Matthew E. Berginski Curriculum Vitae September 2015

Personal Information:

Work Address: 101 Science Drive, CIEMAS Room #1373

Durham, NC 27708

Email: matthew.berginski@gmail.com OR matthew.berginski@duke.edu

Work Phone: 919-660-5125 (Office) OR 919-613-9128 (Lab)

Cell Phone: 859-797-6722

Education:

2006-2013 Ph.D. in Biomedical Engineering with a Certificate in Bioinformatics

and Computational Biology, University of North Carolina at Chapel

Hill and North Carolina State University

2002-2006 B.S. in Biomedical Engineering, Georgia Institute of Technology

Publications (*indicates co-first-authorship):

See Google Scholar for Citation Information

- 1. Creed SJ, Le CP, Hassan M, Pon CK, Albold S, Chan KT, <u>Berginski ME</u>, Huang Z, Bear JE, Lane JR, Halls ML, Ferrari D, Nowell CJ, Sloan EK. β2-adrenoceptor signaling regulates invadopodia formation to enhance tumor cell invasion. Breast Cancer Research, 2015 (HTML|PDF)
- 2. LaCroix AS, Rothenberg KE, <u>Berginski ME</u>, Urs AN, Hoffman BD. Construction, imaging, and analysis of FRET-based tension sensors in living cells. Methods in Cell Biology, 2015 (<u>HTML|PDF</u>)
- 4. Chu PH, Tsygankov D, <u>Berginski ME</u>, Dagliyan O, Gomez SM, Elston TC, Karginov AV, Hahn KM. Engineered kinase activation reveals unique morphodynamic phenotypes and associated trafficking for Src family isoforms. *PNAS*, 2014 (<u>HTML|PDF</u>)
- 5. <u>Berginski ME</u>, Creed SJ, Cochran S, Roadcap DW, Bear JE, Gomez SM. Automated analysis of invadopodia dynamics in live cells. *PeerJ*, 2014 (<u>HTML|PDF</u>) Part of and Cover Image for PeerJ's Top Cancer Papers <u>Collection</u>
- 6. Karginov AV, Tsygangov D, <u>Berginski ME</u>, Chu P, Trudeau ED, Yi JJ, Gomez SM, Elston TC, Hahn KM. Dissecting motility signaling through activation of specific Src-effector complexes. *Nature Chemical Biology*, 2014 (HTML|PDF)
- 7. Lin LK, Fulton LM, <u>Berginski ME</u>, West ML, Taylor NA, Moran TP, Coghill JM, Blazer BR, Bear JE, Serody JS. Intravital imaging of donor allogeneic effector and regulatory T cells with host dendritic cells during GvHD. *Blood*, 2014 (<u>HTML|PDF</u>)

- 8. Vitriol EA, Wise AL, **Berginski ME**, Bamburg JR, and Zheng JQ. Instantaneous Inactivation of Cofilin1 Demonstrates Its Functions of Filament Severing and Depolymerization in Regulating F-actin Networks. Molecular Biology of the Cell, 2013 (HTML|PDF)
- 9. <u>Berginski ME</u>, Gomez SM. The Focal Adhesion Analysis Server: a web tool for analyzing focal adhesion dynamics. *F1000Research*, 2013 (HTML|PDF)
- 10. Sankar CP, Barhoumi R, <u>Berginski ME</u>, Sreenivasappa H, Tranche A, Gomez SM, Rivera GM. Nck enables directional cell migration through the coordination of polarized membrane protrusion with adhesion dynamics. *Journal of Cell Science*, 2013 (<u>HTML|PDF</u>)
- 11. Chen Z, Lessey E, <u>Berginski ME</u>, Cao L, Li J, Trepat X, Itano M, Gomez SM, Kapustina M, Huang C, Burridge K, Truskey G, and Jacobson K. Gleevec, an abl family inhibitor, produces a profound change in cell shape and migration. *PLoS ONE*, 2013. (<u>HTML|PDF</u>)
- 12. Wu C*, Asokan SB*, <u>Berginski ME</u>, Haynes EM, Sharpless NE, Griffith JD, Gomez SM, Bear JE. Arp2/3 Is Critical for Lamellipodia and Response to Extracellular Matrix Cues but Is Dispensable for Chemotaxis. *Cell*, 2012 (<u>HTML</u>|PDF)
- 13. Shen K, Tolbert CE, Guilluy C, Swaminathan VS, <u>Berginski ME</u>, Burridge K, Superfine R, Campbell SL. The vinculin C-terminal hairpin mediates F-actin bundle formation, focal adhesion, and cell mechanical properties. *J Biol Chem*, 2011 (<u>HTML|PDF</u>)
- 14. <u>Berginski ME*</u>, Vitriol EA*, Hahn KM, Gomez SM. High-Resolution Quantification of Focal Adhesion Spatiotemporal Dynamics in Living Cells. *PLoS ONE*, 2011 (HTML|PDF)

Presentations and Posters:

- Construction and use of an Ezrin tension sensor to measure actin-plasma membrane loading, Triangle Cytoskeleton Meeting, Saxapahaw, NC. September 2015
- 2. Comprehensive Spatiotemporal Analysis of Focal Adhesion Dynamics in Living Cells, BMES Meeting, Pittsburgh, PA. October 2009
- 3. Quantitative Analysis of Focal Adhesions in TIRF Microscopy Images, Bioengineering and Bioinformatics Summer Institute, Richmond, VA, Keynote Seminar. August 2009
- 4. Focal Adhesion Dynamics Analysis Through Quantitative Image Processing, RECOMB Systems Biology, Boston, MA. October 2008
- 5. Automatic Characterization of Focal Adhesions in TIRF Microscopy Images, Institute for Biological Engineering Conference, Chapel Hill, NC. March 2008, Received 3rd place in poster competition

Honors and Funding:

2016	NIH F32 Postdoctoral Fellowship (GM119294)
2006-2009	NSF Graduate Research Fellowship
2006-2008	North Carolina State Dean's Fellowship
2002-2006	Graduated Summa Cum Laude from Georgia Institute of Technology

Other Experience:

2006-2013 Tutor with the Learning Center at the University of North Carolina at Chapel Hill

2008-2009	Intern at the Office of Technology Development at the University of North
	Carolina at Chapel Hill
2005-2006	Teaching Assistant in BMED 2300 at the Georgia Institute of Technology