**Appendix S1**

**Supporting Information for**

Optimum growth temperature declines with body size within fish species

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Contents

[Literature search, selection process and criteria 3](#_Toc87960057)

[*Maximum consumption rate* 4](#_Toc87960058)

[*Metabolic rate* 5](#_Toc87960059)

[*Growth rates & optimum temperature for growth over body mass* 5](#_Toc87960060)

[Data overview 12](#_Toc87960061)

[*Maximum consumption & metabolic rate* 12](#_Toc87960062)

[*Growth rate* 13](#_Toc87960063)

[Supplementary methods and analysis 15](#_Toc87960064)

[Model validation and fit 22](#_Toc87960065)

[*Maximum consumption rate – below peak temperatures* 22](#_Toc87960066)

[*Maximum consumption rate – including beyond peak temperatures* 26](#_Toc87960067)

[*Metabolic rate* 30](#_Toc87960068)

[*Optimum growth temperature* 35](#_Toc87960069)

[References 39](#_Toc87960070)

# Literature search, selection process and criteria

This section is an overview of the literature search approach, and below we present the search terms for each rate separately (maximum consumption, metabolism and growth). In addition to search terms, we also applied filters by selecting only the following subjects: ‘marine freshwater biology’, ‘fisheries’, ‘ecology’, ‘zoology’, ‘biology’, ‘physiology’. For growth rates, we also included ‘limnology’ and for maximum consumption we included ‘limnology’ and ‘evolutionary biology’. The use of additional subjects for growth and consumption reflects the lower data availability compared to metabolism. As we suspected that relatively few studies would have considered both body size- and temperature treatments, our goal was to get an as extensive as possible list of studies. Therefore, we also evaluated articles cited by articles found in the search, from published review-type articles and reviews of applications of bioenergetics models such as the Wisconsin model (Deslauriers *et al.* 2017), and if the study was found in the literature search for another rate. The source of the article (WoS search or cited in literature) is indicated in the data sets (Table S1).

Articles were filtered out at three levels of the search: title, abstract and full article. The online repository of this project (<https://github.com/maxlindmark/scaling>) contains .txt files of the complete list of articles found in the literature search. We removed studies from the lists if the titles made it clear the articles did not fulfil all of the following conditions: (1) experimental study, (2) fish as study organism in post-larval life stages, and (3) replicates across both body size and temperature. We treat data as individual-level rates (per fish); however, in some cases they were measured as averages across multiple individuals. In addition to these general criteria, we also had criteria specific for each rate (see below). When several studies were found for the same species, we did not include all but instead chose the study with the largest body size and temperature range (in that order), as there can be large differences in absolute values of some physiological parameters between studies.

For consumption and growth rate, we determined if each data point within species was below or beyond peak temperature for a size group within a species either by using information provided by the authors (e.g., by deriving a polynomial regression of the rate as a function of temperature to find the temperature of peak rate), by fitting Sharpe Schoolfield equations (see main text, Eq. 4) or visually inspecting data for each species separately. Whether a data point is below or above peak or optimum temperatures is indicated by a separate column in the data (Table S1).

## *Maximum consumption rate*

We used the following topic terms for maximum consumption rate (three searches in total): (consumption OR bioenerg\* OR ingestion OR “food-intake”) AND (mass OR weight OR size) AND (temperature\*), as well as: (feeding-rate OR bio-energ\*) AND (mass OR weight OR size) AND (temperature\*) and lastly: (“food intake”) AND (mass OR weight OR size) AND (temperature\*). \* represents any group of characters, including no character. The searches for maximum consumption rate data resulted in 15259 articles (search date: 18 December 2018), with 3449 remaining after filtering by subject categories. The second search (search date: 13 March 2019) resulted in 431 additional titles after filtering by subject categories (of which some where duplicated from the first search) and the third search (search date: 29 June 2020) yielded 626 but no additional articles as they had either been selected already or did not meet the criteria. Articles were filtered out at the abstract and whole article stage if the original reference could not be identified and evaluated, if data were normalized (i.e., using a priori defined scaling relationships to show corrected data rather than measured values), there was no acclimation, or if measurements were not maximum consumption rate. As with the growth data, definitions of ad-libitum feeding may differ between studies – the key for our purpose is that food rations led to satiation and were not limiting. Consumption rates were converted to . These data were compiled in the file consumption\_data.xlsx.

## *Metabolic rate*

We used the following topic terms for metabolic rate data: (metabolism OR "oxygen-consumption" OR "oxygen consumption") AND (mass OR weight OR size) AND (temperature\*). \* represents any group of characters, including no character. The search for metabolic rate experiments resulted in 8405 articles (search date: 6 June 2019), which was reduced to 3458 after applying filters for subject categories. Articles were filtered out at the abstract and whole article stage if the original reference could not be identified and evaluated, if data were normalized (i.e., using a priori defined scaling relationships to normalize data for data a given size rather than measured values), if there was no acclimation or if it was not standard, routine or resting metabolic rate. The latter was defined as oxygen consumption of an unfed fish at no or little spontaneous activity. Metabolic rates were converted to , because it was the most common unit in the data set. These data were compiled in the file metabolism\_data.xlsx.

