The effect of propranolol on paracetamol metabolism in man

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Ten healthy volunteers were treated for 4 days with 160 mg propranolol HCl and placebo in random order. At the end of each treatment salivary antipyrine kinetics and the plasma kinetics and urinary excretion of paracetamol and its major metabolites were measured following a 1500 mg oral dose. Propranolol prolonged the half-life of antipyrine by $11 \pm 5\%$ (mean \pm s.e. mean) and lowered its clearance by $14 \pm 3\%$ (P < 0.05). Propranolol increased the half-life of paracetamol by $25 \pm 12\%$ (P < 0.05) and lowered its clearance by $14 \pm 3\%$ (P < 0.05). Propranolol decreased the partial clearance of paracetamol to its cysteine and mercapturate derivatives by $16 \pm 3\%$ (P < 0.05) and $32 \pm 7\%$ (P < 0.05), respectively. The partial clearance to the glucuronide conjugate was decreased by $27 \pm 6\%$ (P < 0.05), whereas that to sulphate was not changed significantly. Propranolol inhibits paracetamol metabolism predominantly through inhibition of the oxidation and glucuronidation pathways.

Keywords drug metabolism drug interaction propranolol paracetamol antipyrine

Introduction

Since paracetamol is oxidised to a hepatotoxic intermediate metabolite N-acetyl p-benzo-quinoneimine (Corcoran et al., 1980; Meredith & Goulding, 1980), interactions with drugs that induce hepatic mono-oxygenases (Mitchell et al., 1983; Perucca & Richens, 1979; Prescott et al., 1981) and those that inhibit them (Mitchell et al., 1984; Sanchez-Martinez et al., 1985) are of interest.

Propranolol inhibits the metabolism of chlorpromazine (Peet et al., 1980), lignocaine (Ochs et al., 1980) and theophylline (Conrad & Nyman, 1980; Miners et al., 1985) and, in common with some other β-adrenoceptor antagonists, antipyrine (Bax et al., 1983; Greenblatt et al., 1978; Parker et al., 1984). The inhibition of oxidative metabolism by propranolol is related to its lipid solubility (Bax et al., 1981; Deacon et al., 1983), it may be selective for different forms of cytochrome P450 and is possibly concentration dependent (Miners et al., 1984).

In animal studies propranolol in large doses has a protective effect against paracetamolinduced hepatic necrosis (Bax et al., 1983; Legors, 1976). However, in a study of six individuals a relatively small dose of propranolol (80 mg daily for 6 days) did not affect the kinetics of paracetamol (Sanchez-Martinez et al., 1985).

The aim of this study was to re-investigate the effect of propranolol on paracetamol disposition in man. In order to confirm the effect of propranolol on drug metabolising capacity a measurement of antipyrine kinetics was incorporated in the study design.

Method

Subjects

Six males and four females, all clinically healthy, aged between 21 and 32 years and weighing from 51 to 85 kg (median 68 kg) participated in the

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study which was approved by the Bristol and Weston Health Authority Ethics Committee. All were non-smokers, used alcohol in moderation and did not take other drugs, including the oral contraceptive.

Study design

The study was a double-blind, crossover investigation comprising two periods of 5 days each separated by 12 days. Subjects received in a random order either racemic propranolol HCl 80 mg or matched placebo tablets twice daily for 4 days. Subjects refrained from taking alcohol during the study and reported to the clinical laboratory at 09.00 h after an overnight fast on days 4 and 5 of each treatment period.

On day 4 a single oral dose of paracetamol (1500 mg) was administered and the subjects continued to fast for 2 h. Venous blood samples (5 ml) were collected at 0, 0.5, 1, 1.5, 2, 3, 4, 5, 6, 8, 12 and 24 h. A complete urine collection was made for the first 24 h following drug administration.

On day 5 the subjects received a single oral dose of 600 mg antipyrine. Saliva (2 ml) was obtained at 0, 3, 6, 9, 12 and 24 h. All plasma samples were separated immediately and together with aliquots of the urine and saliva samples they were frozen and stored at -20° C until assayed.

