

## Max Wilkinson

Assistant Member, Structural Biology Program  
Sloan Kettering Institute  
Memorial Sloan Kettering Cancer Center  
New York, NY 10065  
United States of America

Phone: +1 857 867 0043  
Email: [wilkinm@mskcc.org](mailto:wilkinm@mskcc.org)  
Citizenship: New Zealand

## Education

**University of Cambridge**, Cambridge, UK 2015 – 2019  
PhD in Biological Sciences at the **MRC Laboratory of Molecular Biology**  
Thesis title: *Structural studies of spliceosome activation and catalysis*.  
Supervisor: Kiyoshi Nagai  
Degree awarded November 30, 2019.

**University of Otago**, Dunedin, New Zealand 2011 – 2014  
BSc (Hons) majoring in Biochemistry, minoring in Chemistry  
First Class Honours (ranked 1<sup>st</sup> amongst all university undergraduates)  
Honours supervisors: Peter Fineran and Kurt Krause

## Positions

**Massachusetts Institute of Technology | Broad Institute** 2021 – present  
HHMI Helen Hay Whitney Postdoctoral Fellow  
Supervisor: Feng Zhang

**MRC Laboratory of Molecular Biology**, Cambridge, UK 2019 – 2021  
Career Development Fellow

## Publications

### Highlighted papers (\* = equal contributions)

1. Faure, G.\*, Saito, M.\*, **Wilkinson, M.E.\***, Quinones-Olvera, N., Xu, P., Flam-Shepherd, D., Kim, S., Reddy, N., Zhu, S., Evgeniou, L., Koonin, E.V., Macrae, R.K., Zhang, F. (2025) TIGR-Tas: A family of modular RNA-guided DNA-targeting systems in prokaryotes and their viruses. *Science* eadv9789.
2. **Wilkinson, M.E.**, Li, D., Gao, A., Macrae, R.K., Zhang, F. (2024) Phage-triggered reverse transcription assembles a toxic repetitive gene from a noncoding RNA. *Science* eadq3977.
3. **Wilkinson, M.E.**, Frangieh, C.J., Macrae, R.K., Zhang, F. (2023) Structure of the R2 non-LTR retrotransposon initiating target-primed reverse transcription. *Science* **380**, 301 – 308.
4. Gao, L.A.\*, **Wilkinson, M.E.\***, Strecker, J.\*, Makarova, K., Macrae, R.K., Koonin, E.V., Zhang, F. (2022) Prokaryotic innate immunity through pattern recognition of conserved viral proteins. *Science* **377**, eabm4096.
5. **Wilkinson, M.E.\***, Fica, S.M.\*, Galej, W.P.\*, Nagai, K. (2021) Structural basis for conformational equilibrium of the catalytic spliceosome. *Mol. Cell* **81**, 1358-1362.
6. **Wilkinson, M.E.\***, Charenton, C.\*, Nagai, K.\* (2020) RNA Splicing by the Spliceosome. *Annu. Rev. Biochem.* **89**.
7. Charenton, C.\*, **Wilkinson, M.E.\***, Nagai, K. (2019) Mechanism of 5' splice site transfer for human spliceosome activation. *Science* **364**, 362–367.
8. **Wilkinson, M.E.**, Fica, S.M., Galej, W.P., Norman, C.M., Newman, A.J., Nagai, K. (2017) Postcatalytic spliceosome structure reveals mechanism of 3'-splice site selection. *Science* **358**, 1283–1288.
9. Galej, W.P., **Wilkinson, M.E.**, Fica, S.M., Oubridge, C., Newman, A.J., Nagai, K. (2016) Cryo-EM structure of the spliceosome immediately after branching. *Nature* **537**, 197–201.