## *Growth rates & optimum temperature for growth over body mass*

Growth rates were taken from data found in the literature search for optimum growth temperatures. Therefore, articles in which growth rates were measured at sub-optimum temperatures only were not included (note this is in contrast to consumption data where “optimum” was not included in the search terms). We used the following topic terms for growth rate data: (growth) AND (mass OR weight OR size) AND (temperature\*) AND (optimum), as well as: (growth) AND (mass OR weight OR size) AND (temperature\*) AND (optim\*). \* represents any group of characters, including no character. The two searches for growth rates resulted in 3313 articles (search date: 22 March 2019), and 3747 articles (search date: 5 May 2019), respectively. After applying additional filters by subject category, we acquired 566 and 893 studies, respectively (of which some are duplicates due to similar search-strings). We removed studies at the abstract and whole article stage where the original reference could not be identified and evaluated, if we could not extract actual growth rates, if there was not a controlled temperature for each growth trial, or if there were not multiple defined size-classes. We used only one observation (data point) per size class and temperature treatment, and in cases where there were two, we used the mean value. In addition, we ensured that no other treatment (e.g., food limitation) confounded the response variable and thus only used data from experiments with satiating food levels. Body mass is either the geometric mean of the initial and final mass of the growth trial or the size class (as reported by the original authors), depending on data availability (see Table S1). It is important to control for feeding rations as it affects the temperature optimum for growth (Brett *et al.* 1969). This was achieved in different ways in the different experimental studies, but commonly involved excess feeding rations once or several times per day. The key description we looked for in the study was that food was not limiting. We treat data as individual-level growth (per fish); however, these were commonly measured as averages for multiple individuals. In the case growth was length-based, we converted it to mass using weight-length relationships from FishBase (Froese *et al.* 2014; Froese & Pauly 2019). We compiled two separate data sets: raw growth rates (growth\_data.xlsx) and temperature at optimum growth (growth\_data\_Topt.xlsx). In the latter, we defined optimum temperature for growth as the fitted optimum temperature by size-class (usually estimated in the original study). Therefore, the optimum temperature may not always correspond to an actual experimental temperature but could be an estimation between two temperature treatments. If the optimum temperature (by size group) was not estimated in the original study, we used the temperature where growth rate was maximized. All growth rates were expressed in unit .

**Table S1** Explanation of data columns (G=growth data, Topt=optimum growth temperature data, C=maximum consumption data, M=metabolism data).

|  |  |  |
| --- | --- | --- |
| Column | Explanation | Datasets |
| *growth\_rate\_%/day* | Main response variable. | G, Topt |
| *opt\_temp\_c* | Main response variable. | Topt |
| *initial\_mass\_g* | Body mass [g] at the onset of the growth trial. | G, Topt |
| *final\_mass\_g* | Body mass [g] at the end of the growth trial. | G, Topt |
| *geom\_mean\_mass\_g* | Geometric mean mass in t1 and t2 of the growth trial. | G, Topt |
| *size\_group* | Representative body mass of size group in the growth trial, in case initial, final or geometric body mass could not be retrieved. | G, Topt |
| *consumption* | Main response variable. | C |
| *metabolic\_rate* | Main response variable. | M |
| *type* | Type of respiration measurement (resting, routine, standard). | M |
| *unit* | Unit of response variable. | C, M |
| *original\_unit* | Original unit of response variable. If different from “***unit***”, see “***notes***” column for information on conversion. | C, M |
| *mass\_g* | Body mass in experiment [g]. Some studies report body masses before and some after the feeding trials. See “***notes***”. | C, M |
| *temp\_c* | Experimental temperature []. | G, C, M |
| *above\_peak\_temp* | Is the experiment conduced at temperature above peak temperature for the given size group? Y/N. | G, C, M |
| *common\_name* | Common name of species. | G, Topt, C, M |
| *species* | Scientific name of species. | G, Topt, C, M |
| *genus* | Genus of species. | G, Topt, C, M |
| *family* | Family of species. | G, Topt, C, M |
| *order* | Order of species. | G, Topt, C, M |
| *habitat* | Species natural habitat, taken from FishBase (Froese & Pauly 2019). | G, Topt, C, M |
| *lifestyle* | Lifestyle of species, taken from FishBase (Froese & Pauly 2019). | G, Topt, C, M |
| *biogeography* | Biogeography of species, taken from FishBase (Froese & Pauly 2019). | G, Topt, C, M |
| *trophic\_level* | Trophic level of species, taken from FishBase (Froese & Pauly 2019). | G, Topt, C, M |
| *w\_maturation\_g* | Body mass [g] at maturation taken from FishBase (Froese & Pauly 2019). If not available, weight was estimated from length using species-specific allometric weight-length, else taken from alternative sources (see ”***notes***”). Used to estimate relative body size across species in the data and to normalized optimum growth temperatures across species. | G, Topt |
| *w\_max\_published\_g* | Max. published weight [g] taken from FishBase (Froese & Pauly 2019). If not available, weight was estimated from length using species-specific allometric weight-length, else taken from alternative sources (see ”***notes***”). Used to estimate relative body size across species in the data. | G, Topt, C, M |
| *env\_temp\_min* | Min. environmental temperature [], taken from FishBase (Froese & Pauly 2019). If not available on FishBase, data were taken from alternative sources (see ”***notes***”). Used to compare experimental temperatures to common temperatures for species. | G, Topt, C, M |
| *env\_temp\_max* | Max. environmental temperature [], taken from FishBase (Froese & Pauly 2019). If not available on FishBase, data were taken from alternative sources (see ”***notes***”). Used to compare experimental temperatures to common temperatures for species. | G, Topt, C, M |
| *env\_temp\_mid* | Median of environmental temperature [], taken from FishBase (Froese & Pauly 2019). If not available on FishBase, data were taken from alternative sources (see ”***notes***”). Used to compare experimental temperatures to common temperatures for species. | G, Topt, C, M |
| *pref\_temp\_mid* | Median of preferred temperature [], taken from FishBase (Froese & Pauly 2019). If not available on FishBase, data were taken from alternative sources (see ”***notes***”). Used to compare experimental temperatures to common temperatures for species. | G, Topt, C, M |
| *notes* | This column contains additional information, including if data were sent by authors, if any column above has data that is not from the main source (i.e. FishBase), how certain metrics were calculated, alternative common names, comments on the experimental protocol, information on conversion to standard “***unit***”, source of the data (literature search or cited in paper from literature search) | G, Topt, C, M |
| *reference* | Source (See Table S2). | G, Topt, C, M |

**Table S2** Species, common name, the data set(s) in which they appear and the sources (G=growth data, Topt=optimum growth temperature data, C=maximum consumption data, M=metabolism data). If more than one data and source, the sources are in order (1 study per species and rate).

