**1. Light-tissue interaction (20%)**

Download the data for the molar extinction coefficients of oxy- and deoxyhemoglobin as a function of wavelength from the Website (https://omlc.org/spectra/hemoglobin/), convert the molar extinction coefficients to absorption coefficients of oxy- and deoxyhemoglobin, and plot the two curves in MATLAB (or similar programming platform).

**2. Optical biosensor (20%)**

What is FRET? Explain how it works.

Ans:

FRET is a kind of dipole to dipole interaction between proteins that occurs at a very small distance, typically less than 10 nm. The application of this interaction is to reveal if proteins are interacting with each other. The basic idea is to attach(fuse) fluorophores to a protein. Now, we excite the first protein-fluorophore pair

**3. Microscopy basics (20%)**

Describe and compare polarized light microscopy, phase contrast microscopy and differential interference contrast microscopy (Basic principles, system configurations and advantages).

Ans:

Talk about Polarized light microsopy…

Phase contrast first works on the principle of what our eye can discern, which is differences in intensity. So light that is slowed down by the thickness of a specimen, also diffracted, lags at a quarter wavelength with the light that was not affected by the specimen. So, if the undisturbed light is sped up by a quarter wavelength there is a destructive interference at the eyepiece that would cause darkness that our eyes are sensitive to and would be able to see.

A system configuration of light

Some of the advantages of the phase contrasting method are:

Differential Interference contrast microscopy is an improvement on the original phase contrast by Zernike’s original idea….

System configuration

Advantages:

Comparisons between all of the above…..

**4. Fluorescence microscopy (20%)**

Describe and compare widefield fluorescence microscopy, confocal laser scanning microscopy and two-photon excitation microscopy (Basic principles, system configurations and advantages).

Ans:

Widefield fluorescence microscopy basically works on the principle of fluorescence which immediately emits light after photon absorption. A typical scenario will see a molecule, fluorophore, attach itself to a molecule or protein of interest and then respond to a stimulus within a prescribed localized biological region. A basic system configuration would see a lamp, lens, aperture, and diaphragm direct light at an excitation filter which would then pass thru typically a dichroic mirror to an objective lens and ultimately to the specimen. This excitation light would then stimulate the specimen which in turn would respond with a weaker emission light passing back thru our emission filter for capture and processing. Some of the advantages of a fluorescent microscope our: development of hundreds of fluorochromes, availability of different contrasting techniques (DIC as an example), highly sensitive and simple to implement.

The basic principle of Confocal laser scanning is that a laser scans across a sample to reconstruct. A system configuration would use a motorized mirror to scan across the sample. Also, a camera would not work as well for detection, so instead the use of photomultiplier tubes would be used to collect and amplify the light. After the PMT the signal would be digitized so that then the image can be reconstructed. An advantage of laser scanning is that you can get clear images in thin optical sections from thick samples. However, a disadvantage of confocal laser scanning is that it is slower than other optical techniques and requires deconvolution of the light

Two-photon microscopy is like confocal in that it focuses on a very small area, this differentiates both two-photon and confocal from wide-field fluorescence which has a wider focus area. So, while it focuses on a small area it is much brighter and sharper than confocal that has to use a pin hole as part of its’ focusing mechanism. The principle and system configuration of two-photon microscopy is that the fluorophore that has attached to the specimen that we wish to excite has to receive and absorb both photons to properly excite our system and have re-emission of another photon for detection. This is a great advantage of two-photon over normal one-photon fluorescence, the focal plane is smaller, so there is a sharper image, and since the area has to capture both photons, extraneous fluorophore emissions, or background signals are not permitted to excite because they do not receive both photons.

**5. Super-resolution microscopy (20%)**

What is the resolution limit of an optical imaging system? Explain why there’s such limit and describe at least one method to break the resolution limit.

Ans:

Part 1: The resolution limit of optical imaging systems is the Abbe diffraction limit which is about

200 nm.

Part 2: The reason that there is a diffraction limit is that the focus size of an object has a limit, this limit is the point spread function (PSF). Now, if the distance between PSF is greater than 200 nm then we can resolve individual PSFs, but if not then the two light objects appear as one and we can no longer resolve.

Now, there are two schools of thought on now to break this diffraction limit, one relies on shrinking the PSF, by pattern illumination. Here methods such as Stimulated emission depletion (STED) and RESOLFT microscopy use this idea of a smaller PSF to get sub-diffraction imaging.

Another method to get beyond the diffraction limit is by the method of stochastic switching of single molecules that is popular. Here the basic idea is to apply a widefield excitation of different fluorophores in the specimen. The switching of light will cause a weak activation in a small subset of all the possible fluorophores. Now their will be a stochastic behavior of which local fluorophores are activated and an accumulation of the images can be used to discern sub-diffraction structures, like organelles. A biomedical imaging technique that uses this method is called Stochastic Optical Reconstruction Microscopy or STORM.