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Sources and consequences of mismatch between leaf disc and whole-leaf leaf mass per area

ABSTRACT

PREMISE: Leaf mass per area (LMA), which is an important functional trait in leaf economic spectrum and plant growth analysis, is measured from leaf discs or whole leaves. These differences in the measurement methods may lead to large differences in the estimates of LMA values.

METHODS: We examined to what extent whole-leaf and disc-based LMA match using 334 woody species from a wide range of biomes (tropics, subtropics, savanna, and temperate), whether the relationship varied by leaf morphology (tissue density, leaf area, leaf thickness), puncher size (0.6- and 1.0-cm diameter), and whether the extent of intraspecifc variation (ITV) for each species matches.

RESULTS: Disc-based estimates of species mean LMA matched well whole-leaf estimates, and whole-leaf LMA tended to be 9.689.69% higher than leaf disc LMA. The whole-leaf to leaf disc LMA ratio was higher for species with higher leaf tissue density and larger leaf, and their variance was greater for species with lower leaf tissue density and thinner leaves. Small leaf punch also inflated the ratio. The extent of ITV only weakly matched between whole-leaf and disc-based estimates ($R^2 = 0.090.08$).

CONCLUSIONS: Our results suggest that simple conversion between whole-leaf and leaf disc LMA is difficult for species obtained with a small leaf punch, but it should be possible for species obtained with a large leaf punch. Accurately representing leaf traits will probably require careful selection between leaf disc and whole-leaf traits depending on the objectives.

⁴¹ Quantifying ITV using leaf disc should be also considered with caution.

KEY WORDS: intraspecific variation, leaf density, leaf economic spectrum, leaf heterogeneity, leaf punch, leaf size, leaf thickness, petiole, specific leaf area, within-leaf variation

INTRODUCTION

The primary function of leaves is to return photosynthetic revenue on the investment that has been made in constructing the leaf (i.e., leaf economic spectrum, Wright et al., 2004; Westoby et al., 2013). Leaf mass per area (LMA or 1/SLA [specific leaf area]), determined by lamina thickness and leaf tissue density (LD), describes how much biomass is invested into given photosynthetic leaf area, which is a key trait in the leaf economic spectrum (Wright et al., 2004; Poorter et al., 2009; Onoda et al., 2017). Generally, resource-acquisitive species (fast-growing species) tend to have low LMA values, showing high photosynthesis, high nutrients, and often have fast leaf turnover (Wright et al., 2004). In contrast, resource-conservative species (slow-growing species) often have higher LMA values with the opposite patterns (Garnier and Laurent, 1994; Wright et al., 2004; Reich, 2014; Díaz et al., 2016). The LMA is also frequently used in plant growth analysis (Evans, 1972; Poorter et al., 2014; Falster

et al., 2018) because relative growth rates (RGR) can be decomposed into the product of
net assimilation rate (NAR), leaf mass ratio (LMR) and LMA (i.e., RGR = NAR × LMR × LMA⁻¹).
Another appealing feature of LMA is that it is relatively easy to measure large numbers of
species (i.e., 'soft' trait, Díaz et al., 2004). Therefore, LMA has been of interests to ecologists
and widely used since the first report more than a century ago (Hanson, 1917). Actually,
LMA has one of the best species coverage in leaf traits of the TRY plant data database
(16,460 species) and has been also the most often requested leaf trait (2,977 out of 7,330
requests), followed by leaf by leaf nitrogen (N) contents per leaf dry mass (12,238 species,
1,938 requests, Kattge et al., 2020 [TRY version 5, status October 1 2019])

