#### **DATA2002**

What can we do when ANOVA assumptions fail?

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What happens when assumptions fail?

Kruskal-Wallis test

# When ANOVA assumptions might be violated

## Assumptions underlying ANOVA (and related methods)

We have learnt much about the comparison of several samples:

- The *F*-test (with p-value computed using the *F*-distribution)
- contrasts (and associated methods, all based on the t-distribution)
- multiple comparisons:
  - Bonferroni
  - Tukey
  - Scheffé

**However**, underlying all of these are the assumptions that

- each sample is from a normal population;
- all population variances are equal.
  - o so all populations are identical up to possible location shifts

What do we do if these assumptions are not reasonable?

#### Possible violations

- There are various ways the assumptions might be violated:
  - the normality might be ok, but **equal variances** might not be;
  - the normality might **not** be ok, but the "identical up to location shifts" assumption might be ok.
- There are a few tools we can appeal to:
  - simulation
  - resampling (together with *conditioning*).

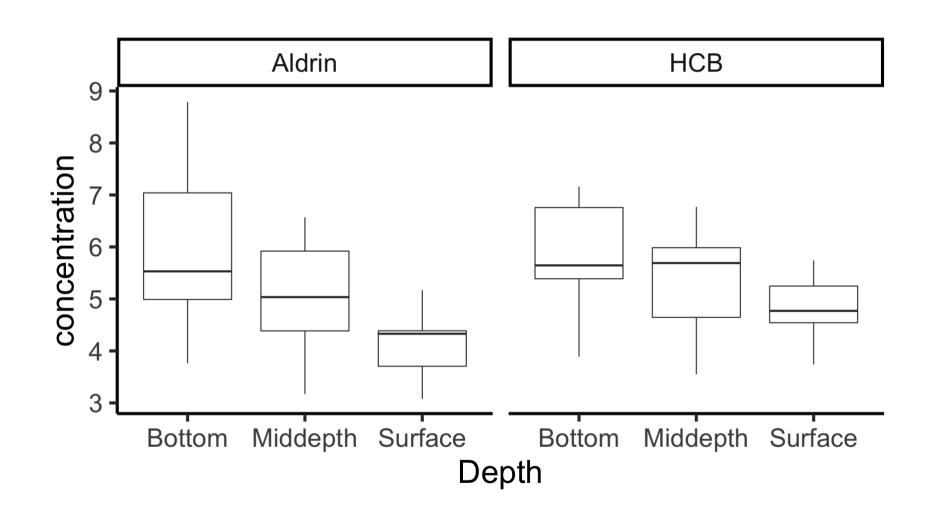
# Relaxing the equal variance assumption

