



Introduction to Nuclei Segmentation with StarDist

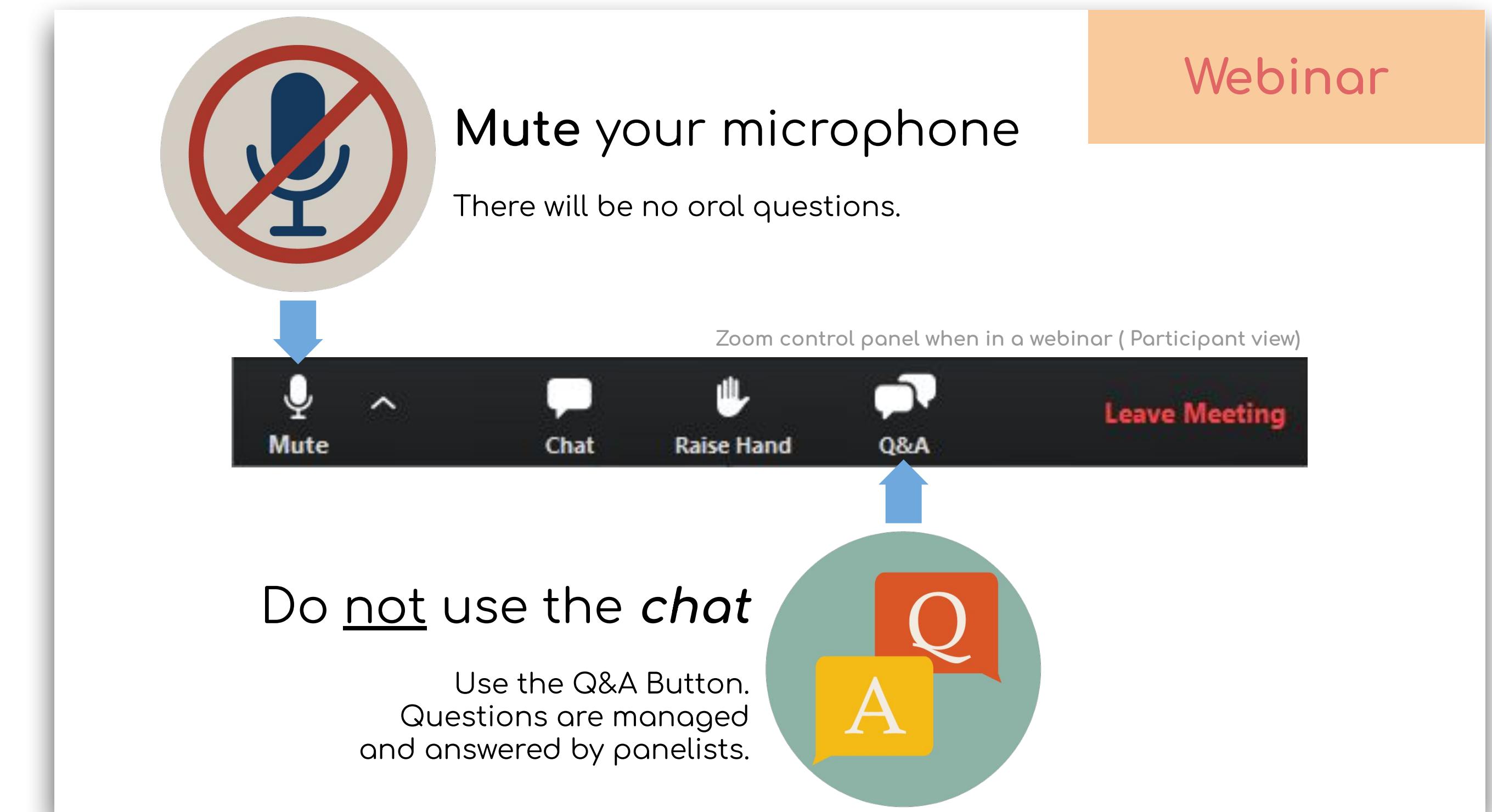


Martin Weigert, Olivier Burri, Siân Culley, Uwe Schmidt
Neubias Academy @ Home, April 28th, 2020

The webinar will start 15:40 CEST

If you are already here:

- 1. Please follow the instructions to the right**
- 2. Please answer the short questions in the poll**
(which should popup on your zoom screen)

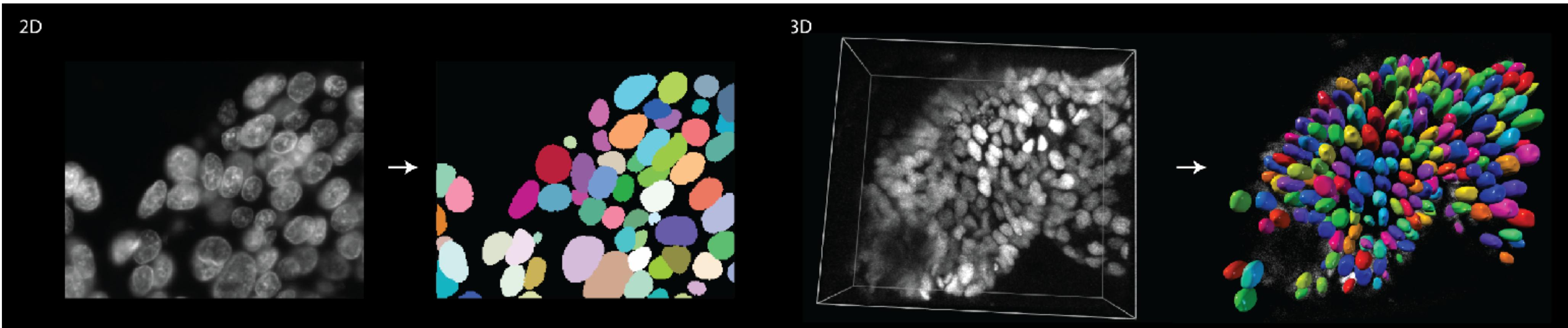




Introduction to Nuclei Segmentation with StarDist



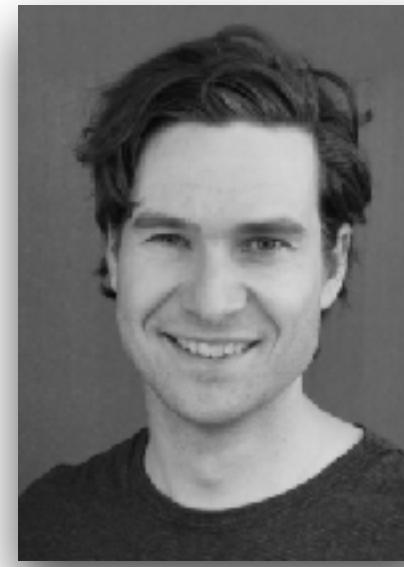
Martin Weigert, Olivier Burri, Siân Culley, Uwe Schmidt
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Webinar Material:

https://github.com/maweigert/neubias_academy_stardist

Speakers/Moderators:



Martin Weigert
EPFL

@martweig



Uwe Schmidt
MPI-CBG, Dresden
 CBG
Max Planck Institute
of Molecular Cell Biology
and Genetics

@uschmidt83



Olivier Burri
EPFL

@ChigureKun



Siân Culley
UCL, London
 UCL

@SuperResoluSian

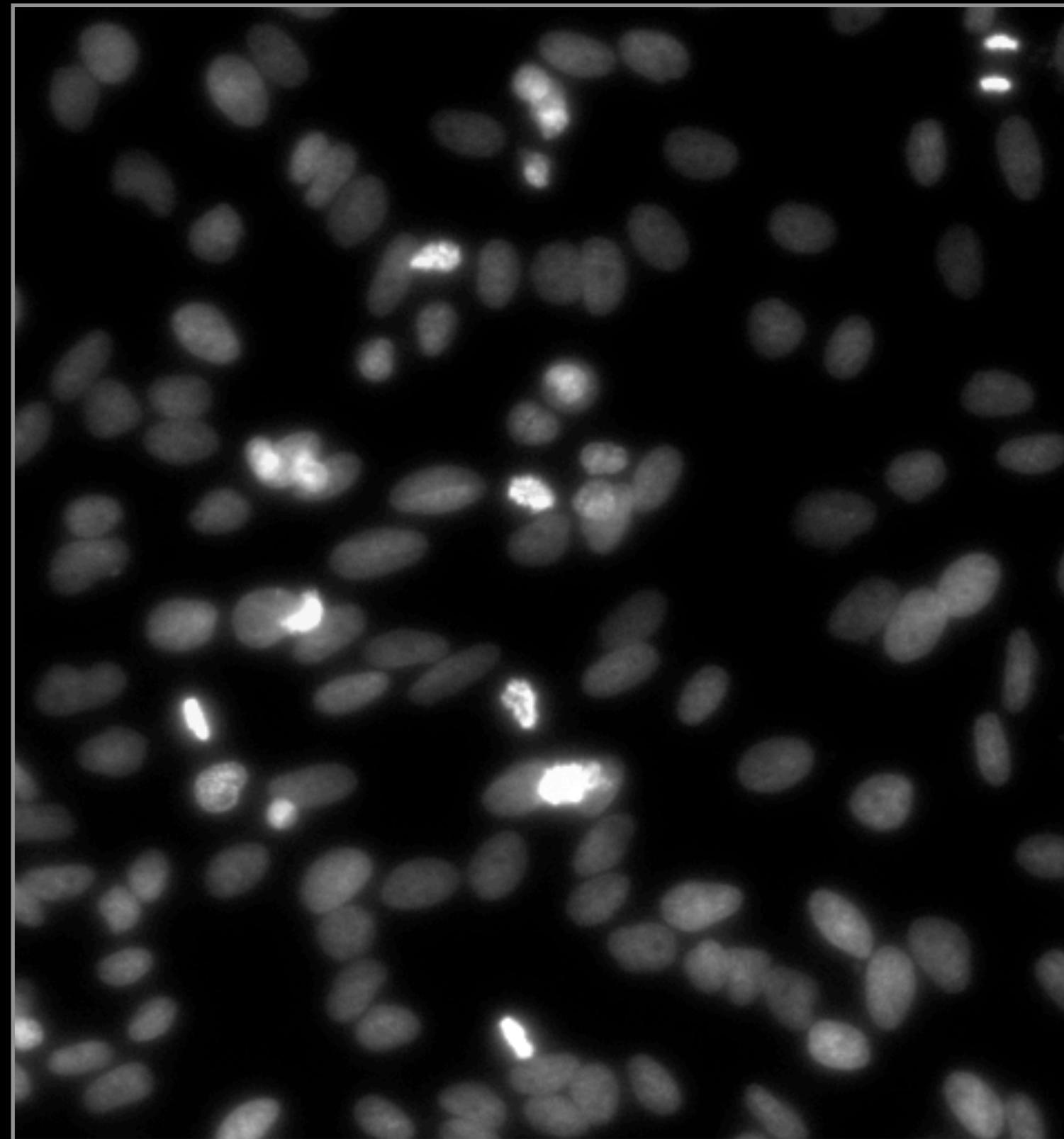
1. Introduction to nuclei segmentation and StarDist
2. Questions & Answers 1
3. How to use StarDist
4. StarDist in a core facility
5. Questions & Answers 2

- 1. Introduction to nuclei segmentation and StarDist**
- 2. Questions & Answers 1**
- 3. How to use StarDist**
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- 5. Questions & Answers 2**

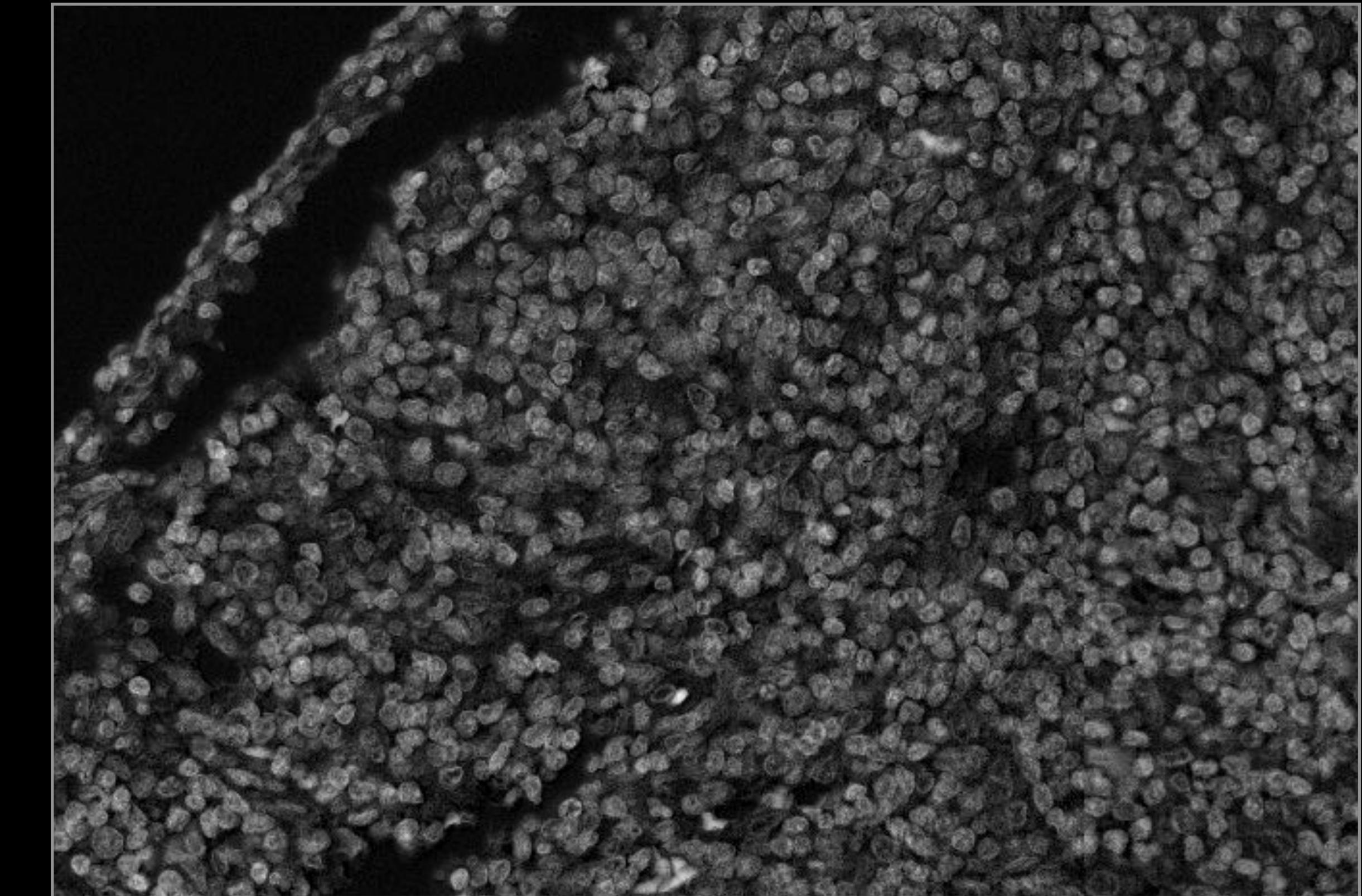
Introduction to nuclei segmentation and StarDist

Cell/Nuclei Segmentation in Microscopy

2D - Fluorescence



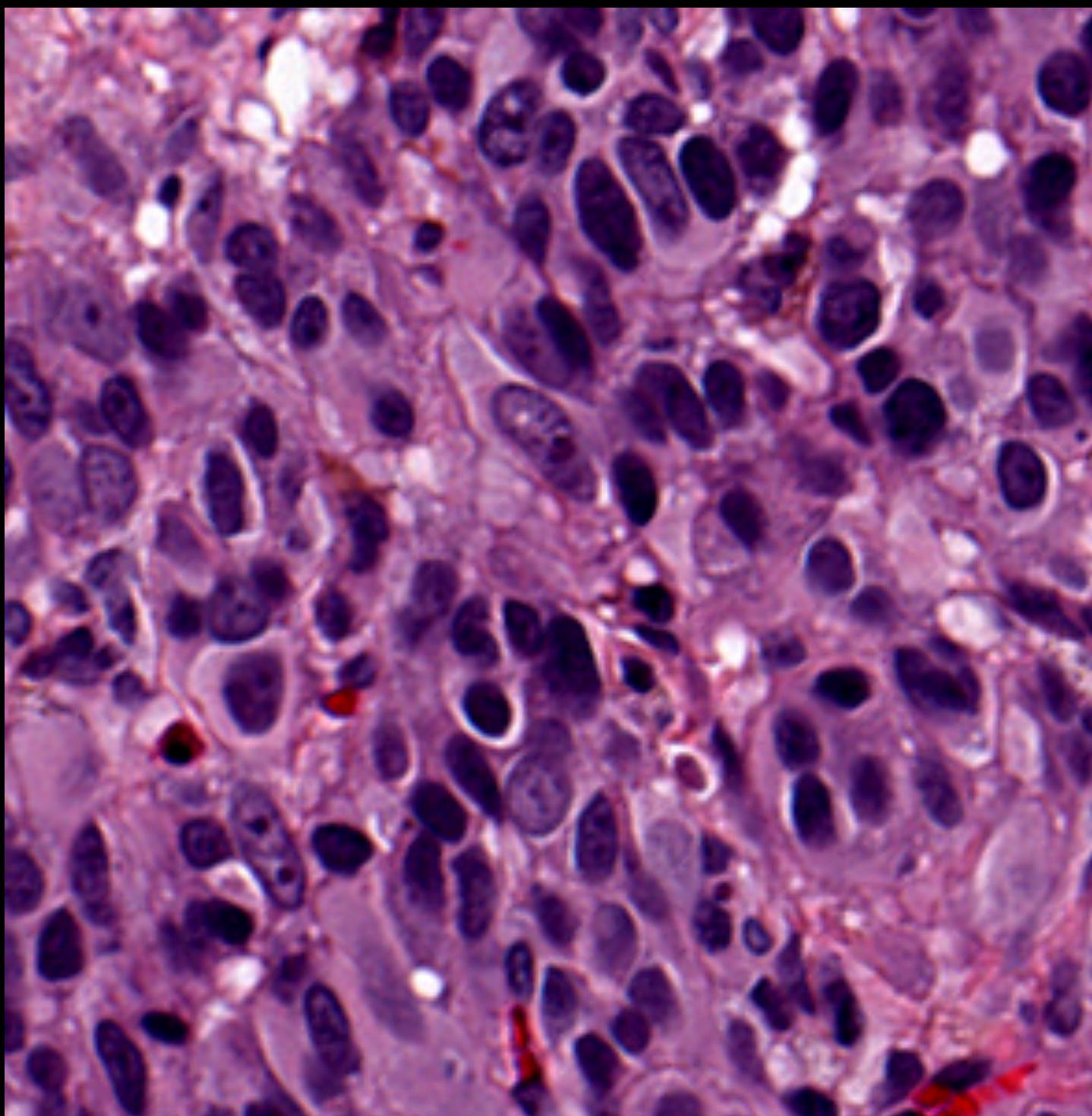
Data from Caicedo et al, 2019



Data from Anna Maria Tsakiroglou (Manchester)

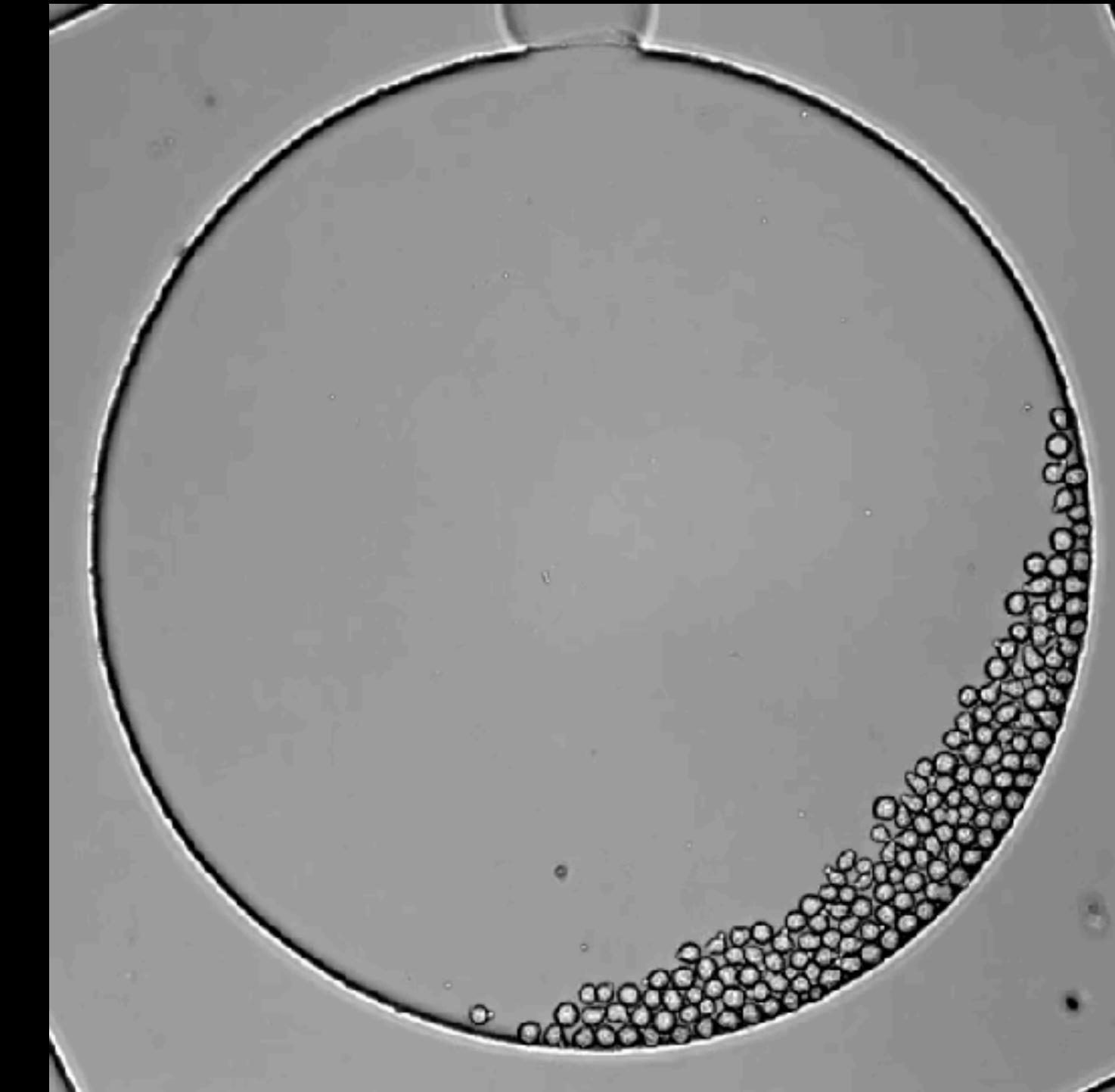
Cell/Nuclei Segmentation in Microscopy

2D - RGB/Histopathology



H&E stain, data from CancerImagingArchive

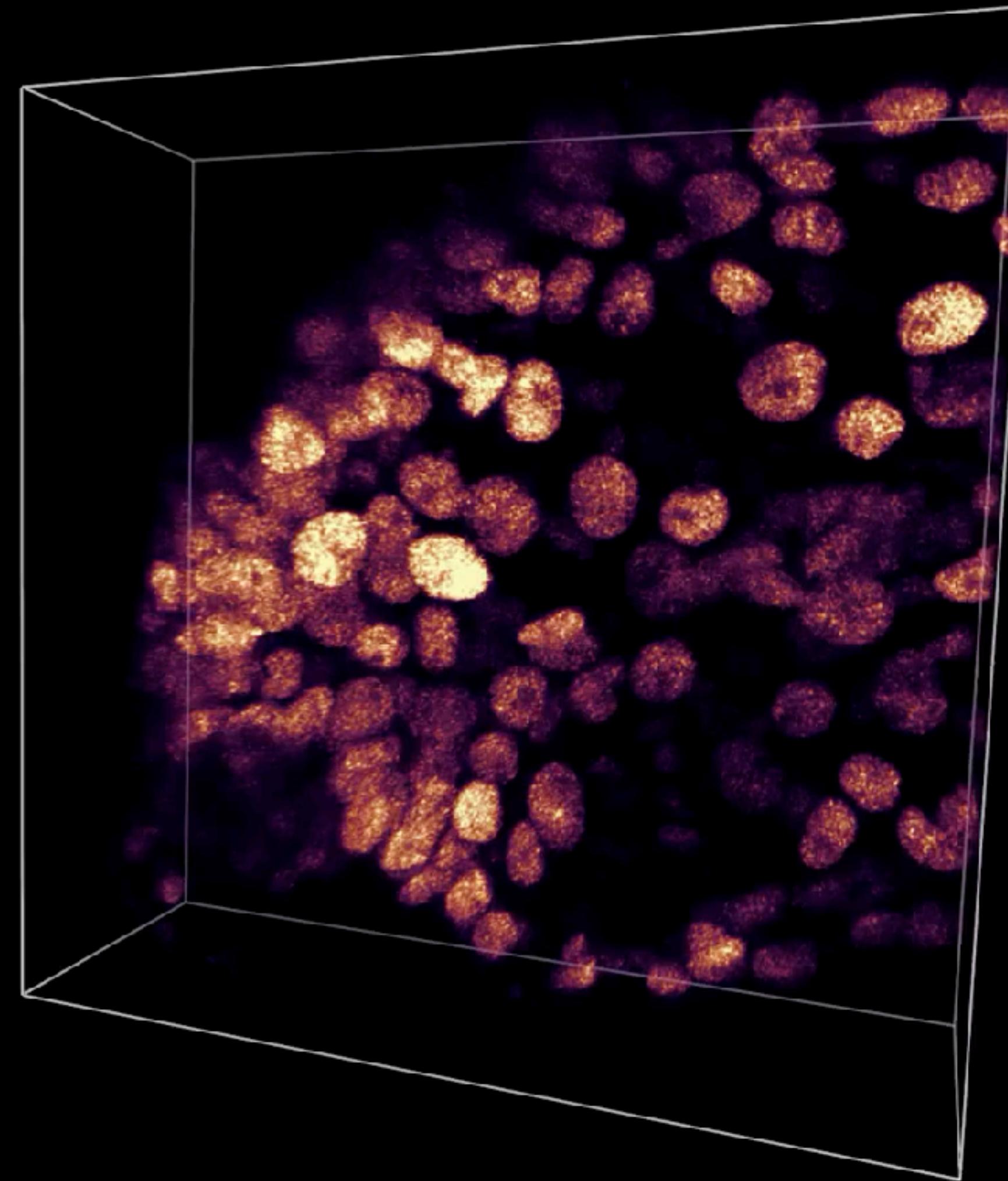
2D + time (Brightfield)



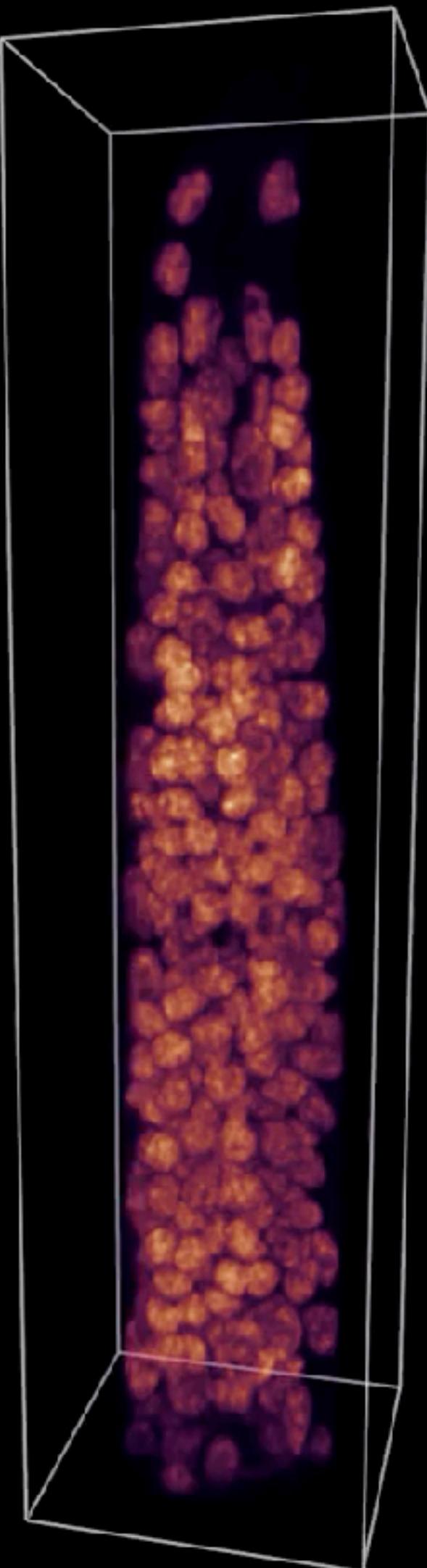
Mouse stem cells, data from cell tracking challenge

Cell/Nuclei Segmentation in Microscopy

3D (+time)



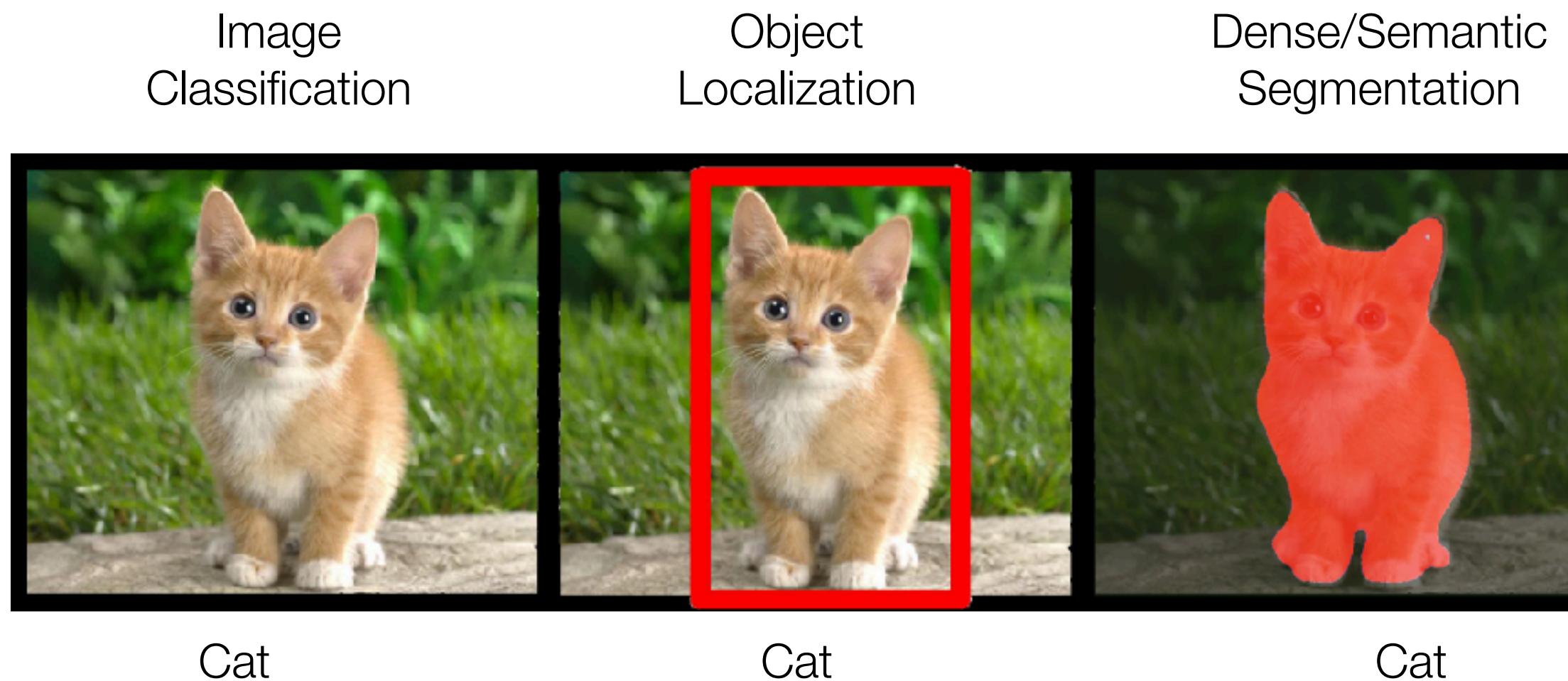
Paryphale Data from Ko Sugiyama



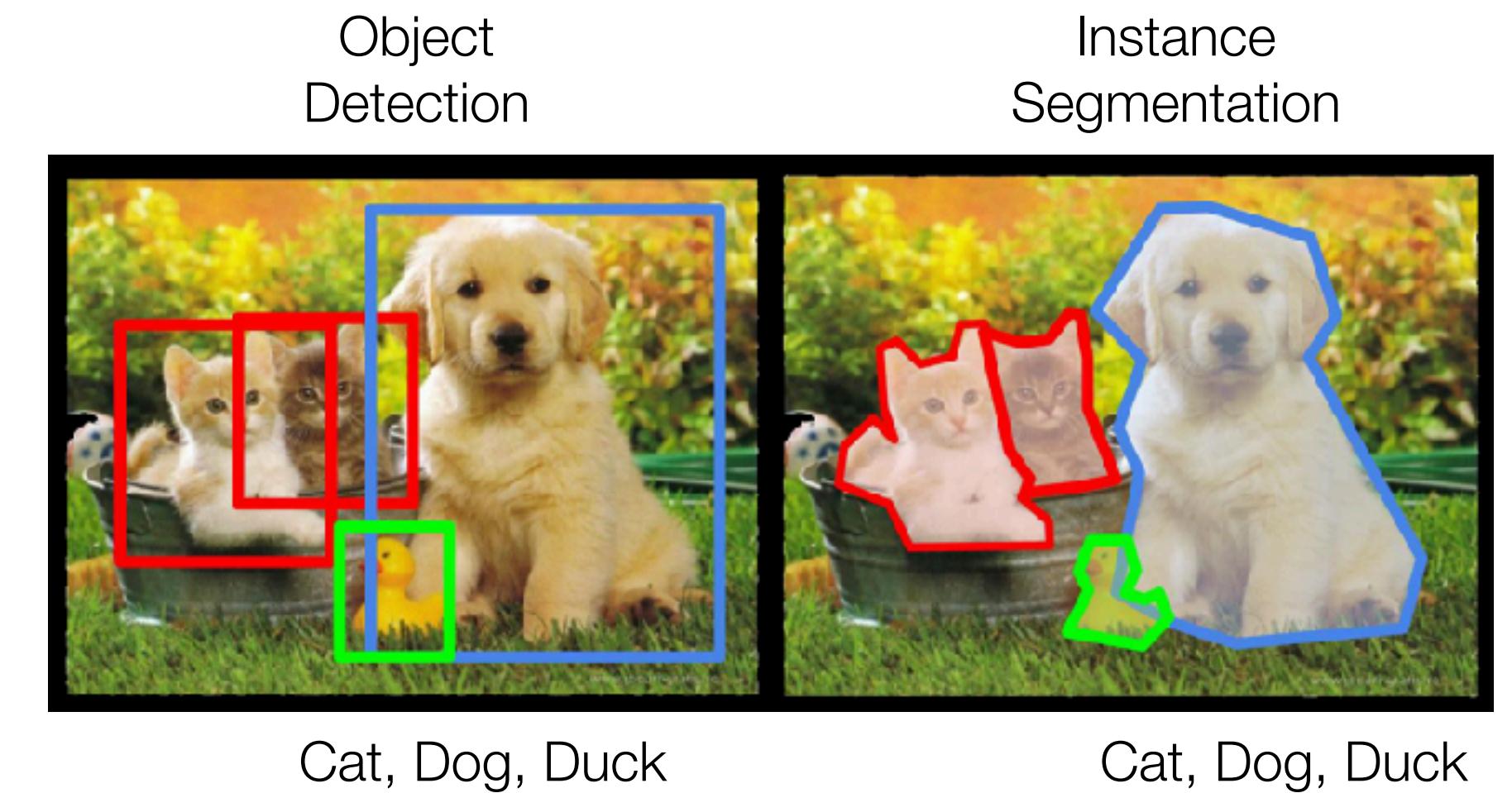
C. elegans Data from Dagmar Kainmüller

Computer Vision - Common Problems

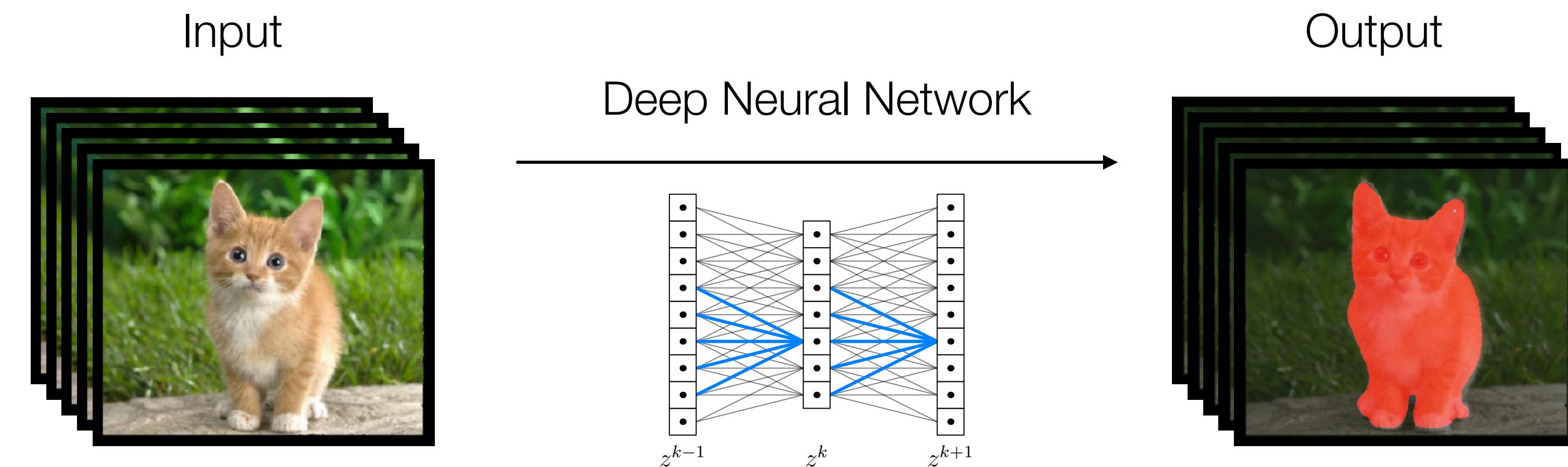
Single Object



Multiple Objects



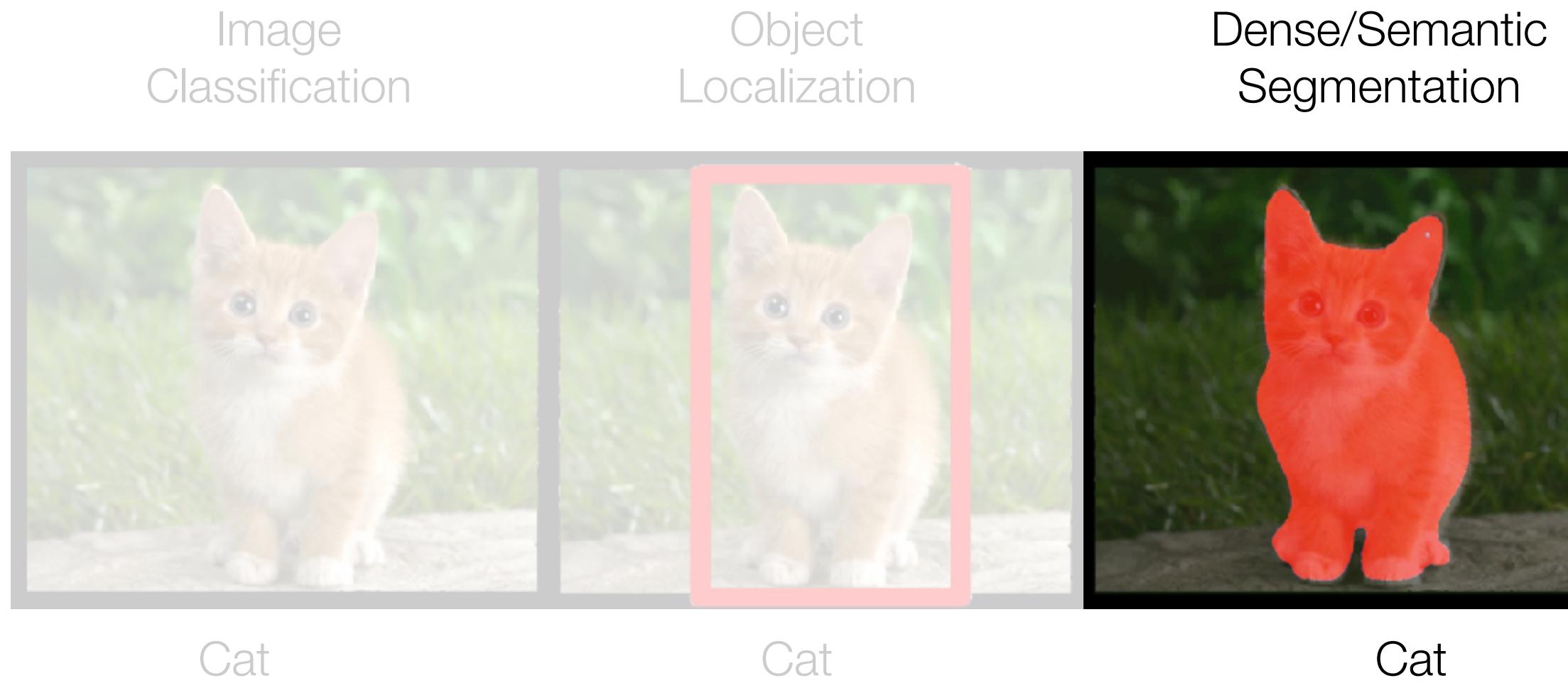
Currently the most successful paradigm: Supervised deep learning