|  |  |  |  |
| --- | --- | --- | --- |
| Species | Common name | Datasets | Source |
| *Pseudopleuronectes yokohamae* | Marbled flounder | G, Topt, C | (Tomiyama *et al.* 2018) |
| *Cyclopterus lumpus* | Lumpfish | G, Topt | (Nytrø *et al.* 2014) |
| *Paralichthys olivaceus* | Japanese flounder (alt. bastard halibut, Japaneese halibut or Olive flounder) | G, Topt, C | (Iwata *et al.* 1994) |
| *Salvelinus alpinus* | Arctic char | G, Topt | (Siikavuopio *et al.* 2013) |
| *Salmo salar* | Atlantic salmon | G, Topt | (Handeland *et al.* 2008) |
| *Lates calcarifer* | Barramundi | G, Topt, C, M | (Bermudes *et al.* 2010)  (Bermudes *et al.* 2010)  (Bermudes *et al.* 2010)  (Glencross & Felsing 2006) |
| *Gadus morhua* | Atlantic cod | G, Topt, M | (Björnsson *et al.* 2007)(Tirsgaard *et al.* 2015) |
| *Hippoglossus hippoglossus* | Atlantic halibut | G, Topt | (Björnsson & Tryggvadóttir 1996) |
| *Scophthalmus maximus* | Turbot | G, Topt | (Árnason *et al.* 2009) |
| *Boreogadus saida* | Arctic cod | G, Topt | (Laurel *et al.* 2017) |
| *Rachycentron canadum* | Cobia | G, Topt, C | (Sun & Chen 2014) |
| *Pelteobagrus fulvidraco* | Yellow catfish | G, Topt,C | (Zhang *et al.* 2017) |
| *Anarhichas minor* | Spotted wolffish | G, Topt | (Imsland *et al.* 2006) |
| *Oncorhynchus mykiss* | Rainbow trout | C, M | (From & Rasmussen 1984) |
| *Perca fluviatilis* | Eurasian perch | C | (Lessmark 1983) |
| *Phoxinus phoxinus* | Eurasian minnow | C, M | (Cui & Wootton 1988) |
| *Coregonus hoyi* | Bloater | C | (Binkowski & Rudstam 1994) |
| *Pomoxis annularis* | White crappie | C | (Hayward & Arnold 1996) |
| *Gambusia affinis* | Western mosquitofish | C | (Chipps & Wahl 2004) |
| *Morone saxatilis* | Striped bass | C | (Duston *et al.* 2004) |
| *Salvelinus fontinalis* | Brook trout | C, M | (Baldwin 1957) (Beamish 1964) |
| *Leuciscus leuciscus* | Dace | C | (Marmulla & Rosch 1990) |
| *Lepomis microlophus* | Redear sunfish | C | (Wang *et al.* 2003) |
| *Channa argus* | Chinese snakehead (alt. Northern snakehead or Snakehead) | C, M | (Liu *et al.* 1998)  (Liu *et al.* 2000) |
| *Siniperca chuatsi* | Mandarin fish | C, M | (Liu *et al.* 1998)  (Liu *et al.* 2000) |
| *Gasterosteus aculeatus* | Three-spined stickleback | C | (Wootton *et al.* 1980) |
| *Salmo trutta* | Brown trout | C | (Elliott 1976) |
| *Epinephelus coioides* | Orange-spotted grouper | C | (Lin *et al.* 2008) |
| *Coregonus albula* | Vendace | M | (Ohlberger *et al.* 2012) |
| *Coregonus fontanae* | Stechlin cisco | M | (Ohlberger *et al.* 2012) |
| *Abramis brama* | Common bream | M | (Ohlberger *et al.* 2012) |
| *Rutilus rutilus* | Common roach | M | (Ohlberger *et al.* 2012) |
| *Salvelinus confluentus* | Bull trout | M | (Mesa *et al.* 2013) |
| *Catostomus commersonii* | White sucker | M | (Beamish 1964) |
| *Cyprinus carpio* | Common carp | M | (Beamish 1964) |
| *Ameiurus nebulosus* | Brown bullhead | M | (Beamish 1964) |
| *Silurus meridionalis* | Southern catfish | M | (Xie & Sun 1990) |
| *Carassius auratus* | Goldfish | M | (Beamish & Mookherjii 1964) |
| *Pomadasys commersonnii* | Spotted grunter | M | (Du Perez *et al.* 1986) |
| *Melanogrammus aeglefinus* | Haddock | M | (Peck *et al.* 2005) |
| *Centropristis striata* | Black sea bass | M | (Slesinger *et al.* 2019) |
| *Anguilla anguilla* | European eel | M | (Degani *et al.* 1989) |
| *Micropterus salmoides* | Largemouth bass | M | (Glover *et al.* 2012) |
| *Cyprinodon macularius* | Desert pupfish | M | (Heuton *et al.* 2018) |
| *Micropogonias undulatus* | Atlantic croaker | M | (Horodysky *et al.* 2011) |
| *Leiostomus xanthurus* | Spot | M | (Horodysky *et al.* 2011) |
| *Coreius guichenoti* | Largemouth bronze gudgeon | M | (Luo & Wang 2012) |
| *Sprattus sprattus* | European sprat | M | (Meskendahl *et al.* 2010) |
| *Plectropomus leopardus* | Leopard coral grouper | M | (Messmer *et al.* 2017) |
| *Galaxias maculatus* | Common galaxias | M | (Milano *et al.* 2016) |
| *Polyodon spathula* | American paddlefish (alt. Mississippi paddlefish) | M | (Patterson *et al.* 2013) |
| *Argyrosomus japonicus* | Mulloway | M | (Pirozzi & Booth 2009) |
| *Lythrypnus dalli* | Bluebanded goby | M | (Rangel & Johnson 2018) |
| *Colossoma macropomum* | Tambaqui (alt. Cachama) | M | (Tomala *et al.* 2014) |
| *Carassius auratus grandoculis* | Round crucian carp (alt. Nigorobuna) | M | (Yamanaka *et al.* 2013) |