LMA values can be determined by either measuring whole-leaf (including or excluding petioles) or leaf disc samples (Pérez-Harguindeguy et al., 2013). According to the TRY database version 5, recorded numbers of observations and species are as follows: 1) a whole leaf in-67 cluding (88,490 observations and 7,068 species) or excluding petioles (64,838 observations 68 and 7,558 species; note that this could include leaf disc samples), and 2) a leaf disc excluding 69 all major veins or petioles (645 observations and 403 species) (Kattge et al., 2020 [TRY version 70 5]). There are not many records that explicitly indicate leaf disc LMA in the TRY database, but leaf disc LMA has been commonly used for ecological studies as well as biochemical studies (e.g., Kraft et al., 2008; Poorter, 2009; Onoda et al., 2011; Osada et al., 2014; Sastry and Barua, 73 2017; Serbin et al., 2019; Campany et al., 2021). Although using whole-leaf traits seems to be more straightforward in the logic of investment costs and returns on investment (Westoby et 75 al., 2013), Poorter (2009) found that leaf disc LMA showed stronger correlations with shade 76 tolerance than whole-leaf LMA, suggesting that lamina traits are more important than wholeleaf traits in certain ecological contexts. Many studies do not make clear what protocol was followed (146,315 observations and 13,101 species), but these differences in the measure-79 ment methods may lead to large discrepancies in the estimates of LMA values because the 80 major vein allocation (major vein, which includes first to second or third-order veins, vol-81 ume per area) has been reported to be one of the main determinants for the variation in LMA (John et al., 2017). Previous works that compared estimates of whole-leaf LMA and leaf disc LMA using tropical tree species showed good correlations between the two ($R^2 = 0.92$ for 84 whole-leaves in Kraft et al., 2008; R^2 = 0.92 for leaf laminas in Onoda et al., 2011). However, 85 the comparisons between the two estimates in other biomes have rarely been examined, 86 even though LMA responses to climate (Poorter, 2009). 87

Since larger leaves tend to invest more of their mass into dense midribs and petiole for support (Niinemets et al., 2006, 2007; Li, Shi, et al., 2022; Li, Zheng, et al., 2022), and thinner leaves have clearly visible and large-diameter veins with less uniform leaf structure (i.e., kitetype leaves; (Grubb, 1986)), discrepancies in the estimates of whole-leaf LMA and leaf disc LMA might be greater for larger and thinner leaves. Higher leaf density (and LMA) is associated with higher vein density (John et al., 2017; Sancho-Knapik et al., 2020), and thus leaf disc LMA avoiding major veins might also underestimate whole-leaf LMA for species with

higher leaf tissue density. If leaf disc LMA largely and consistently underestimates wholeleaf LMA (i.e., differences in means), some calibrations may be required to combine or compare those different estimates (Kraft et al., 2008; Onoda et al., 2011). If divergences between 97 the estimates based on the different methods are large and inconsistent (i.e., differences in 98 variances and low R² values), this is difficult to calibrate and should inflate errors in the subsequent analyses. To date, however, only a database from a single region, Panama Plant 100 Traits Database (Wright et al., 2010), is available in the TRY (Kattge et al., 2020) that have 101 both estimates of LMA from small leaf discs and whole leaves including petioles, limiting of 102 our understanding of under what conditions estimates of LMA from leaf discs are reliable 103 estimates of whole-leaf LMA. 104

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Intraspecific trait variation (ITV), which reflects both heritable genetic variation and phenotypic plasticity, influences plant responses to abiotic and biotic interactions (Westerband et al., 2021). For example, shade leaves tend to have lower LMA than sun leaves because of fewer layers of palisade mesophyll cells (Terashima et al., 2001; Onoda et al., 2008). The extent of ITV within and among communities for LMA can be similar to or greater than interspecific trait variation (e.g., Messier et al., 2010; Kichenin et al., 2013; Fajardo and Siefert, 2018). The coefficient of variation (CV) is often used for quantifying the extent of ITV for each species (Yang et al., 2020; Westerband et al., 2021). Sources of ITV for whole-leaf samples are variation among individuals within the same species and variation among leaves within the same individuals (Messier et al., 2010, 2017). An additional source of trait variation for leaf disc samples is variation among leaf discs within the same leaves. Given that leaf disc samples have more sources of ITV than whole-leaf samples, the effects of discrepancies between whole-leaf LMA and leaf disc LMA might be large when ITV is quantified based on CV. Despite the importance of ITV in community ecology (Siefert et al., 2015; Westerband et al., 2021), the effect of different measurement methods on the extent of ITV has been largely ignored.

In this study, we aimed to investigate the relationship between whole-leaf (including petiole) LMA and leaf disc LMA, whether the relationship varied with leaf tissue density, leaf area and leaf thickness, and whether the extent of ITV for each species match between whole-leaf and leaf disc based estimates. We collected a total leaves from 1459 individuals of 334 woody species from four biomes (tropics, subtropics, savanna, and warm-temperate) to cover the wide range of geography. We evaluate the following hypotheses: (1) whole-leaf LMA would be greater than leaf disc LMA because whole-leaf LMA includes petioles and midribs that have greater dry mass per unit area than laminas (Niinemets et al., 2007), (2) species with higher leaf tissue density, larger and/or thinner leaves would show larger variance and differences in LMA estimates because whole-leaf LMA should be driven more in those species by veins and petioles, whereas those effects are likely to be ignored in leaf disc samples, and (3) the extent of ITV does not match well between whole-leaf and leaf disc based estimates because leaf disc samples have more sources of variation than whole-leaf samples. In addi-

tion to leaf morphology, we also investigate the effects of leaf punch size on the relationship between whole-leaf and leaf disc LMA.

