Kendall Model

Kendall Fitzgerald

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Invertebrates to include in analysis: purple urchin, sunflower sea star, northern kelp crab

Setup

```
#load necessary packages
library(here)
## here() starts at /home/guest/ENV710/group_project
library(tidyverse)
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
                                   2.1.5
## v dplyr 1.1.4
                      v readr
## v forcats 1.0.0 v stringr 1.5.1
## v ggplot2 3.5.1 v tibble
                                   3.2.1
## v lubridate 1.9.3
                    v tidyr
                                   1.3.1
## v purrr
              1.0.2
## -- Conflicts ------ tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()
                   masks stats::lag()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts to become error
library(dplyr)
library(naniar)
library(gtsummary)
library(lme4)
## Loading required package: Matrix
##
## Attaching package: 'Matrix'
## The following objects are masked from 'package:tidyr':
##
##
      expand, pack, unpack
```

```
library(MASS)
##
## Attaching package: 'MASS'
##
## The following object is masked from 'package:gtsummary':
##
##
       select
##
## The following object is masked from 'package:dplyr':
##
##
       select
#load dataset
inverts_kelp <- read_csv("PISCO_kelpforest_swath.1.2.csv")</pre>
## Rows: 222227 Columns: 17
## -- Column specification -----
## Delimiter: ","
## chr (9): campus, method, site, zone, classcode, disease, observer, notes, si...
## dbl (8): survey_year, year, month, day, transect, count, size, depth
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
#tidy data, the addition of recruit and juvenile classcodes does not make a difference so will move for
inverts_kelp <- inverts_kelp %>%
  filter(campus == "UCSC",
         classcode %in% c("PUGPRO", "STRPURAD", "PYCHEL", "MACPYRAD")) %>%
  #only include relevant columns for further analysis
  dplyr::select(campus, survey_year, site, zone, classcode, count, size, disease, depth)
#further tidy data based on relevant rows and sum count data
inverts_summarized <- inverts_kelp %>%
  group_by(campus, site, survey_year, classcode, depth) %>%
  summarise(
   total_count = sum(count, na.rm = TRUE),
   zone = first(zone),
    .groups = "drop"
#group by species (classcode), year (survey_year), and zone
inverts_wide_grouped <- inverts_summarized %>%
  group_by(classcode, survey_year, zone) %>%
  summarise(across(c(total_count), sum, na.rm = TRUE), .groups = "drop")
## Warning: There was 1 warning in 'summarise()'.
## i In argument: 'across(c(total_count), sum, na.rm = TRUE)'.
## i In group 1: 'classcode = "MACPYRAD"', 'survey_year = 1999', 'zone = "INNER"'.
## Caused by warning:
## ! The '...' argument of 'across()' is deprecated as of dplyr 1.1.0.
## Supply arguments directly to '.fns' through an anonymous function instead.
```

```
##
## # Previously
## across(a:b, mean, na.rm = TRUE)
##
## # Now
## across(a:b, \(x) mean(x, na.rm = TRUE))

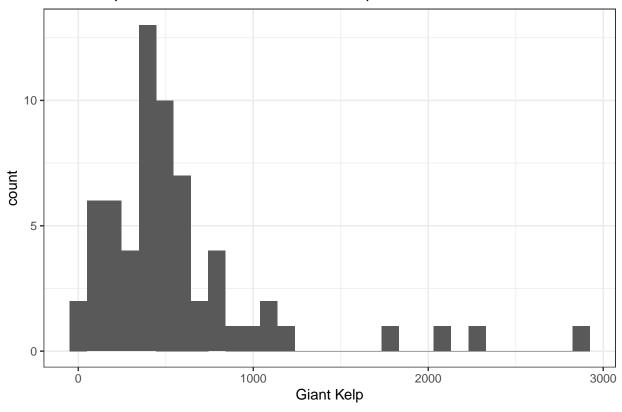
#pivot to wide format to separate out species into separate rows
inverts_wide <- inverts_wide_grouped %>%
    pivot_wider(names_from = classcode, values_from = total_count, values_fill = list(total_count = 0))
```

Step 1. Define Research Question

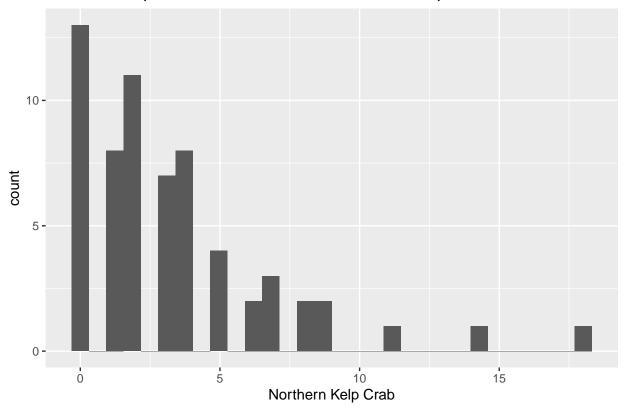
How does the presence of different invertebrates and habitat zone correlate with kelp abundance? Null Hypothesis: Invertebrate presence and habitat zone do not have any correlation with kelp abundance. Alternative Hypothesis: Invertebrate presence and habitat zone do have a correlation with kelp abundance.