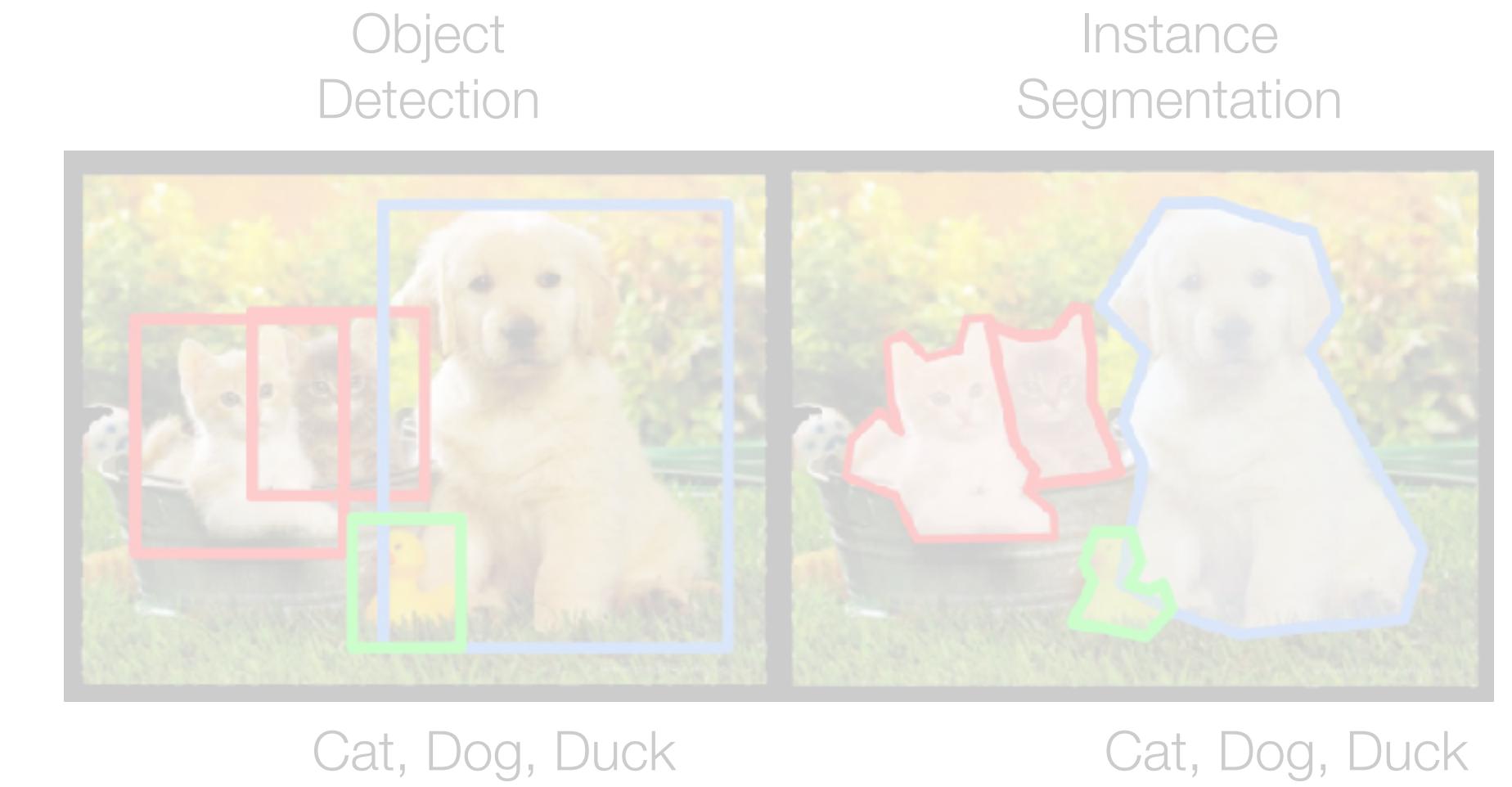
Deep (Convolutional) Neural Networks trained on annotated training data (ground truth, GT)

Computer Vision - Common Problems

Single Object

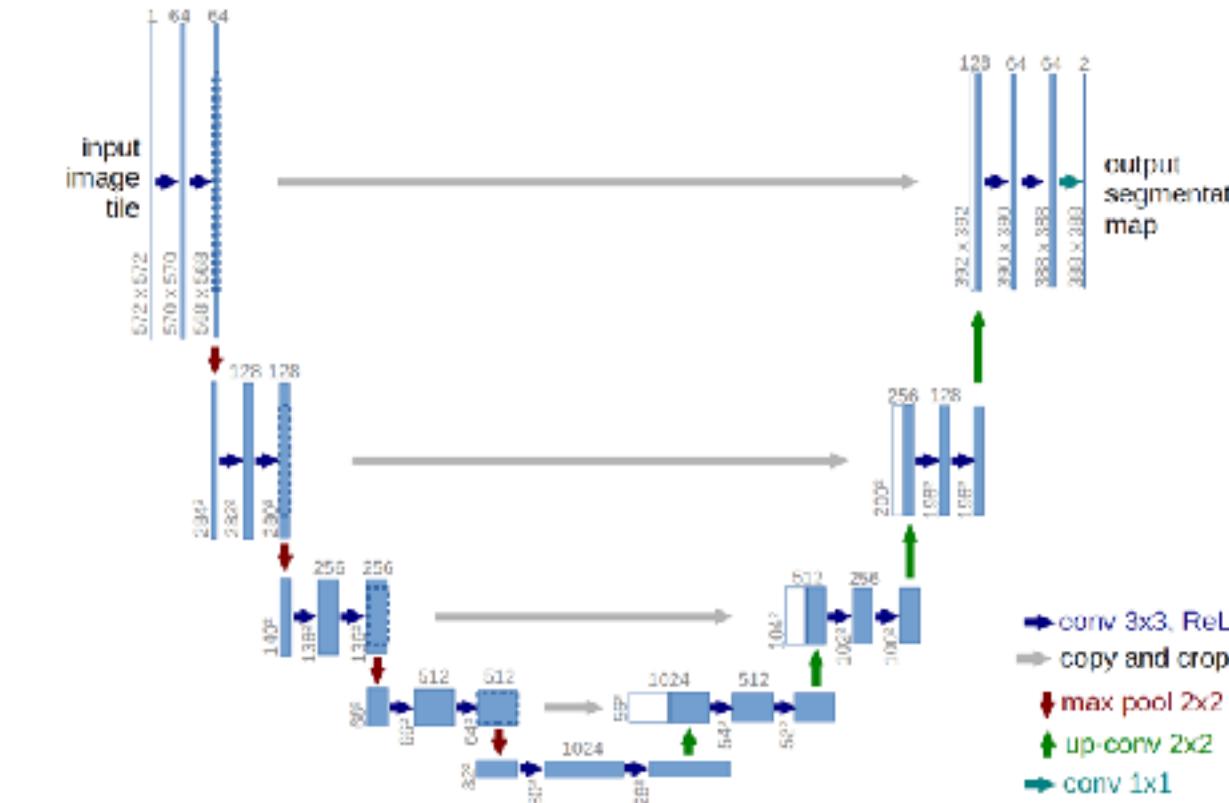


Multiple Objects



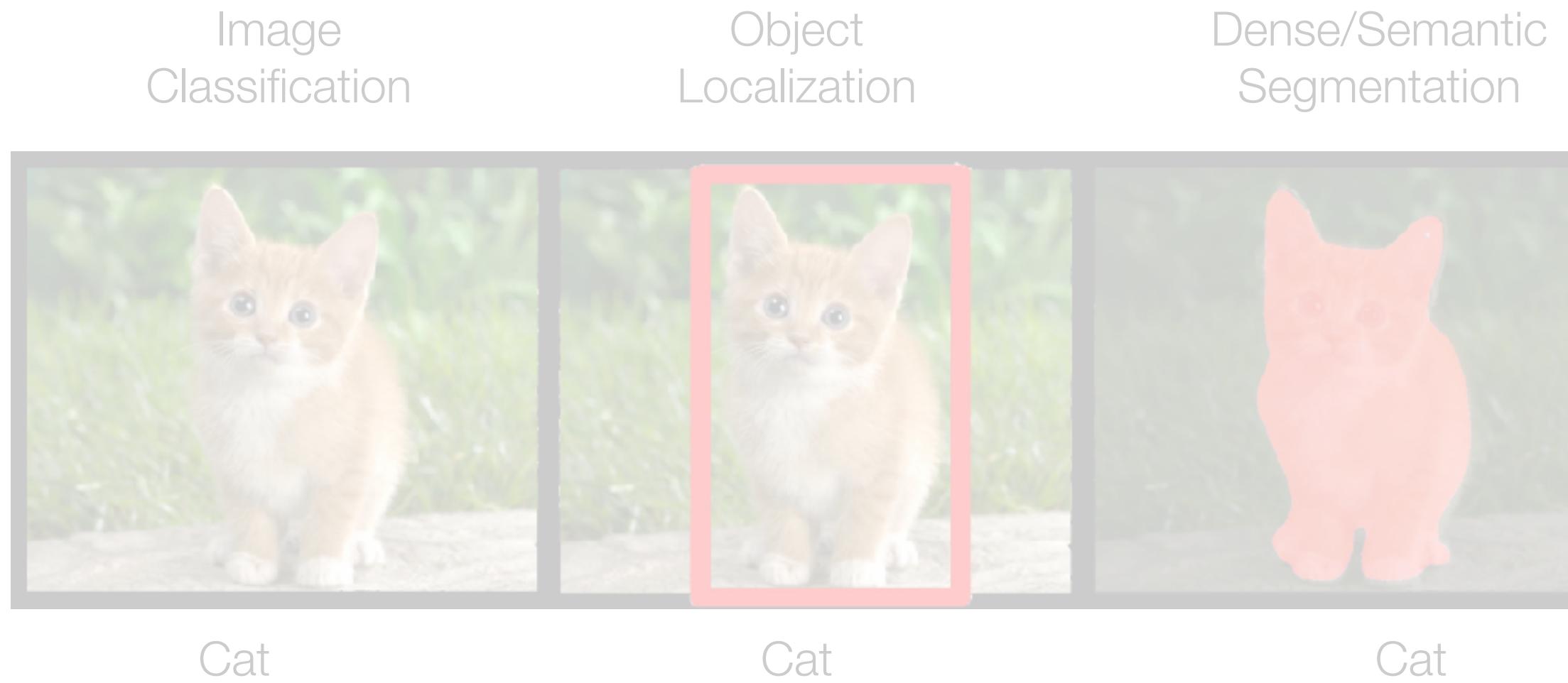
U-Net

Ronneberger et al (2015)

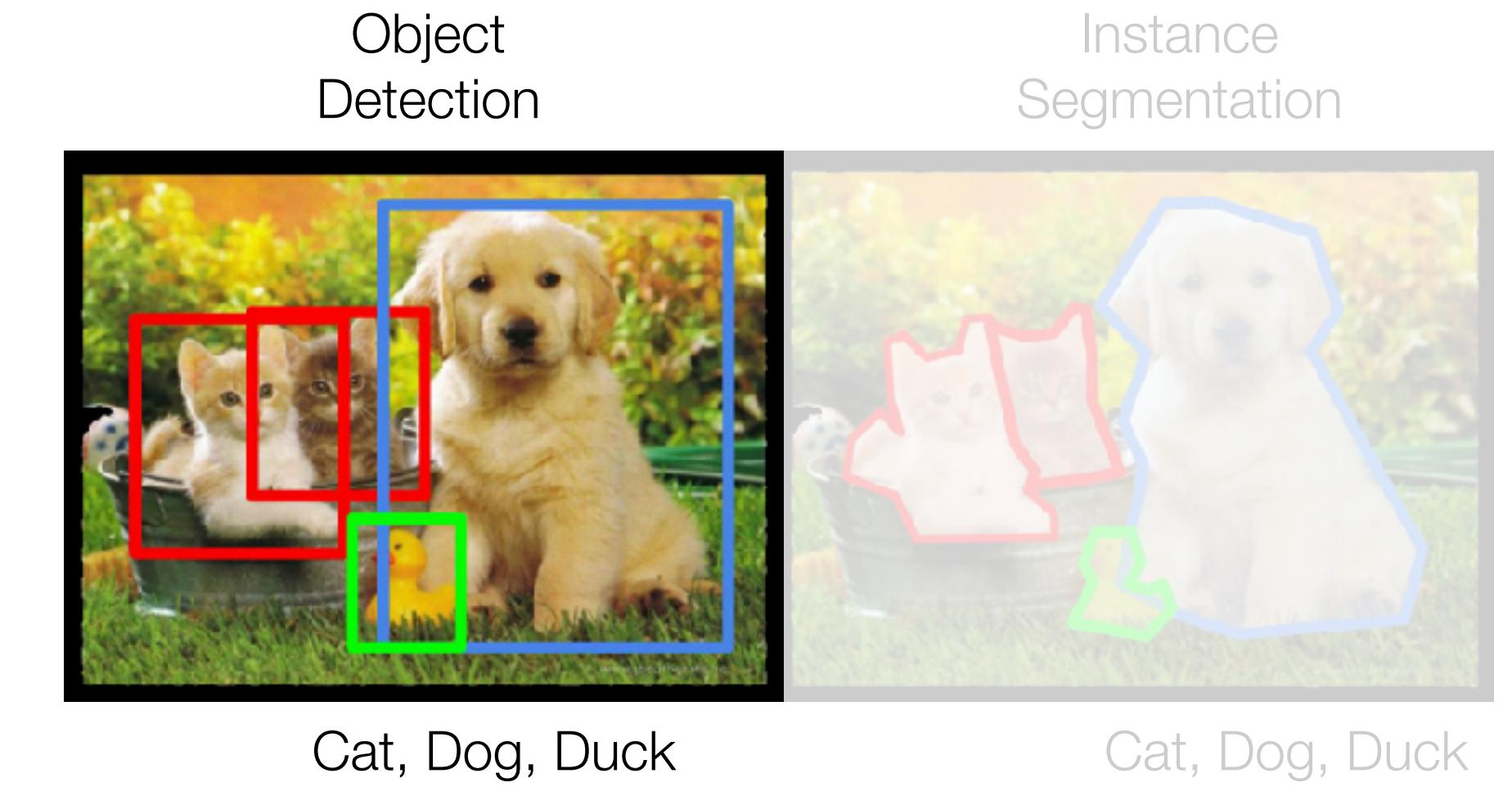


Computer Vision - Common Problems

Single Object

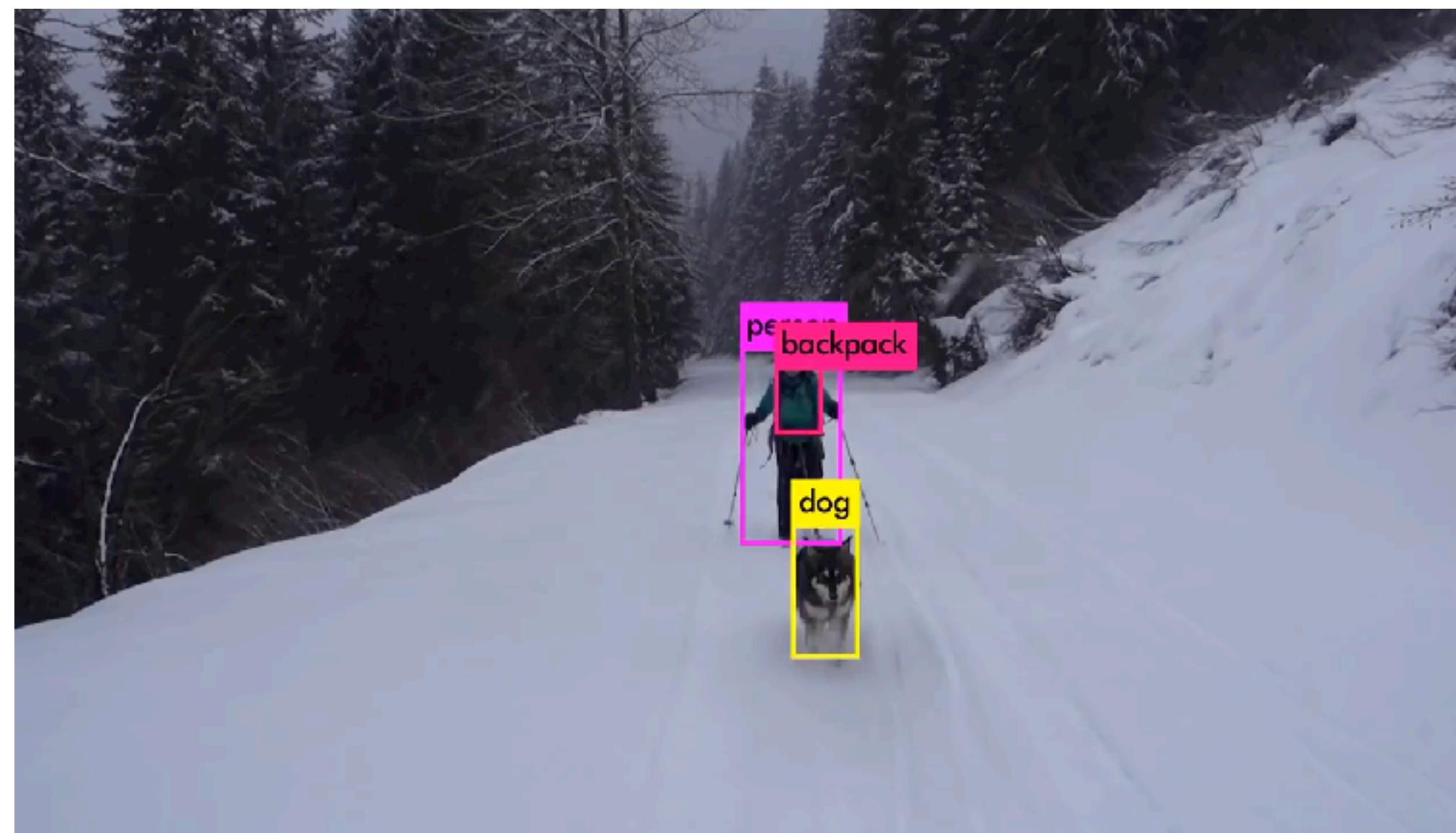


Multiple Objects



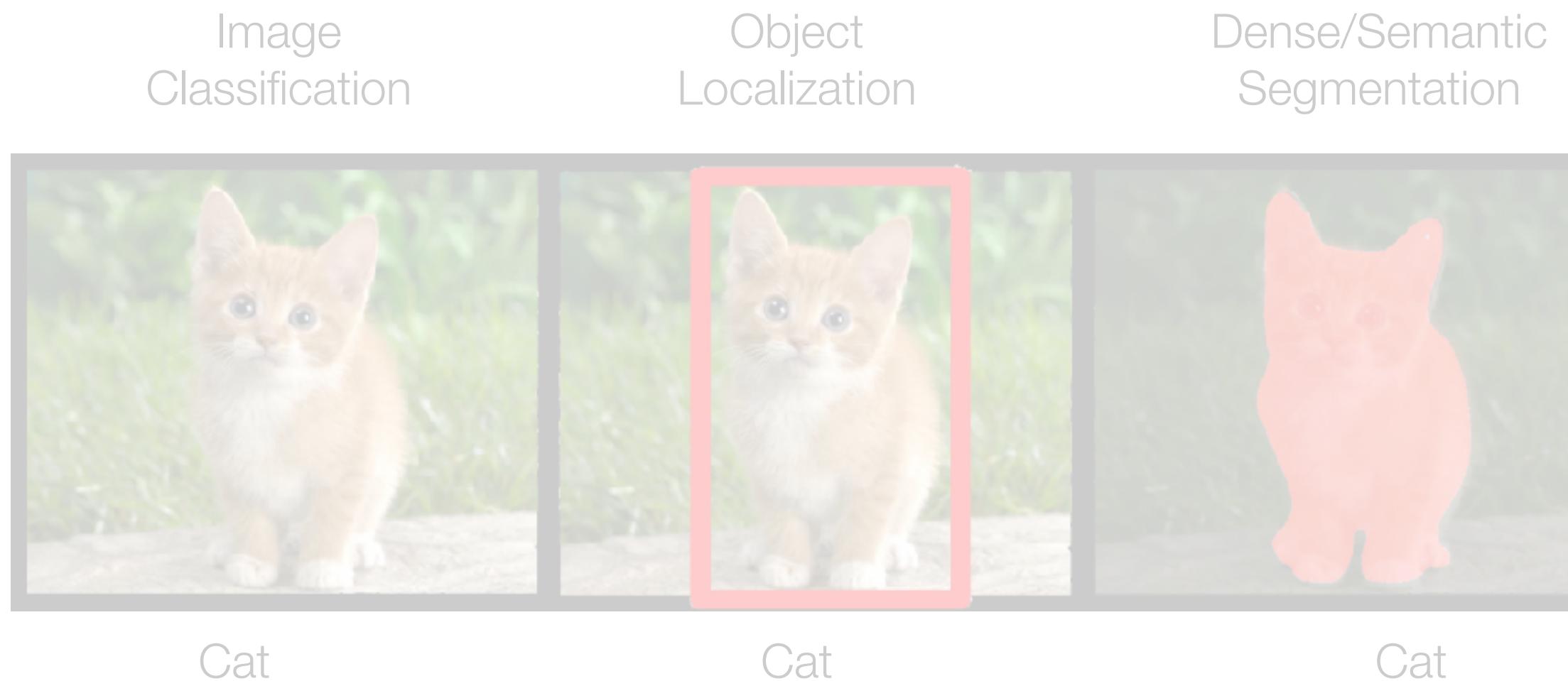
YOLO

Redmond et al (2016)

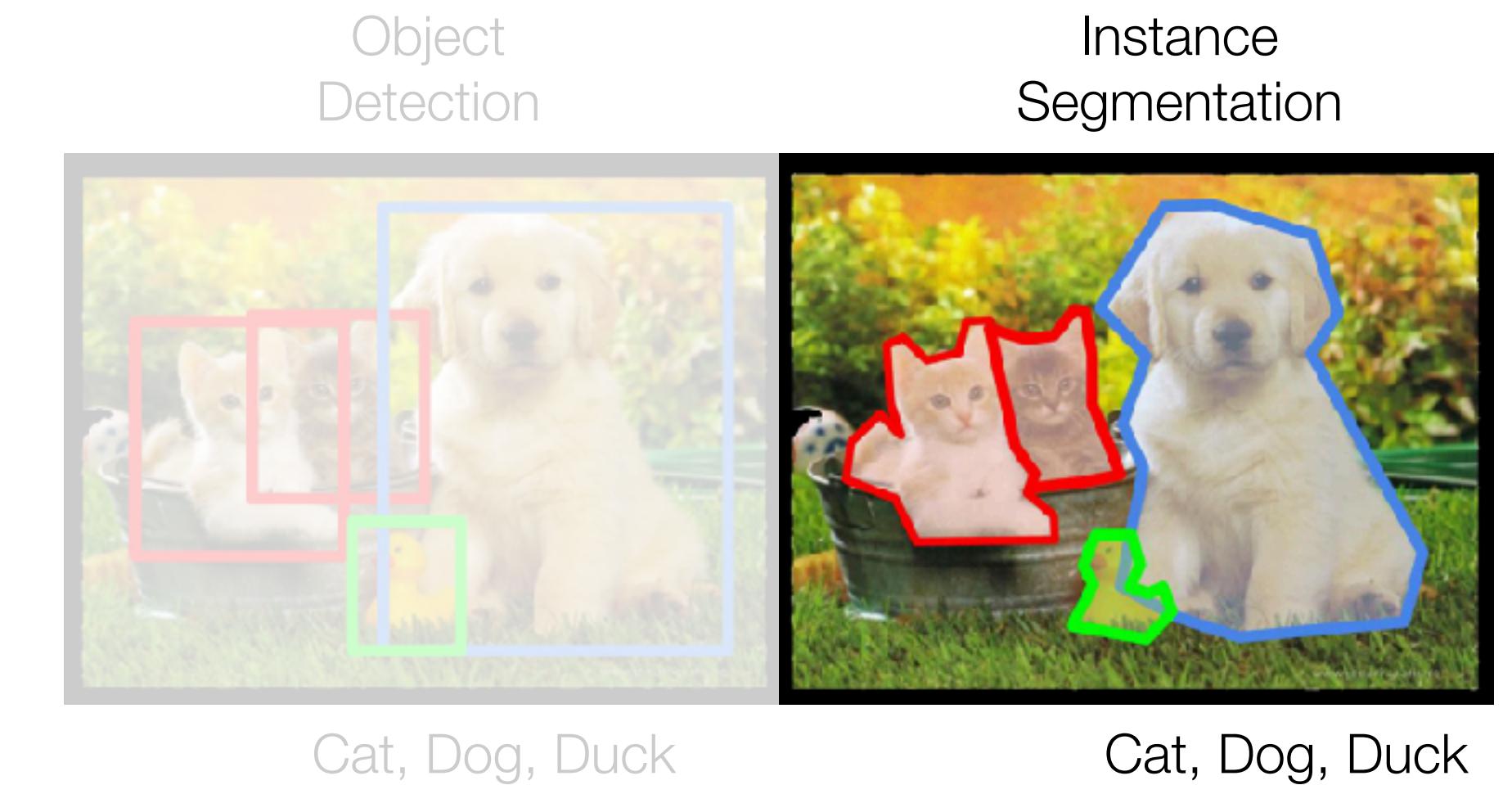


Computer Vision - Common Problems

Single Object



Multiple Objects

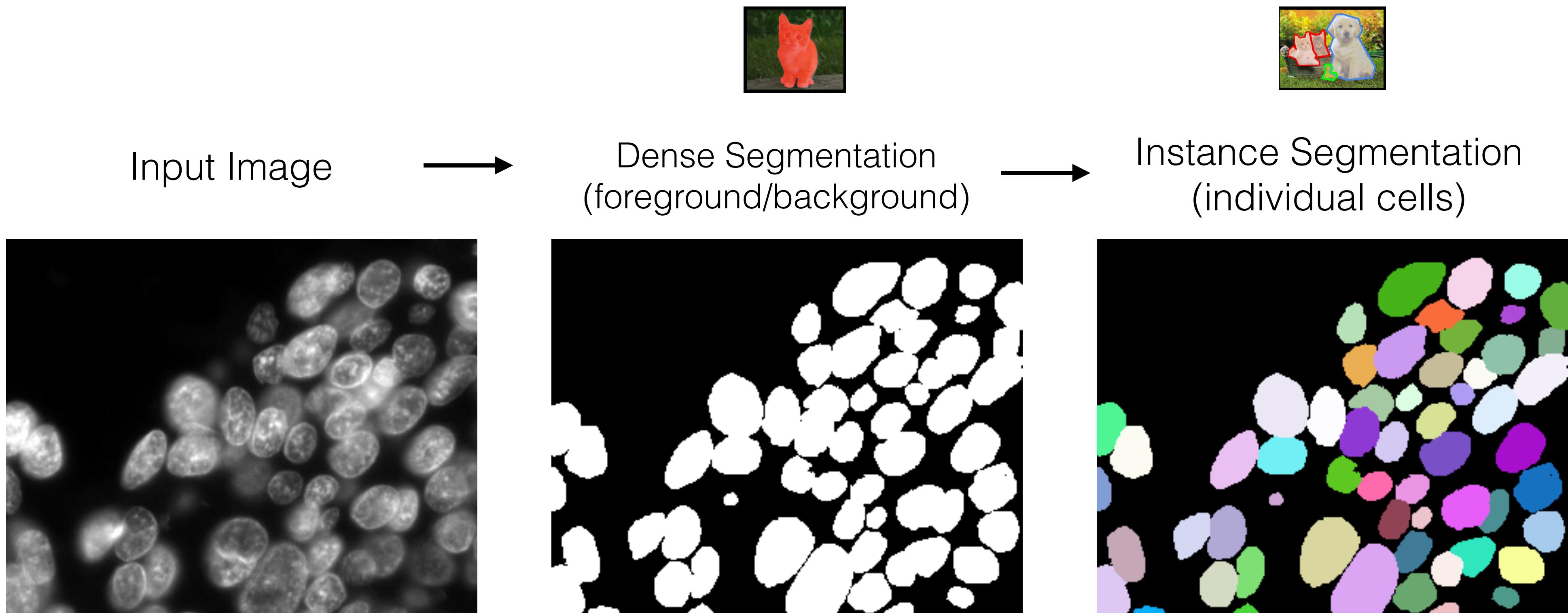


Mask-RCNN

He et al (2017)

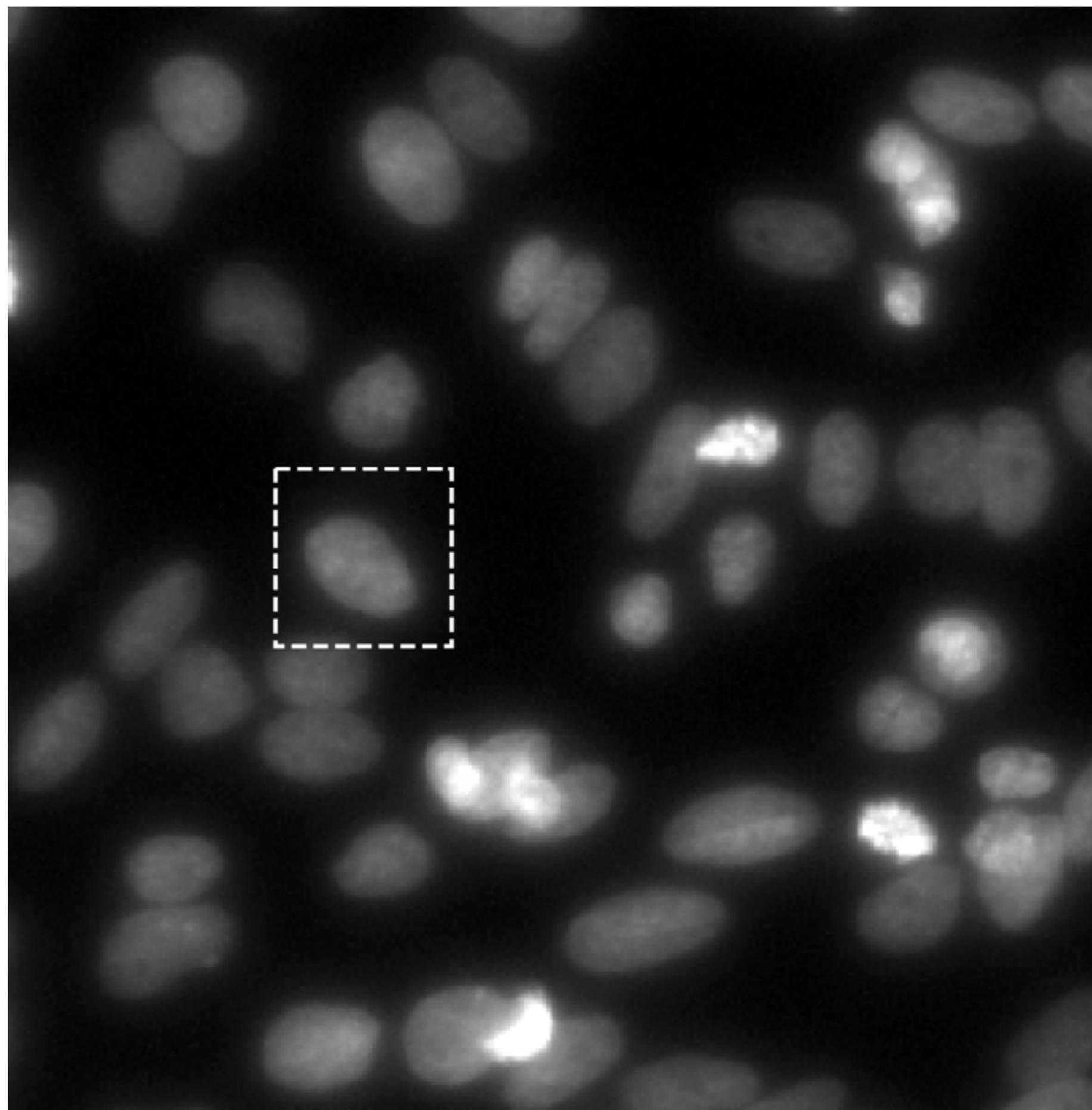


Our Problem: Nuclei Segmentation

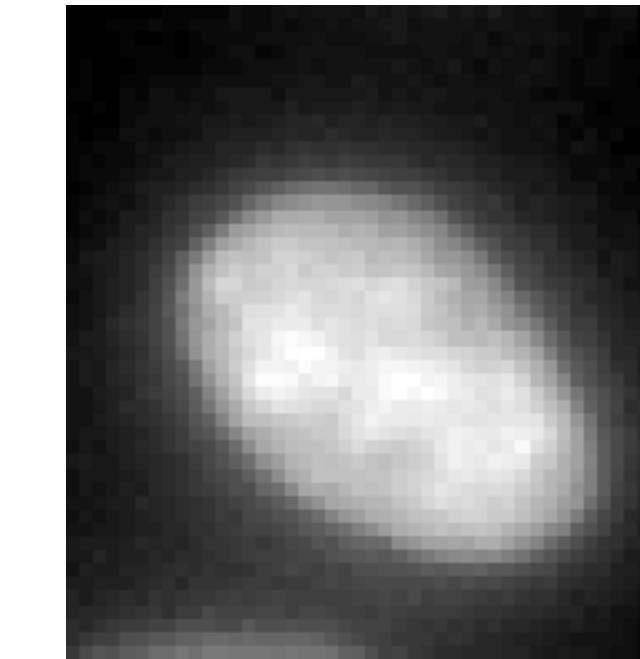


Challenges: many crowded objects, noisy images

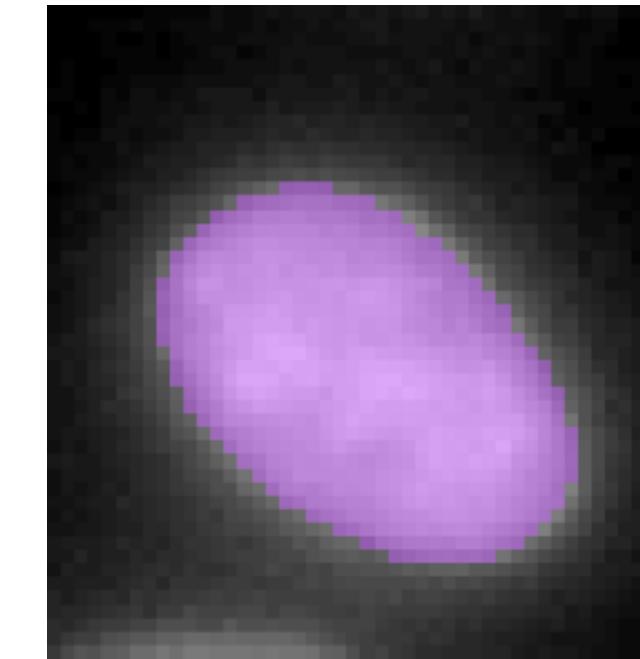
Common (DL) approaches for nuclei segmentation



