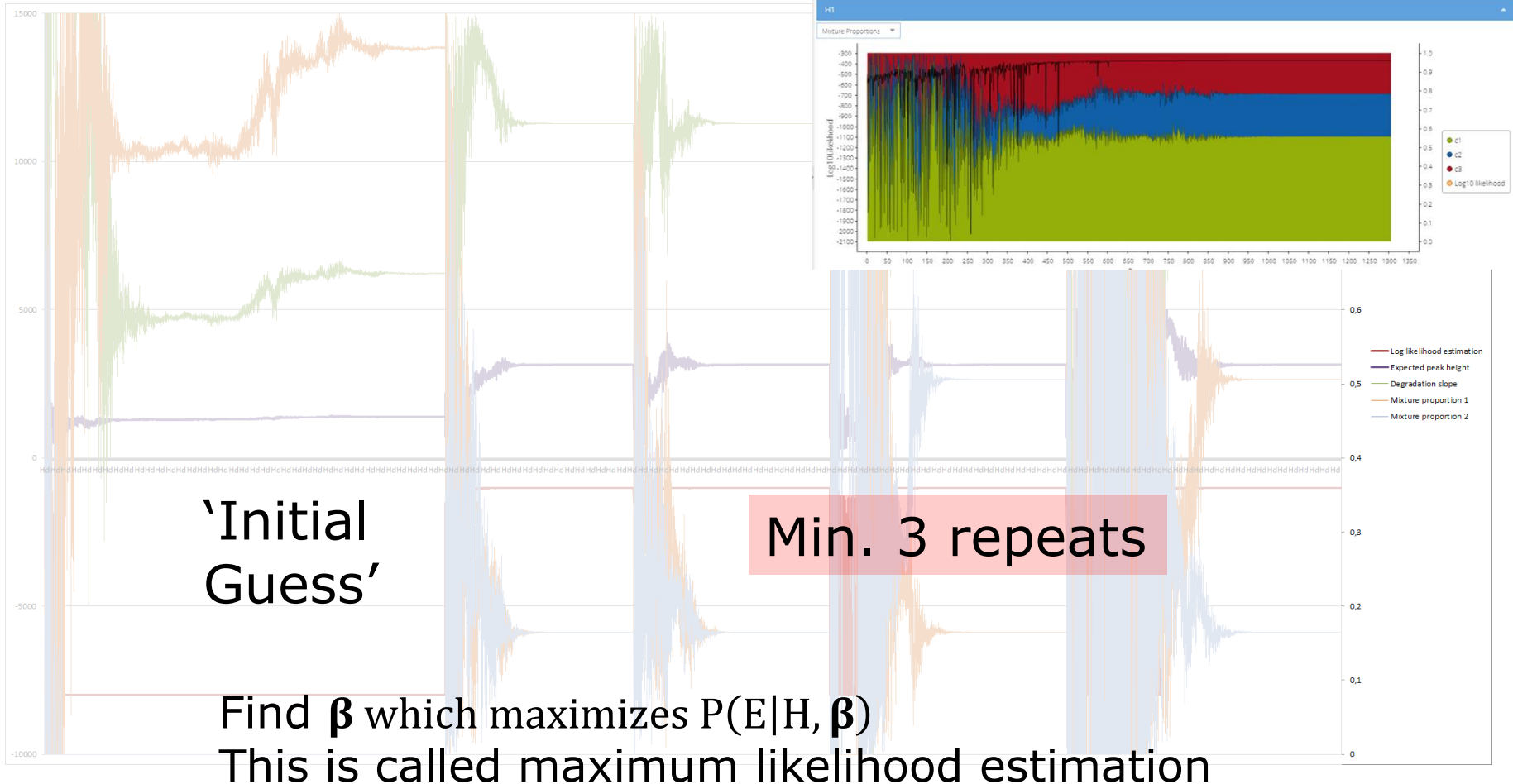
Using evidence  
profile data!

However, these are unknown, thus...  
Trial and error using optimizer



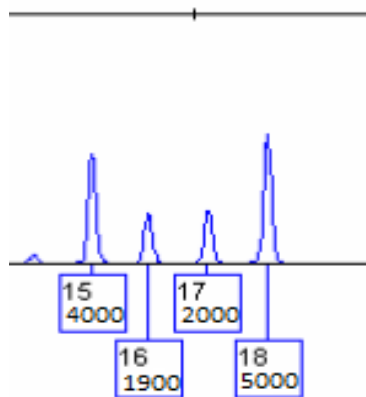


# Optimizer



## Example: Two-person mixture

D3S1358



**D16S539**

Evidence	15, 16, 17, 18
POI	16, 17

Assumption: two-person mixture

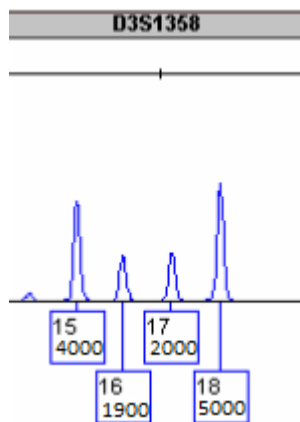
$$LR = \frac{H_p: POI + Unknown \text{ contributed to the sample}}{H_d: Two Unknown persons \text{ (unrelated to the POI) contributed to the sample}}$$



$$LR = \frac{H_p: POI + U}{H_d: U + U}$$

$$LR = \frac{Hp: POI + U}{Hd: U + U}$$

## Define possible genotype combinations



Possible genotype combinations when regarding drop-out and drop-in:

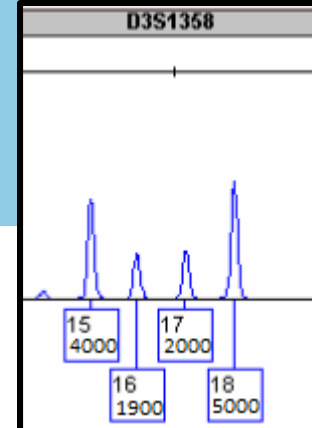
Donor A	Donor B	Drop-in
15/16	17/18	-
15/17	16/18	-
15/18	16/17	-
17/18	15/16	-
16/18	15/17	-
16/17	15/18	-
But also...		
15/15	16/17	18
15/18	16/d.o.	17
d.o.	d.o.	15/16/17/18
Etc...		

All possible genotypes per contributor ( $n=15$ ):

Donor A allele 1	Donor A allele 2
15	15
15	16
15	17
15	18
16	16
16	17
16	18
17	17
17	18
18	18
Q	Q
15	Q
16	Q
17	Q
18	Q

Drop out

With two unknowns:  $15 \times 15 = 225$  combinations

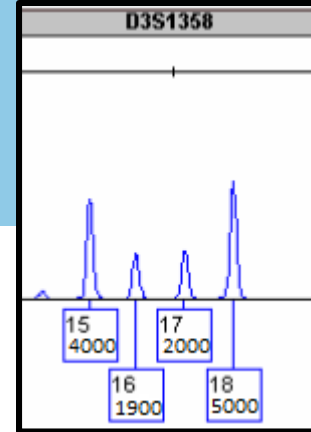


# Determine genotype probabilities

Using allele frequencies

Genotype probability heterozygote	Genotype probability homozygote	Frequency of a dropped-out allele	Drop-in probability	Probability for no drop-in
$2P_aP_b$	$P_a^2$	$P_Q = 1 - P_a - P_b - P_c - P_d$	$PrC * P_a$	$1 - PrC$

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability
15/16	17/18	-		
15/17	16/18	-		
15/18	16/17	-		
17/18	15/16	-		
16/18	15/17	-		
16/17	15/18	-		
But also...				
15/15	16/17	18		
15/18	16/d.o.	17		
d.o.	d.o.	15/16/17/18		
Etc...				



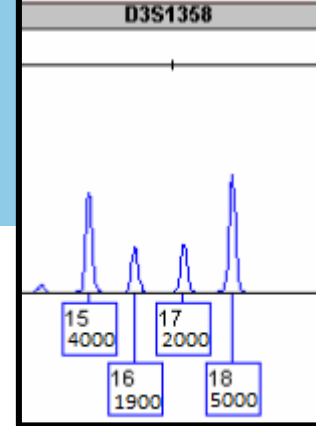
## Likelihoods excl. peak height information

- Calculate **likelihood weights** (per allele)  
Likelihoods **excl.** peak height information (qual model)

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability	Likelihood (product genotype & drop-in probabilities)
15/16	17/18	-	$2P_{15}P_{16} * 2P_{17}P_{18}$	$1 - \text{PrC}$	0.007929
15/17	16/18	-	$2P_{15}P_{17} * 2P_{16}P_{18}$	$1 - \text{PrC}$	0.007929
15/18	16/17	-	$2P_{15}P_{18} * 2P_{16}P_{17}$	$1 - \text{PrC}$	0.007929
17/18	15/16	-	$2P_{17}P_{18} * 2P_{15}P_{16}$	$1 - \text{PrC}$	0.007929
16/18	15/17	-	$2P_{16}P_{18} * 2P_{15}P_{17}$	$1 - \text{PrC}$	0.007929
16/17	15/18	-	$2P_{16}P_{17} * 2P_{15}P_{18}$	$1 - \text{PrC}$	0.007929
But also...					
15/15	16/17	18	$P_{15}^2 * 2P_{16}P_{17}$	$\text{PrC} * P_{18}$	4.1837E-06
15/18	16/d.o.	17	$2P_{15}P_{18} * 2P_{16}P_Q$	$\text{PrC} * P_{17}$	4.7860E-06
d.o.	d.o.	15/16/17/18	$P_Q^4$	$\text{PrC}^4 * P_{15} * P_{16} * P_{17} * P_{18}$	2.5981E-16
Etc...					
					+
					SUM = 0.047950

Allele likelihoods (weights)



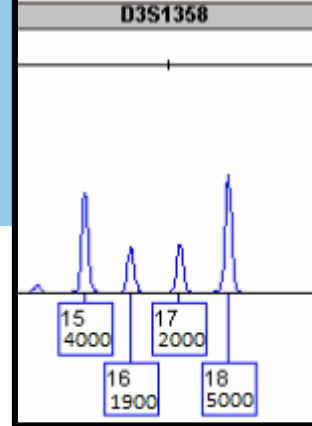


## Compute likelihood weights using peak height model

- Find drop-in, drop-out set and for each genotype combination
- Using gamma distribution model for PH (calculate likelihoods)
- Probability of observing a drop-in at a certain peak height: exponential distribution

Allele likelihoods based on PH model

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability	Likelihood excl PH	PH probability	Likelihood incl PH
15/16	17/18	-	$2P_{15}P_{16} * 2P_{17}P_{18}$	1-PrC	0,007929		
15/17	16/18	-	$2P_{15}P_{17} * 2P_{16}P_{18}$	1-PrC	0,007929		
15/18	16/17	-	$2P_{15}P_{18} * 2P_{16}P_{17}$	1-PrC	0,007929		
17/18	15/16	-	$2P_{17}P_{18} * 2P_{15}P_{16}$	1-PrC	0,007929		
16/18	15/17	-	$2P_{16}P_{18} * 2P_{15}P_{17}$	1-PrC	0,007929		
16/17	15/18	-	$2P_{16}P_{17} * 2P_{15}P_{18}$	1-PrC	0,007929		
But also...							
15/15	16/17	18	$P_{15}^2 * 2P_{16}P_{17}$	$PrC * P_{18}$	4,1837E-06		
15/18	16/d.o.	17	$2P_{15}P_{18} * 2P_{16}P_Q$	$PrC * P_{17}$	4,7860E-06		
d.o.	d.o.	15/16/ 17/18	$P_Q^4$	$PrC^4 * P_{15}^*$ $P_{16} * P_{17} * P_{18}$	2,5981E-16		
Etc...							



# Model

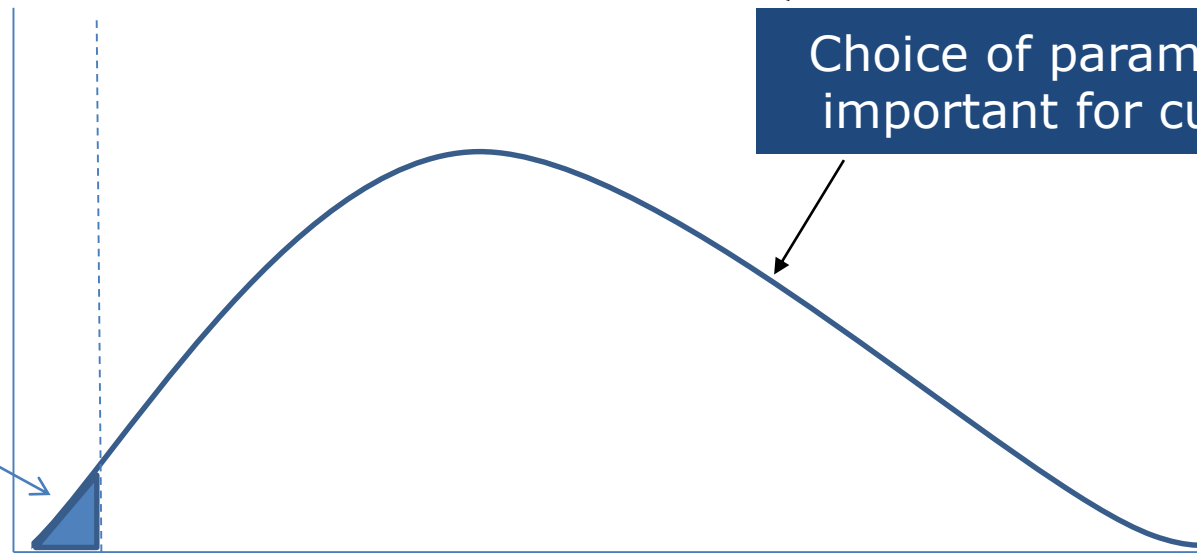
Assumption: Peak heights follow a Gamma distribution

Parameters: Expected peak height, peak height variation, mixture proportions, degradation (and stutter).

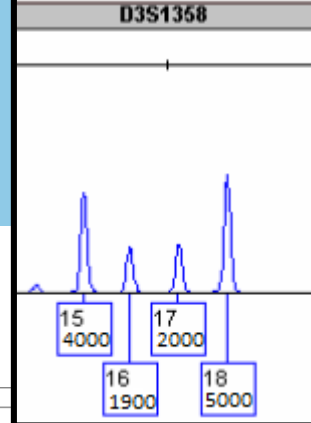
Probability  
density  
function (pdf)

Choice of parameters  
important for curve!