MATERIALS AND METHODS

137 Data sources

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We used newly compiled individual-level plant datasets from a wide range of biomes in China 138 and Japan (Appendix S1, Table S1 (see the Supplementary Data with this article)) to exam-139 ine the relationship between whole-leaf and leaf disc-based estimates of leaf traits. First, 140 the Yunnan dataset is from three forest plots in Yunnan province, Southwest China, which includes a tropical rainforest (TRF), a tropical hot-dry savanna ecosystem (HDS), and a sub-142 tropical evergreen wet forest (STF), with a total of species and 141 species and 852 individ-143 uals. Second, the Yakushima dataset is from a warm-temperate forest in Yakushima island, 144 Japan, with a total of 193 species and 607 individuals. In total, our dataset comprised 334 145 woody species and 1365 individual 1459 individuals that ranged from 8.1 to 24.7 °C in mean annual temperature (MAT) and 732.8 to 4477 mm in mean annual precipitation (MAP) [Ap-147 pendix S1, Table S1; Eguchi (2006); Song et al. (2017); Fei et al. (2018)], which covers the wide 148 climate ranges in the distribution of broad-leaved tree species. The methodologies for trait 149 sampling and measurement are slightly different between the Yunnan and the Yakushima 150 dataset, which we describe below. 151

Measurements of leaf disc and whole-leaf LMA

At the TRF site, we collected leaf samples from trees within reach of a canopy crane (88 m tall with a 60 m long boom). In the HDS, and STF sites, we used a 12-m long pruner to collect samples for most of the target species. In the case of tall individuals that were out of the reach of pruners, we used rope climbing to reach the canopy, then sampled branches with the long pruner. In Yakushima, we used 15-m poles for our leaf sampling. At least six sunexposed healthy leaves were sampled from each of 3 to 6 individuals for each species. Trait values were averaged at the species- and individual-level for the analyses. For compound-leaved species, we referred to a leaflet (the minimum photosynthetic unit) as a single leaf in our study. We excluded species that had tiny leaves or leaflets (length <1 cm) because it was not practical to measure leaf disc LMA.

To determine what extent LMA values were affected by the different methods, LMA were determined (i) LMA based on a whole leaf including petioles and (ii) LMA based on leaf disc avoiding thick veins (first to second veins). Three 0.6-cm diameter discs were taken from the base, middle and tip of each leaf using a hole punch in the Yunnan dataset. Two 1.0-cm diameter discs were taken from each leaf in the Yakushima dataset (see Appendix S2 S1, Figure S1 for more details). Fresh leaf area (LA; cm⁻²) of leaf materials (whole leaves including petioles and midribs) was measured using a scanner and ImageJ software by the R package *LeafArea*

(Katabuchi, 2015). Fresh leaf thickness (LT; mm) was measured using a micrometer in three points at the base, middle and tip of the leaf using a micrometer (Mitutoyo 293-240, Mitu-171 toyo Corporation, Kawasaki, Japan) with a precision at 0.001 mm. Thickness of each leaf 172 disc was also measured for the Yunnan dataset but not for the Yakushima dataset, which 173 enables us to estimate whole-leaf and leaf disc leaf tissue density (LD; g cm⁻³) separately in 174 the Yunnan dataset. Leaf dry mass (whole leaves including petioles and midribs, and leaf discs) was recorded after drying 80°C for over 48 h to constant weight using an electronic 176 scale (Mettler-Toledo MS204TS, Columbus, OHOhio, U.S.A.) with a precision at 0.0001 g. In 177 the subset of Yakushima dataset, leaf lamina and petiole dry mass were also determined 178 separately (80 out of 193 species) to quantify leaf support costs. Dry mass for leaf discs were 179 recorded 2-3 discs together for each leaf. Based on those measurements of whole-leaf and 180 leaf disc samples, LMA (g m⁻²) was calculated as the ratio of leaf dry mass to fresh leaf area, 181 and LD was calculated as the ratio of LMA to LT. We foucs on LMA in our study and we do not 182 present results of LD in the main text, because differences between whole-leaf LD and leaf 183 disc LD only depend on the ratio between whole-leaf LMA and leaf disc LMA and measure-184 ment errors of leaf thickness (Appendix \$3\$1, Figure \$2). 185