Image

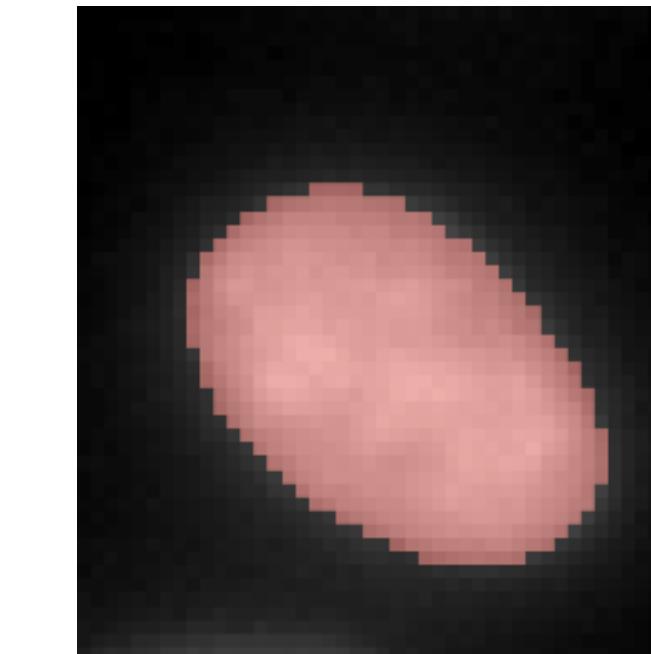


GT



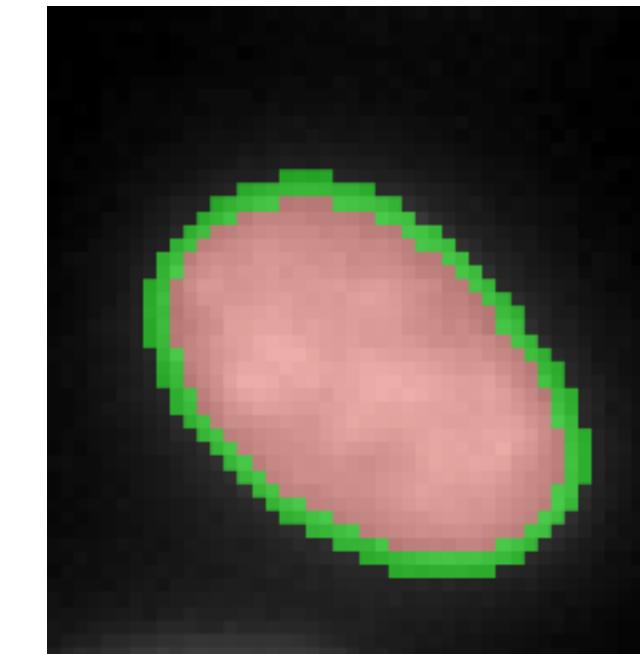
Bottom-Up

First segment, then localize



2 Class U-Net

Ronneberger et al. (2015)

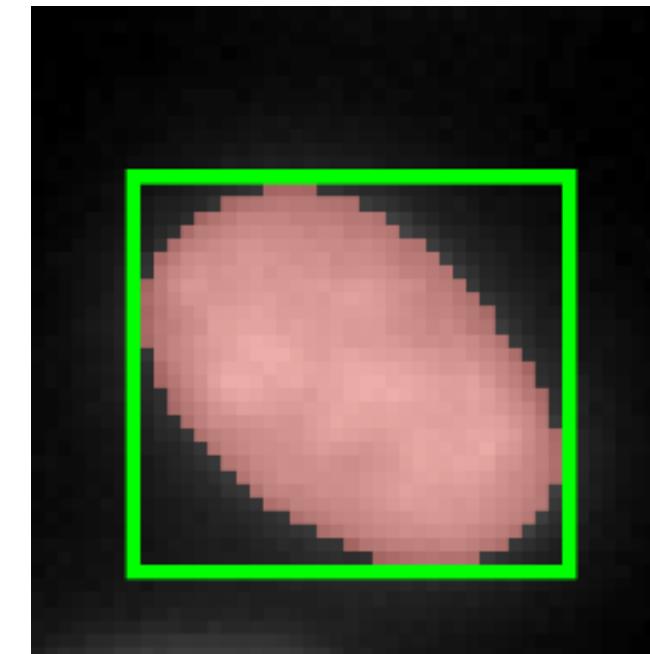


3 Class U-Net

Caicedo et al.(2019)

Top-Down

First localize, then segment

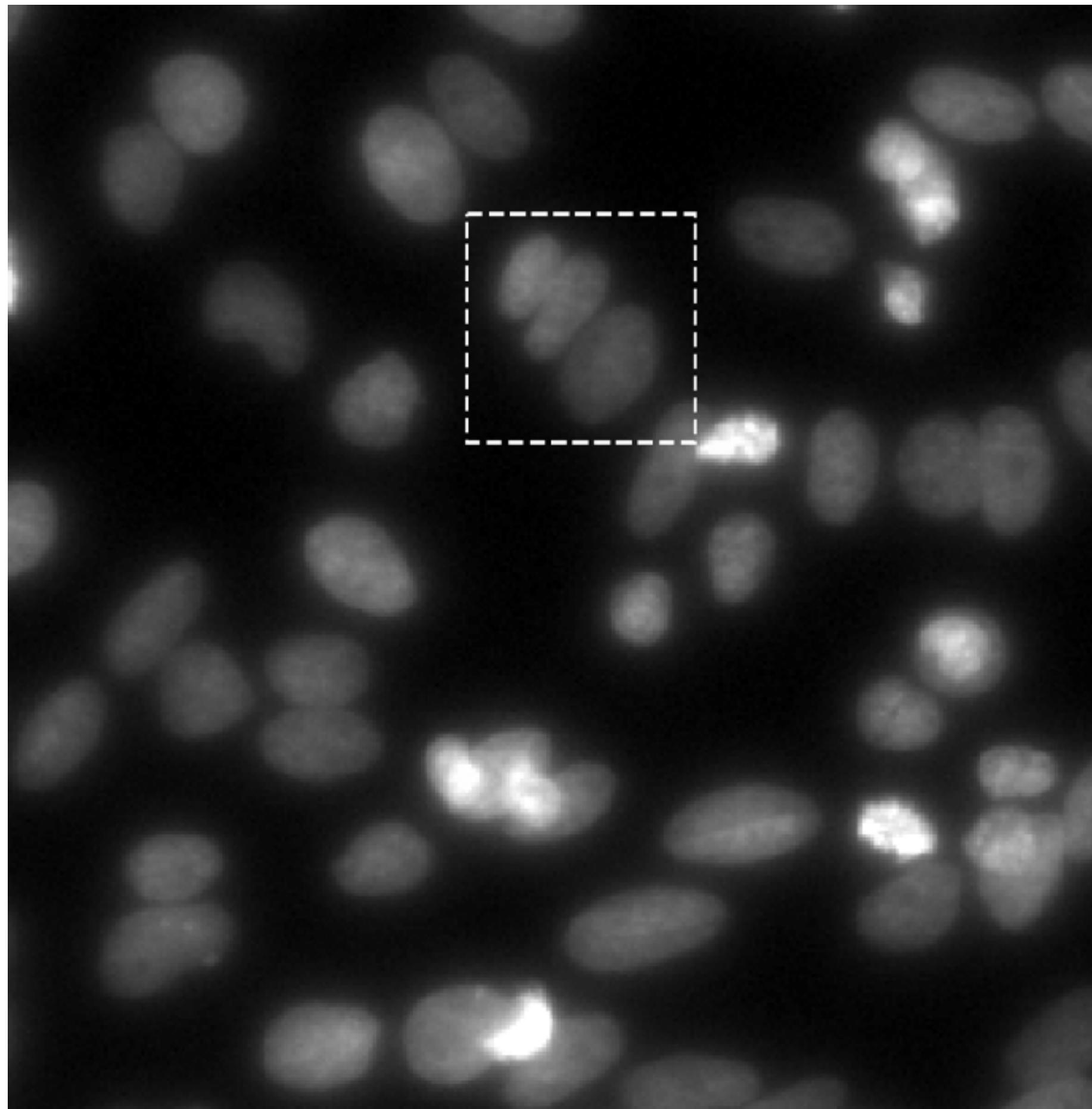


Mask-RCNN

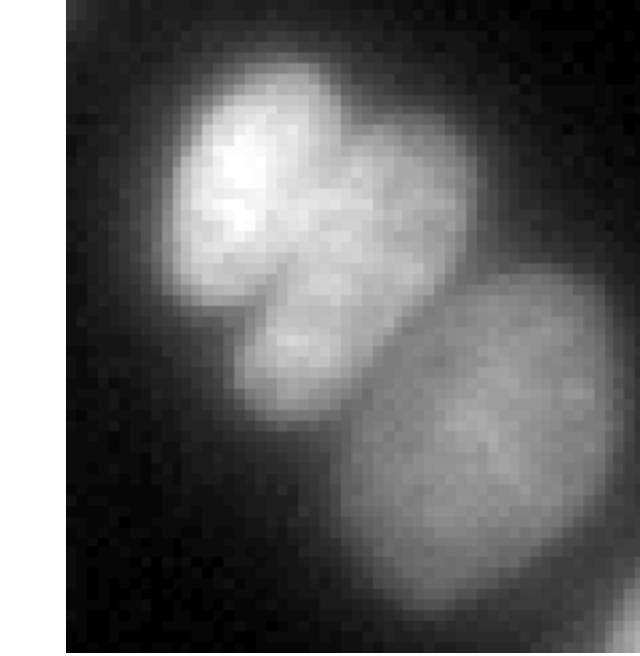
He et al (2017)

- Affinity based methods Wolf, Pape et al. (2018), Hirsch et al. (2020)
- Embedding based methods Neven et al. (2019), Stringer et al. (2020)

Problems for crowded objects



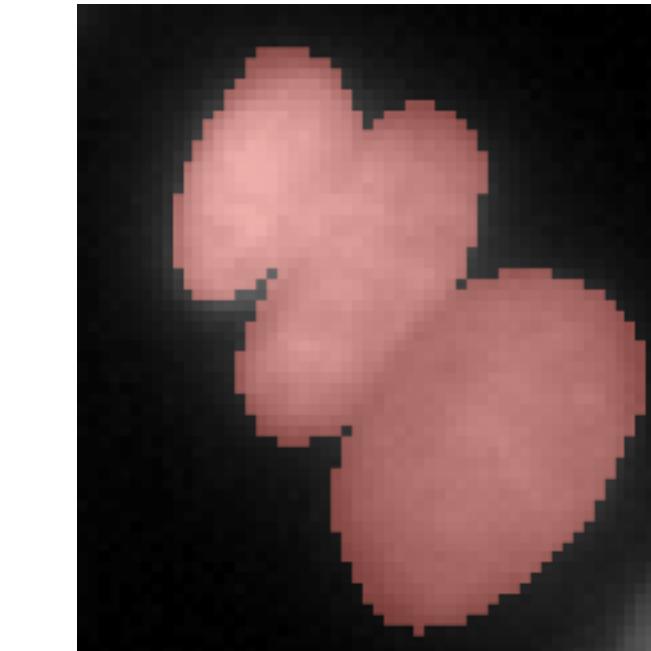
Image



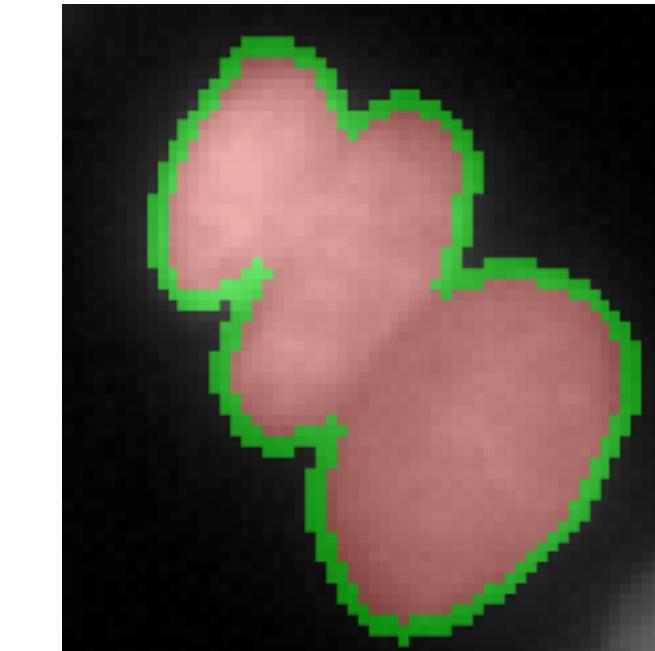
GT

Bottom-Up

First segment, then localize



2 Class U-Net

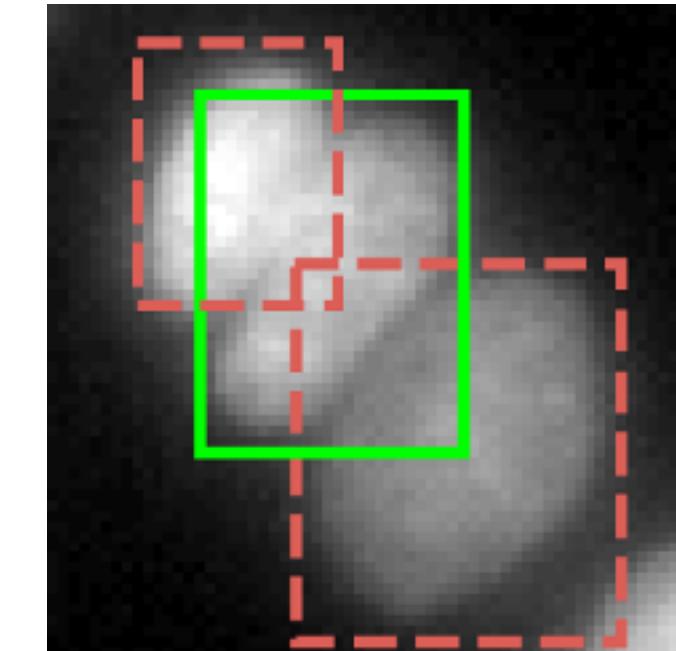


3 Class U-Net

Mislocalization
Fused segmentation maps

Top-Down

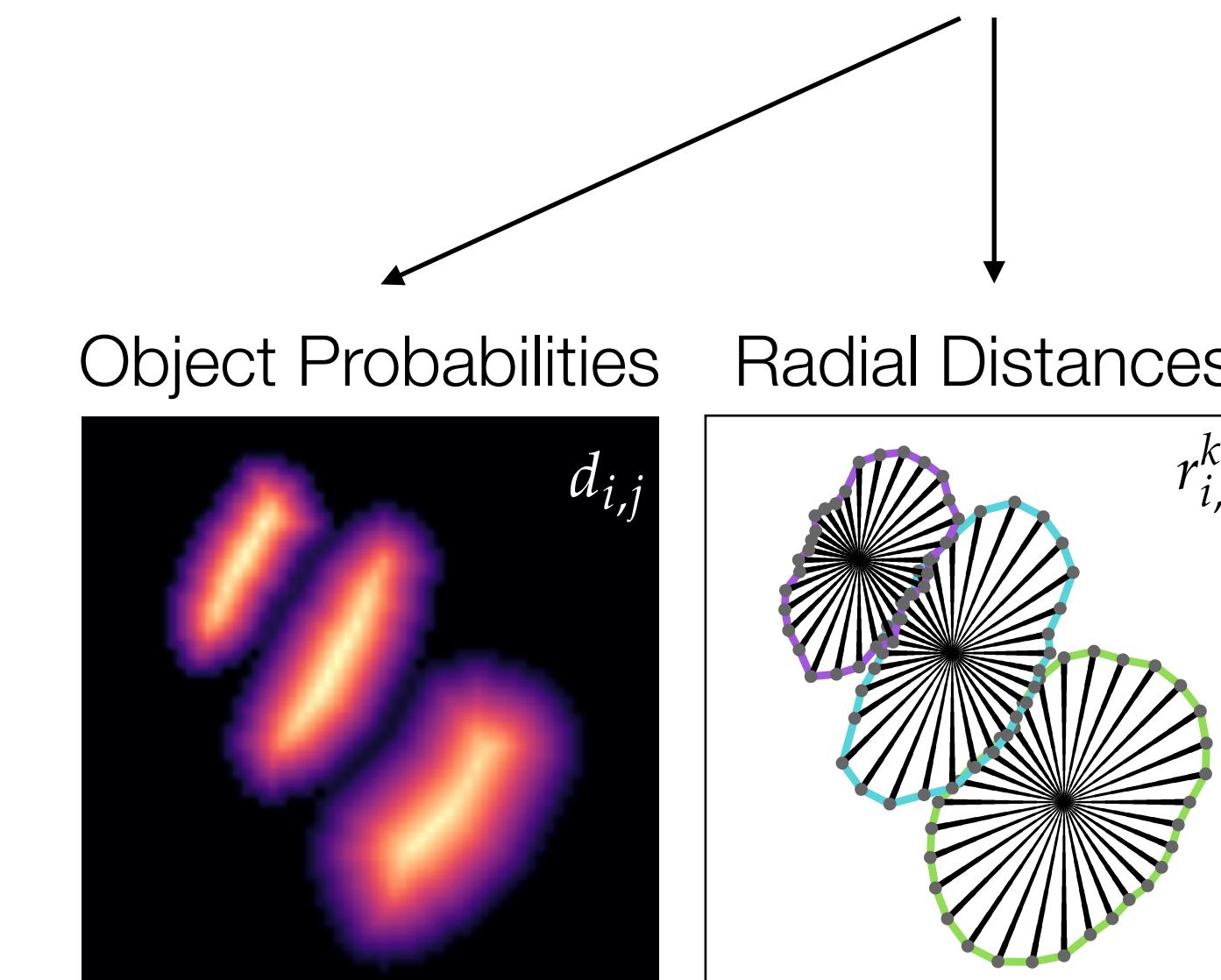
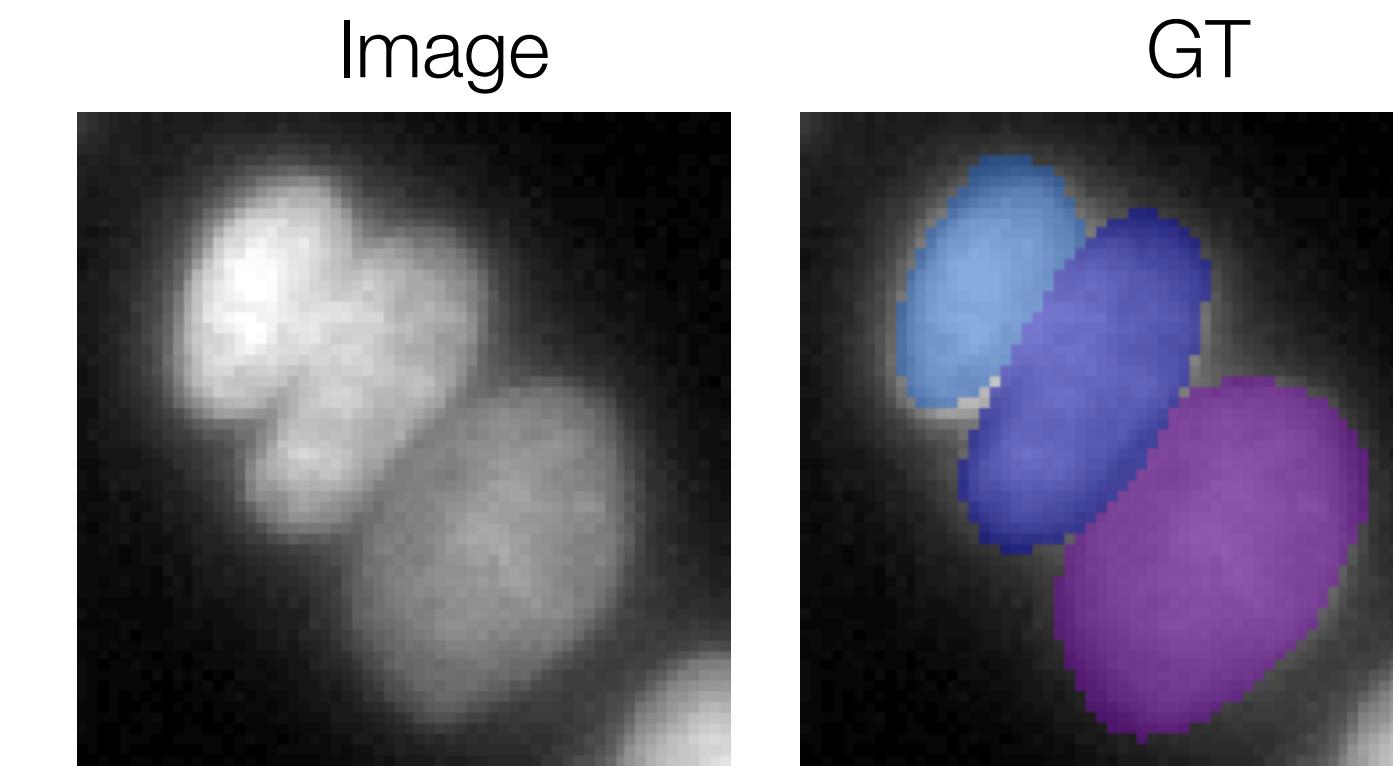
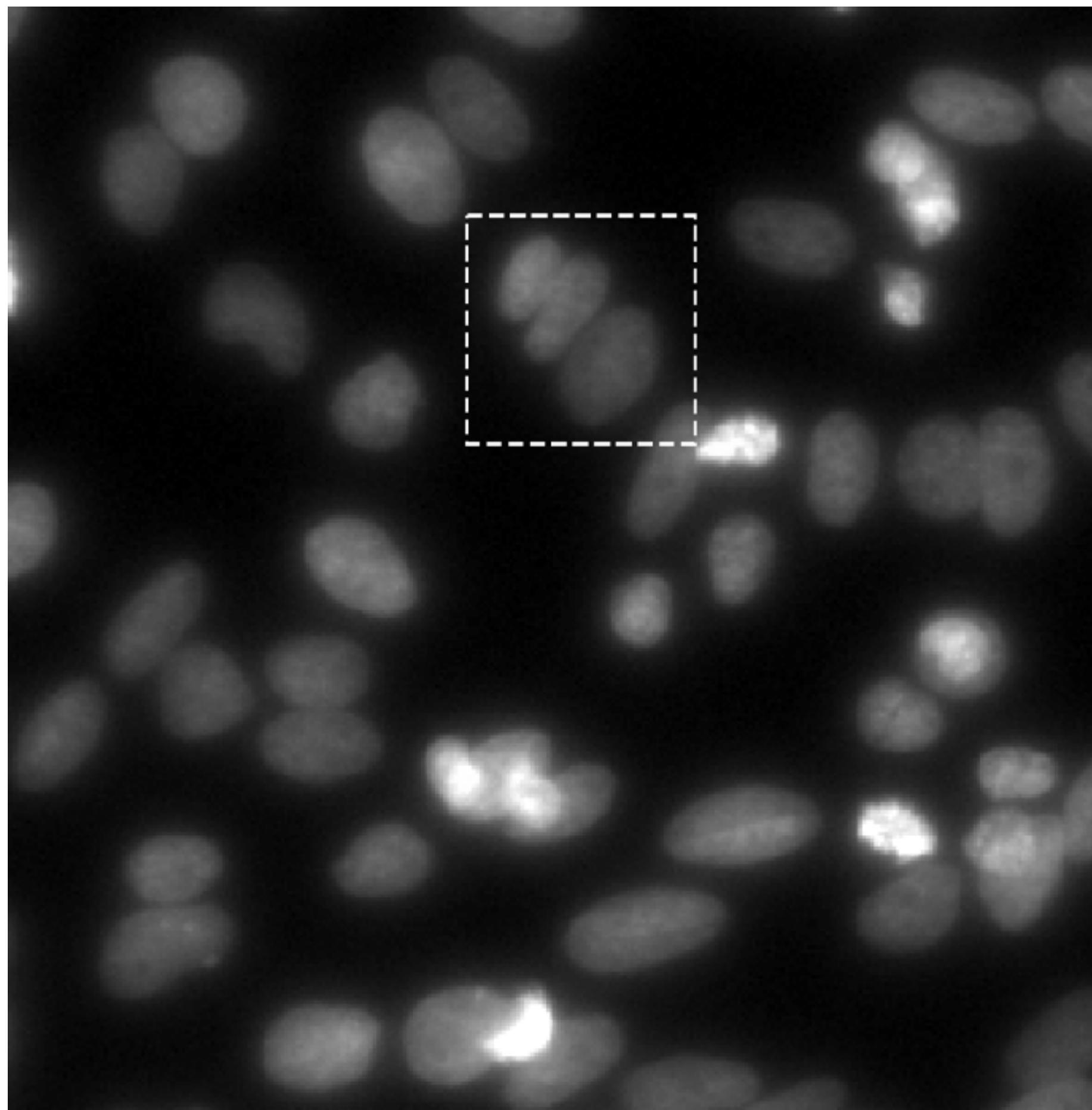
First localize, then segment



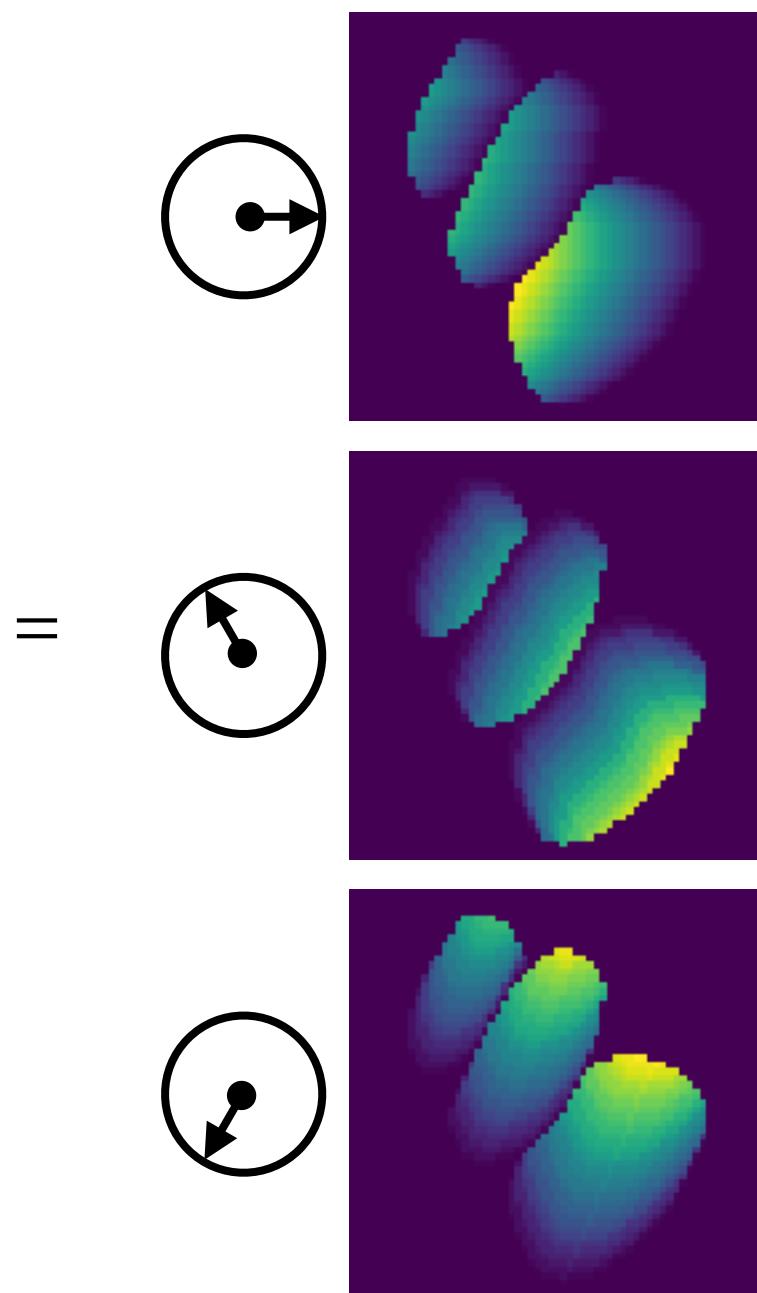
Mask-RCNN

Mislocalization
Bounding box overlap > threshold

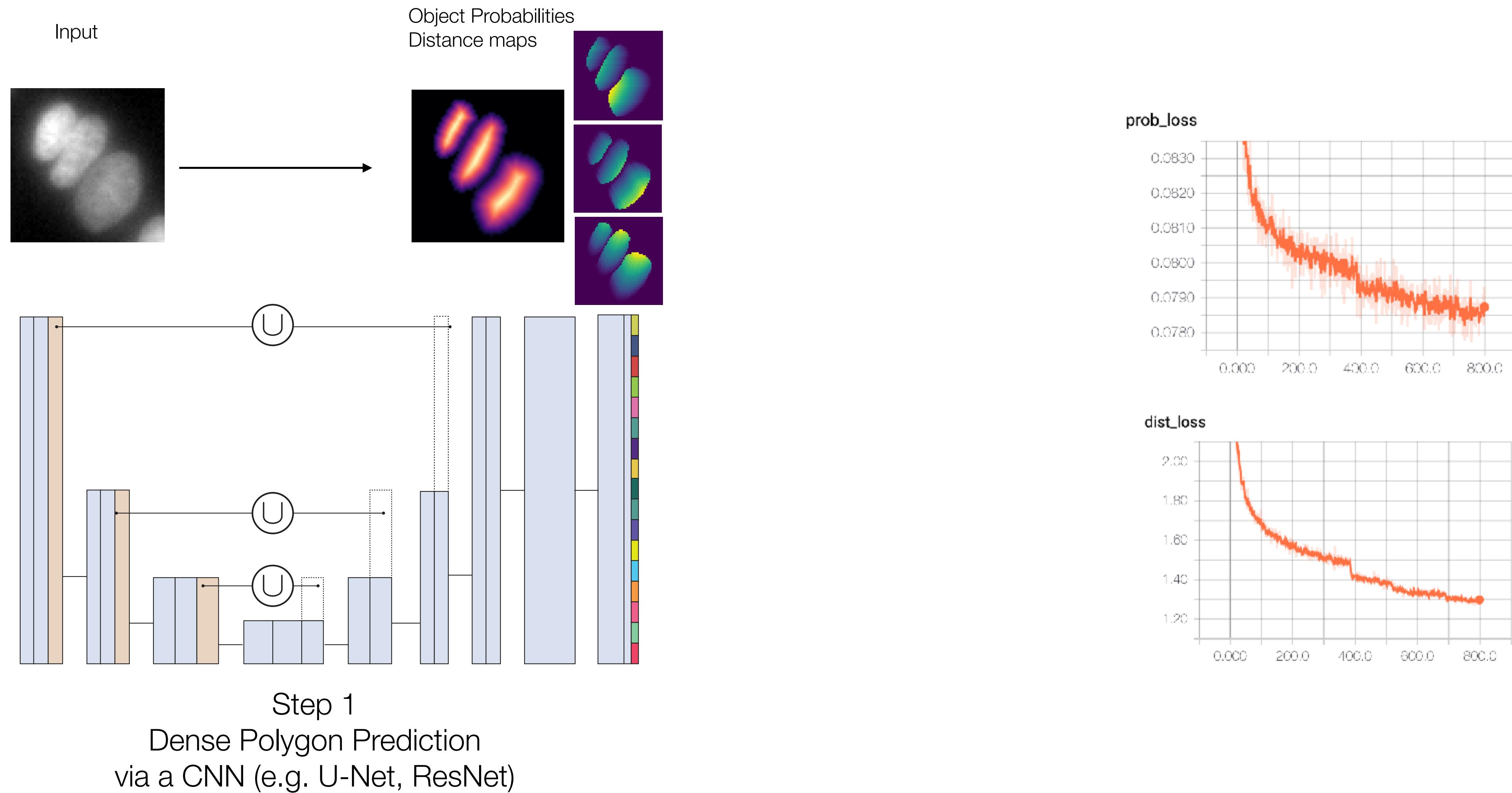
StarDist: Principle



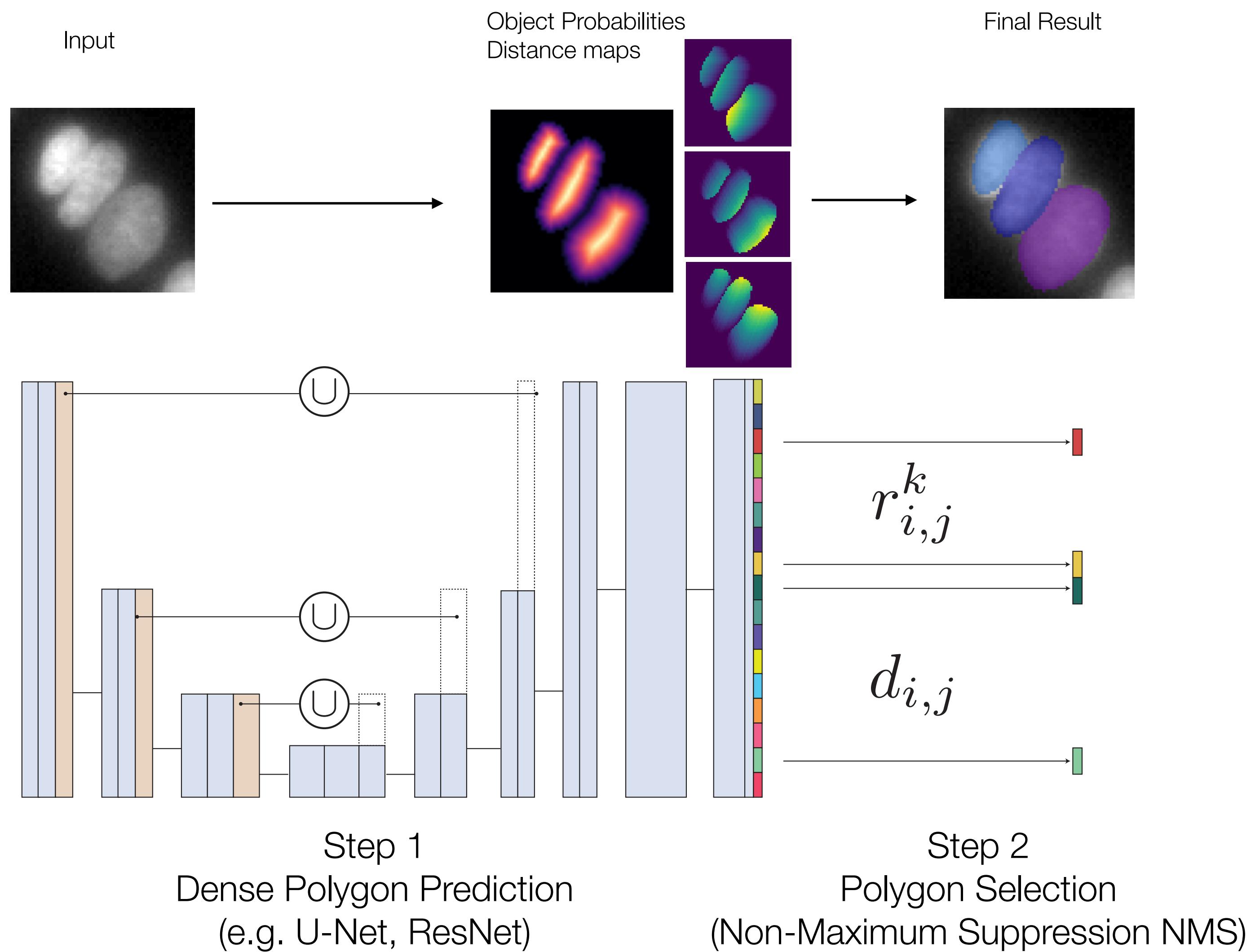
Distance maps (~32)



