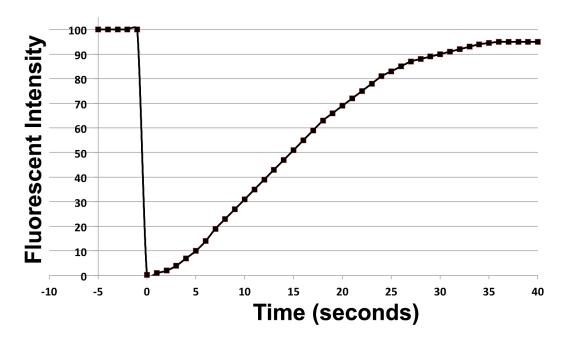
Problem Set #1	Name	
7.06 - Spring 2015	Section	
Question 1 You are using fluores	scence microscopy to image the localization of prot	eins within a cell.
	antages of localizing a protein using a GFP functions is that protein?	sion compared to ar
B. Name 2 advar	ntages of localizing a protein using an antibod GFP fusion?	y against that proteir
However, you protein antiboo	rotein localization using both the GFP fusion and t find that the GFP fusion shows diffuse localizat dies display microtubule localization. You advis the GFP fusion is incorrect. Why might this be	ion, whereas the anti- sor suggests that the
protein. Using to use fluoreso Your advisor s when there are	mize a fluorescent fusion protein to test the loca this fusion, you are able to image viral particles vence microscopy to count the number of virus pasuggests that it will be much harder to correctly one a large number of viral particles compared to the case? Explain your reasoning.	within a cell. You wan articles infecting a cell determine this numbe

## Question 2

You next want to use fluorescent recovery after photobleaching (FRAP) to analyze the dynamics of a plasma membrane-localized Type I protein using a GFP fusion (with the GFP at the C-terminus). You use a laser to bleach a defined area of the plasma membrane. You measure the intensity of the GFP fluorescence in the bleached area before and after photobleaching. Here is the graph that you obtain:



- A. Based on this graph, what is the half-life for recovery of this protein?
- B. What molecular event(s) must occur for the tagged protein to observe fluorescent recovery after photobleaching for this experiment?

You next test the recovery of the GFP after several different perturbations. In each case, indicate whether the turnover would be slower, the same, or faster. Note that only **significant** changes would be detected. Briefly explain your reasoning.

C. Your cells have reduced levels of Cholesterol. The presence of Cholesterol decreases membrane fluidity.

D. You test the turnover of your protein in a cell that is expressing half the normal concentration of the protein.
E. You grow cells on coverslips coated with a protein that binds to your protein and test turnover in regions of the membrane associated with the coverslip.
You next generate a similar fusion for your protein using a variant of GFP termed photoactivatable GFP. This protein is invisible (non-fluorescent) until it is activated by a laser pulse. Using this new fusion protein, you conduct an experiment in which you activate the photoactivatable GFP in a similar sized defined region of the plasma membrane and measure the GFP fluorescence in that region before and afterwards.  F. Draw a graph of what this would look like:
G. What molecular event(s) must occur for the tagged protein to observe the change in fluorescence indicated in your graph?

For the original GFP fusion, you next conduct photobleaching on other areas of the cell. In each case, the indicated site corresponds to both the area you are photobleaching and the area where you are measuring the fluorescence. What molecular events must occur to see fluorescent recovery in each of these cases?

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11.	TOU DICACITUIE	: CHUIC DIASHIA	IIICIIIDIAIIC	tuut not miema	1 16010113 01 66111.

I. You bleach the entire cell.

## Question 3.

**A.** You have identified some proteins that localize to the plasma membrane. Your advisor wants you to demonstrate that these are *stable* membrane proteins, not just peripherally associated with membranes. At this stage, you have no knowledge of their amino acid sequences. What biochemical perturbation would be required to separate a stable protein from the membrane?

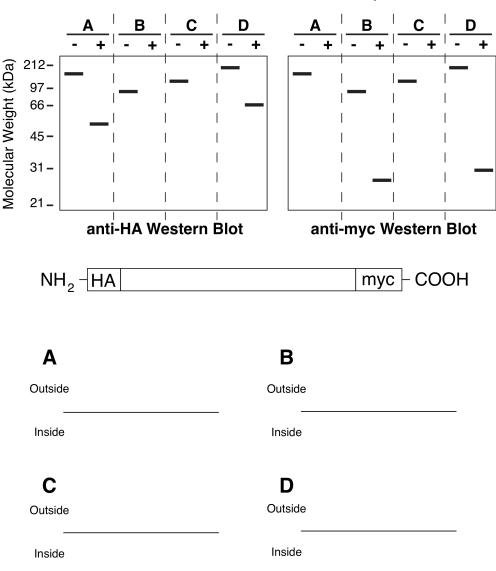
**B.** For each of the following, indicate the primary mechanism by which each of these molecules enters into a cell. Be as specific as possible.

·	Mechanism of Entry
H <sub>2</sub> O	
Cholesterol	
Water	
Gleevac (a small molecule kinase inhibitor	
used to treat cancer)	
Glucose	
A 15 kDa protein	

## Question 4

You are working on four different plasma membrane proteins (A, B, C, and D). To understand the function of these proteins, you decide to test their orientation in the membrane. In each case, you generate a version of the cDNA where you add an HA epitope tag at the N-terminus and a myc epitope tag at the C-terminus (assume that these tags do not disrupt any signal sequences). You then introduce these cDNAs individually into cells. In each case, you split the cells into two populations. For the first, you generate a protein sample from the complete cell ("-" samples). For the second, you treat the intact cells with a protease (Proteinase K) which will digest all accessible protein ("+" samples). In each case, you run the samples on an SDS-PAGE gel, and probe to detect for either the HA or myc tag by Western blot

**A.** Based on the Western blots shown below, draw the most likely orientation of each protein.



**B.** In each case, based on the topology, indicate its corresponding class of membrane protein next to the diagram.