Effect of smoking on caffeine clearance*

The elimination of caffeine from saliva was compared in groups of healthy smokers (n = 13) and nonsmokers (n = 13). Mean caffeine $t\frac{1}{2}$ in smokers (3.5 hr) was shorter than that in the nonsmokers (6.0 hr). The body clearance of caffeine in the smokers $(155 \pm 16 \text{ ml} \cdot \text{kg}^{-1} \cdot \text{hr}^{-1})$ was greater than that in the nonsmokers $(94 \pm 18 \text{ ml} \cdot \text{kg}^{-1} \cdot \text{hr}^{-1})$ (p < 0.05). No significant difference was noted in the apparent volume of distribution in smokers $(720 \pm 67 \text{ ml} \cdot \text{kg}^{-1})$ and nonsmokers $(610 \pm 80 \text{ ml} \cdot \text{kg}^{-1})$. These differences probably reflect the induction of hepatic aryl hydrocarbon hydroxylase (AHH) activity in smokers. The increased clearance of caffeine by smokers may contribute to the higher consumption of coffee reported to occur in this group.

William D. Parsons, M.D.,** and Allen H. Neims, M.D., Ph.D.***

Montreal, Quebec, Canada

Department of Pharmacology and Therapeutics, Roche Developmental Pharmacology Unit, McGill University

Certain observations have led us to postulate a surprisingly close relationship in rats between aryl hydrocarbon hydroxylase activity (AHH) and the metabolism of caffeine¹: (1) pretreatment with 3-methylcholanthrene markedly enhances the animal's capacity to eliminate caffeine, whereas phenobarbital has only a small effect¹; (2) pretreatment with 3-methylcholanthrene increases the catalytic activity of hepatic microsomal preparations toward caffeine²²; and (3) the differential effects of 7,8-benzoflavone and SKF-525A β -diethylaminoethyl 2,2-diphenylvalerate on basal and induced microsomal monooxygenase activities toward 3, 4-benzpyrene³¹ and caffeine²² are analogous.

Cigarette smoke, a complex mixture¹⁶ con-

taining a variety of polycyclic aromatic hydrocarbons, is known to induce AHH in certain human tissues.^{7, 23, 28} To further explore the relationship between AHH and the disposition of caffeine, we studied the effect of cigarette smoking on caffeine clearance in healthy young adults. The results bear directly on the question of whether caffeine might serve as a probe to study the role of AHH and its inducibility in relation to individual susceptibility to chemical carcinogenesis.¹⁰ Furthermore, since cigarette smoking and the consumption of caffeine are among the most common habits of our species, their interaction has significance quite apart from AHH.

Methods

Subjects. Our subjects were 13 smokers and 13 control nonsmokers chosen from laboratory and hospital personnel. Each group consisted of 9 females and 4 males with an average age of 26 (range 20 to 45). No subjects in the nonsmoking group had ever used tobacco regularly. Smoking subjects consumed at least 20 cigarettes a day for a minimum of two years. With the ex-

Supported by the MRC of Canada, Grant MA-5162.

Received for publication Feb. 1, 1978.

Accepted for publication April 11, 1978.

Reprint requests to: Dr. W. D. Parsons, Department of Pharmacology and Therapeutics, McIntyre Medical Sciences Bldg., 3655 Drummond St., Montreal, Quebec H3G 1Y6, Canada.

^{*}A portion of this work was presented at the 1978 meeting of the Canadian Society for Clinical Investigation, Vancouver, B. C.

^{**}Supported by a fellowship from the MRC of Canada.

^{***}Support received from a Fraser Award.

106

117

 165 ± 23

80

152

 158 ± 22

4

25

Mean ± SE

plasma (P)									
Subject	kel (hr^{-1})	aVd (ml	kg^{-1}	Cl (ml·k	$g^{-1} \cdot hr^{-1}$			
	S	P	S	P	S	P			
2	0.461	0.417	482	505	220	210			
3	0.295	0.207	660	886	196	183			
12	0.250	0.266	743	615	185	164			

Table I. Comparison of pharmacokinetic data for caffeine as determined from saliva (S) or plasma (P)

0.101

0.129

 0.224 ± 0.056

Caffeine, 5 mg/kg, was administered orally as caffeine citrate to the five subjects. Saliva and plasma were sampled simultaneously over a 24-hr period. The data were fitted to the first-order, one-compartment equation with use of log-linear least-square regression analysis.

