

Contents

1. Node perturbation analysis of each articulation point interactome in the prenatal and postnatal networks	2
1.1 Interactomes involved in regulation of prenatal development	3
1.2 Interactomes involved in regulation of postnatal development.....	57
2. References (for Table 1).....	128

1. Node perturbation analysis of each articulation point interactome in the prenatal and postnatal networks

We performed single node and multiple node perturbation analyses (OE/KO) at four different simulations i.e. when TF activates gene and miRNA (Simulation 1), when TF represses both miRNA and gene (Simulation 2), when TF represses miRNA and activates gene expression (Simulation 3) and when TF represses gene and activates miRNA expression (Simulation 4). In single node perturbation analysis, we perturbed each miRNA and TF regulating the AP interactome (PC1). In multiple node perturbation analysis, we perturbed genes positively regulating each stage of neurodevelopment either along with all miRNAs and TFs regulating the articulation point interactome (PC2 and PC3) or along with each miRNA/TF regulating the articulation point interactome (PC4 and PC5). We have shown the perturbation results as activation frequency i.e. for every 1000 simulations, the number of times a given stage is upregulated at $t = 150$. We have used the term regulatory state (RS), to show the neurodevelopmental stages which are shown to be regulated in more than one regulatory state (RS). For example, in a particular PC, if a specific stage shows all three regulatory states i.e. upregulation (100%), downregulation (0%) as well as regulation between 0%-100%, we represent such states as RS1. We have four such regulatory states (RS1, RS2, RS3 and RS4), explained in each table. We have presented the results of node perturbation analysis for each articulation point in the prenatal and postnatal network under two headings, single node perturbation analysis and multiple node perturbation analysis as shown below:

Single node perturbation analysis

- ***Perturbation condition 1:*** OE and KO of each miRNA and each TF regulating AP interactome

Multiple node perturbation analysis

- ***Perturbation condition 2:*** OE of genes positively regulating each stage of neurodevelopment (regulated by AP interactome) and OE of miRNAs and TFs regulating AP interactome
- ***Perturbation condition 3:*** KO of genes positively regulating each stage of neurodevelopment (regulated by AP interactome) and OE of miRNAs and TFs regulating AP interactome
- ***Perturbation condition 4:*** OE of genes positively regulating stage of neurodevelopment (regulated by AP interactome) along with OE of each factor (miRNA/TF) regulating AP interactome
- ***Perturbation condition 5:*** KO of genes positively regulating stage of neurodevelopment (regulated by AP interactome) along with OE of each factor (miRNA/ TF) regulating AP interactome

1.1 Interactomes involved in regulation of prenatal development

We analyzed the effect of each perturbation (PC1-PC5) on each stage of neurodevelopment and the overall neurodevelopment process (combined all the stages regulated by AP interactome as one stage). We could not perform node perturbation analysis for the following articulation points, DTNBP1, CNTNAP2, DLG2 and GRIN2A. There were no experimentally validated miRNAs shown to regulate DTNBP1, so no feedforward loops (FFLs) were identified for this articulation point. We could also not perform NP analysis for CNTNAP2, DLG2 and GRIN2A genes, as the FFLs regulating these APs did not regulate genes that interact with these APs

A. DISC1 interactome

During prenatal neurodevelopment, DISC1 interactome has been shown to regulate embryogenesis, neurulation, proliferation, migration, differentiation, neurite growth and synaptogenesis. We have specified rules for each stage of neurodevelopment regulated by DISC1 interactome as well as for the overall neurodevelopment process (all the stages regulated by DISC1 combined as one stage). As GSK3B gene is shown to regulate migration in two different gene expression states, we have given 2 Boolean rules for migration as well as for the overall neurodevelopment process, where DISC1 and GSK3B is expressed regulating neurodevelopment (when GSK3B is expressed) and where DISC1 is expressed and GSK3B is not expressed regulating neurodevelopment (when GSK3B is not expressed).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating DISC1 interactome

Simulations 1-4: OE and KO of each miRNA and each TF downregulated (0%) each of the above-mentioned stages of development and the overall neurodevelopment process regulated by DISC1 interactome.

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) and OE of miRNAs and TFs regulating DISC1 interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment regulated by DISC1 interactome (embryogenesis/ neurulation/ proliferation/ migration (when GSK3B not expressed)/ migration (when GSK3B expressed)/ differentiation/ neurite growth/ synaptogenesis) along with miRNAs and TFs, upregulated (100%) each same stage of development (embryogenesis/ neurulation/ proliferation/ migration / differentiation/

neurite growth/ synaptogenesis). Differentiation stage was also shown to upregulated along with upregulation of each stage.

OE of genes positively regulating neurodevelopment process (when GSK3B is not expressed), upregulated (100%) all the stages of neurodevelopment including the overall neurodevelopment process (when GSK3B is not expressed) whereas OE of genes positively regulating neurodevelopment (when GSK3B is expressed), upregulated (100%) neurodevelopment (when GSK3B is expressed), differentiation, migration, neurulation and synaptogenesis.

Perturbation condition 3: KO of genes positively regulating each stage of development (regulated by DISC1 interactome) and KO of miRNAs and TFs regulating DISC1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (embryogenesis/ neurulation/ proliferation/ migration (when GSK3B expressed)/ migration (when GSK3B not expressed)/ differentiation/ neurite growth/ synaptogenesis/ neurodevelopment (when GSK3B not expressed)/ neurodevelopment (when GSK3B expressed) along with KO of miRNAs and TFs, downregulated (0%) each same stage of neurodevelopment.

Simulations 2 and 4: KO of genes positively regulating each stage of neurodevelopment (except genes positively regulating migration (when GSK3B is expressed) and genes positively regulating neurodevelopment (when GSK3B is expressed)), along with KO of miRNAs and TFs upregulated migration stage (when GSK3B is expressed) (100%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) along with OE of each factor (miRNA/TF) regulating DISC1 interactome

a. Embryogenesis

Simulations 1-4: OE of genes positively regulating embryogenesis and OE of each miRNA/TF upregulated embryogenesis (100%). Similarly, OE of genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated embryogenesis (100%).

Simulation 3: OE of each miRNA/TF along with OE of genes positively regulating neurulation/ proliferation/ migration (when GSK3B is not expressed)/ differentiation/ neurite growth/ synaptogenesis showed overall regulation of embryogenesis process in all three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of either let7e, miR124, miR155, miR26a, miR877, miR190a, miR5011, EP300, TAL1, STAT3, HNF4A, GATA1, KDM2B, KDM5B or PPARG or TEAD4 along with OE of genes positively regulating neurulation/proliferation/migration (GSK3B is not expressed)/differentiation/neurite growth/synaptogenesis downregulated embryogenesis (0%). OE of either TP53 or miR320a or NUCKS1 along with OE of genes positively regulating neurulation/proliferation/migration (when GSK3B is not expressed)/differentiation/neurite growth/synaptogenesis upregulated embryogenesis (100%). OE of each miRNA/ TF (except OE of let7e/ miR124/ miR155/ miR26a/ miR877/ miR190a/ miR5011/ miR320a/ EP300/ TAL1/ STAT3/ HNF4A/ GATA1/ KDM2B/ KDM5B/ PPARG/TP53/NUCKS1) along with OE of genes positively regulating neurulation/proliferation/migration (when GSK3B is not expressed)/differentiation/neurite growth/synaptogenesis regulated embryogenesis (between 0%-100%).

b. Neurulation

Simulations 1-4: OE of genes positively regulating neurulation and OE of each miRNA/ TF upregulated neurulation (100%). Similarly, OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with OE of each miRNA/TF, upregulated neurulation (100%).

c. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation and OE of each miRNA/ TF upregulated proliferation (100%).

Simulations 1, 2 and 4: OE of genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated proliferation (100%).

Simulation 3: OE of each factor along with OE of genes positively regulating neurulation/ embryogenesis/ migration (GSK3B is not expressed)/ neurite growth/ differentiation/ synaptogenesis/ neurodevelopment (GSK3B is not expressed) showed regulation of overall proliferation process in all three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of either miR744 or HNF4A along with OE of genes positively regulating neurulation/ embryogenesis/ neurite growth/ differentiation/ synaptogenesis/ neurodevelopment (GSK3B is not expressed) downregulated proliferation (0%). OE of either miR320a, EP300, TRIM28, STAT3, NUCKS1, NFI, KDM2B, GATA1 or PPARG along with OE of genes positively regulating neurulation/ embryogenesis/ neurite growth/ migration (GSK3B is not expressed)/ differentiation/ synaptogenesis/ neurodevelopment (GSK3B is not expressed), upregulated proliferation (100%). OE of each factor (except miR744/ HNF4A/ miR320a/ EP300/ TRIM28/ STAT3/ NUCKS1/ NFI/ KDM2B/ PPARG) along with OE of genes positively regulating neurulation/ embryogenesis/ neurite growth/

differentiation/ synaptogenesis/ neurodevelopment (when GSK3B is not expressed)

regulated proliferation between 0%-100%.

We also observed that OE of genes positively regulating migration (when GSK3B is not expressed) and OE of either miR124/miR155/miR744/HNF4A, downregulated proliferation (0%). OE of genes positively regulating migration (when GSK3B is not expressed) and OE of either miR190a/ miR5011, regulated proliferation (between 0%-100%). OE of each factor (except OE miR124 or miR155 or miR744 or HNF4A or miR190a or miR5011) along with OE of genes positively regulating migration (when GSK3B not expressed) upregulated proliferation (100%). Thus, OE of each factor along with OE of genes positively regulating migration (when GSK3B not expressed) showed regulation of overall proliferation process in all three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%).

d. Migration (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is not expressed) or OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with OE of each miRNA/ TF, upregulated migration (100%).

e. Migration (GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed) or OE of genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of each miRNA/TF, upregulated migration (100%).

f. Differentiation

Simulations 1-4: OE of miRNA/TF along with OE of genes positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ neurodevelopment (when GSK3B is expressed)/ neurodevelopment (when GSK3B is not

expressed)/ neurulation/ proliferation/ synaptogenesis/ differentiation, upregulated differentiation (100%).

h. Neurite growth

Simulations 1-4: OE of each miRNA/TF along with OE of genes positively regulating neurite growth/ genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated neurite growth (100%).

i. Synaptogenesis

Simulations 1-4: OE of each miRNA/TF along with OE of genes positively regulating synaptogenesis/ genes positively regulating neurodevelopment (when GSK3B is not expressed)/ genes positively regulating neurodevelopment (when GSK3B is expressed), upregulated synaptogenesis (100%).

j. Neurodevelopment (GSK3B is expressed)

Simulations 1-4: OE of each miRNA/TF along with OE of genes positively regulating neurodevelopment (when GSK3B is expressed), upregulated neurodevelopment (100%).

k. Neurodevelopment (GSK3B is not expressed)

Simulations 1-4: OE of each miRNA/TF along with OE of genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) along with KO of each factor (miRNA/TF) regulating DISC1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (embryogenesis, neurulation, proliferation, migration, differentiation, neurite growth and synaptogenesis/ neurodevelopment (when GSK3B is expressed)/ neurodevelopment (when GSK3B is not expressed)) along with KO of each miRNA/TF, downregulated each same stage of neurodevelopment (0%).

Table 1: Perturbation results of DISC1 interactome involved in prenatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)																			
		Embryogenesis				Neurulation				Proliferation				Migration (when GSK3B is not expressed)				Migration (when GSK3B is expressed)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating embryogenesis and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurulation and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	100%	100%	100%	100%	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	RS1	0%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating differentiation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%

	4	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	RS1	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is not expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	RS1	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC denotes Perturbation condition; Sim1 – Simulation 1; Sim2 – Simulation 2; Sim3 – Simulation 3; Sim4 – Simulation 4; 0% – Downregulation; 100% – Upregulation; RS1 represents regulatory state 1, where PC leads to downregulation 0%, regulation between 0%-100% and upregulation 100%. RS2 represents regulatory state 2, where PC leads to upregulation 100% and downregulation 0%

differentiation and factors	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is not expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC denotes Perturbation condition; Sim1 - Simulation 1; Sim2 - Simulation 2; Sim3 - Simulation 3; Sim4 - Simulation 4; 0% - Downregulation; 100% - Upregulation

Summary of perturbation results for DISC1 interactome involved in prenatal regulation

PC1 (Simulations 1-4): Each stage of development regulated by DISC1 interactome was downregulated (0%), as none of the individual factor was shown to be involved in upregulation of stages of development.

PC2 (Simulations 1-4): When genes positively regulating each stage (except migration (when GSK3B is expressed)) regulated by DISC1 interactome was overexpressed along with factors, each same stage along with differentiation stage was upregulated (100%). As all these stages overexpress DISC1, DISC1 upregulates expression of genes involved in differentiation. When genes positively regulating migration (when GSK3B is expressed) along with factors were overexpressed, only migration stage was upregulated (100%). As DISC1 was not expressed in this migration stage, differentiation was not upregulated.

When genes positively regulating neurodevelopment (when GSK3B is expressed) along with factors were overexpressed, neurulation, differentiation, migration (when GSK3B is expressed), synaptogenesis and neurodevelopment (when GSK3 is expressed) stages were upregulated (100%). In all these stages, GSK3B expression was not involved in negative regulation. When genes positively regulating neurodevelopment (when GSK3B is not expressed) was overexpressed along with factors, except migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed), all other stages regulated by DISC1 interactome were upregulated (100%).

PC3 (Simulations 1-4): When genes positively regulating each stage (regulated by DISC1 interactome) was KO along with factors regulating the DISC1 interactome, each same stage was downregulated (0%)

Simulations 2 and 4: At simulations 2 and 4, TF repress gene expression so KO of all factors as well as KO of genes in stages (which express DISC1), upregulated migration (when GSK3B is expressed). DISC1 negatively regulates GSK3B expression.

PC4 (Simulations 1-4): When genes positively regulating each stage (except migration (when GSK3B is expressed)) regulated by DISC1 interactome was overexpressed along with each factor, each same stage as well as the differentiation stage was upregulated (100%). As all these stages OE DISC1, DISC1 upregulates expression of genes involved in differentiation. When genes positively regulating migration (when GSK3B is expressed) along with each factor was overexpressed, only migration stage was upregulated (100%). As DISC1 was not expressed in this migration stage, differentiation was not upregulated.

When genes positively regulating neurodevelopment (when GSK3B is expressed) along with each factor were overexpressed, neurulation, differentiation, migration (when GSK3B is expressed), synaptogenesis and neurodevelopment (when GSK3 is expressed) stages were upregulated (100%). In all these stages, GSK3B expression was not involved in negative regulation. When genes positively regulating neurodevelopment (when GSK3B is not expressed) was overexpressed along with each factor, except migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed), all other stages regulated by DISC1 interactome were upregulated (100%).

Simulation 3: OE of genes positively regulating neurulation/ proliferation/ migration (when GSK3B is not expressed)/ differentiation/ neurite growth/ synaptogenesis along with OE of each factor showed overall regulation of embryogenesis process in more than one regulatory state (RS1). Similarly, OE of genes positively regulating neurulation/ proliferation/ migration (when GSK3B is not expressed)/ differentiation/ neurite growth/ synaptogenesis/ neurodevelopment (when GSK3B is not expressed) along with OE of each factor showed

overall regulation of proliferation process in more than one regulatory state (RS1). These perturbation results show the definite role of certain factors (along with genes in other stages) in regulation of embryogenesis and proliferation stages.

PC5 (Simulations 1-4): When genes positively regulating each stage (regulated by DISC1 interactome) was KO along with each factor regulating the DISC1 interactome, each same stage was downregulated (0%).

Summary of the NP results for DISC1 interactome is given in Table 1 and Table 2.

B. GSK3B interactome

During prenatal development, GSK3B interactome has been shown to regulate embryogenesis, neurulation, proliferation, migration, neurite growth and synaptogenesis. Boolean rules for each of these stages has been given. We have given two different Boolean rules for migration stage, to specify the condition where GSK3B upregulation regulates migration and where GSK3B downregulation regulates migration. So, we also have two different Boolean rules to represent the regulation of overall neurodevelopmental process.

Single node perturbation analysis

Perturbation condition 1: OE and KO of each miRNA and each TF regulating GSK3B interactome

Simulations 1, 2 and 4: OE and KO of each miRNA and each TF downregulated each stage of neurodevelopment (0%) regulated by GSK3B interactome.

Simulation 3: In PC1, the overall neurulation process was shown to be regulated in all three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of each miRNA/TF (except miR744, miR877, miR15a, miR15b, miR16, miR195, miR182, EP300, CREB1, STAT3, FOXM1) regulated neurulation (between 0% and 100%). Similarly, KO of each miRNA (except KO of miR15b/ CREB1) regulated neurulation (between 0% and 100%). OE of miR744/ miR877 downregulated neurulation

(0%). OE of miR15a/ miR15b/ miR16/ miR195/ miR182/ EP300/ CREB1/ STAT3/ FOXM1, upregulated neurulation (100%).

Multiple node perturbation analysis

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) and OE of miRNAs and TFs regulating GSK3B interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (embryogenesis/ neurulation/ proliferation/ migration (when GSK3B is not expressed) /migration (when GSK3B is expressed)/ neurite growth/ synaptogenesis) along with OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of miRNAs and TFs, upregulated embryogenesis, neurulation, proliferation, migration (when GSK3B is not expressed), neurite growth and neurodevelopment (when GSK3B is not expressed) (100%).

a. Synaptogenesis

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed)/ neurodevelopment (when GSK3B is expressed) along with OE of miRNAs and TFs, also upregulated synaptogenesis (100%).

b. Migration (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of miRNAs and TFs, upregulated migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed) (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) and KO of miRNAs and TFs regulating GSK3B interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (embryogenesis/ neurulation/ proliferation/ migration/ neurite growth/ synaptogenesis/ neurodevelopment (when GSK3B is expressed)/ neurodevelopment (when GSK3B is not expressed)) and KO of miRNAs and TFs did not upregulate each same stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) along with OE of each factor (miRNA/ TF) regulating GSK3B interactome

a. Embryogenesis

Simulations 1-4: OE of genes positively regulating embryogenesis along with OE of each miRNA/ TF, upregulated embryogenesis (100%). Similarly, OE of genes positively regulating neurodevelopment (GSK3B not expressed) and OE of each miRNA/TF, upregulated embryogenesis (100%).