Drop out  
probability is  
area from 0 to  
AT

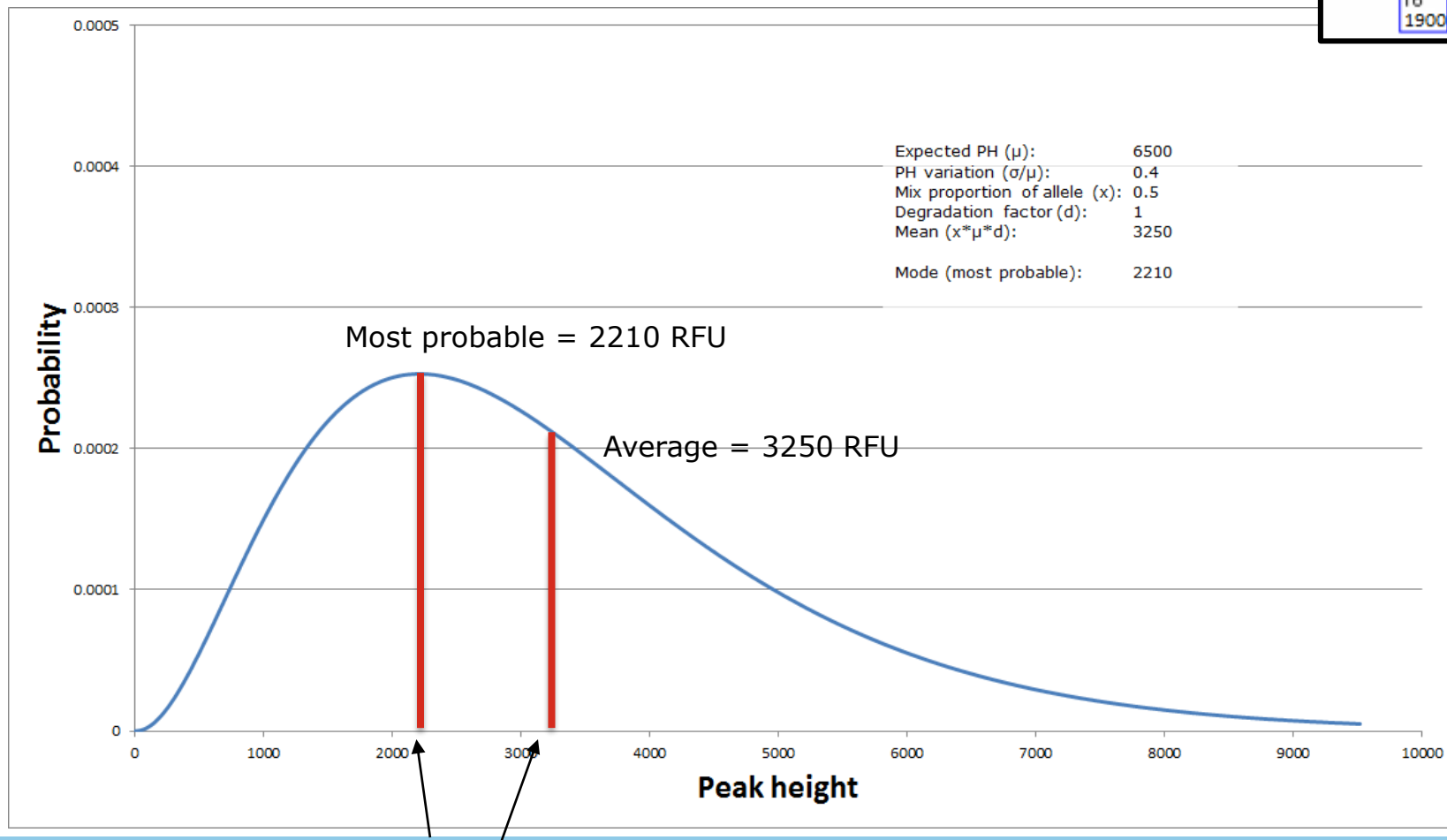


40 RFU analytical threshold (as an example)

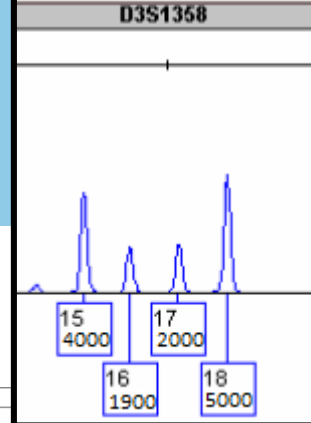


## Model

Peak height variation: 0.4  
Flat bump → More variation

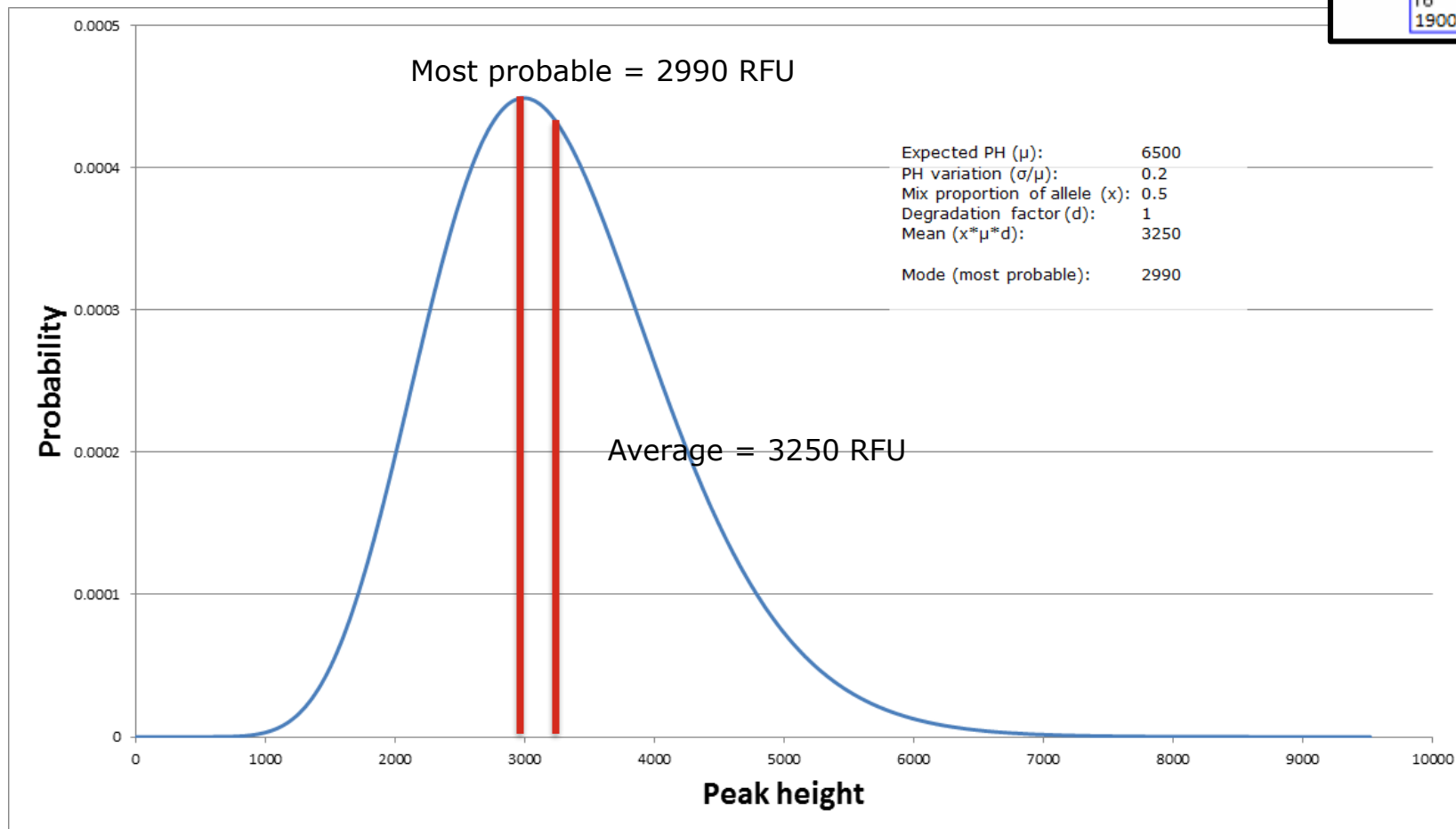


Observed alleles



# Model

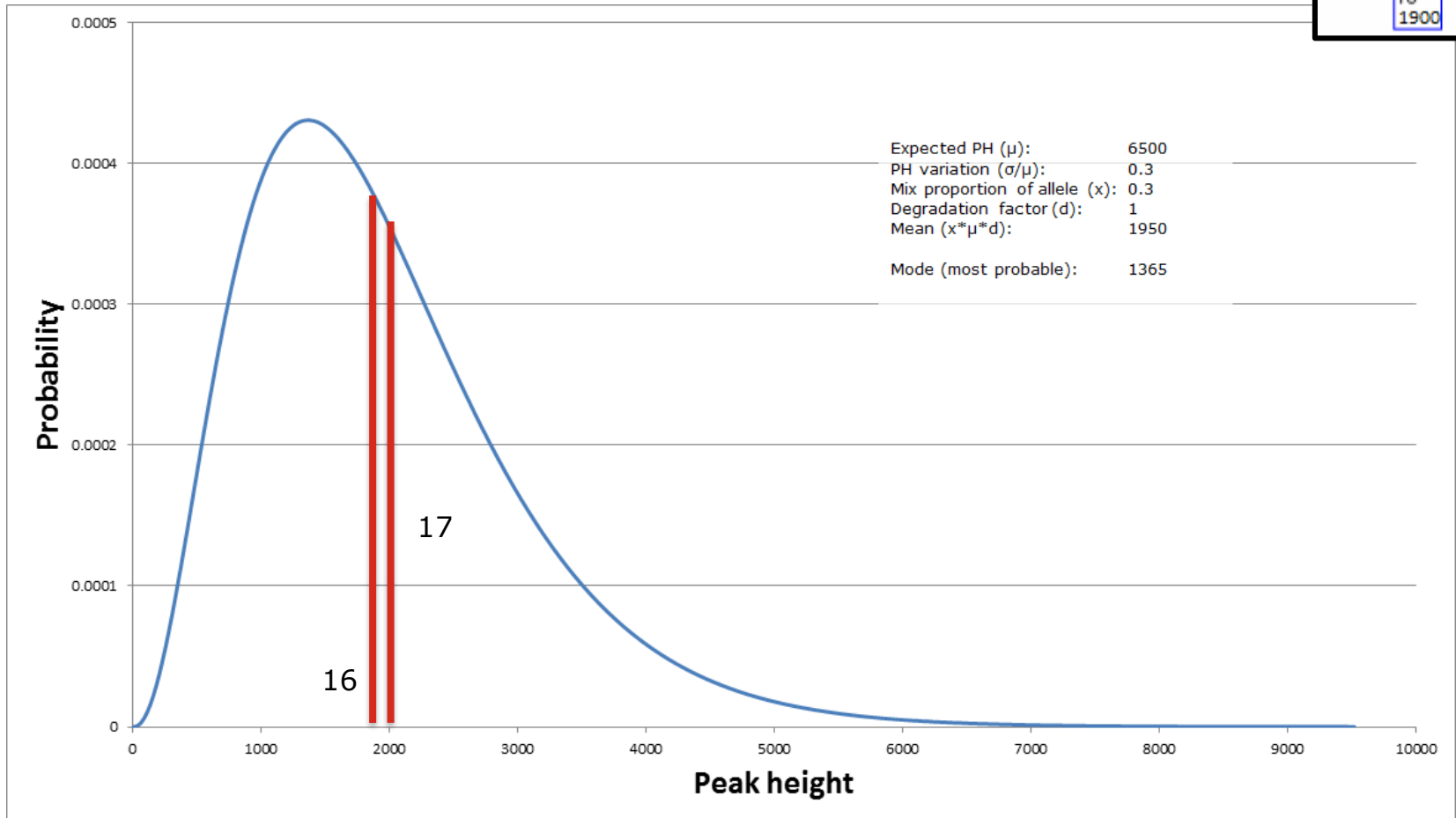
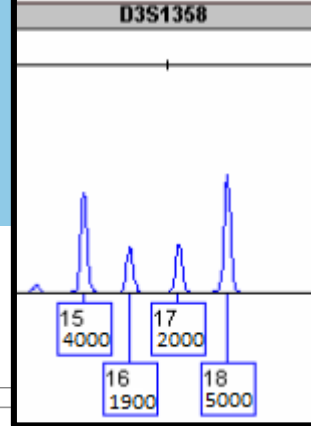
Peak height variation: 0.2  
Steep bump → Less variation





## Probable genotype combination (minor)

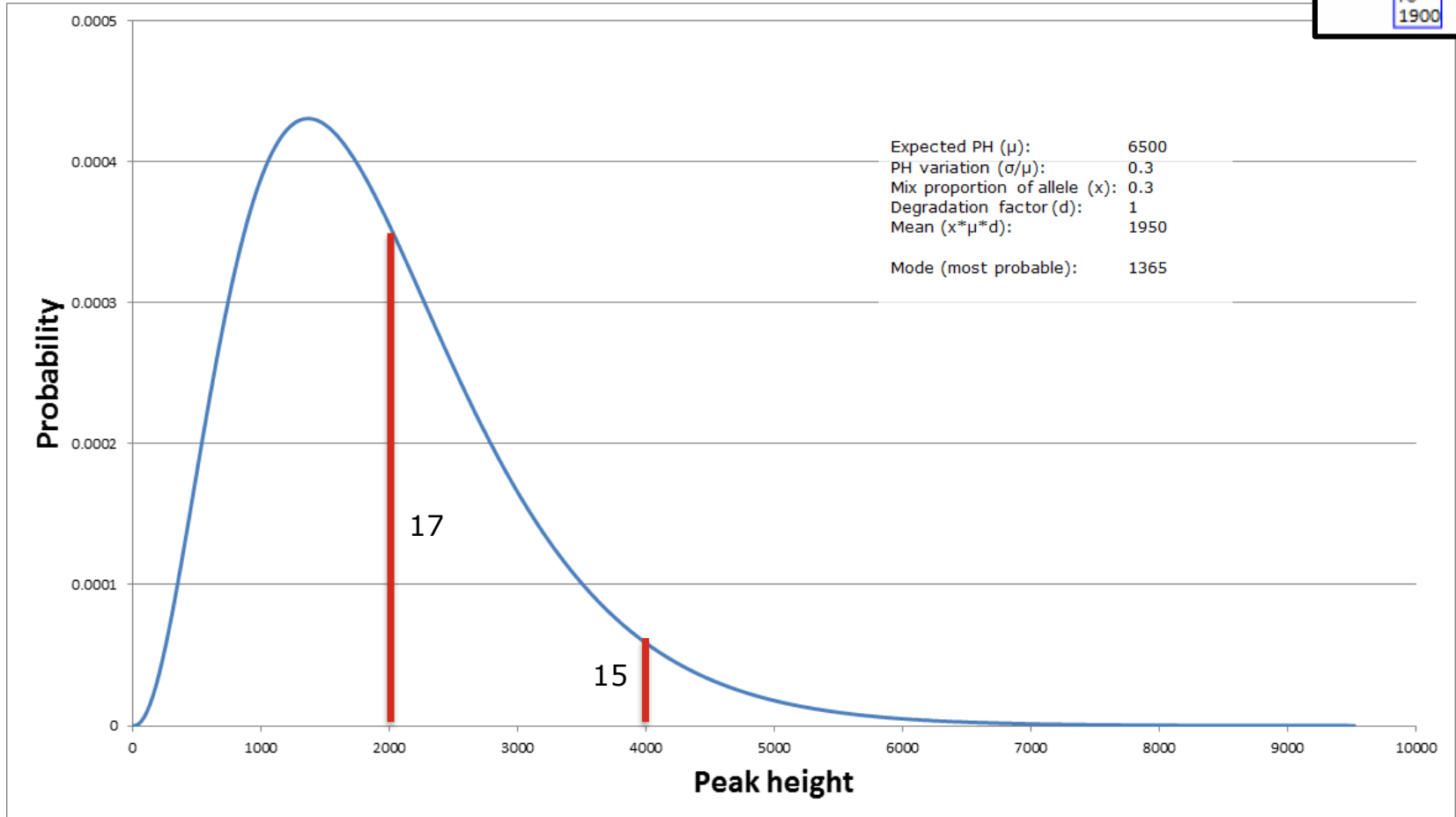
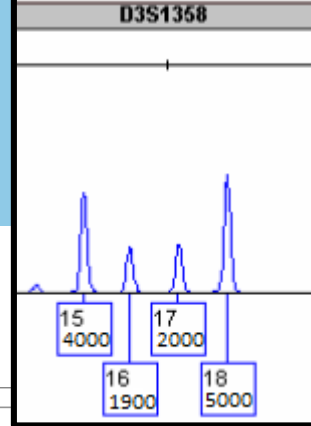
**C1 = 15,18 mixture proportion 0.7**  
**C2 = 16,17 mixture proportion 0.3**





