**Solutions:**

**1.5% iodine potassium iodide (IKI) (100 mL)**

* 1.5 g potassium iodide
* 0.75 g iodine crystal
* 100 mL distilled water (or 70% EtOH)

**Protocol:**

1. Dissolve the potassium iodide with a magnetic stirrer in 80 mL of water (or 70% EtOH)
2. Shake iodine until dissolved and add the remaining water (or 70% EtOH)

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**0.2% phosphotungstic acid (PTA) (100 mL)**

* 20 mL 1% PTA
* 70 mL 100% EtOH

**Protocol:**

1. Add ingredients together and mix well

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**2% formalin (100 mL)**

* 20 mL 10% formalin
* 80 mL PBS (or distilled water)

**Protocol:**

1. Add ingredients together and mix well

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**70% EtOH (100 mL)**

* 70 mL 100% EtOH
* 30 mL distilled water

**Protocol:**

1. Add ingredients together and mix well

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**Sample preparation:**

For smaller test samples

1. Remove the uterus and choose one of the horns.
2. Using a sharp tool (scalpel is best), cut one or several segments from the horn. The size of the segments should only be of 1-2 mm.
3. Place samples in 2% formalin solution on a rocking table at 15 cpm. Due to the small size of the segments, there is no need to pin them in place. Leave samples on the table overnight (minimum 12 h).   
   For the remainder of the protocol, samples must be placed on a rocking table at 15 cpm and must always be fully submerged in solution.
4. Remove samples from 2% formalin solution and place in PBS. Wash the samples in PBS for 2 to 3 hours.
5. Rinse the samples a first time by placing them in 30% EtOH for 2 h.
6. Rinse the samples a second time by placing them in 50% EtOH for 2 h.
7. Rinse the samples a third time by placing them in 70% EtOH for 2 h.
8. Renew the 70% EtOH solution after the rinse and leave overnight (minimum 12 h).
9. Place the samples in stain and on the rocking table at 15 cpm. Leave the samples rocking for several days. 4 days provided good results with PTA stain but the longer the better.