Simulation 3: We observed overall regulation of embryogenesis in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating migration (when GSK3B not expressed)/ proliferation/ neurite growth along with OE of miR320a/SMAD4/NUCKS1/TEAD4 regulated embryogenesis (between 0% and 100%). OE of genes positively regulating migration (when GSK3B not expressed)/ proliferation/ neurite growth along with OE of each miRNA/ TF (except OE of miR320a/SMAD4/NUCKS1/TEAD4) downregulated embryogenesis (0%).

b. Neurulation

Simulation 1: We observed overall regulation of neurulation in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively

regulating neurulation and OE of miR190a/ miR5011/ miR124/ TP53/ HNF4A regulated neurulation (between 0%-100%). Similarly, OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of miR190a/ miR5011/ miR124/ TP53 /HNF4A regulated neurulation (between 0%-100%). OE of genes positively regulating neurodevelopment (when GSK3B not expressed)/ neurulation and OE of each miRNA/TF (except OE of miR190a/ miR5011/ miR124/ TP53 /HNF4A) upregulated neurulation (100%).

Simulation 2: OE of genes positively regulating neurulation/ neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF upregulated neurulation (100%).

Simulation 3: We observed overall regulation of neurulation in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation/ neurulation/ neurodevelopment (when GSK3B is not expressed) and OE of EP300/STAT3/CREB1/FOXO1 downregulated neurulation (0%). OE of miR744/ miR877 along with OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation downregulated neurulation (0%). We also observed that OE of miR15a/ miR15b/ miR16/ miR195/ miR182 along with OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation/ neurulation/ neurodevelopment (when GSK3B is not expressed) upregulated neurulation (100%).

OE of genes positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation/ neurulation/ neurodevelopment (when GSK3B is not expressed) along with OE of each miRNA/TF (except OE of EP300/STAT3/CREB1/FOXO1/ miR15a/ miR15b/ miR16/ miR195/ miR182) regulated neurulation (between 0%-100%).

Similarly, OE of each miRNA (except OE of miR744/ miR877) along with OE of genes

positively regulating embryogenesis/ migration (GSK3B is not expressed)/ neurite growth/ proliferation regulated neurulation (between 0%-100%).

Simulation 4: We observed overall regulation of embryogenesis in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively regulating neurodevelopment (when GSK3B not expressed)/ genes positively regulating neurulation and OE of miR190a/miR5011/miR124/TP53/HNF4A regulated neurulation (between 0% - 100%). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed)/ genes positively regulating neurulation and OE of each miRNA/TF (except miR190a/miR5011/miR124/TP53/HNF4A), upregulated neurulation (100%).

c. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF, upregulated proliferation (100%).

Simulation 3: We observed overall regulation of proliferation process in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth and OE of miR155/ miR124/ TP53/ HNF4A/miR744 downregulated proliferation (0%). OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth and OE of miR190a/ miR5011 regulated proliferation (between 0% - 100%). However, OE of each miRNA/TF (except OE of miR155/ miR124/ TP53/ HNF4A/miR744/ miR190a/ miR5011) along with OE of genes either positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth showed upregulation of proliferation (100%).

d. Migration (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is not expressed)/ neurodevelopment (when GSK3B is not expressed) along with OE of each miRNA/TF, upregulated migration (100%).

e. Migration (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed)/ neurodevelopment (when GSK3B is expressed) along with OE of each miRNA/TF, upregulated migration (100%).

f. Neurite growth

Simulations 1-4: OE of genes positively regulating neurite growth/ neurodevelopment (when GSK3B is not expressed) along with OE of each miRNA/TF, upregulated neurite growth (100%).

g. Synaptogenesis

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed)/ genes positively regulating synaptogenesis/ genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of each miRNA/TF, upregulated synaptogenesis (100%).

h. Neurodevelopment (when GSK3B is not expressed)

Simulation 1: We observed regulation of overall neurodevelopment in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of miR190a/ miR5011/ miR124/ TP53/ HNF4A and OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) regulated neurodevelopment (between 0% - 100%). However, OE of each miRNA/TF (except OE of miR190a/ miR5011/ miR124/ TP53/ HNF4A) and OE of genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated neurodevelopment (100%).

Simulation 2: OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF upregulated neurodevelopment (100%).

Simulation 3: We observed overall regulation of neurodevelopment in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of EP300/ STAT3/ CREB1/ FOXM1/ miR15a/ miR15b/ miR16/ miR195/ miR182 and OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) upregulated neurodevelopment (100%). OE of each miRNA/TF (except OE of EP300/ STAT3/ CREB1/ FOXM1/ miR15a/ miR15b/ miR16/ miR195/ miR182) and OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) regulated neurodevelopment (between 0%-100%).

Simulation 4: We observed overall regulation of neurodevelopment in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of miR190a/ miR5011/ miR124/ TP53/ HNF4A regulated neurodevelopment (between 0% - 100%). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF (except OE of miR190a/ miR5011/ miR124/ TP53/ HNF4A) upregulated neurodevelopment (100%).

i. Neurodevelopment (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed)/ OE of genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of each miRNA/TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of all genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) along with KO of each miRNA and each TF regulating GSK3B interactome

Simulations 1-4: KO of all genes positively regulating each stage of neurodevelopment (embryogenesis/ neurulation/ proliferation/ migration/ neurite growth/ synaptogenesis/ neurodevelopment (when GSK3B is expressed)/ neurodevelopment (when GSK3B is not expressed)) and KO of each miRNA/TF downregulated each same stage of neurodevelopment (0%)

a. Neurulation

Simulation 1: We observed overall regulation of neurulation in two different regulatory states (RS2), upregulation (100%) and downregulation (0%). KO of miR744 and KO of genes positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ migration (when GSK3B is expressed)/ neurite growth/ neurodevelopment (when GSK3B is expressed)/ proliferation/ synaptogenesis upregulated neurulation (100%). KO of each factor (except miR744) and KO of genes positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ migration (when GSK3B is expressed)/ neurite growth/ neurodevelopment (when GSK3B is expressed)/ proliferation/ synaptogenesis downregulated neurulation (0%).

Simulation 2: KO of all genes positively regulating neurulation and KO of each miRNA/TF, downregulated neurulation (0%).

Simulation 3: We observed overall regulation of neurulation in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of miR15b/ CREB1 and KO of genes positively regulating embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation/ neurodevelopment (when GSK3B is not expressed)/ synaptogenesis, downregulated neurulation (0%). KO of each miRNA/TF (except

KO of miR15b and CREB1) and KO of genes embryogenesis/ migration (when GSK3B is not expressed)/ neurite growth/ proliferation/ neurodevelopment (when GSK3B is not expressed)/ synaptogenesis, regulated neurulation (between 0% - 100%).

Simulation 4: We observed overall regulation of neurulation in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of AR and KO of genes positively regulating embryogenesis/ proliferation/ synaptogenesis regulated neurulation (between 0% - 100%). KO of each miRNA/TF (except AR) and KO of genes positively regulating embryogenesis/ proliferation/ synaptogenesis downregulated neurulation (0%).

[illegible]

Table 4: Perturbation results of GSK3B interactome involved in prenatal development (Contd)

[illegible]

neurulation and factors	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is not expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	RS4	100%	RS4	RS4	0%	0%	0%	0%

	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; 100% - upregulation; 0% - downregulation

Summary of perturbation results of GSK3B interactome involved in prenatal regulation

PC1 (Simulations 1, 2 and 4): Perturbation of each factor regulating GSK3B interactome showed that none of the factor was individually involved in upregulation of stages of development.

Simulation 3: Perturbation of each factor showed neurulation to be regulated in all three different regulatory states (RS1). Overall regulation of neurulation was shown to be regulated in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% as well downregulation (0%).

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development regulated by GSK3B interactome along with OE of all factors regulating GSK3B interactome, upregulated each same stage of development. OE of genes positively regulating migration (when GSK3B is expressed) along with OE of all factors, upregulated (100%) synaptogenesis and neurodevelopment (when GSK3B is expressed). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed), upregulated (100%) all the stages except neurodevelopment (when GSK3B is expressed) and migration (when GSK3B is expressed). Similarly, OE of genes positively regulating neurodevelopment (when GSK3B is expressed) upregulated migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed) (100%).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development regulated by GSK3B interactome along with KO of factors regulating GSK3B interactome, downregulated each same stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development regulated by GSK3B interactome (except neurulation, proliferation and embryogenesis) along with OE of each miRNA and each TF upregulated each same stage of development

(100%). OE of genes positively regulating migration (when GSK3B is expressed) along with OE of each factor, upregulated (100%) synaptogenesis and neurodevelopment (when GSK3B is expressed).

Simulations 1 and 4: OE of genes positively regulating neurulation along with OE of each factor, showed overall regulation of neurulation in two different regulatory states (RS4), upregulation (100%) and regulation b/w 0%-100%.

Simulation 3: OE of genes positively regulating neurulation/ proliferation along with OE of each factor, showed regulation of each same stage in three different regulatory states (RS1), upregulation (100%), regulation b/w 0%-100% as well as downregulation (0%). OE of genes positively regulating embryogenesis along with OE of each factor, showed overall regulation of embryogenesis in two different regulatory states (RS4), upregulation (100%) and regulation b/w 0%-100%.

At simulations 1,2 and 4, perturbation of genes involved in proliferation/ embryogenesis along with perturbation of each factor showed upregulation of embryogenesis and proliferation (100%). Similarly, at simulation 2, perturbation of genes involved in neurulation along with perturbation of each factor showed upregulation of neurulation (100%).

Simulations 1, 3 and 4: OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with OE of each factor, showed overall regulation of neurodevelopment process in two different regulatory states (RS4), upregulation (100%) and regulation b/w 0%-100%.

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development regulated by GSK3B interactome (except neurulation and neurodevelopment (when GSK3B is not expressed)) along with KO of each miRNA and each TF downregulated each same stage of development (0%).

Simulations 1, 3 and 4: KO of genes positively regulating neurulation/ neurodevelopment (when GSK3B is not expressed) regulated by GSK3B interactome along with KO of each miRNA and each TF showed overall regulation of neurulation process in more than one regulatory state, RS2 (downregulation (0%) and upregulation (100%)) at simulation 1, RS3 (regulation b/w 0%-100% and downregulation (0%)) at simulations 3 and 4. The overall neurodevelopment process was shown to be affected by genes regulating neurulation.

At simulation 2, KO of genes positively regulating neurulation/ neurodevelopment (when GSK3B is not expressed) regulated by GSK3B interactome showed downregulation of both the stages (0%). Summary of the NP results for GSK3B interactome is given in Table 3 and Table 4.

C. BDNF interactome

During prenatal development, BDNF interactome regulates proliferation, migration, differentiation, neurite growth and synaptogenesis. Self-regulated NOS1 downregulates differentiation, whereas NOS1 expression which is regulated by BDNF, upregulates differentiation. So, two different Boolean rules have been given for differentiation and the overall neurodevelopment process (all the stages regulated by BDNF combined as one stage).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating BDNF interactome

Simulations 1-4: OE and KO of each miRNA and each TF downregulated each of the above-mentioned stages of neurodevelopment and the overall neurodevelopment process regulated by BDNF interactome.

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) and OE of miRNAs and TFs regulating BDNF interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment

(proliferation/migration/ differentiation (when NOS1 is regulated by BDNF) / differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis) along with OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment (when NOS1 is self-regulated) along with OE of miRNAs and TFs, upregulated (100%) each stage of development. OE of genes positively regulating neurodevelopment (when NOS1 is regulated by BDNF) along with OE of miRNAs and TFs upregulated each stage of development (100%) but did not upregulate neurodevelopment process (when NOS1 is self-regulated).

a. Proliferation

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment

migration/ (differentiation (when NOS1 is regulated by BDNF) / differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis) as well as OE of genes positively regulating overall neurodevelopment process (neurodevelopment (when NOS1 is regulated by BDNF), neurodevelopment (when NOS1 is self-regulated) along with OE of miRNAs and TFs, upregulated proliferation (100%).

a. Migration

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment

proliferation/ (differentiation (when NOS1 is regulated by BDNF) / differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis) as well as OE of genes positively regulating overall neurodevelopment process (neurodevelopment (NOS1 is regulated by

BDNF)), neurodevelopment (when NOS1 is self-regulated) along with OE of miRNAs and TFs, upregulated proliferation (100%).

c. Differentiation (when NOS1 is regulated by BDNF)

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis) as well as OE of genes positively regulating overall neurodevelopment process (neurodevelopment (when NOS1 is regulated by BDNF), neurodevelopment (when NOS1 is self-regulated) along with OE of miRNAs and TFs, upregulated differentiation (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) and KO of miRNAs and TFs regulating BDNF interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ differentiation (when NOS1 is regulated by BDNF)/ differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis/ neurodevelopment (when NOS1 is regulated by BDNF)/ neurodevelopment (when NOS1 is self-regulated)) and KO of miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

Perturbation condition 4: OE of all genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) along with OE of each factor (miRNA/TF) regulating BDNF interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ migration/ differentiation (when NOS1 is regulated by BDNF)/ differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis/ neurodevelopment (when NOS1 is self-regulated)/ neurodevelopment (when NOS1 is regulated by BDNF) along with OE of each miRNA/TF upregulated proliferation (100%).

b. Migration

Simulations 1-4: OE of genes positively regulating proliferation/ migration/ differentiation (when NOS1 is regulated by BDNF)/ differentiation (when NOS1 is self-regulated)/ neurite growth/ synaptogenesis/ neurodevelopment (when NOS1 is self-regulated)/ neurodevelopment (when NOS1 is regulated by BDNF) along with OE of each miRNA/TF upregulated proliferation (100%).

c. Differentiation (when NOS1 is regulated by BDNF)

Simulations 1-4: OE of genes positively regulating differentiation (when NOS1 is regulated by BDNF)/ differentiation (when NOS1 is self-regulated)/ neurite growth/ proliferation/ migration/ synaptogenesis/ neurodevelopment (when NOS1 is regulated by BDNF)/ neurodevelopment (when NOS1 is self-regulated) and OE of each miRNA/TF upregulated differentiation (100%).

d. Differentiation (when NOS1 is self-regulated)

Simulations 1-4: OE of genes positively regulating differentiation (when NOS1 is self-regulated) / neurodevelopment (when NOS1 is self-regulated) and OE of each miRNA/TF, upregulated differentiation (100%).

Simulation 3: We observed overall regulation of differentiation in all three regulatory states (RS1), upregulation (0%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating differentiation (when NOS1 is regulated by BDNF)/ neurite growth/ proliferation/ migration/ neurodevelopment (when NOS1 is regulated by BDNF)/ synaptogenesis along with OE of miR22/ SOX2 regulated differentiation (between 0%-100%). OE of genes positively regulating differentiation (when NOS1 is regulated by BDNF)/ neurite growth/ proliferation/ migration/ neurodevelopment (when NOS1 is regulated by BDNF)/ synaptogenesis and OE of miR149/ miR4728/ miR5698/ miR625/ miR6785/ miR6825/ miR6883, downregulated differentiation (0%). However, OE of genes positively

regulating differentiation (when NOS1 is regulated by BDNF)/ neurite growth/ proliferation/ migration/ neurodevelopment (when NOS1 is regulated by BDNF)/ synaptogenesis and OE of each miRNA/TF (except OE of miR149/ miR4728/ miR5698/ miR625/ miR6785/ miR6825/ miR6883/ miR22/ SOX2), upregulated differentiation (100%).

e. Neurite growth

Simulations 1-4: OE of genes positively regulating neurite growth/ neurodevelopment (when NOS1 is regulated by BDNF)/ neurodevelopment (when NOS1 is self-regulated) along with OE of each miRNA/each TF, upregulated neurite growth (100%).

f. Synaptogenesis

Simulations 1-4: OE of genes positively regulating synaptogenesis/ neurodevelopment (when NOS1 is regulated by BDNF)/ neurodevelopment (when NOS1 is self-regulated) along with OE of each miRNA/each TF, upregulated synaptogenesis (100%).

g. Neurodevelopment (when NOS1 is regulated by BDNF)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when NOS1 is regulated by BDNF)/ genes positively regulating neurodevelopment (when NOS1 is self-regulated) and OE of each miRNA/TF, upregulated neurodevelopment (100%).

h. Neurodevelopment (when NOS1 is not expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when NOS1 is self-regulated) and OE of each miRNA/ TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) along with KO of each miRNA and each TF regulating BDNF interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ differentiation/ neurite growth/ synaptogenesis/ neurodevelopment (when NOS1 is regulated by BDNF)/ neurodevelopment (when NOS1 is

self-regulated)) and KO of each miRNA/ TF, downregulated each same stage of neurodevelopment (0%).

Table 5: Perturbation results of BDNF interactome involved in prenatal development

Perturbed stages	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)															
		Proliferation				Migration				Differentiation (when NOS1 is regulated by BDNF)				Differentiation (when NOS1 self-regulated)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	RS1	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration and factors		100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
		0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
		100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	RS1	0%
		0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating differentiation (when NOS1 is regulated by BDNF) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	RS1	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating differentiation (when NOS1 is self-regulated) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	RS1	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%

[illegible]

Table 6: Perturbation results of BDNF interactome involved in prenatal development (Contd)

[illegible]

Genes positively regulating differentiation (when NOS1 is regulated by BDNF) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating differentiation (when NOS1 is self-regulated) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when NOS1 is regulated by BDNF) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when NOS1 is self-regulated) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; Upregulation (100), downregulation (0%)

Summary of perturbation results of BDNF interactome involved in prenatal development

PC1 (Simulations 1-4): Each stage of neurodevelopment regulated by BDNF interactome was shown to be downregulated (0%), showing the individual regulatory effect of each factor on each stage of neurodevelopment. As each factor is involved in FFLs, perturbation of each factor, affects the expression of either miRNA or TF, which in turn affects gene expression. But no factor was shown to be involved in upregulation of the any one of the stages of neurodevelopment regulated by BDNF interactome.