## Less probable genotype combination (minor)

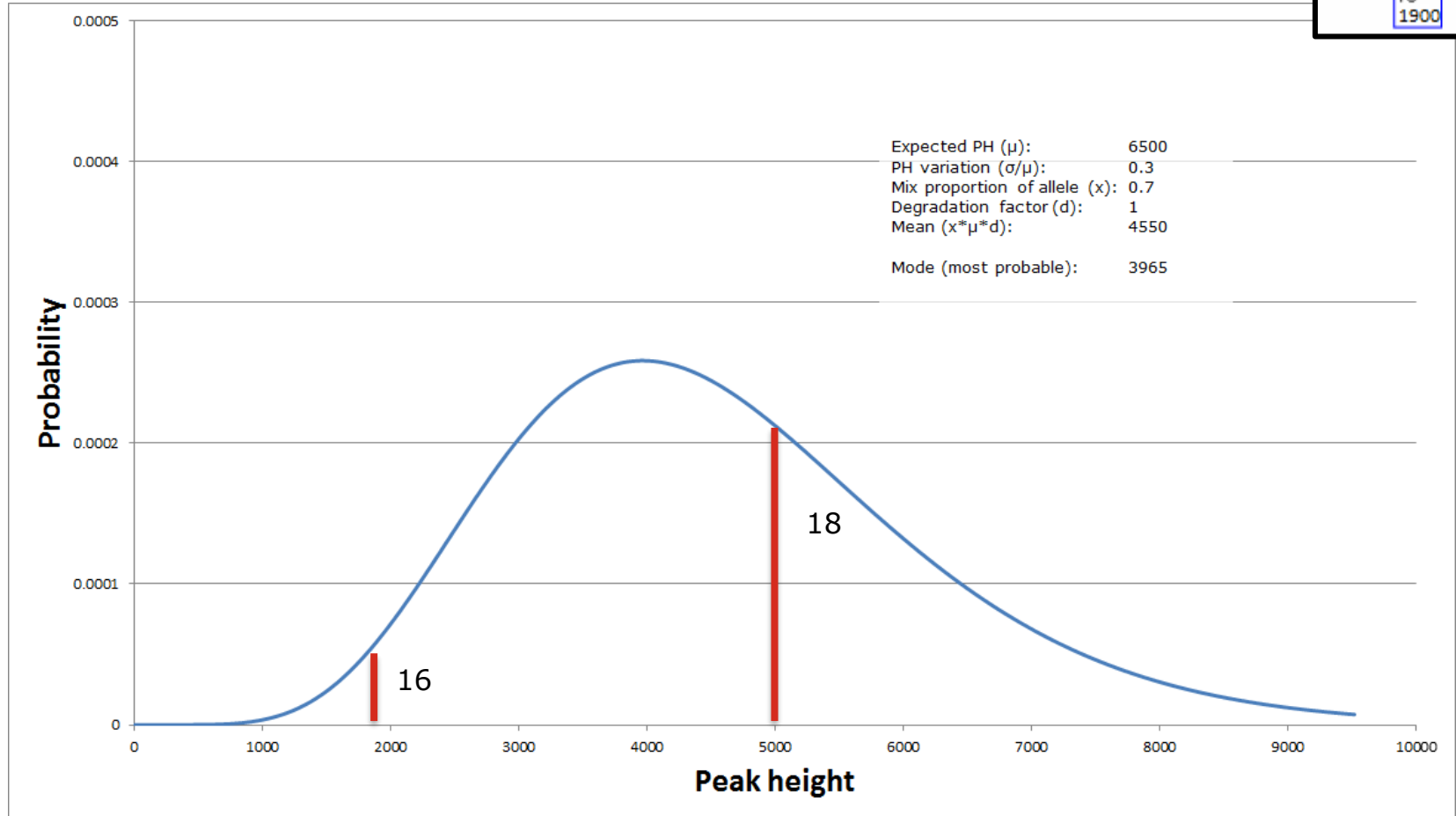
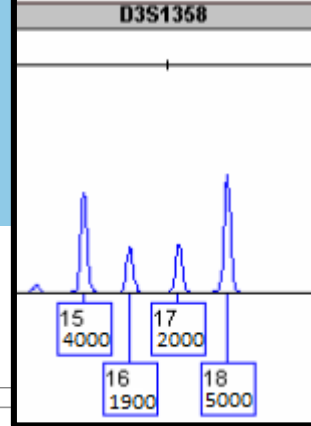
**C1 = 16,18 mixture proportion 0.7**  
**C2 = 15,17 mixture proportion 0.3**





## Less probable genotype combination (major)

**C1 = 16,18 mixture proportion 0.7**  
**C2 = 15,17 mixture proportion 0.3**



# The drop-out model for quantitative models



Netherlands Forensic Institute  
Ministry of Justice and Security

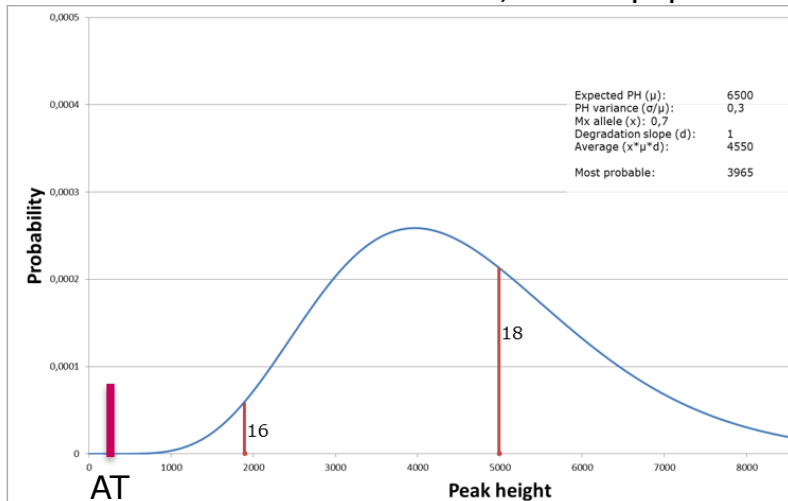
The sizes of the peak heights is important

- Large peak heights gives small drop-out probability
- Small peak heights gives high drop-out probability

The likelihood weight for the Q-allele =  $\Pr(\text{dropout})$

For Major

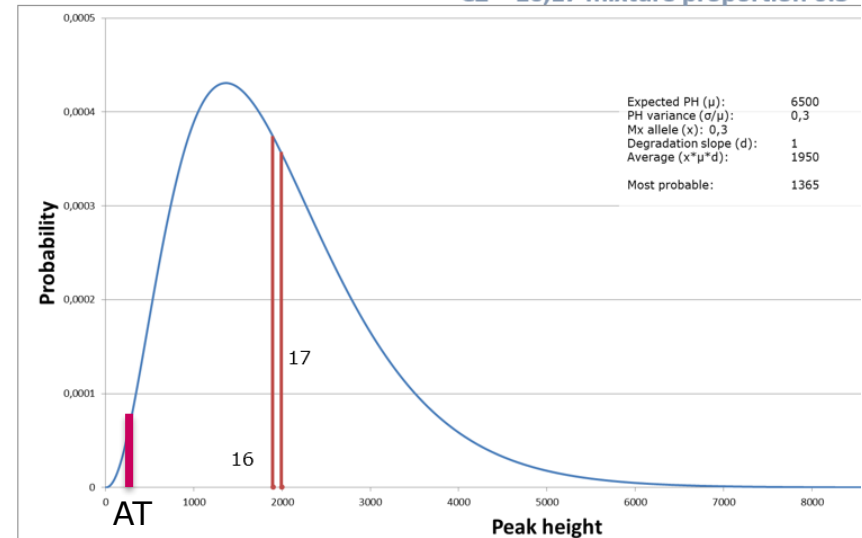
C1 = 16,18 mixture proportion 0.7  
C2 = 15,17 mixture proportion 0.3



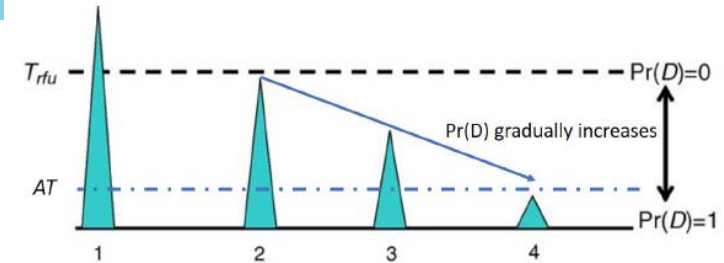
$\Pr(\text{dropout}) = 0$

For minor

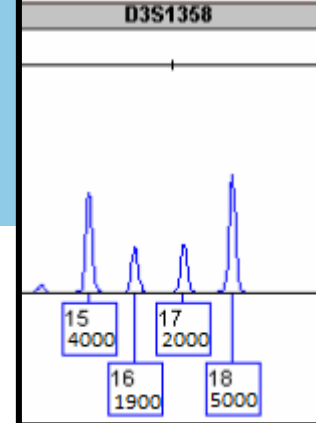
C1 = 15,18 mixture proportion 0.7  
C2 = 16,17 mixture proportion 0.3



$\Pr(\text{dropout}) > 0$

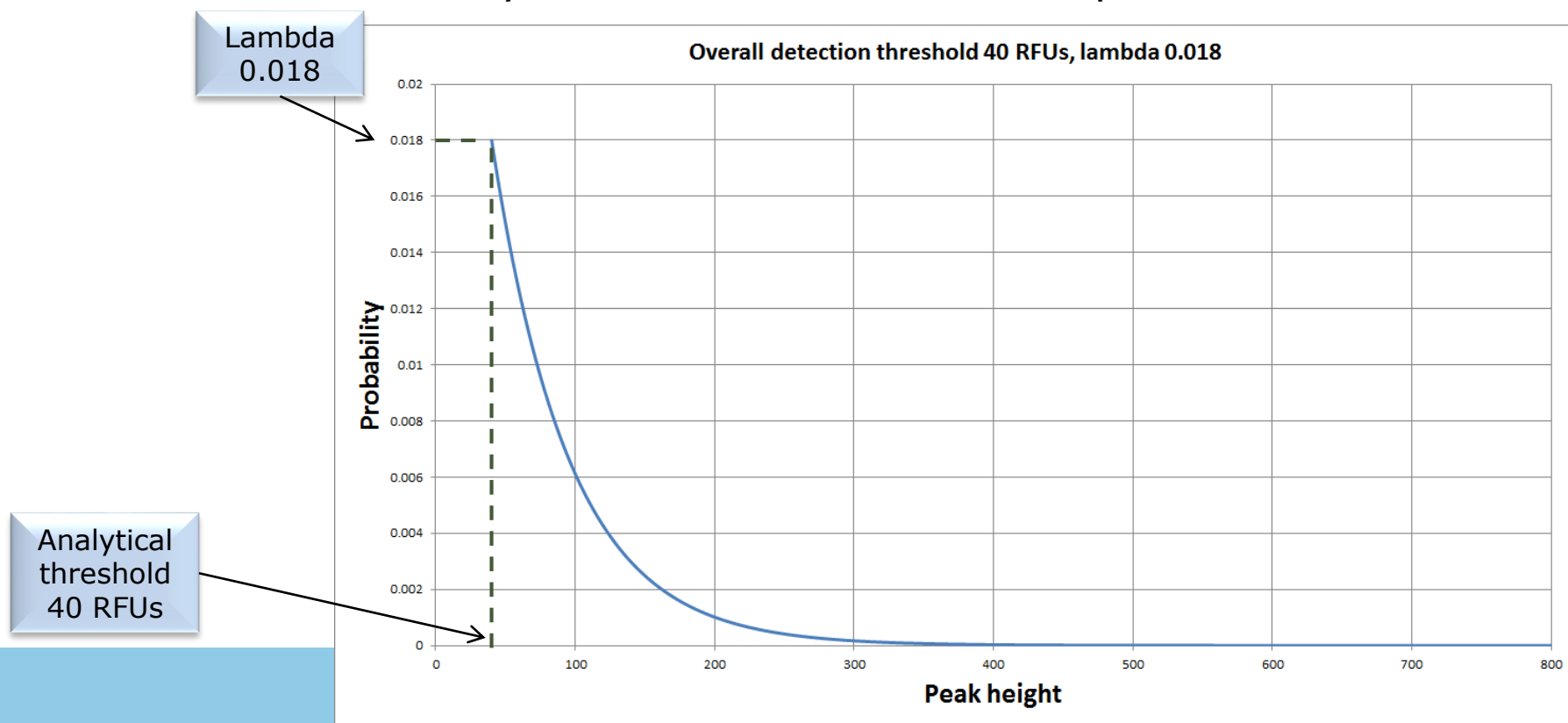


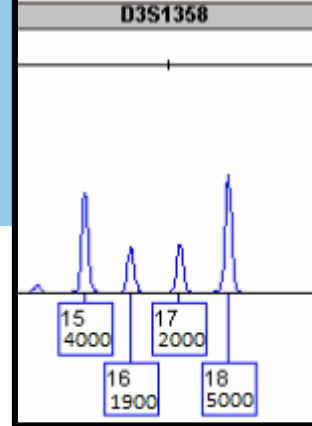




## Drop-in

- Model assumption:  
Drop-in peak heights follow an exponential distribution.
- Defined by lambda ( $\lambda$ ), which can be empirically determined from casework blanks. By default 0.01. In this example 0.018.