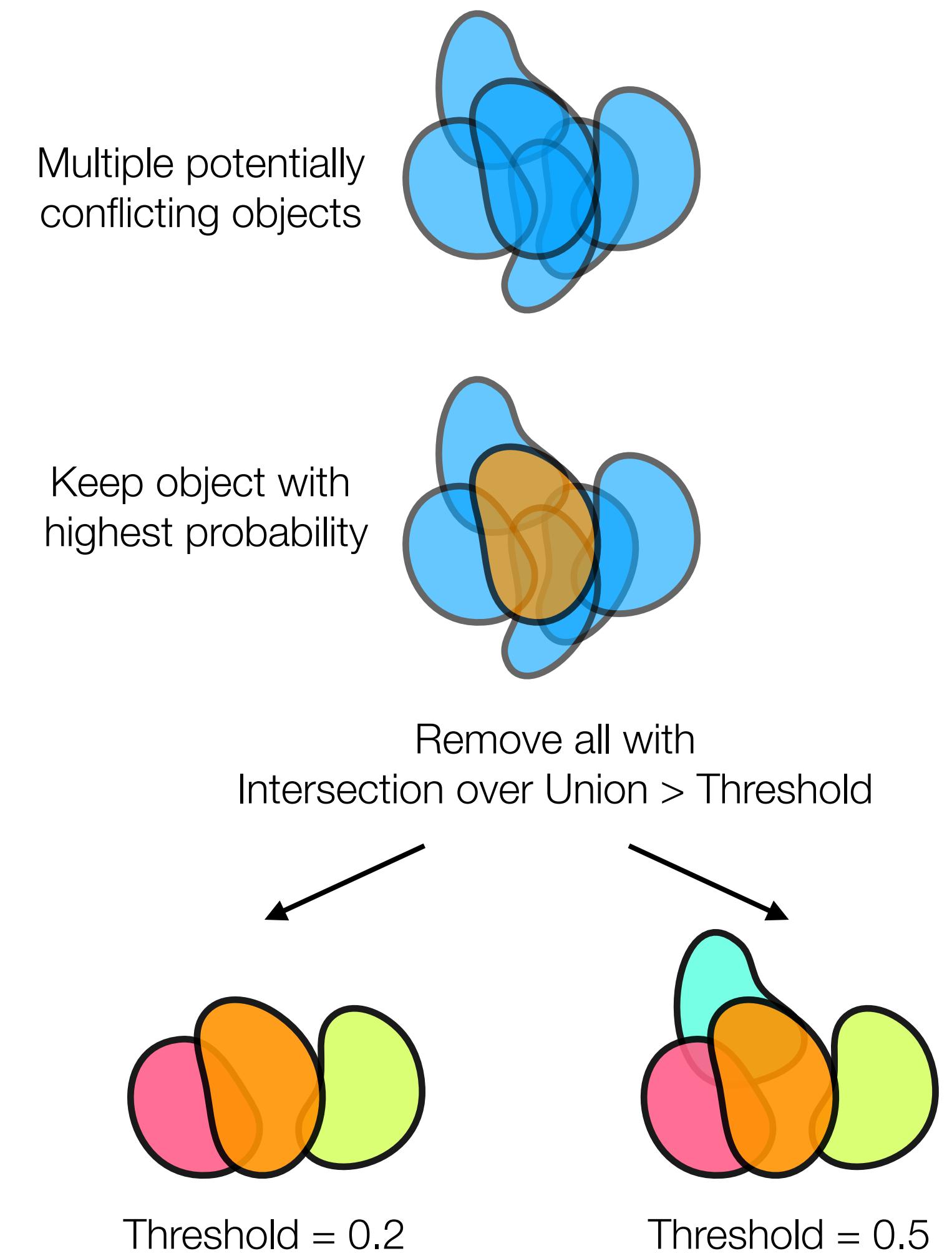
StarDist: Principle



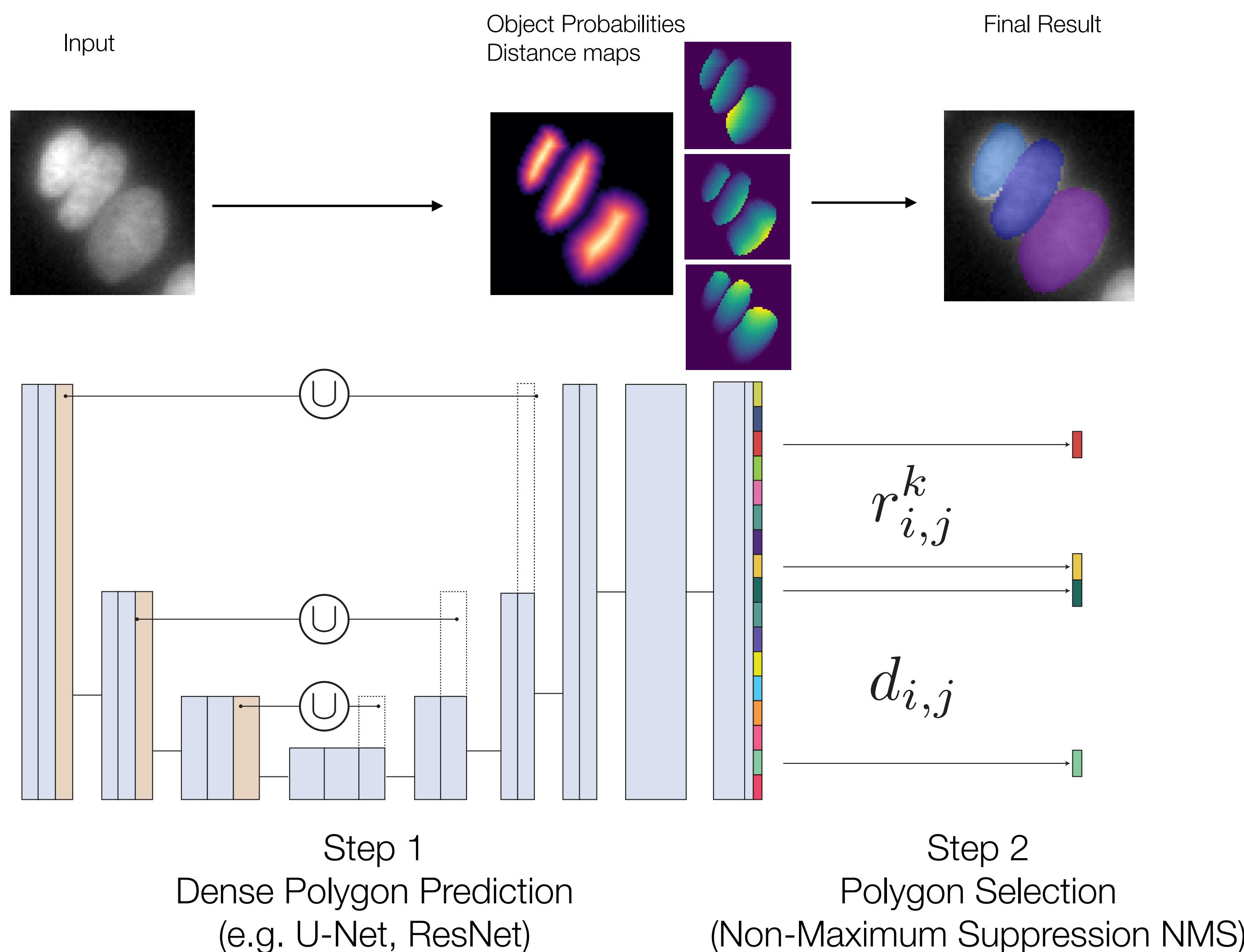
StarDist: Principle



Non-Maximum-Suppression (NMS)

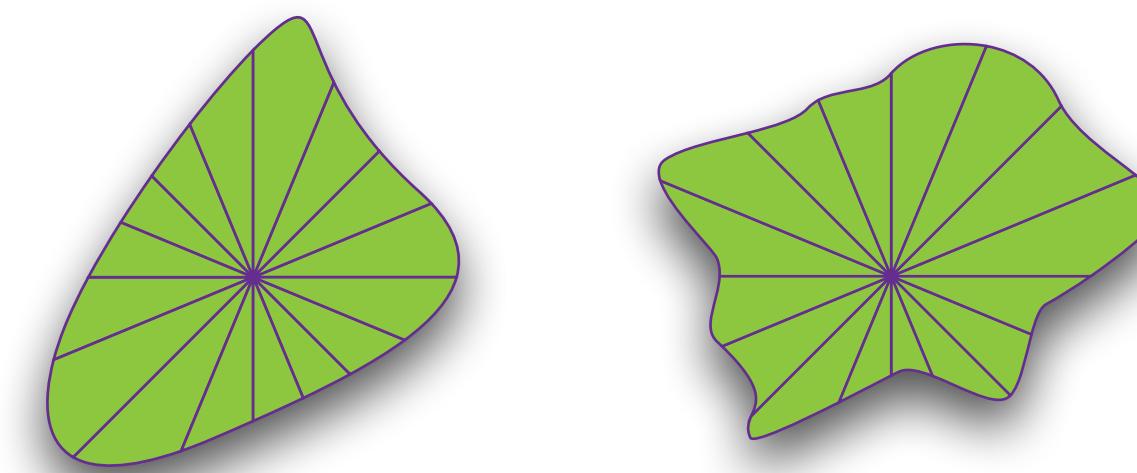


StarDist: Principle

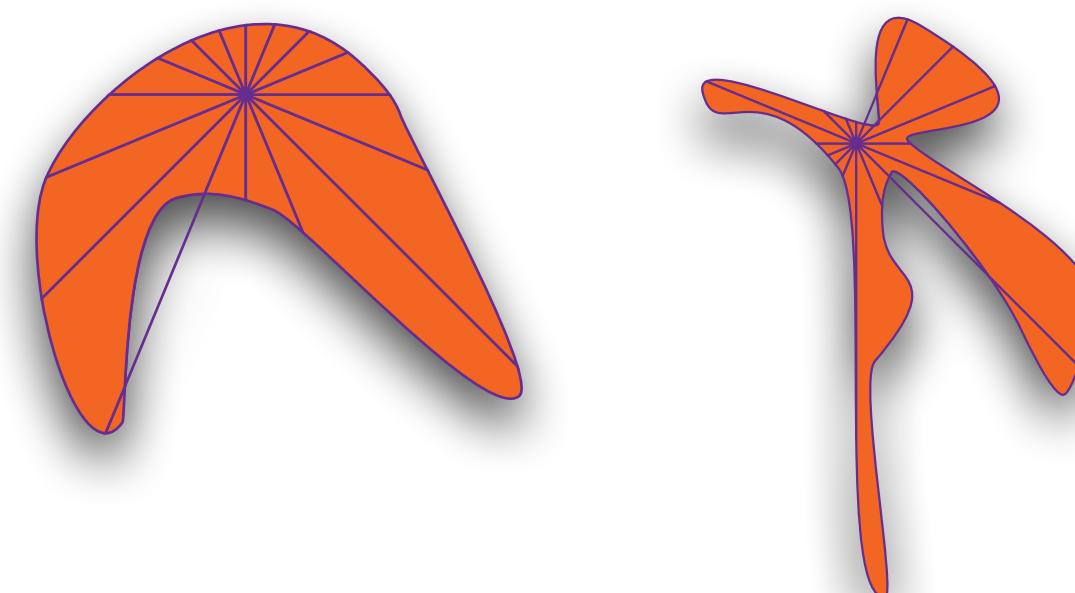


Important:
Assumes objects are star-convex

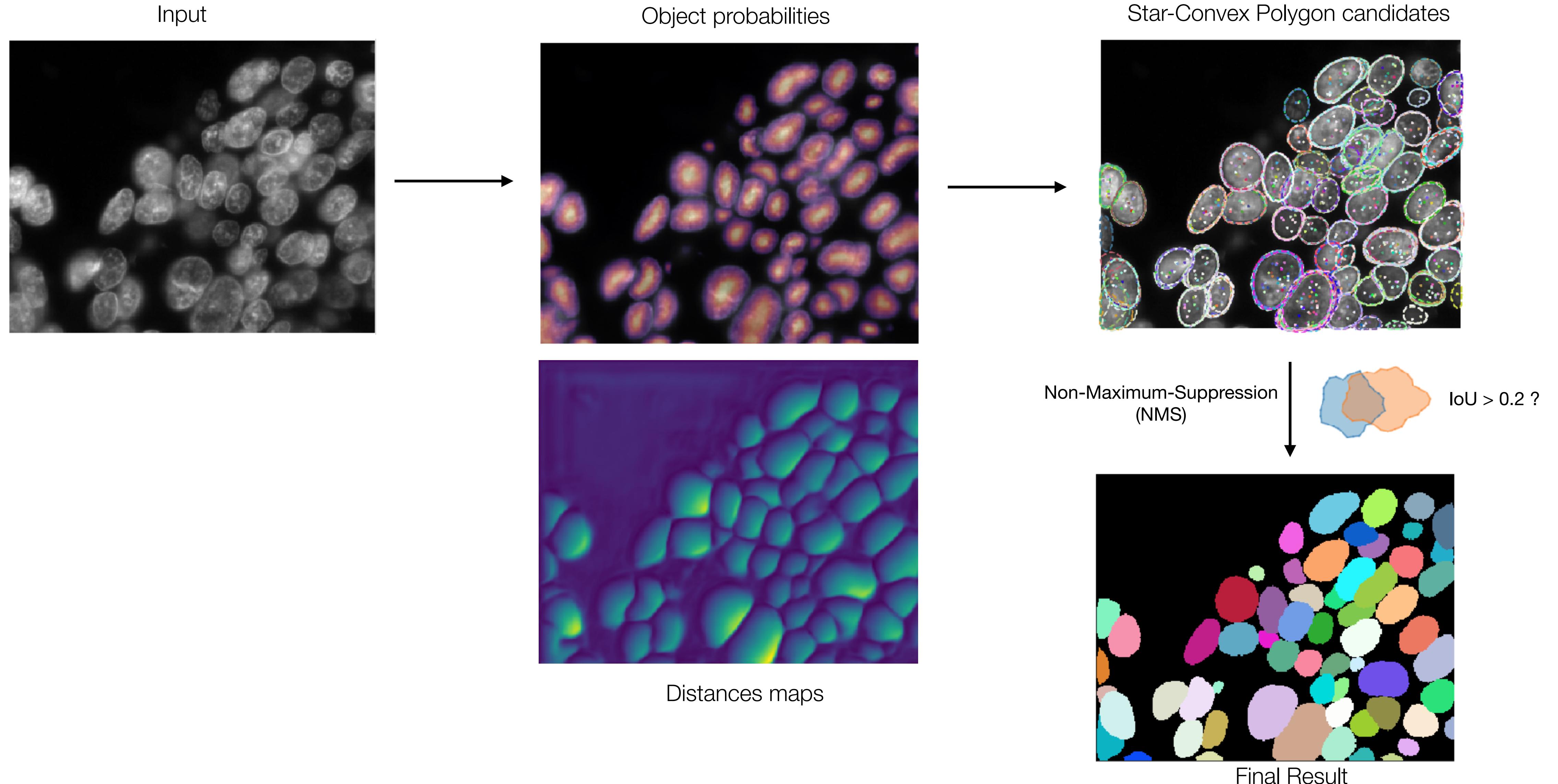
Star-Convex



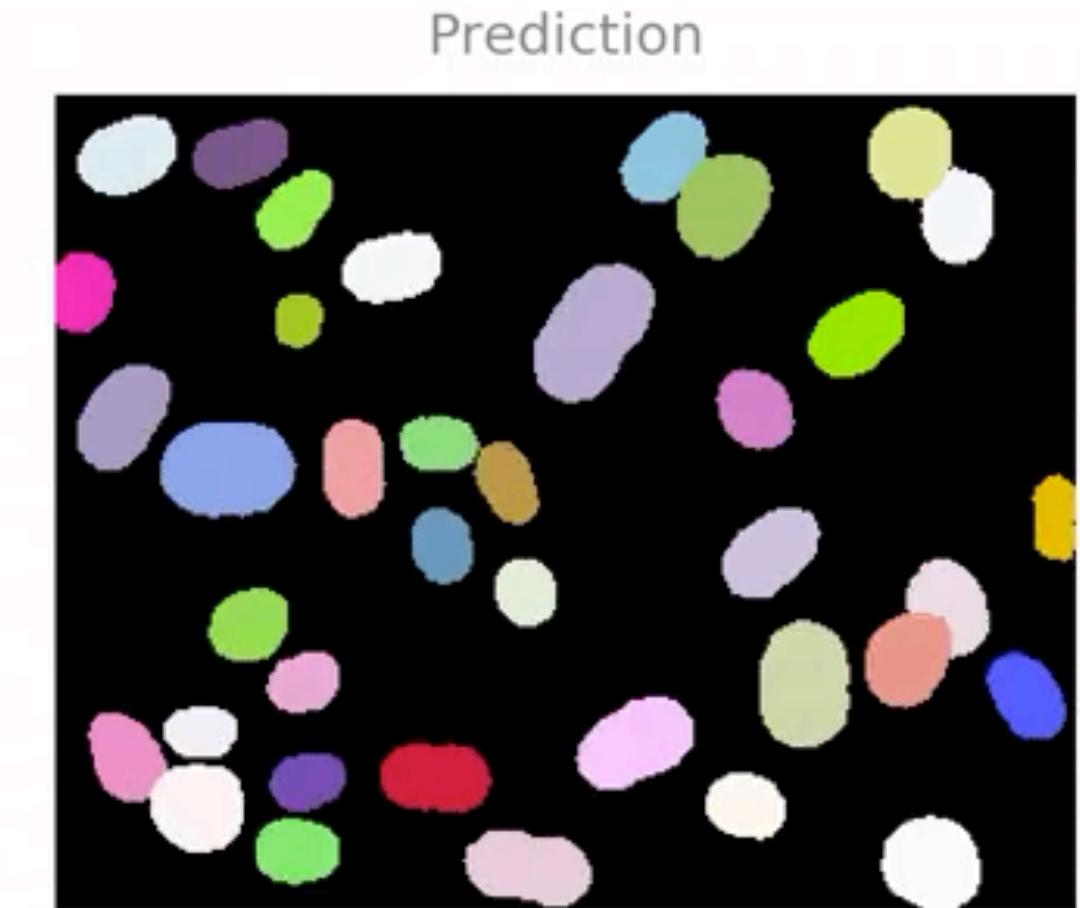
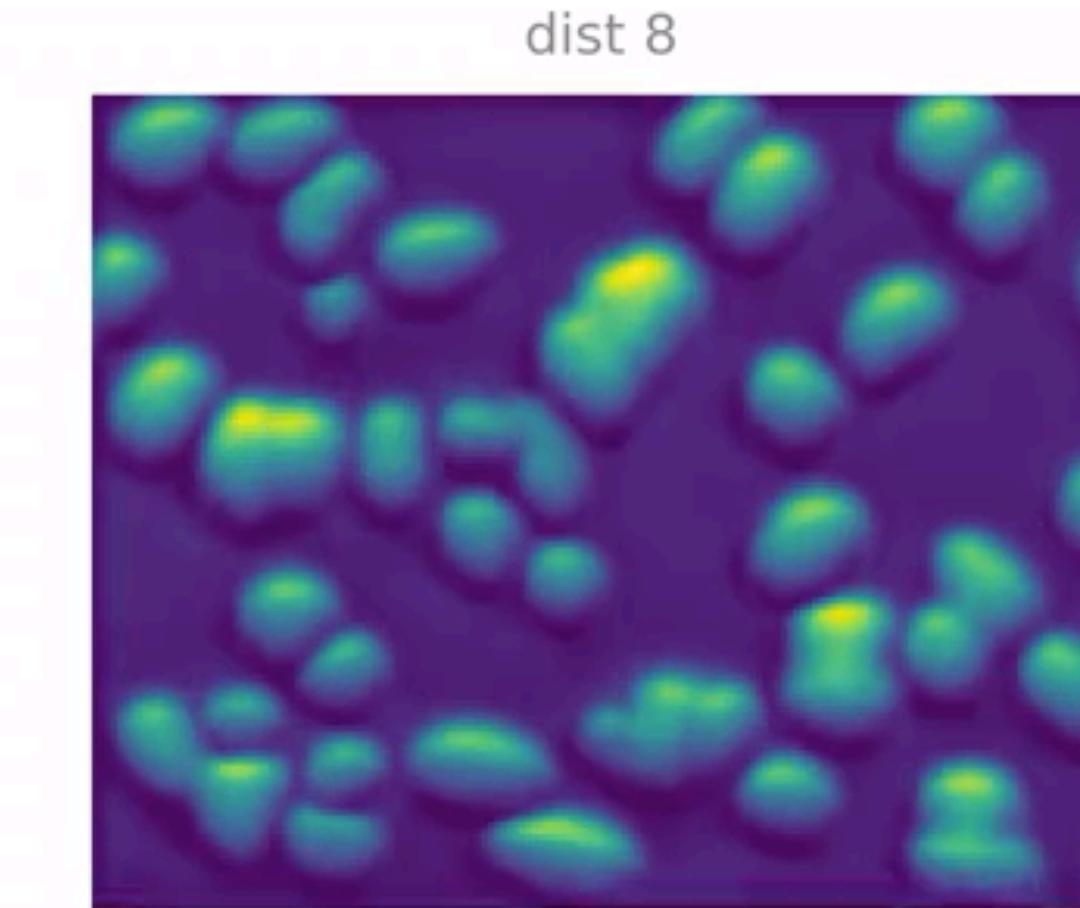
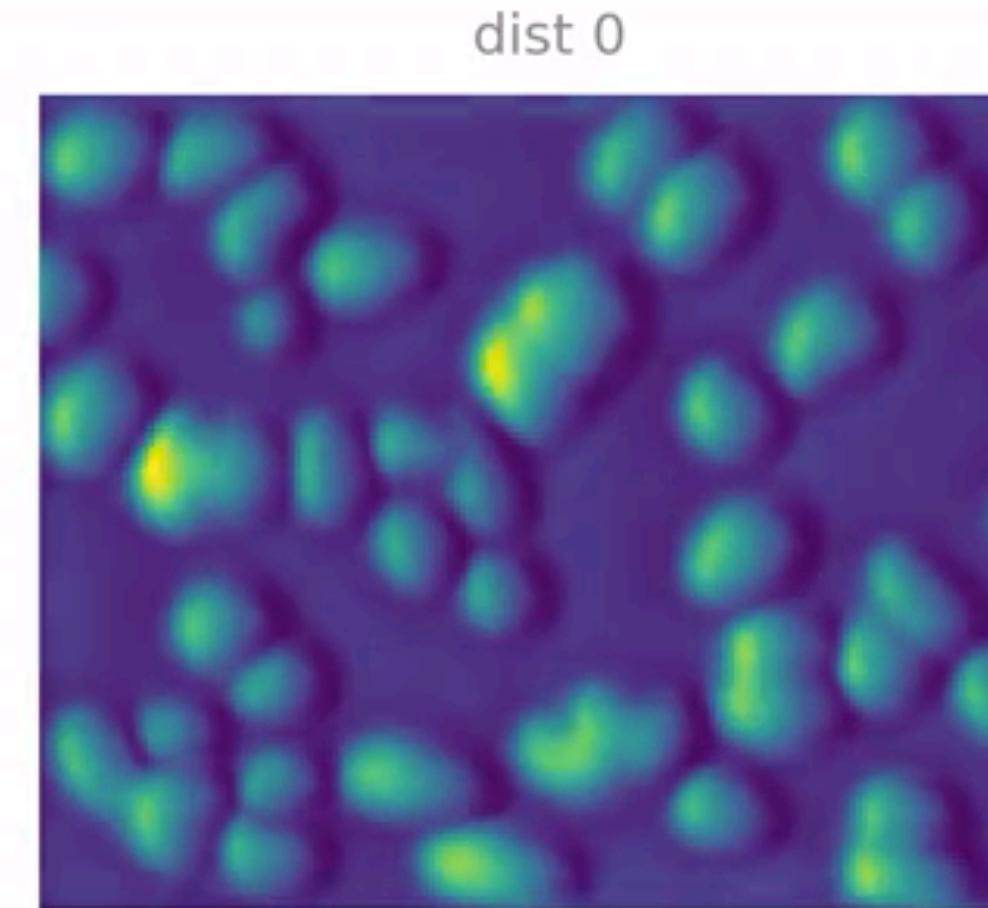
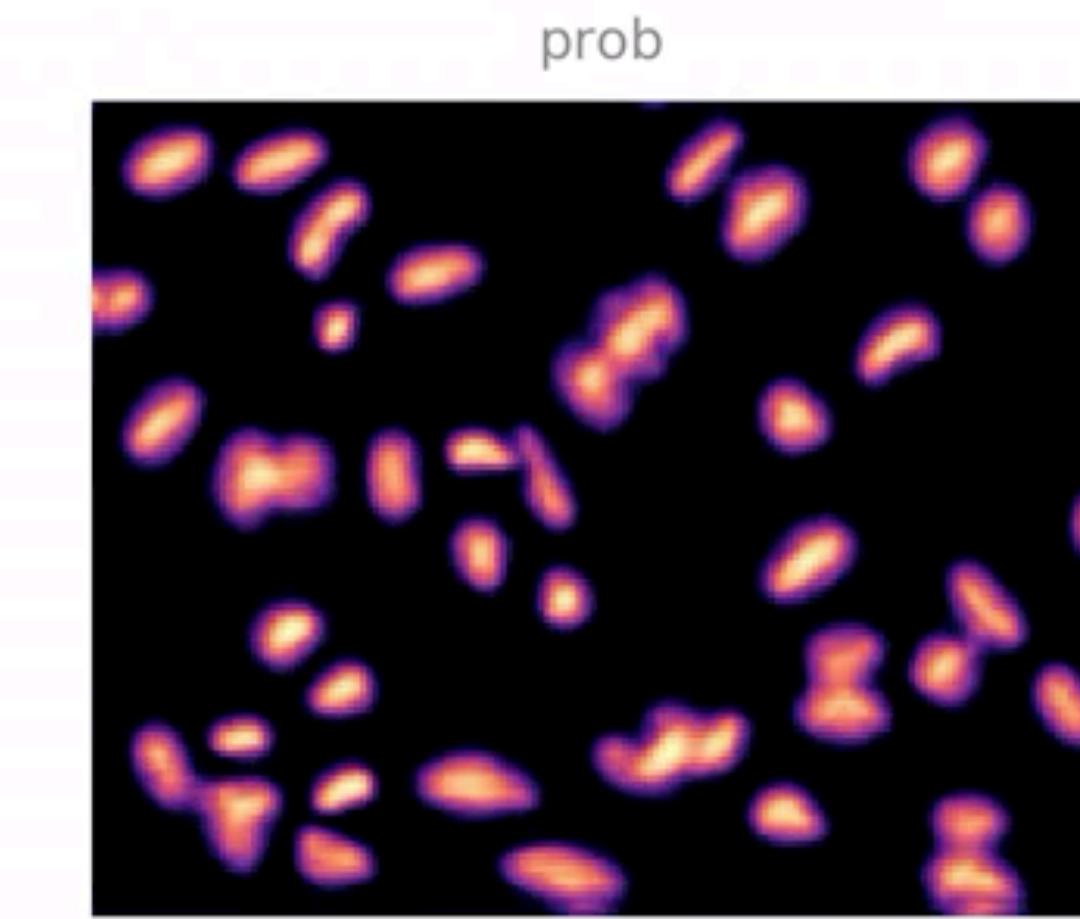
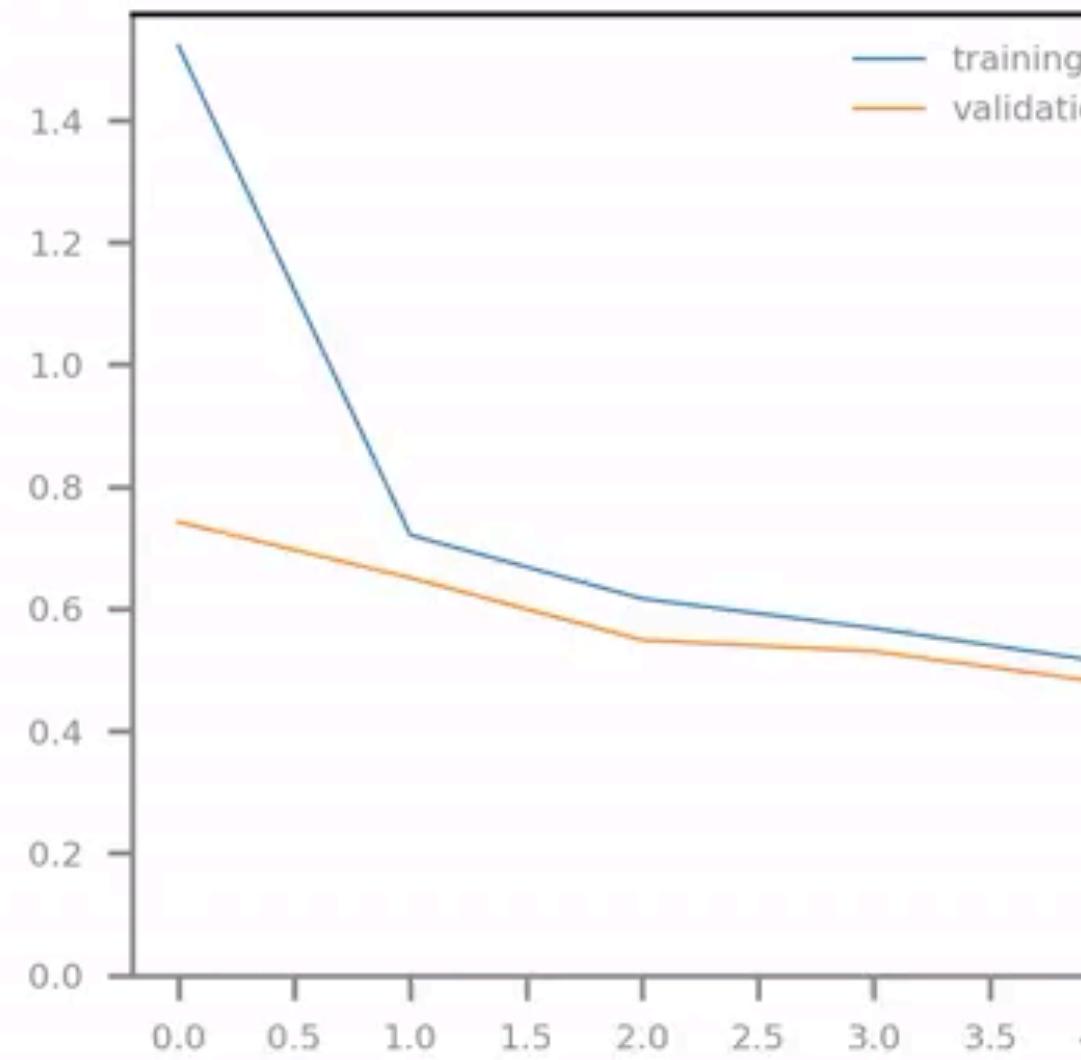
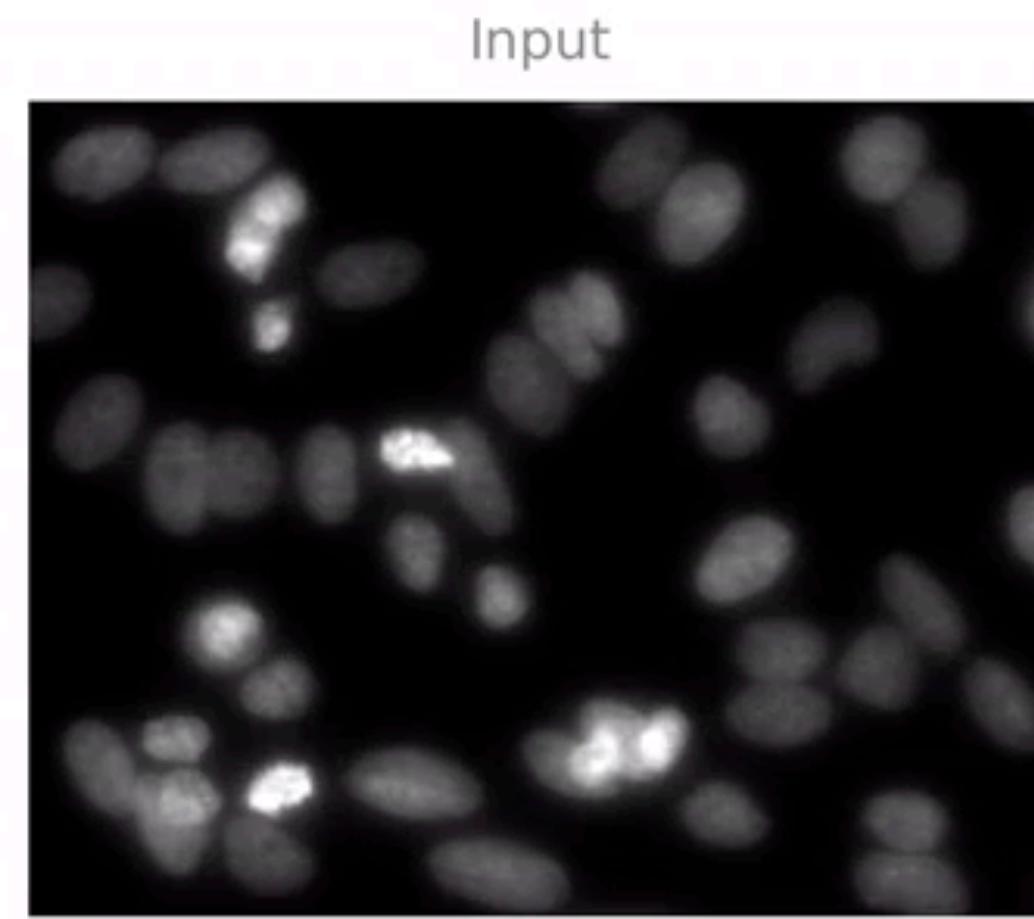
Not Star-Convex



StarDist: Example



Training process

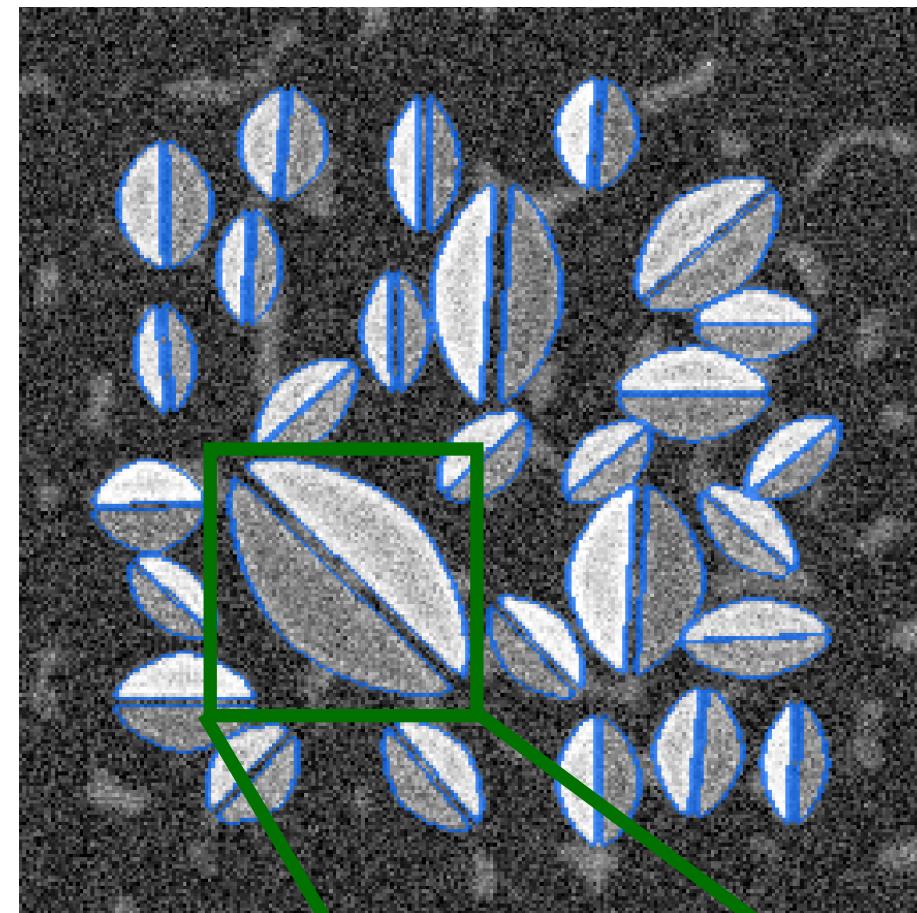


Comparison with common methods

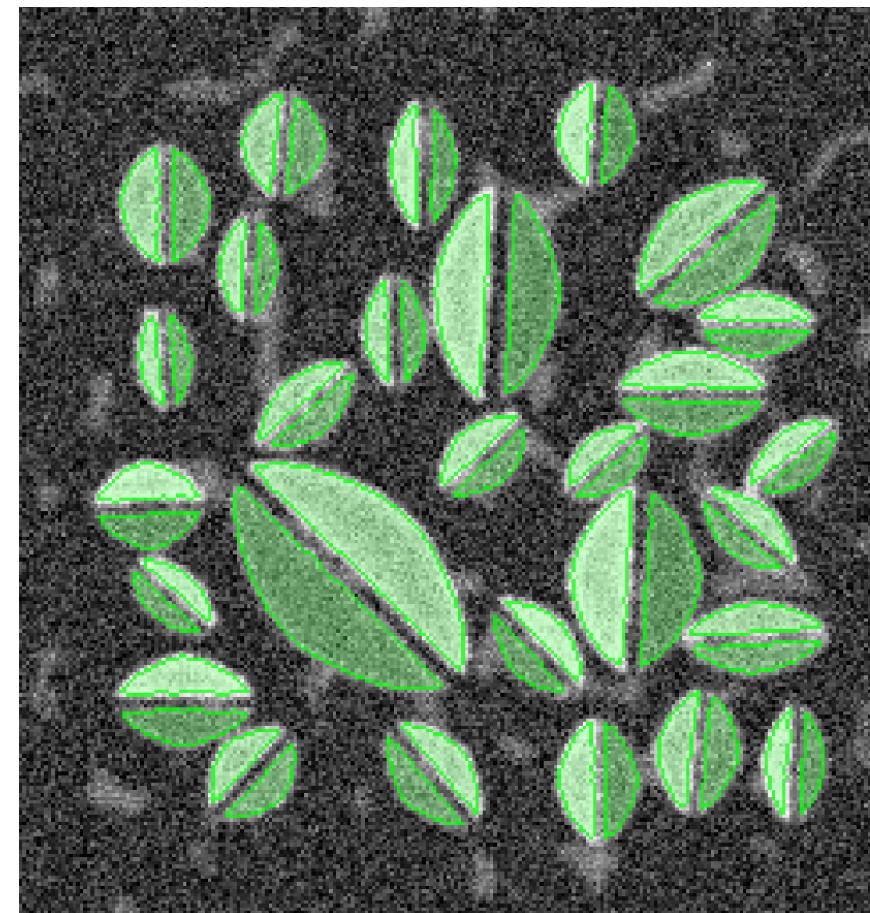
Coffee-Beans (Synthetic)

Predictions

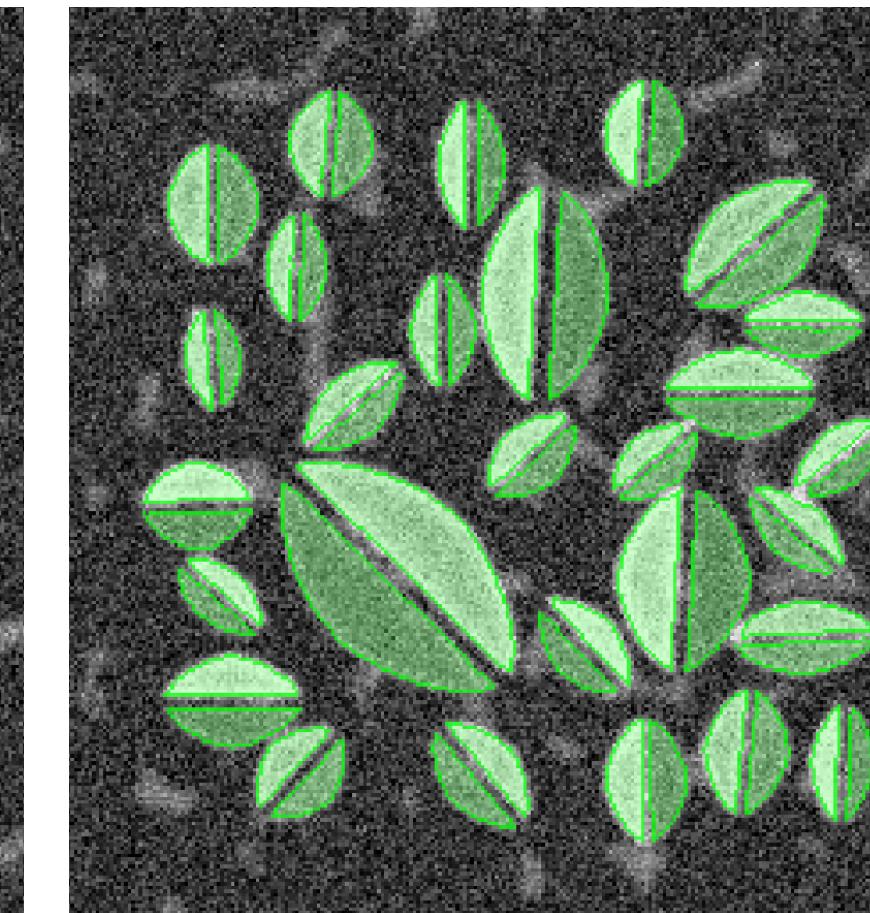
GT



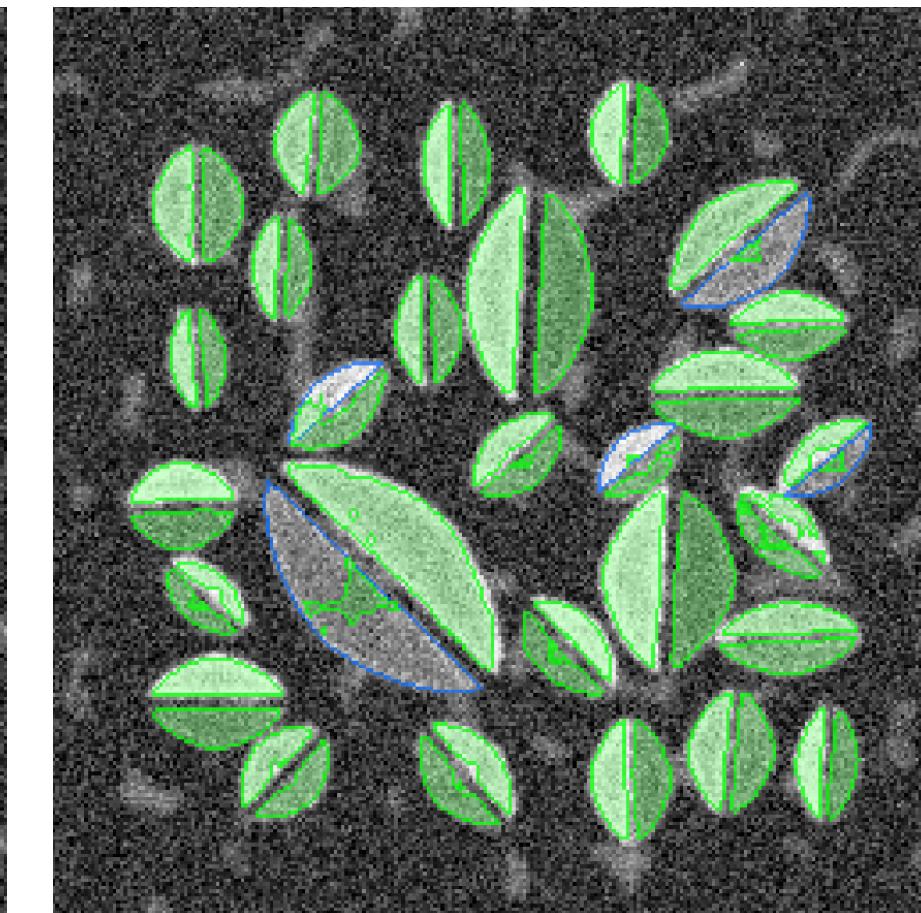
U-Net (2 class)



