R Tutorial-12

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You may click here to access the .qmd file.

In this issue, we

- manipulate the display of factors
- build tables (summaries which provide the distributions of factors)
- carry out inference on a single proportion

Factors

The term **factor** is used in the R community for a variable with finitely many values, particularly if those values split the data into separate categories—think class standing for students, make for automobiles, or number of axles for assessing vehicle tolls on the highway. As the last of these shows, a factor may be quantitative, yet used like a categorical variable.

Sometimes data frames contain variables which are factors, yet have codified the values as numbers—an example might be a variable sex whose values are 0 and 1. The wrist data frame in the fosdata package has variables handed_side and fracture_side, meant to indicate which hand is a patient's dominant hand, and which hand the patient fractured. Since the data frame (or tibble) has 47 columns/variables, we select out just these two:

```
dim(wrist)
```

[1] 105 47

```
wrist |> select(handed_side, fracture_side) |> head()
```

A tibble: 6 x 2 handed side fracture side <dbl> <dbl> 1 1 2 2 1 1 3 1 1 2 4 1 5 1 1 2 6 1

After running help(wrist), we learn that a 1 is used for right hand, 2 for left. We can alter our data to make this more explicit via

```
wristFeaturedVars = wrist |> select(handed_side, fracture_side) |>
    mutate(
    handed_side = factor(handed_side, labels=c("right","left")),
    fracture_side = factor(fracture_side, labels=c("right","left"))
)
head(wristFeaturedVars)
```

```
# A tibble: 6 x 2
  handed_side fracture_side
  <fct>
               <fct>
1 right
               left
2 right
               right
3 right
               right
4 right
               left
5 right
               right
6 right
               left
```

Tables

There are multiple commands (some made available by add-on packages) that can build frequency tables in R. I have eschewed the use of table() (the most obvious, and used regularly in the text), because it does not accept the formula notation varY ~ varX used with mosaic commands. Instead, I have often used tally() (mosaic). In what follows, I will use xtabs(). I suggest you try the next two commands both as written, and with tally() in place of xtabs(), to gain an appreciation for the change.

Running these commands, but drawing data from the original wrist data frame (where factor levels are still given as 1 and 2) makes it clear what advantage is afforded by the mutate() command above.

```
xtabs(~fracture_side, data=wrist)
xtabs(~ fracture_side + handed_side, data=wrist)  # I have blocked evaluation of these
```

One also may appreciate, particularly for the two-way table above, seeing row and column totals, achieved by piping the table to addmargins()

```
xtabs(~ fracture_side + handed_side, data=wristFeaturedVars) |> addmargins()
```

```
handed_side
fracture_side right left Sum
right 41 3 44
left 56 4 60
Sum 97 7 104
```

or obtaining relative frequencies (proportions) in each cell instead of frequencies

```
handTable = xtabs(~ fracture_side + handed_side, data=wristFeaturedVars)
handTable |> proportions()
```

```
handed_side
fracture_side right left
right 0.39423077 0.02884615
left 0.53846154 0.03846154
```

The code in this last chunk demonstrates that

- xtabs() employs raw data to build a summary, the distribution of factors
- the resulting summary, a table, can be stored and used for further processing

Note that proportions(), by default, produced proportions out of the total of 104 hand-fracture patients; that is, adding up all four proportions equals 1. Try out (on your own, as I have suppressed evaluation of these commands) these variants:

```
handTable |> proportions(margin=1)
handTable |> proportions(margin=2)
```

Building a table directly from the numbers for its cells

Many books, newspapers and the like display their data already summarized in table form. You may find you wish to build an R table directly containing the frequencies you see from an article. The website https://www.statology.org/two-way-table-in-r/ displays the following table:

	Baseball	Basketball	Football	Total
Male	13	15	20	48
Female	23	16	13	52
Total	36	31	33	100