# Data overview

## *Maximum consumption & metabolic rate*

A picture containing table

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*Fig. S1. Experimental temperatures (gray) and environmental (min, median and max) temperatures (purple, pink and green, respectively) of species represented in the consumption (left) and metabolism (right) data sets. Missing temperatures means information was not available on FishBase. Experimental temperatures are jittered vertically for visibility.*

## *Growth rate*

Chart, bar chart

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*Fig. S2. The distribution of rescaled masses for individual observations (mass/mass at maturation), where color indicate species.*

Graphical user interface, text, application

Description automatically generated

*Fig. S3. Experimental temperatures (gray) in the growth rate data and environmental (min, median and max) temperatures (purple, pink and green, respectively). Missing temperatures means information was not available on FishBase. Experimental temperatures are jittered vertically for visibility.*

# Supplementary methods and analysis

**Table S3** Description of model parameters (type and their interpretation in brackets) and their prior distributions (see ‘*Model description’* and equations 1-3 in the main text). refers to a normal distribution (mean and standard deviation, s.d.) and to a uniform distribution (interval). For simplicity, only the parameters of the full model are shown here (i.e., with most coefficients varying by species), but note that when a model is fitted with a common rather than species-varying coefficient, for example instead of , we use the same prior for as for .

|  |  |  |  |
| --- | --- | --- | --- |
| Model | Parameter | Description | Prior distribution |
| Log-linear regressions  for consumption and metabolism |  | Hyperparameter (average intercept for standard metabolic rate across species). *Only for the metabolism models.* |  |
|  | Hyperparameter (average intercept for routine and resting metabolic rate across species). *Only for the metabolism models.* |  |
|  | Hyperparameter (average intercept across species). *Only for the consumption models.* |  |
|  | Hyperparameter (average mass coefficient across species). |  |
|  | Hyperparameter (average temperature coefficient across species) |  |
|  | Hyperparameter (average interaction coefficient across species) |  |
|  | Hyperparameter (s.d. of species-intercepts for standard metabolic rate) |  |
|  | Hyperparameter (s.d. of species-intercepts for routine and resting metabolic rate) |  |
|  | Hyperparameter (s.d. of species mass coefficients) |  |
|  | Hyperparameter (s.d. of species temperature coefficients) |  |
|  | Hyperparameter (s.d. of species interaction coefficients) |  |
|  | Parameter (s.d.) |  |
| Sharpe-Schoolfield (unimodal consumption data) |  | Hyperparameter (average consumption at reference temperature [-10 on centered scale] across species) |  |
|  | Hyperparameter (average activation energy across species) |  |
|  | Parameter (common rate of decline with temperature) |  |
|  | Parameter (common temperature at which half the rate is reduced due to high temperatures) |  |
|  | Hyperparameter (s.d. of species-varying activation energies) |  |
|  | Hyperparameter (s.d. of species-varying average consumption) |  |
|  | Parameter (s.d.) |  |
| Linear  models |  | Hyperparameter (average intercept across species) |  |
|  | Hyperparameter (average mass coefficient across species) |  |
|  | Hyperparameter (s.d. of species-intercepts) |  |
|  | Hyperparameter (s.d. of species mass coefficients) |  |
|  | Parameter (s.d.) |  |

**Table S4**. Model comparison for the log-linear regressions of how consumption and metabolism depend on mass and temperature below optimum temperatures (see ‘*Model description’* and equations 1-3 in the main text). The column m\*t indicates whether the model for the rate includes an interactive effect of mass and temperature. The models differ in which coefficients vary among species and which are common, where is the intercept, mass coefficient (mass-exponent on linear scale), temperature coefficient (corresponding to the negative activation energy) and interaction between mass and temperature. The WAIC columns shows WAIC and absolute WAIC in brackets, rounded to the nearest decimal, where WAIC is the difference between each model’s WAIC and the lowest WAIC across models. Bold indicates models with WAIC < 2.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Model | m\*t | Species-varying parameter(s) | WAIC  metabolism | WAIC consumption |
| M1 | Yes |  | **0 (274.6)** | 3.1 (563.7) |
| M2 |  | **0.3 (274.9)** | 2 (562.6) |
| M3a |  | 305.7 (580.3) | 147.1 (707.7) |
| M3b |  | 385.3 (659.9) | 70.5 (631.1) |
| M4 |  | 648.6 (923.2) | 190 (750.6) |
| M5 | No |  | 6.1 (280.6) | **0 (560.6)** |
| M6a |  | 348.2 (622.8) | 165.8 (726.4) |
| M6b |  | 386.6 (661.2) | 73.4 (634) |
| M7 |  | 681.5 (956.1) | 213 (773.6) |

**Table S5**. Comparison of the two models fitted to optimum growth temperature data. The WAIC columns shows WAIC and absolute WAIC in brackets, rounded to the nearest decimal, where WAIC is the difference between each models’ WAIC and the lowest WAIC across models. Bold indicates models with WAIC < 2.

|  |  |  |
| --- | --- | --- |
| Model | Species-varying parameter(s) | WAIC |
| M1 |  | **0 (177.3)** |
| M2 |  | **1 (178.3)** |

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*Fig. S4. Posterior distributions of the global intraspecific mass exponents () and temperature coefficients () for metabolic rate (top) and maximum consumption rate (bottom). For metabolism, the global interaction coefficient () is also shown (estimated and presented on an Arrhenius temperature scale), but for consumption this term was not included in the selected model. Numbers in the top left corner correspond to the posterior median. The axes are the same for each parameter for comparison between the two rates.*

***A screenshot of text

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*Fig. S5. Experimental temperatures (grey) overlap environmental temperatures (green), and optimum growth temperatures (orange) are typically at the upper end or above the environmental range. Horizontal green lines show the minimum and maximum environmental temperature based on either temperature in distribution range (triangles) or modelled distribution maps (circles), both taken from FishBase. The optimum growth temperatures are depicted for all size-classes per species, where the circle size is proportional to number of observations at that temperature.*

*Diagram

Description automatically generated*

*Fig. S6. Posterior median of species-level intercepts (points) and their 95% credible interval (horizontal error bars). Colors indicate the type of metabolism measurement for each species. Vertical solid lines are the posterior medians of the global intercepts (orange for standard metabolic rate, , and green for routine or resting metabolic rate, ), and the dashed vertical lines show the 95% credible intervals for the global parameters.*

# Model validation and fit

Figures showing convergence of species-level parameters can be found on: <https://github.com/maxlindmark/scaling>, in this section only global parameters are visualized.