900

1.020

 761 ± 93

796

1.179

 796 ± 117

ception of three women in each group who were taking oral contraceptives, none of the subjects took any medication regularly. The usual daily intake of caffeine, mostly as coffee, was found by questionnaire to approximate 320 mg in the smokers and 150 mg in the nonsmokers.

0.117

0.117 0.249 ± 0.057

Procedure. Subjects were instructed to abstain from caffeine containing foods and beverages for 24 hours prior to and during the study. On the day of study, they were permitted their usual breakfast excluding caffeine. In the five subjects in whom both plasma and salivary caffeine concentrations were measured, caffeine (5 mg/kg) was administered orally as a solution of caffeine citrate (1 mg/ml). Subjects were instructed to rinse their mouths thoroughly immediately after ingestion of the caffeine. Salivary samples (5 ml mixed saliva) were collected in polyethylene vials at 0.5, 1, 1.5, 2, 4, 6, 8, 10, and 24 hr after ingestion and stored at 4° C until assay within two weeks. Blood samples (2) ml) were taken at the same time as saliva samples through an indwelling intravenous catheter which was flushed periodically with heparinized saline. The initial 0.5 ml of each blood collection was discarded. Plasma was separated and stored at 4° C for assay within two weeks. Only saliva was sampled in the other 21 subjects who were allowed to select 180 ml of coffee (n, 18), tea (n, 2) or cola (n, 1) as a source of the test dose of caffeine. An aliquot of the beverage was assayed for caffeine.

Assay procedure; kinetic and statistical analyses. Caffeine was measured by the radioimmunoassay procedure developed and supplied by Cook and colleagues⁸ of Research Triangle Institute.

The caffeine concentration: time curves were analyzed by assumption of a first-order onecompartment model. The validity of this model was established when it was determined that mean caffeine clearance from plasma in the five subjects described subsequently in Table I was equivalent whether calculated by this model $(158 \pm 22 \text{ ml} \cdot \text{kg}^{-1} \cdot \text{hr}^{-1})$ or the method of area under the curve (159 \pm 23 ml·kg⁻¹·hr⁻¹). The elimination rate constant, kel, was computed by least-square log-linear regression analysis of the descending portion of caffeine concentration as a function of time. The apparent volume of distribution (aVd) was calculated from the dose of caffeine administered divided by the extrapolated caffeine concentration at time 0 after subtraction of the actual 0-time value (a 24-hr abstinence from caffeine was usually sufficient to make this value negligible). The body clearance (Cl) was computed as the product of kel and aVd. Since caffeine is absorbed rapidly and completely, one-compartmental kinetic computations after oral and intravenous dosing are similar.² So also, the small differences in rate of absorption of caffeine from different beverages¹⁹ should not affect our results. The unpaired Student's t test was used in statistical comparisons.

Comparison of the salivary and plasma modes of sampling. Simultaneous salivary and plasma caffeine decay curves were studied in five subjects. Except for a transiently high salivary: plasma concentration ratio in a few subjects during the first 1 to 1.5 hr, the salivary and plasma caffeine concentrations both decrease proportionally in a fashion consistent with first-order elimination. Typical results are

42 Parsons and Neims

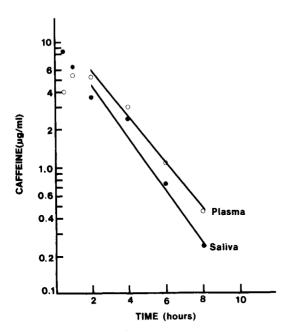


Fig. 1. Time-course of plasma and saliva caffeine concentrations after ingestion of caffeine (5 mg/kg) as caffeine citrate (5 mg/ml). The solid lines represent the least-squares regression line for the data points beginning from 2 hr postingestion.

presented in Fig. 1. The initially high salivary concentrations probably reflect residual caffeine in the mouth, and were ignored in computations. The salivary and plasma caffeine concentrations in each subject were subjected to linear least-squares regression analysis. The lowest correlation coefficient was 0.90, the mean correlation coefficient 0.96, and the y intercept 0.02. The salivary:plasma caffeine concentration ratio for the five subjects was 0.90 ± 0.17 . Similar results have been reported by Cook and co-workers.8 Values for kel, aVd, and C1 were computed separately from the salivary and plasma data of each subject (Table I). Agreement is generally good, and there is no significant difference between the mean value of any of the three parameters whether calculated from salivary or plasma data.