Relationship between whole-leaf and leaf disc LMA

A bivariate trait relationship between disc-based and whole-leaf log-transformed LMA was summarized with variance explained (R^2) based on Pearson correlations and with a standardized major axis (SMA) regression (Warton et al., 2006) across all the species.

A SMA analysis was run using the R package *smatr* (Warton et al., 2012); this included tests for slope heterogeneity and elevation differences between disc-based and whole-leaf estimates of LMA values. Confidence intervals of SMA estimates were based on 2000 bootstraps.

193 Effects of leaf morphology on the relationship between whole-leaf and leaf disc LMA

To further quantify the effect of leaf morphology and leaf punch size on the relationship between whole-leaf and leaf disc LMA, we built a hierarchical Bayesian model. The whole-leaf LMA was assumed to follow a normal distribution (N) with the mean of leaf disc LMA and covariates of leaf tissue density, leaf area and leaf thickness on the log-scale;

$$\ln(LMA_{Wi}) \sim N(\ln(LMA_{Di}) + \tilde{\mu}_i, \sigma_i^2)$$

$$\tilde{\mu}_i = \beta_0 + \beta_1 \ln(LD_i) + \beta_2 \ln(LA_i) + \beta_3 \ln(LT_i) + \beta_4 PS_i$$

$$+ PS_i \times \left\{ \beta_5 \ln(LD_i) + \beta_6 \ln(LA_i) + \beta_7 \ln(LT_i) \right\}$$
(2)

where LMA_{Wi} , LMA_{Di} , LD_i , LA_i , and LT_i are the observed whole-leaf LMA, leaf disc LMA, leaf tissue density, leaf area, and leaf thickness for species i, respectively, The dummy variable PS_i is set to 1 for samples obtained with a small leaf punch (0.6-cm diameter) and 0 for samples obtained with a large leaf punch (1.0-cm diameter). $\tilde{\mu}_i$ is the effect of leaf tissue

density, leaf area, leaf thickness and leaf punch size on the mean estimates for the whole-leaf LMA of species i, σ_i^2 is the variance of whole-leaf LMA for species i, and β are coefficients. Since leaf disc LMA is used as an offset, this model can be used to see the realtionship between whole-leaf to leaf disc LMA ratio and predictors. The standard deviation of whole-leaf LMA (σ_i) was assumed to follow a normal distribution;

$$\ln(\sigma_i) \sim N(\ln(\tilde{\sigma}_i), \omega^2)$$
 (3)

$$\ln(\tilde{\sigma}_i) = \gamma_0 + \gamma_1 \ln(LD_i) + \gamma_2 \ln(LA_i) + \gamma_3 \ln(LT_i) + \gamma_4 P S_i$$

$$+ P S_i \times \left\{ \gamma_5 \ln(LD_i) + \gamma_6 \ln(LA_i) + \gamma_7 \ln(LT_i) \right\}$$
(4)

where $\tilde{\sigma_i}$ is a hyperparameter for σ_i , ω^2 is a scaling hyperparameter, and γ are coefficients. To allow comparisons among parameter estimates, $\ln(LD)$, $\ln(LA)$, and $\ln(LT)$ were scaled to a mean of 0 and standard deviation of 1. Note that this model does not include measurement errors of leaf disc LMA implemented in the previous SMA analysis, because there are too many scaling parameters

Posterior distributions of all parameters were estimated using the Hamiltonian Monte Carlo algorithm (HMC) implemented in Stan (Carpenter et al., 2017) using the weakly-informative priors (Gelman et al., 2008). The Stan code use to fit models is available from Github at: https://github.com/mattocci27/leaf-disc. Convergence of the posterior distribution was assessed with the Gelman-Rubin statistic with a convergence threshold of 1.1 for all parameters (Gelman et al., 2013).

We also performed ordinary least-squares (OLS) regressions to see the relationship between whole-leaf to leaf disc LMA ratio and petiole to leaf dry mass ratio and between leaf area petiole to leaf dry mass ratio, using a subset of the dataset in which we measured leaf lamina and petiole dry mass separately.