Antipyrine was assayed in saliva by h.p.l.c. (Danhof et al., 1979). Paracetamol and its metabolites in plasma and urine were assayed by the h.p.l.c. method of Adriaenssens & Prescott (1978) modified by the use of metacetamol as internal standard. The mobile phase was 0.1 M potassium dihydrogen phosphate, 98% v/v formic acid and propan-2-ol (100:0.16:1.5 v/v/v). This method was able to measure paracetamol and its metabolites at 0.1 µg ml⁻¹ at a signal to noise ratio of 2:1 or better. The inter-assay coefficients of variation for paracetamol and all the metabolites were no greater than 7% at 2 µg ml^{-1} , 4% at 10 µg ml^{-1} and 3% at 50 µg ml^{-1} . The paracetamol metabolites used as standards were donated by Sterling Winthrop Laboratories (Alnwick). The standard metabolite solutions were prepared and stored at -20 °C for 8 weeks during which time repeated assay against freshly prepared solutions showed no evidence of instability. All urine samples were assayed within 6 weeks of collection.

Pharmacokinetic analysis

The plasma elimination half-life (t_{1/2}) of paracetamol was computed by least squares linear

regression of the terminal part of the log drug concentration-time curve. The area under the plasma drug concentration-time profile (AUC) was calculated as follows: the area up to the end of sampling was calculated using the linear trapezoidal rule and the remaining area was extrapolated to infinity using the terminal log drug concentration-time slope. The systemic clearance of paracetamol could not be assessed in the absence of a measurement of bioavailability. However, the oral clearance (CL_{po}) was estimated from:

$$CL_{po} = Dose/AUC$$

The partial clearance of paracetamol to each of its metabolites (CL_m) was calculated from

$$CL_m = Ae(m)$$
 AUC

where Ae(m) = Amount of metabolite recovered in urine.

Statistical analysis

Student's *t*-test for paired data was used to compare measurements on propranolol with those on placebo.

Results

All subjects completed the study without adverse effects. One or more urine collections had been collected incorrectly by three subjects and their urinary results were excluded from the analysis.

Antipyrine kinetics

Propranolol increased the elimination half-life of antipyrine from 11.2 \pm 2.5 h (mean \pm s.d.) to 12.4 \pm 3.5 h (P < 0.05) and decreased its clearance from 58.1 \pm 19.3 ml min⁻¹ to 51.5 \pm 18.3 ml min⁻¹ (P < 0.05) compared with placebo.

Paracetamol and metabolites in plasma and urine

The plasma concentration of paracetamol reached a peak of $27.0 \pm 10.8 \, \mu g \, \mathrm{ml}^{-1}$ after propranolol compared with $18.0 \pm 6.1 \, \mu g \, \mathrm{ml}^{-1}$ following placebo (P < 0.01) (Figure 1). Propranolol increased the half-life of paracetamol from $2.7 \pm 0.6 \, \mathrm{to} \, 3.4 \pm 0.3 \, \mathrm{h} \, (P < 0.05)$ and lowered its oral clearance from $364 \pm 92 \, \mathrm{ml} \, \mathrm{min}^{-1}$ to $313 \pm 28 \, \mathrm{ml} \, \mathrm{min}^{-1} \, (P < 0.05)$.

Propranolol decreased the partial clearance of paracetamol to the oxidised metabolites—the cysteine and mercapturate—by 16 ± 3% and

Table 1 Partial metabolic clearances of paracetamol metabolites and percentage recovery of administered dose in seven healthy volunteers (ml min⁻¹; mean \pm s.d.)

	Glucuronide	Sulphate	Cysteine	Mercapturate	% Recovery
Placebo	241.0 ± 91.0	91.2 ± 19.6	9.2 ± 2.4	12.7 ± 3.7	77.9 ± 16.3
Propranolol	$177.0 \pm 66.9*$	75.2 ± 19.5	$7.7 \pm 2.1*$	$8.0 \pm 3.7*$	88.3 ± 10.4

^{*}P < 0.05 compared with placebo; Student's *t*-test

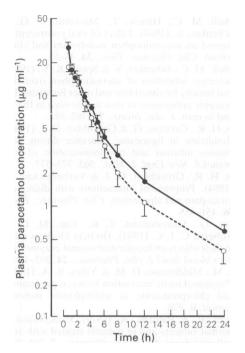


Figure 1 Plasma paracetamol concentrations following oral administration of 1500 mg paracetamol to 10 healthy subjects after treatment with placebo (\circ) or propranolol (\bullet) for 4 days (mean \pm s.e. mean).