The Statology site offers instructions, based on the matrix() command, for building this table. After some modification, I give those instructions here. Keep in mind that we are superceding the work of the xtabs() command and, in fact, we have not acquired an actual data set at all, which is what xtabs() would require.

```
myTable <- matrix(c(13, 15, 20, 23, 16, 13), ncol=3, byrow=TRUE)
rownames(myTable) = c("Male", "Female")
colnames(myTable) = c("Baseball", "Basketball", "Football")
myTable</pre>
```

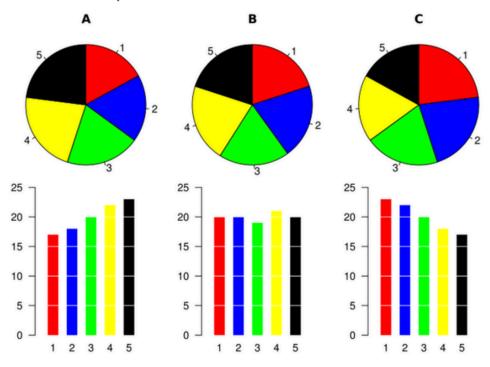
```
Baseball Basketball Football
Male 13 15 20
Female 23 16 13
```

```
# myTable |> addmargins() # Variant to throw in marginal totals
```

Visual displays

There are various ways to display factors/categorical data. One I tend to denigrate is a pie chart. Here is a graphic showing three different distributions of a categorical variable in both pie chart and bar graph form. There are subtle differences in the distributions for Data A, Data B and Data C, and the bar graphs do a better job eliciting these differences.

Don't use pie charts.



As for examples of generating bar graphs using gf_bar(), I recommend my R Tutorial Episode 9, beginning about half way through page 8 with the heading bar charts without and with slicing.

One-proportion inference

Any population proportion p represents a binary focus:

- "What proportion of American citizens are octegenarians?" focuses on American citizens and not so much their ages as whether that age is ≥ 80 (yes/no).
- "What proportion of people have blood type A-positive?" glosses over that there are other blood types, reducing to a yes/no-answer (binary) about being A-pos.

Some facts about proportions

1. A sample proportion is a ratio X/n where n is sample size, and $X = X_1 + X_2 + \cdots + X_n$ where each X_i is 0 or 1, depending on the success or not of a Bernoulli trial. Said another way,

$$\hat{p} = \frac{1}{n} \sum_{i} X_{i}$$

is the mean of a random sample.

2. If the X_i come as an iid sample, then the numerator $X \sim \text{Binom}(n,p)$, where p is the population proportion. In practice, the X_i are often not iid, but if n is relatively small in comparison to the size of the population, we can treat an SRS the same as iid. This enables us to test a null hypothesis

$$\mathbf{H}_0 : p = p_0$$

against a 1- or 2-sided alternative using binom.test() much as we did for the sign test; the only modification is we will use p_0 (the proposed null value), in the function call (the sign test had us using 1/2 for the null value). The command also offers confidence intervals for p in its output.

Example 1: A college football team has a returning kicker who, last season, made only 40% of field goals at distances 50 yards or more. They offer a freshman walk-on the chance to win the job, giving this newbie n = 10 attempts at 50+ yards. The young man gets the position if his test statistic is significant at the 5% level, when his tryout is viewed as an hypothesis test of

$$\mathbf{H}_0 \colon p = 0.4$$
 vs. $\mathbf{H}_a \colon p > 0.4$.

Suppose the player makes 6 of 10; here, X = 6 will serve as his test statistic. Using binom.test(), the relevant command is

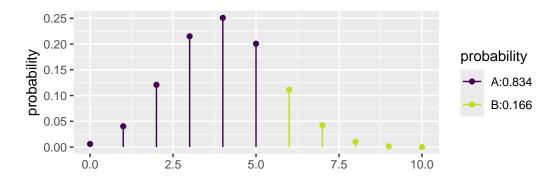
```
binom.test(6, 10, 0.4, alternative="greater")
```

```
data: 6 out of 10
number of successes = 6, number of trials = 10, p-value = 0.1662
alternative hypothesis: true probability of success is greater than 0.4
95 percent confidence interval:
```

The P-value is 0.1662, not statistically significant, which means the evidence is not strong enough (when $\alpha = 0.05$) to reject that this walk-on is successful only 40% of the time at this distance.

