

Title

Author

Date

1 Introduction

In the latest few years, machine learning and deep learning have revolutionized the natural language processing field (NLP)[9]. At the same time some fundamental ideas developed in NLP have been successfully applied to another type of language, the biological one: DNA, RNA and amino acid sequences bringing to excellent results even in the complex task of protein structure prediction[8, 11]. One of these fundamental ideas is word embedding [12] because it transforms words into points in space, therefore easy to process. It is well known that sequence similarity do not always correspond to functional similarity [10].

2 Methods

In this section we describe the pipeline used to analyze the embeddings. As shown in Table 1, the input length is different between the models as well as the output produced. We want to address the following problems: 1) compare different methods to join together the amino acid-specific context-

ual representations in order to have a representation for the whole chunk and subsequently join together the representations of the chunks in order to have a representation for the whole protein; 2) find out if these representations reflect known properties of the proteins.

2.1 Represent protein sequences as continuous vectors

short description of the main characteristics of each embedder

2.1.1 prose

learning with structural supervision

2.1.2 alphafold

- is a deep learning system not a language model

- predict residue-residue distances from sequence families and fold proteins based on the predicted distance constraints

- rely on large datasets of protein sequences that are similar enough to be aligned with

high confidence but contain enough divergence to confidently infer statistical couplings between positions

- unable to learn patterns across large-scale databases of possibly unrelated proteins and have limited ability to draw on the increasing structure and function information available

2.1.3 SeqVec

To build their model Heinzinger et al.[5] adapted the standar ELMo configuration [13] to work with protein sequences modifying the number of tokens and the unroll steps. It is composed by 1 CharCNN and 2 LSTM-Layers. Given a protein sequence of arbitrary length it returns 3072 features for each residue derived by concatenating the outputs of the three layers of ELMo, each describing a token with a vector of length 1024. In order to obtain a smaller representation for each amminoacid i computed the mean of the three layers (as also suggested in the official repository). Given the architecture of the ELMo, these representation are contextual-dependent.

2.2 Combining the (contextual) representations

derived by concatenating the outputs of the three layers of ELMo, each describing a token with a vector of length 1024 We tried four methods to join togheter the amminoacid embeddings in order to produce a fixed size embedding for the chunk: average, maximum, sum and principal component analysis (PCA).

The same operator used to combine the amminoacid embeddings is also used to combine the embeddings of the chunks of the sequence.

Note on the contextual embedding: seqvec: contextual

2.3 Comparison with known informations

Given a set of embeddings of sequences we want to analyze their distribution in the embedding space comparing it with both the distance matrix produced during the multiple sequence alignment with Clustal Omega [14] and higher level annotations as Gene Ontology [3, 1], UniProtKB Keywords and NCBI Taxonomy [4].

2.3.1 Similarity between distance matrices

In order to compare two distance matrices we performed an agglomerative clustering on both, the resulting tree is then cut at each level obtaining flat partitions of all possibles number of clusters. We performed a pairwise comparison of the partitions having the same number of clusters using the adjusted rand score [6]. The mean of these score, starting from two clusters up to $\#elements - 1$ clusters is called mean adjusted rand score (MARS). We compared different distance metrics as well as different methods to perform the hierarchical clustering.

2.3.2 Enrichment analysis

The alignment distance matrix provide an evolutionary related distance between sequences [15], we also wanted to analyze the properties of the embeddings at an higher level. The Gene Ontology (GO) describes our knowledge of the sequence with respect to: molecular function, cellular component and biological process; there are also more specific controlled vocabulary as the UniProt Keywords and hierarchical classifications specific for sequences as the NCBI Taxonomy.

Whatever they are the sets of words to describe the sequences in our datasets, we want to build a distance between sequences among them. Given A and B the sets of annotations of two sequences we computed the distance in two possible ways:

$$d1 = \frac{2 * |A \cap B|}{|A| + |B|}$$
$$d2 = \max\{\frac{|A \cap B|}{|A|}, \frac{|A \cap B|}{|B|}\}$$

Both of them vary between 0 and 1, however $d1$ goes to 1 only when the two sets are equals while $d2$ goes to 1 also when one set is a subset of the other. After calculating one of these distance between all possible pair in the dataset we end up with a similarity matrix, that can be easily transformed in a distance matrix that is possible to compare with the distance matrix derived from the distance between the embeddings using the MARS as described in subsection 2.3.1.

Name	input length (chunk)	embedding dimension
embedding reproduction (rep)[16]	64	64 per chunk
seqvec [5]	1024	1024 per amminoacid
dnabert [7]	512	768 per chunk
prose [2]	512	100 per amino acid
alphafold [8]	1024	384 per ammino acid
evolutionary scale modeling (esm2) [11]	1024	1280 per ammino acid

Table 1: Embedders used in the experiments, their maximum input length and the dimension of the embedding produced.

Name	description	number of sequences	type	avg length
hemoglobin	hemoglobin for various organisms	761	amminoacids	142
mouse	mouse proteome	974	amminoacids	516
bacterium	bacterium proteome	259	amminoacids	427
covid19	covid19 complete genome	77	nucleotides	29831
meningitis	meningitis complete genome	68	nucleotides	2240049

Table 2: Datasets used in the experiments.

3 Results

3.1 Phylogenetic

3.2 Enrichment

3.3 Projections?

3.4 Classification?

3.5 Pointwise representation
similarity?

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