## Drop-in

- If allele 17 is a drop-in:
  - $PrC * P_{17} * ExpDistribution(Height_{17} - AT; 0.018)$  **Unlikely**
  - $0.00426257 * 0.22 * ExpDistribution(2000 - 40; 0.018) = 4.16E^{-19}$
- If allele 17 has a peak height of 80 RFUs instead: **More likely**
  - $0.00426257 * 0.22 * ExpDistribution(80 - 40; 0.018) = 4.56E^{-4}$

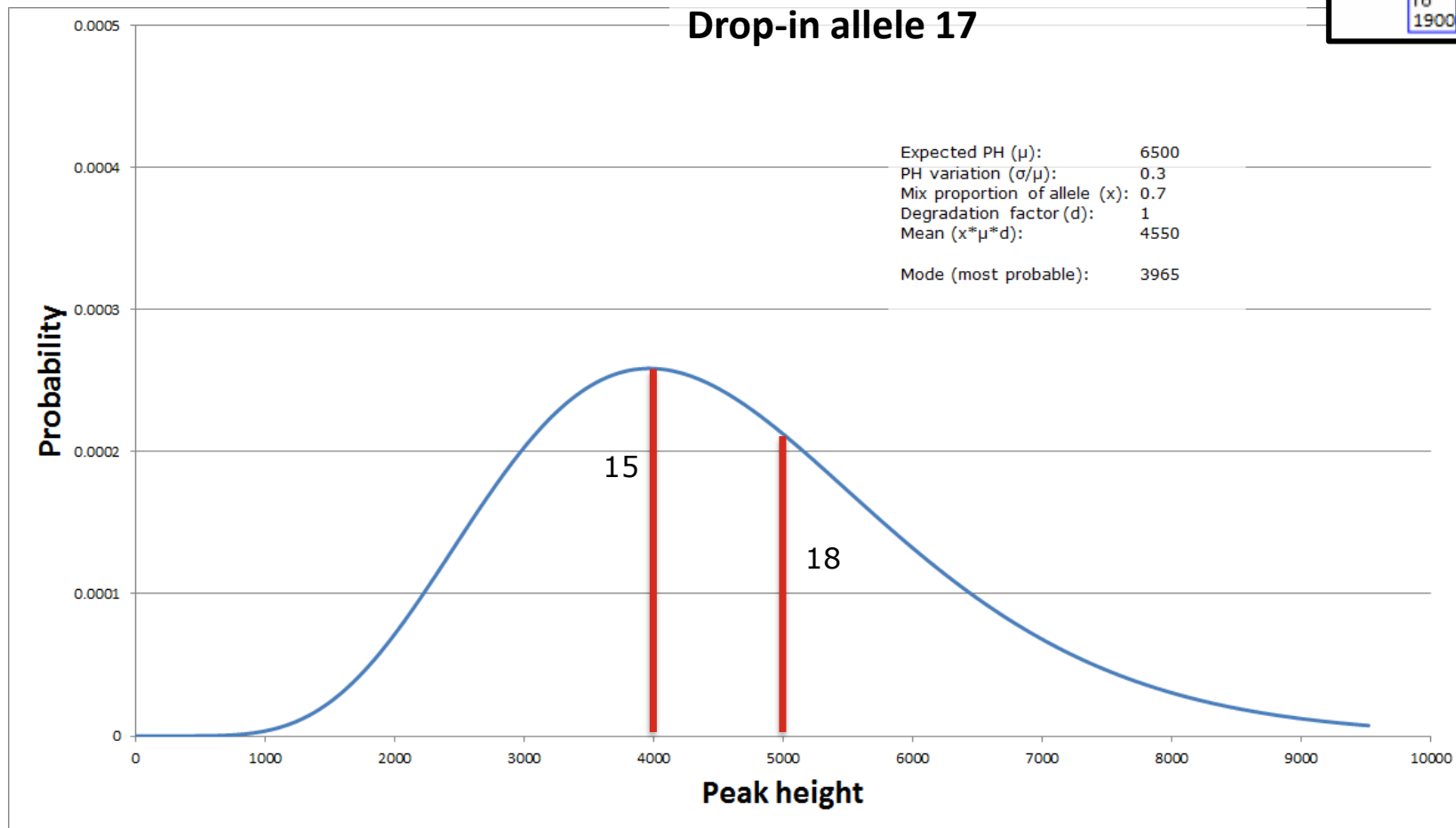
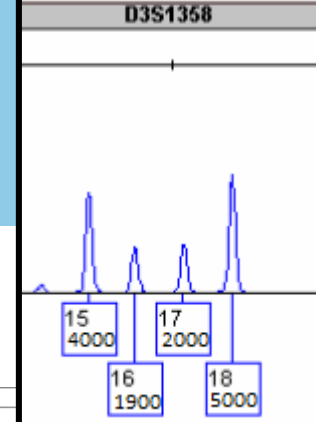


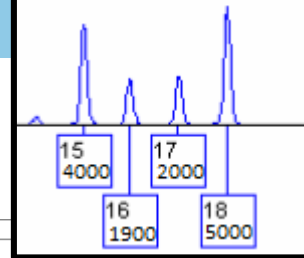
## Probable genotype combination (for major)

**C1 = 15,18 mixture proportion 0.7**

**C2 = 16,16 mixture proportion 0.3**

**Drop-in allele 17**



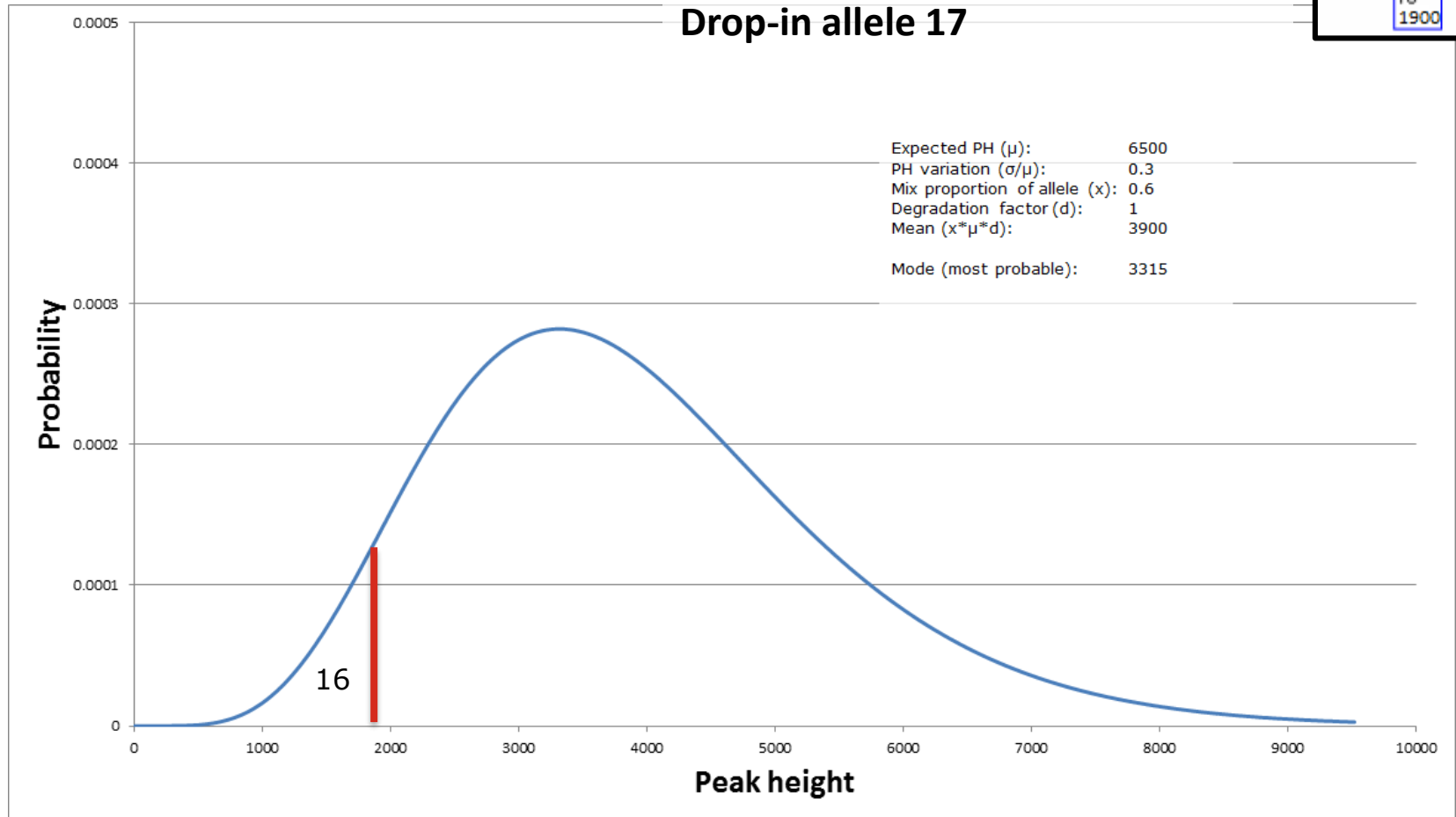


But less probable genotype combination for the minor

**C1 = 15,18 mixture proportion 0.7**

**C2 = 16,16 mixture proportion 0.3**

**Drop-in allele 17**





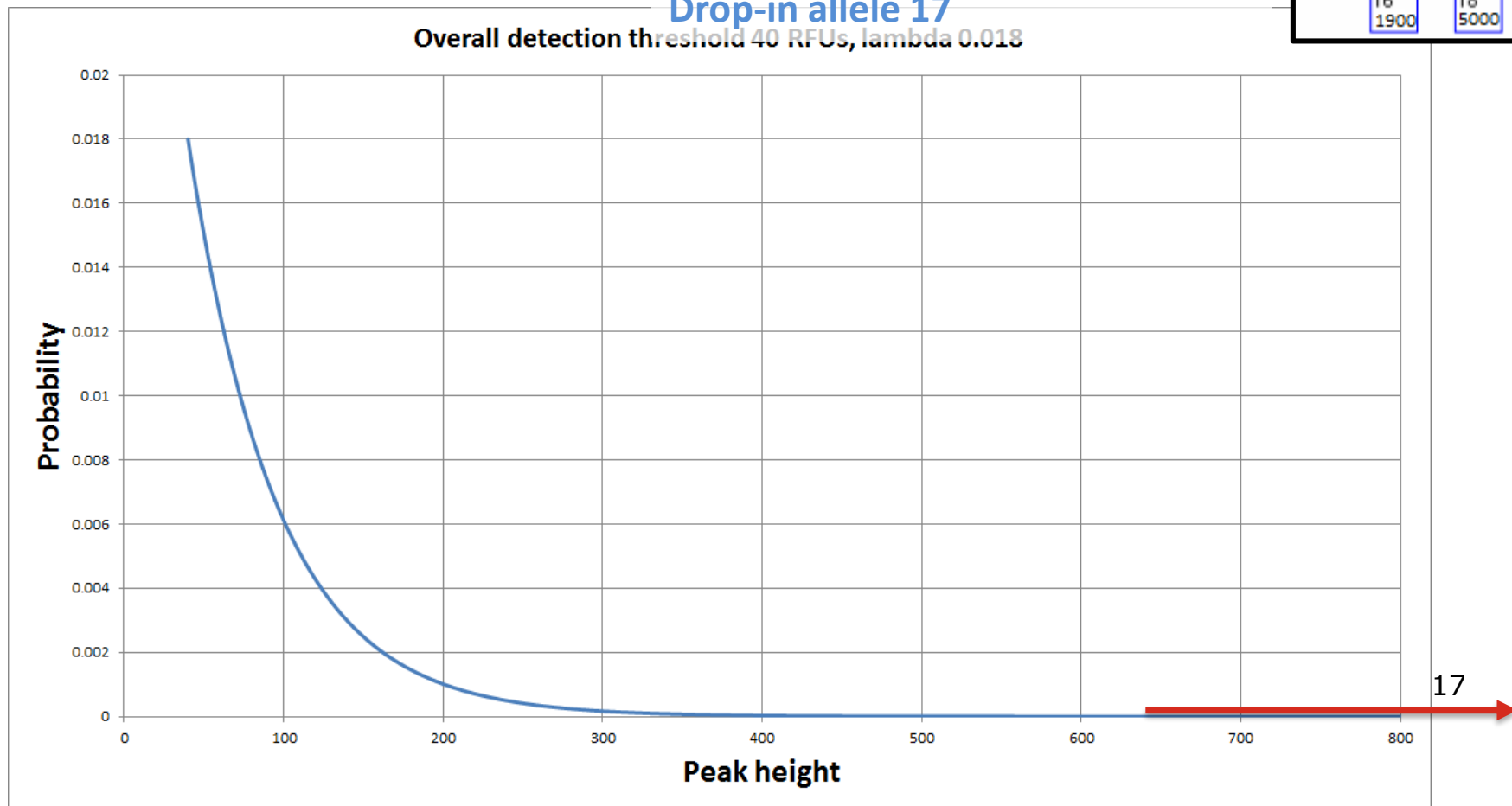
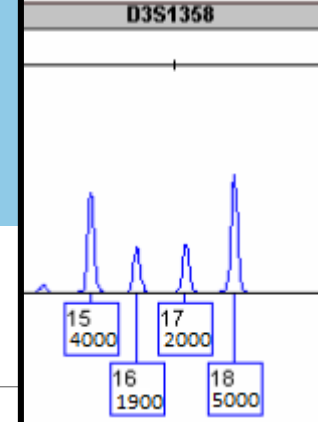
Very unlikely for drop-in

C1 = 15,18 mixture proportion 0.7

C2 = 16,16 mixture proportion 0.3

Drop-in allele 17

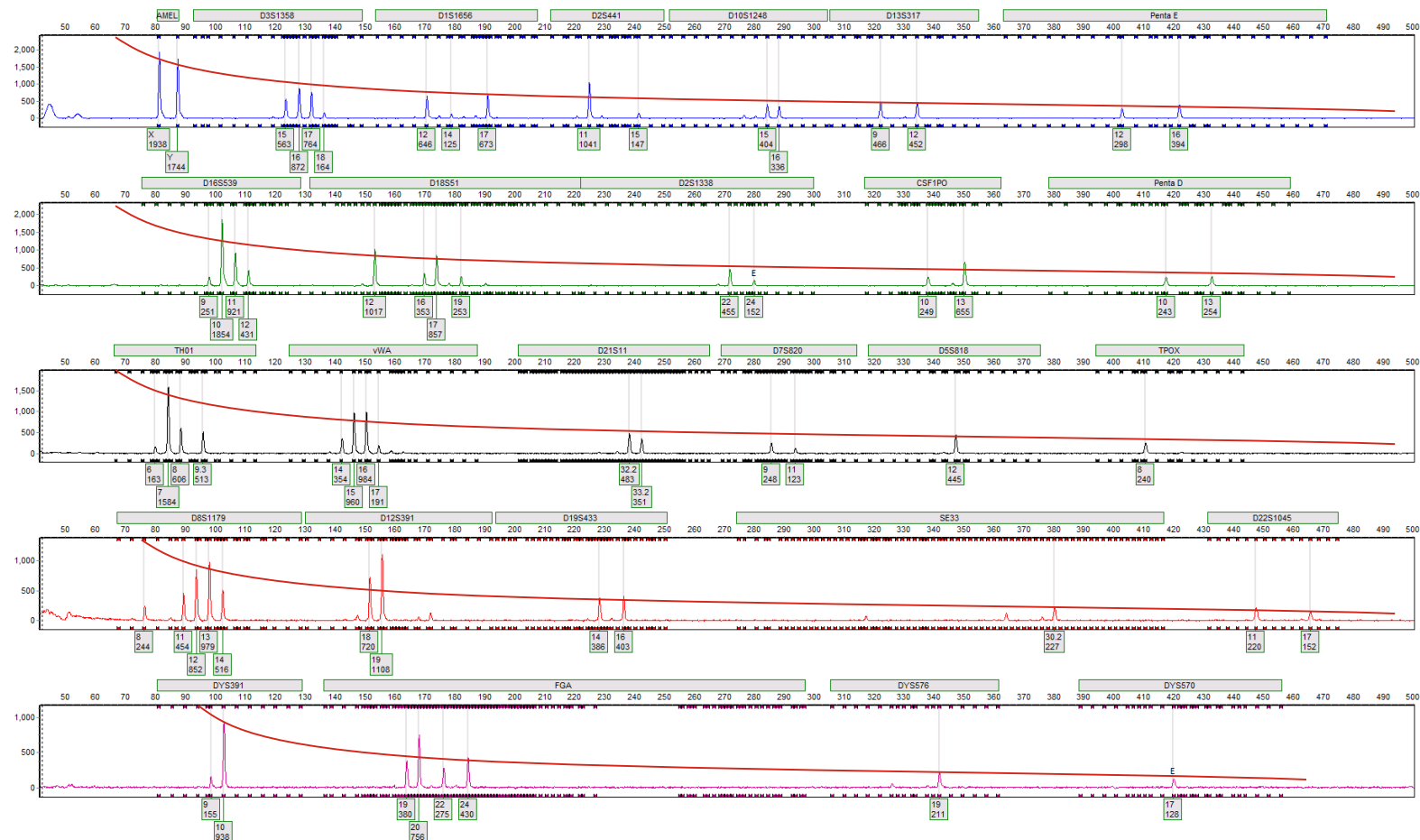
Overall detection threshold 40 RFUs, lambda 0.018