## *Maximum consumption rate – below peak temperatures*

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*Fig. S7. Posterior densities and trace plots for evaluation of chain convergence (by chain, indicated by color), for the global-level parameters for the log-linear maximum consumption rate model at temperatures below peak temperatures.*

Chart, scatter chart

Description automatically generated

*Fig. S8. Potential scale reduction factor () for the log-linear maximum consumption rate model. This factor is based on the comparison of between and within-chain variation for the same parameter. A value close to one implies chains converged to the same distribution. The index of the parameter corresponds to species in alphabetical order.*

Chart

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*Fig. S9. A) Model fit (mean) for the log-linear model of maximum consumption rate at temperatures below temperature peak (by species). Fit is evaluated by simulating data from the likelihood (at each iteration of the MCMC chain), to compare how well it matches the original data. Each simulated data point is assigned a 0 or 1 if it is below or above the mean data point (the vertical line corresponds to the mean in data). The number in the plot corresponds to the mean of the vector of 0’s and 1’s. B) Posterior predictive distribution (orange) and distribution of data (purple). C) Difference between the observed value and the posterior median of the predicted value, plotted against fitted value.*

Chart

Description automatically generated

*Fig. S10. Posterior (black) and prior distribution (red) for the global parameters in the log-linear model for maximum consumption rate, including their % overlap and effective sample size (n.eff).*

## *Maximum consumption rate – including beyond peak temperatures*

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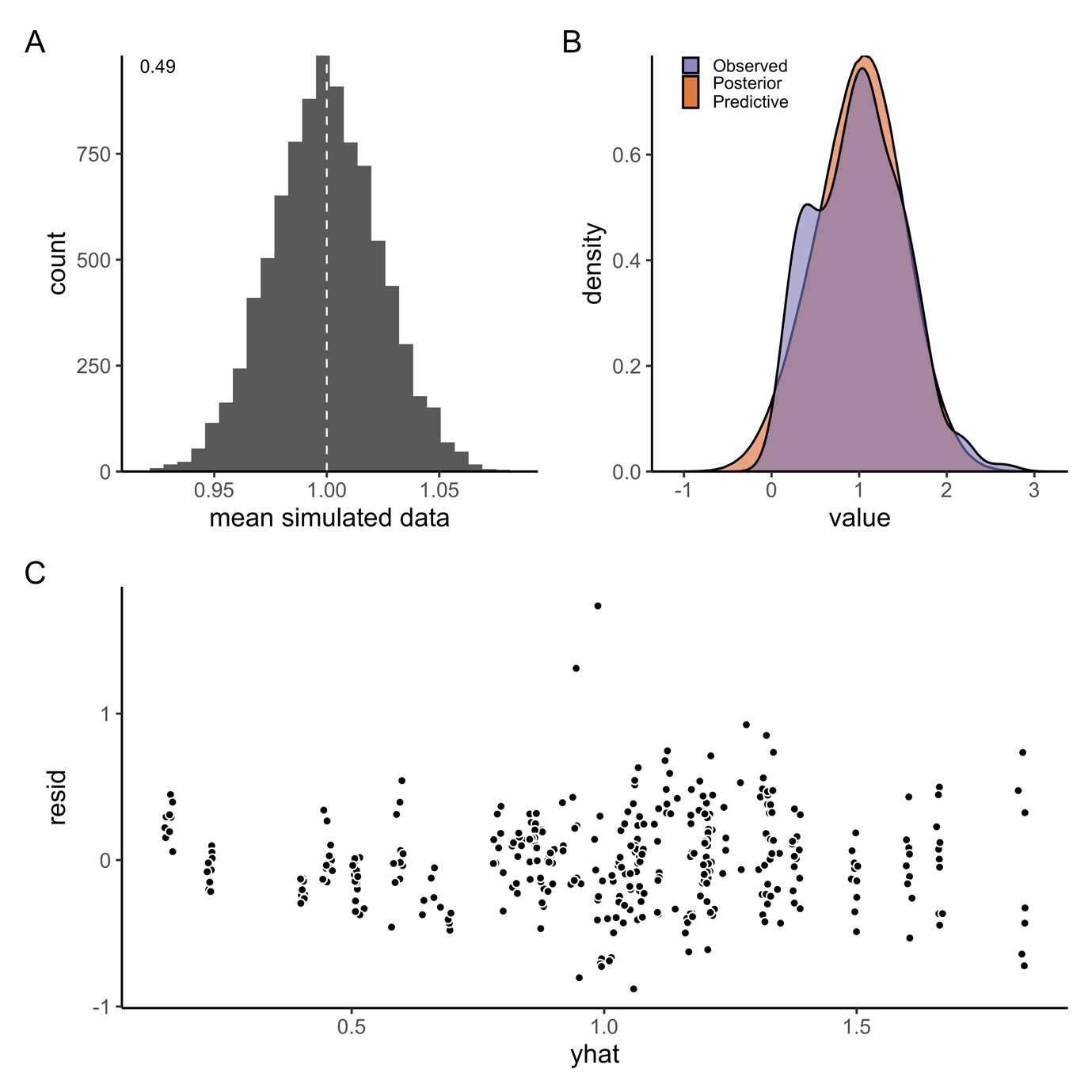
Description automatically generated