Step 2. Examine data and possible correlations

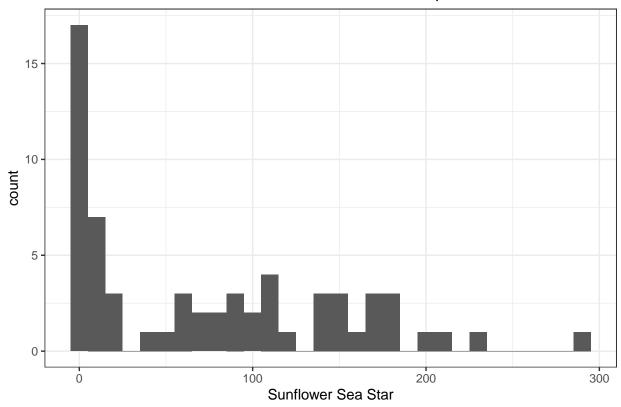
Giant Kelp Abundance in Northern CA Kelp Forests



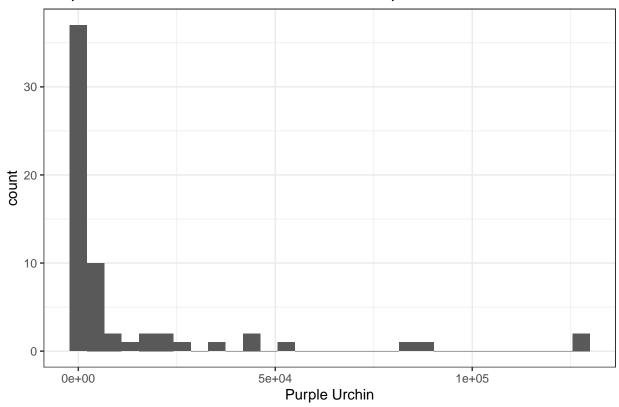
Northern Kelp Crab Abundance in Northern CA Kelp Forests



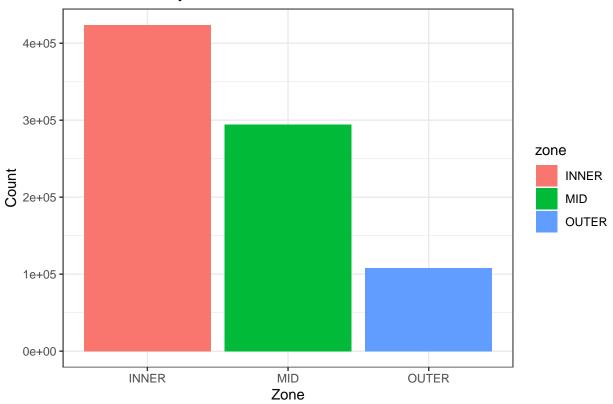
Sunflower Sea Star Abundance in Northern CA Kelp Forests



Purple Urchin Abundance in Northern CA Kelp Forests



Transect Survey Zone Distribution



Relationship figures of species through time

```
#first tidy data so that it can be more easily understood by ggplot; summarize the counts by species an
species_counts <- inverts_summarized %>%
  group_by(classcode, survey_year) %>%
  summarise(total_count = sum(total_count, na.rm = TRUE), .groups = "drop")
#create labels for graph
labels <- c("MACPYRAD"="Giant Kelp",</pre>
                              "PUGPRO"="Northern Kelp Crab",
                              "PYCHEL"="Southern Sea Star",
                              "STRPURAD" = "Purple Urchin")
#create facet-wrapped figure to show species counts by year
species_separate <- ggplot(species_counts, aes(x = survey_year, y = total_count)) +</pre>
  geom_point(aes(color = classcode), size = 2, alpha = 0.7) +
  geom_smooth(aes(color = classcode), method = "loess", se = FALSE) +
  geom_vline(xintercept = 2013, linetype = "longdash", color = "black") +
  facet_wrap(~ classcode, scales = "free_y", labeller = labeller(classcode = labels))+
  labs(title = "Distribution of Kelp Forest Species Abundance Used in Model Over Time",
       x = "Survey Year",
       y = "Total Count") +
  theme bw() +
  theme(strip.text = element_text(face = "bold"),
```

```
axis.text.x = element_text(angle = 45, hjust = 1),
    plot.title = element_text(hjust = 0.5, face = "bold", size = 12),
    legend.position = "none")

species_separate
```