# Degradation

$$\beta \frac{f_{m,a} - 125}{100}$$

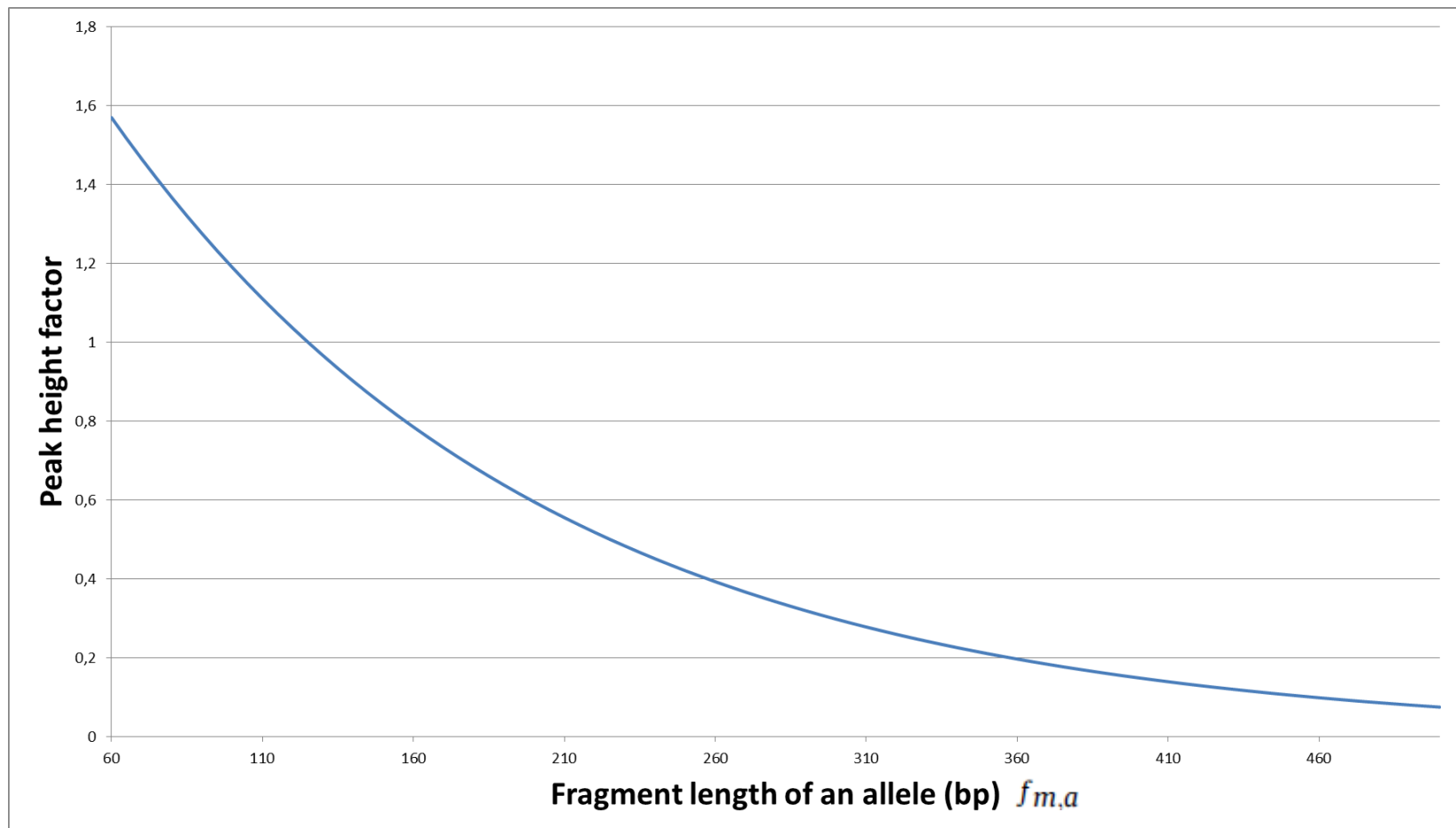




# Degradation model

$\beta=0.5$

$$\beta^{\frac{f_{m,a}-125}{100}}$$

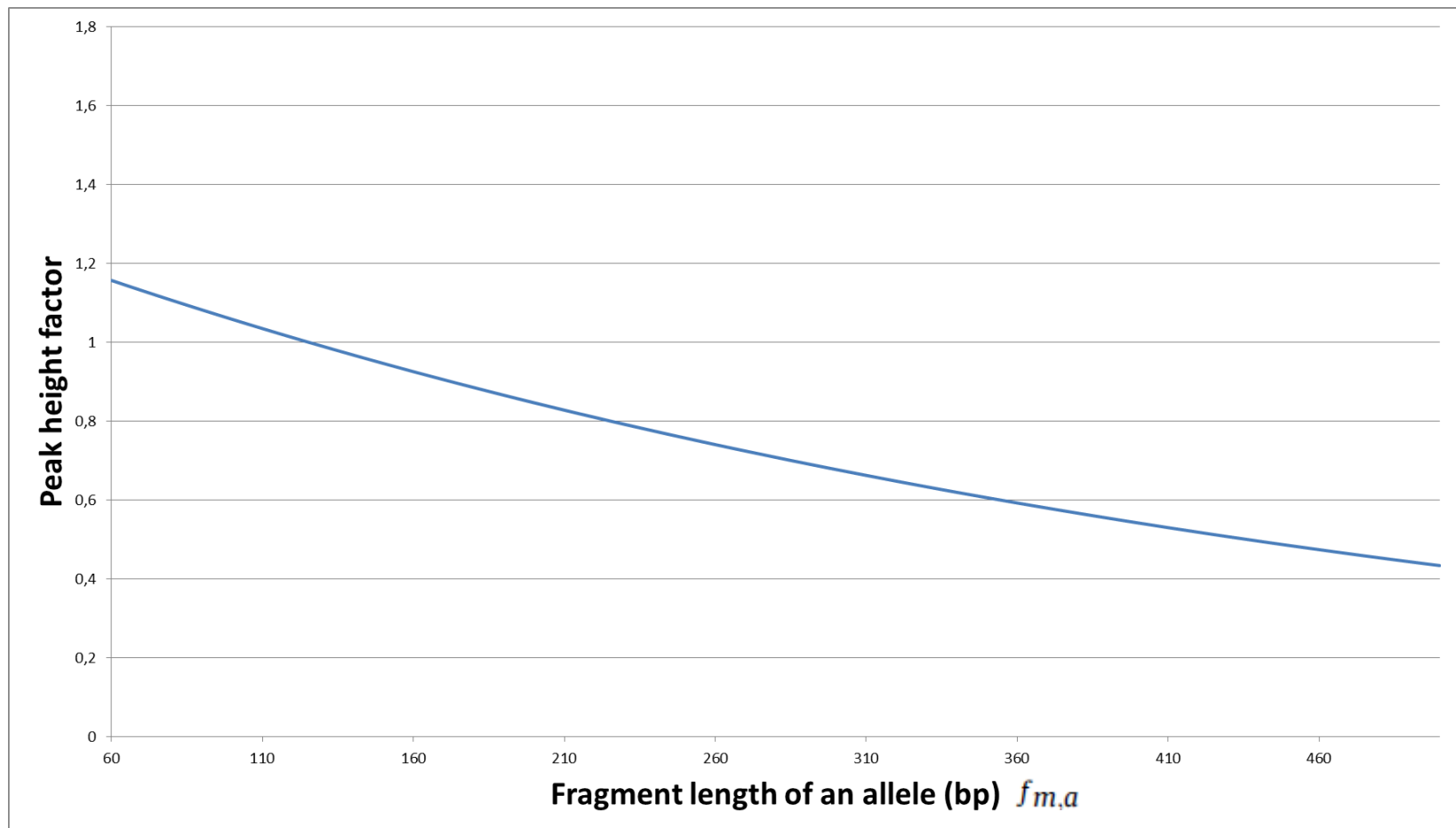




# Degradation model

$\beta=0.8$

$$\beta^{\frac{f_{m,a}-125}{100}}$$



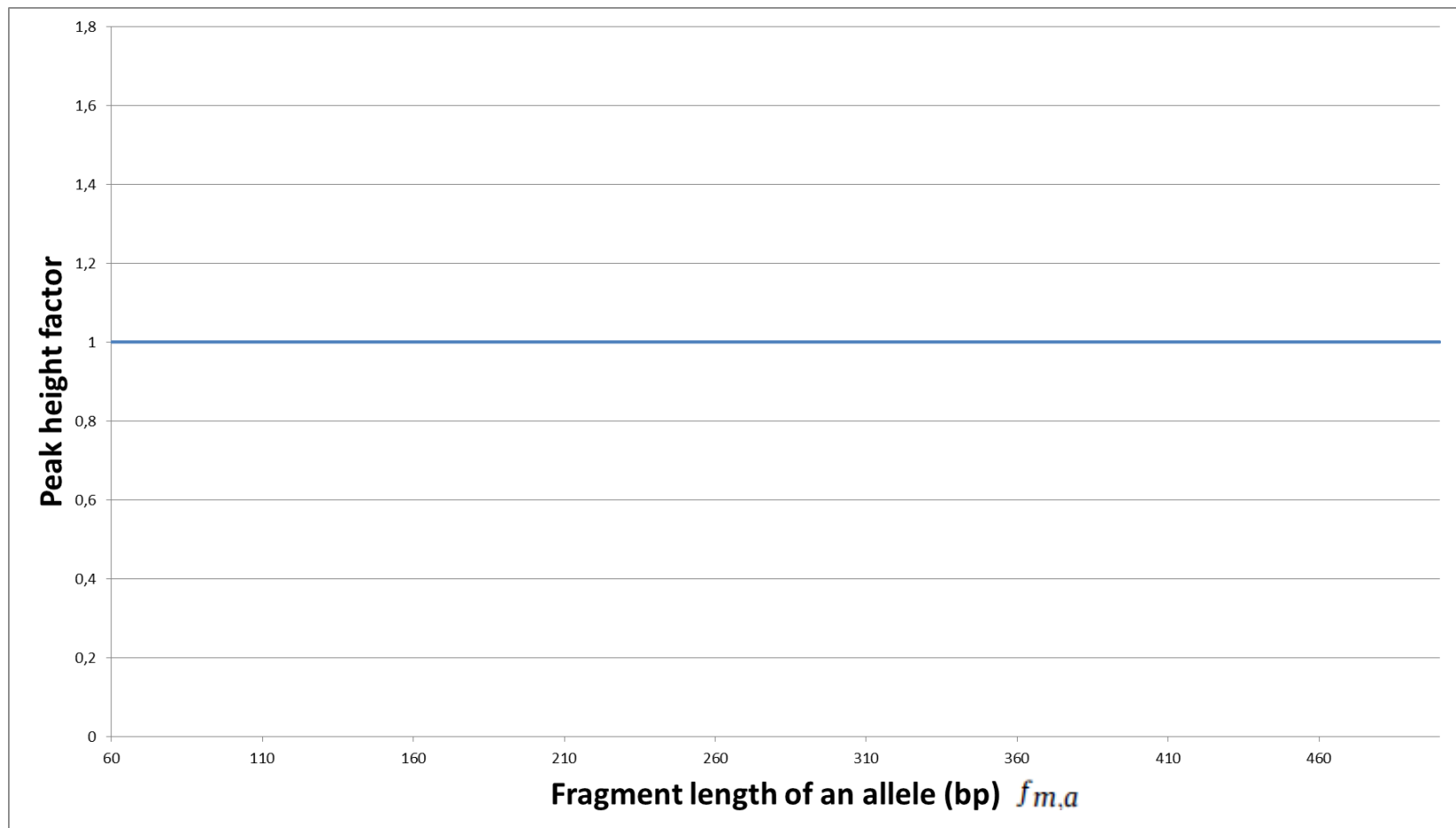




# Degradation model

$$\beta = 1$$

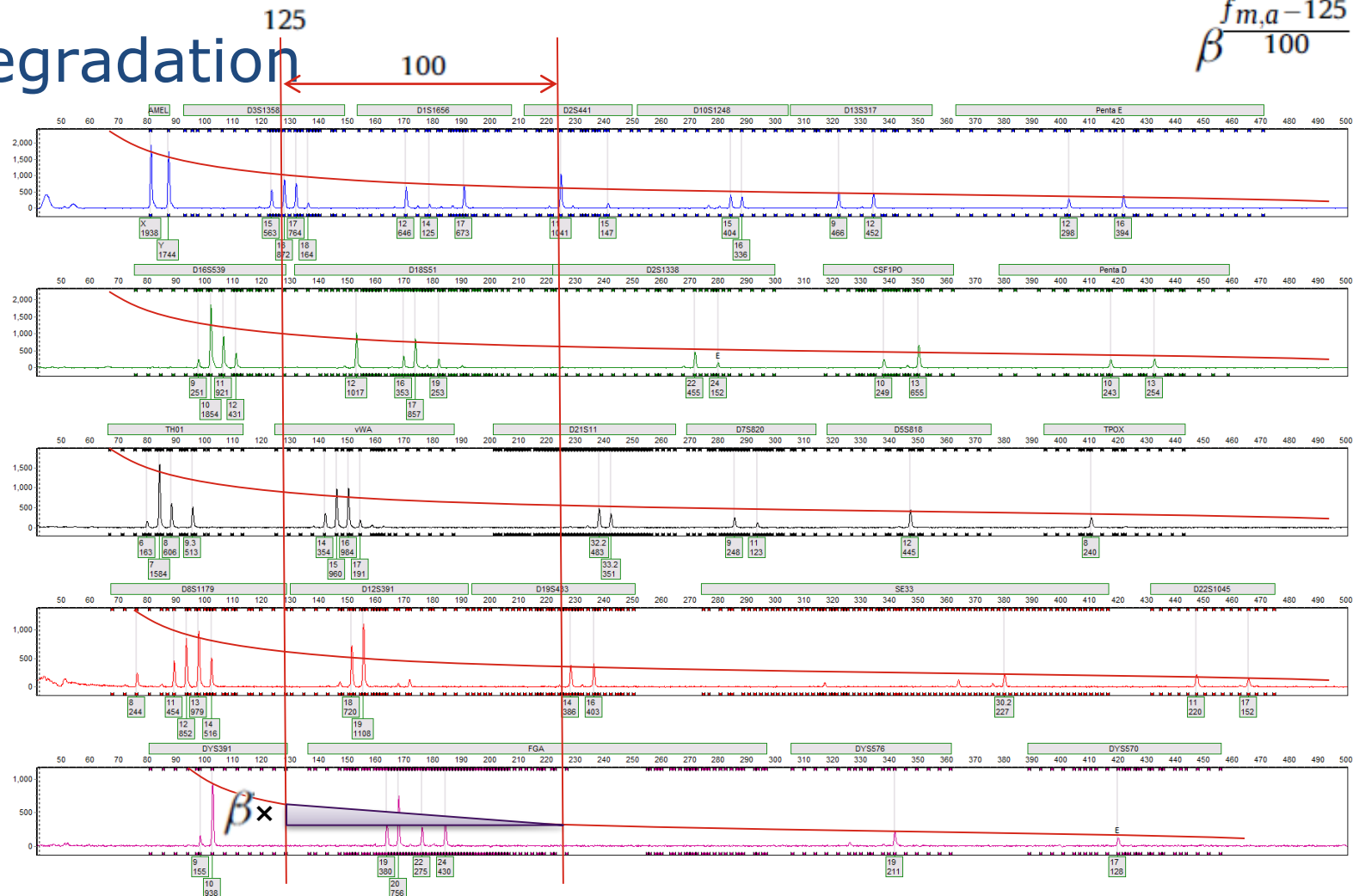
$$\beta^{\frac{f_{m,a} - 125}{100}}$$





# Degradation

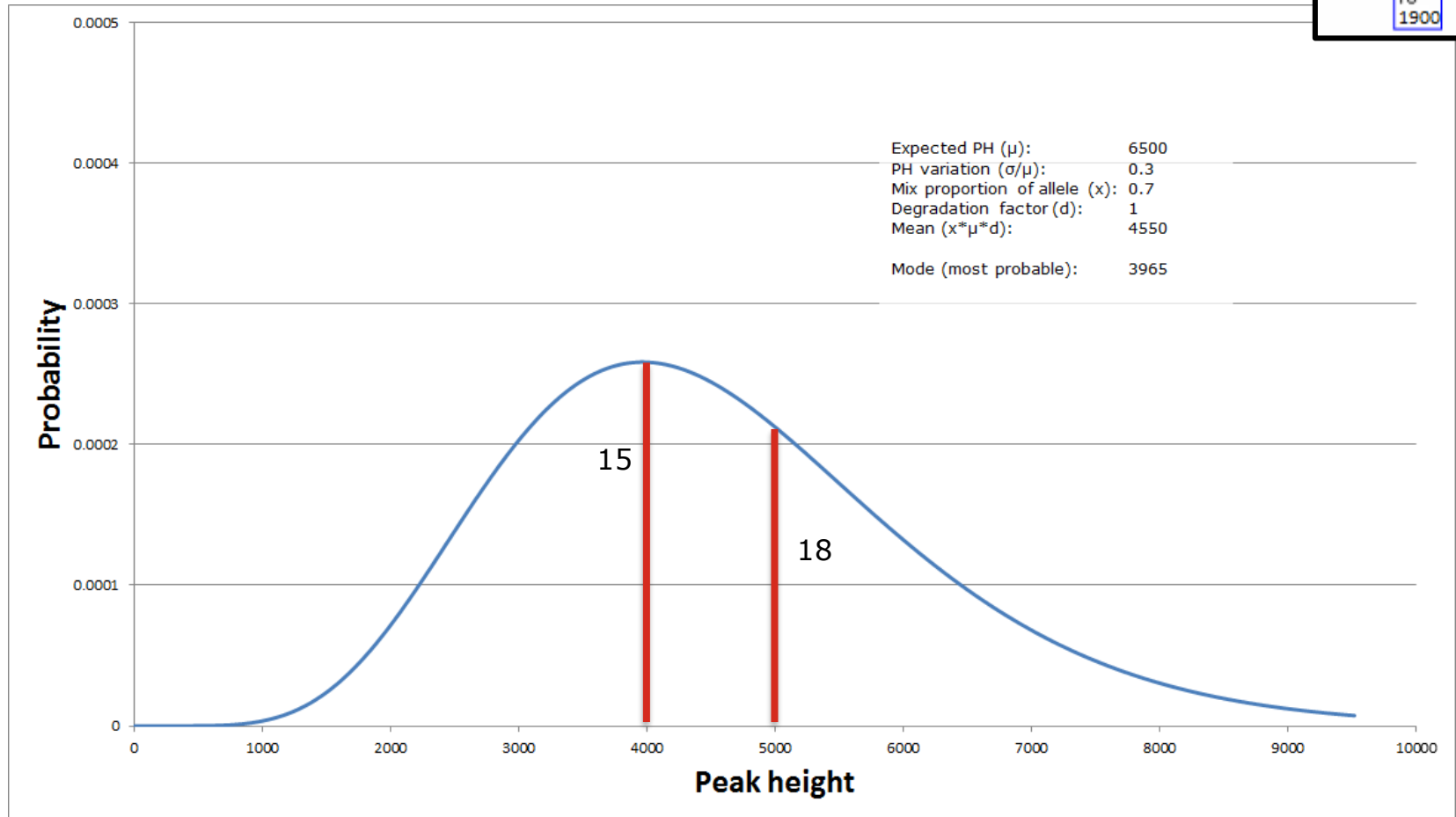
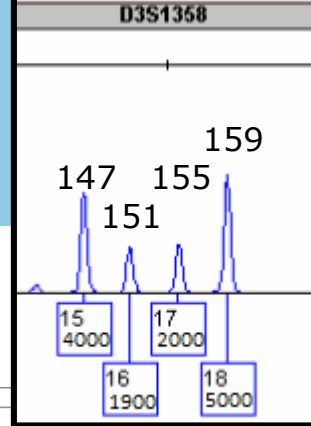
$$\beta \frac{f_{m,a} - 125}{100}$$





Probable?  $\beta=1$

C1 = 15,18 mixture proportion 0.7  
C2 = 16,17 mixture proportion 0.3

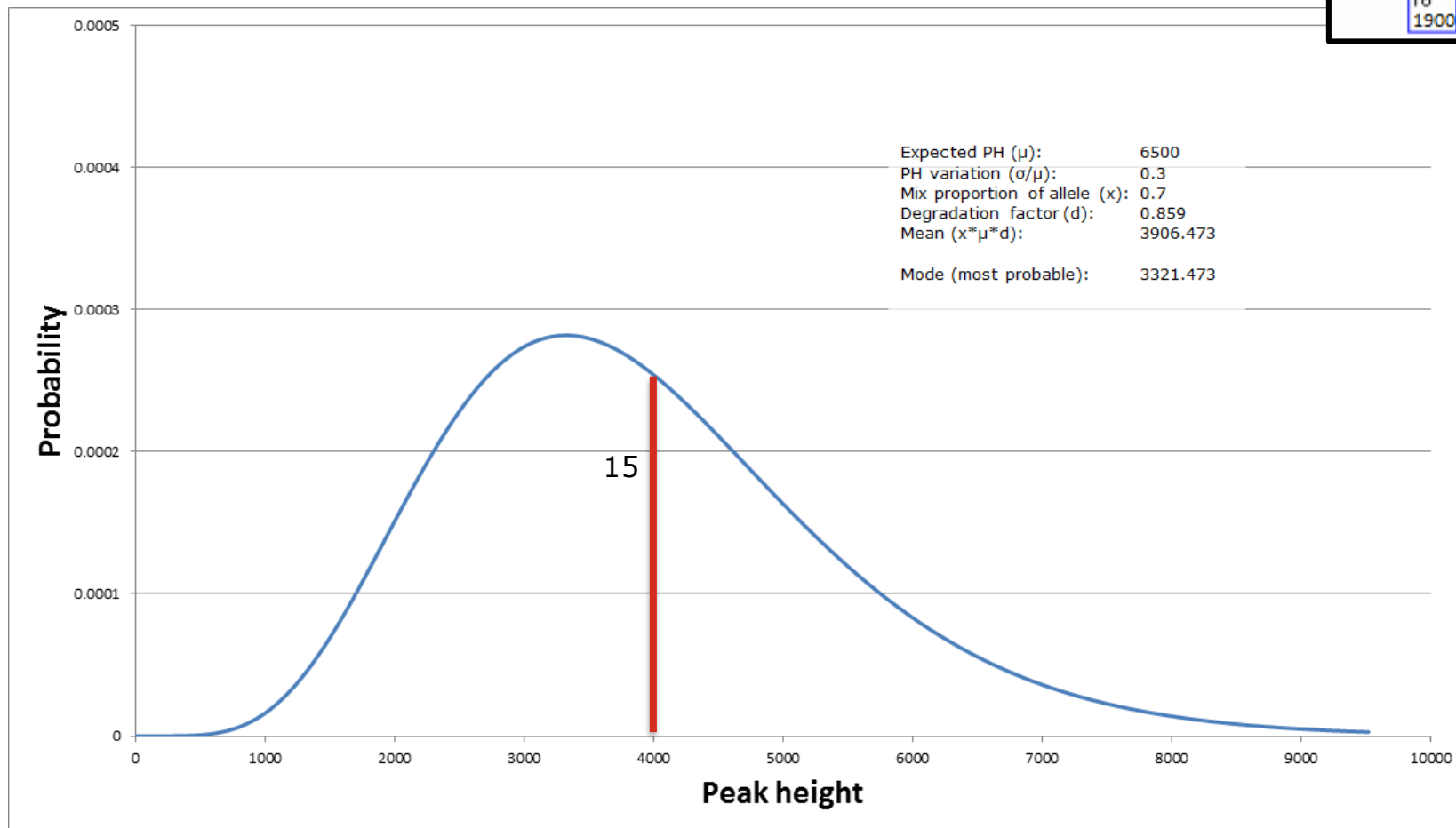
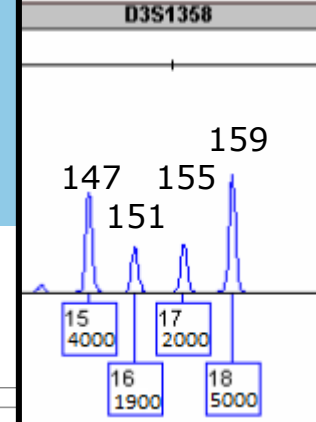




Probable?  $\beta=0.5$

C1 = 15,18 mixture proportion 0.7

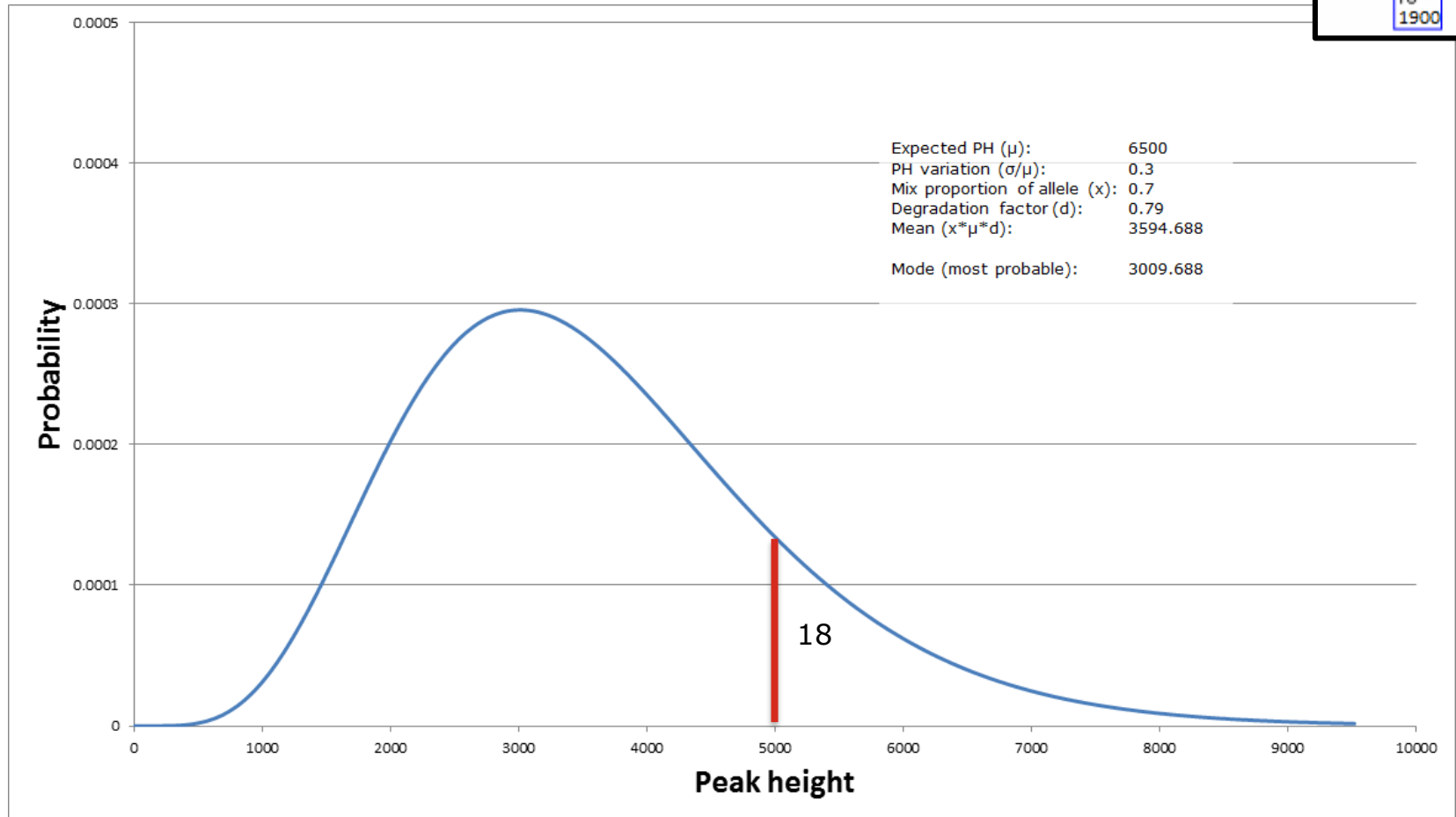
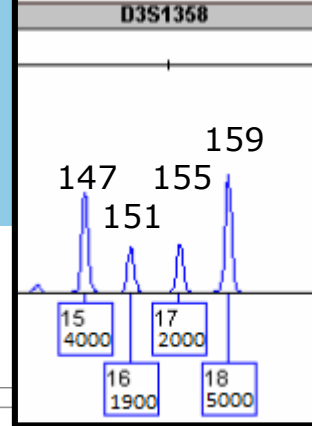
C2 = 16,17 mixture proportion 0.3