PC2 (Simulations 1-4): OE of genes positively regulating each stage of neurodevelopment along with OE of miRNAs and TFs regulating BDNF interactome, upregulated each same stage of development. In addition, differentiation (when NOS1 is regulated by BDNF), proliferation and migration stages were also shown to be upregulated (100%), when genes positively regulating other stages were OE along with factors regulating BDNF interactome. OE of genes positively regulating neurodevelopment (when NOS1 is self-regulated) along with OE of all factors, upregulated each stage of development. But OE of genes positively regulating neurodevelopment (when NOS1 is regulated by BDNF) along with OE of factors upregulated each stage (100%) except differentiation (when NOS1 is self-regulated) and neurodevelopment (when NOS1 is self-regulated).

PC3 (Simulations 1-4): *KO of genes positively regulating each stage of development along with KO of miRNAs and TFs regulating BDNF interactome, upregulated each same stage of development.*

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each factor regulating BDNF interactome, upregulated (100%) each same stage of development. In addition, differentiation (when NOS1 is regulated by BDNF), proliferation and migration stages were also shown to be upregulated (100%), when genes positively regulating other stages were OE along with each factor regulating BDNF interactome. OE of

BDNF in other stages, upregulated proliferation, migration and differentiation stages (when NOS1 is regulated by BDNF). OE of genes positively regulating neurodevelopment (when NOS1 is self-regulated) along with OE of each factor, upregulated each stage of development. But OE of genes positively regulating neurodevelopment (when NOS1 is regulated by BDNF) along with OE of each factor upregulated each stage (100%) except differentiation (when NOS1 is self-regulated) and neurodevelopment (when NOS1 is self-regulated).

Simulation 3: OE of genes positively regulating each stage (except differentiation (when NOS1 is self-regulated) and neurodevelopment (when NOS1 is self-regulated) along with OE of each factor, regulated overall differentiation (when NOS1 is self-regulated) process in all three different regulatory states (RS1), upregulation, downregulation and regulation between 0%-100%. As TF downregulates miRNA expression in simulation 3, NGFR expression is either regulated between 0%-100% or upregulated, which in turn either regulates (b/w 0%-100%) or upregulates (100%) differentiation (when NOS1 is self-regulated). NGFR expression is also downregulated as miRNA repress NGFR expression, downregulating (0%) differentiation (when NOS1 is self-regulated).

At simulations 1, 2 and 4, OE of genes positively regulating each stage (except differentiation (when NOS1 is self-regulated) and neurodevelopment (when NOS1 is self-regulated)) along with OE of each factor, downregulated differentiation (when NOS1 is self-regulated) (0%).

PC5 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each factor regulating BDNF interactome, downregulated (0%) each same stage

of development. Summary of the NP results for BDNF interactome is given in Table 5 and Table 6.

D. GRM5 interactome

GRM5 interactome regulates proliferation, differentiation and synaptogenesis during prenatal development.

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating GRM5 interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating GRM5 interactome downregulated (0%) each stage of neurodevelopment and the overall neurodevelopment process (all the stages regulated by GRM5 are combined as one stage).

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) and OE of miRNAs and TFs regulating GRM5 interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation/ differentiation/ synaptogenesis) and OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment and OE of miRNAs and TFs, upregulated all the stages of neurodevelopment (100%).

a. Proliferation

Simulations 1-4: OE of genes positively regulating differentiation/ neurodevelopment and OE of miRNAs and TFs, upregulated proliferation (100%).

b. Differentiation

Simulations 1-4: OE of genes positively regulating proliferation/ neurodevelopment and OE of miRNAs and TFs, upregulated differentiation (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) and KO of miRNAs and TFs regulating GRM5 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ differentiation/ synaptogenesis) and KO of miRNAs and TFs, downregulated each stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) along with OE of each factor (miRNA/TF) regulating GRM5 interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ differentiation/ neurodevelopment and OE of each miRNA and each TF, upregulated proliferation (100%).

Simulations 1, 3 and 4: Synaptogenesis was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating synaptogenesis and OE of miR4719 regulated proliferation (between 0%-100%). OE of genes positively regulating synaptogenesis and OE of each factor (except OE of miR4719) downregulated proliferation (0%).

b. Differentiation

Simulations 1-4: OE of genes positively regulating proliferation/ differentiation/ neurodevelopment and OE of each miRNA and each TF, upregulated differentiation (100%).

Simulations 1, 3 and 4: Overall differentiation was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating synaptogenesis and OE of miR4719 regulated differentiation (between 0%-100%). OE of genes positively regulating differentiation and OE of each factor (except OE of miR4719) downregulated differentiation (0%).

c. Synaptogenesis

Simulations 1-4: OE of genes positively regulating synaptogenesis/ neurodevelopment and OE of each miRNA and each TF, upregulated synaptogenesis (100%).

d. Neurodevelopment

Simulations 1-4: OE of genes positively regulating neurodevelopment and OE of each miRNA and each TF, upregulated all the stages of neurodevelopment (100%).

Simulations 1, 3 and 4: Neurodevelopment was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating neurodevelopment and OE of miR4719 regulated neurodevelopment (between 0%-100%). OE of genes positively regulating neurodevelopment and OE of each factor (except OE of miR4719) downregulated differentiation (0%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) along with KO of each factor (miRNA/ TF) regulating GRM5 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ neurite growth/ synaptogenesis/ neurodevelopment) and KO of each miRNA/ TF, downregulated each stage of neurodevelopment (0%).

Table 7: Perturbation results of GRM5 interactome involved in prenatal neurodevelopment

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)															
		Proliferation				Differentiation				Synaptogenesis				Neurodevelopment			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating differentiation and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	RS3	0%	RS3	RS3	RS3	0%	RS3	RS3	100%	100%	100%	100%	RS3	0%	RS3	RS3
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; RS3 represents regulatory state 3, where PC leads to downregulation (0%) and regulation between 0%-100%.

Summary of node perturbation results of GRM5 interactome involved in prenatal development

PC1 (Simulations 1-4): Perturbation of each factor showed downregulation (0%) of each stage of development, regulated by GRM5 interactome.

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of miRNAs and TFs regulating GRM5 interactome, upregulated each same stage of development (100%). OE of genes positively regulating differentiation along with OE of all factors upregulated proliferation (100%). OE of genes positively regulating proliferation along with OE of all factors upregulated differentiation (100%). Genes GRM5, ERK1, ERK2 are involved in positive regulation of both proliferation and differentiation stages, so OE of genes positively regulating either proliferation/ differentiation along with OE of all factors, upregulates both stages.

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of all factors, downregulated each same stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each factor, upregulated each same stage of development (100%).

Simulations 1, 3 and 4: OE of genes positively regulating synaptogenesis along with OE of each factor, regulated each stage of development (except synaptogenesis) in more than one regulatory state (RS3), regulation between 0%-100% and downregulation (0%). OE of miR4719 along with OE of genes positively regulating synaptogenesis regulates GRM5 expression, in turn regulating each stage between 0%-100% (except synaptogenesis). OE of each factor (except OE of miR4719) along with OE of genes positively regulating synaptogenesis, downregulated each stage (0%).

At simulation 2, OE of genes positively regulating synaptogenesis along with OE of each factor, downregulated (0%) each stage of development (except synaptogenesis).

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor, downregulated each same stage of development (0%). Summary of the NP results for GRM5 interactome is given in Table 7.

E. NRG1 interactome

NRG1 interactome regulates proliferation, migration, neurite growth and synaptogenesis during prenatal development. NRG1 gene was shown to be regulated by miR124, in turn miR124 has been shown to be regulated by TFs, STAT3, REST, EGR1 and TP53 (TF-miRNA FFLs). The direction of regulation of miRNA by these four TFs is known from experimental evidence, where STAT3 and REST represses miR124 expression and EGR1 and TP53 activates miR124 expression. So, rule for miR124 is same across all four simulations. Furthermore, the Boolean rules for simulation 1 and 3 also remains same, as TF upregulates gene expression in both simulations and the TF regulation of miRNA expression remains same in both the simulations. Similarly, Boolean rules for simulations 2 and 4 are same, as TF downregulates gene expression in both the simulations and the TF regulation of miRNA expression remains same in both the simulations.

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating NRG1 interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating NRG1 interactome, downregulated (0%) each stage of neurodevelopment and the overall neurodevelopment process (all the stages regulated by NRG1 interactome were combined into one stage).

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) and OE of miRNAs and TFs regulating NRG1 interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ neurite growth/ synaptogenesis) and OE of miRNAs and TFs, upregulated each stage of neurodevelopment (100%). OE of genes positively regulating overall neurodevelopment and OE of miRNAs and TFs, upregulated the above-mentioned stages of neurodevelopment (100%).

a. Proliferation

Simulations 1-4: OE of genes positively regulating migration/ neurite growth/ synaptogenesis/ neurodevelopment and OE of miRNAs and TFs, upregulated proliferation (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) and KO of miRNAs and TFs regulating NRG1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ neurite growth/ synaptogenesis) and KO of miRNAs and TFs, downregulated each stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) along with OE of each miRNA and each TF regulating NRG1 interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ migration/ neurite growth/ synaptogenesis/ overall neurodevelopment process and OE of each miRNA/TF, upregulated proliferation (100%).

b. Migration

Simulations 1-4: OE of genes positively regulating migration/neurodevelopment and OE of each miRNA/TF, upregulated migration (100%).

Simulations 1 and 3: We observed overall regulation of migration process in all three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of miR124 and OE of genes positively regulating proliferation/ neurite growth/ synaptogenesis, downregulated migration (0%). Similarly, OE of TP53 and OE of genes positively regulating proliferation/ neurite growth/ synaptogenesis regulated migration (between 0%-100%). OE of each factor (except miR124/TP53) and OE of genes positively regulating proliferation/ neurite growth/ synaptogenesis upregulated migration (100%).

c. Neurite growth

Simulations 1-4: OE of genes positively regulating neurite growth/neurodevelopment and OE of each miRNA and each TF, upregulated neurite growth (100%).

d. Synaptogenesis

Simulations 1-4: OE of genes positively regulating synaptogenesis/neurodevelopment and OE of each miRNA/TF, upregulated synaptogenesis (100%).

e. Neurodevelopment

Simulations 1-4: OE of genes positively regulating neurodevelopment and OE of each miRNA/TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) along with KO of each miRNA and each TF regulating NRG1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ neurite growth/ synaptogenesis/ neurodevelopment) and KO of each miRNA/TF, downregulated each same stage of neurodevelopment (0%).

Table 8: Perturbation results of NRG1 interactome involved in prenatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t=150$)											
		Proliferation				Migration				Neurite growth			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	RS1	0%	RS1	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	RS1	0%	RS1	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	RS1	0%	RS1	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; RS1 represents regulatory state RS1, where PC1 leads to upregulation (100%), downregulation (0%) and regulation between 0%-100%

Table 9: Perturbation results of NRG1 interactome involved in prenatal development (contd)

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t=150$)							
		Synaptogenesis				Neurodevelopment			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%

	4	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment and factors	2	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%

Summary of node perturbation results of NRG1 interactome involved in prenatal development

PC1 (Simulations 1-4): Perturbation of each factor downregulated each stage of development (0%).

PC2 (simulations 1-4): OE of each gene positively regulating each stage of development along with OE of all factors, upregulated each same stage of development (100%) as well as proliferation stage (100%).

PC3 (Simulations 1-4): KO of each gene positively regulating each stage of development along with KO of all factors, downregulated each stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each factor, upregulated (100%) each same stage of development.

Simulations 1 and 3: OE of genes positively regulating each stage, overall regulated migration in three regulatory states (RS1), upregulation (100%) regulation between 0%-100% and downregulation (0%). NRG1 OE in other stages upregulated migration and when miRNA downregulates TF expression, ERBB2 expression is regulated, in turn regulating migration between 0%-100%. When miRNA repress gene expression, migration is downregulated (0%).

Simulations 2 and 4: OE of genes positively regulating each stage, downregulated migration (0%), as TF downregulates gene expression at simulations 2 and 4.

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor, downregulated (0%) each same stage of development. Summary of the NP results for NRG1 interactome is given in Table 8 and Table 9.

F. YWHAE interactome

During prenatal development, YWHAE interactome regulates migration and neurite growth. YWHAE regulates migration at 2 different conditions, when GSK3B is expressed as well as when GSK3B is not expressed. So, two different Boolean rules have been given for migration as well as the overall neurodevelopment process.

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating YWHAE interactome

Simulations 1-4: OE and KO of each miRNA and each TF downregulated (0%) the above-mentioned stages of neurodevelopment and the overall neurodevelopment process (all the stages regulated by YWHAE interactome were combined as one stage)

Multiple node perturbation

Perturbation condition 2: OE of genes regulating each stage of neurodevelopment regulated by YWHAE interactome and OE of miRNAs and TFs regulating YWHAE interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (migration and neurite growth) and OE of miRNAs and TFs, upregulated (100%) each same stage of neurodevelopment. OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of miRNAs and TFs upregulated (100%) migration (when GSK3B is not expressed) and neurodevelopment (when GSK3B is not expressed). OE of genes positively regulating neurodevelopment (when GSK3B is expressed) upregulated

(100%) migration (when GSK3B is expressed), neurite growth and neurodevelopment (when GSK3B is expressed)

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment regulated by YWHAE interactome and KO of miRNAs and TFs regulating YWHAE interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (migration/ neurite growth/ neurodevelopment) and KO of miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) along with OE of each miRNA and each TF regulating YWHAE interactome

a. Migration (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is expressed)/ neurodevelopment (when GSK3B is expressed) and OE of each miRNA/ TF, upregulated migration (100%).

b. Migration (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating migration (when GSK3B is not expressed)/ neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF, upregulated migration (100%).

c. Neurite growth

Simulations 1-4: OE of genes positively regulating neurite growth/ neurodevelopment (when GSK3B is expressed) and OE of each miRNA/TF, upregulated neurite growth (100%).

d. Neurodevelopment (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GSK3B is expressed) and OE of each miRNA/TF, upregulated neurodevelopment (100%).

e. Neurodevelopment (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GSK3B is not expressed)/ migration (when GSK3B is not expressed) and OE of each miRNA/TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) along with KO of each miRNA and each TF regulating YWHAE interactome

Simulations 1-4: KO of all genes positively regulating each stage of neurodevelopment (migration and neurite growth) along with KO of each miRNA/TF, downregulated each same stage of neurodevelopment (0%).

Table 10: Perturbation results of YWHAE interactome involved in prenatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)																			
		Migration (when GSK3B is expressed)				Migration (when GSK3B is not expressed)				Neurite growth				Neurodevelopment (when GSK3B is expressed)				Neurodevelopment (when GSK3B is not expressed)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating migration (when GSK3B is not expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is not expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; Upregulation (100%); Downregulation (0%)

Summary of node perturbation results of YWHAЕ interactome regulating prenatal development

PC1 (Simulations 1-4): Perturbation of each factor downregulated each stage of neurodevelopment (0%).

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development and OE of all factors, upregulated each same stage of development (100%). OE of genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of factors upregulated (100%) neurite growth, migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with OE of factors upregulated migration (when GSK3B is not expressed) and neurodevelopment (when GSK3B is not expressed) (100%).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of all factors downregulated each same stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development and OE of each factor, upregulated each same stage of development (100%). OE of genes positively regulating neurodevelopment (when GSK3B is expressed) along with OE of each factor upregulated (100%) neurite growth, migration (when GSK3B is expressed) and neurodevelopment (when GSK3B is expressed). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with OE of each factor upregulated migration (when GSK3B is not expressed) and neurodevelopment (when GSK3B is not expressed) (100%).

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor downregulated each same stage of development (0%).

Summary of the NP results for YWHAЕ interactome is given in Table 10.

1.2 Interactomes involved in regulation of postnatal development

We performed single node and multiple node perturbation analyses in the postnatal network. We could not perform node perturbation analysis for APs: CNTNAP2 and DTNBP1.

As no miRNAs were shown to regulate DTNBP1, we could not curate miRNA-TF regulatory loops for DTNBP1 gene. The FFLs regulating CNTNAP2 were not shown to regulate the genes interacting with CNTNAP2 gene.

A. DISC1 interactome

During postnatal development, DISC1 interactome regulates proliferation, migration, differentiation, neurite growth and synaptogenesis. We have more than 1 Boolean rule for the following stages of development: neurite growth, synaptogenesis and the overall neurodevelopment process (all the stages are combined as one stage). Neurite growth is upregulated, when DISC1 is expressed (rule 1) and also when DISC1 is not expressed (rule 2).

In synaptogenesis, PDE4B gene plays a major role along with DISC1 in upregulation of synaptogenesis. Synaptogenesis is upregulated, when DISC1 downregulates PDE4B expression (rule 1) and when DISC1 upregulates PDE4B gene expression (rule 2). We have given another separate rule for synaptogenesis, where NRG1 and ERBB4 interacts and upregulates synaptogenesis (rule 3). DISC1 is not included in this rule, as DISC1 inhibits ERBB4 expression. So, we also have given 2 Boolean rules for neurodevelopment, when DISC1 is expressed and downregulates PDE4B expression (rule 1) and when DISC1 is expressed and upregulates PDE4B expression (rule 2). We have not combined the regulation of synaptogenesis by NRG1 and ERBB4 with other stages of development, as ERBB4 is inhibited by DISC1.