32 \pm 7% (P < 0.05), respectively and to the glucuronide by 27 \pm 7% (P < 0.05). However, the partial clearance of paracetamol to sulphate was not changed significantly by propranolol treatment (Table 1).

Discussion

The results of this study suggest that the increase in the plasma half-life and the lowered clearance of paracetamol caused by propranolol are mainly due to inhibition of glucuronidation and oxidative pathways.

The dose of propranolol used in this study was shown to be sufficient to inhibit hepatic drug metabolising enzymes by the significant lowering of antipyrine clearance, although this was less than that reported by others (Bax et al., 1981; Greenblatt et al., 1978).

The decreased oral clearance of paracetamol was in contrast to the findings of Sanchez-Martinez et al. (1985) who reported a lack of effect of propranolol at a dose of 80 mg once a day for 6 days on the kinetics of 1 g oral paracetamol. However, in that study no independent measurement of enzyme inhibition was made. In the present study we have used twice the dose of propranolol and have provided a positive control for enzyme inhibition by the changes in antipyrine clearance. The dose used may be critical as the extent of enzyme inhibition by propranolol is dose dependent (Miners et al., 1985).

The decreased partial clearance to the cysteine and mercapturate metabolites suggests that propranolol significantly inhibits the oxidative pathway of paracetamol. Although the mean decrease in partial clearance to the mercapturate was greater than that to the cysteine derivative this was not significant. This mechanism may explain the observation that propranolol had a protective effect on paracetamol-induced hepatic necrosis in mice (Bax et al., 1983; Legors, 1976). Propranolol significantly impaired the glucuronidation of paracetamol but sulphation was not affected. This contrasts with the finding of Brunk et al. (1974) that propranolol did not inhibit the metabolism of morphine which mainly undergoes conjugation and with those of Ochs et al. (1984) and Hawksworth et al. (1984) who found a decrease in the clearance of diazepam (which undergoes oxidation to the N-desmethyl product) but no effect of propranolol on the kinetics of two benzodiazepines which are glucuronidated, namely lorazepam alprazolam. These differences may reflect the fact that glucuronidation is carried out by several isoenzymes with varying substrate specificity (Dutton, 1966).

There are two possible implications of our findings. Thus, propranolol may afford some protection against paracetamol-induced hepatic damage and it may prolong the effect of paracetamol.

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References

- Adriaenssens, P. I. & Prescott, L. F. (1978). High performance liquid chromatographic estimation of paracetamol metabolites in plasma. *Br. J. clin. Pharmac.*, 6, 87–88.
- Bax, N. D. S., Lennard, M. S. & Tucker, G. T. (1981). Inhibition of antipyrine metabolism by β-adrenoceptor antagonists. Br. J. clin. Pharmac., 12, 779–784.
- Bax, N. D. S., Lennard, M. S., Al-Asady, S., Deacon, C. S., Tucker, G. T. & Woods, H. (1983). Inhibition of drug metabolism by beta adrenoceptor antagonists. *Drugs*, 25 (Suppl. 2), 121–126.
- Brunk, S. F., Delle, M. & Wilson, W. R. (1974). Effect of propranolol on morphine metabolism. *Clin. Pharmac. Ther.*, **28**, 463–467.
- Conrad, K. A. & Nyman, D. W. (1980). Effect of metoprolol and propranolol on theophylline elimination. Clin. Pharmac. Ther., 33, 133-138.
- Corcoran, G. B., Mitchell, J. R., Vasihnav, Y. N. & Horning, E. C. (1980). Evidence that acetaminophen and n-hydroxyacetaminophen form a common arylating intermediate, n-acetyl p-benzoquinoneimine. Mol. Pharmac., 18, 536-542.
- Danhof, M., de Groot van der Vis, E. & Breimer, D. D. (1979). Assay of antipyrine and its primary metabolites in plasma, saliva and urine by high performance liquid chromatography and some preliminary results in man. *Pharmacology*, 18, 210– 223.
- Deacon, C. S., Lennard, M. S., Bax, N. D. S., Woods, H. S. & Tucker, G. T. (1981). Inhibition of oxidative drug metabolism by β-adrenoceptor antagonists is related to their lipid solubility. *Br. J. clin. Pharmac.*, 12, 429–430.
- Dutton, G. J. (1966). The biosynthesis of glucuronide, In Glucuronic acid, ed. Dutton, G. J. New York: Academic Press, inc.
- Greenblatt, D. G., Frankie, K. & Huffman, D. H. (1978). Impairment of antipyrine clearance in human by propranolol. Circulation, 57, 1161-1164.
- Hawksworth, G., Betts, T., Crowe, A., Nyemitei-Addo, I., Parry, K., Petrie, J. C., Raffle, A. & Parsons, A. (1984). Diazepam/β-adrenoceptor antagonist interactions. Br. J. clin. Pharmac., 17, 698-76S.
- Legors, J. (1976). Animal studies. A theoretical basis for treatment. J. int. med. Res., 4 (Suppl.4), 46-54.
- Meredith, T. J. & Goulding, R. (1980). Paracetamol. *Postgrad. med. J.*, **56**, 459-473.
- Miners, J. O., Wing, L. M. H., Lillywhite, K. J. & Smith, K. J. (1984). Failure of therapeutic doses

- of β-adrenoceptor antagonists to alter the disposition of tolbutamide and lignocaine. *Br. J. clin. Pharmac.*, **18**, 853–860.
- Miners, J. O., Wing, L. M. H., Lillywhite, K. J. & Robson, R. A. (1985). Selectivity and dose dependency of the inhibitory effect of propranolol on theophylline metabolism in man. Br. J. clin. Pharmac., 20, 219-223.
- Mitchell, M. C., Hanew, T., Meredith, C. G. & Schenker, S. (1983). Effect of oral contraceptive steroid on acetaminophen metabolism and elimination. Clin. Pharmac. Ther., 34, 48-53.
- Mitchell, M. C., Schenker, S. & Speeg, K. V. (1984). Selective inhibition of acetaminophen oxidation and toxicity by cimetidine and other histamine H₂receptor antagonists in vivo and in vitro in the rat and in man. J. clin. Invest., 73, 383-391.
- Ochs, H. R., Carstens, G. & Greenblatt, D. J. (1980). Reduction in lignocaine clearance during continuous infusion and co-administration of propranolol. New Engl. J. Med., 303, 373-377.
- Ochs, H. R., Greenblatt, D. J. & Verburg-Ochs, B. (1984). Propranolol interactions with diazepam, lorazepam and alprazolam. Clin. Pharmac. Ther., 36, 451-455.
- Parker, G., Daneshmend, T. K., Enc. M. D. & Roberts, C. J. C. (1984). Do beta blockers differ in their effects on hepatic microsomal enzymes and liver blood flow? J. clin. Pharmac., 24, 395–400.
- Peet, M., Middlemass, D. M. & Yates, R. A. (1980).
 Pharmacokinetic interaction between propranolol and chlorpromazine in schizophrenic patients.
 Lancet, ii, 978.
- Perucca, E. & Richens, A. (1979). Paracetamol in normal subjects and in patients treated with antiepileptic drugs. Br. J. clin. Pharmac., 7, 201-206.
- Prescott, L. F., Critchley, J. A. J. H., Balali-Mood, M. & Pentland, B. (1981). Effect of microsomal enzyme induction on paracetamol metabolism in man. Br. J. clin. Pharmac., 12, 149-153.
- Sanchez-Martinez, V., Tucker, G. T., Jackson, P. R., Lennard, M. S., Bax, N. D. S. & Woods, H. F. (1985). Lack of effect of propranolol on the kinetics of paracetamol in man. *Br. J. clin. Pharmac.*, 20, 548P.

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