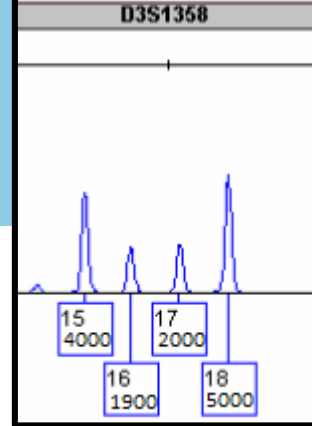




Probable?  $\beta=0.5$

C1 = 15,18 mixture proportion 0.7  
C2 = 16,17 mixture proportion 0.3



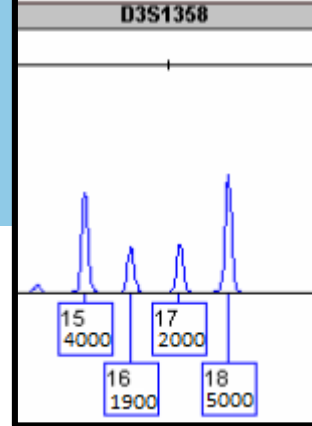


## Calculate likelihoods including PH model

➤ Sum up product within all genotype combinations

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability	Likelihood incl PH
15/16	17/18	-	$2P_{15}P_{16} * 2P_{17}P_{18}$	1-PrC	2.6944E-52
15/17	16/18	-	$2P_{15}P_{17} * 2P_{16}P_{18}$	1-PrC	2.5123E-50
15/18	16/17	-	$2P_{15}P_{18} * 2P_{16}P_{17}$	1-PrC	3.8438E-15
17/18	15/16	-	$2P_{17}P_{18} * 2P_{15}P_{16}$	1-PrC	9.3019E-42
16/18	15/17	-	$2P_{16}P_{18} * 2P_{15}P_{17}$	1-PrC	9.9761E-44
16/17	15/18	-	$2P_{16}P_{17} * 2P_{15}P_{18}$	1-PrC	6.5203E-79
But also...					
15/15	16/17	18	$P_{15}^2 * 2P_{16}P_{17}$	$PrC * P_{18}$	1.5107E-89
15/18	16/d.o.	17	$2P_{15}P_{18} * 2P_{16}P_Q$	$PrC * P_{17}$	1.4539E-117
d.o.	d.o.	15/16/17/18	$P_Q^4$	$PrC^4 * P_{15} * P_{16} * P_{17} * P_{18}$	0
Etc...					