#### Wolf River data

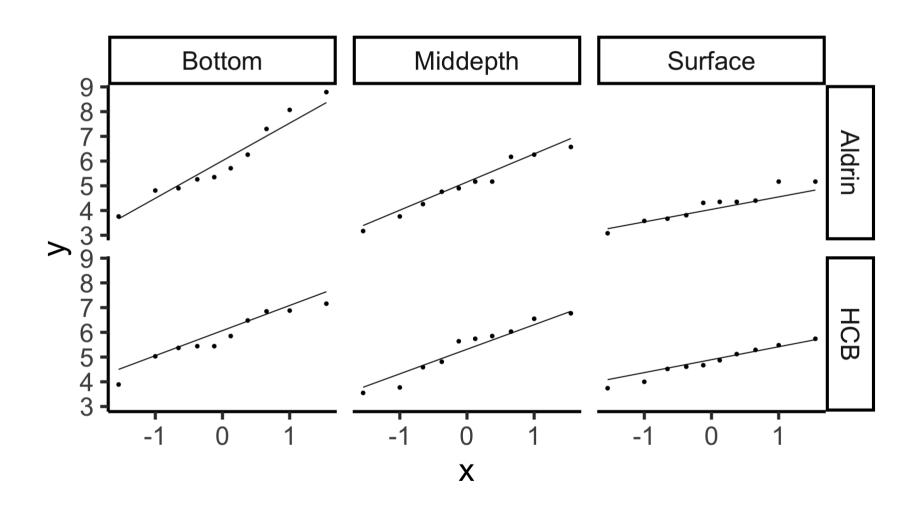
- The data in the file wolfriver.csv contains 30 measurements (10 at 3 depths: Bottom = 1; Middepth = 2; Surface = 3) on each of two chemicals, Aldrin and HCB.
  - these were taken downstream from an abandoned dump site near the Wolf River in Tennessee (Jaffe, Parker, and Wilson, 1982).
- It was believed the concentration might not be constant across different depths.
- We shall be *mainly* interested in each chemical separately, although considering them jointly is also a bit interesting...

```
library(tidyverse)
 wolf = read csv("https://raw.githubusercontent.com/DATA2002/data/master/wolfriver.csv")
 glimpse(wolf)
## Rows: 30
## Columns: 3
## $ Aldrin <dbl> 3.08, 3.58, 3.81, 4.31, 4.35, 4.40, 3.67, 5...
## $ HCB <dbl> 3.74, 4.61, 4.00, 4.67, 4.87, 5.12, 4.52, 5...
## $ Depth <dbl> 3, 3, 3, 3, 3, 3, 3, 3, 3, 2, 2, 2, 2, 2...
wolf = wolf %>% mutate(
                                                     wolf %>% count(Depth)
  Depth = case when(
    Depth == 1 ~ "Bottom",
                                                    ## # A tibble: 3 × 2
    Depth == 2 ~ "Middepth",
                                                    ## Depth n
    Depth == 3 ~ "Surface"
                                                    ## <chr> <int>
                                                    ## 1 Bottom
                                                                     10
                                                    ## 2 Middepth
                                                                   10
                                                    ## 3 Surface
                                                                   10
                                                     wolf_long = wolf %>%
                                                       gather(key = "chemical",
                                                              value = "concentration", -Depth)
```

```
ggplot(wolf_long, aes(x = Depth, y = concentration)) +
  geom_boxplot() + facet_wrap(~chemical)
```



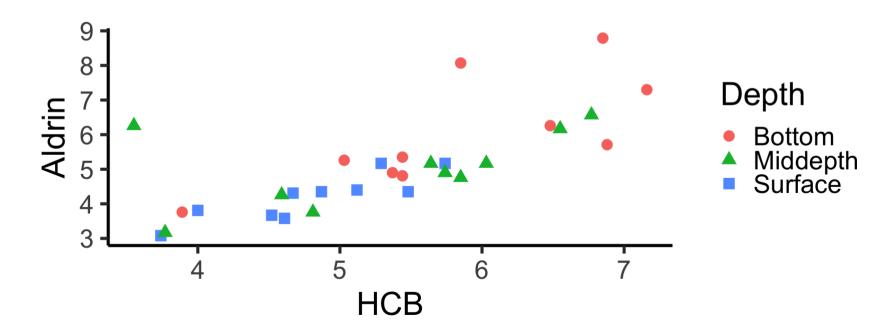
```
ggplot(wolf_long, aes(sample = concentration)) +
  geom_qq() + geom_qq_line() + facet_grid(chemical ~ Depth)
```



### Assumptions?

- Both sets of boxplots suggest a different spread in each group
  - normality is probably ok (points close to line in QQ plots)
- A scatterplot (tracking both chemicals together) reveals something interesting:

```
ggplot(wolf) + aes(x = HCB, y = Aldrin, shape = Depth, colour = Depth) +
  geom_point(size = 5)
```



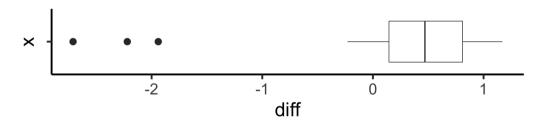
#### **Outliers** ?!