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating DISC1 interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating DISC1 interactome downregulated (0%) proliferation, migration, differentiation, neurite growth (when DISC1 is expressed), synaptogenesis (when PDE4B is downregulated), synaptogenesis (when PDE4B is upregulated), neurodevelopment (when PDE4B is upregulated) and neurodevelopment (when PDE4B is downregulated).

a. Neurite growth (when DISC1 is not expressed)

Simulations 1 and 3: OE and KO of each miRNA and each TF (except OE of HNF4A or miR124), upregulated (100%) neurite growth (when DISC1 is not expressed). OE of HNF4A or miR124 downregulated (0%) neurite growth (when DISC1 is not expressed).

Simulation 1: OE of HNF4A downregulated (0%) neurite growth (when DISC1 is not expressed).

Simulation 3: OE of HNF4A regulated neurite growth (when DISC1 is not expressed) b/w 0%-100%.

At simulation 1, we observed overall regulation of neurite growth in two different regulatory states (RS2), upregulation (100%) and downregulation (0%).

At simulation 3, we observed overall regulation of neurite growth in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%).

Simulations 2 and 4: OE and KO of each miRNA and each TF, downregulated (0%) neurite growth (when DISC1 is not expressed).

b. Synaptogenesis (when NRG1 is expressed)

Simulations 1 and 3: We observed overall regulation of synaptogenesis in two different regulatory states (RS1), upregulation (100%), downregulation (0%) and regulation between 0%-100%. OE and KO of each miRNA and each TF (except OE of HNF4A or miR124 and KO of AR), upregulated (100%) synaptogenesis (when NRG1 is expressed). OE of HNF4A or miR124 downregulated (0%) synaptogenesis (when NRG1 is expressed). KO of AR regulated synaptogenesis (when NRG1 is expressed) between 0%-100%.

Simulations 2 and 4: We observed overall regulation of synaptogenesis in two different regulatory states (RS4), downregulation (0%) and regulation between 0%-100%. OE and KO of each miRNA and each TF (except AR KO), downregulated (0%) synaptogenesis (when NRG1 is expressed). KO of AR showed regulation of synaptogenesis (when NRG1 is expressed) between 0%-100%.

Multiple node perturbation analysis

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) and OE of miRNAs and TFs regulating DISC1 interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation/ migration/differentiation/ neurite growth (when DISC1 is expressed)/ neurite growth (when DISC1 is not expressed)/ synaptogenesis (when PDE4B is downregulated)/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated)) and OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment (when PDE4B is upregulated)/ genes positively regulating neurodevelopment (when PDE4B is downregulated) and OE of miRNAs and TFs, upregulated (100%) all the stages of

development regulated by DISC1 interactome, except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed) stages.

OE of synaptogenesis (when PDE4B is upregulated) along with OE of miRNAs and TFs upregulated (100%) synaptogenesis (when PDE4B is downregulated) and similarly OE of genes positively regulating synaptogenesis (when PDE4B is downregulated) along with OE of miRNAs and TFs upregulated (100%) synaptogenesis (when PDE4B is upregulated).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) and KO of miRNAs and TFs regulating DISC1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ differentiation/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is downregulated)/ synaptogenesis (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated)) and KO of miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

a. Neurite growth (when DISC1 is not expressed)

Simulations 1 and 3: KO of genes positively regulating proliferation, migration, differentiation, synaptogenesis (when PDE4B is upregulated), synaptogenesis (when PDE4B is downregulated), synaptogenesis (when NRG1 is expressed) and KO of miRNAs and TFs, downregulated (0%) neurite growth (when DISC1 is not expressed).

Simulations 2 and 4: KO of genes positively regulating proliferation, migration, differentiation, synaptogenesis (when PDE4B is upregulated), synaptogenesis (when PDE4B is downregulated), synaptogenesis (when NRG1 is expressed) and KO of miRNAs and TFs, upregulated (100%) neurite growth (when DISC1 is not expressed).

b. Synaptogenesis (when NRG1 is expressed)

Simulations 1 and 3: KO of genes positively regulating proliferation, migration, differentiation, synaptogenesis (when PDE4B is upregulated), synaptogenesis (when PDE4B is downregulated), neurite growth (when DISC1 is not expressed), neurite growth (when DISC1 is expressed) and KO of miRNAs and TFs, downregulated (0%) synaptogenesis (when NRG1 is expressed).

Simulations 2 and 4: KO of genes positively regulating proliferation, migration, differentiation, synaptogenesis (when PDE4B is upregulated), synaptogenesis (when PDE4B is downregulated), neurite growth (when DISC1 is not expressed), neurite growth (when DISC1 is expressed) and KO of miRNAs and TFs, upregulated (100%) synaptogenesis (NRG1 is expressed).

Perturbation condition 4: OE of all genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) along with OE of each factor (miRNA/TF) regulating DISC1 interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is upregulated) along with OE of each miRNA/ TF, upregulated proliferation (100%).

Simulation 3: Proliferation was shown to be regulated in three different regulation states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating differentiation/ migration/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated) and OE of HNF4A regulates proliferation (between 0%-100%).

OE of genes positively regulating differentiation/ migration/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated) and OE of miR744/ miR155/ miR124 downregulated proliferation (0%).

OE of genes positively regulating differentiation/ migration/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated) and OE of each miRNA /TF (except OE of HNF4A/ miR124/ miR155/ miR744), upregulated proliferation (100%).

b. Migration

Simulations 1-4: OE of genes positively regulating proliferation/ migration/ differentiation/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/ TF, upregulated migration (100%).

c. Differentiation

Simulations 1-4: OE of genes positively regulating proliferation/ migration/ differentiation/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when DISC1 is expressed)/ synaptogenesis (when DISC1 is not expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/ TF, upregulated differentiation (100%).

d. Neurite growth (when DISC1 is expressed)

Simulations 1-4: OE of genes positively regulating neurite growth (when DISC1 is expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/TF, upregulated neurite growth (100%).

e. Neurite growth (when DISC1 is not expressed)

Simulations 1-4: OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of each miRNA/ TF regulating DISC1 interactome, upregulated neurite growth (100%).

Simulation 1: OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of HNF4A downregulated neurite growth (0%).

Simulation 3: OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of HNF4A regulated neurite growth (between 0%-100%).

Simulations 1 and 3: OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of miR124 downregulated neurite growth (0%).

At simulation 1, we observed overall regulation of neurite growth in two different regulatory states (RS2), upregulation (100%) and downregulation (0%). OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of each factor (except miR124/ HNF4A) upregulated neurite growth (100%).

At simulation 3, we observed overall regulation of neurite growth in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of each factor (except miR124/ HNF4A) upregulated neurite growth (100%).

f. Synaptogenesis (when PDE4B is upregulated)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/TF, upregulated synaptogenesis (100%).

g. Synaptogenesis (when PDE4B is downregulated)

Simulations 1-4: OE of genes positively regulating synaptogenesis (PDE4B is downregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/TF, upregulated synaptogenesis (100%).

h. Synaptogenesis (when NRG1 is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when NRG1 is expressed) along with OE of each miRNA/TF, regulated synaptogenesis (100%).

Simulation 1: OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of HNF4A downregulated synaptogenesis (0%).

Simulation 3: OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of HNF4A regulated synaptogenesis (between 0%-100%).

Simulations 1 and 3: OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of miR124 downregulated synaptogenesis (0%).

At simulation 1, we observed overall regulation of synaptogenesis in two different regulatory states (RS2), upregulation (100%) and downregulation (0%). OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of each factor (except miR124/ HNF4A) upregulated synaptogenesis (100%).

At simulation 3, we observed overall regulation of synaptogenesis in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating neurite growth (when DISC1 is not expressed) along with OE of each factor (except miR124/ HNF4A) upregulated synaptogenesis (100%).

i. Neurodevelopment (when PDE4B is upregulated)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/TF upregulated neurodevelopment (100%).

j. Neurodevelopment (when PDE4B is downregulated)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) along with OE of each miRNA/TF upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by DISC1 interactome) along with KO of each factor (miRNA/ TF) regulating DISC1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ migration/ differentiation/ neurite growth (when DISC1 is expressed)/ synaptogenesis (when PDE4B is downregulated)/ synaptogenesis (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated)) and KO of each miRNA/TF, downregulated each same stage of neurodevelopment (0%).

a. Neurite growth (when DISC1 is not expressed)

Simulations 1 and 3: KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ synaptogenesis (when NRG1 is expressed) and KO of each miRNA/TF, upregulated (100%) neurite growth (when DISC1 is not expressed).

Simulations 2 and 4: KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ synaptogenesis (when NRG1 is expressed) and KO of each miRNA/TF, downregulated (0%) neurite growth (when DISC1 is not expressed).

b. Synaptogenesis (when NRG1 is expressed)

Simulations 1 and 3: KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurite growth (when DISC1 is expressed)/ neurite growth (when DISC1 is not expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) and KO of each miRNA/TF (except AR KO), upregulated (100%) synaptogenesis (when NRG1 is expressed). KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurite growth (when DISC1 is expressed)/ neurite growth (when DISC1 is not expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) and KO of AR showed regulation of synaptogenesis (when NRG1 is expressed) between 0%-100%.

Simulations 2 and 4: KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurite growth (when DISC1 is expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) and KO of each miRNA/TF (except AR KO), downregulated (0%) synaptogenesis (when NRG1 is expressed). KO of genes positively regulating proliferation/ migration/ differentiation/ synaptogenesis (when PDE4B is upregulated)/ synaptogenesis (when PDE4B is downregulated)/ neurite growth (when DISC1 is expressed)/ neurodevelopment (when PDE4B is upregulated)/ neurodevelopment (when PDE4B is downregulated) and KO of AR showed regulation of synaptogenesis (when NRG1 is expressed) between 0%-100%.

Table 11: Perturbation results of DISC1 interactome involved in postnatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)																			
		Proliferation				Migration				Differentiation				Neurite growth (when DISC1 is expressed)				Neurite growth (when is DISC1 not expressed)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	RS2	0%	RS1	0%
Genes positively regulating proliferation and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%
Genes positively regulating migration and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%
Genes positively regulating differentiation and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%
	4	0%	0%	RS1	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%
Genes positively regulating neurite growth (when DISC1 is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	RS1	0%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth (when DISC1 is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when PDE4B is	2	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%

[illegible]

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; RS1 represents regulatory state 1, where PC leads to upregulation (0%), regulation between 0%-100% and downregulation (0%). RS2 represents regulatory state 2, where PC leads to regulation between 0%-100% and downregulation (0%)

Table 12: Perturbation results of DISC1 interactome involved in postnatal development (contd)

[illegible]

[illegible]

	5	0%	0%	0%	0%	0%	0%	0%	0%	RS4	RS3	RS4	RS3	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when NRG1 is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when PDE4B is expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	RS4	RS3	RS4	RS3	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when PDE4B is not expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	RS4	RS3	RS4	RS3	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4

Summary of node perturbation results for DISC1 interactome regulating postnatal development

PC1 (Simulations 1-4): Perturbation of each factor downregulated each stage (except neurite growth (when DISC1 is not expressed), synaptogenesis (when NRG1 is expressed)) of development (0%).

Simulation 1: Perturbation of each factor regulated neurite growth (when DISC1 is not expressed) in two different regulatory states (RS2), downregulation (0%) as well as upregulation (100%).

Simulation 3: Perturbation of each factor regulated neurite growth (when DISC1 is not expressed) in three different regulatory states (RS1), regulation between 0%-100%, downregulation (0%) as well as upregulation (100%).

At simulations 2 and 4, perturbation of each factor, downregulated (0%) neurite growth (when DISC1 is not expressed) and synaptogenesis (NRG1 is expressed).

Simulations 1 and 3: Perturbation of each factor regulated synaptogenesis (when NRG1 is expressed) in three different regulatory states (RS1), regulation between 0%-100%, downregulation (0%) as well as upregulation (100%).

Simulations 2 and 4: Perturbation of each factor regulated synaptogenesis (when NRG1 is expressed) in two different regulatory states (RS1), regulation between 0%-100%, and upregulation (100%).

PC2 (Simulations 1-4): OE of genes positively regulating each stage along with OE of factors upregulated each same stage of development (100%). OE of genes positively regulating each stage (except neurite growth (when DISC1 is not expressed)) and synaptogenesis (when NRG1 is expressed) along with OE of factors, upregulated migration and differentiation (100%). OE of genes positively regulating synaptogenesis (when PDE4B is upregulated) along with OE of factors, upregulated synaptogenesis (when PDE4B is downregulated) (100%). OE

of genes positively regulating synaptogenesis (when PDE4B is downregulated) along with OE of factors, downregulated synaptogenesis (when PDE4B is upregulated) (100%).

OE of genes positively regulating neurodevelopment (when PDE4B is upregulated) along with OE of factors, upregulated (100%) each stage of development (except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed)). Similarly, OE of genes positively regulating neurodevelopment (when PDE4B is downregulated) along with OE of factors, upregulated (100%) each stage of development (except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed)).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of factors, downregulated each stage of development (0%).

Simulations 2 and 4: KO of genes positively regulating each stage of development (except neurite growth (when DISC1 not expressed), neurite growth (DISC1 expressed), neurodevelopment (when PDE4B is upregulated) and neurodevelopment (when PDE4B is downregulated)) along with KO of all factors, upregulated neurite growth (when DISC1 is not expressed) (100%). Similarly, KO of genes positively regulating each stage of development (except synaptogenesis (when NRG1 is expressed)) and KO of all factors, upregulated synaptogenesis (when NRG1 is expressed) (100%).

PC4 (Simulations1-4): OE of genes positively regulating each stage along with OE of each factor upregulated each same stage of development (100%). OE of genes positively regulating each stage (except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed) along with OE of each factor, upregulated migration and differentiation (100%). OE of genes positively regulating synaptogenesis (PDE4B is upregulated) along with OE of each factor, upregulated synaptogenesis (when PDE4B is downregulated) (100%). OE of genes positively regulating synaptogenesis (when

PDE4B is downregulated) along with OE of each factor, downregulated synaptogenesis (when PDE4B is upregulated) (100%).

OE of genes positively regulating neurodevelopment (when PDE4B is upregulated) along with OE of each factor, upregulated (100%) each stage of development (except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed)).

Similarly, OE of genes positively regulating neurodevelopment (when PDE4B is downregulated) along with OE of each factor, upregulated (100%) each stage of development (except neurite growth (when DISC1 is not expressed) and synaptogenesis (when NRG1 is expressed)).

Simulation 1: OE of genes positively regulating neurite growth (when DISC1 is not expressed) and OE of each factor showed overall regulation of synaptogenesis (when NRG1 is expressed) in two different regulatory states (RS2), downregulation (0%) and upregulation (100%). OE of genes positively regulating synaptogenesis (when NRG1 is expressed) and OE of each factor regulated neurite growth (when DISC1 is not expressed) in two different regulatory states (RS2), downregulation (0%) and upregulation (100%).

Simulation 3: OE of genes positively regulating neurite growth (when DISC1 is not expressed) and OE of each factor showed overall regulation of synaptogenesis (when NRG1 is expressed) in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when NRG1 is expressed) and OE of each factor regulated neurite growth (when DISC1 is not expressed) in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%).

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor, downregulated each stage of development (0%).

Simulations 1 and 3: We observed regulation of synaptogenesis (when NRG1 is expressed) in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. KO of genes positively regulating each stage of development (except neurite growth (when DISC1 not expressed) along with KO of each factor (except AR KO), upregulated synaptogenesis (when NRG1 is expressed) (100%). KO of genes positively regulating each stage of development (except neurite growth (when DISC1 not expressed) and KO of each factor, showed upregulation of neurite growth (100%).

Summary of the NP results for DISC1 interactome is given in Table 11 and Table 12.

B. GSK3B interactome

GSK3B interactome regulates proliferation, neurite growth and synaptogenesis.

Synaptogenesis is upregulated, when GSK3B is expressed as well when GSK3B is not expressed, so two Boolean rules were given for the stage, synaptogenesis. All the stages of development regulated by GSK3B interactome were also combined as single stage, neurodevelopment.

Single gene perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating GSK3B interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating GSK3B interactome, downregulated (0%) neurite growth and the overall neurodevelopment process.

Simulation 2: OE and KO of each miRNA and each TF regulating GSK3B interactome, downregulated (0%) proliferation, neurite growth, synaptogenesis (when GSK3B is expressed), synaptogenesis (when GSK3B is not expressed) and the overall neurodevelopment process.

a. Proliferation

Simulation 1: Proliferation was shown to be regulated in two regulatory states (RS3), regulation between 0%-100% and downregulation (0%). AR KO regulates proliferation (between 0%-100%). OE of each miRNA/TF and KO of each miRNA/TF (except AR KO), downregulated proliferation (0%).

Simulation 3: Proliferation was shown to be regulated in two regulatory states (RS2), upregulation (100%) and downregulation (0%). OE of miR124/ miR744/miR155/ HNF4A downregulates proliferation (0%). OE of each miRNA/TF (except OE of miR124/ miR744/ miR155/ HNF4A) upregulates proliferation (100%). KO of each miRNA/TF upregulates proliferation (100%).

b. Synaptogenesis (when GSK3B is not expressed)

Simulation 1: Synaptogenesis was shown to be regulated in two regulatory states (RS3), regulation between 0%-100% and downregulation (0%). OE of miR124 and KO of AR or miR155 regulated synaptogenesis (between 0%-100%). OE of each miRNA/TF (except miR124 OE) and KO of each miRNA/TF (except KO of AR or miR155), downregulated synaptogenesis (0%).

Simulation 3: Synaptogenesis was shown to be regulated in three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). KO of TCF3 regulated synaptogenesis (between 0%-100%). KO of each factor (except TCF3 KO), upregulated synaptogenesis (100%). OE of miR124 or KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3 also regulated synaptogenesis (between 0%-100%). OE of miR155/ miR26b downregulates synaptogenesis (0%). OE of each miRNA/TF (except OE of miR124/ KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3/ miR155/ miR26b), upregulated synaptogenesis (100%).