'geom_smooth()' using formula = 'y ~ x'

Distribution of Kelp Forest Species Abundance Used in Model Over Time

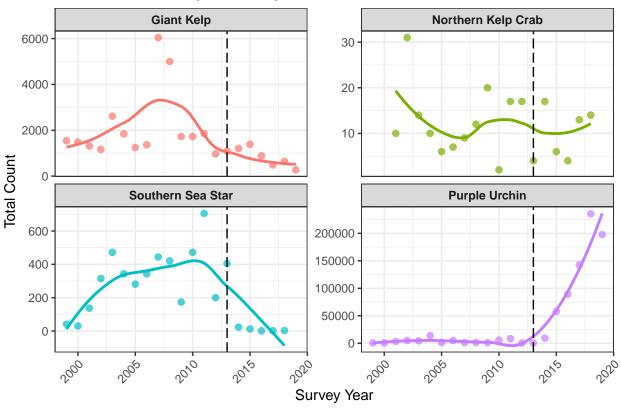


Figure 1: This figure shows the abundance of species used in the model over time. The black dotted line sitting at the year 2013 indicates the first large marine heatwave during that time period. This heatwave significantly contributed to the alteration of the kelp forest ecosystem.

Performing correlation tests between independent variables

```
#make dataframe with continuous predictor variables only
predictors <- inverts_wide %>%
   dplyr::select(PUGPRO, STRPURAD, PYCHEL)

#calculate the correlation matrix
cor_matrix <- cor(predictors, use = "complete.obs")

# View the correlation matrix
cor_matrix</pre>
```

```
## PUGPRO STRPURAD PYCHEL
## PUGPRO 1.00000000 -0.02495791 0.2427995
## STRPURAD -0.02495791 1.00000000 -0.4112285
## PYCHEL 0.24279953 -0.41122852 1.0000000
```

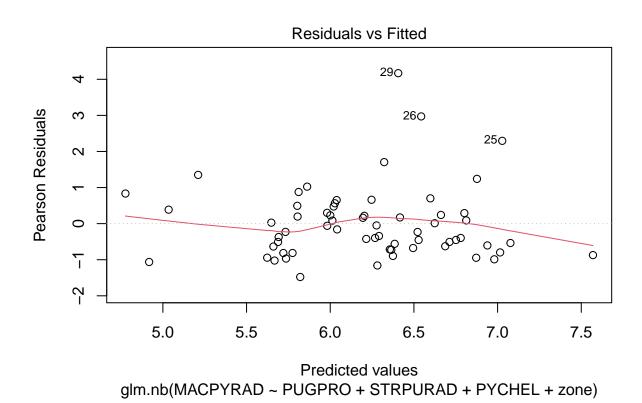
Because correlation does not exceed 0.6 between all variables, the test showed little to moderate correlation between the continuous predictor variables. Since this is the case, we can move forward with step 3.