222 Relationship between disc-based and whole-leaf estimates of intraspecific variation

The extent of intraspecific variation (ITV) was calculated using Bao's CV estimator (Yang et al., 2020);

$$CV = CV_1 - \frac{CV_1^3}{N} + \frac{CV_1}{4N} + \frac{CV_1^2 \gamma_1}{2N} + \frac{CV_1 \gamma_2}{8N}$$
 (5)

where $CV_1=s/\bar{x}$, s is the sample standard deviation and \bar{x} is the sample mean, N is the sample size, and γ_1 and γ_2 are the skewness and kurtosis of the trait distribution. Bao's CV estimator is a robust method to quantify ITV based on a coefficient of variation (CV) when sample size is small (Yang et al., 2020). We calculated CV for species that have at least 5 individuals as recommended in Westerband et al. (2021). Ideally, 20 individuals for each

species are required for robust estimation of CV [i.e., <5% differences from the true ITV; Yang et al. (2020)].

SMA analyses were run as described above. We also performed the paired t-test to compare the extent of ITV between whole-leaf LMA and leaf disc LMA. All statistical analyses were conducted in R version 4.1.3 (R Core Team, 2022) - using the R package *targets* version 0.12.0 for workflow management (Landau, 2021)

RESULTS

There was a significant shift in elevation between whole-leaf and leaf disc LMA (Fig.1, SMA slope: 0.975 [95%CI: 0.938, 1.013], SMA intercept: 0.0881 [95%CI: 0.016, 0.16]), suggesting that the whole-leaf estimates of LMA were 9.689.69% [95%CI: 8.05, 11.48.06, 11.3] greater than disc-based estimates in the arithmetic scale.

Leaf tissue density, leaf area, leaf thickness and leaf punch size influenced the relationship 241 between the whole-leaf LMA and leaf disc LMA (Fig. 2 and Appendix \$4\$1, Figure \$3). Species 242 with lower leaf tissue density had greater variance in the estimate of whole-leaf to leaf disc 243 LMA ratio (Fig. 2a). When a small leaf punch was used, this pattern was more pronounced 244 and leaf disc and whole-leaf LMA estimate diverged for species with higher leaf tissue den-245 sity (Fig. 2a). Species with larger leaves had greater estimate for the whole-leaf to leaf disc 246 LMA ratio (Fig. 2b), which was only significant for leaves obtained by a large punch. Species with thinner leaves had greater variance in the estimate of whole-leaf to leaf disc LMA ratio 248 (Fig. 2c), which was only significant for leaves obtained by a small punch. A small punch also 249 inflated variance in the estimate of whole-leaf to leaf disc LMA ratio (Fig. 2a-c and Appendix 250 S4S1, Figure S3). Leaf disc and whole-leaf LMA estimate diverged for species with greater 251 investment in petiole dry mass (Fig. 3a). Large-leaved species tended to have greater invest-252 ment for petiole dry mass than small-leaved species (Fig. 3b).

The extent of intraspecific trait variation (CV; coefficient of variation) only weakly matched between whole-leaf and disc-based estimates (Fig.4). Although there were no statistical differences in the SMA slope or elevation (SMA slope: $\frac{1.116}{1.112}$ [95%CI: $\frac{0.954}{0.954}$, $\frac{1.304}{0.951}$, $\frac{1.302}{0.951}$, SMA intercept: $\frac{0.102}{0.096}$ [95%CI: $\frac{0.174}{0.377}$ -0.18, 0.372]), leaf disc LMA had greater CVs than whole-leaf LMA ($t = \frac{3.19}{3.2}$, $d.f. = \frac{144143}{0.951}$, $d.f. = \frac{144143}{0.951}$,

59 DISCUSSION

Leaf disc LMA explained interspecific variations in whole-leaf LMA ($R^2 = 0.87$, n = 334) quite well, and the strength of their relationship was approximately same or slightly weaker than previously reported ($R^2 = 0.92$, n = 409, for whole-leaves in Kraft et al., 2008; $R^2 = 0.92$, n = 364

for leaf laminas in Onoda et al., 2011). Standardized major axis (SMA) regression analysis showed significantly greater intercepts when all the species were pooled (9.689.69% differences). Together, these findings suggest that disc-based estimates can be generally used as good proxies for their whole-leaf estimates with appropriate calibration (see discussion below). However, leaf tissue density, leaf area, leaf thickness and leaf punch size also influenced the relationship between the whole-leaf LMA and leaf disc LMA (Fig. 2 and Appendix S4S1, Figure S3).