• There are 3 possible "outliers" (actually *bivariate* outliers, really):

```
wolf %>% arrange(HCB-Aldrin)
```

```
## # A tibble: 30 × 3
     Aldrin
##
              HCB Depth
      <dbl> <dbl> <chr>
##
      6.26 3.55 Middepth
##
       8.07
            5.85 Bottom
##
       8.79
             6.85 Bottom
       5.26
             5.03 Bottom
##
##
       7.3
             7.16 Bottom
##
       5.35
             5.44 Bottom
##
       5.17 5.29 Surface
##
       3.76
             3.89 Bottom
##
       3.81
                   Surface
## 10
       6.57 6.77 Middepth
## # ... with 20 more rows
```

```
wolf %>% mutate(
  diff = HCB - Aldrin
) %>%
  ggplot() + aes(x = "", y = diff) +
  geom_boxplot(outlier.size = 5) +
  coord_flip()
```



# Relaxing the "common variance" assumption: all pairwise comparisons

- We could consider assuming normality, but dropping the "common variance" assumption.
- A simple way to do so is to consider all pairwise **Welch tests**, and apply a Bonferroni correction.
- Recall **Welch tests** only assume that each sample is normal, with possibly different variances  $\sigma_X^2$  and  $\sigma_Y^2$  and different means, and all random variables are independent.

$$T=rac{ar{X}-ar{Y}}{\sqrt{rac{S_X^2}{m}+rac{S_Y^2}{n}}},$$

which is **approximately**  $t_{d^*(m,n,\sigma_X,\sigma_Y)}$  under  $H_0$ , for a known function  $d^*(\dots)$ .

• The Welch tests is the what R defaults to in the t.test() function.

# Welch test pairwise comparisons (unadjusted)

#### Middepth vs Surface

```
t.test(wolf$Aldrin[wolf$Depth=="Middepth"],wolf$Aldrin[wolf$Depth=="Surface"])$p.value
```

## [1] 0.06053252

#### **Middepth vs Bottom**

```
t.test(wolf$Aldrin[wolf$Depth=="Middepth"],wolf$Aldrin[wolf$Depth=="Bottom"])$p.value
```

**##** [1] 0.119901

#### **Surface vs Bottom**

```
t.test(wolf$Aldrin[wolf$Depth=="Surface"],wolf$Aldrin[wolf$Depth=="Bottom"])$p.value
```

## [1] **0.**005471484

- Since we are doing 3 pairwise comparisons, we mutliply the "unadjusted" p-values by 3 to get "adjusted-for-multiplicity" p-values.
- The smallest of these can be used as a test that all (population) means are equal:

```
t.test(wolf$Aldrin[wolf$Depth=="Surface"], wolf$Aldrin[wolf$Depth=="Bottom"])$p.value

## [1] 0.005471484

3 * t.test(wolf$Aldrin[wolf$Depth=="Surface"], wolf$Aldrin[wolf$Depth=="Bottom"])$p.value

## [1] 0.01641445
```

• This is a perfectly valid p-value for testing the "global" or "overall" hypothesis that all means are equal (assuming all 3 populations are normal, but with possibly different variances).

```
pairwise.t.test(wolf$Aldrin, wolf$Depth, p.adjust.method = "none", pool.sd = FALSE)
##
       Pairwise comparisons using t tests with non-pooled SD
##
##
## data: wolf$Aldrin and wolf$Depth
##
##
            Bottom Middepth
## Middepth 0.1199 -
## Surface 0.0055 0.0605
##
## P value adjustment method: none
 pairwise.t.test(wolf$Aldrin, wolf$Depth, p.adjust.method = "bonferroni", pool.sd = FALSE)
##
       Pairwise comparisons using t tests with non-pooled SD
##
##
## data: wolf$Aldrin and wolf$Depth
##
##
            Bottom Middepth
## Middepth 0.360
## Surface 0.016 0.182
##
```

## P value adjustment method: bonferroni

#### Simultaneous confidence intervals

• To obtain a set of 3 simultaneous Bonferroni-style 95% confidence intervals, we compute 3 individual  $(1-(0.05/3)) \times 100 = 98.3\%$  intervals