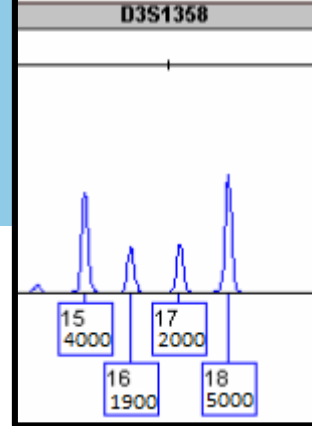
3.0378E-112	1.4539E-117
0	0
	+
	3.8438E-15



## Calculate likelihoods multiple replicates

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability	Likelihood incl PH Replicate 1
15/16	17/18	-	$2P_{15}P_{16} * 2P_{17}P_{18}$	1-PrC	2.6944E-52
15/17	16/18	-	$2P_{15}P_{17} * 2P_{16}P_{18}$	1-PrC	2.5123E-50
15/18	16/17	-	$2P_{15}P_{18} * 2P_{16}P_{17}$	1-PrC	3.8438E-15
17/18	15/16	-	$2P_{17}P_{18} * 2P_{15}P_{16}$	1-PrC	9.3019E-42
16/18	15/17	-	$2P_{16}P_{18} * 2P_{15}P_{17}$	1-PrC	9.9761E-44
16/17	15/18	-	$2P_{16}P_{17} * 2P_{15}P_{18}$	1-PrC	6.5203E-79
But also...					
15/15	16/17	18	$P_{15}^2 * 2P_{16}P_{17}$	PrC* $P_{18}$	1.5107E-89
15/18	16/d.o.	17	$2P_{15}P_{18} * 2P_{16}P_Q$	PrC* $P_{17}$	1.4539E-117
d.o.	d.o.	15/16/17/18	$P_Q^4$	PrC <sup>4</sup> * $P_{15} * P_{16} * P_{17} * P_{18}$	0
Etc...					

3.0378E-112	1.4539E-117
0	0
	+
	3.8438E-15



## Calculate profile likelihood

Donor A	Donor B	Drop-in	Genotype probability	Drop-in probability	Likelihood incl PH Replicate 1	Likelihood incl PH Replicate 2	Likelihood
15/16	17/18	-	$2P_{15}P_{16} * 2P_{17}P_{18}$	1-PrC	2.6944E-52	2.6475E-45	7.1334E-97
15/17	16/18	-	$2P_{15}P_{17} * 2P_{16}P_{18}$	1-PrC	2.5123E-50	2.8226E-46	7.0912E-96
15/18	16/17	-	$2P_{15}P_{18} * 2P_{16}P_{17}$	1-PrC	3.8438E-15	1.2048E-14	4.6310E-29
17/18	15/16	-	$2P_{17}P_{18} * 2P_{15}P_{16}$	1-PrC	9.3019E-42	2.9156E-41	2.7121E-82
16/18	15/17	-	$2P_{16}P_{18} * 2P_{15}P_{17}$	1-PrC	9.9761E-44	2.7347E-40	2.7282E-83
16/17	15/18	-	$2P_{16}P_{17} * 2P_{15}P_{18}$	1-PrC	6.5203E-79	6.4067E-72	4.1774E-150
But also...							
15/15	16/17	18	$P_{15}^2 * 2P_{16}P_{17}$	PrC* $P_{18}$	1.5107E-89	1.4552E-83	2.1984E-172
15/18	16/d.o.	17	$2P_{15}P_{18} * 2P_{16}P_Q$	PrC* $P_{17}$	1.4539E-117	2.9109E-115	4.2322E-232
d.o.	d.o.	15/16/17/18	$P_Q^4$	PrC <sup>4</sup> * $P_{15}^*$ $P_{16}^*P_{17}^*P_{18}^*$	0	0	0
Etc...							+
							4.6311E-29

### ➤ Multiply between loci

Locus 1	Locus 2	Locus 3	Locus 4	Locus 5	Locus 6	Locus n	Overall likelihood Hd
4.6311E-29 X	..... X	..... X	..... X	..... X	..... X	.....	$P(E H, \beta)$



## General formula for probabilistic genotyping models

Product rule

Summing over all possible  
unknown genotypes

$$LR = \frac{P(E|H_p, \beta_p)}{P(E|H_d, \beta_d)} = \frac{\prod_m \sum_g w(E_m, g | \beta_p) P(g | H_p)}{\prod_m \sum_g w(E_m, g | \beta_p) P(g | H_p)}$$

$m$  = marker index

$g$  = genotype combination (for marker  $m$ )

$\beta_p, \beta_d$  unknown model parameters

Must be estimated

$w$  are the likelihood weights (product over all alleles)



## A lot of calculations...

### 2 person mixture:

$(15 + 225 \text{ genotypes}) * (23 \text{ loci}) * \pm 5.000 \text{ optimization steps}$   
 $= 27.600.000 \text{ LE calculations} \rightarrow \text{less than a minute}$

### 4 person mixture:

$(91.125 + 4.100.625 \text{ genotypes}) * (23 \text{ loci}) * \pm 5.000 \text{ optimization steps}$   
 $= 482.051.250.000 \text{ LE calculations} \rightarrow \text{several days (a day)}$

### 5 person mixture:

$(4.100.625 + 1.252.332.576) * (23 \text{ loci}) * \pm 5.000 \text{ optimization steps}$   
 $= 144.489.818.115.000 \text{ LE calculations} \rightarrow \text{takes a long time...}$

*Calculation time is much reduced  
in the latest versions of  
EuroForMix and DNASTatistX! 😊*



## Exploratory approach

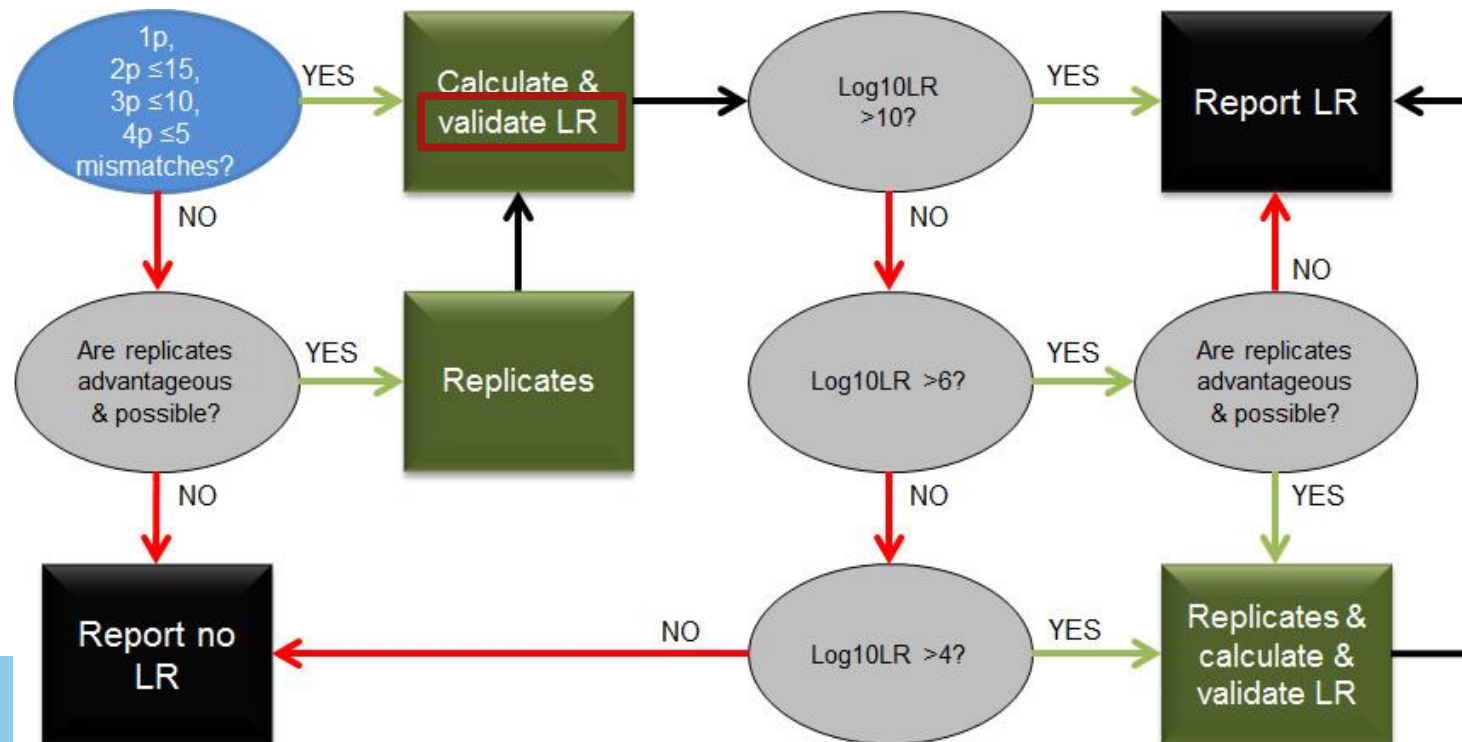
- Preferred approach is always to maximize the quality of the sample first
- Statistical tool is not a substitute for sound bio-chemistry
- The LR does not tell you if a proposition is true or not – it can only tell you if one is more likely than the other
- Not a substitute to careful consideration of the circumstances of the case to help formulate the propositions
- Rapid evaluation of multiple sets of propositions is facilitated by software



# Guidelines for use in forensic casework

- Based on results from research, casework experience, including efficiency and usefulness of calculations.
- Clearly, the case circumstances, availability of other stains and profiles within the case are factors considered in the process.

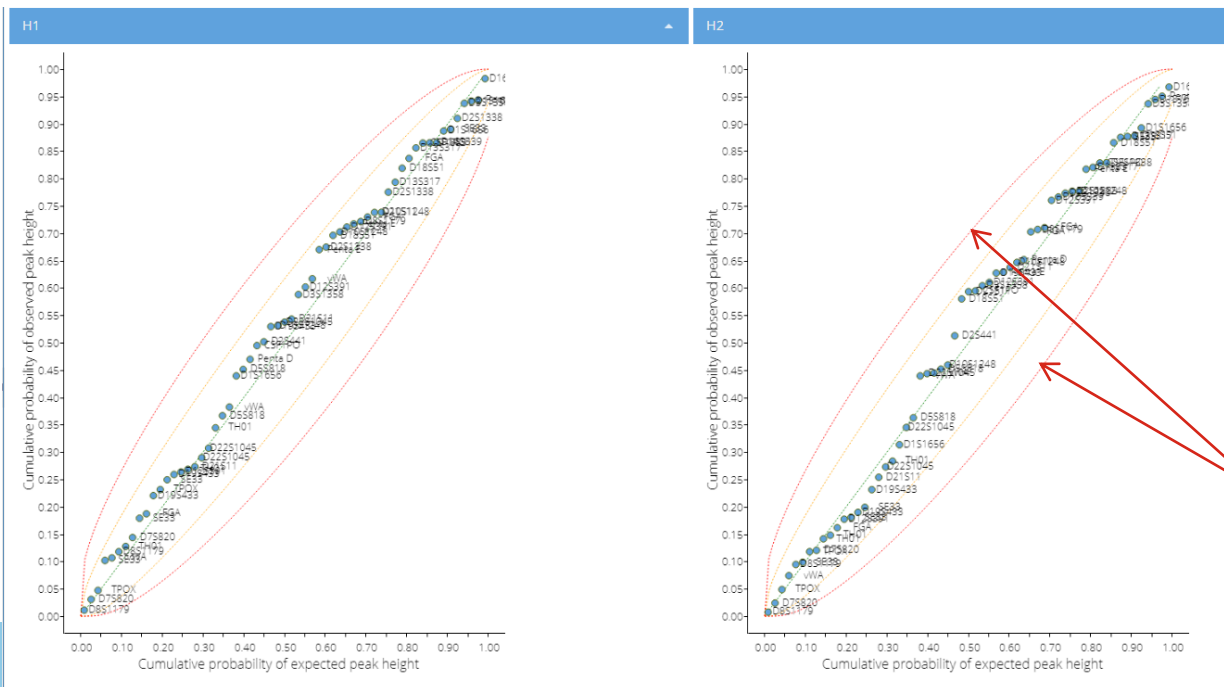
Summarized guidelines to be used as helpful tool in the decision making process are as follows:



# Model validation

- The model validation plots are important quality checks.
- In this model validation, the cumulative probabilities for the expected peak height are plotted against the cumulative probabilities for the observed peak heights, resulting in a PP (probability-probability) plot.

Validation	
Model fitting:	Passed
Iterations H1:	3
Iterations H2:	3



The default Bonferroni corrected significance level is set at 0.01.

When at least four values are outside the envelope (0.01-line), the model validation is scored as 'failed'.

# Model validation status

- Model validation may fail due to various reasons

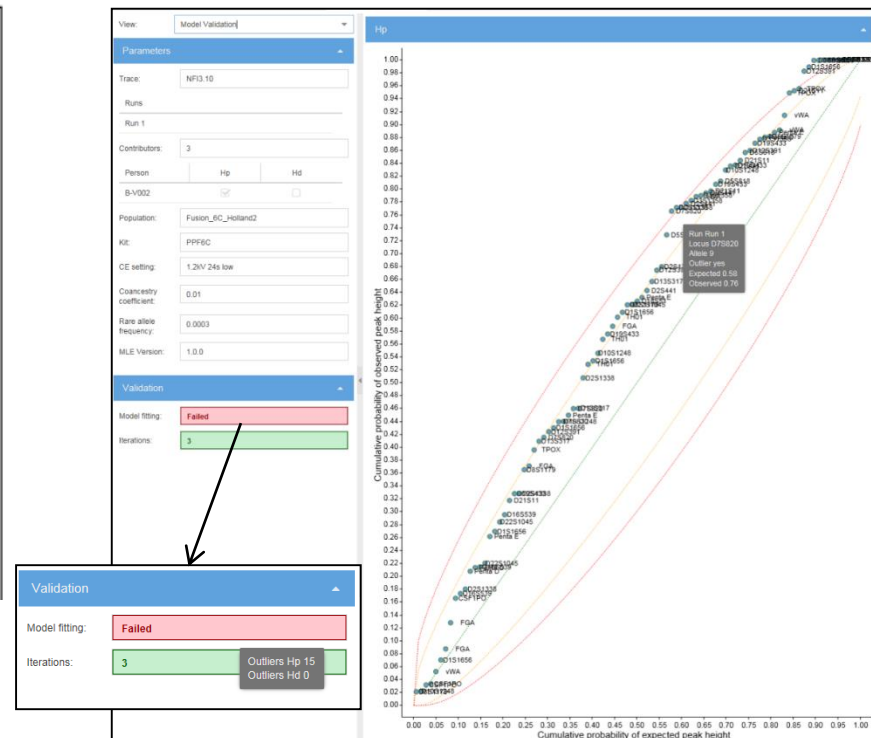
The model validation status can be returned as Passed, Warning, Failed, or Unavailable:

Passed	The model fitting has no or one outlier(s) under H1 and H2 separately.
Warning	The model fitting includes 2 or 3 outliers under H1 and/or H2 separately.
Failed	The model fitting includes more than 3 outliers under H1 and/or H2 separately.