Cigarette smoking and caffeine elimination. Individual values for kel, a Vd, and C1 are presented in Table II for nonsmokers and in Table III for smokers. The mean caffeine t½ in smokers was 3.5 hr, while that in the nonsmokers was 6.0 hr. There was an increase in

caffeine C1 from 94 \pm 8 ml/kg/hr (nonsmokers) to 155 \pm 16 ml/kg/hr (smokers) (p < 0.05). This difference reflected mainly an increase in kel (0.130 \pm 0.013 to 0.230 \pm 0.029 hr⁻¹; p < 0.05) in the smokers. The increase in aVd in smokers (610 \pm 80 to 720 \pm 67 ml/kg⁻¹) was not significant. The distributions of caffeine C1 values in the smokers and nonsmokers were similar (Fig. 2).

As noted above, caffeine consumption, especially as coffee, is higher in smokers than in nonsmokers. Despite this association, the correlation coefficient between coffee consumption and caffeine clearance is only 0.23, indicating that coffee consumption is not the major determinant of clearance in these subjects.

Discussion

A series of in vivo and in vitro experiments in rats in our laboratory, 1, 22 as well as a recent report by Welch, Hsu, and DeAngelis,29 have indicated a close relationship between AHH and the capacity to metabolize and eliminate caffeine. The object of our investigation was to explore the situation in man by studying the effect of cigarette smoking on caffeine clearance. Caffeine clearance was found to be 55% higher in smokers than in nonsmokers (p < 0.05), but the extent to which this result can be taken as evidence for a relationship between AHH and caffeine metabolism in human beings depends on two assumptions: that metabolism is rate-limiting for caffeine clearance and that the predominant effect of cigarette smoking on xenobiotic metabolism is induction of AHH.

With reference to the first assumption, it is likely that the increased clearance of caffeine in smokers reflects increased metabolism because the disposition of caffeine is essentially one-compartmental with only a small fraction of an ingested dose excreted unchanged by adults. The enhancement of drug metabolism in association with cigarette smoking has been invoked before to explain the increased clearance of theophylline^{13, 14} and antipyrene, ^{12, 27} the increased biotransformation of pentazocine¹⁵ and phenacetin, ^{17, 21} and the decreased actions of certain other drugs³⁻⁵ in smokers. The decreased bioavailability of phenacetin in cigarette smokers has been ascribed more speci-

Table II. Pharmacokinetic data for caffeine in nonsmoking, healthy subjects

Subject	t½ (hr)	$kel \ (hr^{-1})$	$aVd (ml \cdot kg^{-1})$	$(ml \cdot kg^{-1} \cdot hr^{-1})$
14	4.2	0.167	370	61
15	3.0	0.234	116	270
16	7.6	0.091	580	52
17	3.8	0.183	386	70
18	6.4	0.109	330	36
19	7.7	0.090	786	70
20	9.4	0.074	288	21
21	6.7	0.103	610	63
22	4.2	0.162	682	110
23	7.7	0.090	161	145
24	7.1	0.097	784	76
25	5.9	0.117	1,020	117
26	3.7	0.185	730	135
Mean ± SE	6.0	0.130 ± 0.013	610 ± 80	94 ± 18

Caffeine (0.7 to 2.2 mg/kg) was administered as 180 ml of coffee, tea, or cola drink except to those subjects described in Table I. Saliva was sampled 8 times over a 24-hr period and data were analyzed using log-linear least-square regression analysis.

Table III. Pharmacokinetic data for caffeine in smoking, healthy subjects

$Cl (ml \cdot kg^{-1} \cdot hr^{-1})$
159
220
196
106
69
113
218
138
268
82
164
185
105
155 ± 16

Caffeine (0.7 to 1.9 mg/kg) was administered as 180 ml of coffee, tea, or cola drink except to those subjects described in Table I. Saliva was sampled 8 times over a 24-hr period and data were analyzed using a log-linear least-square regression analysis.