### Other papers

10. Edmonds, K.K., **Wilkinson, M.E.**, Strebing, D., Chen, H., Lash, B., Schaefer, C.C., Zhu, S., Liu, D., Zilberzwige-Tal, S., Ladha, A., Walsh, M.L., Frangieh, C.J., Vaz Reay, N.A., Macrae, R.K., Wang, X.,

- Zhang, F. (2025) Structure and biochemistry-guided engineering of an all-RNA system for DNA insertion with R2 retrotransposons. *Nature Communications* **16**, 6079.
11. Zilberzwige-Tal., S., Altae-Tran, H., Kannan, S., **Wilkinson, M.E.**, Vo., S.C-D-T., Strebinger, D., Edmonds, K.H.K., Yao, C-C.J., Mears, K.S., Shmakov, S.A., Makarova, K.S., Macrae, R.K., Koonin, E.V., Zhang, F. (2025) Reprogrammable RNA-targeting CRISPR systems evolved from RNA toxin-antitoxins. *Cell* **188**, 1–16.
  12. Senn, K., Lipinski, K., Zeps, N.J., Griffin, A.F., **Wilkinson, M.E.**, Hoskins, A. (2024) Control of 3'-splice site selection by the yeast splicing factor Fyv6. *eLife* **13**, RP100449.
  13. Xu, P., Saito, M., Faure, G., Maguire, S., Vo, S.C.D.R., **Wilkinson, M.E.**, Kuang, H., Wang, B., Rice, W.J., Macrae, R.K., Zhang, F. (2024) Structural insights into the diversity and DNA cleavage mechanism of Fanzor. *Cell* **187**, 5238–5252.
  14. Birkholz, N., Kamata, K., Feussner, M., **Wilkinson, M.E.**, Samaniego, C.C., Migur, A., Kimanius, D., Ceelen, M., Went, S.C., Usher, B., Blower, T.R., Brown, C.M., Beisel, C.L., Weinberg, Z., Fagerlund, R.D., Jackson, S.A., Fineran, P.C. (2024) Phage anti-CRISPR control by an RNA-and DNA-binding helix-turn-helix protein. *Nature* **631**, 670–677.
  15. Frangieh, C.J., **Wilkinson, M.E.**, Strebinger, D., Strecker, J., Walsh, M.L., Faure, G., Yushenova, I., Macrae, R.K., Arkhipova, I.R., Zhang, F. (2024) Internal initiation of reverse transcription in a Penelope-like retrotransposon. *Mobile DNA* **15**, 12.
  16. Madigan, V., Zhang, Y., Raghavan, R., **Wilkinson, M.E.**, Faure, G., Puccio, E., Segel, M., Lash, B., Macrae, R.K., Zhang, F. (2024) Human paraneoplastic antigen Ma2 (PNMA2) forms icosahedral capsids that can be engineered for mRNA delivery. *Proc. Natl. Acad. Sci. U. S. A.* **121**, e2307812120.
  17. Garcia-Rodriguez, F.M., Martinez-Abarca, F., **Wilkinson, M.E.**, Toro, N. (2024) Similar mechanisms of retron-mediated anti-phage defense for different families of tailed phages. *bioRxiv* 2024.02.09.579579
  18. Kimanius, D., Jamali, K., **Wilkinson, M.E.**, Lövestam, S., Velazhahan, V., Nakane, T., Scheres, S.H.W. (2024) Data-drive regularisation lowers the size barrier of cryo-EM structure determination. *Nature Methods* **21**, 1216–1221.
  19. Strecker, J., Demircioglu, F.E., Li, D., Faure, G., **Wilkinson, M.E.**, Gootenberg, J.S., Abudayyeh, O.O., Nishimasu, H., Macrae, R.K., Zhang, F. (2022) RNA-activated protein cleavage with a CRISPR-associated endopeptidase. *Science* **378**, 874 – 881.
  20. Hirano, S., Kappel, K., Altae-Tran, H., Faure, G., **Wilkinson, M.E.**, Kannan, S., Demircioglu, F.E., Yan, R., Shiozaki, M., Yu, Z., Makarova, K., Koonin, E.V., Macrae, R.K., Zhang, F. (2022) Structure of the OMEGA nickase IsrB in complex with  $\omega$ RNA and target DNA. *Nature* **610**, 575 – 581.
  21. **Wilkinson, M.E.\***, Kumar, A.\*, Casañal, A. (2019) Methods for merging data sets in electron cryo-microscopy. *Acta Crystallogr. D* **75**, 782–791.
  22. Fica, S.M., Oubridge, C., **Wilkinson, M.E.**, Newman, A.J., Nagai, K. (2019) A human postcatalytic spliceosome structure reveals essential roles of metazoan factors for exon ligation. *Science* **363**, 710–714.
  23. **Wilkinson, M.E.\***, Lin, P.C.\*, Plaschka, C.\*, Nagai, K. (2018) Cryo-EM studies of pre-mRNA splicing: from sample preparation to model visualization. *Annu. Rev. Biophys.* **47**, 175–199.
  24. Fagerlund, R.D.\*, **Wilkinson, M.E.\***, Klykov, O.\*, Barendregt, A., Pearce, F.G., Kieper, S.N., Maxwell, H.W.R., Capolupo, A., Heck, A.J.R., Krause, K.L., Bostina, M., Scheltema, R.A., Staals, R.H.J., Fineran, P.C. (2017) Spacer capture and integration by a type I-F Cas1-Cas2-3 CRISPR adaptation complex. *Proc. Natl. Acad. Sci. U. S. A.* **114**, E5122–E5128.
  25. Fica, S.M., Oubridge, C., Galej, W.P., **Wilkinson, M.E.**, Bai, X.C., Newman, A.J., Nagai, K. (2017) Structure of a spliceosome remodeled for exon ligation. *Nature* **542**, 377–380.
  26. **Wilkinson, M.E.**, Nakatani, Y., Staals, R.H., Kieper, S.N., Opel-Reading, H.K., McKenzie, R.E., Fineran, P.C., Krause, K.L. (2016) Structural plasticity and in vivo activity of Cas1 from the type I-F CRISPR-Cas system. *Biochem. J.* **473**, 1063–1072.