270 Effects of leaf punch size and leaf morphology

The effect of leaf punch size was the most important source of the variation (Fig. 2, Ap-271 pendix \$4\$1, Figure \$3), and samples obtained with a small leaf punch showed greater variance in whole-leaf to leaf disc LMA ratio. Small leaf disc size inflates trait varition within 273 leaves because dry mass will be inflated when major veins are unintentionally included in 274 the samples. Species with lower leaf tissue density and thin leaves showed greater variance 275 in whole-leaf to leaf disc LMA ratio especially when a small punch was used (Fig.2ac: small 276 leaf punch, Fig. 2a: large leaf punch). Kraft et al. (2008) found high R^2 values ($R^2 = 0.92$) be-277 tween whole-leaf and leaf disc LMA based on leaf discs of approximately 1.0-cm diameter for sapling leaves under closed canopy that are softer and thinner than adult leaves (Kita-279 jima and Poorter, 2010; Oktavia and Jin, 2020). Altough a leaf punch of 1.0-cm diameter was 280 used only in the Yakushima dataset in our study, the variance in the whole-leaf to leaf disc 281 LMA ratio were small for the Yakushima dataset, regardless of leaf tissue density or leaf thick-282 ness (Fig. 2ac). On the other hand, a previous study found that leaf disc LMA obtained with 283 a very small leaf punch (0.3-cm diameter) and whole-leaf LMA had different relationships 284 between shade tolerance (Poorter, 2009), suggesting that leaf disc LMA and whole-leaf LMA 285 can be inherently different when small disc samples were used. The relatively low R^2 values 286 in the Yunnan dataset (Appendix \$551, Figure \$4) compared to the previous study and the 287 Yakushima dataset may suggest not only the effect of distinct veins on variance in LMA (John 288 et al., 2017) or distinct veins with kite-type appearances (Grubb, 1986) but also technical difficulty in measuring tiny weight of discs in lower LMA leaves. We found that estimates of 290 whole-leaf and leaf disc LMA diverged when total dry mass of leaf discs was small (Appendix 291 S6S1, Figure S5), and thus species with lower leaf tissue density and thin leaves, which have 292 smaller dry mass (Appendix \$7\$1, Table \$2), tended to show grater variation in whole leaf 293 whole-leaf to leaf disc LMA ratio especially when we used a smaller leaf punch.

Species with higher leaf tissue density and large leaves showed greater whole-leaf LMA than leaf disc LMA (Fig. 2ab: large leaf punch). We also observed that leaf disc and whole-leaf LMA estimate diverged for species with greater investment in petiole dry mass (Fig. 3a), and large-leaved species tended to have greater investment for petiole dry mass than small-leaved species (Fig. 3b). This is consistent with the pattern of large leaves disproportionately investing in more leaf veins and/or petioles (Niinemets et al., 2006, 2007; Sack et al., 2012) and the pattern of higher leaf tissue density associated with higher vein density (John et al.,

2017; Sancho-Knapik et al., 2020) because those driving factors of LMA are likely to be ignored in leaf disc samples. The estimated effects of leaf tissue density and leaf area was also positive for species obtained with a small leaf punch (Appendix S4S1, Figure S3), but those effects were not significant. This is probably because the variation in whole-leaf to leaf disc LMA ratio was too large to show clear effects of leaf tissue density and leaf area on the mean estimates (β in Eq. 2) when a small leaf punch was used.