#### Middepth vs Surface

```
t.test(wolf$Aldrin[wolf$Depth=="Middepth"],
     wolf$Aldrin[wolf$Depth=="Surface"],
     conf.level = 1-(0.05/3))$conf.int
```

```
## [1] -0.2721228 1.9321228
## attr(,"conf.level")
## [1] 0.9833333
```

#### **Middepth vs Bottom**

#### **Surface vs Bottom**

```
t.test(wolf$Aldrin[wolf$Depth=="Surface"],
     wolf$Aldrin[wolf$Depth=="Bottom"],
     conf.level = 1-(0.05/3))$conf.int
```

```
## [1] -3.3395315 -0.3244685
## attr(,"conf.level")
## [1] 0.9833333
```

• Of course, this does not include 0 because the *adjusted* p-value < 0.05!

# Relaxing the normality assumption

# A p-value for the "global" hypothesis under weaker assumptions

- Under the formal ANOVA assumptions:
  - each population is normal
  - variances are the same

the null hypothesis reduces to

All observations come from the same normal distribution

A weaker set of assumptions at least under the null hypothesis is that

All observations come from the same distribution

## The powerful tool of *conditioning*

- A common tool in testing is to condition on an "ancillary" statistic:
  - "ancillary statistic" just means a statistic that does not tell us anything useful.
- A familiar example is the **sign test**:
  - $\circ$  we usually *condition* on the number N of non-zeroes (i.e. we *ignore the ties*)
- Then, p-values are in fact *conditional* probabilities, e.g. for a one-sided sign test based on the number S of positive signs, the p-value is

$$P(S \geq s \mid N=n) = P(B(n,0.5) \geq s)$$

## Conditioning on the combined sample

- If we combine all the groups into one combined sample (i.e. throw away the labels) then the remaining "data" tells us **nothing** about differences between groups i.e. what we are interested in.
- In this sense, the *combined sample* is an "ancillary statistic".
- Once we condition on the combined sample, the only remaining "randomness" is the allocation of observations to groups.
- Under the null hypothesis of "no differences between groups" all possible allocations are equally likely.

### Enumerating all possible allocations: exact p-values

- We can (in principle) compute an *exact conditional* p-value for **any** "**sensible**" **statistic** under this particular null hypothesis.
- There are actually

$$\frac{N!}{n_1!n_2!\dots n_q!}$$

different possible allocations of the N total observations into groups of size  $n_1, n_2, \ldots, n_g$ .

- We can (in principle) compute the value of the statistic under each possible allocation.
- Since each such value is *equally likely under the null hypothesis*, we can use this "sampling distribution" to compute a p-value.

- Suppose the statistic is T, the observed value is  $t_0$  and larger values indicate more evidence against the null hypothesis.
- The *exact* conditional p-value is a *simple proportion*:

$$P(T \geq t_0 \mid ext{combined sample}) = rac{ ext{no. allocations with } T \geq t_0}{ ext{total no. allocations}} \,.$$

- Unfortunately, unless the sample sizes are very small,
  - the total number of allocations is MASSIVE;
  - computing the value of the statistic over all possible allocations is not feasible.
- Fortunately, we can *estimate* this proportion by taking a sufficiently large random sample from the "population of all possible allocations":
  - this is a binomial/hypergeometric (depending on whether we sample with or without replacement) proportion estimation problem!

#### Permutation tests

- In R, if the data is represented as a data frame with
  - observations in one column and
  - groups indicated by a factor in another column

then is it **easy** to obtain a "random" allocation:

- simply randomly permute the observation vector, keeping the factor vector fixed.
- Do this a large number of times.
- The "observed proportion" of the times the statistic exceeds  $t_0$  becomes an *estimate* of the "exact" p-value.
- This general procedure is known as a *permutation test*.