## Fellowships, scholarships, and awards

HHMI Helen Hay Whitney Fellowship <i>postdoctoral funding</i>	2022 –
RNA Society/Scaringe Young Scientist Award <i>worldwide, for significant contributions to RNA research</i>	2019
Max Perutz Student Prize <i>for outstanding PhD work at the MRC LMB</i>	2018
Trinity College Krishnan-Ang Studentship	2015
Cambridge-Rutherford Memorial Scholarship <i>full PhD funding for three New Zealanders at the University of Cambridge</i>	2014
Prince of Wales Prize <i>for top undergraduate student at the University of Otago</i>	2014
Edson Prize in Biochemistry (Fourth Year)	2014
Sir George Grey Senior Scholarship for the top science graduate	2013
Edson Prize in Biochemistry (Third Year)	2013
F G Soper Prize in Chemistry	2012
New Zealand Institute of Chemistry prize	2011
University of Otago Leaders of Tomorrow Scholarship	2011
New Zealand Qualifications Authority Premier Scholarship <i>for top ten high school students nationwide</i>	2010
Papanui High School Dux	2010

## Conference presentations

2024 Symposium on the Immune System of Bacteria, 16–18 April 2024. *Selected talk*  
6<sup>th</sup> International Conference on CRISPR Technologies, 17–19 October 2023. *Invited talk*  
Microsymposium on RNA Biology, Vienna, Austria, 3–5 May 2023. *Oral presentation*  
IUBMB/FEBS/PABMB Global Biochemistry Summit, Lisbon, Portugal, 9–14 July 2022. *Oral presentation*  
British Crystallographic Association Spring Meeting, Leeds, 11–14 April 2022. *Invited talk*  
Cryo-electron microscopy in structural biology, Rome, Italy, 10–11 October 2019. *Invited talk*  
24<sup>th</sup> Annual Meeting of the RNA Society, Krakow, Poland, 11–15 June 2019. *Invited talk*  
84<sup>th</sup> Cold Spring Harbor Symposium on Quantitative Biology: RNA Control & Regulation, Cold Spring Harbor, NY, USA, 29 May to 3 June 2019. *Oral presentation*  
6<sup>th</sup> UK RNA Splicing Workshop, Rydal Hall, Cumbria, UK, 25–27 January 2019. *Oral presentation*  
14<sup>th</sup> Nucleic Acids Forum, Royal Society of Chemistry, London, UK, 6 July 2018. *Invited talk*  
23<sup>rd</sup> Annual Meeting of the RNA Society, Berkeley, USA, 29 May to 2 June 2018. *Poster presentation*  
1<sup>st</sup> BioPharma Expo, Tokyo, Japan, 28–30 June 2017. *Invited talk*  
EMBO Conference – Molecular Machines: Integrative Structural and Molecular Biology, Heidelberg, Germany, 20–23 November 2016. *Oral presentation*  
21<sup>st</sup> Annual Meeting of the RNA Society, Kyoto, Japan, 28 June to 2 July 2016. *Poster presentation*  
Queenstown Molecular Biology Meeting, Queenstown, New Zealand, 29 August to 4 September 2015. *Poster presentation*