Simulation 4: Synaptogenesis was shown to be regulated in two regulatory states (RS3), regulation between 0%-100% and downregulation (0%). KO of AR or miR155 regulates synaptogenesis (between 0%-100%). OE of miR124 regulates synaptogenesis (between 0%-100%). OE of each miRNA/TF (except miR124 OE) and KO of each miRNA/TF (except KO of AR / miR155), downregulated synaptogenesis (0%).

c. Synaptogenesis (when GSK3B is expressed)

Simulation 1: Synaptogenesis was shown to be regulated in two regulatory states (RS3), regulation between 0%-100% and downregulation (0%). KO of AR and OE of miR124 regulated synaptogenesis (between 0%-100%). OE of each miRNA/TF (except miR124 OE) and KO of each miRNA/TF (except KO of AR), downregulated synaptogenesis (0%).

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) and OE of miRNAs and TFs regulating GSK3B interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation/ neurite growth / synaptogenesis (when GSK3B is expressed)/ synaptogenesis (when GSK3B is not expressed)) and OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating overall neurodevelopment along with OE of miRNAs and TFs, upregulated neurite growth, proliferation, synaptogenesis (when GSK3B is not expressed) and neurodevelopment (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) and KO of miRNAs and TFs regulating GSK3B interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ neurite growth / synaptogenesis (when GSK3B is expressed)/ synaptogenesis

(when GSK3B is not expressed)) and KO of miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

a. Proliferation

Simulations 2 and 4: KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed) along with KO of miRNAs and TFs, upregulated proliferation (100%).

b. Synaptogenesis (when GSK3B is not expressed)

Simulations 2 and 4: KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed)/ proliferation along with KO of miRNAs and TFs, upregulated synaptogenesis (when GSK3B is not expressed) (100%).

c. Synaptogenesis (when GSK3B is expressed)

Simulations 2 and 4: KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) and KO of miRNAs and TFs upregulated synaptogenesis (when GSK3B is expressed) (100%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) along with OE of each factor (miRNA/ TF) regulating GSK3B interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF regulating GSK3B interactome, upregulated proliferation (100%).

Simulation 3: Genes regulating other stages regulated proliferation in two regulatory states (RS2), upregulation (100%) and downregulation (0%). OE of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is not expressed) and OE of miR124/ miR155/ HNF4A/ miR744, downregulated proliferation (0%). OE of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is not expressed) and OE of each miRNA/ TF (except

OE of miR124/ miR155/ HNF4A/ miR744) regulating GSK3B interactome, upregulated proliferation (100%).

b. Neurite growth

Simulations 1-4: OE of genes positively regulating neurite growth/ neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF, upregulated neurite growth (100%).

c. Synaptogenesis (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (GSK3B is not expressed)/ neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF, upregulated synaptogenesis (100%).

Simulations 1 and 4: Genes positively regulating neurite growth/ proliferation regulated synaptogenesis in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating neurite growth/ proliferation and OE of miR124, regulated synaptogenesis (between 0%-100%). OE of genes positively regulating neurite growth/ proliferation and OE of each factor (except OE of miR124), regulated synaptogenesis (between 0%-100%).

Simulation 3: Genes positively regulating neurite growth/ proliferation regulated synaptogenesis in three different regulatory states (RS1), upregulation (0%), downregulation (0%) and regulation between 0%-100%. OE of genes positively regulating neurite growth/ proliferation and OE of miR124/ KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3, regulated synaptogenesis (between 0%-100%). OE of genes positively regulating neurite growth/ proliferation and OE of miR155/ miR26b, downregulated synaptogenesis (0%). OE of genes positively regulating neurite growth/ proliferation and OE of each factor (except OE

of miR155/ miR26b/ miR124/ KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3) showed upregulation of synaptogenesis (100%).

d. Synaptogenesis (when GSK3B is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GSK3B is expressed) and OE of each miRNA/ TF, upregulated synaptogenesis (100%).

e. Neurodevelopment (when GSK3B is not expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) and OE of each miRNA/TF regulating GSK3B interactome upregulated neurodevelopment (100%).

Simulation 3: Neurodevelopment was shown to be regulated in three different regulatory states (RS1), upregulation (0%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating neurite growth and OE of KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3, regulated neurodevelopment (0%-100%). OE of genes positively regulating neurite growth and OE of miR124/ miR155/ miR26b, downregulated neurodevelopment (0%). OE of genes positively regulating neurite growth and OE of each miRNA/TF (except OE of KLF4/ miR15a/ miR16/ miR182/ miR744/ miR96/ STAT3/ miR124/ miR155/ miR26b), upregulated neurodevelopment (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by GSK3B interactome) along with KO of each factor (miRNA/ TF) regulating GSK3B interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation/ neurite growth / synaptogenesis (when GSK3B is expressed)/ synaptogenesis (when GSK3B is not expressed)) and KO of each miRNA/ TF regulating GSK3B interactome, downregulated each same stage of neurodevelopment (0%).

a. Proliferation

Simulation 1: Proliferation was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed)/ synaptogenesis (when GSK3B is not expressed) and KO of AR regulated proliferation (0%-100%). KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed)/ synaptogenesis (when GSK3B is not expressed) and KO of each miRNA/ TF (except AR KO) downregulated proliferation (0%).

b. Synaptogenesis (when GSK3B is not expressed)

Simulation 1: Synaptogenesis was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of genes positively regulating neurite growth/ proliferation/ synaptogenesis (when GSK3B is expressed) and KO of AR/ miR155 regulated synaptogenesis (0%-100%). KO of genes positively regulating neurite growth/ proliferation/ synaptogenesis (when GSK3B is expressed) and KO of each miRNA/TF (except KO of AR/ miR155) downregulated synaptogenesis (0%).

Simulation 3: Synaptogenesis was shown to be regulated in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. KO of genes positively regulating neurite growth/ proliferation and KO of TCF3 regulated synaptogenesis between 0%-100%. KO of genes positively regulating neurite growth/ proliferation and KO of each miRNA/TF (except KO of TCF3) upregulated synaptogenesis (100%). KO of genes positively regulating synaptogenesis (when GSK3B is expressed) and KO of each miRNA/TF, upregulated synaptogenesis (100%).

Simulation 4: Synaptogenesis was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of genes positively

regulating neurite growth/ proliferation and KO of AR/ miR155 regulated synaptogenesis (0%-100%). KO of genes positively regulating neurite growth/ proliferation and KO of each miRNA/TF (except AR/ miR155), downregulated synaptogenesis (0%).

c. Synaptogenesis (when GSK3B is expressed)

Simulation 1: Synaptogenesis was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. KO of genes positively regulating neurite growth/ proliferation/ neurodevelopment (when GSK3B is not expressed)/ synaptogenesis (when GSK3B is not expressed) and KO of AR regulated synaptogenesis (0%-100%). KO of genes positively regulating neurite growth/ proliferation/ neurodevelopment (when GSK3B is not expressed)/ synaptogenesis (when GSK3B is not expressed) and KO of each miRNA/TF (except AR KO) downregulated synaptogenesis (0%).

Simulations 2 and 4: KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) and KO of each miRNA/TF, upregulated synaptogenesis (100%).

Simulation 3: KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) and KO of each miRNA and each TF (except KO of TCF3), upregulated synaptogenesis (100%). KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) and KO of TCF3 regulated synaptogenesis (between 0%-100%).

Table 13: Perturbation results of GSK3B interactome involved in postnatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)																			
		Proliferation				Neurite growth				Synaptogenesis (when GSK3B is not expressed)				Synaptogenesis (when GSK3B is expressed)				Neurodevelopment (when GSK3B is not expressed)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	RS3	0%	RS2	0%	0%	0%	0%	0%	RS3	0%	RS1	RS3	RS3	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	RS3	0%	RS1	RS3	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	RS3	0%	RS4	RS3	RS3	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	RS2	0%	100%	100%	100%	100%	RS3	0%	RS1	RS3	0%	0%	0%	0%	0%	0%	RS1	0%
	5	RS3	0%	0%	0%	0%	0%	0%	0%	RS3	0%	RS4	RS3	RS3	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GSK3B is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	RS2	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	RS3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	RS3	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GSK3B is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	5	RS3	0%	0%	0%	0%	0%	0%	0%	RS3	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GSK3B is not expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	100%	0%	100%	0%	0%	0%	0%
	4	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	RS3	100%	RS4	100%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; RS2 represents regulatory state, where PC leads to upregulation (100%) and downregulation (0%). RS3 represents regulatory state 3, where PC leads to upregulation (0%), regulation between 0%-100% and downregulation (0%). RS4 represents regulatory state 4, where PC leads to regulation between 0%-100% and upregulation (100%)

Summary of node perturbation results of GSK3B interactome involved in regulation of postnatal development

PC1 (Simulations 1-4): Perturbation of each factor regulating GSK3B interactome

downregulated each stage of development (except proliferation, synaptogenesis (when GSK3 is not expressed) and synaptogenesis (when GSK3B is expressed) (0%).

Simulation 1: Perturbation of each factor resulted in overall regulation of proliferation and synaptogenesis (when GSK3B is not expressed) in two different regulatory states (RS3), regulation between 0%-100% and downregulation (0%). Synaptogenesis (when GSK3B is expressed) was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

Simulation 2: Perturbation of each factor showed downregulation (0%) of proliferation, synaptogenesis (when GSK3B is not expressed) and synaptogenesis (when GSK3B is expressed).

Simulation 3: Perturbation of each factor resulted in overall regulation of proliferation in two different regulatory states (RS2), upregulation (100%) and downregulation (0%). Synaptogenesis (when GSK3B is not expressed) was shown to be regulated in three different regulatory states (RS1), upregulation (100%), downregulation (0%) and regulation between 0%-100%.

Simulation 4: Synaptogenesis (when GSK3B is not expressed) was shown to be regulated in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of factors regulating GSK3B interactome, upregulated each same stage of development (100%). OE of genes positively regulating neurodevelopment (when GSK3B is

not expressed) along with OE of factors, upregulated each stage of development (except synaptogenesis (when GSK3B is expressed)).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of factors regulating GSK3B interactome, downregulated each same stage of development (0%).

Simulations 2 and 4: KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed) along with KO of factors, upregulated proliferation (100%). Similarly, KO of each stage (except synaptogenesis (when GSK3B is not expressed) and neurodevelopment (when GSK3B is not expressed), upregulated (100%) synaptogenesis (when GSK3B is not expressed). We also observed that KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with KO of factors, upregulated (100%) synaptogenesis (when GSK3B is expressed).

PC4 (Simulations 1-4): OE of each factor along with OE of genes positively regulating each stage of development, upregulated each same stage of development (100%).

Simulations 1 and 4: OE of genes positively regulating neurite growth/ proliferation showed overall regulation of synaptogenesis (when GSK3B is not expressed) in two different regulatory states (RS3), regulation between 0%-100% as well as downregulation (0%).

Simulation 3: OE of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is not expressed) showed overall regulation of proliferation in two different regulatory states (RS2), upregulation (100%) as well as downregulation (0%). Similarly, OE of genes positively regulating neurite growth/ proliferation showed regulation of synaptogenesis (when GSK3B is not expressed) in two different regulatory states (RS2), upregulation (100%) as well as downregulation (0%). OE of genes positively regulating neurodevelopment (when GSK3B is not expressed) showed regulation of neurodevelopment

in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% as well as downregulation (0%).

PC5 (Simulations 1-4): KO of each factor along with KO of genes positively regulating each stage of development, downregulated each same stage of development (0%).

Simulation 1: KO of genes positively regulating neurite growth/ synaptogenesis (when GSK3B is expressed)/ synaptogenesis (when GSK3B is not expressed) along with KO of each factor, showed overall regulation of proliferation in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. Similarly, KO of genes positively regulating neurite growth/ proliferation/ synaptogenesis (when GSK3B is expressed) along with KO of each factor showed overall regulation of synaptogenesis (when GSK3B is not expressed) in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

KO of genes positively regulating neurite growth/ proliferation/ synaptogenesis (when GSK3B is not expressed)/ neurodevelopment (when GSK3B is not expressed) and KO of each factor showed overall regulation of synaptogenesis (when GSK3B is expressed) in two different regulatory states, RS3, downregulation (0%) and regulation between 0%-100%.

Simulation 3: KO of genes positively regulating neurite growth/ proliferation along with KO of each factor showed overall regulation of synaptogenesis (when GSK3B is not expressed) in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. KO of genes positively regulating synaptogenesis (when GSK3B is expressed) with KO of each factor, upregulated synaptogenesis (when GSK3B is not expressed) (100%). Similarly, KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) and KO of each factor, showed overall regulation of synaptogenesis (when GSK3B is expressed) in

two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%.

Simulations 2 and 4: KO of genes positively regulating neurodevelopment (when GSK3B is not expressed) along with KO of each factor, upregulated synaptogenesis (when GSK3B is expressed).

Simulation 4: KO of genes positively regulating neurite growth/ proliferation and KO of each factor showed overall regulation of synaptogenesis (when GSK3B is not expressed) in three different regulatory states (RS3), regulation between 0%-100% and upregulation (100%).

Summary of the NP results for GSK3B interactome is given in Table 13Table 13.

C. BDNF interactome

During postnatal period, BDNF interactome regulates differentiation, neurite growth and synaptogenesis. We have specified two different Boolean rules for neurite growth and synaptogenesis. BDNF activates miR134 expression and upregulates neurite growth (rule 1) as well as BDNF represses miR134 expression and upregulates neurite growth (rule 2). Similarly, when BDNF is expressed, synaptogenesis is upregulated (rule 1) as well as when BDNF is not expressed synaptogenesis is upregulated (rule 2). So, for the overall neurodevelopment process (all stages combined into one stage), we also have given two different Boolean rules, neurodevelopment (when miR134 is expressed) and neurodevelopment (when miR134 is repressed).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating BDNF interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating BDNF interactome, downregulated (0%) each stage of neurodevelopment and the overall neurodevelopment process (all the stages regulated by BDNF combined into one stage).

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) and OE of miRNAs and TFs regulating BDNF interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (differentiation, neurite growth (when miR134 is activated), neurite growth (when miR134 is repressed), synaptogenesis (BDNF is expressed), synaptogenesis (BDNF is not expressed), neurodevelopment (when miR134 is expressed) and neurodevelopment (when miR134 is repressed)) along with OE of miRNAs and TFs regulating BDNF interactome, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of miRNAs and TFs, upregulated (100%) differentiation, neurite growth (when miR134 is activated), neurite growth (when miR134 is repressed), synaptogenesis (when BDNF is expressed), neurodevelopment (when miR134 is activated) and neurodevelopment (when miR134 is repressed). OE of genes positively regulating neurodevelopment (when miR134 is repressed) along with OE of miRNAs and TFs, upregulated (100%) neurodevelopment (when miR134 is repressed), differentiation, synaptogenesis (when BDNF is expressed) and neurite growth (when miR134 is repressed).

a. Differentiation

Simulations 1-4: OE of genes positively regulating neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed) along with OE of miRNAs and TFs, upregulated differentiation (100%).

b. Neurite growth (miR134 is not expressed)

Simulations 1-4: OE of genes positively regulating neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed)/ synaptogenesis

(when BDNF is expressed) along with OE of miRNAs and TFs, upregulated neurite growth (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) and KO of miRNAs and TFs regulating BDNF interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (differentiation, neurite growth (when miR134 is activated), neurite growth (when miR134 is repressed), synaptogenesis (when BDNF is expressed), synaptogenesis (when BDNF is not expressed), neurodevelopment (when miR134 is expressed) and neurodevelopment (when miR134 is repressed)) along with KO of miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) along with OE of each factor (miRNA/ TF) regulating BDNF interactome

a. Differentiation

Simulation 1: Genes positively regulating each stage of development (except synaptogenesis (when BDNF is not expressed) showed overall regulation of differentiation in all three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of STAT3, downregulated differentiation (0%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR-1-3p/ miR15a/

KLF4/ TCF3 upregulated differentiation (100%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA/TF (except miR-1-3p/ miR15a/ KLF4/ TCF3/ STAT3) regulated differentiation (between 0%-100%).

Simulation 2: Genes positively regulating each stage of development (except synaptogenesis (when BDNF is not expressed) showed overall regulation of differentiation in two regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively regulating differentiation/neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR16, regulated differentiation (between 0%-100%). OE of genes positively regulating differentiation/neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA/TF (except miR16), upregulated differentiation (100%).

Simulation 3: Genes positively regulating each stage of development (except synaptogenesis (when BDNF is not expressed) showed overall regulation of differentiation in three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ neurodevelopment (when miR134 is not expressed)/ neurodevelopment (when miR134 is

expressed)/ synaptogenesis (when BDNF is expressed) and OE of miR-1-3p/ miR15a upregulated differentiation (100%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ neurodevelopment (when miR134 is not expressed)/ neurodevelopment (when miR134 is expressed)/ synaptogenesis (when BDNF is expressed) and OE of miR16 or KLF4, regulated differentiation (between 0%-100%). OE of genes positively regulating differentiation/neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA/TF (except OE of miR16/ KLF4/ miR-1-3p/ miR15a), downregulated differentiation (0%).

Simulation 4: Genes positively regulating each stage of development (except synaptogenesis (when BDNF is not expressed) showed overall regulation of differentiation in two regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR-1-3p / miR15a/ KLF4/ TCF3/ STAT3 upregulated differentiation (100%). OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA/TF (except OE of miR-1-3p / miR15a/ KLF4/ TCF3/ STAT3) regulated differentiation (between 0%-100%).

b. Neurite growth (when miR134 is expressed)

Simulations 1-4: Genes positively regulating neurite growth (when miR134 is expressed)/ neurodevelopment (when miR134 is expressed) and OE of each miRNA/TF regulating BDNF interactome, upregulated neurite growth (100%).

c. Neurite growth (when miR134 is not expressed)

Simulations 1-4: Genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed) and OE of each miRNA/TF regulating BDNF interactome, upregulated neurite growth (100%).

d. Synaptogenesis (when BDNF is expressed)

Simulation 1: Synaptogenesis was shown to be regulated in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with TCF3 OE, upregulated synaptogenesis (100%). However, OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with STAT3 OE, downregulated synaptogenesis (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each TF (except TCF3/STAT3 OE), regulated synaptogenesis (between 0%-100%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA regulated synaptogenesis (between 0%-100%).

