Instructions for Obtaining a Seed Sequence for Organelle Assembly

We will be using NOVOPlasty (Dierckxsens et al., 2017) to assemble organelle genomes for our species! As we talked about at the end of Week 8 Lab, NOVOPlasty is a seed-extend based assembler. This means that the software starts with a "seed" sequence and attempts to find overlapping reads that can extend the sequence in both directions. The seed is just a string of DNA nucleotides that belong to the organelle genome of a species that is closely related to your sample. The seed sequence does not become part of the assembly, it is just used as a starting point to retrieve the first sequencing read from the raw sequencing read data set. Remember that our sequencing read files will include reads from both nuclear and organellar genomes. We don't want to include reads from the nuclear genome in our organelle assemblies!

The protocol for obtaining seed sequences will differ slightly depending on whether your species is a plant or an animal. For animals, you will only be assembling the mitochondria genome. For plants, you must assemble the chloroplast genome before you can assemble the mitochondrial genome.

Animals

Look for the sequence of a conserved mitochondrial gene in a species closely related to your species! A good place to start would be the cytochrome c oxidase subunit I (COI) gene. See the instructions below to learn how to do this.

Plants

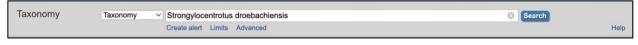
Look for a chloroplast-specific gene in a closely related species. In plants, the chloroplast and mitochondria have many similar sequences due to intergenomic transfer. It is important to find a sequence that is only found in the chloroplast, and not in the mitochondria. A good place to start would be the RuBisCO enzyme. Data on the RuBisCO enzyme for your species might be under several different synonymous names:

- Ribulose-1,5-bisphosphate carboxylase-oxygenase
- Rubisco
- RuBPCase
- RuBPco

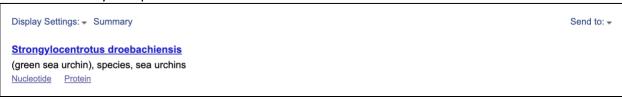
How to Find a Seed Sequence

I will use *Strongylocentrotus droebachiensis*, the green sea urchin, to demonstrate how to follow this protocol.

1. Search for your species on NCBI in the "Taxonomy" database



2. Click on your species



3. Look at the info under "Entrez records". Entrez is a database for molecular sequence data. Is there an assembly available (a row titled "Assembly" with a number greater than zero next to it)? If so, skip to step 4. In the example below, there is not an assembly available for *Strongylocentrotus droebachiensis*.

Look at the "Lineage" information. We are going to go up the lineage of your organism to hopefully find the most closely related species with an assembled genome. Click the last link in the lineage list (this most likely corresponds to a genus).

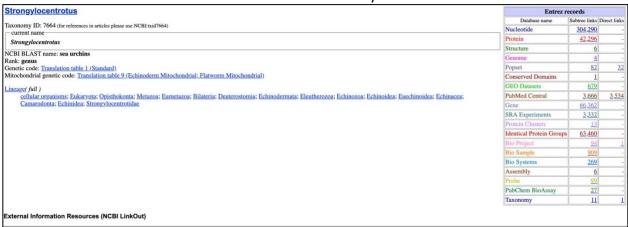


After clicking the link, you will see all of the organisms that fall under that lineage. Click on the genus name one more time. In this case, it is *Strongylocentrotus*.



4. Look at the info under "Entrez" records. Is there an assembly available? If the answer is no, repeat steps 3 and 4 until you get to a taxonomic group that has an assembly available.

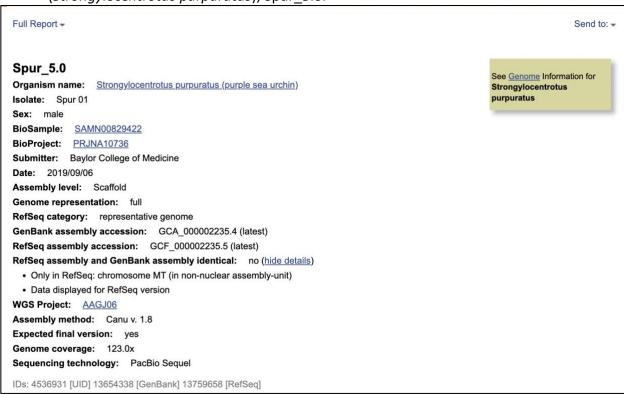
In this case, the Entrez records indicate that there is at least one assembly available! Click on the underlined number next to "Assembly."