*Fig. S11. Posterior densities and trace plots for evaluation of chain convergence (by chain, indicated by color), for the global-level parameters for the Sharpe-Schoolfield model fitted to maximum consumption rate data with temperatures including beyond peak temperatures.*

Chart, scatter chart

Description automatically generated

*Fig. S12. Potential scale reduction factor () for the Sharpe-Schoolfield model fitted to maximum consumption rate data (including data beyond peak). This factor is based on the comparison of between and within-chain variation for the same parameter. A value close to one implies chains converged to the same distribution. The index of the parameter corresponds to species in alphabetical order.*



*Fig. S13. A) Model fit (mean) for the Sharpe-Schoolfield model fitted to maximum consumption rate data including temperatures beyond peak (by species). Fit is evaluated by simulating data from the likelihood (at each iteration of the MCMC chain), to compare how well it matches the original data. Each simulated data point is assigned a 0 or 1 if it is below or above the mean data point (the vertical line corresponds to the mean in data). The number in the plot corresponds to the mean of the vector of 0’s and 1’s. B) Posterior predictive distribution (orange) and distribution of data (purple). C) Difference between the observed value and the posterior median of the predicted value, plotted against fitted value.*

Chart

Description automatically generated

*Fig. S14. Posterior (black) and prior distribution (red) for the global parameters in the Sharpe-Schoolfield model for maximum consumption rate including data beyond peak, including their % overlap (rounded) and effective sample size (n.eff).*

## *Metabolic rate*

Shape, polygon

Description automatically generated with medium confidence

*Fig. S15. Posterior densities and trace plots for evaluation of chain convergence (by chain, indicated by color), for the global-level parameters for the metabolic rate model at temperatures below peak temperatures.*

Chart, scatter chart

Description automatically generated

*Fig. S16. Potential scale reduction factor () for the metabolic rate model. This factor is based on the comparison of between and within-chain variation for the same parameter. A value close to one implies chains converged to the same distribution. The index of the parameter corresponds to species in alphabetical order. Note that species with routine metabolism do not have estimates for a standard metabolic rate intercept and vice versa, hence, not all parameters in the graph have values.*

Chart, scatter chart

Description automatically generated

*Fig. S17. A) Model fit (mean) for the log-linear model of metabolic rate. Fit is evaluated by simulating data from the likelihood (at each iteration of the MCMC chain), to compare how well it matches the original data. Each simulated data point is assigned a 0 or 1 if it is below or above the mean data point (the vertical line corresponds to the mean in data). The number in the plot corresponds to the mean of the vector of 0’s and 1’s. B) Posterior predictive distribution (orange) and distribution of data (purple). C) Difference between the observed value and the posterior median of the predicted value, plotted against fitted value.*

Diagram, polygon

Description automatically generated

*Fig. S18. Posterior (black) and prior distribution (red) for the global parameters in the model for metabolic rate, including their % overlap and effective sample size (n.eff).*

## *Optimum growth temperature*

A picture containing text, transport

Description automatically generated

*Fig. S19. Posterior densities and trace plots for evaluation of chain convergence (by chain, indicated by color), for the global-level parameters for the model.*

Chart, scatter chart

Description automatically generated

*Fig. S20. Potential scale reduction factor () for the model. This factor is based on the comparison of between and within-chain variation for the same parameter. A value close to one implies chains converged to the same distribution. The index of the parameter corresponds to species in alphabetical order.*

Chart, histogram

Description automatically generated

*Fig. S21. A) Model fit (mean) for the model of optimum growth temperature as a function of body mass. Fit is evaluated by simulating data from the likelihood (at each iteration of the MCMC chain), to compare how well it matches the original data. Each simulated data point is assigned a 0 or 1 if it is below or above the mean data point (the vertical line corresponds to the mean in data). The number in the plot corresponds to the mean of the vector of 0’s and 1’s. B) Posterior predictive distribution (orange) and distribution of data (purple). C) Difference between the observed value and the posterior median of the predicted value, plotted against fitted value.*

Chart, histogram

Description automatically generated

*Fig. S22. Posterior (black) and prior distribution (red) for the global parameters in the model for , including their % overlap and effective sample size (n.eff).*

# References

Árnason, T., Björnsson, B., Steinarsson, A. & Oddgeirsson, M. (2009). Effects of temperature and body weight on growth rate and feed conversion ratio in turbot (*Scophthalmus maximus*). *Aquaculture*, 295, 218–225.

Baldwin, N.S. (1957). Food Consumption and Growth of Brook Trout at Different Temperatures. *Transactions of the American Fisheries Society*, 86, 323–328.

Beamish, F.W.H. (1964). Respiration of fishes with special emphasis on standard oxygen consumption II. Influence of weight and temperature on respiration of several species’. *Canadian Journal of Zoology/Revue Canadienne de Zoologie*, 42, 177–188.

Beamish, F.W.H. & Mookherjii, P.S. (1964). Respiration of fishes with special emphasis on standard oxygen consumption: I. influence of weight and temperature on respiration of goldfish, *Carassius auratus* l. *Can. J. Zool.*, 42, 161–175.

Bermudes, M., Glencross, B., Austen, K. & Hawkins, W. (2010). The effects of temperature and size on the growth, energy budget and waste outputs of barramundi (*Lates calcarifer*). *Aquaculture*, 306, 160–166.

Binkowski, F.P. & Rudstam, L.G. (1994). Maximum Daily Ration of Great Lakes Bloater. *Transactions of the American Fisheries Society*, 123, 335–343.

Björnsson, B., Steinarsson, A. & Árnason, T. (2007). Growth model for Atlantic cod (*Gadus morhua*): Effects of temperature and body weight on growth rate. *Aquaculture*, 271, 216–226.

