Introduction:

Cell Ranger provides prebuilt human and mouse reference packages for use with the pipeline,

downloadable here (https://support.10xgenomics.com/single-cell-vdj/software/downloads/latest). Our

reference packages are based on the T cell receptor (TRA, TRB) and B cell immunoglobin (IGH, IGL,

IGK) gene annotations in Ensembl version 94 for the human and mouse references. Our reference

also includes multiple corrections to various V, D, J, and C genes based on empirical observations

and to correct clear errors such as frameshifts, leader peptide truncations, and nucleotides that are

never observed in rearrangements. These changes are documented in the release notes of each

version of Cell Ranger. Also see Prebuilt References for more information about how these references

were created.

If you would like to use your own genome FASTA or gene GTF annotations, Cell Ranger supports the

use of customer-generated Ensembl-based references. Cell Ranger also includes support for

generating a V(D)J reference from the IMGT database.

There are two ways to generate a V(D)J reference:

Making a Genome-based Reference Package (e.g., using Ensembl)

Making a V(D)J Segment-based Reference Package (e.g., using IMGT)

Making a Genome-based Reference Package

The cellranger mkvdjref tool can be used to generate a custom reference package from a genome

sequence FASTA File and a gene annotation GTF.

Download cellranger:

Under: /share/data/software/cellranger/

curl -o cellranger-6.1.2.tar.gz

"https://cf.10xgenomics.com/releases/cell-exp/cellranger-

6.1.2.tar.gz?Expires=1648621580&Policy=eyJTdGF0ZW1lbnQiOlt7llJlc291cmNlljoiaHR0cHM6Ly9jZi4xM

HhnZW5vbWljcy5jb20vcmVsZWFzZXMvY2VsbC1leHAvY2VsbHJhbmdlci02LjEuMi50YXluZ3oiLCJDb25k

aXRpb24iOnsiRGF0ZUxlc3NUaGFuljp7lkFXUzpFcG9jaFRpbWUiOjE2NDg2MjE1ODB9fX1dfQ_&Signat

ure=RAATXYUck9peRrJIRrXGAA3Oz2mrK-j-WTpkFAMflWKdDAK3GFImviaj4XqsTx64GtCVhWvfds30E-

RGScYMVATqnyYm7yTkcA~ObrKTe2Tb8ob6DoVuWAaUNBxnO92e9ql2Ka7FbvRDgJjPWTyfwpMsYHEybBWM86epljqJijmftjTUXjakwR9NMdfYraOyYVpRci7yPTxqZZKSmh6Z2mCPVFHYlCufsfcGK4ldlO2obdTk6SKOWjvMdNWJ3D9
bTrOxJv8V6TRN~92Ge4n44PzT3zUFcAU2fiZSgPr5jW4axMWlfFm0wRQnGh1-2p9lpTCfZGpplGJqekWvrbw___&Key-Pair-Id=APKAI7S6A5RYOXBWRPDA"

Downloading Pre-built Indexes and a sample dataset for testing cellranger

https://support.10xgenomics.com/single-cell-geneexpression/software/pipelines/latest/using/tutorial ct

```
Download sample data and genome index as follows:
                                 https://cf.10xgenomics.com/samples/cell-
wget
exp/3.0.0/pbmc 1k v3/pbmc 1k v3 fastqs.tar
tar -xvf pbmc 1k v3 fastqs.tar
pbmc 1k v3 fastqs/
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L001 R2 001.fastq.gz
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L002 I1 001.fastq.gz
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L001 R1 001.fastq.gz
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L002 R1 001.fastq.gz
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L002 R2 001.fastq.gz
pbmc 1k v3 fastqs/pbmc 1k v3 S1 L001 I1 001.fastq.gz
       https://cf.10xgenomics.com/supp/cell-exp/refdata-cellranger-GRCh38-
wget
3.0.0.tar.gz
tar -zxvf refdata-cellranger-GRCh38-3.0.0.tar.gz
```

Create your own genome index using cellranger:

Under:/share/data/RNA Seg/reference genomes/10X genomics

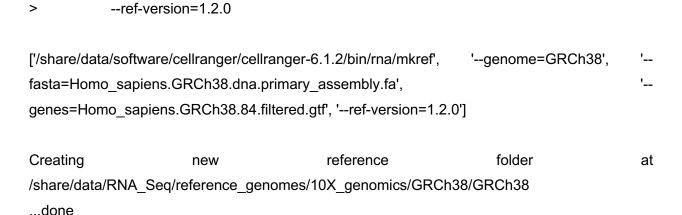
https://support.10xgenomics.com/single-cell-gene-expression/software/release-notes/build#grch38 1.2.0

```
wget
                                                           ftp://ftp.ensembl.org/pub/release-
84/fasta/homo sapiens/dna/Homo sapiens.GRCh38.dna.primary assembly.fa.gz
gunzip Homo sapiens.GRCh38.dna.primary assembly.fa.gz
wget ftp://ftp.ensembl.org/pub/release-84/gtf/homo sapiens/Homo sapiens.GRCh38.84.gtf.gz
gunzip Homo sapiens.GRCh38.84.gtf.gz
cellranger mkgtf Homo sapiens.GRCh38.84.gtf Homo sapiens.GRCh38.84.filtered.gtf \
          --attribute=gene_biotype:protein_coding \
          --attribute=gene biotype:lincRNA \
         --attribute=gene biotype:antisense \
          --attribute=gene biotype:IG LV gene \
         --attribute=gene_biotype:IG_V_gene \
          --attribute=gene biotype:IG V pseudogene \
          --attribute=gene biotype:IG D gene \
         --attribute=gene_biotype:IG_J_gene \
          --attribute=gene_biotype:IG_ J pseudogene \
          --attribute=gene_biotype:IG_C_gene \
          --attribute=gene biotype:IG C pseudogene \
          --attribute=gene biotype:TR V gene \
         --attribute=gene biotype:TR V pseudogene \
          --attribute=gene biotype:TR D gene \
          --attribute=gene biotype:TR J gene \
```

```
--attribute=gene biotype:TR J pseudogene \
--attribute=gene biotype:TR C gene
```