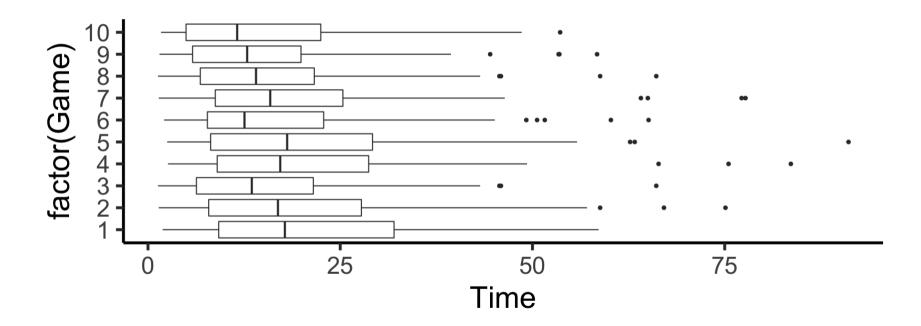
## Rugby analysis

\*

- In the early 1990's the rules for International Rugby were changed, apparently to make the game "more continuous", that is, for passages of "play" to be longer between stoppages.
  - The data is in the file rugby.txt.
- The lengths of time of passages of play were recorded in 10 games featuring the New Zealand national team (the "All Blacks"):
  - the first 5 games under the old rules
  - the last 5 games under the new rules
- Boxplots appear on the next slide.



```
rugby = read_tsv("http://www.statsci.org/data/oz/rugby.txt")
ggplot(rugby) + aes(x = factor(Game), y = Time) +
  geom_boxplot() + coord_flip() +
  theme_classic(base_size = 30)
```



Looks skewed, not normal...

#### F-test?

## [1] 3.886706

- Let's try doing a permutation test using the F-statistic.
- It is convenient to use the R function anova(aov(...)) or broom::tidy(aov(...)) instead of summary(aov(...)) since the ANOVA table is returned as numbers in a matrix:

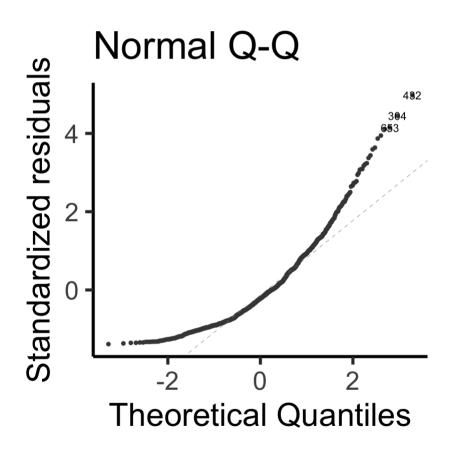
```
rugby_anova = aov(Time ~ factor(Game), data = rugby)
 anova(rugby anova)
## Analysis of Variance Table
##
## Response: Time
                Df Sum Sq Mean Sq F value Pr(>F)
##
## factor(Game) 9 6904 767.08 3.8867 7.335e-05 ***
## Residuals 969 191241 197.36
## Signif. codes:
## 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
 anova(rugby anova)[1,4]
```

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# Check for normality

```
library(ggfortify)
autoplot(rugby_anova, which = 2)
```



# The **broom** package

```
rugby_anova = aov(Time~factor(Game), data = rugby)
mod_sum = broom::tidy(rugby_anova)
mod_sum
## # A tibble: 2 × 6
    term
                       sumsq meansq statistic p.value
   <chr> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 factor(Game)
                   9 6904. 767. 3.89 0.0000733
## 2 Residuals
              969 191241. 197.
                                      NA
                                            NA
mod_sum$statistic[1]
## [1] 3.886706
mod\_sum \%>\% kable(format = "markdown", digits = c(0,0,0,1,3,4))
```

term	df	sumsq	meansq	statistic	p.value
factor(Game)	9	6904	767.1	3.887	1e-04
Residuals	969	191241	197.4	NA	NA



• The R function sample(), with only one vector argument, returns a *permutation* of that vector:

```
x = 1:5
sample(x)
```