308 Conversion between whole-leaf and leaf disc LMA

When the objective is to use leaf disc LMA as an alternative for whole-leaf LMA, using an ap-309 propriate size of leaf punch (e.g., 1.0-cm in diameter) is important for obtaining stable estimates of leaf disc LMA that matches well with whole-leaf LMA. Our results suggest that simple conversion between whole-leaf and leaf disc LMA is difficult for species obtained with a small 312 leaf punch. The effects of leaf tissue density and leaf area are predictable when we used a 313 large leaf punch (Fig. 2ab: large leaf punch), suggesting that conversion between leaf disc 314 and whole-leaf LMA should be possible using leaf area as a covariate (measuring leaf tissue 315 density requires whole-leaf LMA itself). The ordinary least-squares (OLS) regressions with 316 leaf area as a covariate as following: (1) $log_{10}LMA_w = 0.997 \times log_{10}LMA_d + 0.0589 \times log_{10}LA$, (2) $\log_{10} LMA_d = 0.15 + 0.945 \times \log_{10} LMA_w - 0.063 \times \log_{10} LA$ ($R^2 = 0.94$, AIC = -321.7, n = 193), 318 where LMA_w, LMA_d, and LA are the whole-leaf LMA, leaf disc LMA and leaf area, respectively. 319 Thei The OLS regressions without leaf area as a covariate as following: (3) log₁₀LMA_w = 0.124 320 $+0.958 \times \log_{10} LMA_d$ or (4) $\log_{10} LMA_d = 0.973 \times \log_{10} LMA_w$ ($R^2 = 0.93$, AIC = -284.2, n = 193). 321 [Note that OLS regressions minimize errors of either whole-leaf LMA or leaf disc LMA, and 322 thus those four models are not identical, whereas SMA regressions are not appropriate for 323 predictions (Warton et al., 2006)] Nonetheless, including leaf area only slightly improves the 324 R^2 values for the Yakushima dataset where a large punch was used. This is because the large 325 discrepancy in the whole-leaf to leaf disc LMA for large leaves can be explained by scaling re-326 lationships between leaf mass and leaf area called "diminishing returns" (Niklas et al., 2007) 327 which refers to increases in leaf dry mass including petioles do not result in the proportional increase in leaf area including petioles (Li, Shi, et al., 2022). This effect is generally strong 329 for mature leaves (Jiao et al., 2022). The "diminishing returns" can be either described as 330 the scaling slope of leaf area in the model 1 or the scaling slope of leaf-disc LMA and the in-331 tercept in the model 3. When one needs to mix leaf disc LMA and whole-leaf LMA in a single 332 analysis (e.g., meta-analysis) or quickly measure leaf disc LMA as a proxy for whole-leaf LMA, we recommend to convert LMA values using one of the models above. 334

Intraspecific variation

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Even though we used the same species, the same individuals and the same trait (LMA) to estimate the extents of intraspecific trait variation (ITV), the explained variance of the relationship between whole-leaf and disc-based estimate was surprisingly small (Fig. 3; $R^2 = \frac{0.090.08}{0.090.08}$). This is probably because leaf disc samples have more sources of ITV than wholeleaf samples (i.e., variation among leaf discs within the same leaves). On average, ITV based on leaf disc LMA was greater than ITV based on whole-leaf LMA as we expected. Although there are not many studies that quantified trait variation within a single leaf, a previous study that used a giant leaved species Alocasia macrorrhiza, suggests that LMA varied within leaves due to reduced water supply and demand from midrib to outer regions (Li et al., 2013). Because calculating the mean values generally diminishes the effect of within-sample variation, the relationship between disc-based estimates of individual mean LMA and its wholeleaf estimates will be stronger than the relationship between leaf disc CV and whole-leaf CV. Thus, leaf disc and whole-leaf estimates of species means will show the strongest relationship. The mismatch of ITV between whole-leaf and disc-based estimates could be ignored in the type of ITV studies that attempt to determine sources of trait variation (e.g., site, across species, within species) which often use mixed linear models and variance partitioning (e.g., Messier et al., 2010; Lepš et al., 2011; Siefert et al., 2015). However, it is likely to change patterns significantly in the type of ITV studies that attempt to quantify the extent of ITV which usually use CV (i.g., Albert et al., 2010; Bastias et al., 2017; Yang et al., 2020). This caveat might apply to other leaf chemical traits such as leaf N contents because those traits have both disc and whole-leaf based estimates but those two estimates are usually not clarified in trait databases (e.g., TRY). Since we used limited sample numbers (at least 5 samples per species) and most of them were obtained with a small punch, our study design may inflate errors in estimating ITV (Yang et al., 2020). Future studies should include more samples to improve the comparisons of leaf disc and whole-leaf based ITV.