## Other invited talks

<b>Memorial Sloan Kettering Cancer Centre, USA</b> Topic: Reverse transcription	January 2025
<b>University of Michigan, USA</b> Topic: Reverse transcription	December 2024
<b>Johns Hopkins University, USA</b> Topic: Reverse transcription	September 2024
<b>IMBA, Vienna BioCenter, Vienna, Austria</b> Topic: Reverse transcription	September 2024
<b>The Rockefeller University, USA</b> Topic: LINE retrotransposons	February 2024
<b>MRC LMB, Cambridge, UK</b> Topic: LINE retrotransposons	June 2023
<b>University of Wisconsin-Madison, USA</b> Topic: LINE retrotransposons	April 2023
<b>University of Otago, Dunedin, New Zealand</b> Topic: phage defence systems	April 2022
<b>Trinity Graduate Biologists seminar</b> Topic: spliceosomes	October 2019
<b>BA Seminar, Trinity College, Cambridge</b> Topic: spliceosomes	January 2019
<b>MRC LMB Lab Talks</b> Topic: spliceosomes	October 2018
<b>SciLifeLab, Stockholm, Sweden</b> Topic: spliceosomes	September 2018
<b>MRC LMB Structural Studies Colloquium</b> Topic: spliceosomes	March 2018

<b>University of Otago, Dunedin, New Zealand</b> Topic: spliceosomes	February 2018
<b>University of Cambridge RNA club</b> Topic: spliceosomes	January 2018
<b>University of Tokyo</b> Topic: spliceosomes	June 2017
<b>First Trinity Forum</b> Topic: spliceosomes. <b>Prize</b> for best 1 minute talk.	June 2017
<b>Trinity Graduate Biologists seminar</b> Topic: spliceosomes	January 2017
<b>Osaka University</b> Topic: spliceosomes	July 2016

## Teaching and mentoring

Undergraduate Research Opportunities Programme mentor, MIT	2022 – 2025
MRC LMB student parent programme	2017 – 2018
Trinity College Cambridge student parent programme	2016 – 2017
Tutor at Carrington College, University of Otago	2012 – 2013

## References

*Prof. Feng Zhang*

Broad Institute; McGovern Institute of Brain Research, MIT

[zhangoffice@broadinstitute.org](mailto:zhangoffice@broadinstitute.org)

Phone: +1 617 714 7000

Postdoctoral Advisor

*Dr. Kelly Nguyen*

MRC Laboratory of Molecular Biology, Cambridge, UK

[knguyen@mrc-lmb.cam.ac.uk](mailto:knguyen@mrc-lmb.cam.ac.uk)

Phone: +44 1223 267190

Group leader at MRC LMB, PhD mentor, can provide reference in lieu of my deceased PhD supervisor  
Kiyoshi Nagai

*Dr. Clemens Plaschka*

Research Institute of Molecular Pathology, Vienna, Austria

[clemens.plaschka@imp.ac.at](mailto:clemens.plaschka@imp.ac.at)

Phone: +43 179730 3451

Group leader at IMP, PhD mentor, can provide reference in lieu of my deceased PhD supervisor  
Kiyoshi Nagai

*Prof. Peter Fineran*

University of Otago, Dunedin, New Zealand

[peter.fineran@otago.ac.nz](mailto:peter.fineran@otago.ac.nz)

Phone: +64 3 479 7735

Professor of Molecular Microbiology at University of Otago, undergraduate supervisor and current collaborator