## [1] 3 4 5 1 2

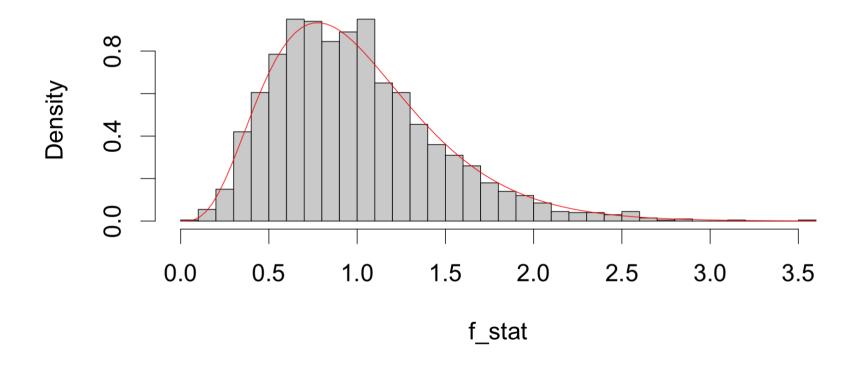
 The following loop takes a sample of size B from all possible permutations and computes the value of the F-statistic:

```
B = 2000
f_stat = vector(mode = "numeric", length = B)
for (i in 1:B){
   permuted_anova = aov(sample(rugby$Time) ~ factor(rugby$Game))
   f_stat[i] = broom::tidy(permuted_anova)$statistic[1]
}
```



```
hist(f_stat, probability = TRUE, breaks = 40)
curve(df(x, 9, 969), add = TRUE, col = "red")
```

#### **Histogram of f\_stat**



(FF)

- The F-distribution density is drawn over the histogram and the fit looks pretty good.
- Our *estimate* of the *exact conditional p-value* is obtained as follows:

```
t_0 = broom::tidy(rugby_anova)$statistic[1]
t_0

## [1] 3.886706

mean(f_stat >= t_0)
```

- So of the 2000 random permutations 0 gave an F-ratio bigger than (or equal to) 3.8867058.
- We have avoided making any normality assumption here!
- This says a lot about the robustness of the F-test...

## [1] 0

 $\circ$  the *conditional* distribution is very close to the corresponding F-distribution.

# Using ranks

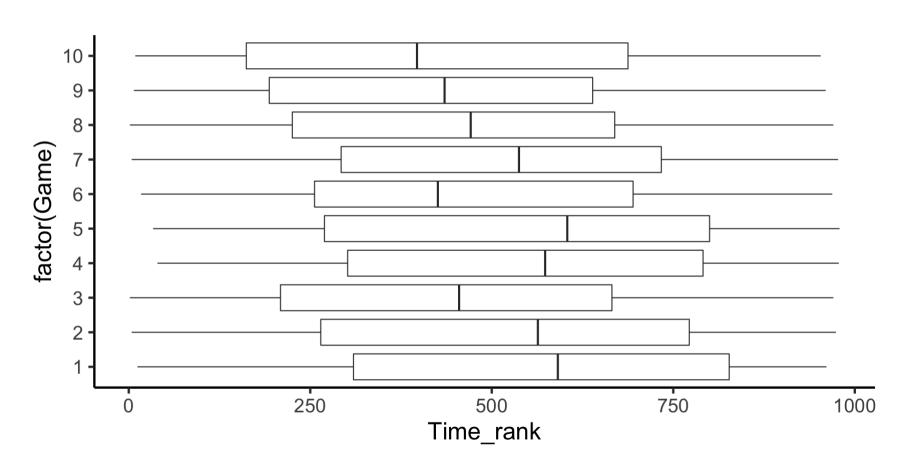
#### Kruskal-Wallis test

- This is performed by
  - replacing each observation by its "global" rank;
  - $\circ$  then computing the *F*-ratio as usual *on the ranks*.
- A p-value can be obtained
  - using a permutation test approach or
  - $\circ$  a "large-sample"  $\chi^2$  approximation can also be used



# Rugby data (ranks)

```
rugby = rugby %>% ungroup() %>% mutate(Time_rank = rank(Time))
ggplot(rugby, aes(x = factor(Game), y = Time_rank)) +
  geom_boxplot() + coord_flip() + theme_classic(base_size = 24)
```





# Rugby data (ANOVA on the ranks)

Perform the usual ANOVA on the ranks.