Björnsson, B. & Tryggvadóttir, S.V. (1996). Effects of size on optimal temperature for growth and growth efficiency of immature Atlantic halibut (*Hippoglossus hippoglossus* L.). *Aquaculture*, 142, 33–42.

Brett, J.R., Shelbourn, J.E. & Shoop, C.T. (1969). Growth rate and body composition of fingerling sockeye salmon, *Oncorhynchus nerka*, in relation to temperature and ration size. *J. Fish. Res. Bd. Can.*, 26, 2363–2394.

Chipps, S.R. & Wahl, D.H. (2004). Development and Evaluation of a Western Mosquitofish Bioenergetics Model. *Transactions of the American Fisheries Society*, 133, 1150–1162.

Cui, Y. & Wootton, R.J. (1988). Bioenergetics of growth of a cyprinid, *Phoxinus phoxinus*: the effect of ration, temperature and body size on food consumption, faecal production and nitrogenous excretion. *J Fish Biology*, 33, 431–443.

Degani, G., Gallagher, M.L. & Meltzer, A. (1989). The influence of body size and temperature on oxygen consumption of the European eel, *Anguilla anguilla*. *J Fish Biology*, 34, 19–24.

Deslauriers, D., Chipps, S.R., Breck, J.E., Rice, J.A. & Madenjian, C.P. (2017). Fish Bioenergetics 4.0: An R-Based Modeling Application. *Fisheries*, 42, 586–596.

Du Perez, H., H., McLachlan, A. & Marais, J.F.K. (1986). Oxygen consumption of a shallow water teleost, the spotted grunter, *Pomadysis commersonni* (Lacépéde, 1802). *Comparative Biochemistry and Physiology*, 84a, 61–70.

Duston, J., Astatkie, T. & MacIsaac, P.F. (2004). Effect of body size on growth and food conversion of juvenile striped bass reared at 16–28 °C in freshwater and seawater. *Aquaculture*, 234, 589–600.

Elliott, J.M. (1976). The Energetics of Feeding, Metabolism and Growth of Brown Trout (*Salmo trutta* L.) in Relation to Body Weight, Water Temperature and Ration Size. *The Journal of Animal Ecology*, 45, 923.

Froese, R. & Pauly, D. (2019). *Editors. FishBase*. World Wide Web electronic publication. www.fishbase.org, (12/2019).

Froese, R., Thorson, J.T. & Reyes, R.B. (2014). A Bayesian approach for estimating length‐weight relationships in fishes. *Journal of Applied Ichthyology*, 30, 78–85.

From, J. & Rasmussen, G. (1984). A growth model, gastric evacuation, and body composition in rainbow trout, *Salmo gairdneri* Richardson, 1836. *Dana*, 3, 61–139.

Glencross, B.D. & Felsing, M. (2006). Influence of fish size and water temperature on the metabolic demand for oxygen by barramundi, *Lates calcarifer* (Bloch), in freshwater. *Aquaculture Res*, 37, 1055–1062.

Glover, D.C., DeVries, D.R. & Wright, R.A. (2012). Effects of temperature, salinity and body size on routine metabolism of coastal largemouth bass *Micropterus salmoides*. *Journal of Fish Biology*, 81, 1463–1478.

Handeland, S.O., Imsland, A.K. & Stefansson, S.O. (2008). The effect of temperature and fish size on growth, feed intake, food conversion efficiency and stomach evacuation rate of Atlantic salmon post-smolts. *Aquaculture*, 283, 36–42.

Hayward, R.S. & Arnold, E. (1996). Temperature Dependence of Maximum Daily Consumption in White Crappie: Implications for Fisheries Management. *Transactions of the American Fisheries Society*, 125, 132–138.

Heuton, M., Ayala, L., Morante, A., Dayton, K., Jones, A.C., Hunt, J.R., *et al.* (2018). Oxygen consumption of desert pupfish at ecologically relevant temperatures suggests a significant role for anaerobic metabolism. *J Comp Physiol B*, 188, 821–830.

Horodysky, A.Z., Brill, R.W., Bushnell, P.G., Musick, J.A. & Latour, R.J. (2011). Comparative metabolic rates of common western North Atlantic Ocean sciaenid fishes. *Journal of Fish Biology*, 79, 235–255.

Imsland, A.K., Foss, A., Sparboe, L.O. & Sigurdsson, S. (2006). The effect of temperature and fish size on growth and feed efficiency ratio of juvenile spotted wolffish *Anarhichas minor*. *J Fish Biology*, 68, 1107–1122.

Iwata, N., Kikuchi, K., Honda, H., Kiyono, M. & Kurokura, H. (1994). Effects of temperature on the growth of Japanese flounder. *Fisheries science*, 60, 527–531.

Laurel, B.J., Copeman, L.A., Spencer, M. & Iseri, P. (2017). Temperature-dependent growth as a function of size and age in juvenile Arctic cod (*Boreogadus saida*). *ICES Journal of Marine Science*, 74, 1614–1621.

Lessmark, O. (1983). Competition between perch (*Perca fluviatilis*) and roach (*Rutilus rutilus*) in south Swedish lakes. PhD Thesis. Limnologiska Institutionen, Lunds Universitet (Sweden).

Lin, X., Xie, S., Su, Y. & Cui, Y. (2008). Optimum temperature for the growth performance of juvenile orange-spotted grouper (*Epinephelus coioides* H.). *Chin. J. Ocean. Limnol.*, 26, 69–75.

Liu, J., Cui, Y. & Liu, J. (1998). Food consumption and growth of two piscivorous fishes, the mandarin fish and the Chinese snakehead. *Journal of Fish Biology*, 53, 1071–1083.

Liu, J., Cui, Y. & Liu, J. (2000). Resting metabolism and heat increment of feeding in mandarin fish (*Siniperca chuatsi*) and Chinese snakehead (*Channa argus*). *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 127, 131–138.