fically to induction of drug-metabolizing enzymes in the intestine.¹⁷

With reference to the second assumption, two recent observations are pertinent: firstly, caffeine elimination is remarkably slow in patients with liver failure, 24 indicating that this organ is probably of major importance in caffeine's biotransformation and, secondly, Pelkonen and colleagues 23 have found that AHH activity in liver biopsy specimens from smokers is increased relative to that of nonsmokers. These observations are consistent with the hypothesis that smokers clear caffeine more rapidly than

nonsmokers predominantly because of induction of hepatic AHH. Since AHH is also present in human lung, ³³ skin, ¹⁸ and lymphocytes⁶, ³² and has been shown to be induced by cigarette smoking in pulmonary alveolar macrophage⁷ and placenta, ²⁸ it is interesting to speculate that organs other than liver might contribute to the metabolism of caffeine. Indeed, the contribution of extrahepatic metabolism may be accentuated in smokers since extrahepatic AHH in mouse²⁵ and rat²⁶, ³⁰ is more inducible than that of liver. Finally, we cannot exclude the possibility that a complex mixture such as cigarette

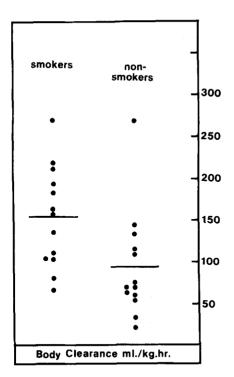


Fig. 2. Body clearance of caffeine in smokers and nonsmokers. Mean clearance values are indicated by the horizontal lines.

smoke⁴ induces monoxygenase activities in addition to AHH and that such forms contribute to the increased clearance of caffeine in smokers.

One confounding factor in studies of cigarette smoking in man is that smokers consume more coffee than nonsmokers. We found a positive correlation between coffee consumption and caffeine clearance, but it was not statistically significant. It is intriguing to speculate that the smoker consumes more coffee than the nonsmoker in part to compensate for increased caffeine clearance. However, since caffeine itself can induce mixed-function oxidase activity toward certain substrates in animals, 20 and the same could apply to other components of coffee, the causal relationship between consumption and clearance remains to be clarified. Such considerations would seem to merit attention in the design and interpretation of epidemiologic and behavioral studies dealing with smoking or coffee consumption.

We wish to thank Ms. L. Bonet and Mrs. R. Jain for their expert technical assistance and Dr. C. E. Cook for supplying the radioimmunoassay for caffeine.

References

- Aldridge A, Parsons WD, Neims AH: Stimulation of caffeine metabolism in the rat by 3-methylcholanthrene, Life Sci 21:967-974, 1977.
- Axelrod J, Reichenthal J: The fate of caffeine in man and a method for its estimation in biological material, J Pharmacol Exp Ther 107:519-523, 1952.
- Boston Collaborative Drug Surveillance Program: Clinical depression of the central nervous system due to diazepam and chlordiazepoxide in relation to cigarette smoking and age, N Engl J Med 288:277-280, 1973.
- Boston Collaborative Drug Surveillance Program: Decreased clinical efficacy of propoxyphene in cigarette smokers, CLIN PHARMACOL THER 14:259-263, 1973.
- 5. Boston Collaborative Drug Surveillance Program: Drowsiness due to chlorpromazine in relation to cigarette smoking, Arch Gen Psychiatry 31:211-213, 1974.
- Busbee DL, Shaw CR, Cantrell ET: Aryl hydrocarbon hydroxylase induction in human leucocytes, Science 178:315-316, 1972.
- Cantrell ET, Warr GA, Busbee DL, Martin RR: Induction of aryl hydrocarbon hydroxylase in human pulmonary alveolar macrophages by cigarette smoking, J Clin Invest 52:1881-1884, 1973.
- 8. Cook, CE, Tallent CR, Amerson E, Myers MW, Kepler JA, Taylor GC, Christensen HD: Caffeine in saliva and plasma by a radioimmune assay procedure, J Pharmacol Exp Ther 199:679-686, 1976.
- Cornish HH, Christman AA: A study of the metabolism of theobromine, theophylline and caffeine in man, J Biol Chem 228:315-323, 1957.
- Gelboin HV: Cancer susceptibility and carcinogen metabolism, N Engl J Med 297:385-386, 1977
- 11. Gilbert RM: Caffeine as a drug of abuse, in Gibbins RJ, Israel Y, Kalant H et al, editors: Research advances in alcohol and drug problems, New York, 1976, John Wiley & Sons, Inc., vol. 3.
- Hart P, Farrell GC, Cooksley WGE, Powell LW: Enhanced drug metabolism in cigarette smokers, Br Med J 2:147-149, 1976.
- 13. Hunt SN, Jusko WJ, Yurchak AM: Effect of smoking on theophylline disposition, CLIN PHARMACOL THER 19:546-551, 1976.
- 14. Jenne J, Nagasawa H, McHugh R: Decreased theophylline half-life in cigarette smokers, Life Sci 17:195-198, 1975.
- Keeri-Szanto M, Pomeroy JR: Atmospheric pollution and pentazocine metabolism, Lancet 1: 947-949, 1971.
- Kilburn KH: Effects of tobacco smoke on biological systems, Scand J Respir Dis (Suppl.) 19:63-78, 1974.