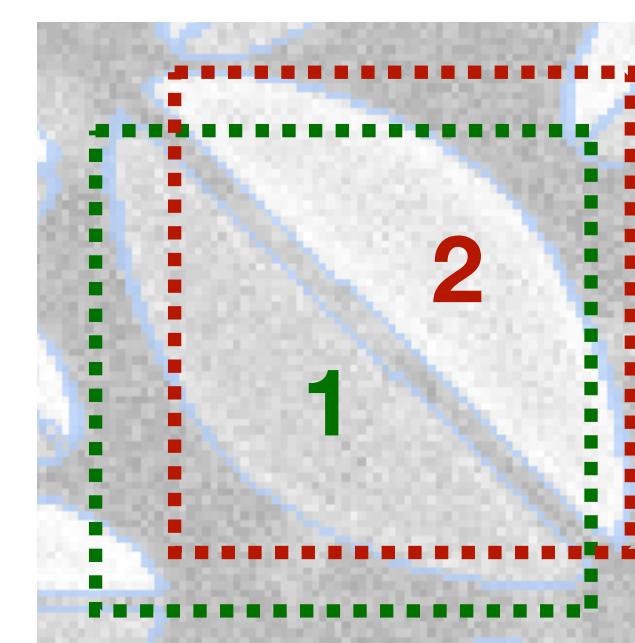
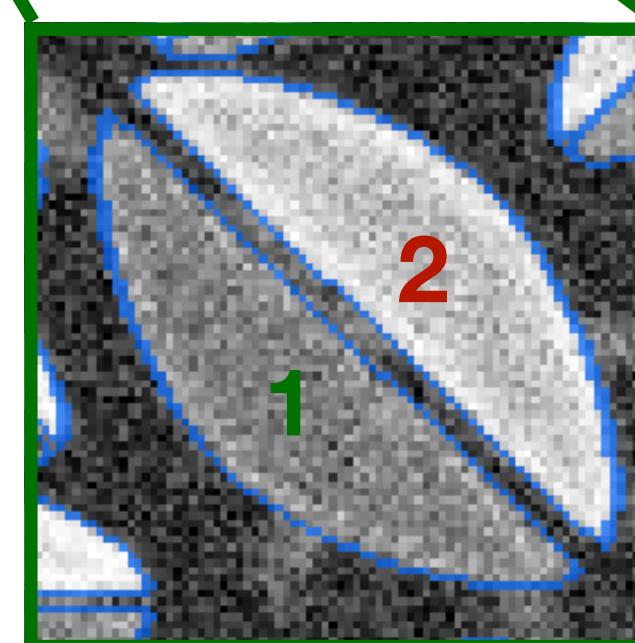
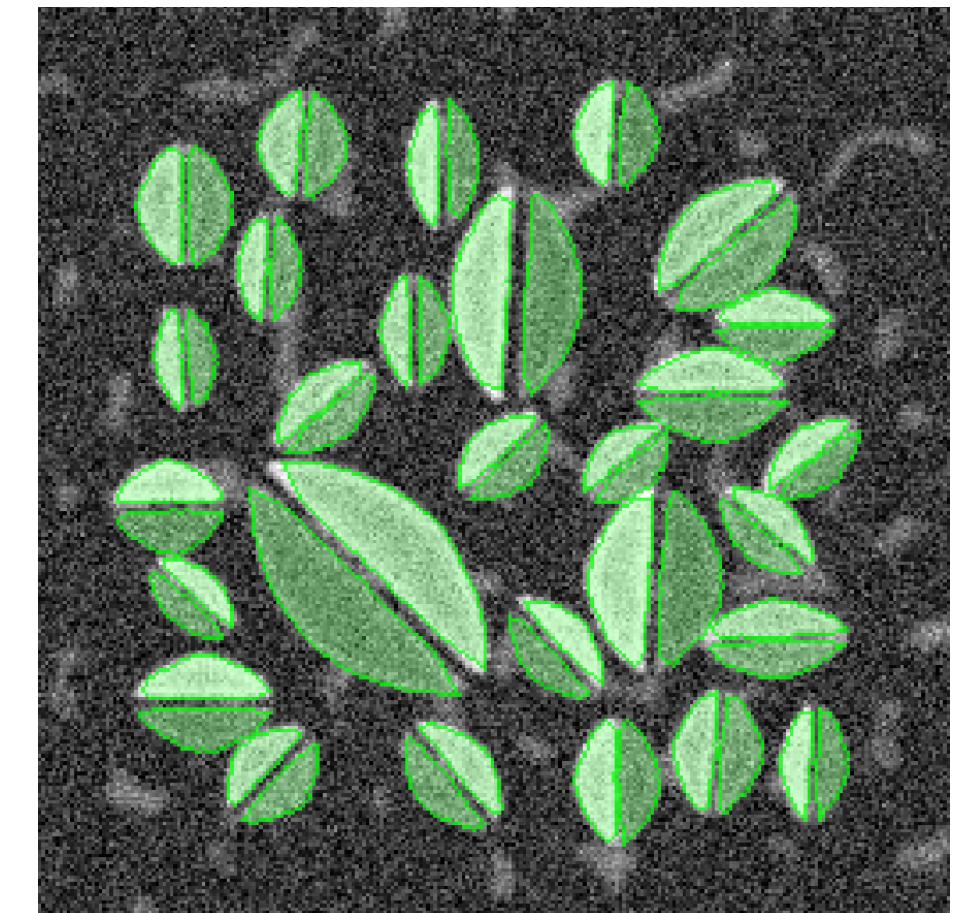
U-Net (3 class)



Mask-RCNN

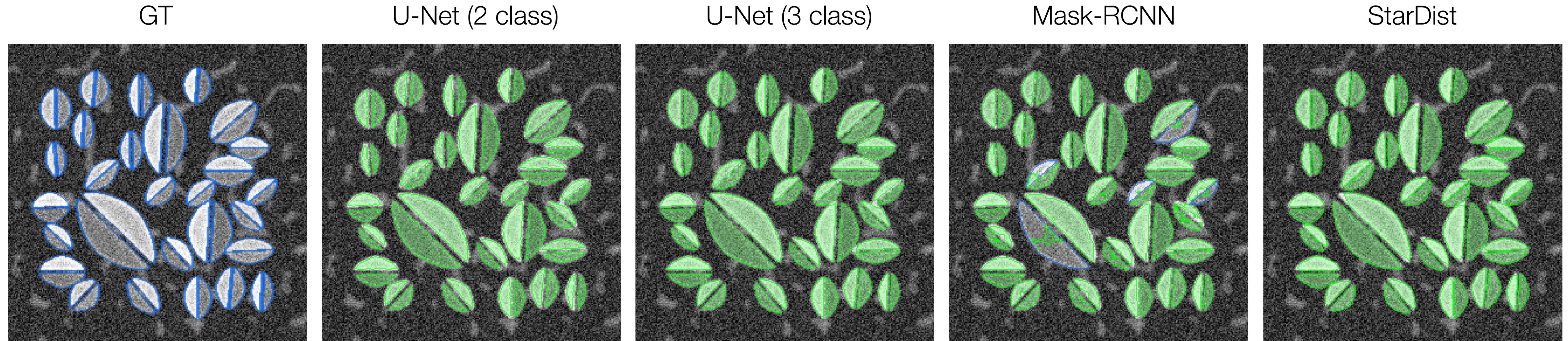


StarDist

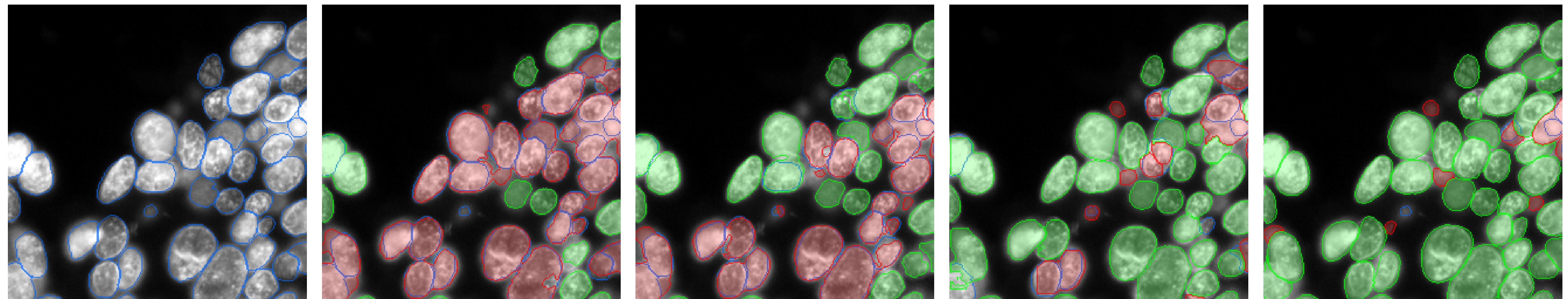


Comparison with common methods

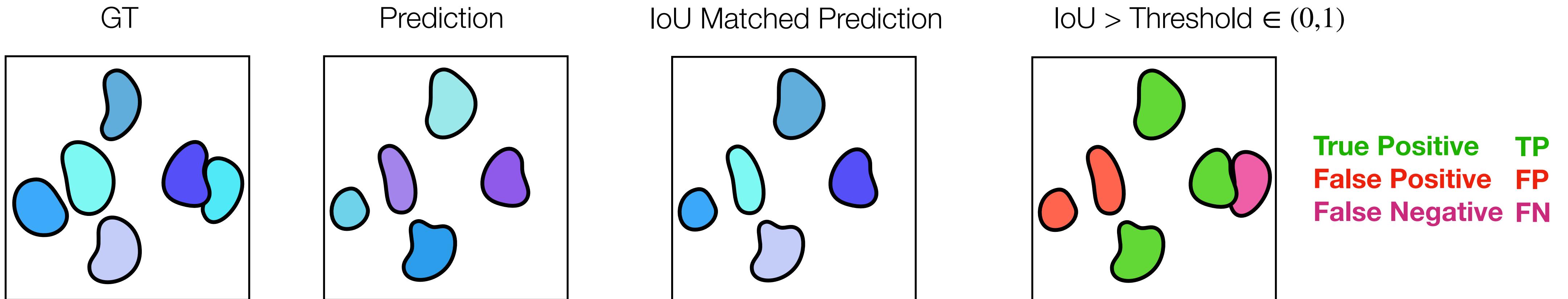
Coffee-Beans (Synthetic)



DSB 2018 (Kaggle Challenge) Caicedo et al (2019)

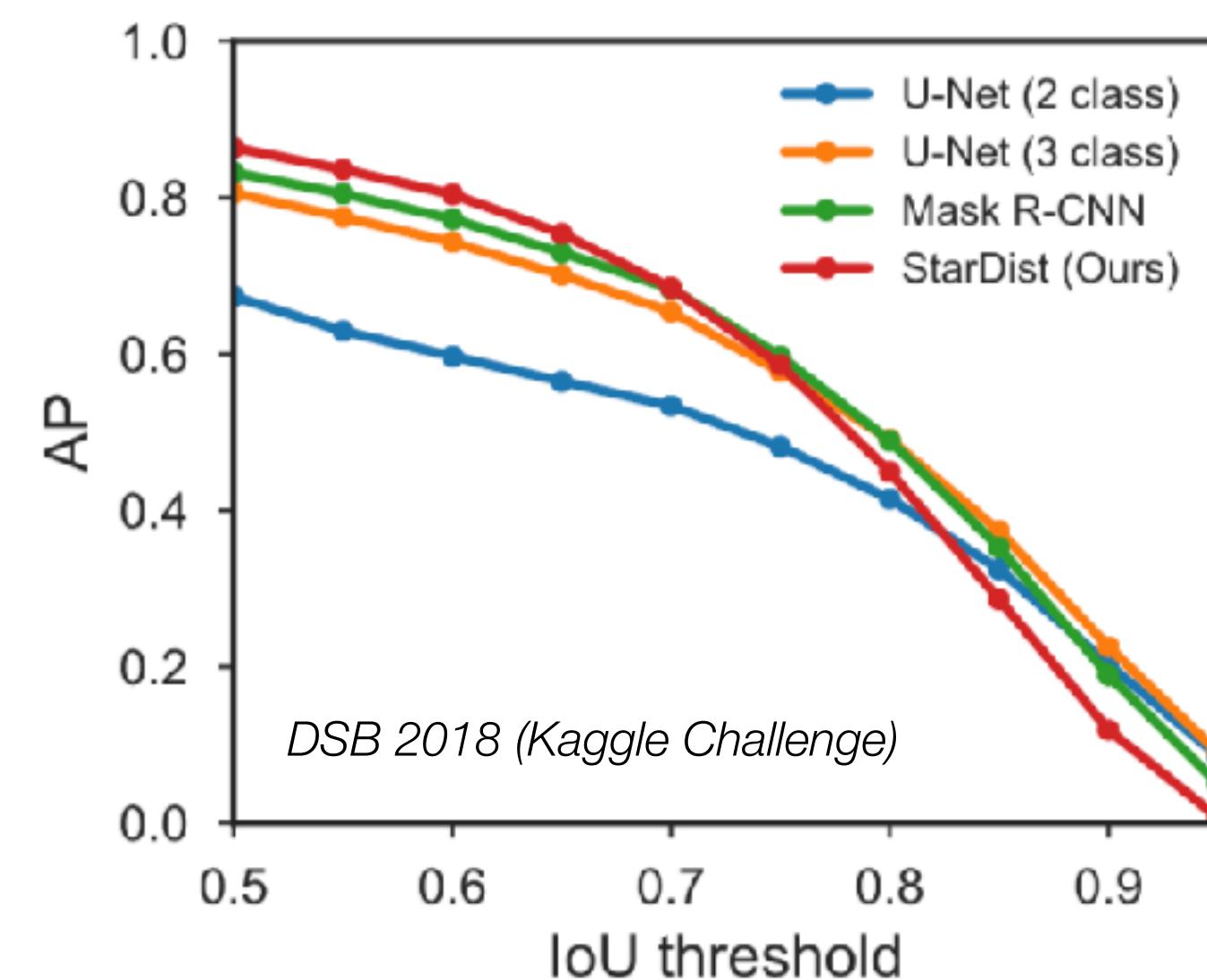


Accuracy Measures



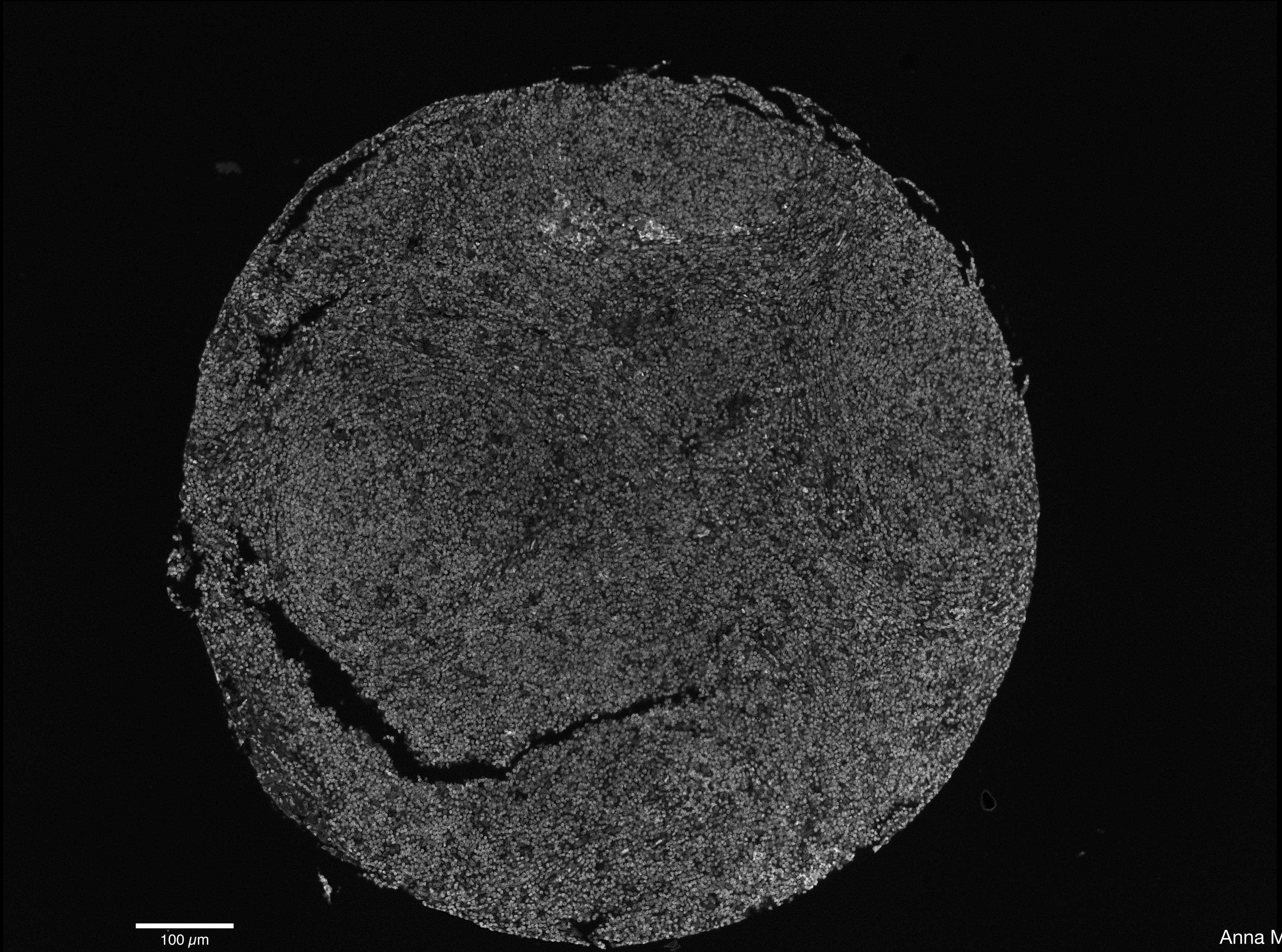
low threshold: already slightly overlaps are counted as TP (high AP)

high threshold: only almost complete overlaps are counted as TP (low AP)



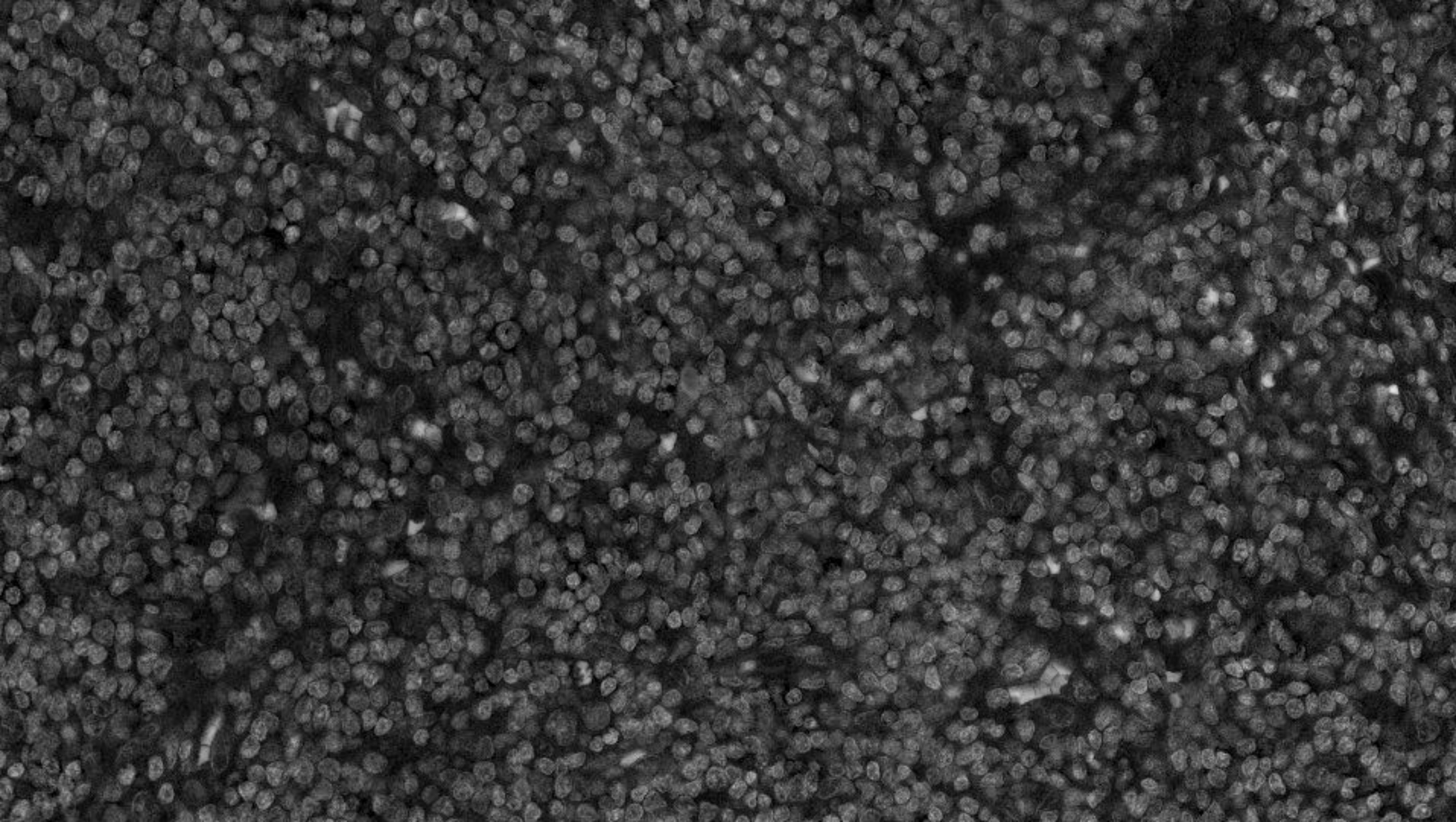
Precision Recall Average Precision (AP) Accuracy	$\frac{TP}{TP + FP}$ $\frac{TP}{TP + FN}$ $\frac{TP}{TP + FP + FN}$
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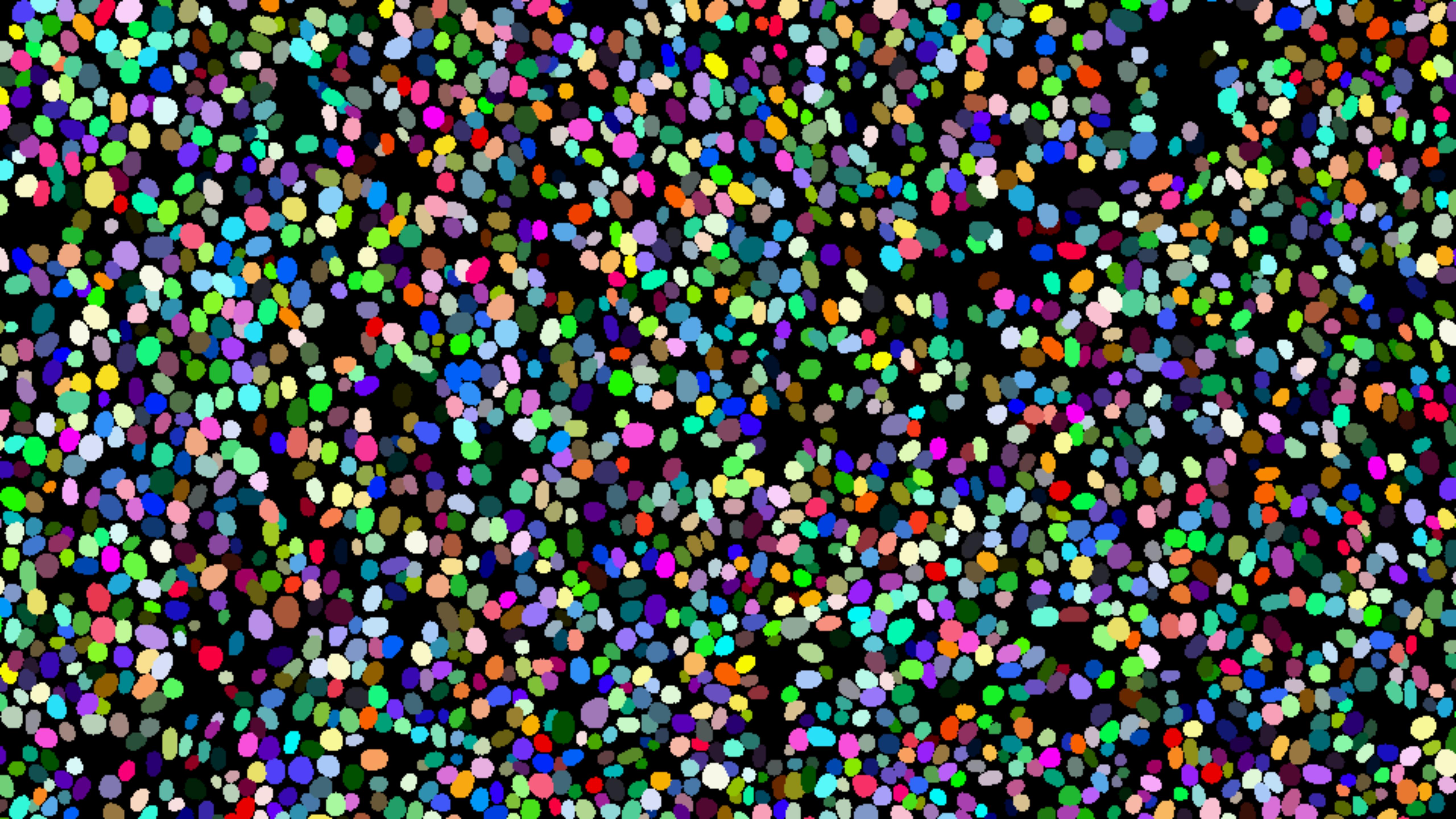
Example: crowded lymphoma cells



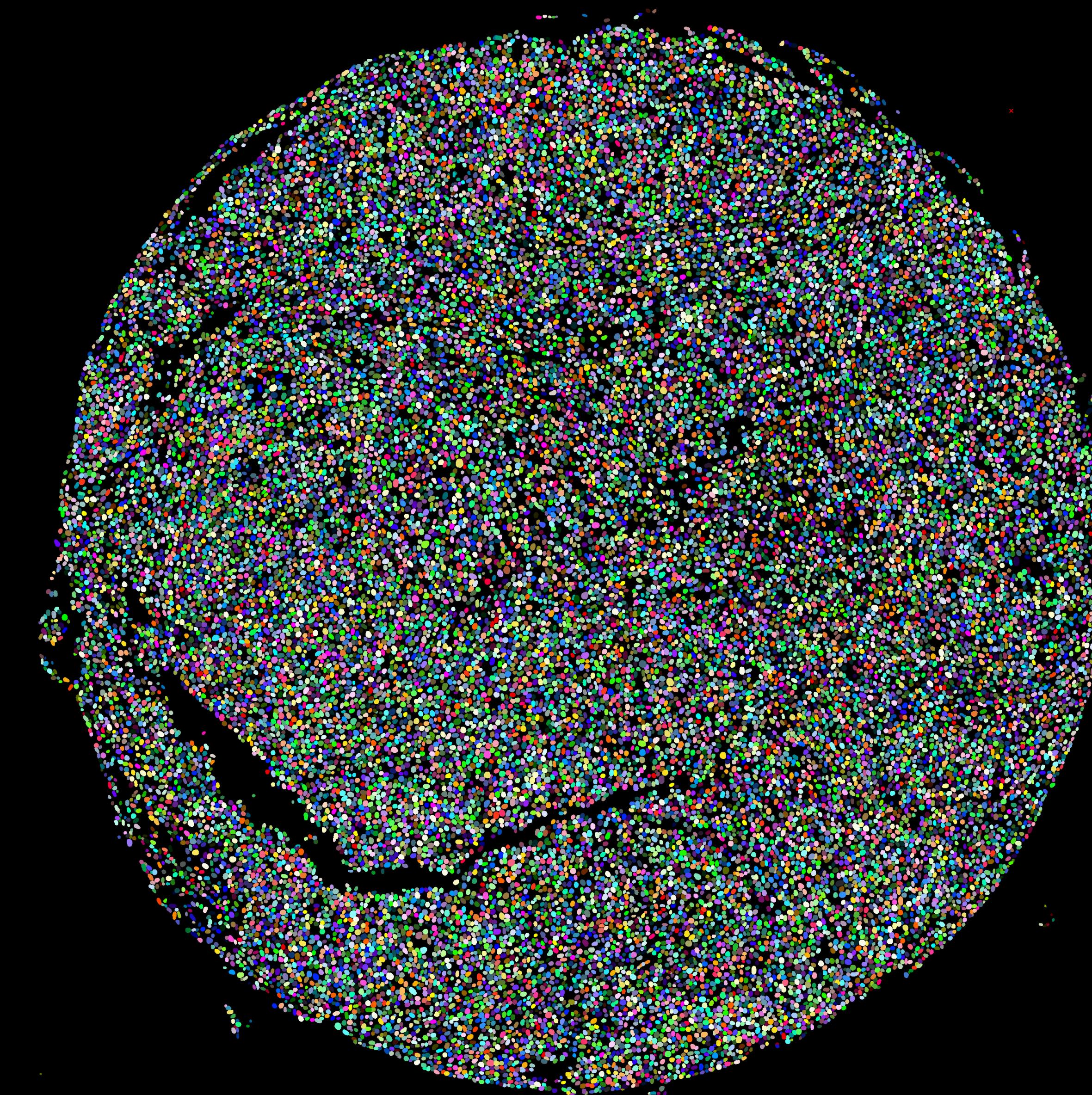
100 μm

Anna Maria Tsakiroglou (Manchester)

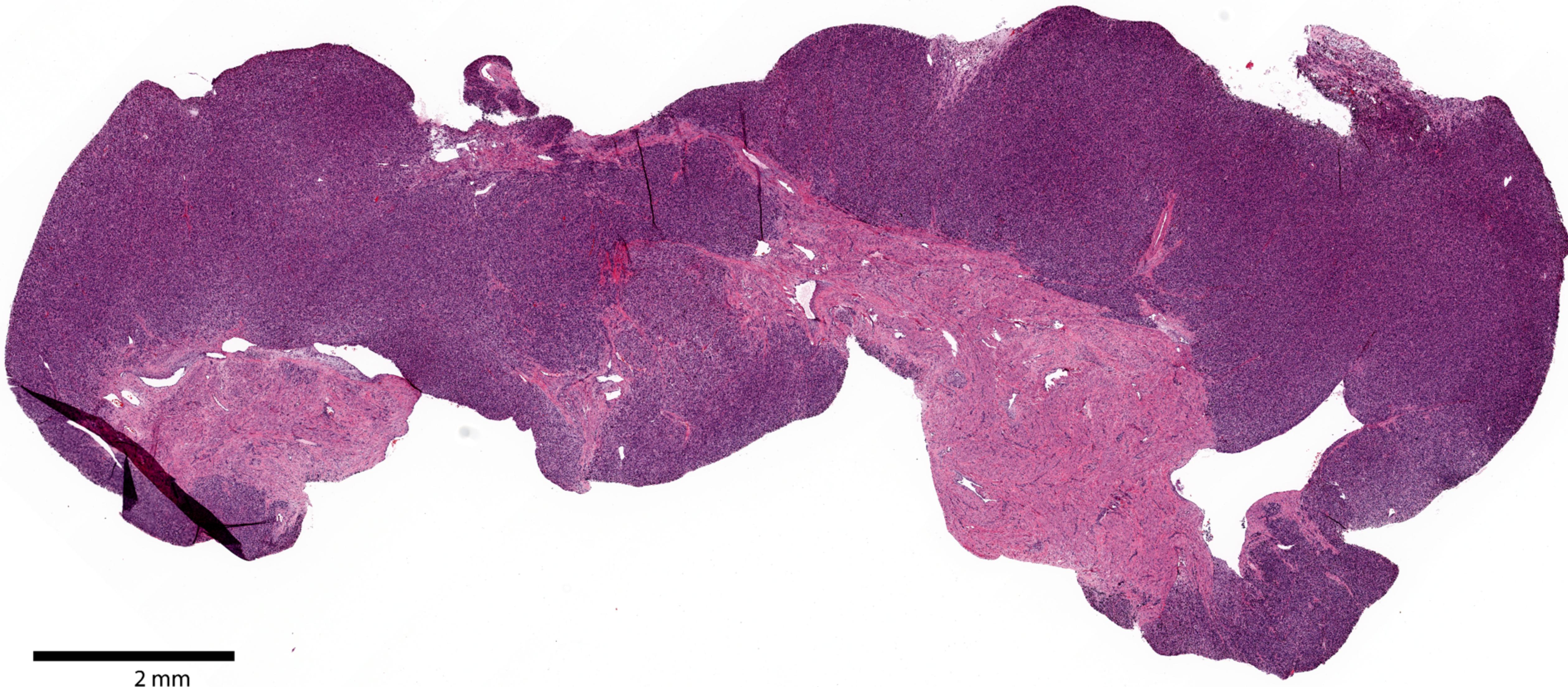




Example: crowded lymphoma cells

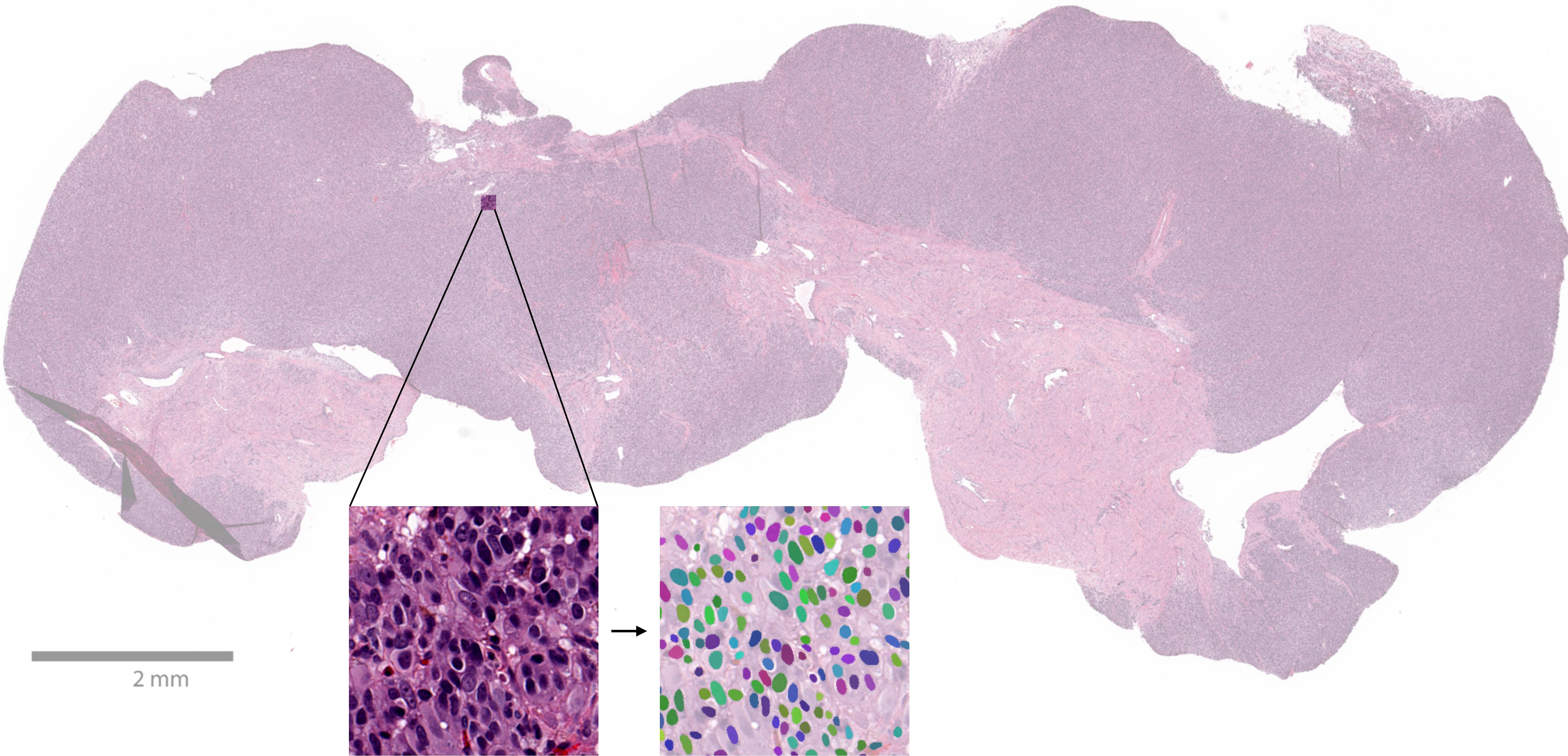


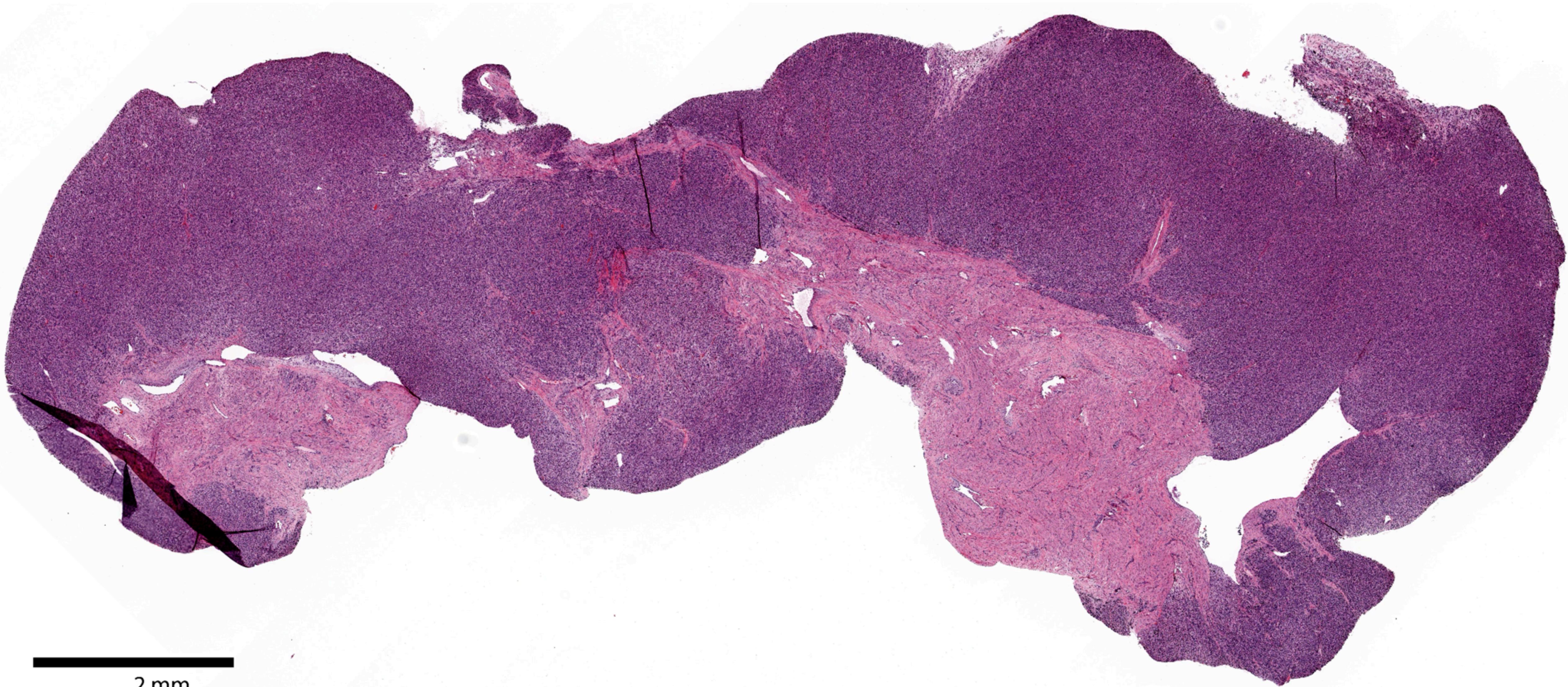
Example: Histopathology (H&E stain)