Luo, Y.P. & Wang, Q.Q. (2012). Effects of body mass and temperature on routine metabolic rate of juvenile largemouth bronze gudgeon *Coreius guichenoti*. *Journal of Fish Biology*, 80, 842–851.

Marmulla, G. & Rosch, R. (1990). Maximum daily ration of juvenile fish fed on living natural zooplankton. *J Fish Biology*, 36, 789–801.

Mesa, M.G., Weiland, L.K., Christiansen, H.E., Sauter, S.T. & Beauchamp, D.A. (2013). Development and evaluation of a bioenergetics model for bull trout. *Transactions of the American Fisheries Society*, 142, 41–49.

Meskendahl, L., Herrmann, J.-P. & Temming, A. (2010). Effects of temperature and body mass on metabolic rates of sprat, *Sprattus sprattus* L. *Mar Biol*, 157, 1917–1927.

Messmer, V., Pratchett, M.S., Hoey, A.S., Tobin, A.J., Coker, D.J., Cooke, S.J., *et al.* (2017). Global warming may disproportionately affect larger adults in a predatory coral reef fish. *Global Change Biology*, 23, 2230–2240.

Milano, D., Vigliano, P. & Beauchamp, D. (2016). Effect of body size and temperature on respiration of *Galaxias maculatus* (Pisces: Galaxiidae). *New Zealand Journal of Marine and Freshwater Research*, 51, 295–303.

Nytrø, A.V., Vikingstad, E., Foss, A., Hangstad, T.A., Reynolds, P., Eliassen, G., *et al.* (2014). The effect of temperature and fish size on growth of juvenile lumpfish (*Cyclopterus lumpus* L.). *Aquaculture*, 434, 296–302.

Ohlberger, J., Mehner, Thomas., Staaks, Georg. & Hölker, Franz. (2012). Intraspecific temperature dependence of the scaling of metabolic rate with body mass in fishes and its ecological implications. *Oikos*, 121, 245–251.

Patterson, J.T., Mims, S.D. & Wright, R.A. (2013). Effects of body mass and water temperature on routine metabolism of American paddlefish *Polyodon spathula*: routine metabolism of *polyodon* *spathula*. *J Fish Biol*, 82, 1269–1280.

Peck, M.A., Buckley, L.J. & Bengtson, D.A. (2005). Effects of temperature, body size and feeding on rates of metabolism in young-of-the-year haddock. *Journal of Fish Biology*, 66, 911–923.

Pirozzi, I. & Booth, M.A. (2009). The effect of temperature and body weight on the routine metabolic rate and postprandial metabolic response in mulloway, *Argyrosomus japonicus*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 154, 110–118.

Rangel, R.E. & Johnson, D.W. (2018). Metabolic responses to temperature in a sedentary reef fish, the bluebanded goby (*Lythrypnus dalli*, Gilbert). *Journal of Experimental Marine Biology and Ecology*, 501, 83–89.

Siikavuopio, S.I., Foss, A., Saether, B.-S., Gunnarsson, S. & Imsland, A.K. (2013). Comparison of the growth performance of offspring from cultured versus wild populations of arctic charr, *Salvelinus alpinus* (L.), kept at three different temperatures. *Aquac Res*, 44, 995–1001.

Slesinger, E., Andres, A., Young, R., Seibel, B., Saba, V., Phelan, B., *et al.* (2019). The effect of ocean warming on black sea bass (*Centropristis striata*) aerobic scope and hypoxia tolerance. *PLoS ONE*, 14, e0218390.

Sun, L. & Chen, H. (2014). Effects of water temperature and fish size on growth and bioenergetics of cobia (*Rachycentron canadum*). *Aquaculture*, 426–427, 172–180.

Tirsgaard, B., Behrens, J.W. & Steffensen, J.F. (2015). The effect of temperature and body size on metabolic scope of activity in juvenile Atlantic cod *Gadus morhua* L. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 179, 89–94.

Tomala, D., Chavarria, J. & Angeles, B. (2014). Evaluacion de la tasa de consumo de oxigeno de *Colossoma macropomum* en relacion al peso corporal y temperatura del agua. *lajar*, 42, 971–979.

Tomiyama, T., Kusakabe, K., Otsuki, N., Yoshida, Y., Takahashi, S., Hata, M., *et al.* (2018). Ontogenetic changes in the optimal temperature for growth of juvenile marbled flounder *Pseudopleuronectes yokohamae*. *Journal of Sea Research*, 141, 14–20.

Wang, H.P., Hayward, R.S., Whitledge, G.W. & Fischer, S.A. (2003). Prey-size Preference, Maximum Handling Size, and Consumption Rates for Redear Sunfish *Lepomis microlophus* Feeding on Two Gastropods Common to Aquaculture Ponds. *J World Aquaculture Soc*, 34, 379–386.

Wootton, R.J., Allen, J.R.M. & Cole, S.J. (1980). Effect of body weight and temperature on the maximum daily food consumption of *Gasterosteus aculeatus* L. and *Phoxinus phoxinus* (L.): selecting an appropriate model. *J Fish Biology*, 17, 695–705.

Xie, Xiaojun. & Sun, Ruyung. (1990). The Bioenergetics of the Southern Catfish (*Silurus meridionalis* Chen). I. Resting Metabolic Rate as a Function of Body Weight and Temperature. *Physiological Zoology*, 63, 1181–1195.

Yamanaka, H., Takahara, T., Kohmatsu, Y. & Yuma, M. (2013). Body size and temperature dependence of routine metabolic rate and critical oxygen concentration in larvae and juveniles of the round crucian carp *Carassius auratus grandoculis* Temminck & Schlegel 1846. *J. Appl. Ichthyol.*, 29, 891–895.

Zhang, L., Zhao, Z.-G. & Fan, Q.-X. (2017). Effects of water temperature and initial weight on growth, digestion and energy budget of yellow catfish *Pelteobagrus fulvidraco* (Richardson, 1846). *J Appl Ichthyol*, 33, 1108–1117.