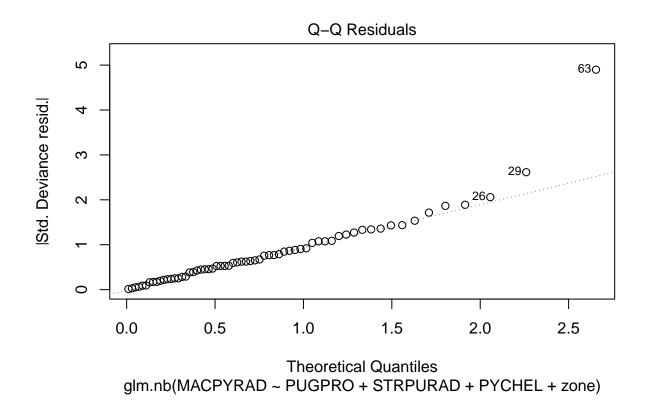
Step 3 - Fit regression model

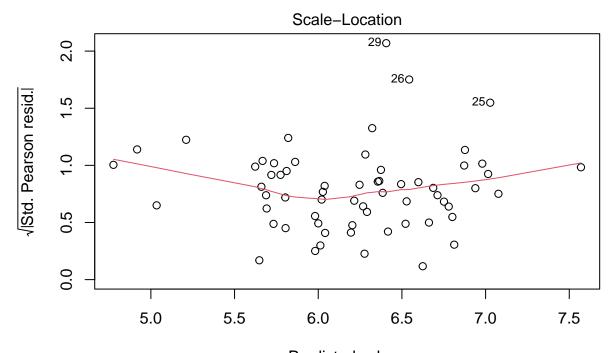
Step 4 - Evaluate Model Diagnostics

```
#Examine overall model output
summary(kelp_glm_nb)
##
## glm.nb(formula = MACPYRAD ~ PUGPRO + STRPURAD + PYCHEL + zone,
##
      data = inverts_wide, init.theta = 2.201056427, link = log)
##
## Coefficients:
##
                Estimate Std. Error z value Pr(>|z|)
## (Intercept) 5.975e+00 2.240e-01 26.667 < 2e-16 ***
## PUGPRO
              -3.216e-02 2.574e-02 -1.250
                                              0.211
## STRPURAD
              -8.380e-06 3.573e-06 -2.345
                                              0.019 *
## PYCHEL
               5.521e-03 1.321e-03
                                     4.178 2.94e-05 ***
## zoneMID
              1.507e-01 2.112e-01
                                    0.713
                                              0.476
## zoneOUTER -5.740e-02 2.227e-01 -0.258
                                              0.797
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for Negative Binomial(2.2011) family taken to be 1)
##
```

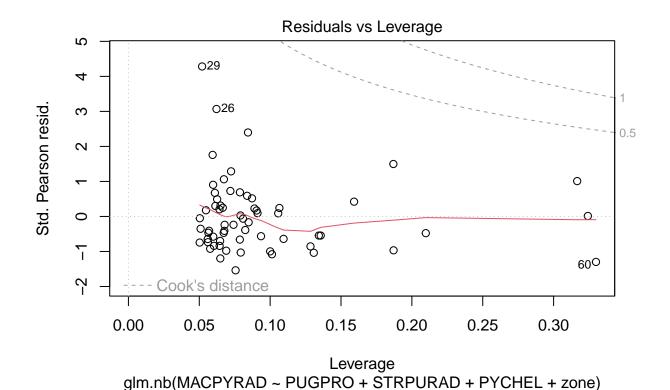
```
Null deviance: 106.503 on 62 degrees of freedom
## Residual deviance: 70.837 on 57 degrees of freedom
  AIC: 906.23
##
  Number of Fisher Scoring iterations: 1
##
##
##
                 Theta:
                         2.201
##
                         0.383
##
             Std. Err.:
##
    2 x log-likelihood:
                         -892.233
##
\#plot\ model\ output\ residuals
plot(kelp_glm_nb)
```





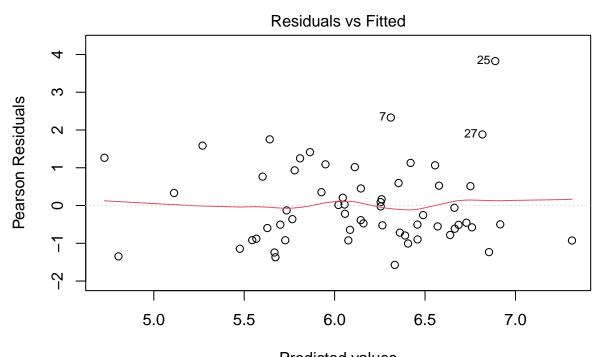


Predicted values glm.nb(MACPYRAD ~ PUGPRO + STRPURAD + PYCHEL + zone)