## 2 Residuals 969 75737977. 78161.

NA

NA

#### Kruskal-Wallis test statistic

- ullet The traditional approach to the Kruskal-Wallis test uses a test statistic that is computed as a ratio (like the F-test)
  - the numerator is *exactly* the Treatment Sum of Squares of the ranks
  - the denominator is the sample variance of all the ranks!
    - this denominator is not random (it is the same regardless of the allocation).

$$T = rac{ ext{Treatment SS of the ranks}}{ ext{Varance of all the ranks}}$$

• When  $H_0$  is true, it has an *approximate*  $\chi^2_{q-1}$  distribution.



```
rugby_rank_anova = aov(Time_rank ~ factor(Game), data = rugby)
rank_Treat_SS = broom::tidy(rugby_rank_anova)$sumsq[1]
rank_Treat_SS

## [1] 2452917

rank_Treat_SS/var(rugby$Time_rank)
```

## [1] 30.68072

## The R function kruskal.test()

• This statistic is computed (and a resultant "approximate" p-value is obtained) via the R function kruskal.test():

```
##
##
## Kruskal-Wallis rank sum test
##
## data: Time by factor(Game)
## Kruskal-Wallis chi-squared = 30.681, df = 9, p-value
## = 0.0003358
```

## Kruskal-Wallis procedure

- 1. **Hypotheses:**  $H_0$ : the response variable is distributed identically for all groups vs  $H_1$ : the response variable is systematically higher for at least one group
- 2. **Assumptions:** Observations are independent within each group and groups are independent of each other. The different groups follow the same distribution (differing only by the location parameter).
- 3. **Test statistic:** like ANOVA applied to the ranks (not examinable), see here for details. Under the null hypothesis the Kruskal-Wallis test statistic approximately follows a  $\chi^2$  distribution with g-1 degrees of freedom where g is the number of groups.
- 4. **p-value:**  $P(T \ge t_0) = P(\chi_{g-1}^2 \ge t_0)$ .
- 5. **Decision:** If the p-value is less than  $\alpha$  we reject the null hypothesis and conclude that the population mean of at least one group is significantly different to the others. If the p-value is larger than  $\alpha$  we do not reject the null hypothesis and conclude that there is no significant difference between the population means.

#### Permuted ranks

- The permutation test approach is valid for any "sensible" statistic;
  - o it only assumes the same distribution in each group under the null hypothesis.
- What of the "sensible statistic"?
  - If the data are truly normal, the F-statistic makes sense;
  - is it still "sensible" if the normality assumption is being relaxed?
  - could also do a permutation test using the Kruskal-Wallis statistic



# Rugby data (KW permutation test)

```
set.seed(1)
 B = 2000
 Game = rugby$Game
 Time = rugby$Time
 kw_stat = vector("numeric", length = B)
 for (i in 1:B){
   aov_rank = aov(rank(sample(Time)) ~ factor(Game))
   kw_stat[i] = broom::tidy(aov_rank)$statistic[1]
 original_rank_mod = aov(rank(Time)~factor(Game))
 t0 = broom::tidy(original_rank_mod)$statistic[1]
 t0
## [1] 3.486988
mean(kw_stat >= t0)
## [1] 5e-04
```

#### References

Jaffe, P., F. Parker, and D. Wilson (1982). "Distribution of toxic substances in rivers". In: *Journal American Society of Civil Engineers, Environmental Engineering Division* 108, pp. 639-649.

Lenth, R. (2018). *emmeans: Estimated Marginal Means, aka Least-Squares Means*. R package version 1.2.3. URL: https://CRAN.R-project.org/package=emmeans.