Run cellranger command:

```
cellranger mkref --genome=GRCh38 \
         --fasta=Homo_sapiens.GRCh38.dna.primary_assembly.fa \
         --genes=Homo sapiens.GRCh38.84.filtered.gtf \
         --ref-version=1.2.0
Output log from the above command:
cellranger mkref --genome=GRCh38 \
           --fasta=Homo sapiens.GRCh38.dna.primary assembly.fa \
           --genes=Homo sapiens.GRCh38.84.filtered.gtf \
>
```



Writing genome FASTA file into reference folder...

...done

>

Indexing genome FASTA file...

...done

Writing genes GTF file into reference folder...

...done

```
Generating STAR genome index (may take over 8 core hours for a 3Gb genome)...

Mar 29 15:41:13 ..... started STAR run

Mar 29 15:41:13 ... starting to generate Genome files

Mar 29 15:42:42 ... starting to sort Suffix Array. This may take a long time...
```

Mar 29 15:42:51 ... sorting Suffix Array chunks and saving them to disk...

Run cellranger using the test dataset:

Working commands:

```
cellranger count --id=run_count_1kpbmcs --fastqs=/share/data/RNA_Seq/10X/Raw_fastq/pbmc_1k_v3_fastqs/ --sample=pbmc_1k_v3 --transcriptome=/share/data/RNA_Seq/reference_genomes/10X_genomics/human/refdata-cellranger-GRCh38-3.0.0/
```

```
echo /share/data/software/cellranger/cellranger-6.1.2/bin/cellranger count --id=sample_name_5k --transcriptome=/share/data/RNA_Seq/reference_genomes/10X_genomics/human/refdata-cellranger-GRCh38-3.0.0/ --fastqs=/share/data/RNA_Seq/10X/Raw_fastq/pbmc_1k_v3_fastqs/ -expect-cells=5000 | qsub -S /bin/sh -V -N pbmc_cellranger -o /share/data/RNA_Seq/10X/Raw_fastq/pbmc_1k_v3_fastqs/ -e /share/data/RNA_Seq/10X/Raw_fastq/ -j y -cwd -I h_vmem=10G -pe smp 16
```

Id name is used to dump the output in the output directory: In this case, it is run_count_1kpbmcs. Use a meaningful name as per your project requirements.

```
[komalj@login refdata-cellranger-GRCh38-3.0.0]$ 11
total 12K
drwxr-xr-x 7 komalj binf 127 Mar 31 16:36 .
drwxrwxr-x 3 komalj binf
                           69 Apr
                                   1 09:37 ...
drwxr-xr-x 2 komalj binf
                           54 Nov
                                      2018 fasta
drwxr-xr-x 2 komalj binf
                           30 Nov 6
                                      2018 genes
drwxr-xr-x 2 komalj binf
                           33 Nov
                                   6
                                      2018 pickle
-rw-r--r-- 1 komalj binf
                          424 Nov
                                   6
                                      2018 reference.json
drwxrwxr-x 4 komalj binf 4.0K Mar 31 16:36 run count 1kpbmcs
```

Note: The command generates the output in the directory from where cellranger is run. Make sure to run cellranger in the project directory. Inside the script, cd into the directory.

Result interpretation of cellranger count (Useful links)

https://assets.ctfassets.net/an68im79xiti/163qWiQBTVi2YLbskJphQX/e90bb82151b1cdab 6d7e9b6c845e6130/CG000329 TechnicalNote InterpretingCellRangerWebSummaryFiles RevA.pdf

https://support.10xgenomics.com/single-cell-geneexpression/software/pipelines/latest/output/summary

https://bauercore.fas.harvard.edu/10x-chromium-system

https://kb.10xgenomics.com/hc/en-us/articles/360019665352-What-is-the-difference-between-TotalSeq-A-B-and-C-

TotalSeq data analysis:

https://www.biolegend.com/en-us/totalseq/mas

https://assets.ctfassets.net/an68im79xiti/7KPpQNrqSsAeSGYDr6Xeub/06fa19442477db83
6fde8928c89d5dda/CG000383 TechNote ChromiumNextGEMSingle Cell 3 v3.1 CellM
ultiplexing Rev A.pdf

Hashing using antibodies:

https://genomebiology.biomedcentral.com/articles/10.1186/s13059-018-1603-1