Conclusions

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The present study was designed to examine the extent to which whole leaf whole-leaf and leaf disc LMA matched. Although our study was not designed to evaluate the effects of diameter sizes of leaf punches, the sizes of leaf punches as well as leaf morphology are the important sources of variance between whole-leaf and leaf disc LMA. Our study shows that mean values of leaf disc LMA is generally a good proxy for mean values of whole-leaf LMA when an appropriate size of leaf punch (e.g., 1.0-cm in diameter) was used. Those LMA values can be converted using OLS regressions. Additionally, although the mean values of leaf disc and whole-leaf LMA matched well, quantifying trait variation using leaf discs should be considered with caution, because variation among leaf discs within the same leaves (including measurement errors) seems to be considerably large but cannot be avoided with the typical sampling numbers (2-3 discs per leaf). Leaf discs, which can target leaf lamina avoiding leaf veins, are useful and essential for biochemical determinations such as measurements of Rubisco activity (Parry et al., 2002), because the photosynthetic activity occurs most in lamina rather than in veins for C₃ plants (Hibberd and Quick, 2002; Gao et al., 2018). Leaf disc LMA is also the preferred method for quantifying leaf lamina density, which avoid effects of veins. Accurately representing leaf traits will probably require careful selection between leaf disc and whole-leaf traits depending on the objectives.

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AUTHOR CONTRIBUTIONS

PM, MK, and YJC conceived the study; PM, YO, and YJC collected data; MK and CZ performed the analysis; MK and YJC led the writing of the paper; and all authors contributed to revisions.

DATA AVAILABILITY STATEMENT

Dataand codes for, codes and computing environments to reproduce this manuscript are archived and available on Zenodo (XXX)on Zenodo at https://doi.org/10.5281/zenodo. 6665083 and also available on Github at https://github.com/mattocci27/leaf-disc.

SUPPORTING INFORMATION

- Additional supporting information may be found online in the Supporting Information section at the end of the article.
- 402 **APPENDIX S1.** Supplementary tables and figures, including Tables S1–S2 and Figures S1–S5.
- Table S1. Site information.

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- ⁴⁰⁵ **APPENDIX Table S2.** Summary of a linear mixed model for log-transformed total dry mass of leaf discs.
- Figure \$1. Detailed sampling protocol for whole-leaf and leaf disc LMA.
- APPENDIX S3Figure S2. Generalization for a relationship between ratios of disc-based and whole-leaf estimates of leaf mass per area (LMA), leaf tissue density (LD), and leaf thickness (LT).
- APPENDIX S4Figure S3. Standardized regression coefficients modelig the effects of leaf tissue density, leaf area, leaf thickness, punch size, and their interactios (a) on the mean estimates of whole-leaf LMA, and (b) on the estimated variance of whole-leaf LMA.
- APPENDIX S5Figure S4. Relationships between species mean leaf mass per area (LMA) determined by using whole-leaves and leaf discs for species obtained with leaf punches of different diameters.
- 417 **APPENDIX S6Figure S5.** Relationships between whole-leaf LMA: disc leaf LMA and total dry
 418 mass for the leaf disc.
- 419 **APPENDIX S7.** Summary of a linear mixed model for log-transformed total dry mass of leaf 420 discs.

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Figure legends

- Fig. 1. Relationships between species mean leaf mass per area (LMA) determined by using whole-leaves and leaf discs. Dashed lines indicate 1:1 lines. Blue solid lines indicate standardized major axis (SMA) regressions. The 95% confidence intervals are resented represented as the shaded area. The correlation is significant (*P* < 0.001).
- Fig. 2. Model predictions of the whole-leaf to leaf disc LMA ratio as a function of leaf punch sizes and (a) leaf tissue density, (b) leaf area, and (c) leaf thickness. Dashed line indicates 1:1 ratio of leaf disc and whole-leaf estimates. Solid lines indicate the posterior means ($\tilde{\mu}$ in Eq. 1-2), and the shade regions shows \pm the posterior means of standard deviations ($\tilde{\sigma}$ in Eq. 3-4).
- Fig. 3. Relationships between petiole to leaf dry mass ratio and (a) leaf area and (b) wholeleaf to leaf disc LMA ratio. Blue solid lines indicate ordinary least squares (OLS) regressions.

 The 95% confidence intervals are presented as the shaded area. All the correlations are significant (P < 0.001). All-Note that all the samples were obtained with a diameter 1.0-cm leaf
 punch.
- Fig. 4. Relationships between coefficient of variation (Bao's CV estimator, Eq. 5) in LMA values determined by using whole leaves and leaf discs. The correlation is significant (*P* < 0.001). Details as in Fig. 1.