Whole Slide H&E (Sarcoma), (15mm x 6mm, 32k x 14k pixels, 1.35 GB)

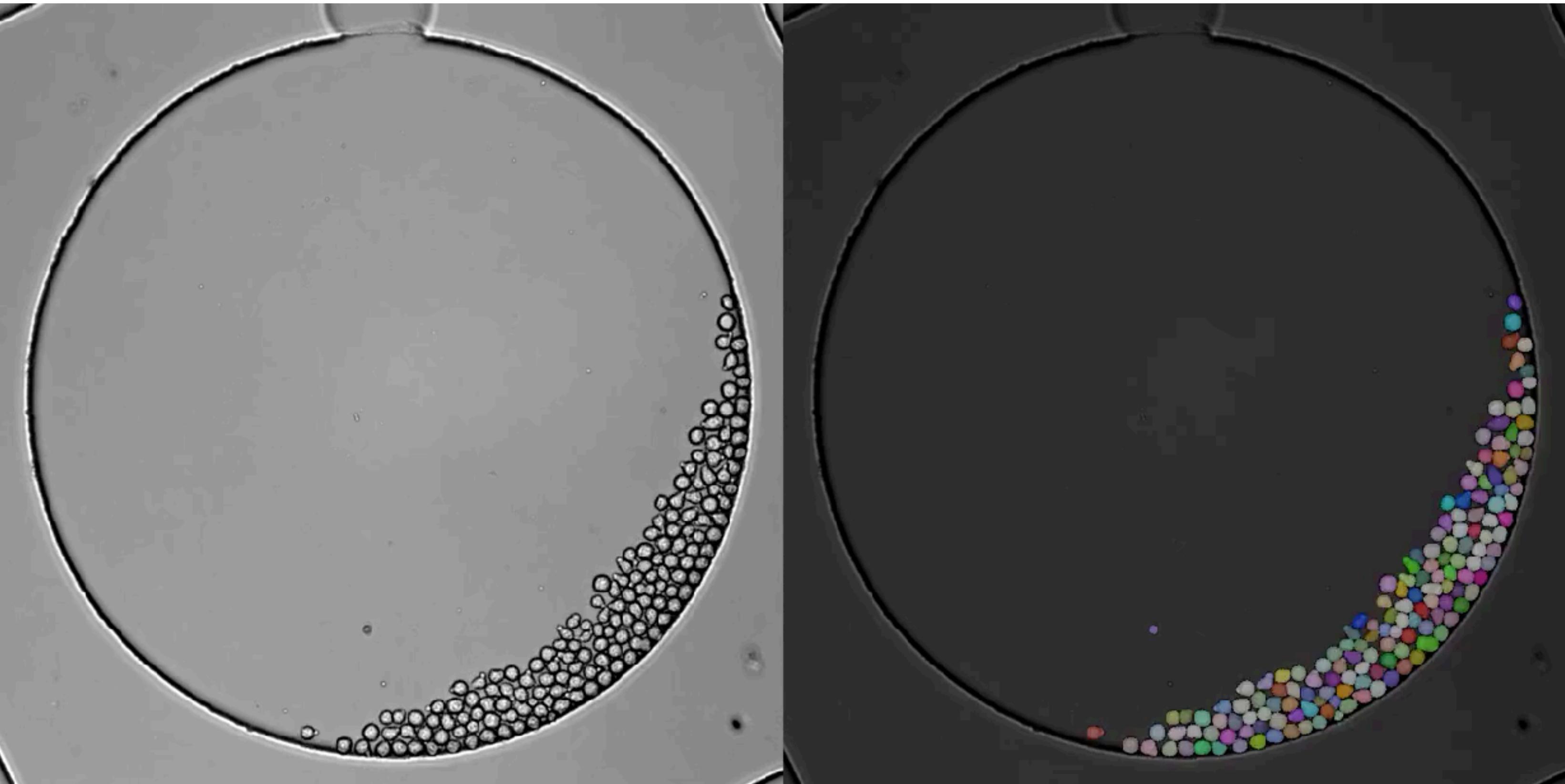
Example: Histopathology (H&E stain)





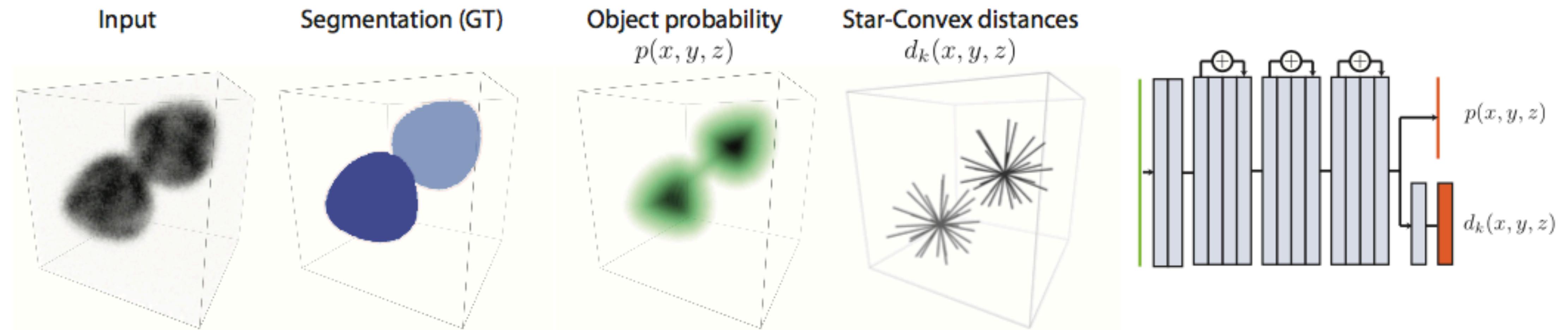
2 mm

Example: Brightfield



StarDist for 3D images

Similar approach:



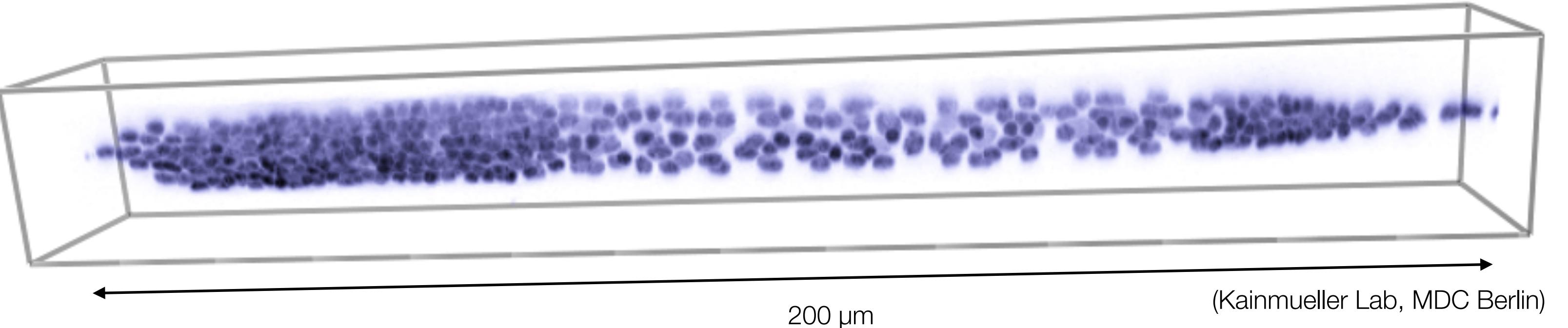
Additional Considerations:

- Ray choice: Fibonacci Lattice on sphere ~ 96 points
- Data anisotropy: Adjust rays according to GT anisotropy
- Non-Maximum Suppression of large sets of polyhedra (> 2 Mio candidates)

Examples 3D

C. elegans (L1)

- 28 Stacks
- ~15k annotated cells
- almost isotropic resolution



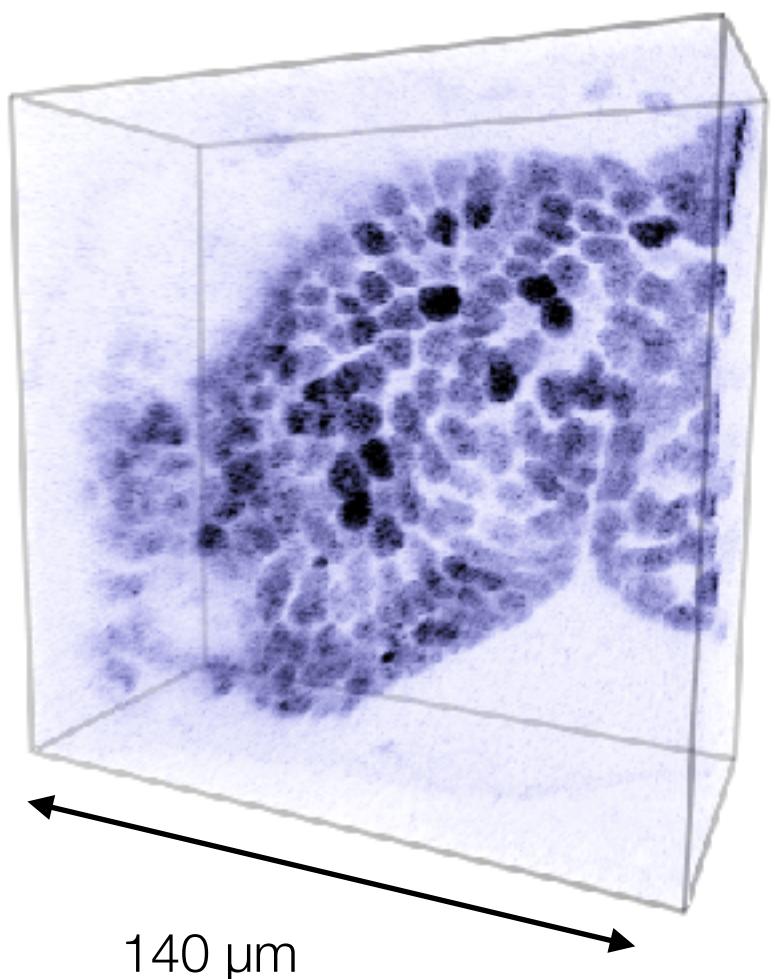
(Kainmueller Lab, MDC Berlin)



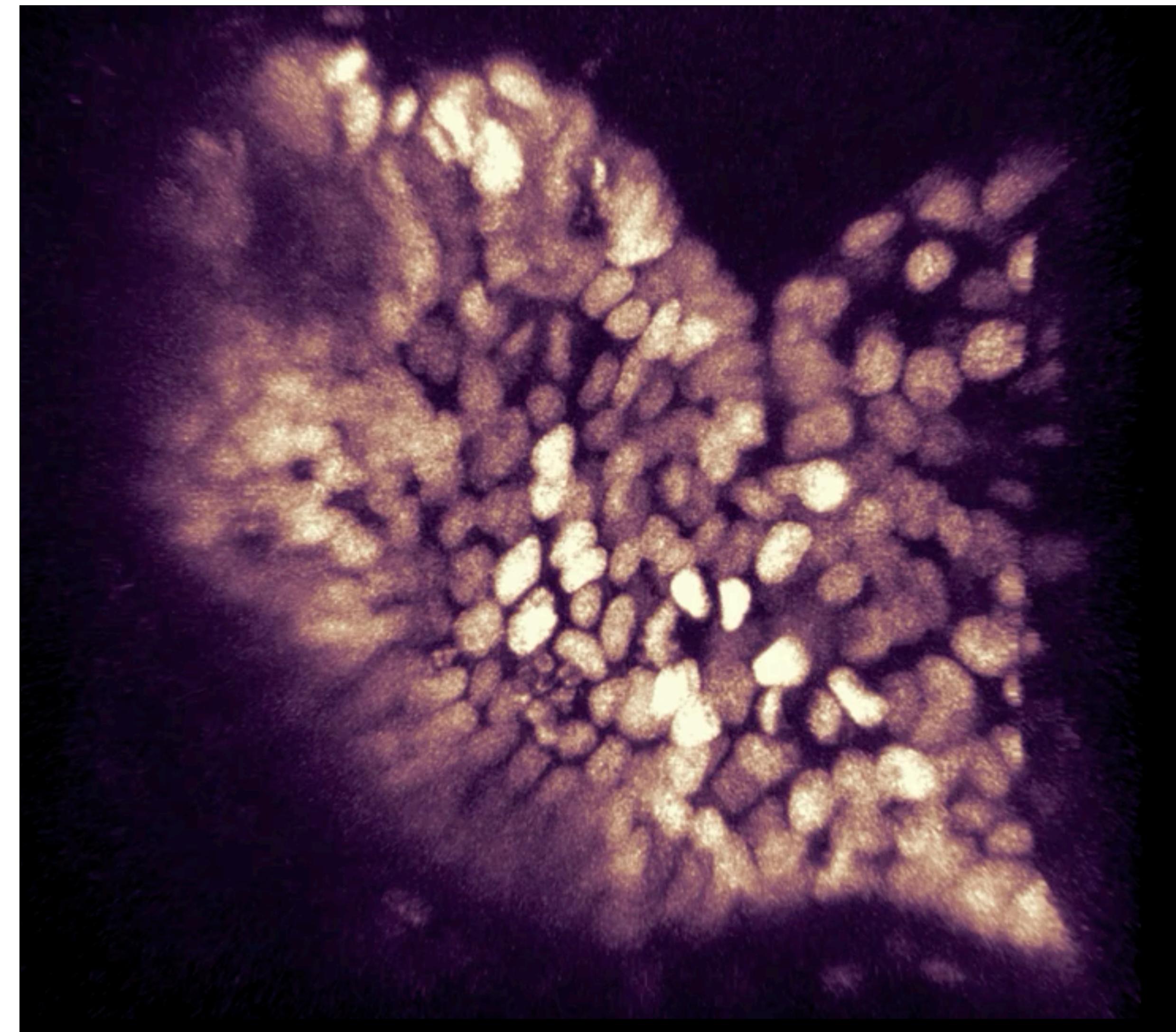
Examples 3D

Parhyale hawaiensis

- 6 stacks
- ~1500 annotated cells
- highly anisotropic resolution



(Ko Sugarawa, IGF Lyon)



Questions & Answers 1

How to use StarDist

How to use StarDist



Main python library

```
pip install stardist
```

<https://github.com/mpicbg-csbd/stardist>

- Training and prediction for 2D/3D images
- Neural network backend keras/tensorflow via csbdeep
- Sensible training defaults
- Multi-Core NMS, tiled prediction
- Image normalization
- Segmentation/Detection measures
- Model export to Fiji

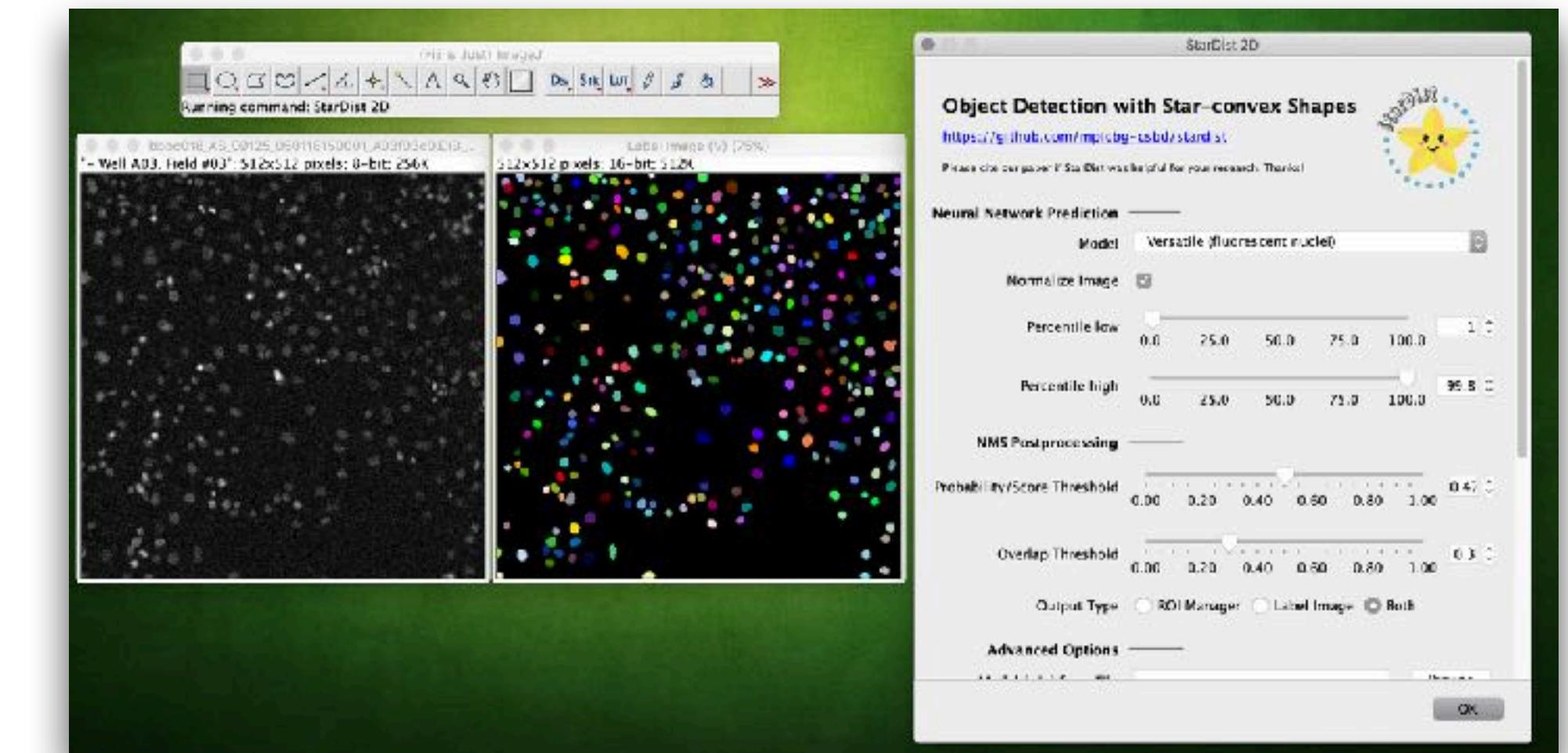
```
from stardist.models import StarDist2D, StarDist3D
model = StarDist2D(config, name = "mymodel")
Using default values: prob_thresh=0.5, nms_thresh=0.4.
model.train(X,Y,validation_data=(Xv,Yv))
Epoch 1/400
 53/100 [=====>.....] - ETA: 26s - loc_
t_loss: 8.6035 - prob_kld: 0.3578 - dist_relevant_mae: 8
labels, _ = model.predict_instances(img)
```



Fiji Plugin

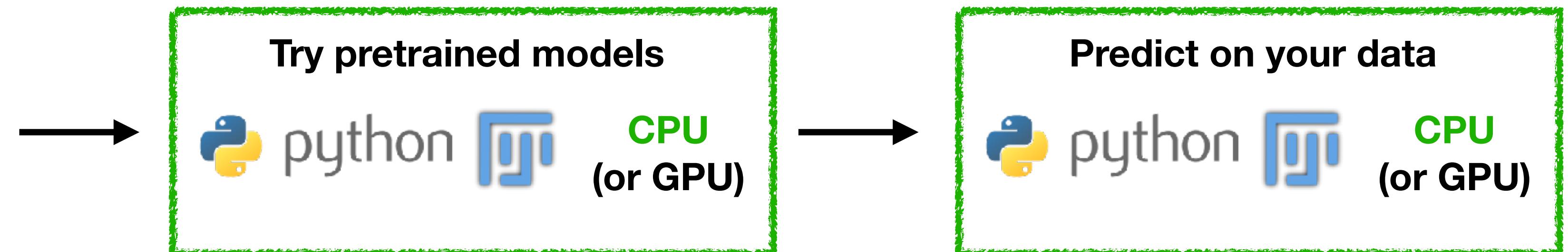
<https://imagej.net/StarDist>

- Prediction for 2D images by already trained models
- Scriptable
- CPU and GPU support (via CSBDeep-Fiji by Deborah Schmidt, MPI-CBG)

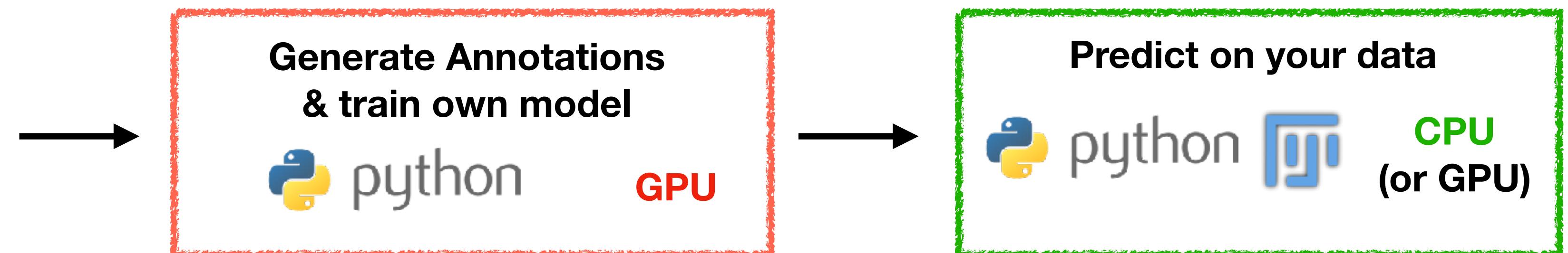


How to use StarDist on your own data

- 2D data
- similar to pretrained images (H&E, fluorescent nuclei)

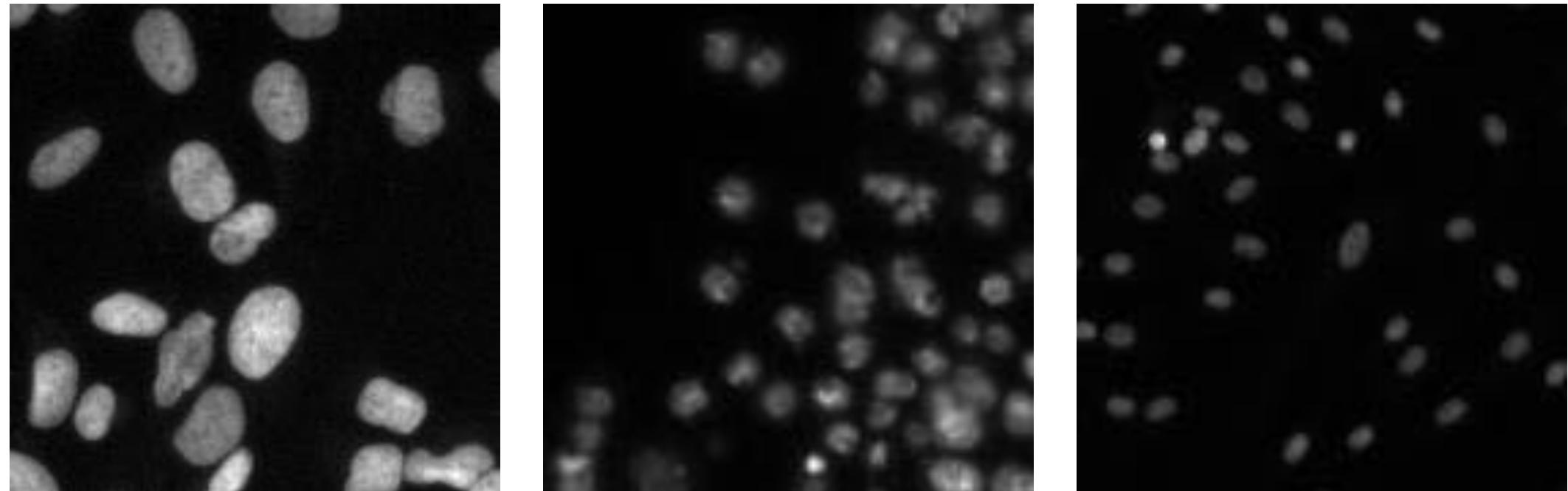


- 2D data dissimilar to pretrained
- 3D data



Pretrained Models (2D)

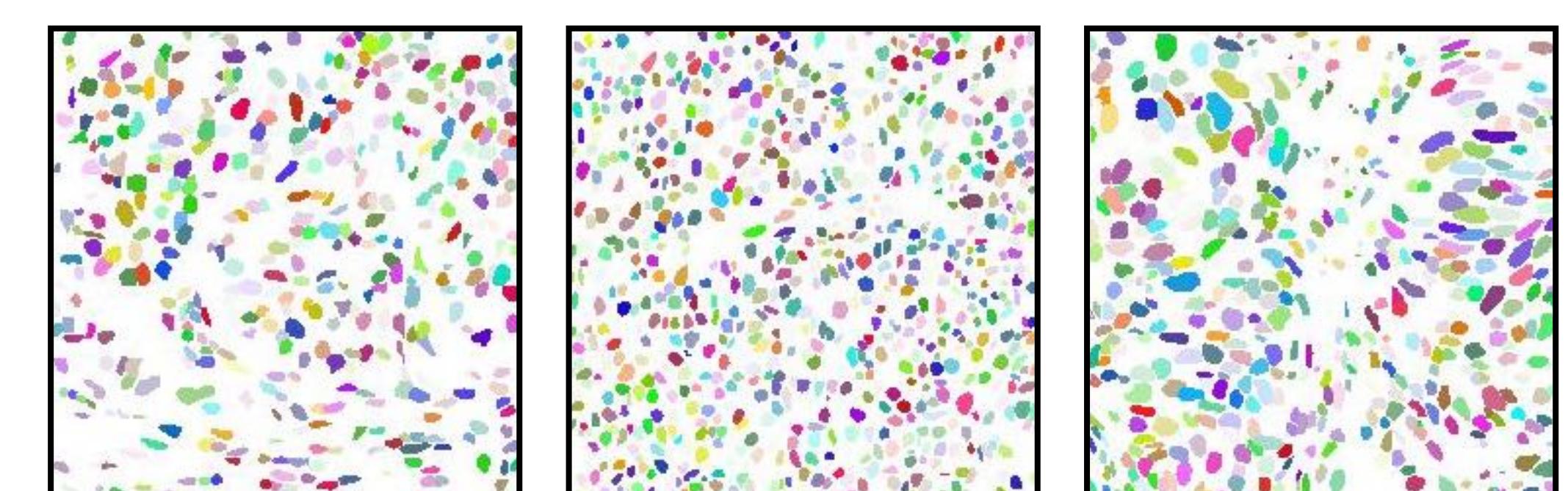
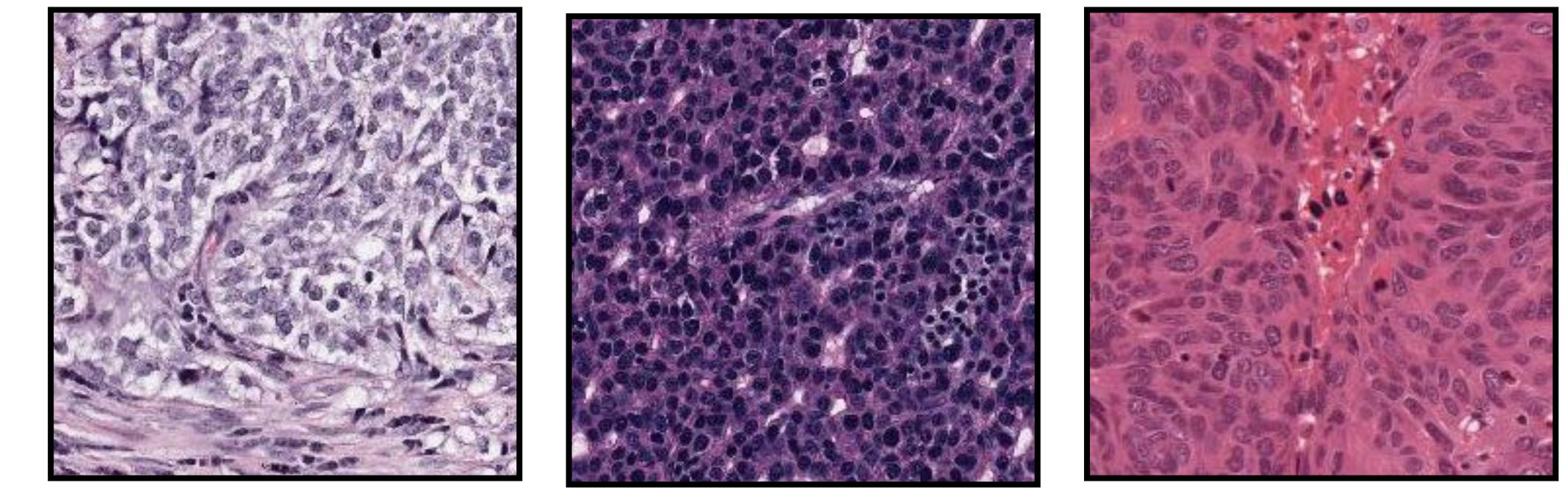
Fluorescence Microscopy
Single Channel



Data Science Bowl 2018
Caicedo et al. (2018)

~ 600 images (2D)
~ 20k annotations

Histopathology
RGB H&E



MoNuSeg
Kumar et al (2017)

~ 30 Images (2D)
~ 22k annotations

Pretrained Models (2D)



```
from stardist.models import StarDist2D

StarDist2D.from_pretrained()

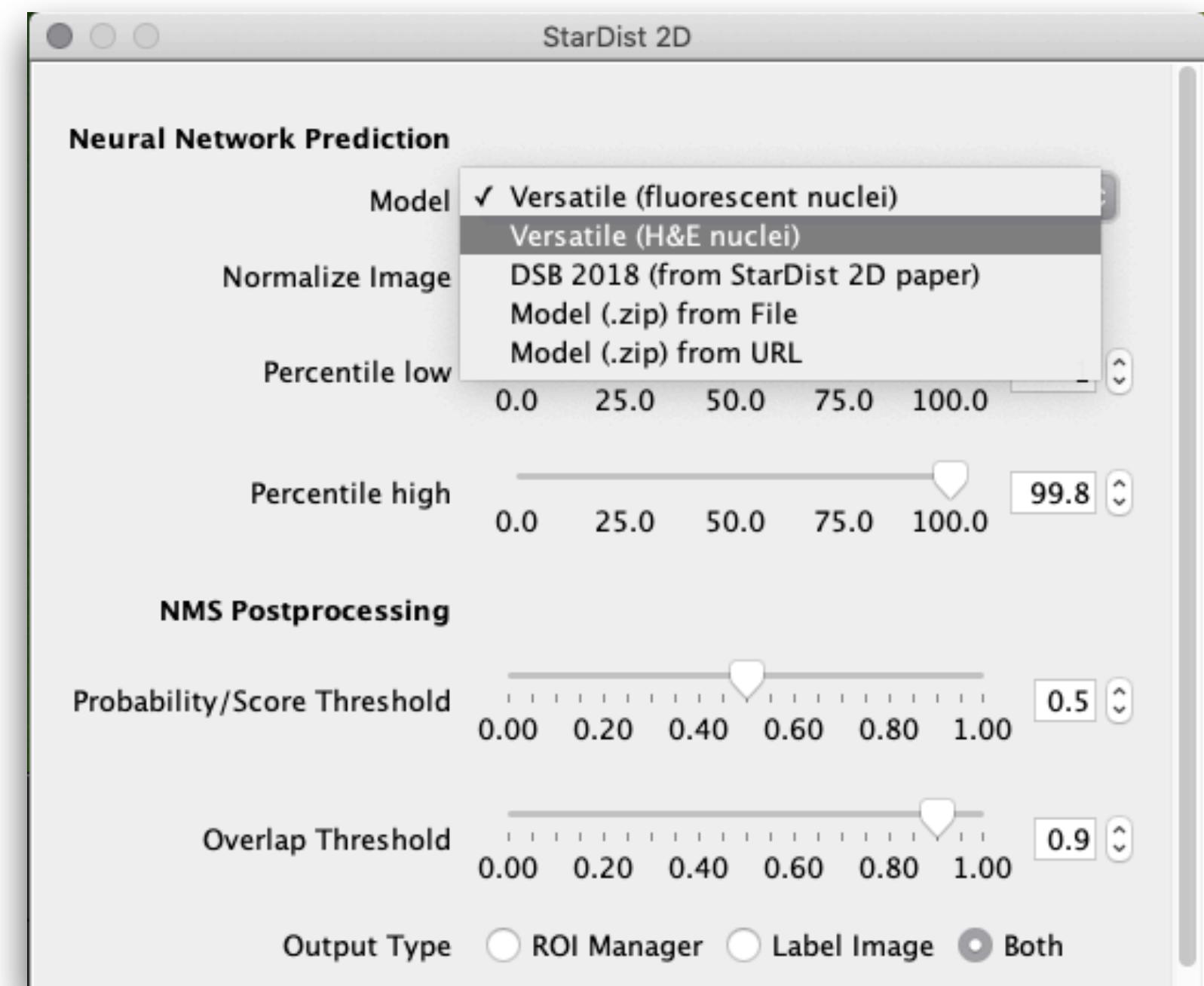
There are 4 registered models for 'StarDist2D':

Name           Alias(es)
_____
'2D_versatile_fluo'  'Versatile (fluorescent nuclei)'
'2D_versatile_he'    'Versatile (H&E nuclei)'
'2D_paper_dsb2018'   'DSB 2018 (from StarDist 2D paper)'
'2D_demo'           None

model = StarDist2D.from_pretrained('2D_versatile_fluo')

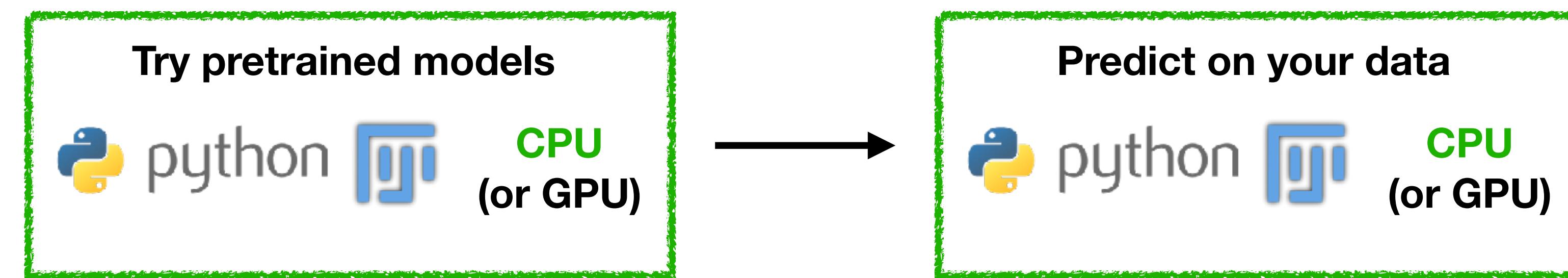
labels, _ = model.predict_instances(img)

Found model '2D_versatile_fluo' for 'StarDist2D'.
Loading network weights from 'weights_best.h5'.
Loading thresholds from 'thresholds.json'.
Using default values: prob_thresh=0.479071, nms_thresh=0.3.
```