Simulation 2: Synaptogenesis was shown to be regulated in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%. OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR16 regulated synaptogenesis (between 0%-100%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA (except miR16 OE) upregulated synaptogenesis (100%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each TF upregulated synaptogenesis (100%).

Simulation 3: OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA and each TF, downregulated synaptogenesis (0%).

Simulation 4: Synaptogenesis was shown to be regulated in two different regulatory states (RS3) regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR22 (repressed by MYC) / miR22 (activated by MYC) / CTCF/ MYC (activates miR22) / MYC (represses miR22)/ REST/ SOX2/ TCF3 downregulated synaptogenesis (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each miRNA/TF (except OE of miR22 (repressed by MYC) / miR22 (activated by MYC) / CTCF/

MYC (activates miR22) / MYC (represses miR22)/ REST/ SOX2/ TCF3) regulated synaptogenesis (between 0% - 100%).

e. Synaptogenesis (when BDNF is not expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when BDNF is not expressed) along with OE of each miRNA and each TF upregulated synaptogenesis (100%).

f. Neurodevelopment (when miR134 is expressed)

Simulation 1: Neurodevelopment was shown to be regulated in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of TCF3, upregulated neurodevelopment (100%). However, OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of STAT3, downregulated neurodevelopment (0%). OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of STAT3/ TCF3, regulated neurodevelopment (between 0%-100%). OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of each miRNA, regulated neurodevelopment (between 0%-100%).

Simulations 2, 3 and 4: OE of genes positively regulating neurodevelopment (when miR134 is expressed) and OE of each miRNA/TF, downregulated neurodevelopment (0%), as HDAC1 gene (self-regulated) represses BDNF gene expression.

g. Neurodevelopment (when miR134 is not expressed)

Simulation 1: Neurodevelopment was shown to be regulated in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) and OE of TCF3, upregulated neurodevelopment (100%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is

expressed)/ neurodevelopment (when miR134 is not expressed) and OE of STAT3, downregulated neurodevelopment (0%). OE of each TF (except TCF3/STAT3) and OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed), regulated neurodevelopment (between 0%-100%). OE of each miRNA and OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (miR134 is expressed)/ neurodevelopment (when miR134 is not expressed), regulated neurodevelopment (between 0%-100%).

Simulation 2: Neurodevelopment was shown to be regulated in two different regulatory states (RS3), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) and OE of miR16, regulated neurodevelopment (between 0%-100%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) and OE of each miRNA/TF (except miR16), upregulated neurodevelopment (100%).

Simulation 3: OE of genes positively regulating neurodevelopment (when miR134 is not expressed) and OE of each miRNA and each TF, downregulated neurodevelopment (0%), as HDAC1 gene (self-regulated) represses BDNF gene expression.

Simulation 4: Neurodevelopment was shown to be regulated in two different regulatory states (RS3) regulation between 0%-100% and downregulation (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of miR22 (repressed by MYC) / miR22 (activated by MYC) / CTCF/ MYC (activates miR22) / MYC

(represses miR22)/ REST/ SOX2/ TCF3 OE downregulated neurodevelopment (0%). OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) and OE of each miRNA/TF (except miR22 (repressed by MYC) / miR22 (activated by MYC) / CTCF/ MYC (activates miR22) / MYC (represses miR22)/ REST/ SOX2/ TCF3), regulated neurodevelopment (b/w 0% - 100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by BDNF interactome) along with KO of each factor (miRNA/ TF) regulating BDNF interactome

Simulations 1-4: KO of all genes positively regulating each stage of neurodevelopment (differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ synaptogenesis (when BDNF is not expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed)) along with KO of each miRNA/TF, downregulated each same stage of neurodevelopment (0%).

synaptogenesis (when BDNF is expressed) and factors	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when BDNF is not expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	RS1	RS3	0%	RS3
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when miR134 is expressed) and factors	2	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	RS1	0%	0%	0%	RS1	RS3	0%	RS3
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when miR134 is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	RS1	RS3	0%	RS3
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4. RS1 represents regulatory state 1, where PC leads to upregulation (100%), regulation between 0%-100% and downregulation (0%). RS3 represents regulatory state regulatory state 3, where PC condition leads to downregulation (0%) and regulation between 0%-100%.

Summary of node perturbation results of BDNF interactome regulating postnatal development

PC1 (Simulations 1-4): Perturbation of each factor, downregulated each stage of neurodevelopment (0%).

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of miRNAs and TFs, upregulated each same stage of development (100%).

Simulations 1-4: OE of genes positively regulating neurodevelopment (when miR134 is not expressed along with OE of factors, upregulated (100%) differentiation, synaptogenesis (when BDNF is expressed), neurodevelopment (miR134 is not expressed) and neurite growth (when miR134 is not expressed). Similarly, OE of genes positively regulating neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed) along with OE of factors upregulated differentiation (100%). OE of genes positively regulating neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed) along with OE of factors, upregulated (100%) neurite growth (when miR134 is not expressed).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of miRNAs and TFs, downregulated each same stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each miRNA/TF, upregulated each same stage of development (100%).

Simulations 1 and 3: OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor, showed

overall regulation of differentiation in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%).

Simulations 2 and 4: OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor, regulated differentiation in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%.

Simulations 1-4: OE of genes positively regulating differentiation/ neurite growth (when miR134 is expressed)/ neurite growth (when miR134 is not expressed)/ synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor, upregulated neurite growth (when miR134 is not expressed) (100%).

Simulation 1: OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor showed regulation of synaptogenesis and neurodevelopment (when miR134 is not expressed) in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating neurodevelopment (when miR134 is expressed) along with OE of each factor showed regulation of neurodevelopment in three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%).

Simulation 2: OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor showed regulation of synaptogenesis as well as

neurodevelopment (when miR134 is not expressed) in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%.

Simulation 3: OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor, downregulated synaptogenesis and neurodevelopment (when miR134 is not expressed) (0%).

Simulation 4: OE of genes positively regulating synaptogenesis (when BDNF is expressed)/ neurodevelopment (when miR134 is expressed)/ neurodevelopment (when miR134 is not expressed) along with OE of each factor showed regulation of synaptogenesis as well as neurodevelopment (when miR134 is not expressed) in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor, downregulated each same stage of development (0%).

Summary of the NP results for BDNF interactome is given in Table 14 and Table 15.

D. GRM5 interactome

During postnatal development, GRM5 interactome regulates proliferation, neurite growth and synaptogenesis. For performing node perturbation analysis, we have given two different Boolean rules for neurite growth stage, where HOMER1 is expressed along with GRM5 and regulates neurite growth. In the other rule, HOMER1 is not expressed and GRM5 is expressed. Similarly, GRM5 expression upregulates as well as downregulates synaptogenesis, so we have given two different Boolean rules for the stage, synaptogenesis. So, for the overall neurodevelopment process (combining all stages into one stage), we have given three different Boolean rules, where neurodevelopment is upregulated when GRM5 is

expressed (rule 1), neurodevelopment is upregulated when GRM5 and HOMER1 is expressed (rule 2) and neurodevelopment is upregulated when GRM5 is repressed (rule 3).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating GRM5 interactome

Simulations 1-4: OE and KO of each miRNA and each TF downregulated proliferation, neurite growth (when GRM5 is expressed), neurite growth (when GRM5 and HOMER1 is expressed), synaptogenesis (when GRM5 is expressed), synaptogenesis (when GRM5 is not expressed), neurodevelopment (when GRM5 is repressed), neurodevelopment (when GRM5 and HOMER1 is expressed) and neurodevelopment (when GRM5 is expressed) (0%).

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) and OE of miRNAs and TFs regulating GRM5 interactome

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation, neurite growth (when GRM5 is expressed), neurite growth (when GRM5 and HOMER is expressed), synaptogenesis (when GRM5 is expressed), synaptogenesis (when GRM5 is not expressed), neurodevelopment (when GRM5 and HOMER1 is expressed), neurodevelopment (when GRM5 is repressed) and neurodevelopment (when GRM5 is expressed)) along with OE of miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). Similarly, OE of genes positively regulating neurodevelopment (when GRM5 is expressed) along with OE of miRNAs and TFs, upregulated proliferation, neurite growth (when GRM5 is expressed), synaptogenesis (when GRM5 is expressed) and neurodevelopment (when GRM5 is expressed) (100%).

Perturbation condition 3: KO of genes regulating each stage of neurodevelopment (regulated by GRM5 interactome) and KO of miRNAs and TFs regulating GRM5 interactome)

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation, neurite growth (when GRM5 is expressed), neurite growth (when GRM5 and HOMER is expressed), synaptogenesis (when GRM5 is expressed), synaptogenesis (when GRM5 is not expressed), neurodevelopment (when GRM5 and HOMER1 is expressed), neurodevelopment (when GRM5 is repressed) along with KO of miRNAs and TFs regulating GRM5 interactome, downregulated each same stage of neurodevelopment (0%).

Perturbation condition 4: OE of genes regulating each stage of neurodevelopment (regulated by GRM5 interactome) along with OE of each factor (miRNA/ TF) regulating GRM5 interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ neurodevelopment (when GRM5 is expressed) and OE of each miRNA/ TF, upregulated proliferation (100%).

Simulations 1 and 3: OE of genes positively regulating neurite growth (when GRM5 is expressed)/ synaptogenesis (when GRM5 is expressed) and OE of REST/ RUNX2/ NEUROD2 regulated proliferation (between 0%-100%). OE of genes positively regulating neurite growth (when GRM5 is expressed)/ synaptogenesis (when GRM5 is expressed) and OE of each factor (except REST/ RUNX2/ NEUROD2), downregulated proliferation (0%). Thus, proliferation is shown to be regulated by other stages of development in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

b. Neurite growth (when GRM5 and HOMER1 is expressed)

Simulations 1-4: OE of genes positively regulating neurite growth (when GRM5 and HOMER1 is expressed)/ neurodevelopment (when GRM5 and HOMER1 is expressed) along with OE of each miRNA/ TF, upregulated neurite growth (100%).

c. Neurite growth (when GRM5 is expressed)

Simulations 1-4: OE of genes positively regulating neurite growth (when GRM5 is expressed)/ neurodevelopment (when GRM5 is expressed along with OE of each miRNA/ TF, upregulated neurite growth (100%).

d. Synaptogenesis (when GRM5 is expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GRM5 is expressed)/ synaptogenesis (when GRM5 is expressed) along with OE of each miRNA/ TF, upregulated synaptogenesis (100%).

e. Synaptogenesis (when GRM5 is not expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRM5 is repressed) along with OE of each miRNA/ TF, upregulated synaptogenesis (100%).

f. Neurodevelopment (when GRM5 and HOMER1 is expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GRM5 and HOMER1 is expressed) along with OE of each miRNA/ TF, upregulated neurodevelopment (100%).

g. Neurodevelopment (when GRM5 is expressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GRM5 is expressed) along with OE of each miRNA/ TF, upregulated neurodevelopment (100%).

h. Neurodevelopment (when GRM5 is repressed)

Simulations 1-4: OE of genes positively regulating neurodevelopment (when GRM5 is repressed) along with OE of each miRNA/ TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of all genes positively regulating each stage of neurodevelopment (regulated by GRM5 interactome) along with KO of each factor (miRNA/ TF) regulating GRM5 interactome

Simulations 1-4: KO of all genes positively regulating each stage of neurodevelopment (proliferation, neurite growth (when GRM5 is expressed), neurite growth (when GRM5 and HOMER is expressed), synaptogenesis (when GRM5 is expressed), synaptogenesis (when

GRM5 is not expressed), neurodevelopment (when GRM5 and HOMER1 is expressed), neurodevelopment (when GRM5 is repressed) along with KO of each factor (miRNA/ TF), downregulated each same stage of neurodevelopment (0%).

factors	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurite growth (when GRM5 is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GRM5 is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GRM5 is not expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GRM5 and HOMER1 is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GRM5 is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment (when GRM5 is repressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; Upregulation (100%), downregulation (0%)

Summary of node perturbation results of GRM5 interactome involved in postnatal development

PC1 (Simulations 1-4): Perturbation of each factor downregulated each of development.

Perturbation of each factor showed upregulation (100%) of neurite growth (when GRM5 is not expressed).

PC2 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of all factors regulating GRM5 interactome, upregulated each same stage of development (100%). OE of genes positively regulating neurodevelopment (when GRM5 is expressed) along with OE of all factors, upregulated (100%) each stage of development (synaptogenesis (when GRM5 is not expressed) and neurodevelopment (when GRM5 is not expressed)).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of all factors regulating GRM5 interactome, downregulated each same stage of development (0%).

PC4 (Simulations 1-4): OE of genes positively regulating each stage of development along with OE of each factor, showed upregulation of each same stage of development (100%). OE of genes positively regulating neurodevelopment (when GRM5 is expressed) along with OE of each factor, upregulated (100%) each stage of development (except synaptogenesis (when GRM5 is not expressed) and neurodevelopment (when GRM5 is not expressed)). OE of genes positively regulating neurodevelopment (when GRM5 and HOMER1 is expressed) along with OE of each factor, showed upregulation of neurite growth (when GRM5 and HOMER is expressed) (100%).

Simulations 1 and 3: OE of genes positively regulating neurite growth (when GRM5 is expressed), synaptogenesis (when GRM5 is expressed) along with OE of each factor, showed

overall proliferation process to be regulated in two regulatory states (RS3), downregulation (0%) and regulation between 0%-100%.

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of all factors regulating GRM5 interactome, downregulated each same stage of development (0%).

Summary the NP results for GRM5 interactome is given in Table 16 and Table 17.

E. NRG1 interactome

NRG1 regulates differentiation, myelination and synaptogenesis. NRG1 gene has been shown to be regulated only by miR124, in turn miR124 is regulated by TFs, STAT3, REST and EGR1. The direction of regulation of miRNA by these four TFs has been experimentally shown. Therefore, Boolean rules for simulation 1 and 3 remains same, as TF upregulates gene expression in both simulations and the TF regulation of miRNA expression also does not change in both the simulations. Similarly, Boolean rules for simulations 2 and 4 are same, as TF downregulates gene expression in both the simulations and the TF regulation of miRNA expression does not change in both the simulations. We have given two different Boolean rules for synaptogenesis, where DISC1 expression upregulates synaptogenesis (rule 1) and NRG1 expression upregulates synaptogenesis (rule 2).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating NRG1 interactome

Simulations 1-4: OE of each miRNA and each TF, downregulated (0%) each stage of development (proliferation, differentiation, myelination, synaptogenesis (DISC1 is expressed), synaptogenesis (when NRG1 is expressed), neurodevelopment). KO of each miRNA downregulated proliferation, differentiation, myelination and synaptogenesis (when NRG1 is expressed) (0%). KO of each TF downregulated proliferation, differentiation,

myelination, synaptogenesis (when DISC1 is expressed) and synaptogenesis (when NRG1 is expressed) (0%).

a. Synaptogenesis (when DISC1 is expressed)

Simulations 1 and 3: We observed regulation of synaptogenesis in two different regulatory states (RS2), downregulation (0%) and upregulation (100%). KO of miR124 upregulated (100%) synaptogenesis (when DISC1 is expressed). KO of each factor (except miR124), downregulated (0%) synaptogenesis (when DISC1 is expressed).

Perturbation condition 2: OE of genes regulating each stage of neurodevelopment (regulated by NRG1 interactome) and OE of miRNAs and TFs regulating NRG1 interactome)

Simulations 1-4: OE of genes positively regulating each stage of neurodevelopment (proliferation, differentiation, myelination, synaptogenesis (when DISC1 is expressed), synaptogenesis (when NRG1 is expressed) and neurodevelopment) and OE of all miRNAs and TFs, upregulated each same stage of neurodevelopment (100%). OE of genes positively regulating neurodevelopment along with OE of miRNAs and TFs, upregulated proliferation, differentiation, myelination, synaptogenesis (when NRG1 is expressed), neurodevelopment (100%).

a. Proliferation

Simulations 1-4: OE of genes positively regulating differentiation/myelination/synaptogenesis (when NRG1 is expressed) along with OE of miRNAs and TFs, upregulated proliferation (100%).

b. Differentiation

Simulations 1-4: OE of genes positively regulating proliferation/myelination/ synaptogenesis (when NRG1 is expressed) along with OE of miRNAs and TFs, upregulated differentiation (100%).

c. Myelination

Simulations 1-4: OE of genes positively regulating proliferation/ differentiation/ synaptogenesis (when NRG1 is expressed) along with OE of miRNAs and TFs upregulated myelination (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) and KO of miRNAs and TFs regulating NRG1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation, differentiation, myelination, synaptogenesis (when DISC1 is expressed), synaptogenesis (when NRG1 is expressed), neurodevelopment (when DISC1 is expressed), neurodevelopment) and KO of all miRNAs and TFs, downregulated each same stage of neurodevelopment (0%).

a. Synaptogenesis (DISC1 is expressed)

Simulations 2 and 4: KO of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and KO of all miRNAs and TFs, upregulated synaptogenesis (when DISC1 is expressed) (100%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) along with OE of each factor (miRNA/ TF) regulating NRG1 interactome

a. Proliferation

Simulations 1-4: OE of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/ TF, upregulated proliferation (100%).

b. Differentiation

Simulations 1-4: OE of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/ TF, upregulated differentiation (100%).

c. Myelination

Simulations 1 and 3: OE of genes positively regulating proliferation/ differentiation along with OE of each miRNA/TF (except OE of EGR1) showed upregulation of myelination (100%). OE of genes positively regulating differentiation along with OE of EGR1 showed regulation of myelination between 0%-100%. OE of genes positively regulating myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/ TF, upregulated myelination (100%).