To see, visually, what is going on, look at the graph produced by the command

1 - xpbinom(5, 10, 0.4)



[1] 0.1662386

This shows the null distribution Binom(10,0.4), coloring results "5 or less" one way, and results "6 or more" another. In a world where this young man's rate of success at 50+ yards is truly 40%, you will see him make "6 or more" out of 10 about 16.6% of the time. Our P-value omits things in the left tail, because it was a 1-sided (right-tailed) alternative hypothesis. You should be able to see that the command

$$1 - pbinom(5, 10, 0.4)$$

is a direct way of obtaining the P-value.

If one wishes to have a 90% confidence interval for p, the proportion of kicks at 50-plus yards this newbie makes, binom.test() can be tweaked to report this, too. (The above command gave a 1-sided 95% confidence interval, by the way.) We obtain (evaluation has been suppressed) a (2-sided) 90% confidence interval using

I put 0.4 in for the null value, but confidence intervals are constructed without taking into account null values. So you should get the same 90% confidence interval even if you change 0.4 to 0.1, 0.5, or whatever in the command above.

Example 2: It is said that, in the United States, 35.7% of people are of blood type A-positive. Let's take as our research question: Does the proportion p of South Koreans with blood type A+match that in the U.S.? That is, we wish to test

$$\mathbf{H}_0$$
: $p = 0.357$ vs. \mathbf{H}_a : $p \neq 0.357$.

Now you collect a random sample (probably something akin to an SRS). Suppose 63 out of 211 Koreans sampled have blood type A+. We use the (default) two-sided alternative in our call to binom.test():

```
binom.test(63, 211, 0.357)
```

```
data: 63 out of 211
number of successes = 63, number of trials = 211, p-value = 0.08445
alternative hypothesis: true probability of success is not equal to 0.357
95 percent confidence interval:
0.2376893 0.3652278
sample estimates:
probability of success
0.2985782
```

The P-value, 0.08445, is not statistically significant at the 5% level, so we cannot reject the null hypothesis (at that level) that South Koreans also have blood type A+ at a rate of 35.7%. (Note that Wikipedia indicates 32% of South Koreans have A+ blood. Assuming Wikipedia is correct in its value of the population proportion p, then our sample led us into a Type II error.)

Important Note: The P-value reported by binom.test() is calculated as

```
referenceProbability = dbinom(63, 211, 0.357)
binomProbabilities = dbinom(0:211, 211, 0.357)
sum(binomProbabilities[binomProbabilities <= referenceProbability])</pre>
```

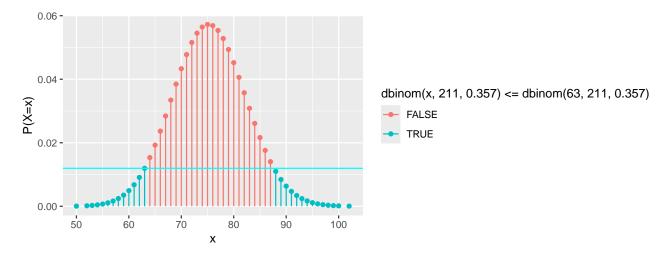
[1] 0.08445173

which is *not* the same as

```
2 * pbinom(63, 211, 0.357)
```

[1] 0.08648912

See the graphical depiction.



The reason they differ is that the Binom(211, 0.357) distribution is not truly symmetric, though it is not badly skewed. The more skewed your binomial (null) distribution, the greater this discrepancy will be.