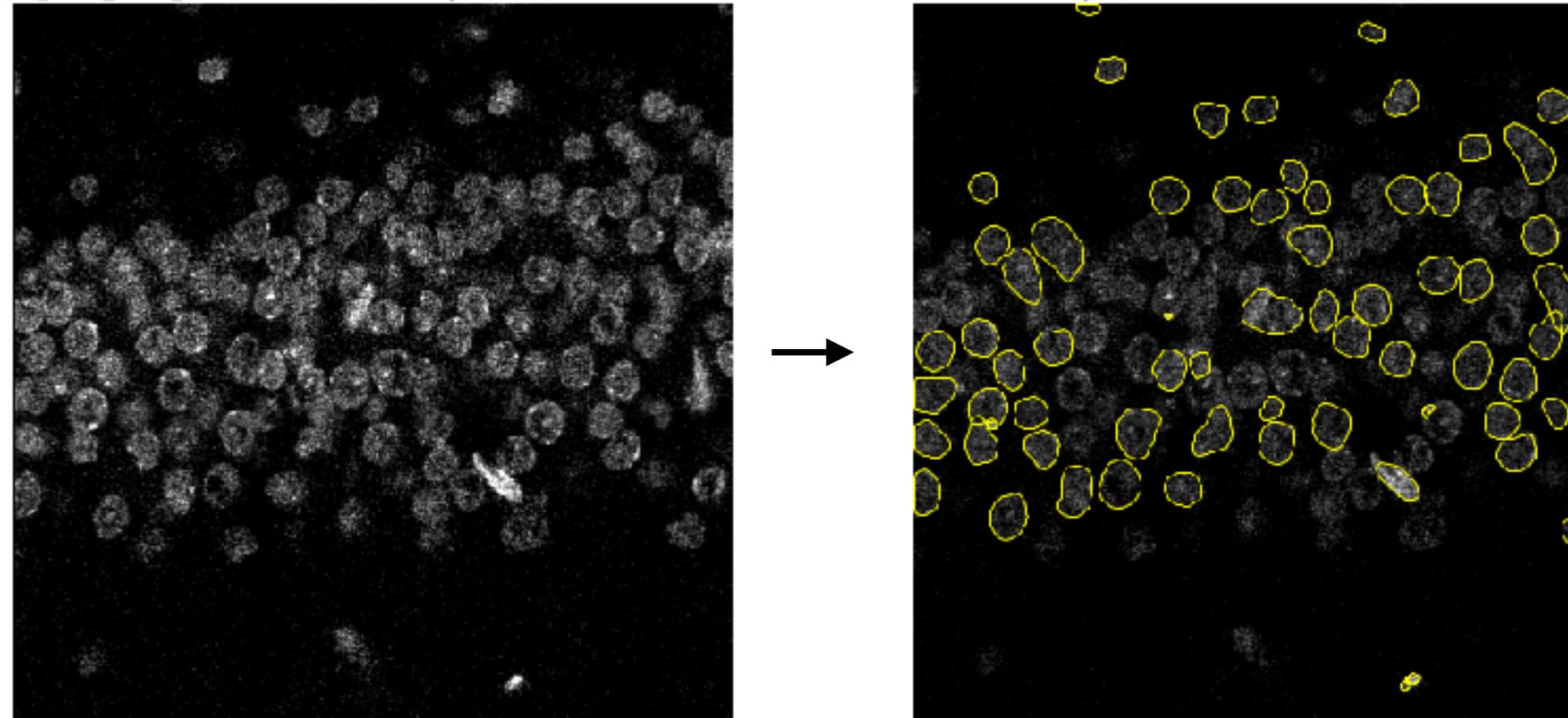
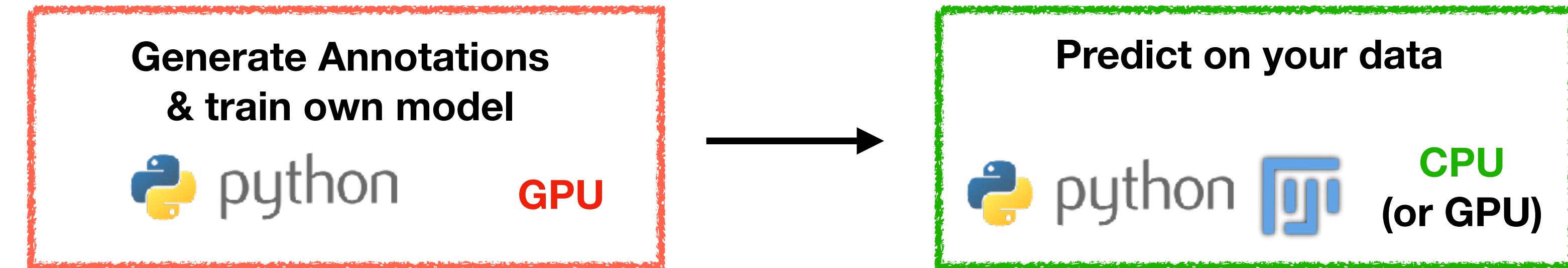
- Try different scalings
- Invert intensity, RGB -> grayscale,
- Play around with the prob and overlap (NMS) threshold

Demo: Pretrained models in Python and Fiji (2D)



Training of custom models

If the pretrained models do not work on your 2D images (or your data is 3D) you need to training your own model



Data from Silvia Monari, EPFL

- Needs user annotated Image/label pairs
- GPU workstation
- Python
- Training time from scratch:
 - 2D: 30min-2h
 - 3D: 4h-12h

Training of custom models

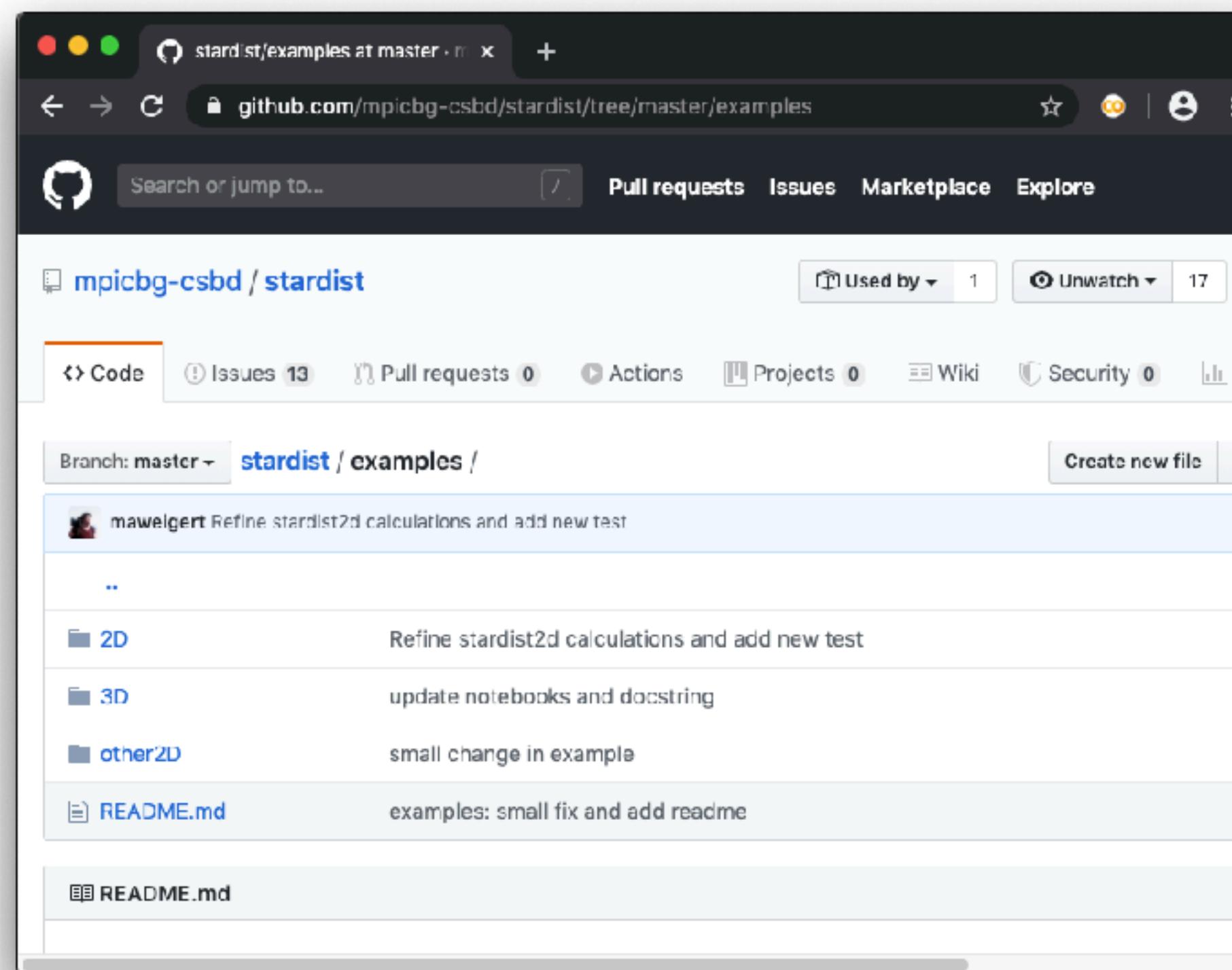
Demo Jupyter Notebooks for 2D and 3D data that you can adapt and run on your (or your facilities) GPU-workstation



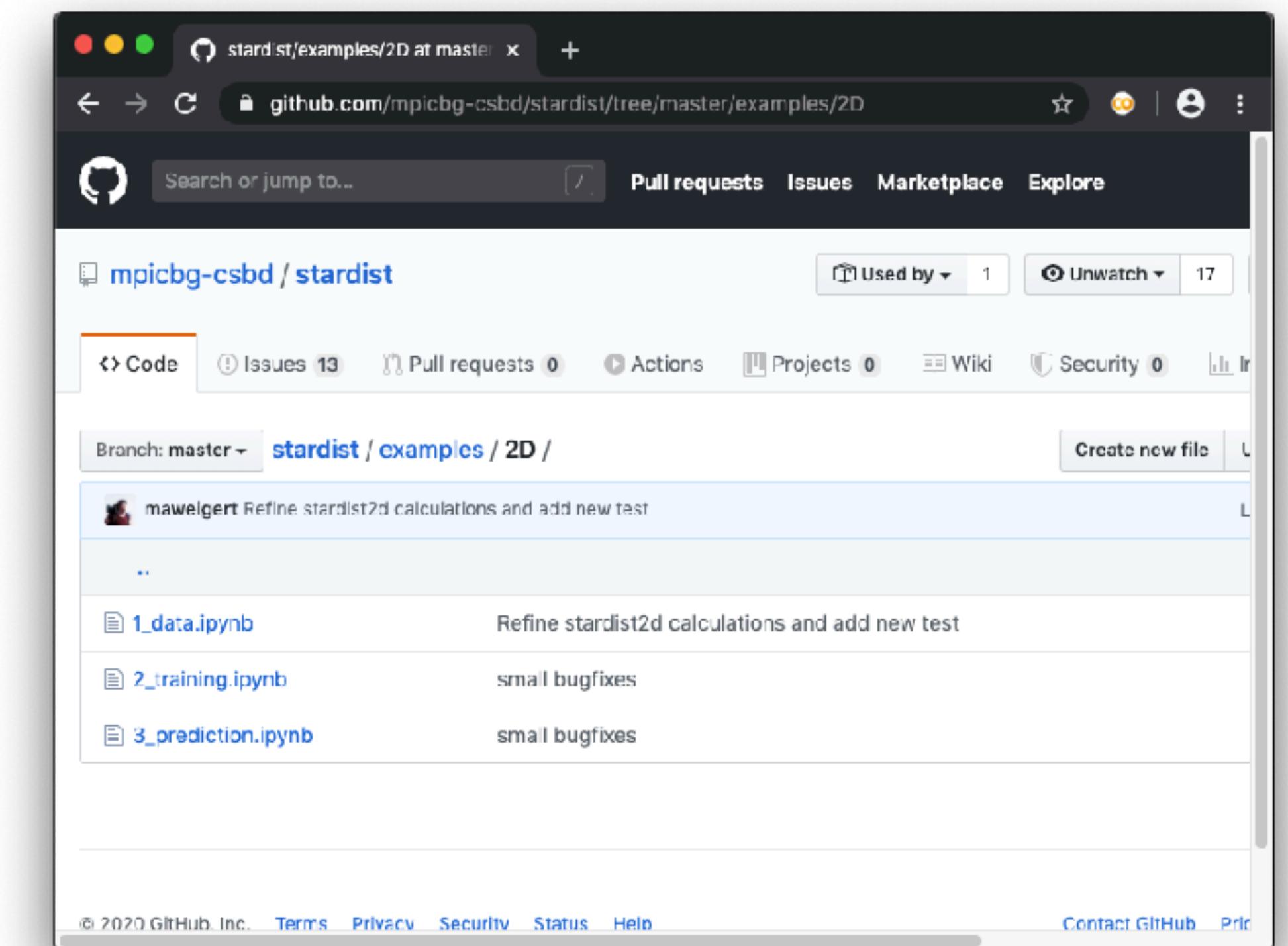
<https://github.com/mpicbg-csbd/stardist/tree/master/examples>

Jupyter Notebooks examples in 2D and 3D

1. Data preparation/inspection
2. Training of the StarDist model
3. Prediction on new images



This screenshot shows the GitHub repository page for 'stardist/examples'. The repository is owned by 'mpicbg-csbd' and has a 'stardist' parent repository. It contains 13 issues and 0 pull requests. The 'Code' tab is selected, showing a list of files and folders. The 'stardist/examples' folder is expanded, revealing '2D', '3D', and 'other2D' subfolders, each with a brief description. A 'README.md' file is also present.

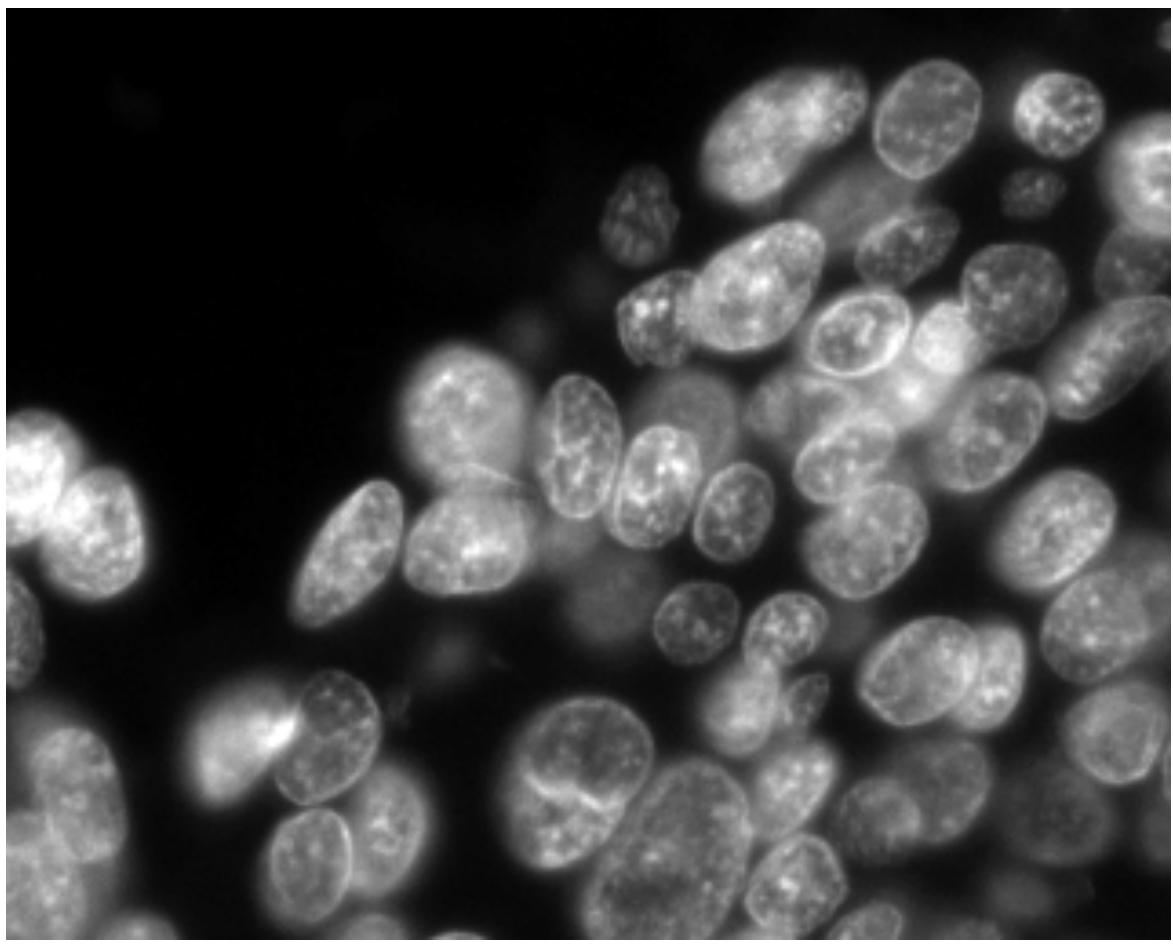


This screenshot shows the GitHub repository page for 'stardist/examples/2D'. It is a subdirectory of the main 'examples' repository. It contains 13 issues and 0 pull requests. The 'Code' tab is selected, showing a list of files. It includes '1_data.ipynb', '2_training.ipynb', and '3_prediction.ipynb', each with a brief description. A 'README.md' file is also present at the top level of this directory.

Training data generation

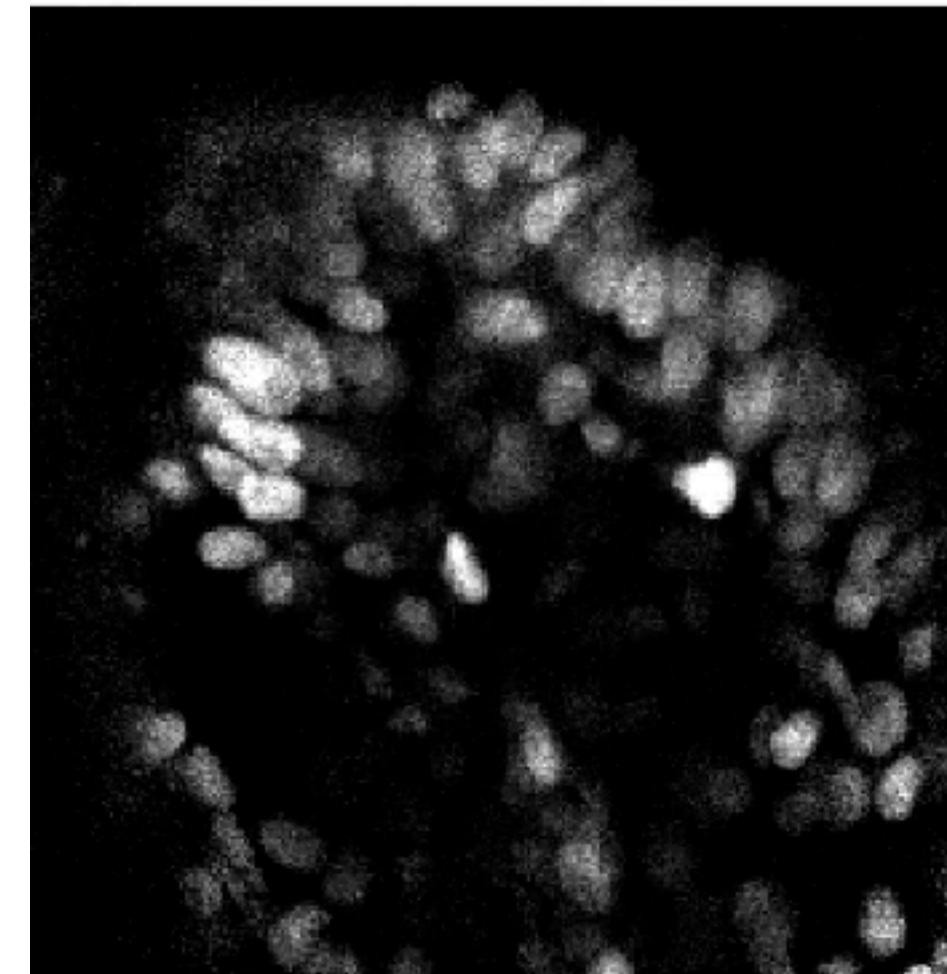
Generate corresponding Image/Mask pairs:

2D

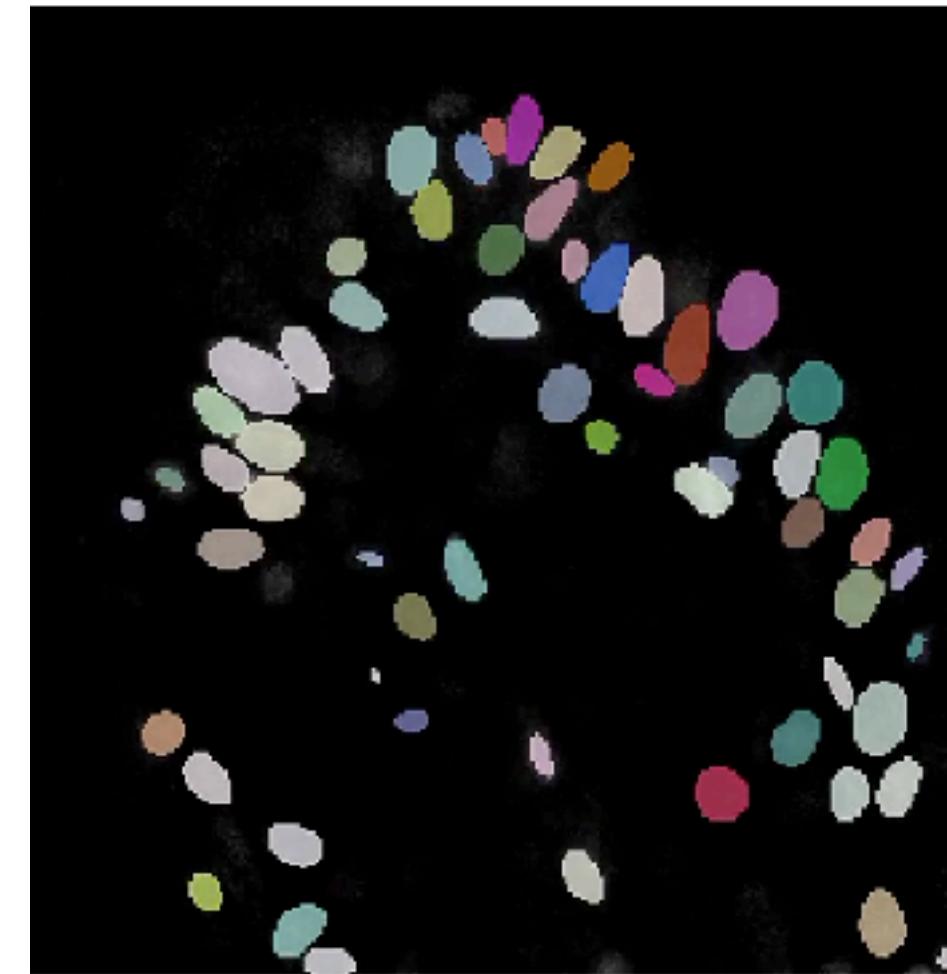
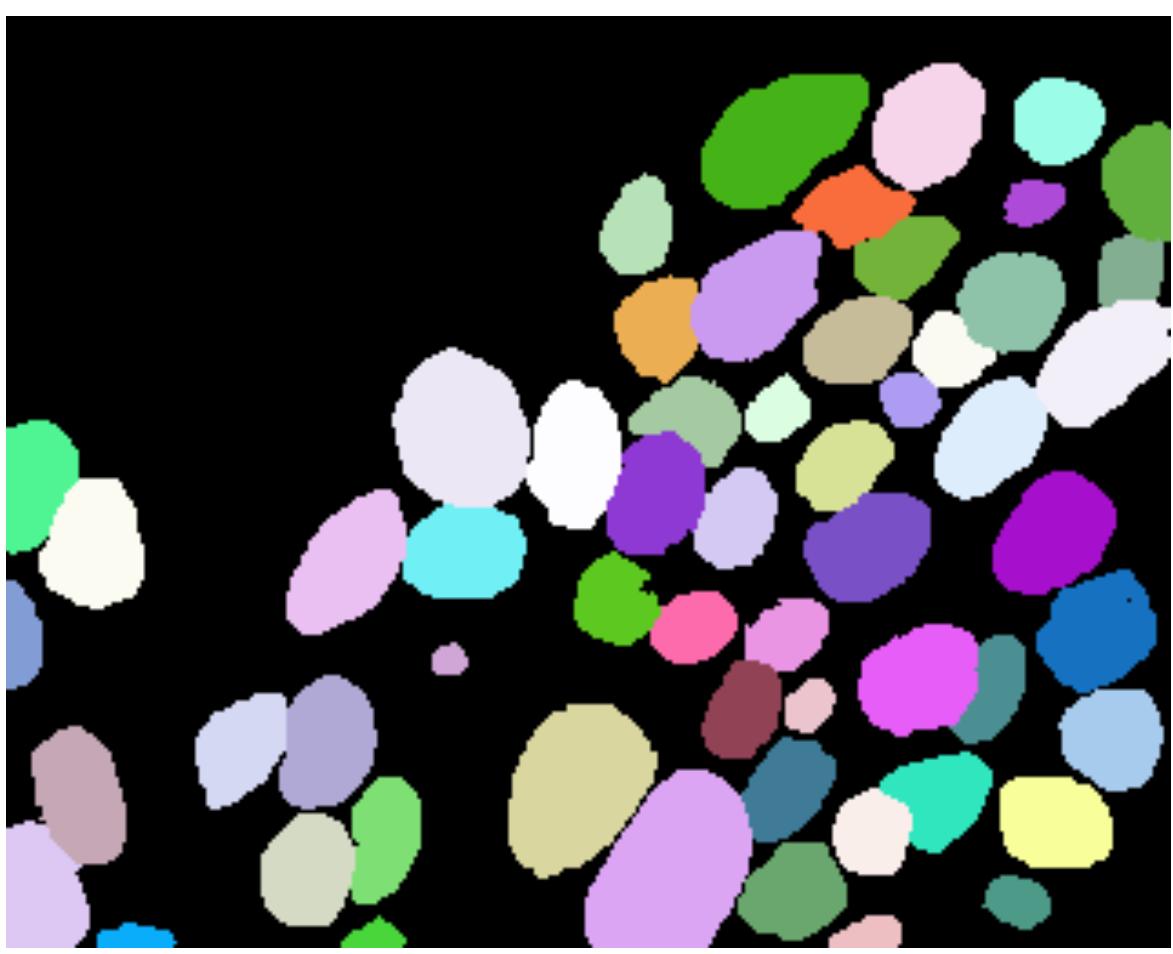


Images

3D



Masks



- Image/Mask: tif format
- Mask: Every cell pixel has to have a different label (dense labeling)
- Use crops from different regions/timepoints
- Size: at least 128^2 (2D) or 96^3 (3D)
- Number: $N > 10$ (2D) and $N > 4$ (3D)

Training data generation

```

data/
  └── train
    ├── images
    │   ├── img_1.tif
    │   ├── img_2.tif
    │   ├── img_3.tif
    │   ├── img_4.tif
    │   ├── img_5.tif
    │   └── img_6.tif
    └── masks
        ├── mask_1.tif
        ├── mask_2.tif
        ├── mask_3.tif
        ├── mask_4.tif
        ├── mask_5.tif
        └── mask_6.tif
  └── test
    ├── images
    │   ├── img_6.tif
    │   ├── img_7.tif
    │   └── img_8.tif
    └── masks
        ├── mask_6.tif
        ├── mask_7.tif
        └── mask_8.tif

```

Generate corresponding Image/Mask pairs:

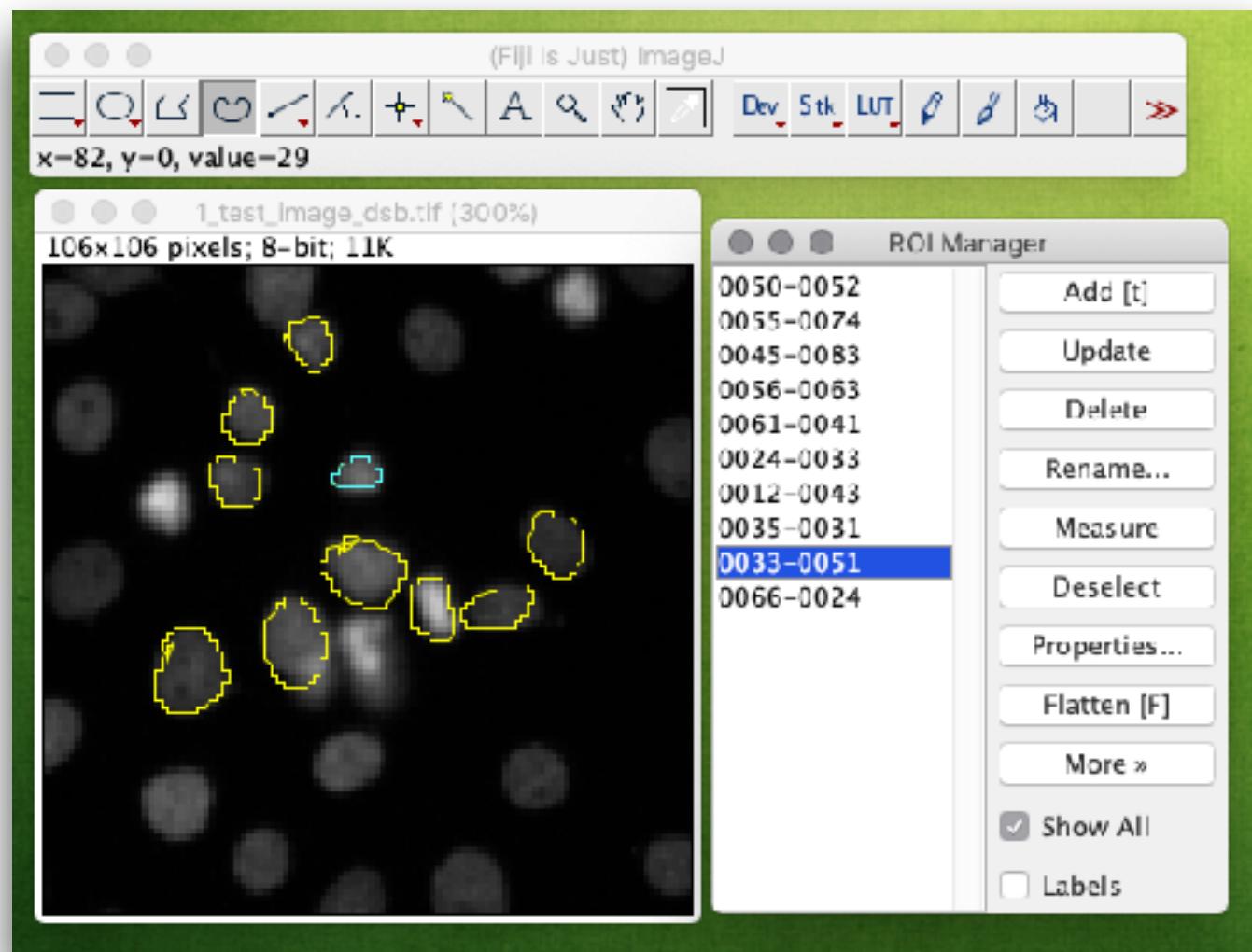
- Image/Mask: tif format
- Mask: Every cell pixel has to have a different label (dense labeling)
- Use crops from different regions/timepoints, e.g.
2D: > 10 of size 128x128
3D: > 4 of size 32x96x96
- Split into (non-overlapping) train and test for validation

Annotation Software (2D)



Fiji/ImageJ

- Draw Roi per object (Roi-Manager)
- Convert to label mask (e.g. see [here](#))



<http://fiji.sc/>

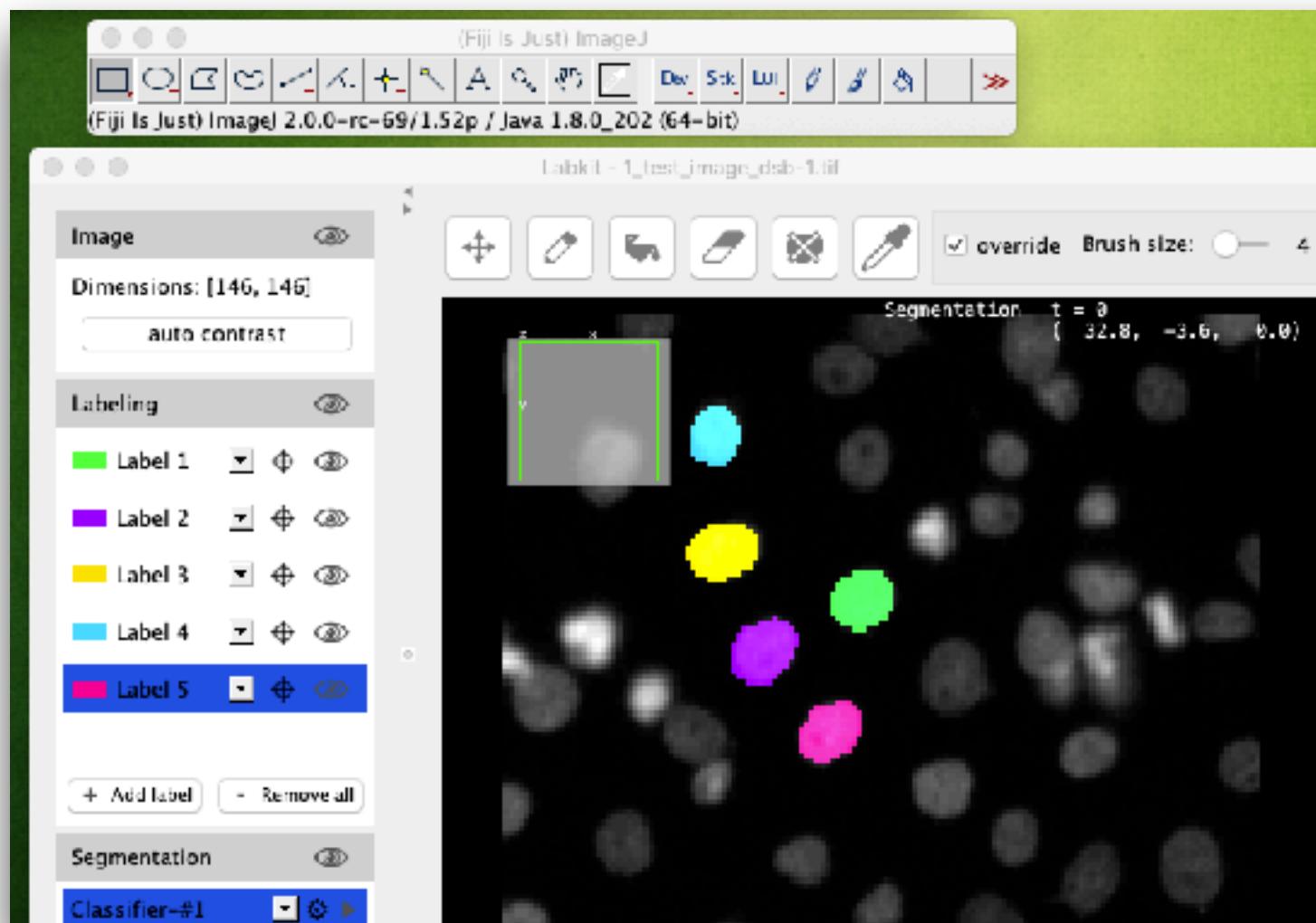
Schindelin et al. (2012)

Schneider, Rasband et al (2012)