Simulations 2 and 4: OE of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/ TF, upregulated myelination (100%).

c. Synaptogenesis (when NRG1 is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/TF, upregulated synaptogenesis (100%).

d. Synaptogenesis (when DISC1 is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when DISC1 is expressed) and OE of each miRNA/TF, upregulated synaptogenesis (100%).

Simulations 1 and 3: Synaptogenesis was shown to be regulated in all three regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). OE of genes positively regulating proliferation/ differentiation/ myelination and OE of miR124, downregulated synaptogenesis (0%). OE of genes positively regulating proliferation/ differentiation/ myelination and OE of EGR1 regulated synaptogenesis (between 0%-100%). OE of genes positively regulating proliferation/

differentiation/myelination and OE of each miRNA/TF (except OE of miR124 or EGR1) upregulated synaptogenesis (100%).

e. Neurodevelopment

Simulations 1-4: OE of genes positively regulating synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each miRNA/TF, upregulated neurodevelopment (100%).

Perturbation condition 5: KO of all genes positively regulating each stage of neurodevelopment (regulated by NRG1 interactome) along with KO of each factor (miRNA/TF) regulating NRG1 interactome

Simulations 1-4: KO of genes positively regulating each stage of neurodevelopment (proliferation, differentiation, myelination, synaptogenesis (when NRG1 is expressed), synaptogenesis (when DISC1 is expressed) and neurodevelopment) along with KO of each miRNA/ TF regulating NRG1 interactome, downregulated each same stage of neurodevelopment (0%).

a. Synaptogenesis (when DISC1 is expressed)

Simulations 1 and 3: KO of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment and each miRNA/TF KO upregulated synaptogenesis (100%).

Table 18: Perturbation results of NRG1 interactome involved in postnatal development

[illegible]

neurodevelopment and factors	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4					100%	100%	100%	100%	100%	100%	100%	100%
	5					0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Simulation 3; Sim4: Simulation 4; RS1 represents regulatory state 1, where PC leads to upregulation (100%), regulation between 0%-100% and downregulation (0%); RS2 represents regulatory state 2, where PC leads to upregulation (100%) and downregulation (0%); RS4 represents regulatory state 4, where PC leads to upregulation (100%) and regulation between 0%-100%

Table 19: Perturbation results of NRG1 interactome involved in postnatal development (contd)

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t = 150$)							
		Synaptogenesis (when DISC1 is expressed)				Neurodevelopment			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	RS2	0%	RS2	0%	0%	0%	0%	0%
Genes positively regulating proliferation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%
	4	RS1	0%	RS1	0%	0%	0%	0%	0%
	5	100%	0%	100%	0%	0%	0%	0%	0%
Genes positively regulating differentiation and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%
	4	RS1	0%	RS1	0%	0%	0%	0%	0%
	5	100%	0%	100%	0%	0%	0%	0%	0%
Genes positively regulating myelination and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%
	4	RS1	0%	RS1	0%	0%	0%	0%	0%
	5	100%	0%	100%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when NRG1 is expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	100%	0%	100%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%

	5	100%	0%	100%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when DISC1 is expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating neurodevelopment and factors	2	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	100%	0%	100%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%
	5	100%	0%	100%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Simulation 3; Sim4: Simulation 4; RS1 represents regulatory state 1, where PC leads to upregulation (100%), regulation between 0%-100% and downregulation (0%); RS2 represents regulatory state 2, where PC leads to upregulation (100%) and downregulation (0%); RS4 represents regulatory state 4, where PC leads to upregulation (100%) and regulation between 0%-100%

Summary of node perturbation results regulating NRG1 interactome involved in postnatal development

PC1 (Simulations 1-4): Perturbation of each factor, downregulated each stage (except synaptogenesis (when NRG1 is expressed) of development (0%).

Simulations 1 and 3: Perturbation of each factor showed overall regulation of synaptogenesis in two different regulatory states (RS2), downregulation (0%) and regulation between 0%-100%.

PC2 (Simulations 1-4): OE of genes positively each stage along with OE of factors upregulated each same stage of development (100%). OE of genes positively regulating neurodevelopment, showed upregulation (100%) of each stage (except synaptogenesis (when DISC1 is expressed)). OE of genes positively regulating differentiation/ myelination/ synaptogenesis (when NRG1 is expressed) along with OE of factors, upregulated proliferation (100%). OE of genes positively regulating proliferation/ myelination/ synaptogenesis (when NRG1 is expressed) along with OE of factors, upregulated differentiation (100%). Similarly, OE of genes positively regulating proliferation/ differentiation/ synaptogenesis (when NRG1 is expressed) along with OE of factors, showed upregulation of myelination (100%).

PC3 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of all factors regulating NRG1 interactome, downregulated each same stage of development (0%).

Simulations 2 and 4: KO of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment along with KO of all factors, upregulated (100%) synaptogenesis (when DISC1 is expressed).

PC4 (Simulations 1-4): OE of genes positively each stage along with OE of each factor upregulated each same stage of development (100%). OE of genes positively regulating

neurodevelopment, showed upregulation (100%) of each stage (except synaptogenesis (when DISC1 is expressed)). OE of genes positively regulating differentiation/ myelination/ synaptogenesis (when NRG1 is expressed) along with OE of each factor, upregulated differentiation (100%). OE of genes positively regulating proliferation/ myelination/ synaptogenesis (when NRG1 is expressed) along with OE of each factor, upregulated differentiation (100%). Similarly, OE of genes positively regulating proliferation/ differentiation/ synaptogenesis (when NRG1 is expressed) along with OE of each factor, showed upregulation of myelination (100%).

OE of genes positively regulating synaptogenesis (when NRG1 is expressed)/ neurodevelopment and OE of each factor, showed upregulation of neurodevelopment (100%).

Simulations 1 and 3: OE of genes positively regulating proliferation/ myelination/ differentiation along with OE of each factor, showed overall regulation of synaptogenesis (when DISC1 is expressed) in all three different regulatory states (RS1), upregulation (100%), regulation between 0%-100% and downregulation (0%). Similarly, OE of genes positively regulating proliferation/ differentiation along with OE of each factor, showed overall regulation of myelination in two different regulatory states (RS4), upregulation (100%) and regulation between 0%-100%.

PC5 (Simulations 1-4): KO of genes positively regulating each stage of development along with KO of each factor regulating NRG1 interactome, downregulated each same stage of development (0%).

Simulations 1 and 3: KO of genes positively regulating proliferation/ differentiation/ myelination/ synaptogenesis (when NRG1 is expressed)/ neurodevelopment (when NRG1 is

expressed) along with KO of each factor, upregulated (100%) synaptogenesis (when DISC1 is expressed).

Summary of the NP results for NRG1 interactome is given in Table 18Table 18 and Table 19Table 19.

F. YWHAE interactome

During prenatal development, YWHAE interactome regulates neurite growth

Single gene perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating YWHAE interactome

Simulations 1-4: OE and KO of each miRNA and each TF regulating YWHAE interactome, downregulated neurite growth (0%).

Multiple gene perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) and OE of miRNAs and TFs regulating YWHAE interactome

Simulations 1-4: OE of all genes positively regulating neurite growth and OE of all miRNAs and TFs upregulated neurite growth (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) and OE of miRNAs and TFs regulating YWHAE interactome

Simulations 1-4: KO of all genes positively regulating neurite growth and KO of all miRNAs and TFs downregulated neurite growth (0%).

Perturbation condition 4: OE of all genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) along with OE of each factor (miRNA/TF) regulating YWHAE interactome

Simulations 1-4: OE of all genes positively regulating neurite growth and OE of each miRNA/TF upregulated neurite growth (100%).

Perturbation condition 5: KO of all genes positively regulating each stage of neurodevelopment (regulated by YWHAE interactome) along with KO of each factor (miRNA/ TF) regulating YWHAE interactome

Simulations 1-4: KO of genes positively regulating neurite growth and KO of each miRNA/ TF downregulated neurite growth (0%).

Table 20: Perturbation results of YWHAE interactome involved in postnatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t = 150$ (Activation frequency for 1000 simulations at $t = 150$)			
		Neurite growth			
		Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%
Genes positively regulating neurite growth and factors	2	100%	100%	100%	100%
	3	0%	0%	0%	0%
	4	100%	100%	100%	100%
	5	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; Upregulation (100%), Downregulation (0)

Summary of node perturbation results of YWHAE interactome involved in postnatal development

YWHAE interactome regulated neurite growth during postnatal development. At all four simulations, perturbation of each factor (PC1) regulating YWHAE interactome showed downregulation of neurite growth (0%). Similarly, KO of genes positively regulating neurite growth along with KO of all factors (PC3)/ each factor (PC5), also showed downregulation of neurite growth (0%). But, OE of genes positively regulating neurite growth along with OE of all factors (PC2)/ each factor (PC4), showed upregulation of neurite growth (100%).

Summary of the NP results for YWHAE interactome is given in Table 20.

G. DLG2 interactome

During postnatal development, DLG2 interactome is shown to regulate synaptogenesis. DLG2 interactome has been shown to regulated by miR873 and EP300 (TF-miRNA FFLs).

Single node perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating DLG2 interactome

Simulations 1-4: OE and KO of miR873 and EP300 downregulated synaptogenesis (0%)

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by DLG2 interactome) and OE of miRNAs and TFs regulating DLG2 interactome

Simulations 1-4: OE of genes positively regulating synaptogenesis and OE of miR873 and EP300 regulating DLG2 interactome, upregulated synaptogenesis (100%).

Perturbation condition 3: KO of genes positively regulating each stage of neurodevelopment (regulated by DLG2 interactome) and KO of miRNAs, TFs regulating DLG2 interactome

Simulations 1-4: KO of genes positively regulating synaptogenesis and KO of miR873 and EP300 regulating DLG2 interactome, downregulated synaptogenesis (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by DLG2 interactome) along with OE of each factor (miRNA/ TF) regulating DLG2 interactome

Simulations 1-4: OE of genes positively regulating synaptogenesis and OE of EP300/ miR873, upregulated synaptogenesis (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by DLG2 interactome) along with KO of each factor (miRNA/TF) regulating DLG2 interactome

Simulations 1-4: KO of genes positively regulating synaptogenesis and KO of EP300/ miR873, downregulated synaptogenesis (0%).

Table 21: Perturbation results of DLG2 interactome involved in postnatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t = 150$ (Activation frequency for 1000 simulations at $t = 150$)

		Synaptogenesis			
		Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%
Genes positively regulating synaptogenesis and factors	2	100%	100%	100%	100%
	3	0%	0%	0%	0%
	4	100%	100%	100%	100%
	5	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1; Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4; Upregulation (100%), Downregulation (0%)

Summary of node perturbation results of DLG2 interactome involved in postnatal development

DLG2 interactome regulated synaptogenesis during postnatal development. At all four simulations, perturbation of each factor (PC1) regulating DLG2 interactome showed downregulation of synaptogenesis (0%). Similarly, KO of genes positively regulating synaptogenesis along with KO of all factors (PC3)/ each factor (PC5), also showed downregulation of synaptogenesis (0%). But, OE of genes positively regulating synaptogenesis along with OE of all factors (PC2)/ each factor (PC4), showed upregulation of synaptogenesis (100%). Summary of the NP results for DLG2 interactome is given in Table 21.

h. GRIN2A interactome

During postnatal development, GRIN2A interactome regulates synaptogenesis. We have given two different Boolean rules for synaptogenesis, where GRIN2B expression upregulates synaptogenesis (rule 1) and GRIN2A expression downregulates synaptogenesis (rule 2).

Single gene perturbation

Perturbation condition 1: OE and KO of each miRNA and each TF regulating GRIN2A interactome

Simulations 1-4: OE and KO of each miRNA and each TF downregulated (0%) synaptogenesis (when GRIN2A is expressed) and synaptogenesis (when GRIN2B is expressed). OE and KO of

each miRNA and each TF (except KO of miR411/HNF4A) downregulated synaptogenesis (when GRIN2A is not expressed) (0%).

a. Synaptogenesis (when GRIN2A is not expressed)

Simulations 1 and 3: We observed that the overall synaptogenesis process to be regulated in two different regulatory states (RS3), regulation between 0%-100% and downregulation (0%). KO of miR411 regulated synaptogenesis (between 0%-100%).

Simulation 2 and 4: We observed that the overall synaptogenesis process is regulated in two different regulatory states (RS3), regulation between 0%-100% and downregulation (0%). KO of HNF4A regulates synaptogenesis (between 0%-100%)

Multiple node perturbation

Perturbation condition 2: OE of genes positively regulating each stage of neurodevelopment (regulated by GRIN2A interactome) and OE of miRNAs and TFs regulating GRIN2A interactome

a. Synaptogenesis (when GRIN2B is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2B is expressed) and OE of miRNAs and TFs regulating GRIN2A interactome, upregulated synaptogenesis (100%).

b. Synaptogenesis (when GRIN2A is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2A is expressed) and OE of miRNAs and TFs regulating GRIN2A interactome, upregulated synaptogenesis (100%).

c. Synaptogenesis (when GRIN2A is not expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2A is not expressed) and OE of miRNAs and TFs regulating GRIN2A interactome, upregulated synaptogenesis (100%).

Perturbation condition 3: KO of genes regulating each stage of neurodevelopment (regulated by GRIN2A interactome) and KO of miRNAs and TFs regulating GRIN2A interactome

a. Synaptogenesis (when GRIN2B is expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (GRIN2B is expressed) and KO of miRNAs and TFs, downregulated synaptogenesis (0%).

b. Synaptogenesis (when GRIN2A is expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (when GRIN2A is expressed) and KO of miRNAs and TFs, downregulated synaptogenesis (0%).

c. Synaptogenesis (when GRIN2A is not expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (when GRIN2A is not expressed) and KO of miRNAs and TFs, downregulated synaptogenesis (0%).

Perturbation condition 4: OE of genes positively regulating each stage of neurodevelopment (regulated by GRIN2A interactome) along with OE of each factor (miRNA/TF) regulating GRIN2A interactome

a. Synaptogenesis (when GRIN2B is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2B is expressed) and OE of each miRNA and each TF, upregulated synaptogenesis (100%).

b. Synaptogenesis (when GRIN2A is expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2A is expressed) and OE of each miRNA and each TF, upregulated synaptogenesis (100%).

c. Synaptogenesis (GRIN2A is not expressed)

Simulations 1-4: OE of genes positively regulating synaptogenesis (when GRIN2A is not expressed) and OE of each miRNA and each TF, upregulated synaptogenesis (100%).

Perturbation condition 5: KO of genes positively regulating each stage of neurodevelopment (regulated by GRIN2A interactome) along with KO of each factor (miRNA/ TF) regulating GRIN2A interactome

a. Synaptogenesis (when GRIN2B is expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (when GRIN2B is expressed)

and KO of each miRNA and TF, downregulated synaptogenesis (0%).

b. Synaptogenesis (GRIN2A is expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (when GRIN2A is expressed)

and KO of each miRNA and each TF, downregulated synaptogenesis (0%).

c. Synaptogenesis (GRIN2A is not expressed)

Simulations 1-4: KO of genes positively regulating synaptogenesis (when GRIN2A is not expressed) and KO of each miRNA and each TF, downregulated synaptogenesis (0%).

Table 22: Perturbation results of GRIN2A interactome involved in postnatal development

Perturbed nodes	PC	Effect of perturbation on each stage at $t=150$ (Activation frequency for 1000 simulations at $t=150$)											
		Synaptogenesis (when GRIN2B is not expressed)				Synaptogenesis (when GRIN2A is expressed)				Synaptogenesis (when GRIN2A is not expressed)			
		Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4	Sim1	Sim2	Sim3	Sim4
Factors	1	0%	0%	0%	0%	0%	0%	0%	0%	RS3	RS3	RS3	RS3
Genes positively regulating synaptogenesis (when GRIN2B is expressed) and factors	2	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	100%	100%	100%	100%	0%	0%	0%	0%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GRIN2A is expressed) and factors	2	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	100%	100%	100%	100%	0%	0%	0%	0%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Genes positively regulating synaptogenesis (when GRIN2A is not expressed) and factors	2	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	3	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	0%	0%	0%	0%	0%	0%	0%	0%	100%	100%	100%	100%
	5	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%

PC: Perturbation condition; Sim1: Simulation 1, Sim2: Simulation 2; Sim3: Simulation 3; Sim4: Simulation 4

Summary of node perturbation results of GRIN2A interactome regulating postnatal development

GRIN2A interactome regulated synaptogenesis during postnatal development. From the curated experimental evidence, we observed that synaptogenesis is regulated in three different conditions, when GRIN2A and GRIN2B is expressed, when GRIN2A is not expressed and when GRIN2B is not expressed.

PC1 (Simulations 1-4): Perturbation of each factor, downregulated (0%) synaptogenesis (when GRIN2B is not expressed) and synaptogenesis (when GRIN2A is expressed).

We observed overall regulation of synaptogenesis (when GRIN2A is not expressed) in two different regulatory states (RS3), downregulation (0%) and regulation between 0%-100%. GRM3 (one of the positively regulating gene involved in synaptogenesis) has been shown to regulated by miR411-HNF4A composite FFLS. When either one of the factors is KO (miR411 in simulations 1 and 3 and HNF4A in simulations 2 and 4) synaptogenesis was shown to be regulated between 0%-100%.

Regulation of synaptogenesis (at PC2, PC3, PC4 and PC5)

OE of genes positively regulating synaptogenesis (when GRIN2A is expressed)/ synaptogenesis (when GRIN2A is not expressed)/ synaptogenesis (when GRIN2B is not expressed) along with OE of all factors (PC2) or OE of each factor (PC4), showed upregulation (100%) of synaptogenesis (when GRIN2A is expressed)/ synaptogenesis (when GRIN2A is not expressed)/ synaptogenesis (when GRIN2B is not expressed)

KO of genes positively regulating synaptogenesis (when GRIN2A is expressed)/ synaptogenesis (when GRIN2A is not expressed)/ synaptogenesis (when GRIN2B is not expressed) along with KO of all factors (PC3) or KO of each factor (PC5), showed downregulation (0%) of

synaptogenesis (when GRIN2A is expressed)/ synaptogenesis (when GRIN2A is not expressed)/
synaptogenesis (when GRIN2B is not expressed).