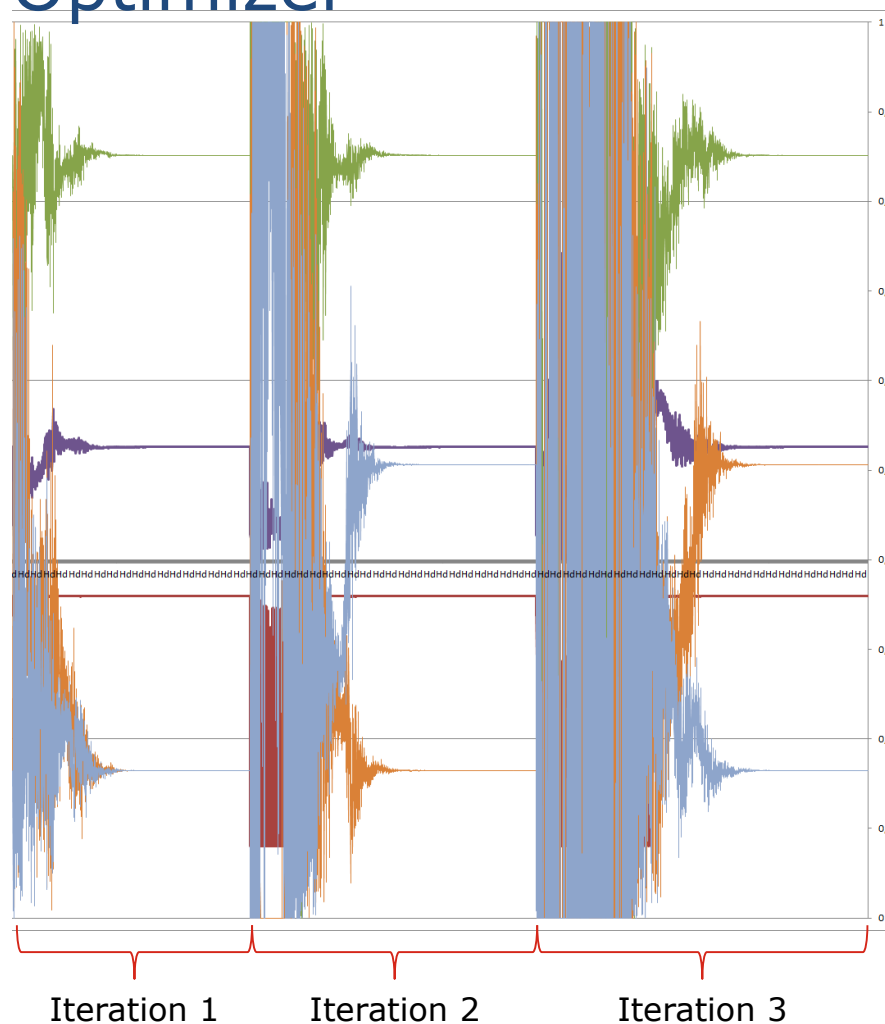
Model fitting (i.e. validation) may fail when peak heights cannot be explained according to the propositions and parameters, and can occur as a result of e.g.:

- Incorrect parameter setting such as
  - o Lack of degradation model application when data does show degradation
    - Advised action: rerun using degradation model turned on.
  - o Underassigned number of contributors (and/or drop-in values) to enable explaining the observed peaks
    - Advised action: rerun using higher number of contributors.
- H1 analyses with a non-contributor as POI (note that model validation often, but not always, fails with a non-contributor)
  - Advised action: none if LR is exclusionary. Results can be reported.
- A peak of improbable height
  - Advised action: check profile.
- Analyses with replicates of extraordinary peak height variation
  - Advised action: Report the LR based on the individual replicates (if model validation passes), but not using replicates analyzed jointly.
- Other than above
  - Advised action: Check results (e.g. EPG vs LR per locus, equality of mixture proportions under H1 and H2, kit settings) and perform a rerun. If model validation still fails, it is advised not to report this LR value.

- If failing model validation cannot be explained/ solved, it is advised not to report the LR

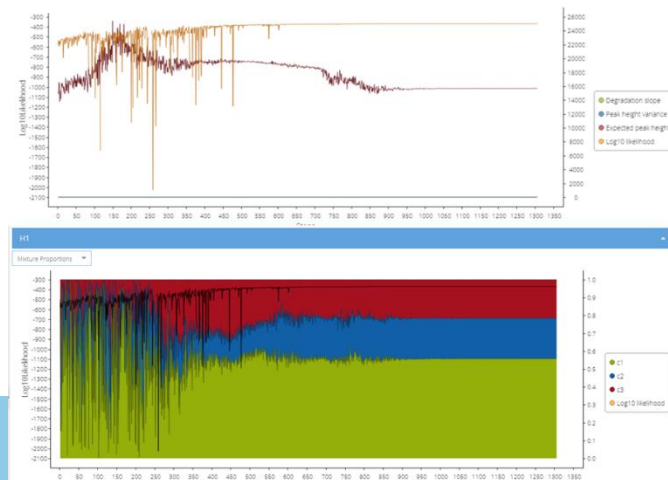
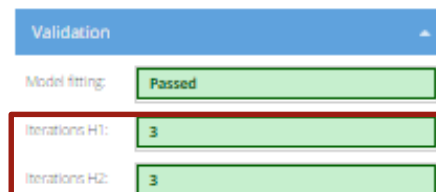


# Optimizer






- Log likelihood estimation
- Expected peak height
- Degradation slope
- Mixture proportion 1
- Mixture proportion 2

- Optimizer results are accepted in DNAXs if 3 times sufficiently similar results are obtained.



## Optimizer iterations

- User manual: The number of Iterations defines the number of repetitions needed to obtain a sufficiently similar optimum. The number of similar results needed is an application parameter and can be set by an administrator (see Settings: [Change application settings](#)). The MLE calculation is repeated until the predefined number of similar results (default 3) is reached or when the maximum number of repetitions is met (default 10). The maximum number of repetitions is also an application setting that can be changed by the administrator in the application settings.
- In DNASStatistX, the MLE calculation is repeated until three sufficiently similar results are found with a maximum of ten repetitions.

	The number of similar results is met (the default number of required similar results is 3).
	The predefined number of similar results was not met after the maximum number of iterations.
	No sufficiently similar results were found after the maximum number of iterations.

- Iterations may fail under H1 and/or H2 and indicate that no three (default) optimum parameter values were found to be sufficiently similar out of ten iterations.
- With failing iterations it is advised to rerun the calculation.

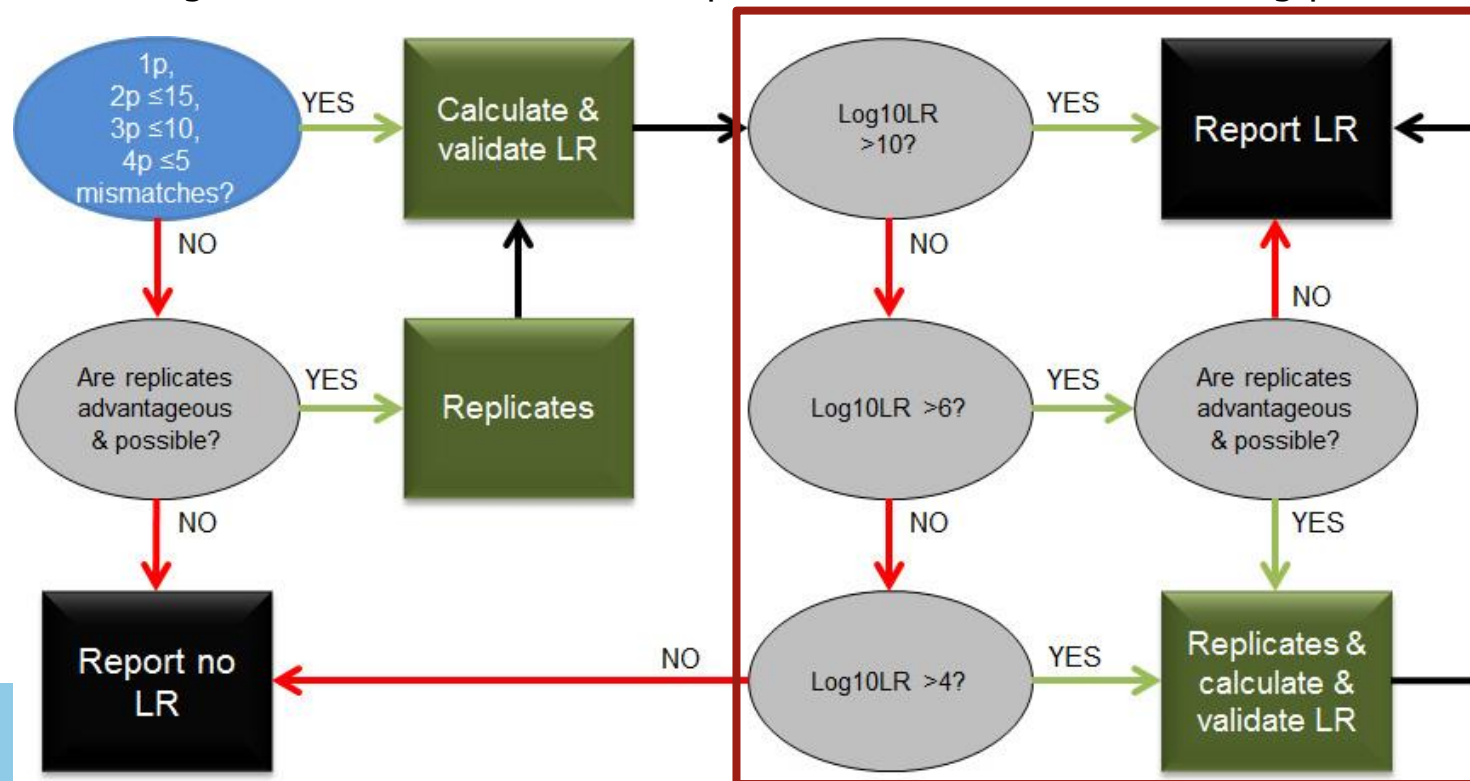




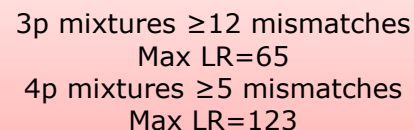
# Guidelines for use in forensic casework

- Based on results from research, casework experience, including efficiency and usefulness of calculations.
- Clearly, the case circumstances, availability of other stains and profiles within the case are factors considered in the process.

Summarized guidelines to be used as helpful tool in the decision making process are as follows:



## Trends with 2p, 3p and 4p PPF6C profiles



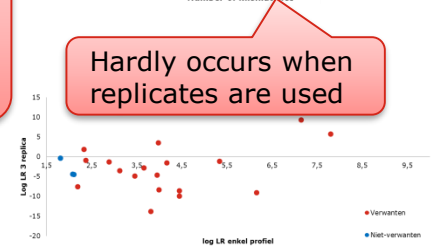
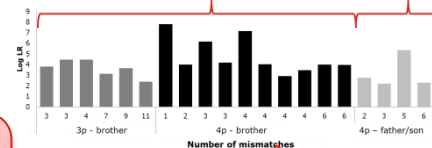
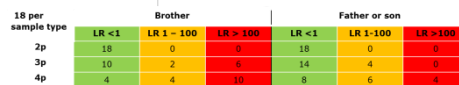
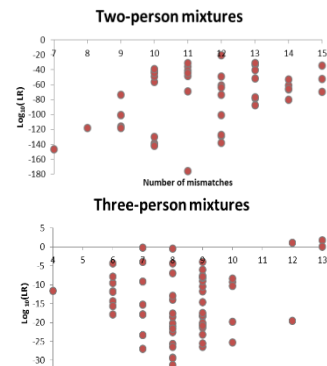
Simulated brothers and fathers/sons that are selected based on a high overlap with the stain profile can yield large LRs

False positives

False  
negatives

Some 3p & 4p samples with  $\geq 5$  drop-outs

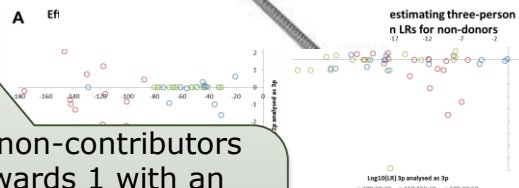
Hardly occurs when replicates are used



True positives

All non-donors  
(non-relatives) for  
2p samples

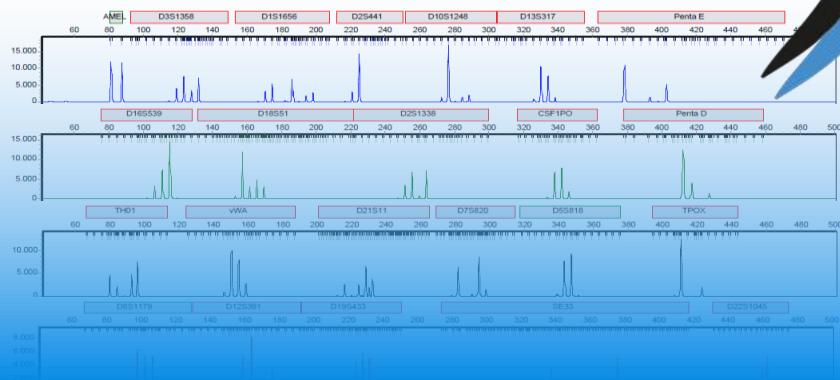
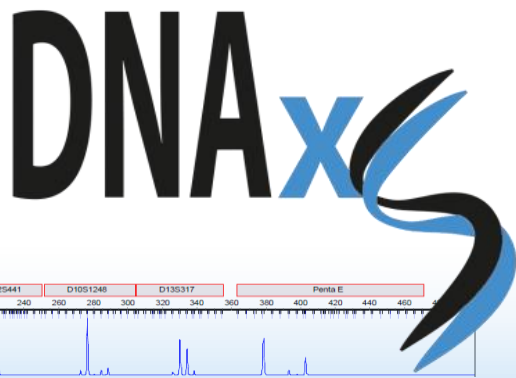
LRs for non-contributors tend towards 1 with an overestimated number of contributors (not seen when mixtures contain equal contributions).





Netherlands Forensic Institute  
Ministry of Justice and Security

# Interpretation of (mixed) DNA profiles within the NFI



*Corina Benschop*

*13/11/2020*