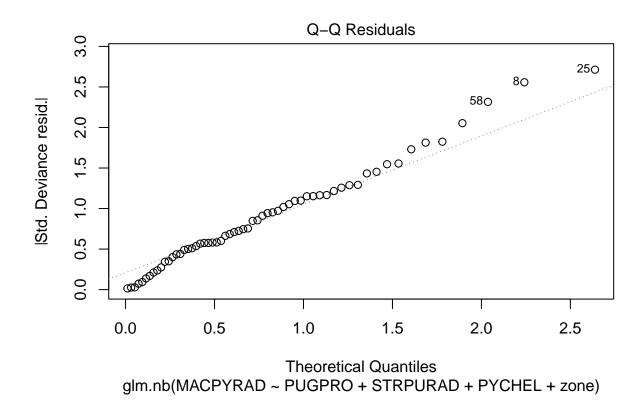


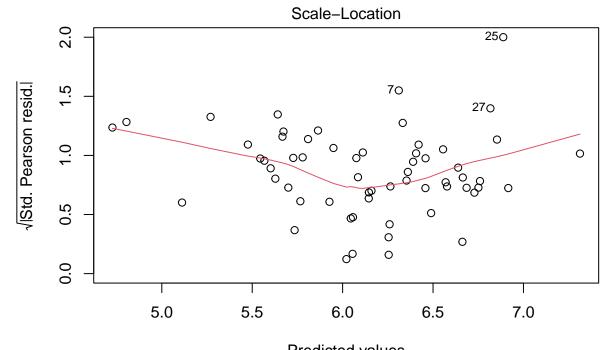
```
##
## Call:
## glm.nb(formula = MACPYRAD ~ PUGPRO + STRPURAD + PYCHEL + zone,
##
       data = inverts_final, init.theta = 3.948276584, link = log)
##
## Coefficients:
                 Estimate Std. Error z value Pr(>|z|)
##
## (Intercept) 6.026e+00 1.677e-01 35.936 < 2e-16 ***
## PUGPRO
               -1.561e-02
                          1.947e-02
                                      -0.802 0.422712
## STRPURAD
                          2.689e-06
                                      -3.604 0.000314 ***
               -9.691e-06
## PYCHEL
                4.155e-03
                          1.006e-03
                                       4.131 3.62e-05 ***
## zoneMID
                1.757e-01
                          1.581e-01
                                       1.112 0.266152
## zoneOUTER
               -2.947e-01 1.727e-01
                                     -1.707 0.087836 .
## Signif. codes:
                  0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
##
```

```
(Dispersion parameter for Negative Binomial(3.9483) family taken to be 1)
##
##
       Null deviance: 121.172
                               on 59
                                      degrees of freedom
                               on 54 degrees of freedom
## Residual deviance: 62.554
   AIC: 828.22
##
##
## Number of Fisher Scoring iterations: 1
##
##
##
                 Theta:
                         3.948
                         0.701
##
             Std. Err.:
##
    2 x log-likelihood:
                        -814.223
#re-examine model residuals
plot(kelp_glm_refit)
```

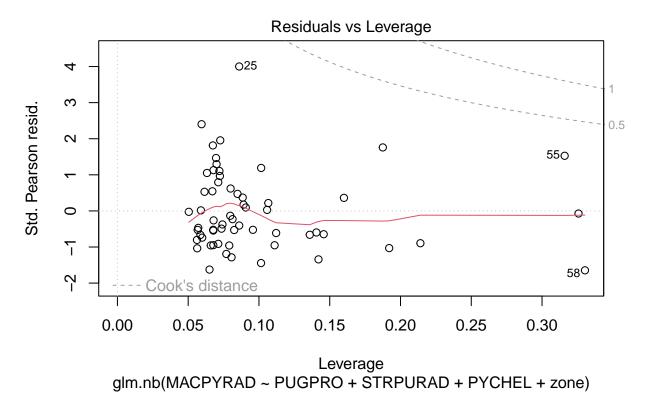


Predicted values glm.nb(MACPYRAD ~ PUGPRO + STRPURAD + PYCHEL + zone)





Predicted values glm.nb(MACPYRAD ~ PUGPRO + STRPURAD + PYCHEL + zone)



#perform Akaike Information Criterion test to compare the two models
AIC(kelp_glm_nb, kelp_glm_refit)

```
## df AIC
## kelp_glm_nb 7 906.2329
## kelp_glm_refit 7 828.2233
```

The results of the Akaike Information Criterion (AIC) test showed that the re-fit model had a lower AIC score and, therefore, was a better fit of the data.

Step 5 - Interpret the model and communicate the results

The Negative Binomial regression model suggests that the abundance of Giant Kelp in study areas of Northern California kelp forests is significantly influenced by the presence of Sunflower Sea Stars (B = 0.004, p = 3.62e-05) and Purple Urchins (B = -9.691e-06, p = 0.0003). Northern kelp crabs (B = -0.0156, p = 0.42), Mid Zone (B = 0.176, p = 0.266), and Outer Zone (B = -0.295, p = 0.0878) by contrast are not significant predictors for Giant Kelp abundance. The significant negative relationship between Giant Kelp and Purple Urchins shows that as urchin populations increase, kelp abundance decreases which makes logical sense as urchins predate on kelp. When left unchecked, they can turn entire kelp forests into "Urchin Barrens" with little to no kelp population. The strong positive relationship between Sunflower Sea Star abundance and kelp abundance also supports this narrative, as Sea Stars primarily predate on urchins and keep their population in check, which in turn helps the kelp population.