Fiji + LabKit

- Directly draw label mask



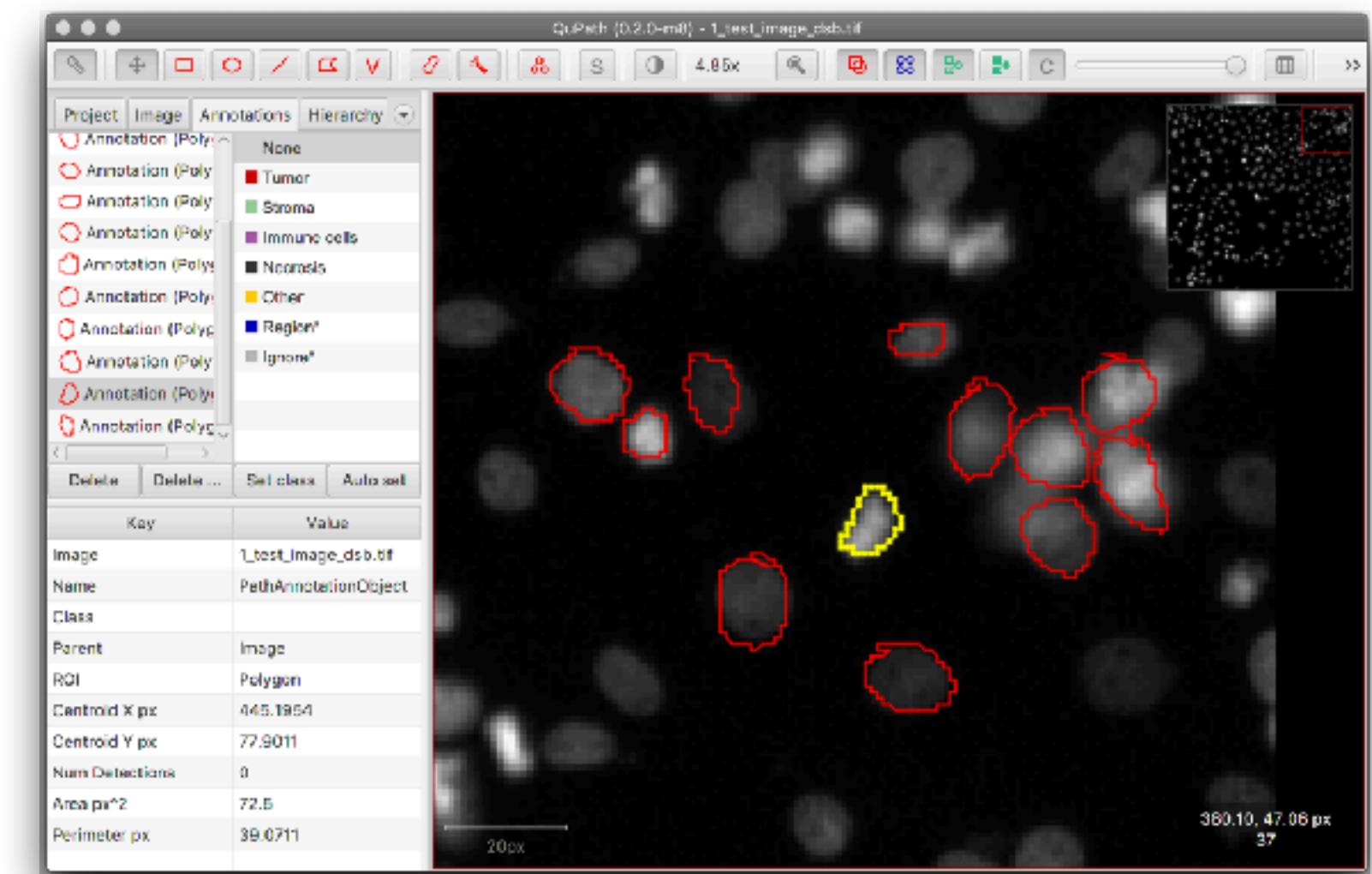
<https://imagej.net/Labkit>

M. Arzt, MPI-CBG



QuPath

- Draw ROI per object
- Convert to label mask



<https://qupath.github.io>

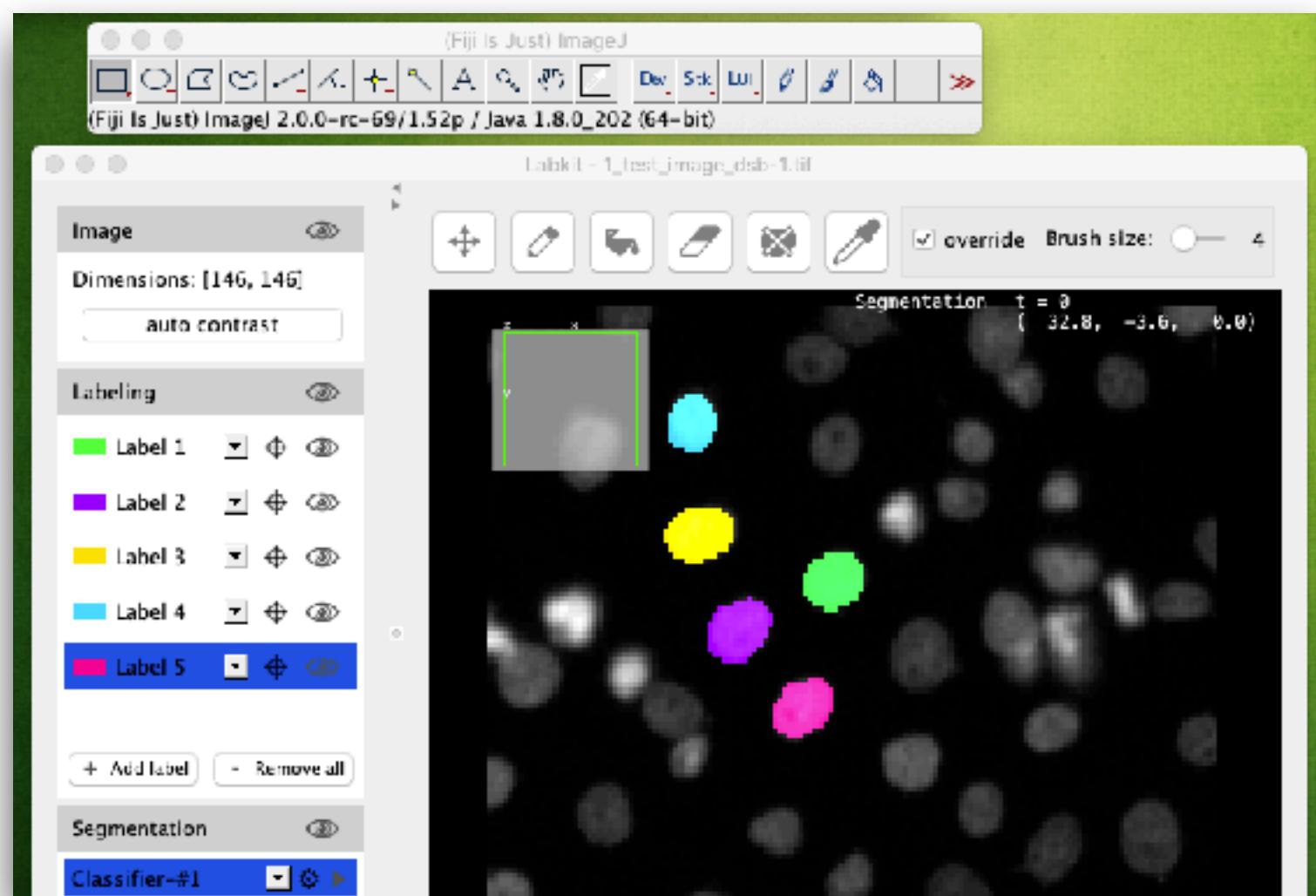
Bankhead et al. (2017)

Annotation Software (3D)



Fiji + LabKit

- Directly draw label mask
- Reorder z -> t (to not miss a plane)



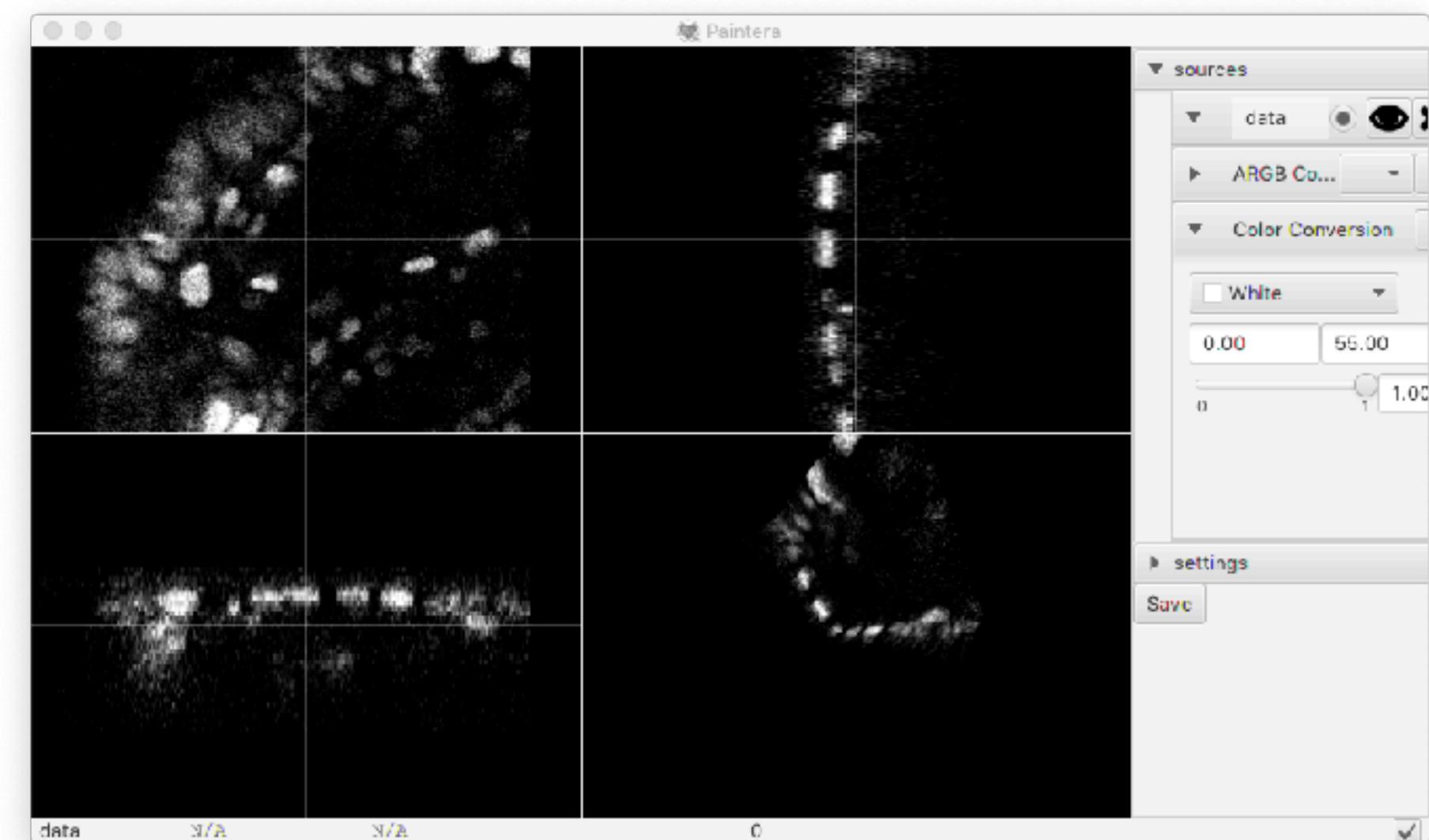
<https://imagej.net/Labkit>

M. Arzt, MPI-CBG



Painter

- For very large volumes (> GBs)
- Powerful but slightly steeper learning curve



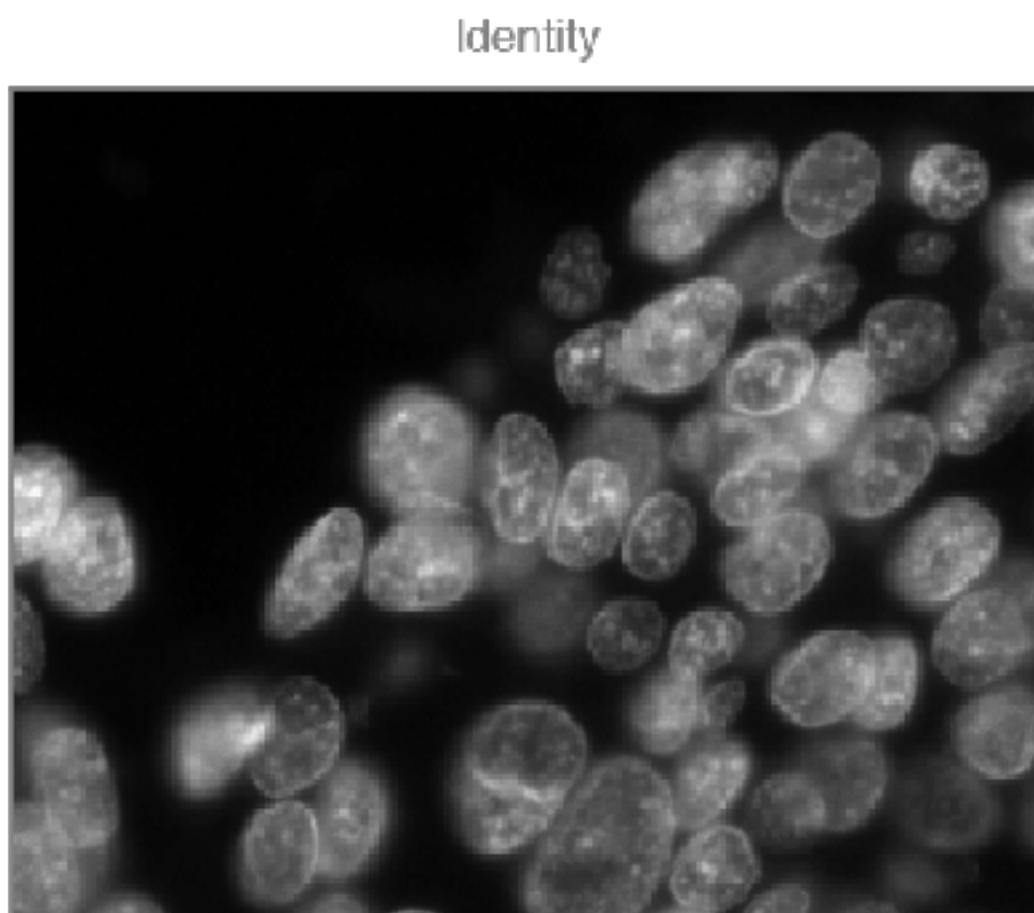
<https://github.com/saalfeldlab/paintera>

P. Hanslovsky, S. Saalfeld et al Janelia

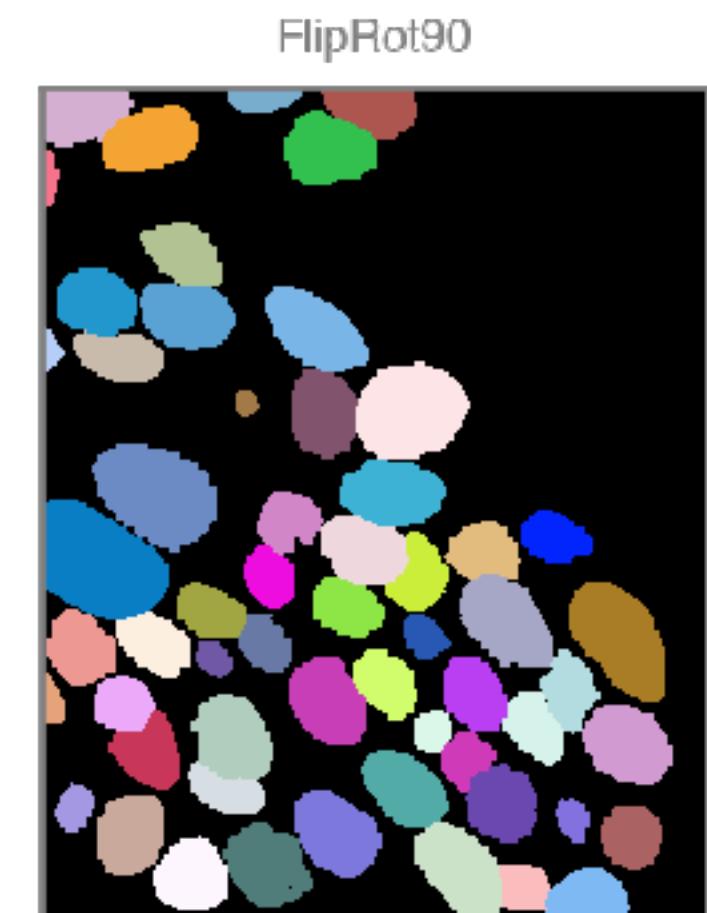
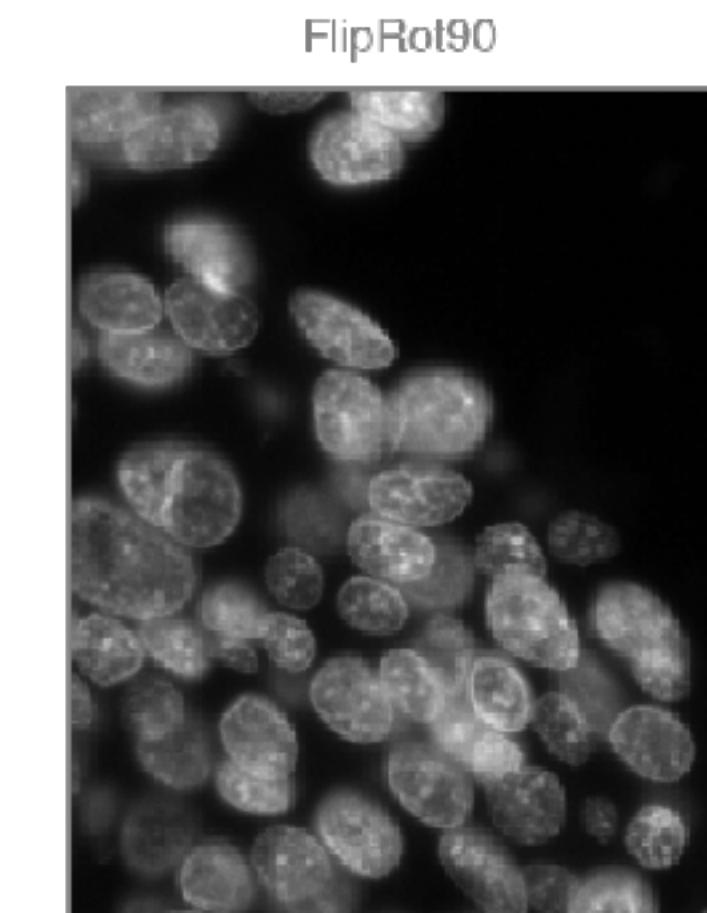
Data Augmentation

Artificially create more training data by transforming existing images/masks into different, yet plausible versions

Original



Flip/90 degree Rotation

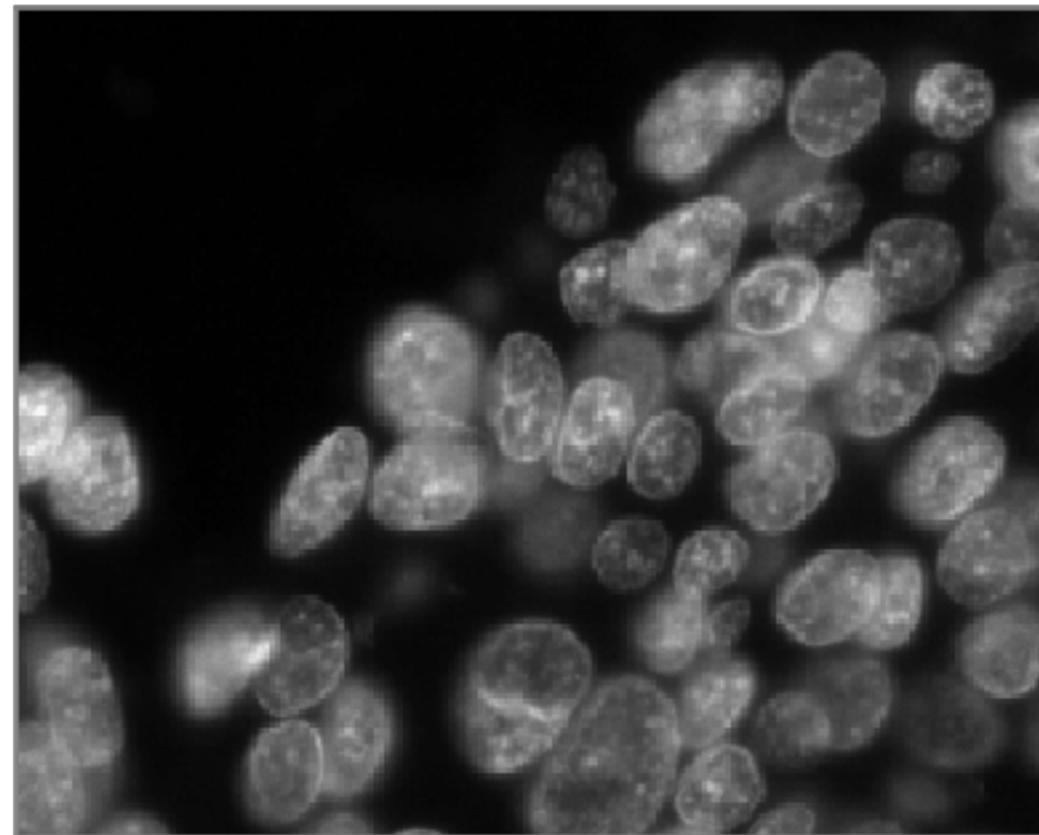


Data Augmentation

Artificially create more training data by transforming existing images/masks into different, yet plausible versions

Original

Identity

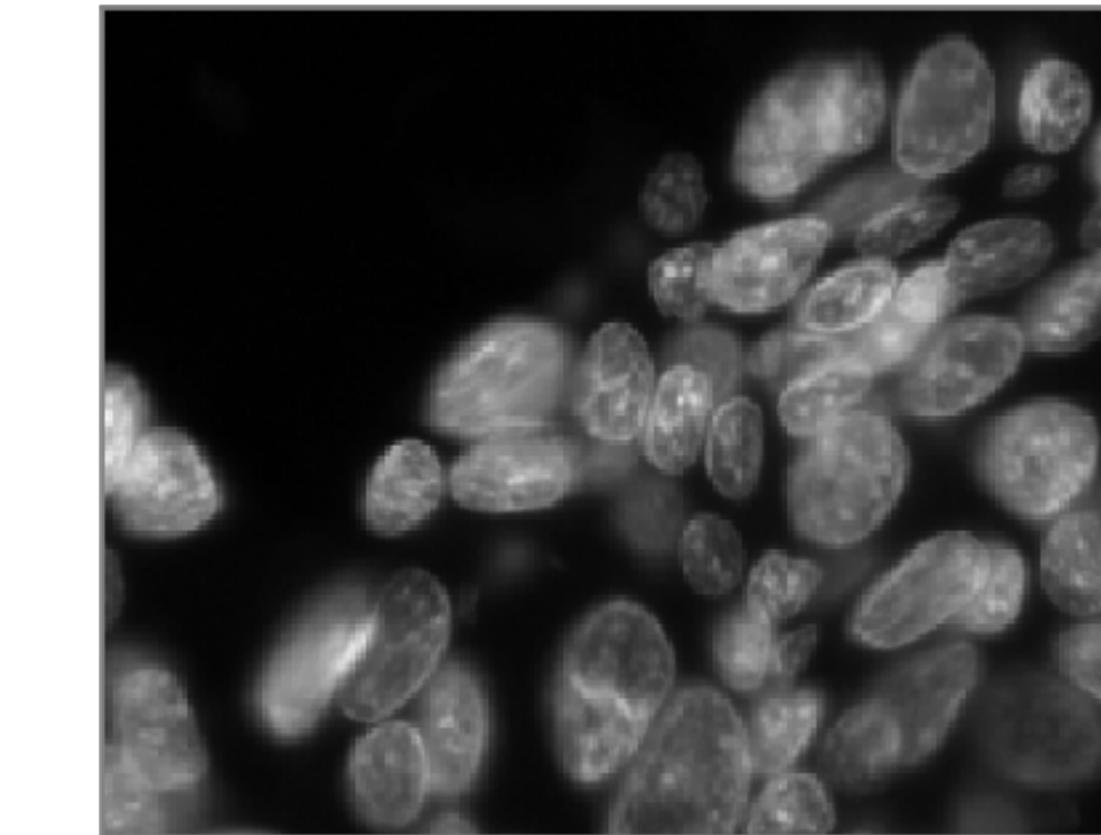


Identity

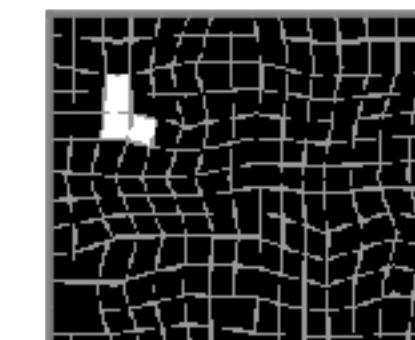
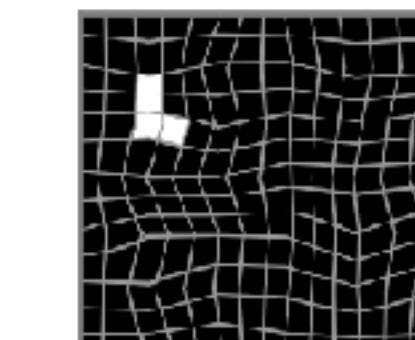
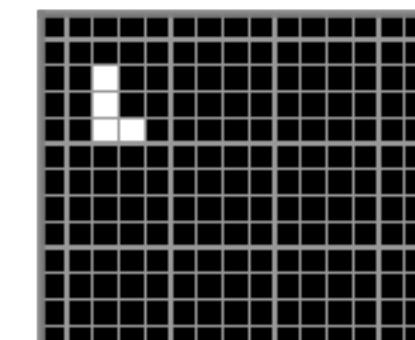
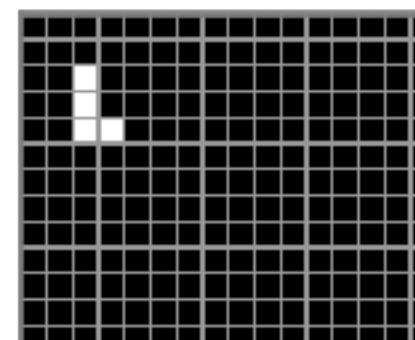
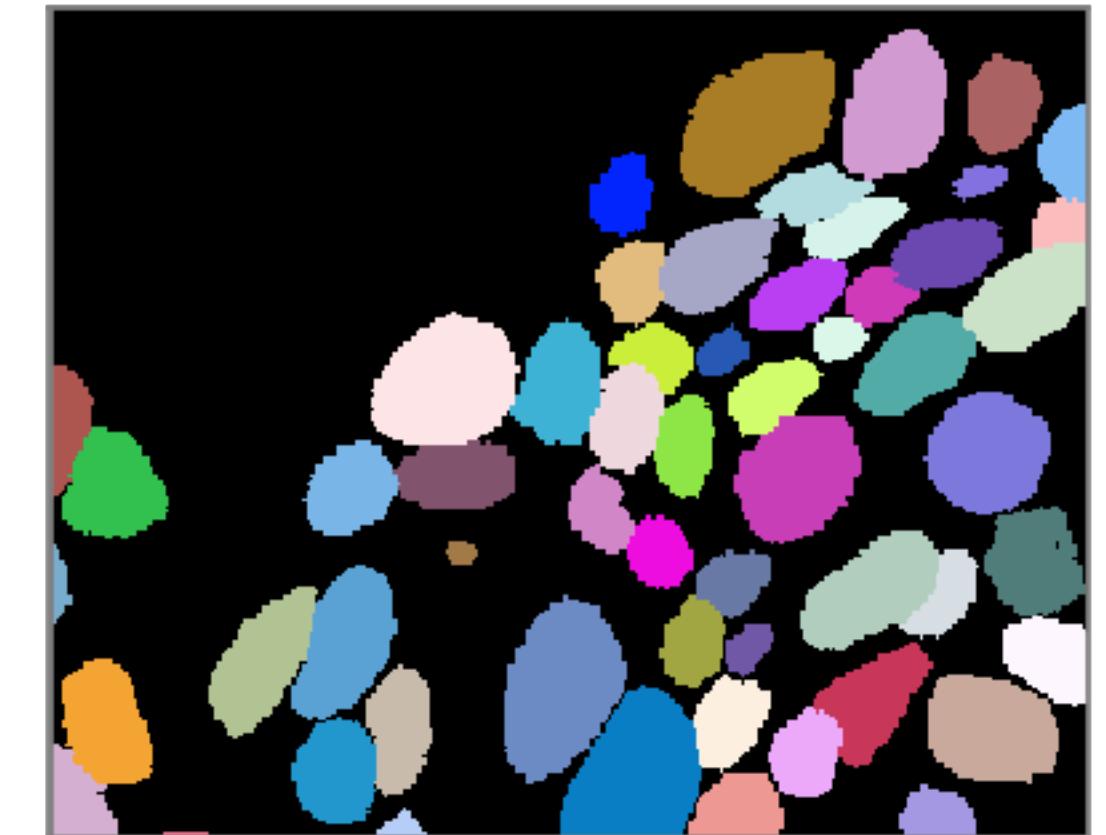


Elastic deformation

Elastic



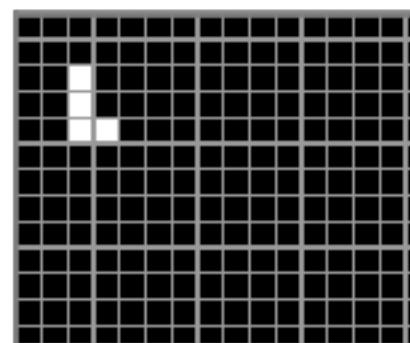
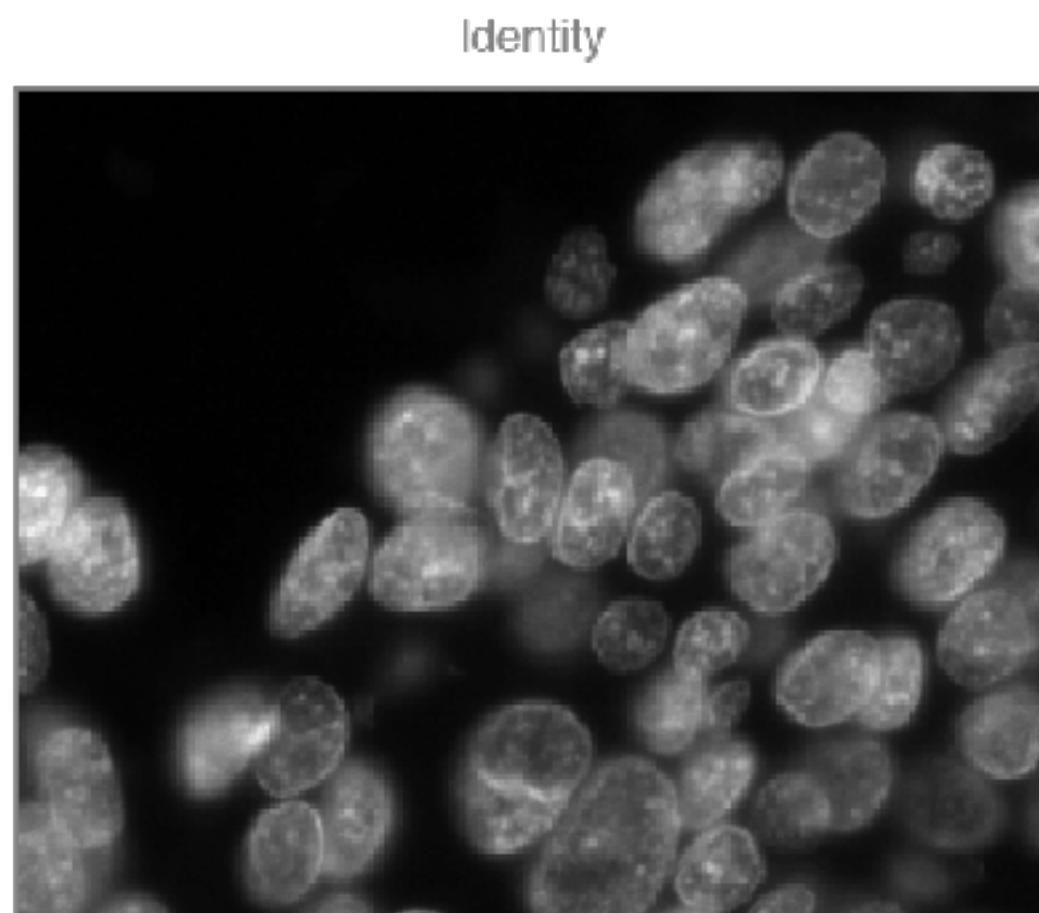
Elastic



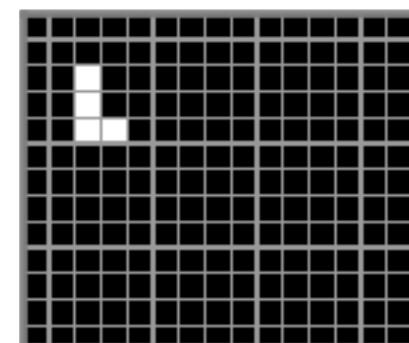
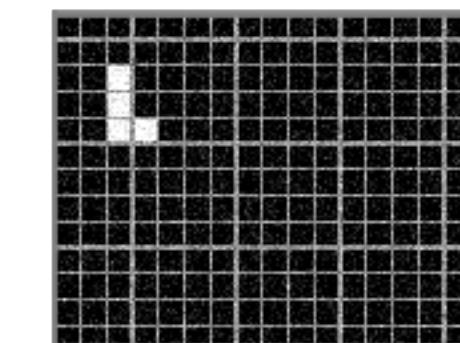
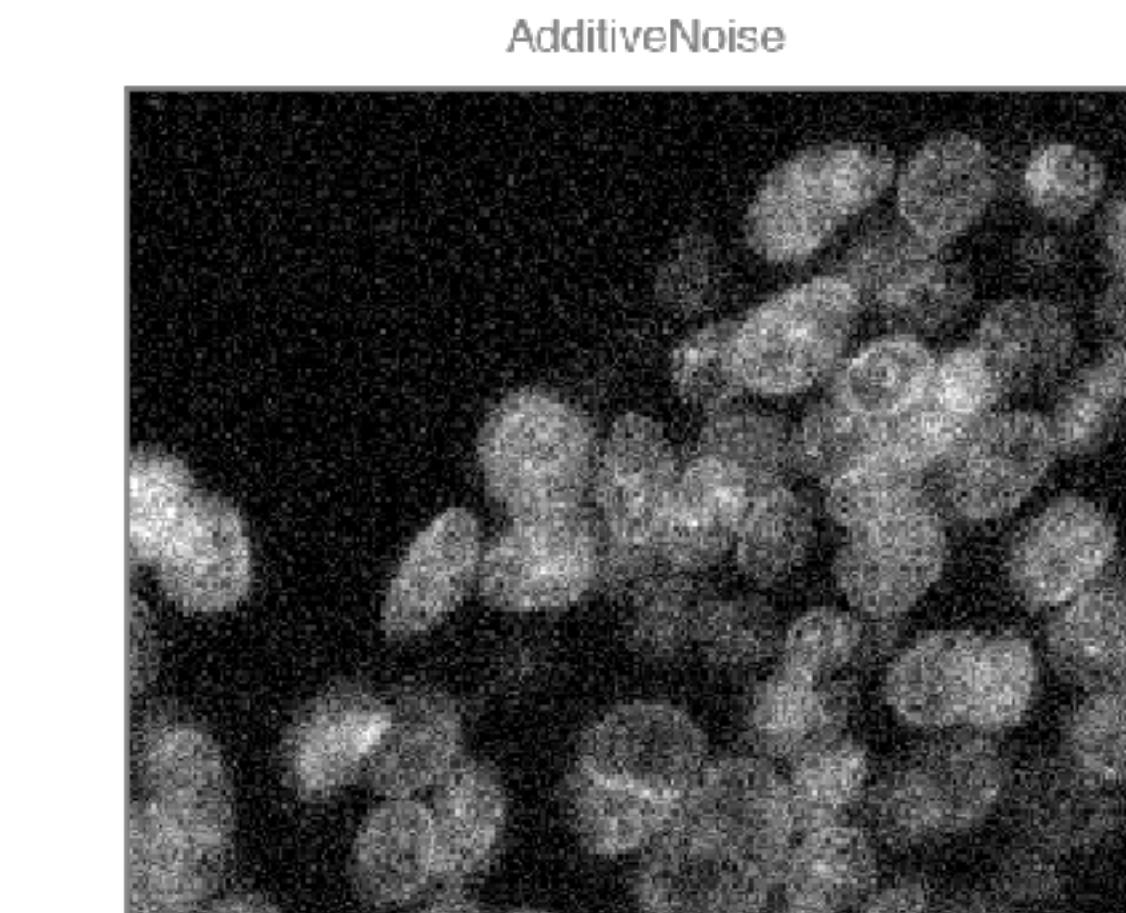
Data Augmentation

Artificially create more training data by transforming existing images/masks into different, yet plausible versions

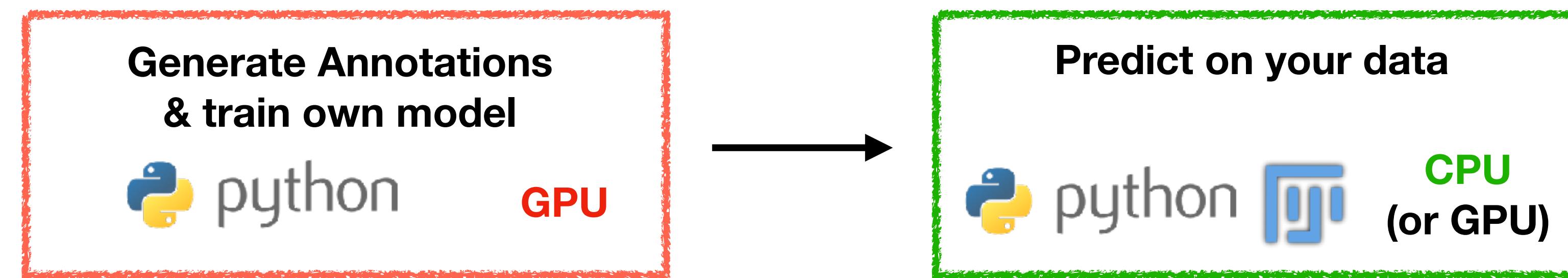
Original



Noise/Intensity shift



Demo: Training of custom models (python)



Webinar Demo on Google Colab
(Click to try it out!)

Where to ask questions / get help?

Github project issues page

- Technical questions
- Bugs, unexpected behavior
- Missing functionality

<https://github.com/mpicbg-csbd/stardist/issues>

The screenshot shows the GitHub project issues page for mpicbg-csbd/stardist. The URL in the address bar is https://github.com/mpicbg-csbd/stardist/issues. The page has a dark theme. At the top, there are navigation links for Pull requests, Issues, Marketplace, and Explore. Below that, the repository name mpicbg-csbd / stardist is displayed. The main content area shows a list of issues. A filter bar at the top of the list allows filtering by 'Used by' (1), 'Code' (0), 'Issues' (13), 'Pull requests' (0), 'Actions' (0), 'Projects' (0), and 'Wiki' (0). A search bar below the filter bar contains the query 'is:issue is:open'. The list of issues includes:

- 13 Open (checkbox)
- 28 Closed (checkbox)
- Author dropdown
- Label dropdown
- Pagination dropdown

One specific issue is highlighted: #46: Cannot find pre-trained model "versatile" from Fiji in '.h5` format. It was opened 6 days ago by VolkerH. The issue description is: "Cannot find pre-trained model "versatile" from Fiji in '.h5` format".

Image.sc forum

- Usage questions
- Best practices
- Problems with training/data

[https://forum.image.sc \(use tag “stardist”!\)](https://forum.image.sc/t/batch-processing-time-series-with-stardist-in-fiji/36507)

The screenshot shows a forum thread titled 'Batch Processing Time Series with StarDist in Fiji' on the image.sc platform. The URL in the address bar is https://forum.image.sc/t/batch-processing-time-series-with-stardist-in-fiji/36507. The page has a light theme. At the top, there is a header with the image.sc logo and the title 'Batch Processing Time Series with StarDist in Fiji'. Below the header, there is a navigation bar with a 'Usage & Issues' tab selected, showing 'fiji, stardist' under it. The main content area shows a comment from a user named Cameron.Nowell, whose profile picture is a pink circle with a white letter 'C'. The comment was posted 10 days ago. The text of the comment is:

Hi All,
I am finally getting around to play with StarDist in earnest and have hit a snag in batch processing in Fiji with it.
The data is 3 channel time series of celltrack/PI/AnnexinV for looking at apoptosis. We have a Fiji macro that has served us well over the years but StarDist is much faster and more robust at pulling out the nuclei.
I am hitting a few issues when trying to batch process it however.
I am using Stardist with default settings and if I run without activating batch mode it all works fine with Label image output or ROI manager output.

StarDist in a core facility

Questions & Answers 2