Summary of the NP results for GRIN2A interactome is given in Table 22Table 22.

2. References (for Table 1)

1. Ota, M. *et al.* A polymorphism of the ABCA1 gene confers susceptibility to schizophrenia and related brain changes. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 1877–1883 (2011).
2. Zhang, J. *et al.* Upregulation of adenosine A2A receptors induced by atypical antipsychotics and its correlation with sensory gating in schizophrenia patients. *Psychiatry Res* **200**, 126–132 (2012).
3. Jagannathan, K. *et al.* Genetic associations of brain structural networks in schizophrenia: a preliminary study. *Biol. Psychiatry* **68**, 657–666 (2010).
4. Mathur, A., Law, M. H., Megson, I. L., Shaw, D. J. & Wei, J. Genetic association of the AKT1 gene with schizophrenia in a British population. *Psychiatr. Genet.* **20**, 118–122 (2010).
5. Schwab, S. G. *et al.* Further evidence for association of variants in the AKT1 gene with schizophrenia in a sample of European sib-pair families. *Biol. Psychiatry* **58**, 446–450 (2005).
6. Ohi, K. *et al.* The AKT1 gene is associated with attention and brain morphology in schizophrenia. *World J. Biol. Psychiatry* **14**, 100–113 (2013).
7. Yasuda, Y. *et al.* AKT1 GENE IS ASSOCIATED WITH ATTENTION AND BRAIN MORPHOLOGY IN PATIENTS WITH SCHIZOPHRENIA. *Schizophrenia Research* **117**, 456–457 (2010).

8. Tan, H.-Y. *et al.* Genetic variation in AKT1 is linked to dopamine-associated prefrontal cortical structure and function in humans. *J. Clin. Invest.* **118**, 2200–2208 (2008).
9. Yuan, A. *et al.* ANK3 as a risk gene for schizophrenia: new data in Han Chinese and meta analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 997–1005 (2012).
10. Guo, X. *et al.* Association analysis of ANK3 gene variants with schizophrenia in a northern Chinese Han population. *Oncotarget* **7**, 85888–85894 (2016).
11. Nie, F. *et al.* Genetic analysis of SNPs in CACNA1C and ANK3 gene with schizophrenia: A comprehensive meta-analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **168**, 637–648 (2015).
12. Cassidy, C. *et al.* Association of a risk allele of ANK3 with cognitive performance and cortical thickness in patients with first-episode psychosis. *J Psychiatry Neurosci* **39**, 31–39 (2014).
13. Liu, W. *et al.* Association of APOE gene with schizophrenia in Chinese: a possible risk factor in times of malnutrition. *Schizophr. Res.* **62**, 225–230 (2003).
14. Al-Asmary, S. M., Kadasah, S., Arfin, M., Tariq, M. & Al-Asmari, A. Apolipoprotein E polymorphism is associated with susceptibility to schizophrenia among Saudis. *Arch Med Sci* **11**, 869–876 (2015).
15. Kecmanović, M. *et al.* Schizophrenia and apolipoprotein E gene polymorphism in Serbian population. *Int. J. Neurosci.* **120**, 502–506 (2010).
16. Akanji, A. O., Ohaeri, J. U., Al-Shammri, S. N. & Fatania, H. R. Apolipoprotein E polymorphism and clinical disease phenotypes in Arab patients with schizophrenia. *Neuropsychobiology* **60**, 67–72 (2009).

17. Kampman, O. *et al.* Apolipoprotein E polymorphism is associated with age of onset in schizophrenia. *J. Hum. Genet.* **49**, 355–359 (2004).
18. Dean, B. *et al.* Increased levels of apolipoprotein E in the frontal cortex of subjects with schizophrenia. *Biol. Psychiatry* **54**, 616–622 (2003).
19. Joob, R. *et al.* Apolipoprotein E genotype in schizophrenia. *Am. J. Med. Genet.* **67**, 235 (1996).
20. Nao, J. *et al.* Adverse Effects of the Apolipoprotein E ϵ 4 Allele on Episodic Memory, Task Switching and Gray Matter Volume in Healthy Young Adults. *Front Hum Neurosci* **11**, 346 (2017).
21. Knickmeyer, R. C. *et al.* Common variants in psychiatric risk genes predict brain structure at birth. *Cereb. Cortex* **24**, 1230–1246 (2014).
22. Porter, T. *et al.* KIBRA is associated with accelerated cognitive decline and hippocampal atrophy in APOE ϵ 4-positive cognitively normal adults with high A β -amyloid burden. *Sci Rep* **8**, 2034 (2018).
23. Mas, S. *et al.* A functional variant provided further evidence for the association of ARVCF with schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 1052–1059 (2010).
24. Mas, S. *et al.* ARVCF single marker and haplotypic association with schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **33**, 1064–1069 (2009).
25. Sanders, A. R. *et al.* Haplotypic association spanning the 22q11.21 genes COMT and ARVCF with schizophrenia. *Mol. Psychiatry* **10**, 353–365 (2005).
26. Boardman, J. P. *et al.* Common genetic variants and risk of brain injury after preterm birth. *Pediatrics* **133**, e1655-1663 (2014).

27. Kähler, A. K. *et al.* Candidate gene analysis of the human natural killer-1 carbohydrate pathway and perineuronal nets in schizophrenia: B3GAT2 is associated with disease risk and cortical surface area. *Biol. Psychiatry* **69**, 90–96 (2011).
28. Skibinska, M. *et al.* Val66Met functional polymorphism and serum protein level of brain-derived neurotrophic factor (BDNF) in acute episode of schizophrenia and depression. *Pharmacol Rep* **70**, 55–59 (2018).
29. Xia, H. *et al.* Suicide attempt, clinical correlates, and BDNF Val66Met polymorphism in chronic patients with schizophrenia. *Neuropsychology* **32**, 199–205 (2018).
30. Zhang, C. *et al.* [Association of Val66Met polymorphism of brain-derived neurotrophic factor gene with cognitive impairment and clinical symptoms in first episode schizophrenia]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **34**, 592–596 (2017).
31. Dong, Z. *et al.* Association of DISC1, BDNF, and COMT polymorphisms with exploratory eye movement of schizophrenia in a Chinese Han population. *Psychiatr. Genet.* **26**, 258–265 (2016).
32. Mezquida, G. *et al.* Association of the brain-derived neurotrophic factor Val66Met polymorphism with negative symptoms severity, but not cognitive function, in first-episode schizophrenia spectrum disorders. *Eur. Psychiatry* **38**, 61–69 (2016).
33. Kim, S.-W. *et al.* Gender-specific Associations of the Brain-derived Neurotrophic Factor Val66Met Polymorphism with Neurocognitive and Clinical Features in Schizophrenia. *Clin Psychopharmacol Neurosci* **14**, 270–278 (2016).
34. Ursini, G. *et al.* BDNF rs6265 methylation and genotype interact on risk for schizophrenia. *Epigenetics* **11**, 11–23 (2016).

35. Zhang, X. Y. *et al.* BDNF polymorphisms are associated with schizophrenia onset and positive symptoms. *Schizophr. Res.* **170**, 41–47 (2016).
36. Kheirollahi, M., Kazemi, E. & Ashouri, S. Brain-Derived Neurotrophic Factor Gene Val66Met Polymorphism and Risk of Schizophrenia: A Meta-analysis of Case-Control Studies. *Cell. Mol. Neurobiol.* **36**, 1–10 (2016).
37. Ahmed, A. O., Mantini, A. M., Fridberg, D. J. & Buckley, P. F. Brain-derived neurotrophic factor (BDNF) and neurocognitive deficits in people with schizophrenia: a meta-analysis. *Psychiatry Res* **226**, 1–13 (2015).
38. Zakharyan, R. & Boyajyan, A. Brain-derived neurotrophic factor blood levels are decreased in schizophrenia patients and associate with rs6265 genotypes. *Clin. Biochem.* **47**, 1052–1055 (2014).
39. Chen, S.-L. *et al.* The BDNF Val66Met polymorphism and plasma brain-derived neurotrophic factor levels in Han Chinese patients with bipolar disorder and schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **51**, 99–104 (2014).
40. Pełka-Wysiecka, J. *et al.* BDNF rs 6265 polymorphism and COMT rs 4680 polymorphism in deficit schizophrenia in Polish sample. *Pharmacol Rep* **65**, 1185–1193 (2013).
41. golimbet, V. E., Alfimova, M. V., Korovaitseva, G. I. & Lezheiko, T. V. [The moderating effect of the Va166Met polymorphism of brain-derived neurotrophic factor gene on the clinical and psychological features of patients with schizophrenia]. *Mol. Biol. (Mosk.)* **48**, 81–88 (2014).
42. Zhai, J. *et al.* Association of the brain-derived neurotrophic factor gene G196A rs6265 polymorphisms and the cognitive function and clinical symptoms of schizophrenia. *Int J Clin Exp Pathol* **6**, 1617–1623 (2013).

43. Li, W. *et al.* Association of BDNF gene polymorphisms with schizophrenia and clinical symptoms in a Chinese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **162B**, 538–545 (2013).
44. Suchanek, R., Owczarek, A. & Kowalski, J. Association study between BDNF C-281A polymorphism and paranoid schizophrenia in Polish population. *J. Mol. Neurosci.* **46**, 217–222 (2012).
45. Watanabe, Y., Nunokawa, A. & Someya, T. Association of the BDNF C270T polymorphism with schizophrenia: updated meta-analysis. *Psychiatry Clin. Neurosci.* **67**, 123–125 (2013).
46. Sotiropoulou, M. *et al.* BDNF serum concentrations in first psychotic episode drug-naïve schizophrenic patients: associations with personality and BDNF Val66Met polymorphism. *Life Sci.* **92**, 305–310 (2013).
47. Eisenberg, D. P. *et al.* Brain-derived neurotrophic factor (BDNF) Val(66)Met polymorphism differentially predicts hippocampal function in medication-free patients with schizophrenia. *Mol. Psychiatry* **18**, 713–720 (2013).
48. Galaktionova, D. I. *et al.* [An association study of polymorphisms in HTR2A, BDNF and SLC6A4 genes with paranoid schizophrenia and suicidal behavior]. *Zh Nevrol Psikhiatr Im S S Korsakova* **112**, 39–44 (2012).
49. Vyas, N. S. & Puri, B. K. Evidence for an association between brain-derived neurotrophic factor Val66Met gene polymorphism and general intellectual ability in early-onset schizophrenia. *Isr J Psychiatry Relat Sci* **49**, 137–142 (2012).

50. Sun, M. *et al.* [Association study of brain-derived neurotrophic factor Val66Met polymorphism and clinical characteristics of first episode schizophrenia]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **29**, 155–158 (2012).
51. Lu, W. *et al.* Association between BDNF Val66Met polymorphism and cognitive performance in antipsychotic-naïve patients with schizophrenia. *J. Mol. Neurosci.* **47**, 505–510 (2012).
52. Zhang, X. Y. *et al.* Cognitive and serum BDNF correlates of BDNF Val66Met gene polymorphism in patients with schizophrenia and normal controls. *Hum. Genet.* **131**, 1187–1195 (2012).
53. Zai, C. C. *et al.* Association study of BDNF and DRD3 genes in schizophrenia diagnosis using matched case-control and family based study designs. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **34**, 1412–1418 (2010).
54. Spalletta, G. *et al.* BDNF Val66Met polymorphism is associated with aggressive behavior in schizophrenia. *Eur. Psychiatry* **25**, 311–313 (2010).
55. Zhou, D. H. *et al.* The study of BDNF Val66Met polymorphism in Chinese schizophrenic patients. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **34**, 930–933 (2010).
56. Squassina, A. *et al.* NRG1 and BDNF genes in schizophrenia: an association study in an Italian case-control sample. *Psychiatry Res* **176**, 82–84 (2010).
57. Rizos, E. N. *et al.* Association of serum BDNF and val66met polymorphism of the brain-derived neurotrophic factor in a sample of first psychotic episode patients. *Psychiatriki* **20**, 297–304 (2009).

58. Golimbet, V. E., Korovaïtseva, G. I., Abramova, L. I., Kasparov, S. V. & Uvarova, L. G. [Association between the Val66Met polymorphism of brain-derived neurotrophic factor gene and schizophrenia in Russians]. *Mol. Biol. (Mosk.)* **42**, 599–603 (2008).
59. Takahashi, T. *et al.* Association between the brain-derived neurotrophic factor Val66Met polymorphism and brain morphology in a Japanese sample of schizophrenia and healthy comparisons. *Neurosci. Lett.* **435**, 34–39 (2008).
60. Han, D. H. *et al.* Effects of brain-derived neurotrophic factor-catecholamine-O-methyltransferase gene interaction on schizophrenic symptoms. *Neuroreport* **19**, 1155–1158 (2008).
61. Alfimova, M. V. *et al.* [Investigation of association of the brain-derived neurotrophic factor (BDNF) and a serotonin receptor 2A (5-HTR2A) genes with voluntary and involuntary attention in schizophrenia]. *Zh Nevrol Psikhiatr Im S S Korsakova* **108**, 62–69 (2008).
62. Huang, T.-L. & Lee, C.-T. Associations between brain-derived neurotrophic factor G196A gene polymorphism and clinical phenotypes in schizophrenia patients. *Chang Gung Med J* **30**, 408–413 (2007).
63. Chao, H. M., Kao, H.-T. & Porton, B. BDNF Val66Met variant and age of onset in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **147B**, 505–506 (2008).
64. Zintzaras, E. Brain-derived neurotrophic factor gene polymorphisms and schizophrenia: a meta-analysis. *Psychiatr. Genet.* **17**, 69–75 (2007).
65. Xu, M.-Q., St Clair, D., Ott, J., Feng, G.-Y. & He, L. Brain-derived neurotrophic factor gene C-270T and Val66Met functional polymorphisms and risk of schizophrenia: a moderate-scale population-based study and meta-analysis. *Schizophr. Res.* **91**, 6–13 (2007).

66. Gratacòs, M. *et al.* Brain-derived neurotrophic factor Val66Met and psychiatric disorders: meta-analysis of case-control studies confirm association to substance-related disorders, eating disorders, and schizophrenia. *Biol. Psychiatry* **61**, 911–922 (2007).
67. Qian, L. *et al.* Brain-derived neurotrophic factor and risk of schizophrenia: an association study and meta-analysis. *Biochem. Biophys. Res. Commun.* **353**, 738–743 (2007).
68. Numata, S. *et al.* Interaction between catechol-O-methyltransferase (COMT) Val108/158Met and brain-derived neurotrophic factor (BDNF) Val66Met polymorphisms in age at onset and clinical symptoms in schizophrenia. *J Neural Transm (Vienna)* **114**, 255–259 (2007).
69. Numata, S. *et al.* Brain-derived neurotrophic factor (BDNF) Val66Met polymorphism in schizophrenia is associated with age at onset and symptoms. *Neurosci. Lett.* **401**, 1–5 (2006).
70. Chen, C.-Y., Lu, R.-B., Yeh, Y.-W., Shih, M.-C. & Huang, S.-Y. Association study of catechol-O-methyltransferase gene polymorphisms with schizophrenia and psychopathological symptoms in Han Chinese. *Genes Brain Behav.* **10**, 316–324 (2011).
71. Gourion, D. *et al.* Age at onset of schizophrenia: interaction between brain-derived neurotrophic factor and dopamine D3 receptor gene variants. *Neuroreport* **16**, 1407–1410 (2005).
72. Neves-Pereira, M. *et al.* BDNF gene is a risk factor for schizophrenia in a Scottish population. *Mol. Psychiatry* **10**, 208–212 (2005).
73. Szekeres, G., Juhász, A., Rimanóczy, A., Kéri, S. & Janka, Z. The C270T polymorphism of the brain-derived neurotrophic factor gene is associated with schizophrenia. *Schizophr. Res.* **65**, 15–18 (2003).

74. Nanko, S. *et al.* Brain-derived neurotrophic factor gene and schizophrenia: polymorphism screening and association analysis. *Schizophr. Res.* **62**, 281–283 (2003).
75. Koolschijn, P. C. M. P. *et al.* Effects of brain-derived neurotrophic factor Val66Met polymorphism on hippocampal volume change in schizophrenia. *Hippocampus* **20**, 1010–1017 (2010).
76. Varnäs, K. *et al.* Brain-derived neurotrophic factor polymorphisms and frontal cortex morphology in schizophrenia. *Psychiatr. Genet.* **18**, 177–183 (2008).
77. Ho, B.-C., Andreasen, N. C., Dawson, J. D. & Wassink, T. H. Association between brain-derived neurotrophic factor Val66Met gene polymorphism and progressive brain volume changes in schizophrenia. *Am J Psychiatry* **164**, 1890–1899 (2007).
78. Ho, B.-C. *et al.* Cognitive and magnetic resonance imaging brain morphometric correlates of brain-derived neurotrophic factor Val66Met gene polymorphism in patients with schizophrenia and healthy volunteers. *Arch. Gen. Psychiatry* **63**, 731–740 (2006).
79. Agartz, I. *et al.* BDNF gene variants and brain morphology in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141B**, 513–523 (2006).
80. Szeszko, P. R. *et al.* Brain-derived neurotrophic factor val66met polymorphism and volume of the hippocampal formation. *Mol. Psychiatry* **10**, 631–636 (2005).
81. Wassink, T. H., Nelson, J. J., Crowe, R. R. & Andreasen, N. C. Heritability of BDNF alleles and their effect on brain morphology in schizophrenia. *Am. J. Med. Genet.* **88**, 724–728 (1999).
82. