ELSEVIER

Contents lists available at ScienceDirect

### Drug Metabolism and Pharmacokinetics

journal homepage: http://www.journals.elsevier.com/drug-metabolism-andpharmacokinetics



Regular article

# Prediction of inter-individual variability on the pharmacokinetics of CYP1A2 substrates in non-smoking healthy volunteers



Kenta Haraya <sup>a, \*</sup>, Motohiro Kato <sup>b</sup>, Koji Chiba <sup>c, d</sup>, Yuichi Sugiyama <sup>d</sup>

- <sup>a</sup> Chugai Pharmabody Research Pte. Ltd., Singapore
- <sup>b</sup> Chugai Pharmaceutical Co., Ltd., Shizuoka, Japan
- <sup>c</sup> Laboratory of Clinical Pharmacology, Yokohama University of Pharmacy, Yokohama, Japan
- <sup>d</sup> Sugiyama Laboratory, RIKEN Innovation Center, Research Cluster for Innovation, RIKEN, Yokohama, Japan

#### ARTICLE INFO

Article history:
Received 27 December 2015
Received in revised form
10 March 2016
Accepted 15 March 2016
Available online 24 March 2016

Keywords: CYP1A2 Inter-individual variability Monte Carlo simulation Human pharmacokinetics Drug development

#### ABSTRACT

The activity of CYP1A2, a major drug-metabolizing enzyme, is known to be affected by various environmental factors. Our study aimed to predict inter-individual variability of AUC/Dose of CYP1A2 substrates in non-smoking healthy volunteers using the Monte Carlo simulation. Inter-individual variability in hepatic intrinsic clearance of CYP1A2 substrates (CL<sub>int,h,1A2</sub>) was estimated using dispersion model based on the inter-individual variability (N = 96) of the AUC of caffeine, a major CYP1A2 substrate. The estimated coefficient of variation (CV) of CL<sub>int,h,1A2</sub> was 55%, similar to previously reported CL<sub>int,h,2D6</sub> (60%) but larger than CL<sub>int,h,3A4</sub> (33%). Then, this estimated CV was validated by predicting the CVs of AUC/Dose of tizanidine and phenacetin, which are mainly metabolized by CYP1A2 and have negligible renal clearance. As a result, reported CVs were successfully predicted within 2.5–97.5 percentile range of predicted values. Moreover, CVs for AUC/Dose of the CYP1A2 substrates theophylline and lidocaine, which are affected by other CYPs and renal clearance, were also successfully predicted. The interindividual variability of AUC/Dose of CYP1A2 substrates was successfully predicted using 55% CV for CL<sub>int,h,1A2</sub>, and the results, along with those reported by our group for other CYPs, support the prediction of inter-individual variability of pharmacokinetics in the clinical setting.

© 2016 The Japanese Society for the Study of Xenobiotics. Published by Elsevier Ltd. All rights reserved.

### 1. Introduction

In drug development, it is important to predict the clearance of a clinical candidate drug in human, and various methodologies for the prediction of clearance have been reported [1–5]. Generally, a mean value for clearance is predicted, and subsequently the accuracy for the prediction is evaluated by predicting the results of one clinical trial. However, in actuality, clearance in human has interindividual variability, which, if large, may result in unexpected side effects and failure of the treatment. If the inter-individual variability in the pharmacokinetics of a clinical candidate drug in clinical trials can be predicted beforehand, it will greatly contribute to selecting the optimal clinical candidate.

Previously, our group proposed a methodology using the Monte Carlo simulation to predict the inter-individual variability of the AUC for CYP3A4, CYP2C19, and CYP2D6 substrates and the urine

netic parameters that had been collected from literature. Kato et al. successfully predicted the inter-individual variability of the AUC for CYP3A4 substrates by setting the CV of CL<sub>int,h</sub> of CYP3A4 at 33%, after comparing the reported CV of CYP3A4 content in human liver microsome and the reported CV of AUC of CYP3A4 substrates [6]. Ito et al. estimated the CV of CL<sub>int,h</sub> of CYP2D6 substrates for an extensive metabolizer (EM) and an intermediate metabolizer (IM) as 60% from the variability in urine MR of CYP2D6 substrates [7], and successfully predicted the inter-individual variability in urine MR of CYP2D6 substrates from the estimated 60% CV. Also, Chiba et al. estimated the CV of CL<sub>int,h</sub> of CYP2D6 substrates for each CYP2D6 polymorphism from the variability in urine MR of CYP2D6 substrates and successfully predicted the inter-individual variability of AUC of CYP2D6 substrates for each race [8].

metabolic ratio (MR) for CYP2D6 substrates from the mean value and the variability of physiological parameters and pharmacoki-

In this study, we focused on the inter-individual variability of the AUC of CYP1A2 substrates. CYP1A2 is a major hepatic CYP [9] and its activity has been known to be affected by various polymorphisms and environmental factors [10–13]. Therefore, large inter-individual

E-mail address: haraya.kenta@chugai-pharm.co.jp (K. Haraya).

<sup>\*</sup> Corresponding author.

variability in the  $CL_{int,h}$  of CYP1A2 substrates can be expected. In particular, smoking is a typical environmental factor that induces the activity of CYP1A2 [14–17] and was reported to reduce the AUC of well-known CYP1A2 substrates caffeine and tizanidine by approximately 50% [18,19]. However, in most cases, detailed information on smoking behavior (frequency of smoking, duration of smoking, etc.) was not described in the literature, so the effect of smoking could not be incorporated into the inter-individual variability of  $CL_{int,h}$  of CYP1A2 substrates. Therefore, in this study the inter-individual variability in the AUC of CYP1A2 substrates was predicted in non-smokers.

In this study, caffeine, tizanidine, and phenacetin were selected as typical CYP1A2 substrates. First, the CV of CL<sub>int,h</sub> of CYP1A2 substrates was estimated from the AUC/Dose (AUC/D) data for caffeine, to which the first-pass effect contributes slightly after oral

Uchimura et al.'s method [20]. Only AUC/D data after a single oral administration in healthy non-smokers were used. Since the ratio of renal clearance in systemic clearance for tizanidine, phenacetin, and lidocaine was below 1%, it was assumed that renal clearance was negligible. PubMed (http://www.ncbi.nlm.nih.gov/pubmed/) was used to search the literature.

When multiple AUC/D data were available for a substrate, the overall mean value and SD were estimated from each mean value and SD using the following equations.

Weighted mean (WM) was calculated from

$$WM = \frac{\sum_{i=1}^{n} (N_i \cdot m_i)}{\sum_{i=1}^{n} N_i}$$

Overall SD was calculated from

$$SD = \sqrt{\frac{\sum_{i=1}^{n} \left[ \left\{ (SD_i)^2 \times N_i \cdot (N_i - 1) + (N_i \cdot m_i)^2 \right\} \middle/ N_i \right] - WM^2 \bullet \sum_{i=1}^{n} N_i}{\sum_{i=1}^{n} N_i - 1}}$$

administration. Then, using the estimated CV, the inter-individual variability in the AUC/D of tizanidine and phenacetin was predicted. Moreover, the accuracy of the predicted value was confirmed by predicting the inter-individual variability of the AUC/D of the CYP1A2 substrates theophylline, which is affected by renal clearance, and lidocaine, which is affected by other CYPs.

### 2. Materials and method

### 2.1. Selection of CYP1A2 substrates

Caffeine, tizanidine, and phenacetin were selected as major CYP1A2 substrates that are mainly metabolized in liver and have more than 0.9 of contribution of CYP1A2 in liver metabolism ( $f_{\rm m}$  CYP1A2). Theophylline and lidocaine were selected as CYP1A2 substrates that have contribution of renal clearance and other CYPs.

### 2.2. Data collection

The mean value and variability (standard deviation (SD), standard error (SE), coefficient of variation (CV), confidence interval (CI)) of physiological parameters (liver volume, hepatic blood flow rate, body weight, and albumin concentration) were collected from literature. The mean value and variability of pharmacokinetic parameters (AUC/D, plasma unbound fraction ( $f_p$ ), renal clearance (CL<sub>r</sub>), ratio of blood-to-plasma concentration ( $R_B$ ), and excretion fraction of drug in urine ( $A_e$ )) of CYP1A2 substrates were also collected from literature (Tables 1 and 2). Since the  $R_b$  of tizanidine has not been reported in literature, it was estimated from  $f_p$  using

where  $N_i$  is the number of subjects,  $m_i$  is the mean value in the ith study, and  $SD_i$  is the SD value in the ith study.

### 2.3. Estimation of $f_{m \text{ CYP1A2}}$ for CYP1A2 substrates

The  $f_{m\ CYP1A2}$  was estimated using furafylline [21,22], a known selective CYP1A2 inhibitor, in the following way. In the case of substrate which has a small contribution of renal clearance, more than 0.9 of  $f_{m\ CYP1A2}$ , and information of AUC changed by furafylline in human,  $f_{m\ CYP1A2}$  was estimated from following equation.

$$f_{m\;\text{CYP1A2}} = 1 - \frac{\text{AUC}}{\text{AUC}_f}$$

where AUC<sub>f</sub> is the AUC of the CYP1A2 substrate dosed concomitantly with furafylline. When an increase above 900% in the AUC of the CYP1A2 substrate was observed for concomitant dosing, it was assumed that the activity of CYP1A2 was completely inhibited by furafylline. In fact, because reports gave an increase of over 900% in the AUCs of caffeine and phenacetin dosed concomitantly with furafylline [23,24], the values for f<sub>m CYP1A2</sub> of caffeine and phenacetin were estimated as 1. For substrates to which the contribution of renal clearance is large or for which concomitant dosing with furafylline increased the AUC either by less than 900% or there was no report of a change in AUC,  $f_{m\ CYP1A2}$  was estimated from in vitro CYP1A2 inhibition data or recombinant CYP metabolism data. The f<sub>m CYP1A2</sub> values of tizanidine, theophylline, and lidocaine were estimated in this way. Since approximately 90% of tizanidine metabolism was inhibited by furafylline in an in vitro study [25], the  $f_{m \text{ CYP1A2}}$  of tizanidine was estimated as 0.9. When  $f_{m \text{ CYP1A2}}$  was 0.9

 Table 1

 Pharmacokinetic parameters of CYP1A2 substrates used in this study.

Substrate	$f_p$	$R_{B}$	$f_{m, 1A2}$	$CL_r$ mean (ml/min)	CL <sub>r</sub> CV (%)	A <sub>e</sub> (%)	Reference
Caffeine	0.8	1	1	1.85	36.3	2	[20,23,60]
Tizanidine	0.7	0.85 <sup>a</sup>	1	_b	_b	<1	[25,61,62]
Phenacetin	0.47	1.01	1	_b	_b	<1	[24,61,63,64]
Theophylline	0.44	0.82	0.65	8.8	26.1	17	[20,26,65]
Lidocaine	0.296	0.84	0.68	_b	_b	<1	[27,63,66]

a Calculated using the reported method.

b Urinary excretion < 1%.

**Table 2** Weighted AUC/D and CV values obtained from literature of CYP1A2 substrates.

Substrate	Number of groups	Number of subjects	AUC/D mean (min/l)	AUC/D CV (%)	CL <sub>int,h</sub> (ml/min/kg)	Reference
Caffeine	9	96	13.7	53.9	1.33	[18,67-72]
Tizanidine	10	167	0.105	86.0	85.0	[19,73-80]
Phenacetin	4	61	0.679	80.2	40.6	[81-83]
Theophylline	13	158	18.7	32.8	1.42	[65,84-94]
Lidocaine	6	45	0.362	52.1	88.3	[95-100]

and over,  $f_{m\ CYP1A2}$  was assumed as 1 in this study. The  $f_{m\ CYP1A2}$  and  $f_{m\ other\ CYPs}$  for the ophylline [26] and lidocaine [27] were estimated using normalized intrinsic clearance (CL<sub>int</sub>) from the reported expression data for each CYP in human liver microsomes (HLM) in an *in vitro* recombinant CYPs metabolism study [28].

### 2.4. Estimation of variability of CLint.h.1A2 from caffeine data

The CV of  $CL_{int,h,1A2}$  was estimated by a Monte Carlo simulation using data on the WM of AUC/D and CV of caffeine, which were collected from literature (Fig. 1). The dispersion model was used for the Monte Carlo simulation. One thousand sets of the mean value and CV of the AUC/D were generated using each pharmacokinetic parameter of caffeine and an arbitrary CV of  $CL_{int,h,1A2}$ . Since AUC/D of caffeine of 96 subjects was collected from literature, 96 of AUC/D were generated in 1 set of simulation. The optimal CV of  $CL_{int,h,1A2}$  was estimated by finding comparable CV in simulation with weighted mean of CV in literatures.

AUC/D after oral administration was calculated from the following equation.

$$AUC/Dose = \frac{F_a \cdot F_g \cdot F_h}{CL_h + CL_r}$$

where  $F_a$  and  $F_g$  were assumed to be 1 and to have no interindividual variability.

F<sub>h</sub> was calculated using the dispersion model from the following equation.

$$Fh = \frac{4a}{(1+a)^2 \cdot \exp\{(a-1)/2/D_N\} - (1-a)^2 \cdot \exp\{-(a+1)/2/D_N\}}$$

$$a = (1 + 4R_N \cdot D_N)^{1/2}$$

$$R_N = f_B \cdot \frac{CL_{\text{int},h}}{Qh}$$

where  $f_B$  is the blood unbound fraction,  $Q_h$  is the hepatic blood flow rate, and  $D_N$  is the dispersion number.  $D_N$  was assumed to be 0.17 [29,30].

CL<sub>h</sub> was calculated from the following equation.

$$F_h = 1 - \frac{CL_h}{Q_h}$$

The plasma unbound fraction  $(f_p)$  was calculated from the equation  $f_p = 1/(1 + nPt/K_d)$ , where n is the number of binding sites, Pt is the albumin concentration, and  $K_d$  is the dissociation constant, and n and  $K_d$  were assumed to have no inter-individual variability. The simulation used reported values for  $Q_h$  (1.22 mL/min/mL liver, CV: 12.9%) [31], liver volume (19.5 mL/kg, CV: 11.4%) [31], albumin concentration (42.4 g/L, CV: 10.3%) [32–35], and body weight (Asian: 66.2 kg, CV: 12.4%, Caucasian: 78.8 kg, CV: 11.7%) [36].

The Monte Carlo simulation was performed in a virtual population of 96 subjects and generated 1000 sets of AUC/D and CV, and

a log-normal distribution of  $CL_{int,h}$  was assumed. The simulation was conducted according to Kato et al.'s method and used the RAND function of Microsoft Excel 2007 to generate uniform random numbers ranging from 0 to 1. Two normal random numbers  $(Z_i$  and  $Z_{i+1})$  were generated from the following equations.

$$Z_i = (-2*lnu_i)^{0.5}*cos(2*\pi*u_{i+1})$$

$$Z_{i+1} = (-2*lnu_i)^{0.5}*sin(2*\pi*u_{i+1})$$

No correlation among parameters was assumed.

Above simulation was programmed using Visual Basic for Applications (VBA) of Microsoft Excel 2007, and the Monte Carlo simulation was conducted by generating an arbitrary distribution of each parameter from generated random numbers. All simulation was conducted under assumption of central limit theorem.

### 2.5. Simulation of AUC/D of CYP1A2 substrates

A Monte Carlo simulation was conducted to generate 1000 sets of the mean value and CV of AUC/D for each CYP1A2 substrate using the dispersion model in a virtual population with the number of subjects which were collected from literature. When the information on R<sub>B</sub> was not available in literature, it was estimated from f<sub>D</sub> using the reported method [20]. Since Kato et al. reported the CV of CLint.h.3A4 to be 33%, this value was used in this study. Since the CV of CL<sub>int.h.2E1</sub> has not been reported, it was estimated from data on chlorzoxazone, which is a CYP2E1 specific substrate, using the same method as that used to estimate the CV of CL<sub>int,h,1A2</sub>. Compared with other CYPs for which our group has predicted the inter-individual variability, CYP2E1 is a minor isotype, which means there are only a few substrates that are mainly catalyzed by CYP2E1 and data sources for them were limited. Therefore, in this study, the CV of CLint,h,2E1 was only estimated from the data of chlorzoxazone. Chlorzoxazone has been used in vitro and in vivo to prove the presence of CYP2E1 [37,38]. In addition, the urinary excretion of unchanged drug for chlorzoxazone is below 1%, its clearance is low, and its  $f_{m\ CYP2E1}$  is 1. Therefore, chlorzoxazone can be considered as the optimal substrate to estimate the CV of  $CL_{int,h,2E1}$ . The  $f_p$ ,  $R_B$ , and  $f_m$  CYP2E1 were set as 0.04, 0.51, and 1 respectively based on information from literature [39,40]. Since its urinary excretion of unchanged drug is below 1%, renal clearance was set as 0 [41].

The log-normal distribution of  $CL_{int,h}$  was assumed. The log-transferred mean  $(\mu)$  and SD  $(\sigma)$  were calculated from the arithmetic mean and CV collected from literatures using the following equations.

$$\mu = ln(\textit{arithmetric mean}) - \frac{\sigma^2}{2}$$

$$\sigma = \sqrt{\ln\!\left(\left(\frac{CV}{100}\right)^2 + 1\right)}$$

To confirm the effect of changes in  $CL_{int,h}$  on the AUC/D and its CV, the AUC/D and CV were simulated using the pharmacokinetic parameters of caffeine under different  $CL_{int,h}$  values (Fig. 4A). It was assumed that 100% of drug was metabolized in the liver, so the simulation was conducted by removing renal clearance from the pharmacokinetic parameters of caffeine. The Monte Carlo simulation generated 1000 sets in a virtual population of 96 subjects using 33% and 55% CV for  $CL_{int,h}$ .

### 3. Results

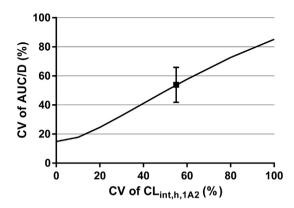
## 3.1. Estimation of pharmacokinetic parameters for CYP1A2 substrates

The pharmacokinetic parameters of 5 CYP1A2 substrates estimated from literature are listed in Table 1. The  $f_{m\ CYP1A2}$  of caffeine, tizanidine, and phenacetin was estimated as 0.96, 0.90, and 0.99 respectively by assuming that the activity of CYP1A2 was completely inhibited by furafylline. If AUC of CYP1A2 substrates increased 10 fold by concomitant dosing with furafylline,  $f_{m\ CYP1A2}$  was calculated as 0.90. However, if the concentration of furafylline was insufficient to inhibit the CYP1A2 activity completely, it can be considered that  $f_{m\ CYP1A2}$  is above 0.90; therefore, the values for  $f_{m\ CYP1A2}$  of caffeine, tizanidine, and phenacetin were set as 1.

The  $f_{m\ CYP1A2}$ ,  $f_{m\ CYP3A4}$ , and  $f_{m\ CYP2E1}$  of theophylline were estimated as 0.65, 0.20, and 0.15, respectively, and the  $f_{m\ CYP1A2}$  and  $f_{m\ CYP3A4}$  of lidocaine were estimated as 0.68 and 0.32, respectively.

### 3.2. Estimation of variability of CL<sub>int.h.1A2</sub>

The WM and CV were calculated from information in the literature on the AUC/D and its variability for caffeine in healthy non-smokers (Table 2). The CV of the WM of AUC/D was calculated as



**Fig. 2.** Estimation of CV of CL<sub>int,h,1A2</sub> from caffeine data set. Closed diamond indicates observed CV of AUC/D of caffeine. Solid curve indicates the simulation curve of relationship between CV of CL<sub>int,h</sub> and CV of AUC/D with 2.5–97.5 percentile range. 1000 simulations were conducted in a virtual population with 96 subjects.

53.9% from the AUC/D and its CV values in 9 populations with a total of 96 subjects. Then, the CV of AUC/D was simulated using an arbitrary CV for  $CL_{int,h,1A2}$  ranging from 0% to 100%. As a result, the optimal CV of  $CL_{int,h,1A2}$  for the CV of the WM of AUC/D that had been collected from literature was estimated as 55% (Fig. 2).

### 3.3. Prediction of variability of AUC/D for major CYP1A2 substrates

To confirm the accuracy of the predicted value for CV of  $CL_{in-t,h,1A2}$  that was estimated from caffeine data, the CVs of AUC/D for the CYP1A2 substrates were predicted using the estimated CV of  $CL_{int,h,1A2}$  of 55%. First, AUC/D and its CV for caffeine in each report were compared with the results simulated by setting the CV of

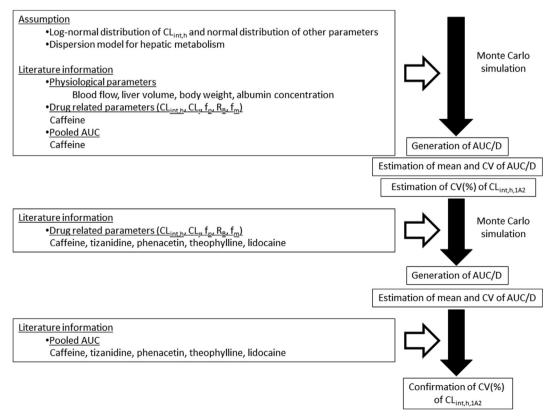
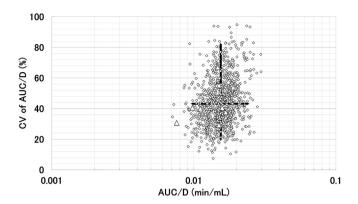


Fig. 1. Analytical scheme for estimation of the inter-individual variability.



**Fig. 3.** Simulation of AUC variability of caffeine after oral administration. Open triangles indicate AUC/D and CV from literature. Open diamonds indicate simulated AUC/D and CV. Open circle indicates the mean AUC/D and CV of simulation with 2.5–97.5 percentile range. The range in the number of subjects in literature was from 6 to 16.

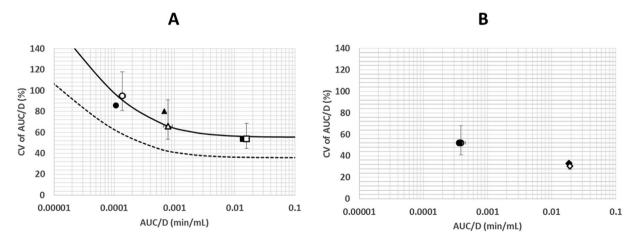
CLint.h.1A2 at 55%. One thousand sets of AUC/D of caffeine were simulated in a virtual population of 6 subjects, which is the minimum number of subjects observed in the collected literature. As a result, most reported values (8 out of 9) were predicted within the 2.5-97.5 percentile range of simulated results (Fig. 3). Then, tizanidine and phenacetin were selected as major CYP1A2 substrates with f<sub>m CYP1A2</sub> over 0.9. The data for healthy non-smokers in 10 populations with a total of 167 subjects for tizanidine, and in 4 populations with a total of 61 subjects for phenacetin, were collected from literature. The CVs of the WM of AUC/D for tizanidine and phenacetin that were collected from literature were 86% and 80.2%, respectively (Table 2). AUC/D and its CV were simulated using a CV of  $CL_{int,h,1A2}$  of 55% while changing  $CL_{int,h,1A2}$ . As a result, the simulated results (caffeine: CV; 53.6%, 2.5-97.5 percentile range; 44.5-68.5%, tizanidine: CV; 94.8%, 2.5-97.5 percentile range; 81.0-117.8%, phenacetin: CV; 66.1%, 2.5-97.5 percentile range; 53.4–91.3%) were comparable with the CVs of the WM of reported AUC/D for caffeine, tizanidine, and phenacetin (Fig. 4A). The simulation successfully predicted the reported variability in the AUC/D for CYP1A2 substrates. Moreover, results simulated using a 33% CV of CL<sub>int.h.1A2</sub> were clearly underestimated (Fig. 4A). Hence,

the study supports the ability of this simulation to clearly distinguish the difference between the variability values of CYP1A2 and 3A4. Additionally, smaller AUC/D showed larger CV in simulation, and the same trend was observed in the reported AUC/D and CV for caffeine, tizanidine, and phenacetin. Moreover, the CV of the WM for each reported AUC/D for caffeine, tizanidine, and phenacetin was predicted by the Monte Carlo simulation using the CV of CL<sub>int,h,1A2</sub> at 55%. As a result, the CV of the WM of each reported AUC/D for caffeine, tizanidine, and phenacetin was successfully predicted within the 2.5–97.5 percentile range of simulation (Fig. 4A). As shown in Fig. 4A, high hepatic first-pass effect of high clearance drugs will make the inter-individual variability of the AUC/D for these drugs also high. In this study, inter-individual variability was accurately predicted for a wide range of low to high clearance drugs.

# 3.4. Prediction of variability of AUC/D for CYP1A2 substrates with contribution of other CYPs and renal clearance

The confirmation of prediction accuracy was conducted using theophylline and lidocaine which are CYP1A2 substrates with contribution of renal clearance and/or other CYPs. The data in 13 populations with a total of 158 subjects for theophylline, and in 6 populations with a total of 45 subjects for lidocaine, were collected from the literature on healthy non-smokers. CVs of the WM of AUC/D for theophylline and lidocaine collected from literature were 32.8% and 52.1%, respectively (Table 2).

To estimate the CV of CL<sub>int,h,2E1</sub>, the AUC/D and its CV for chlorzoxazone, a CYP2E1 specific substrate, were collected from literature. The data in 5 populations with a total of 80 subjects for chlorzoxazone were collected from literature on healthy volunteers [39,42–45], and the WM of AUC/D and its CV were calculated as 3.93 min/l and 45.6%, respectively. Then, the CV of AUC/D was simulated using an arbitrary CV for CL<sub>int,h,2E1</sub> ranging from 0% to 100%. As a result, the optimal CV of CL<sub>int,h,2E1</sub> for the CV of the WM of AUC/D that had been collected from literature was estimated as 39% (Supplement Fig. 1). This CV was moderate compared with our reported CVs for other CYPs. Since various polymorphisms of CYP2E1 have been reported, it is possible that polymorphisms of CYP2E1 contribute to the CV of CL<sub>int,h,2E1</sub> [46,47]. Theophylline is



**Fig. 4.** Simulation of AUC variability of CYP1A2 substrates after oral administration. Graphs show (A) the major CYP1A2 substrates caffeine (squares), phenacetin (triangles), and tizanidine (circles) and (B) CYP1A2 substrates with contribution of other elimination, theophylline (diamonds) and lidocaine (circles). Closed symbols indicate observed AUC/D and CV. Open symbols indicate simulated AUC/D variability of 55% for CL<sub>int,h,1A2</sub> and 33% for CL<sub>int,h,3A4</sub> (previously reported). The contribution of elimination on theophylline is 83% in hepatic metabolism and 17% in urine excretion. The contribution ratio of CYPs in hepatic metabolism on theophylline is 65% by CYP1A2, 20% by CYP3A4, and 15% by CYP2E1. The contribution of elimination on lidocaine is 100% in hepatic metabolism, of which 68% is by CYP1A2 and 32% by CYP3A4.

partially metabolized by CYP2E1 in the liver. Therefore, a CV of  $CL_{int,h,2E1}$  of 39% was used for the predictions for theophylline.

The CV of the WM of each reported AUC/D for theophylline and lidocaine was predicted by the Monte Carlo simulation using a CV of CL<sub>int,h,1A2</sub> of 55%. As a result, the CV of WM of each reported AUC/D for theophylline and lidocaine was successfully predicted within the 2.5–97.5 percentile range of simulation (theophylline: CV; 30.6%, 2.5–97.5 percentile range; 27.5–34.4%, lidocaine: CV; 52.3%, 2.5–97.5 percentile range; 41.0–67.9%) (Fig. 4B).

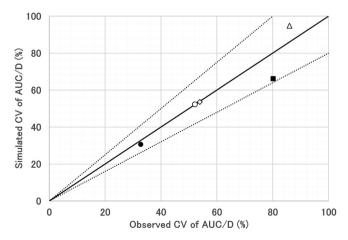
## 3.5. Prediction accuracy of variability of AUC/D for CYP1A2 substrates

To confirm the prediction accuracy, the reported CV of WM of AUC/D for 5 CYP1A2 substrates was compared to the simulated results (Fig. 5). Reported CV of WM of AUC/D for all substrates was successfully predicted within 80–120% of the simulated results. Hence, our methodology was confirmed to have good prediction accuracy.

### 4. Discussion

The purpose of this study is to evaluate a method for predicting the inter-individual variability of AUC/D for CYP1A2 substrates by setting the CV of  $\text{CL}_{\text{int,h,1A2}}$  based on *in vivo* AUC/D data. In this study, since it was assumed that the amount of CYP enzyme was proportional to  $\text{CL}_{\text{int}}$ , the inter-individual variability of the amount of CYP enzyme was determined by that of  $\text{CL}_{\text{int}}$ . Therefore, the interindividual variability of  $\text{CL}_{\text{int}}$  could be transferred from that of AUC and reflected the inter-individual variability of the amount of CYP enzyme. Once the inter-individual variability of the amount of each CYP enzyme is determined, the inter-individual variability of  $\text{CL}_{\text{int}}$  can be applied to  $\text{CL}_{\text{int}}$  of other substrates.

In this study, the top-down approach based on *in vivo* data was applied to predict the inter-individual variability of the AUC/D for CYP1A2 substrates. In contrast, the commercially available software SimCyp uses the bottom-up approach based on *in vitro* data to predict the inter-individual variability of pharmacokinetics [48]. The inter-individual variability in SimCyp was estimated using *in vitro* data on the amount of enzymes in HLM, and the estimated CV for CYP1A2 was 67%, which was larger than our result. Since previous reports show that the stability of HLM in *in vitro* studies



**Fig. 5.** Prediction accuracy of inter-individual variability of AUC/D in CYP1A2 substrates after oral administration. Observed data of caffeine (open diamond), phenacetin (closed square), tizanidine (open triangle), theophylline (closed circle), and lidocaine (open circle) are shown. The solid line indicates 100% accuracy and dotted lines indicate 120% and 80% accuracy.

can be strongly affected by freezing and thawing, as well as by storage conditions, we selected the top-down approach based on *in vivo* data [49,50].

CYP1A2, which is a major CYP in human liver that accounts for approximately 13% of total CYP activity in that organ [9], is involved in the metabolism of more than 100 drugs in human and is one of the key metabolizing enzymes involved in drug elimination [51]. Since over 15-fold [52] and 40-fold [53] inter-individual variability of CYP1A2 mRNA in human liver has been observed, there is a possibility that this inter-individual variability has a great impact on the drugs metabolized by CYP1A2. As described above, it has been reported that various genetic polymorphisms and environmental factors affect the activity of CYP1A2.

According to the NCBI dbSNP database (http://www.ncbi.nlm. nih.gov), over 200 single nucleotide polymorphisms (SNPs) have been found in the CYP1A2 gene. Moreover, various variant alleles were also found. The variant alleles reported to have over 10% prevalence are \*1A, \*1B, \*1C, \*1D, \*1F, \*1L, and \*1V. Although various genetic polymorphisms of CYP1A2 have been reported [15,16,54,55], there has been no report to indicate that genetic polymorphism has a clear impact on the pharmacokinetics of CYP1A2 substrates in healthy non-smokers; therefore, the effect of genetic polymorphism was not incorporated into this study.

It has also been reported that broccoli consumption [56], panfried meat consumption [57], and menstrual cycle phases [58] are environmental factors that possibly affect the activity of CYP1A2. As described above, since smoking has been reported as the most influential factor on the activity of CYP1A2, inter-individual variability in non-smokers was investigated in this study.

Since the  $f_{m\ CYP1A2}$  of caffeine was almost 1 and caffeine showed a low hepatic first-pass effect, caffeine was selected to estimate the CV of  $CL_{int,h,1A2}$ . Because drugs with a low hepatic first-pass effect minimize the variability of that effect, they can be considered to estimate the CV of  $CL_{int,h,1A2}$  more accurately. The inter-individual variability of other CYPs that have been reported by our group are 33% CV for CYP3A4 [6], 60% CV for CYP2D6 in EM [7], 66% CV for CYP2C19 in EM [59], and 18.1% CV for CYP2C9 in EM. The interindividual variability of CYP1A2 (55%) was similar to that of CYP2D6 but otherwise relatively high among CYPs. As described above, since it has been reported that various genetic polymorphisms and environmental factors affect the activity of CYP1A2, it cannot be denied that these factors have a composite effect on the inter-individual variability of CYP1A2 in this study.

As shown in Fig. 4A, high clearance drugs showed high interindividual variability of the AUC/D due to hepatic first-pass effect after oral administration. Kato et al. investigated the inter-individual variability of the AUC/D for CYP3A4 substrates after both intravenous and oral administration [6]. In the case of intravenous administration, since the hepatic clearance of a high clearance drug can be limited by hepatic blood flow, smaller inter-variability of the AUC/D for high clearance drugs was observed. In contrast, the interindividual variability of the AUC/D after oral administration showed similar trend with our result in this study. These results suggest that the dosing route could be an important information to consider the inter-individual variability of the AUC/D.

As described above, high clearance drug shows high intervariability of the AUC/D after oral administration. However, although lidocaine showed smaller AUC/D than phenacetin, lidocaine showed smaller inter-variability of the AUC/D than phenacetin. This could be due to the contribution of the inter-variability of CL<sub>int,h,3A4</sub> (33%) for lidocaine metabolism. Since the inter-variability of CL<sub>int,h,3A4</sub> (33%) was smaller than that of CL<sub>int,h,1A2</sub> (55%), the inter-variability of overall CL<sub>int,h</sub> for lidocaine should be smaller than 55%. Moreover, since no correlation was assumed between CL<sub>int,h,3A4</sub> and CL<sub>int,h,1A2</sub> in the Monte Carlo simulation, the standard

deviation of the sum of generated  $CL_{int,h,3A4}$  and  $CL_{int,h,1A2}$  should be smaller than the sum of each standard deviation. This is according to propagation of error. Therefore, the drugs with multiple metabolic pathways could show smaller inter-variability of AUC/D. The selection of the candidate drugs with multiple metabolic pathways can potentially reduce the inter-individual variability of AUC/D.

It is desirable that a clinical candidate drug has small interindividual variability of pharmacokinetics for stable efficacy and safety. Since relatively high CV of CL<sub>int,h</sub> was estimated for CYP1A2, CYP2D6, and CYP2C19, drugs that are metabolized by these CYPs will require a wide-ranging efficacy concentration and a broad safety margin in clinical development. Also, as shown in Fig. 4A, since high clearance drugs are expected to have large interindividual variability of AUC/D caused by the hepatic first-pass effect, selecting candidate drugs with low clearance is one approach to reducing the inter-individual variability of AUC/D in human.

#### 5. Conclusions

In this study, we propose a methodology that can predict the inter-individual variability of AUC for CYP1A2 substrates in human. Since the estimated 55% CV of CL<sub>int,h,1A2</sub> is relatively high among CYPs, it can be considered that prediction of inter-individual variability of AUC for CYP1A2 substrates in human will strongly contribute to improving the success rate and shortening the length of clinical trials. In the future, inter-individual variability of pharmacokinetics in human will be comprehensively predictable by this method of estimating the inter-individual variability, for not only for CYPs, but also for transporters in the liver and kidney, and not only in healthy volunteers but also in children, the elderly, and in populations with a specific disorder. Also, this methodology of predicting inter-individual variability of pharmacokinetics can be expanded to predict the inter-individual variability of efficacy and safety for various drugs. Moreover, we consider it will contribute to efficient clinical development and reduction of unexpected side effect.

### **Conflict of interest**

The authors have no conflict of interest.

### Acknowledgments

This work was fully supported by Chugai Pharmaceutical Co., Ltd.

### Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.dmpk.2016.03.003.

### References

- Sanoh S, Horiguchi A, Sugihara K, Kotake Y, Tayama Y, Ohshita H, et al. Prediction of in vivo hepatic clearance and half-life of drug candidates in human using chimeric mice with humanized liver. Drug Metab Dispos 2012;40:322–8.
- [2] Zanelli U, Caradonna NP, Hallifax D, Turlizzi E, Houston JB. Comparison of cryopreserved HepaRG cells with cryopreserved human hepatocytes for prediction of clearance for 26 drugs. Drug Metab Dispos 2012;40:104–10.
- [3] Obach RS. Prediction of human clearance of twenty-nine drugs from hepatic microsomal intrinsic clearance data: an examination of in vitro half-life approach and nonspecific binding to microsomes. Drug Metab Dispos 1999;27:1350–9.
- [4] Hallifax D, Foster JA, Houston JB. Prediction of human metabolic clearance from in vitro systems: retrospective analysis and prospective view. Pharm Res 2010;27:2150–61.

- [5] Ogawa K, Kato M, Houjo T, Ishigai M. A new approach to predicting human hepatic clearance of CYP3A4 substrates using monkey pharmacokinetic data. Xenobiotica 2012;43:468–78.
- [6] Kato M, Chiba K, Ito T, Koue T, Sugiyama Y. Prediction of interindividual variability in pharmacokinetics for CYP3A4 substrates in humans. Drug Metab Pharmacokinet 2010;25:367–78.
- [7] Ito T, Kato M, Chiba K, Okazaki O, Sugiyama Y. Estimation of the interindividual variability of cytochrome 2D6 activity from urinary metabolic ratios in the literature. Drug Metab Pharmacokinet 2010:25:243–53.
- [8] Chiba K, Kato M, Ito T, Suwa T, Sugiyama Y. Inter-individual variability of in vivo CYP2D6 activity in different genotypes. Drug Metab Pharmacokinet 2012;27:405—13.
- [9] Shimada T, Yamazaki H, Mimura M, Inui Y, Guengerich FP. Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. J Pharmacol Exp Ther 1994;270: 414–23
- [10] Ingelman-Sundberg M, Sim SC, Gomez A, Rodriguez-Antona C. Influence of cytochrome P450 polymorphisms on drug therapies: pharmacogenetic, pharmacoepigenetic and clinical aspects. Pharmacol Ther 2007;116:496–526.
- [11] Gunes A, Dahl ML. Variation in CYP1A2 activity and its clinical implications: influence of environmental factors and genetic polymorphisms. Pharmacogenomics 2008;9:625–37.
- [12] Zhou SF, Di YM, Chan E, Du YM, Chow VD, Xue CC, et al. Clinical pharmacogenetics and potential application in personalized medicine. Curr Drug Metab 2008:9-738–84
- [13] Zhou SF, Liu JP, Chowbay B. Polymorphism of human cytochrome P450 enzymes and its clinical impact. Drug Metab Rev 2009;41:89–295.
- [14] Djordjevic N, Ghotbi R, Bertilsson L, Jankovic S, Aklillu E. Induction of CYP1A2 by heavy coffee consumption in Serbs and Swedes. Eur J Clin Pharmacol 2008;64:381–5
- [15] Ghotbi R, Christensen M, Roh HK, Ingelman-Sundberg M, Aklillu E, Bertilsson L. Comparisons of CYP1A2 genetic polymorphisms, enzyme activity and the genotype—phenotype relationship in Swedes and Koreans. Eur J Clin Pharmacol 2007;63:537—46.
- [16] Gunes A, Ozbey G, Vural EH, Uluoglu C, Scordo MG, Zengil H, et al. Influence of genetic polymorphisms, smoking, gender and age on CYP1A2 activity in a Turkish population. Pharmacogenomics 2009:10:769—78.
- [17] Petersen MS, Halling J, Damkier P, Nielsen F, Grandjean P, Weihe P, et al. Caffeine N3-demethylation (CYP1A2) in a population with an increased exposure to polychlorinated biphenyls. Eur J Clin Pharmacol 2006;62: 1041–8
- [18] Carrillo JA, Christensen M, Ramos SI, Alm C, Dahl ML, Benitez J, et al. Evaluation of caffeine as an in vivo probe for CYP1A2 using measurements in plasma, saliva, and urine. Ther Drug Monit 2000;22:409—17.
- [19] Backman JT, Schroder MT, Neuvonen PJ. Effects of gender and moderate smoking on the pharmacokinetics and effects of the CYP1A2 substrate tizanidine. Eur J Clin Pharmacol 2008;64:17–24.
- [20] Uchimura T, Kato M, Saito T, Kinoshita H. Prediction of human blood-to-plasma drug concentration ratio. Biopharm Drug Dispos 2010;31:286–97.
- [21] Eagling VA, Tjia JF, Back DJ. Differential selectivity of cytochrome P450 inhibitors against probe substrates in human and rat liver microsomes. Br J Clin Pharmacol 1998;45:107—14.
- [22] Sesardic D, Boobis AR, Murray BP, Murray S, Segura J, de la Torre R, et al. Furafylline is a potent and selective inhibitor of cytochrome P450IA2 in man. Br J Clin Pharmacol 1990;29:651–63.
- [23] Tarrus E, Cami J, Roberts DJ, Spickett RG, Celdran E, Segura J. Accumulation of caffeine in healthy volunteers treated with furafylline. Br J Clin Pharmacol 1987;23:9–18.
- [24] Boobis AR, Lynch AM, Murray S, de la Torre R, Solans A, Farre M, et al. CYP1A2-catalyzed conversion of dietary heterocyclic amines to their proximate carcinogens is their major route of metabolism in humans. Cancer Res 1994;54:89–94.
- [25] Granfors MT, Backman JT, Laitila J, Neuvonen PJ. Tizanidine is mainly metabolized by cytochrome p450 1A2 in vitro. Br J Clin Pharmacol 2004;57: 349–53.
- [26] Gu L, Gonzalez FJ, Kalow W, Tang BK. Biotransformation of caffeine, paraxanthine, theobromine and theophylline by cDNA-expressed human CYP1A2 and CYP2E1. Pharmacogenetics 1992;2:73—7.
- [27] Wang JS, Backman JT, Taavitsainen P, Neuvonen PJ, Kivisto KT. Involvement of CYP1A2 and CYP3A4 in lidocaine N-deethylation and 3-hydroxylation in humans. Drug Metab Dispos 2000;28:959–65.
- [28] Kawakami H, Ohtsuki S, Kamiie J, Suzuki T, Abe T, Terasaki T. Simultaneous absolute quantification of 11 cytochrome P450 isoforms in human liver microsomes by liquid chromatography tandem mass spectrometry with in silico target peptide selection. J Pharm Sci 2011;100:341–52.
- [29] Roberts MS, Rowland M. Correlation between in-vitro microsomal enzyme activity and whole organ hepatic elimination kinetics: analysis with a dispersion model. J Pharm Pharmacol 1986;38:177–81.
- [30] Naritomi Y, Terashita S, Kagayama A, Sugiyama Y. Utility of hepatocytes in predicting drug metabolism: comparison of hepatic intrinsic clearance in rats and humans in vivo and in vitro. Drug Metab Dispos 2003;31:580–8.
- [31] Wynne HA, Cope LH, Mutch E, Rawlins MD, Woodhouse KW, James OF. The effect of age upon liver volume and apparent liver blood flow in healthy man. Hepatology 1989;9:297–301.

- [32] Kerremans AL, Gribnau FW, Tan Y, van Ginneken CA. Pharmacokinetic and pharmacodynamic studies of tienilic acid in healthy volunteers. Eur J Clin Pharmacol 1982;22:515–21.
- [33] Gorski JC, Hall SD, Becker P, Affrime MB, Cutler DL, Haehner-Daniels B. In vivo effects of interleukin-10 on human cytochrome P450 activity. Clin Pharmacol Ther 2000;67:32—43.
- [34] Pirttiaho HI, Sotaniemi EA, Pelkonen RO, Pitkanen U. Hepatic blood flow and drug metabolism in patients on enzyme-inducing anticonvulsants. Eur J Clin Pharmacol 1982:22:441–5.
- [35] Davis D, Grossman SH, Kitchell BB, Shand DG, Routledge PA. The effects of age and smoking on the plasma protein binding of lignocaine and diazepam. Br J Clin Pharmacol 1985;19:261–5.
- [36] Myrand SP, Sekiguchi K, Man MZ, Lin X, Tzeng RY, Teng CH, et al. Pharmacokinetics/genotype associations for major cytochrome P450 enzymes in native and first- and third-generation Japanese populations: comparison with Korean, Chinese, and Caucasian populations. Clin Pharmacol Ther 2008;84:347–61.
- [37] Lucas D, Menez JF, Berthou F. Chlorzoxazone: an in vitro and in vivo substrate probe for liver CYP2E1. Methods Enzymol 1996;272:115—23.
- [38] Peter R, Bocker R, Beaune PH, Iwasaki M, Guengerich FP, Yang CS. Hydroxylation of chlorzoxazone as a specific probe for human liver cytochrome P-450IIE1. Chem Res Toxicol 1990;3:566—73.
- [39] de Vries JD, Salphati L, Horie S, Becker CE, Hoener BA. Variability in the disposition of chlorzoxazone. Biopharm Drug Dispos 1994;15:587–97.
- [40] Lucas D, Ferrara R, Gonzalez E, Bodenez P, Albores A, Manno M, et al. Chlorzoxazone, a selective probe for phenotyping CYP2E1 in humans. Pharmacogenetics 1999;9:377–88.
- [41] Conney AH, Burns JJ. Physiological disposition and metabolic fate of chlor-zoxazone (paraflex) in man. J Pharmacol Exp Ther 1960;128:340–3.
- [42] Park JY, Kim KA, Park PW, Ha JM. Effect of high-dose aspirin on CYP2E1 activity in healthy subjects measured using chlorzoxazone as a probe. J Clin Pharmacol 2006:46:109—14.
- [43] Prompila N, Wittayalertpanya S, Komolmit P. A study on the pharmacokinetics of chlorzoxazone in healthy Thai volunteers. J Med Assoc Thai 2007:90:160-6.
- [44] Girre C, Lucas D, Hispard E, Menez C, Dally S, Menez JF. Assessment of cytochrome P4502E1 induction in alcoholic patients by chlorzoxazone pharmacokinetics. Biochem Pharmacol 1994:47:1503—8.
- [45] Desiraju RK, Renzi Jr NL, Nayak RK, Ng KT. Pharmacokinetics of chlorzoxazone in humans. J Pharm Sci 1983;72:991–4.
- [46] Huang X, Chen L, Song W, Niu J, Han X, Feng G, et al. Systematic functional characterization of cytochrome P450 2E1 promoter variants in the Chinese Han population. PLoS One 2012;7:e40883.
- [47] Qian J, Song Z, Lv Y, Huang X. CYP2E1 T7632A and 9-bp insertion polymorphisms and colorectal cancer risk: a meta-analysis based on 4,592 cases and 5,918 controls. Tumour Biol 2013;34:2225–31.
- [48] Howgate EM, Rowland Yeo K, Proctor NJ, Tucker GT, Rostami-Hodjegan A. Prediction of in vivo drug clearance from in vitro data. I: impact of interindividual variability. Xenobiotica 2006;36:473–97.
- [49] Yamazaki H, Inoue K, Turvy CG, Guengerich FP, Shimada T. Effects of freezing, thawing, and storage of human liver samples on the microsomal contents and activities of cytochrome P450 enzymes. Drug Metab Dispos 1997;25: 168–74.
- [50] Pearce RE, McIntyre CJ, Madan A, Sanzgiri U, Draper AJ, Bullock PL, et al. Effects of freezing, thawing, and storing human liver microsomes on cytochrome P450 activity. Arch Biochem Biophys 1996;331:145–69.
- [51] Zhou SF, Wang B, Yang LP, Liu JP. Structure, function, regulation and polymorphism and the clinical significance of human cytochrome P450 1A2. Drug Metab Rev 2010;42:268–354.
- [52] Ikeya K, Jaiswal AK, Owens RA, Jones JE, Nebert DW, Kimura S. Human CYP1A2: sequence, gene structure, comparison with the mouse and rat orthologous gene, and differences in liver 1A2 mRNA expression. Mol Endocrinol 1989;3:1399–408.
- [53] Schweikl H, Taylor JA, Kitareewan S, Linko P, Nagorney D, Goldstein JA. Expression of CYP1A1 and CYP1A2 genes in human liver. Pharmacogenetics 1993;3:239–49.
- [54] Aklillu E, Carrillo JA, Makonnen E, Hellman K, Pitarque M, Bertilsson L, et al. Genetic polymorphism of CYP1A2 in Ethiopians affecting induction and expression: characterization of novel haplotypes with single-nucleotide polymorphisms in intron 1. Mol Pharmacol 2003;64:659–69.
- [55] Soyama A, Saito Y, Hanioka N, Maekawa K, Komamura K, Kamakura S, et al. Single nucleotide polymorphisms and haplotypes of CYP1A2 in a Japanese population. Drug Metab Pharmacokinet 2005;20:24–33.
- [56] Kall MA, Vang O, Clausen J. Effects of dietary broccoli on human in vivo drug metabolizing enzymes: evaluation of caffeine, oestrone and chlorzoxazone metabolism. Carcinogenesis 1996;17:793–9.
- [57] Sinha R, Rothman N, Brown ED, Mark SD, Hoover RN, Caporaso NE, et al. Panfried meat containing high levels of heterocyclic aromatic amines but low levels of polycyclic aromatic hydrocarbons induces cytochrome P4501A2 activity in humans. Cancer Res 1994;54:6154—9.
- [58] Kashuba AD, Bertino Jr JS, Kearns GL, Leeder JS, James AW, Gotschall R, et al. Quantitation of three-month intraindividual variability and influence of sex and menstrual cycle phase on CYP1A2, N-acetyltransferase-2, and xanthine oxidase activity determined with caffeine phenotyping. Clin Pharmacol Ther 1998;63:540-51.

- [59] Chiba K, Shimizu K, Kato M, Nishibayashi T, Terada K, Izumo N, et al. Prediction of inter-individual variability in the pharmacokinetics of CYP2C19 substrates in humans. Drug Metab Pharmacokinet 2014;29:379–86.
- [60] Carbo M, Segura J, De la Torre R, Badenas JM, Cami J. Effect of quinolones on caffeine disposition. Clin Pharmacol Ther 1989;45:234–40.
- [61] Obach RS, Lombardo F, Waters NJ. Trend analysis of a database of intravenous pharmacokinetic parameters in humans for 670 drug compounds. Drug Metab Dispos 2008;36:1385–405.
- [62] Shellenberger MK, Groves L, Shah J, Novack GD. A controlled pharmacokinetic evaluation of tizanidine and baclofen at steady state. Drug Metab Dispos 1999:27:201–4.
- [63] Shibata Y, Takahashi H, Chiba M, Ishii Y. Prediction of hepatic clearance and availability by cryopreserved human hepatocytes: an application of serum incubation method. Drug Metab Dispos 2002;30:892–6.
- [64] Mineshita S, Toyoshima A, Yazaki T. Influence of phenacetin and its metabolites on renal function. Nihon Jinzo Gakkai Shi 1989;31:629–33.
- [65] Brion N, Naline E, Beaumont D, Pays M, Advenier C. Lack of effect of terfenadine on theophylline pharmacokinetics and metabolism in normal subjects. Br J Clin Pharmacol 1989;27:391–5.
- [66] Lindberg R, Kanto J, Hovi-Viander M, Laurikainen E. Serum concentrations and urinary excretion of lidocaine and its two desethylated metabolites after spinal anaesthesia using lidocaine or lidocaine with phenylephrine. Acta Anaesthesiol Scand 1985;29:811—3.
- [67] Amchin J, Zarycranski W, Taylor KP, Albano D, Klockowski PM. Effect of venlafaxine on CYP1A2-dependent pharmacokinetics and metabolism of caffeine. J Clin Pharmacol 1999;39:252–9.
- [68] Seng KY, Fun CY, Law YL, Lim WM, Fan W, Lim CL. Population pharmacokinetics of caffeine in healthy male adults using mixed-effects models. J Clin Pharm Ther 2009;34:103—14.
- [69] Healy DP, Polk RE, Kanawati L, Rock DT, Mooney ML. Interaction between oral ciprofloxacin and caffeine in normal volunteers. Antimicrob Agents Chemother 1989:33:474–8.
- [70] Rodopoulos N, Norman A. Assessment of dimethylxanthine formation from caffeine in healthy adults: comparison between plasma and saliva concentrations and urinary excretion of metabolites. Scand J Clin Lab Investig 1996:56:259-68
- [71] Gelal A, Guven H, Balkan D, Artok L, Benowitz NL. Influence of menthol on caffeine disposition and pharmacodynamics in healthy female volunteers. Eur J Clin Pharmacol 2003;59:417–22.
- [72] Culm-Merdek KE, von Moltke LL, Harmatz JS, Greenblatt DJ. Fluvoxamine impairs single-dose caffeine clearance without altering caffeine pharmacodynamics. Br J Clin Pharmacol 2005;60:486–93.
- [73] Henney 3rd HR, Shah J. Relative bioavailability of tizanidine 4-mg capsule and tablet formulations after a standardized high-fat meal: a single-dose, randomized, open-label, crossover study in healthy subjects. Clin Ther 2007;29:661–9.
- [74] Backman JT, Karjalainen MJ, Neuvonen M, Laitila J, Neuvonen PJ. Rofecoxib is a potent inhibitor of cytochrome P450 1A2: studies with tizanidine and caffeine in healthy subjects. Br J Clin Pharmacol 2006;62:345–57.
- [75] Granfors MT, Backman JT, Neuvonen M, Ahonen J, Neuvonen PJ. Fluvoxamine drastically increases concentrations and effects of tizanidine: a potentially hazardous interaction. Clin Pharmacol Ther 2004;75:331–41.
- [76] Backman JT, Granfors MT, Neuvonen PJ. Rifampicin is only a weak inducer of CYP1A2-mediated presystemic and systemic metabolism: studies with tizanidine and caffeine. Eur J Clin Pharmacol 2006;62:451–61.
- [77] Granfors MT, Backman JT, Laitila J, Neuvonen PJ. Oral contraceptives containing ethinyl estradiol and gestodene markedly increase plasma concentrations and effects of tizanidine by inhibiting cytochrome P450 1A2. Clin Pharmacol Ther 2005;78:400—11.
- [78] Granfors MT, Backman JT, Neuvonen M, Neuvonen PJ. Ciprofloxacin greatly increases concentrations and hypotensive effect of tizanidine by inhibiting its cytochrome P450 1A2-mediated presystemic metabolism. Clin Pharmacol Ther 2004;76:598–606.
- [79] Henney 3rd HR, Fitzpatrick A, Stewart J, Runyan JD. Relative bioavailability of tizanidine hydrochloride capsule formulation compared with capsule contents administered in applesauce: a single-dose, open-label, randomized, two-way, crossover study in fasted healthy adult subjects. Clin Ther 2008;30: 2263-71
- [80] Karjalainen MJ, Neuvonen PJ, Backman JT. Celecoxib is a CYP1A2 inhibitor in vitro but not in vivo. Eur J Clin Pharmacol 2008;64:511–9.
- [81] Xiaodong S, Gatti G, Bartoli A, Cipolla G, Crema F, Perucca E. Omeprazole does not enhance the metabolism of phenacetin, a marker of CYP1A2 activity, in healthy volunteers. Ther Drug Monit 1994;16:248–50.
- [82] Bartoli A, Xiaodong S, Gatti G, Cipolla G, Marchiselli R, Perucca E. The influence of ethnic factors and gender on CYP1A2-mediated drug disposition: a comparative study in Caucasian and Chinese subjects using phenacetin as a marker substrate. Ther Drug Monit 1996;18:586–91.
- [83] Dong SX, Ping ZZ, Xiao WZ, Shu CC, Bartoli A, Gatti G, et al. Effect of active and passive cigarette smoking on CYP1A2-mediated phenacetin disposition in Chinese subjects. Ther Drug Monit 1998;20:371–5.
- [84] Gillum JG, Sesler JM, Bruzzese VL, Israel DS, Polk RE. Induction of theophylline clearance by rifampin and rifabutin in healthy male volunteers. Antimicrob Agents Chemother 1996;40:1866—9.
- [85] Moller SE, Larsen F, Pitsiu M, Rolan PE. Effect of citalopram on plasma levels of oral theophylline. Clin Ther 2000;22:1494–501.

- [86] Meyer MC, Jarvi EJ, Straughn AB, Pelsor FR, Williams RL, Shah VP. Bio-equivalence of immediate-release theophylline capsules. Biopharm Drug Dispos 1999;20:417–9.
- [87] Plezia PM, Thornley SM, Kramer TH, Armstrong EP. The influence of enteral feedings on sustained-release theophylline absorption. Pharmacotherapy 1990;10:356–61.
- [88] West RJ, Boehm G, Dwyer M, Williams DB, Sansom LN, Penna AC. Bioavailability of a new sustained-release theophylline capsule in fasted and nonfasted healthy subjects: single and multiple dosing studies. Biopharm Drug Dispos 1990;11:165–77.
- [89] Bhargava VO, Schaaf LJ, Berlinger WG, Jungnickel PW. Effect of an enteral nutrient formula on sustained-release theophylline absorption. Ther Drug Monit 1989;11:515—9.
- [90] Schulz HU, Karlsson S, Sahner-Ahrens I, Steinijans VW, Beier W. Effect of drug intake prior to or after meals on serum theophylline concentrations: single-dose studies with Euphylong. Int J Clin Pharmacol Ther Toxicol 1987;25:222–8.
- [91] Cohen IA, Johnson CE, Berardi RR, Hyneck ML, Achem SR. Cimetidine—theophylline interaction: effects of age and cimetidine dose. Ther Drug Monit 1985;7:426–34.
- [92] Yao C, Kunze KL, Kharasch ED, Wang Y, Trager WF, Ragueneau I, et al. Fluvoxamine—theophylline interaction: gap between in vitro and in vivo inhibition constants toward cytochrome P4501A2. Clin Pharmacol Ther 2001;70: 415–24.

- [93] Lohmann A, Dingler E, Sommer W. Influence of food on the bioavailability of theophylline from a sustained-released theophylline preparation. Arzneimittelforschung 1991;41:732–4.
- [94] Lalonde RL, Bottorff MB, Straughn AB. Comparison of high pressure liquid chromatography and fluorescence polarization immunoassay methods in a theophylline pharmacokinetic study. Ther Drug Monit 1985;7:442–6.
- [95] Tonsuwannont W, Praisontarangkul OA, Manorot M, Klangwarnwong D. Pharmacokinetics of oral lidocaine and nifedipine in patients with liver cirrhosis. J Med Assoc Thai 1996;79:309–19.
- [96] de Boer AG, Breimer DD, Mattie H, Pronk J, Gubbens-Stibbe JM. Rectal bioavailability of lidocaine in man: partial avoidance of "first-pass" metabolism. Clin Pharmacol Ther 1979;26:701–9.
- [97] Isohanni MH, Neuvonen PJ, Olkkola KT. Effect of fluvoxamine and erythromycin on the pharmacokinetics of oral lidocaine. Basic Clin Pharmacol Toxicol 2006;99:168–72.
- [98] Isohanni MH, Neuvonen PJ, Olkkola KT. Effect of erythromycin and itraconazole on the pharmacokinetics of oral lignocaine. Pharmacol Toxicol 1999;84:143–6.
- [99] Perucca E, Richens A. Reduction of oral bioavailability of lignocaine by induction of first pass metabolism in epileptic patients. Br J Clin Pharmacol 1979:8:21–31.
- [100] Bax ND, Tucker GT, Lennard MS, Woods HF. The impairment of lignocaine clearance by propranolol major contribution from enzyme inhibition. Br J Clin Pharmacol 1985:19:597—603.