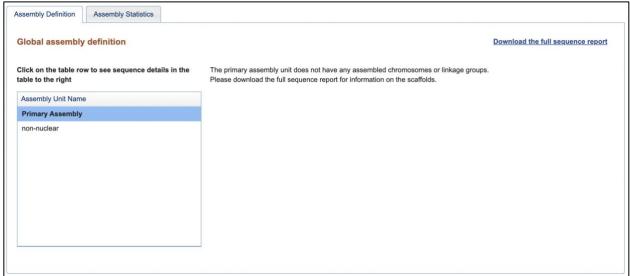
5. A list of published genome assemblies will appear. Click on one of the listed assemblies. If many options appear, note that you can filter using options on the left side bar.



Shown below is the information for the latest genome assembly of the purple sea urchin (*Strongylocentrotus purpuratus*), Spur_5.0.



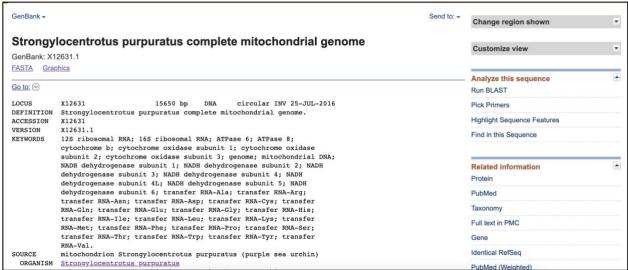
6. Scroll down to the bottom of the page to the section titled: "Global assembly definition"



7. Click the "non-nuclear option." For animals, look to see if there is a mitochondria option. For plants, look to see if there is a chloroplast option. Click the underlined link under "GenBank ID." If there are not any choices, go back to step 5 and choose a different assembly. If there are no other assemblies available, repeat steps 3 and 4, following the lineage further back to check for assemblies in distantly related taxonomic groups.



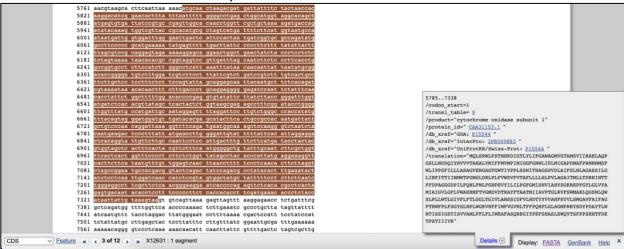
8. Shown below is the information for the assembled *Strongylocentrotus purpuratus* mitochondrial genome. If you scroll down, you will see all the genes in the assembly listed. For animals, try to find a COI gene. For plants, try to find a RuBisCO gene. You might have to skim through the genes or keyword search different terms. For example, the COI gene could be under "cytochrome c" or "COI."



9. Once you've found the gene you want, click on the underlined "gene" or "CDS" link. CDS stands for coding sequence. Clicking this link will show you where your sequence is in the genome assembly.

```
CDS
    5785..7338
    /codon_start=1
    /transl_table=9
    /product="cytochrome oxidase subunit 1"
    /protein id="CAA31153.1"
    /db xref="GOA:P15544"
    /db xref="InterPro: IPR000883"
    /db xref="UniProtKB/Swiss-Prot:P15544"
    /translation="MQLSRWLFSTNHKDIGTLYLIFGAWAGMVGTAMSVIIRAELAQP
    GSLLNDDQIYNVVVTAHALVMIFFMVMPIMIGGFGNWLIPLMIGAPDMAFPRMNNMSF
    WLIPPSFILLLASAGVENGAGTGWTIYPPLSSNITHAGSSVDLAIFSLHLAGASSILG
    LINFITTIINMRTPGMSLDRLPLFVWSVFVTAFLLLLSLPVLAGAITMLLTDRNINTT
    FFDPAGGGDPILFOHLFWLFGHPEVYILILPGFGMISHVIAHYSGKREPFGYLGLVYA
    MIAIGVLGFLVWAHHMFTVGMDVDTRAYFTAATMIIAVPTGLKVFSWMAKLQGSNLQW
    SLPLLWTLGIVFLFTLGGLTGIVLANSSIDFVLHDTYYVVAHFHYVLSMGAVFAIFAG
    FTHWFPLFSGYSLHPLWGKVHFFIMFVGVNLTFFPOHFLGLAGMPRRYSDYPDAYTLW
    NTISSIGSTISVVAMLFFLFLIWEAFASQREGITPEFSHASLEWQYTSFPPSHHTFDE
    TPSTIIIVK"
```

10. On the bottom right, click the "Display: FASTA" option. This will take you to a new screen with the selected sequence in FASTA format.



FASTA -

Strongylocentrotus purpuratus complete mitochondrial genome

GenBank: X12631.1 GenBank Graphics

>X12631.1:5785-7338 Strongylocentrotus purpuratus complete mitochondrial genome ${\tt CCTGAGCTGGCATGGTAGGCACAGCTATGAGTGTGATTATCCGTGCCGAGTTGGCACAACCTGGTTCGCT}$ GCTAAAAGATGACCAGATATACAAAGTGGTCGTTACCGCACATGCGCTAGTCATGATTTTCTTCATGGTA ATGCCAATAATGATTGGTGGATTTGGGAATTGACTCATTCCACTAATGATCGGTGCGCCAGATATGGCCT TCCCCGCATGAAAAATATGAGTTTTTGACTTATTCCCCCTTCTTTTATATTACTTTTAGCGTCCGCAGG AGTAGAAAAAGGAGCAGGAACTGGCTGAACTATCTACCCTCCTCTCTAGTAAAATAACACACGCCGGT AGGTCCGTTGATTTAGCAATCTTCTCCCTTCACCTGGCCGGTGCCTCTTCCATCTTGGGCCTCATTAAAT TTATAACAACAATTATTAATATGCGGACACCGGGGATGTCTTTGGATCGTCTTTATTCGTCTGATC CGTCTTTGTCACTGCCTTCTTGCTCCTTCTTCTCCAGTATTAGCAGGAGCAATTACAATGCTTCTC ${\tt ACAGATCGTAAAATAAACACAACTTTCTTTGACCCTGCAGGAGGGGGAGATCCAATTCTATTTCAACACC}$ TATTCTGGCTTTTTGGACACCCCGAGGTGTATATTCTTATCTTACCGGGATTTGGTATGATCTCACACGT ACTTCACTGCCGCCACAATGATTATTGCTGTCCCAACAGGATTAAAGGTTTTCAGATGAATGGCAAAGCT CCAAGGGTCTAATCTACAATGAAGACTCCCTTTATTATGAACCTTGGGGATTGTATTTTATTCACATTA GGAGGACTCACAGGTATTGTTCTTGCCAATTCCTCCATTGACTTTGTTCTTCATGATACCTACTACGTGG GTCAACTTAACCTTTTTCCCTCAACACTTCTTAGGTCTAGCCGGAATGCCACGACGGTACTCAGACTATC CAGACGCCTATACACTTTGAAATACTATCTCCTCAATTGGATCAACCATCTCCGTAGTGGCTATGCTATT TCACTAGAGTGACAATACACCTCCTTTCCCCCTTCTCACCACACCTTCGATGAAACACCCTCTACCATAA TTATTGTAAAGTAA

11. Copy and paste the sequence into a text editor (either Sublime or Atom, do not use TextEdit). Make sure to include the header that beings with the ">" symbol.



12. Save this file as seed mito.fasta or seed chloro.fasta. Remember where you saved it!