- 3. Since \hat{p} is a mean, the Central Limit Theorem says that it will have an approximately normal distribution when n is large enough. Given that
- the count of successes $X \sim \text{Binom}(n, p)$ has expected value np and variance np(1-p), and
- $\hat{p} = \frac{X}{n}$,

it follows that \hat{p} has an approximate sampling distribution $\operatorname{Norm}\left(p,\sqrt{\frac{p(1-p)}{n}}\right)$, or equivalently that

$$\frac{\hat{p} - p}{\sqrt{\frac{p(1-p)}{n}}} \sim \text{Norm}(0, 1).$$

Using the normal approximation

Statistics books have, for many years, assumed people wanted to use a normal approximation for the sampling distribution of \hat{p} . This was largely because they could print a standard normal table, such as the one at https://math.arizona.edu/~jwatkins/normal-table.pdf (a substitute for pnorm()), and could not count on people having the computing power wrapped into pbinom(). They offered this rule of thumb, as a test for whether n is large enough to justify a normal approximation:

One can treat
$$\hat{p} \sim \text{Norm}\left(p, \sqrt{\frac{p(1-p)}{n}}\right)$$
 so long as $np \geq 10$ and $n(1-p) \geq 10$.

Let's see this in action.

Example 3:

Once again, suppose there are 63 of 211 South Koreans with Type A+ blood. We wish, as before, to test hypotheses

$$\mathbf{H}_0$$
: $p = 0.357$ vs. \mathbf{H}_a : $p \neq 0.357$.

We note that, assuming the null hypothesis is true,

$$np = (211)(0.357) = 75.327$$
 and $n(1-p) = (211)(0.643) = 135.673$,

so the rule-of-thumb for a normal approximation is justified. The reference distribution for \hat{p} is

$$Norm\left(0.357, \sqrt{\frac{(0.357)(0.643)}{211}}\right) = Norm(0.357, 0.03298362).$$

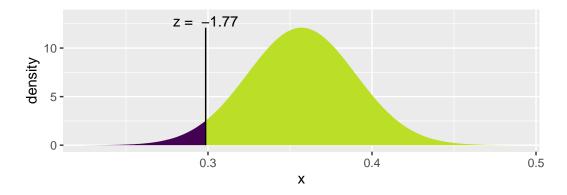
Taking our test statistic to be $\hat{p} = 63/211$ as our test statistic, the area in the right tail is

xpnorm(63/211, 0.357, 0.03298362)

If $X \sim N(0.357, 0.03298)$, then

$$P(X \le 0.2986) = P(Z \le -1.771) = 0.03826$$

$$P(X > 0.2986) = P(Z > -1.771) = 0.9617$$



[1] 0.03826067

As our alternative hypothesis is two-tailed, and normal distributions are always symmetric, we may take our P-value as twice this number:

2*pnorm(63/211, 0.357, 0.03298362)

[1] 0.07652134

It would be too strong to say the use of a normal approximation to the sampling distribution of \hat{p} has fallen out of favor; it is still taught in most intro statistics courses. For those who use it, there is an obvious improvement, called **continuity correction** (named so because we are replacing a discrete distribution with a continuous one), which leads to selecting the numerator for \hat{p} to be half-way between two integers. In our case, applying continuity correction yields the P-value

2*pnorm(63.5/211, 0.357, 0.03298362)

[1] 0.08924516

prop.test()

The prop.test() command is as easy to use as binom.test(). It

- automates the use of a normal approximation, yet
- provides a warning alongside the results when the rule of thumb is not met, and
- does continuity correction by default.

```
prop.test(63, 211, p=0.357)
```

1-sample proportions test with continuity correction

Like binom.test(), prop.test() produces a confidence interval (at the 95% level, by default). Since prop.test() is based on a normal approximation, its upper/lower bounds are not a match to those produced by binom.test(). The results of binom.test() (both confidence intervals and P-values) are more accurate than those of prop.test().