- Kuntzman R, Pantuck EJ, Kaplan SA, Conney AH: Phenacetin metabolism: Effects of hydrocarbons and cigarette smoking, CLIN PHAR-MACOL THER 22:757-764, 1977.
- Levin W, Conney AH, Alvares AP, Merkatz I, Kappas A: Induction of benzo(a)pyrene hydroxylase in human skin, Science 176:419-420, 1972.
- 19. Marks V, Kelly JF: Absorption of caffeine from tea, coffee and coca cola, Lancet 1:827, 1973.
- Mitoma C, Lombrozo L, Le Valley SC, Dehn F: Nature of the effect of caffeine on the drugmetabolizing enzymes, Arch Biochem Biophys 134:434-441, 1969.
- Pantuck EJ, Hsiao KC, Maggio A, Nakamura K, Kuntzman R, Conney AH: Effect of cigarette smoking on phenacetin metabolism, CLIN PHARMACOL THER 15:9-17, 1974.
- Parsons WD, Aldridge A: Stimulation of caffeine metabolism in the rat by 3-methylcholanthrene, Fed Proc vol. 35 Abst. No. 2538, 1976.
- Pelkonen O, Kaltiala EH, Karki NT, Jalonen K, Pyorala K: Properties of benzpyrene hydroxylase from human liver and comparison with the rat, rabbit and guinea-pig enzymes, Xenobiotica 5:501-509, 1975.
- Statiand BE, Desmas T, Danes M: Caffeine accumulation associated with alcoholic liver disease, N Engl J Med 295:110-111, 1976.
- Van Cantfort J, Gielen J: Induction by cigarette smoke of aryl hydrocarbon hydroxylase activity in the rat kidney and lung, Int J Cancer 19:538-545, 1977.
- Vestal RE, Norris AH, Tobin JD, Cohen BH, Shock NW, Andres R: Antipyrine metabolism in

- man: Influence of age, alcohol, caffeine and smoking, CLIN PHARMACOL THER **18:**425-432, 1975.
- Welch RM, Harrison YE, Gommi BW, Poppers PJ, Finster M, Conney AH: Stimulatory effect of cigarette smoking on the hydroxylation of 3,4benzpyrene and N-demethylation of 3-methyl-4-monomethylaminoazobenzene by enzymes in human placenta, CLIN PHARMACOL THER 10:100-109, 1969.
- Welch RM, Hsu SY, DeAngelis RL: Effect of Aroclor 1254, phenobarbital, and polycyclic aromatic hydrocarbons on the plasma clearance of caffeine in the rat, CLIN PHARMACOL THER 22:791-798, 1977.
- Welch RM, Loh A, Conney AH: Cigarette smoke: Stimulatory effect on metabolism of 3,4-benzpyrene by enzymes in rat lung, Life Sci 10:215-221, 1971.
- 31. Wiebel FJ, Gelboin HV, Bunn-Hoi NP, Stout MG, Burnham WS: Flavones and polycyclic hydrocarbons as modulators of aryl hydrocarbon (benzo(a)pyrene) hydroxylase, *in* Ts'o, P.O.P., and DiPaolo JA, editors: Chemical carcinogenesis. Part A, New York, 1974, Marcel-Dekker, Inc., pp. 249-270, 1974.
- 32. Whitlock JP, Cooper HL, Gelboin HV: Aryl hydrocarbon (benzopyrene) hydroxylase is stimulated in human lymphocytes by mitogens and benz(a)anthracene, Science 177:618-619, 1972.
- 33. Yang SK, Gelboin HV, Trump BF, Autrup H, Harris CC: Metabolic activation of benzo(a)pyrene and binding to DNA in cultured human bronchus, Cancer Res 37:1210-1215, 1977.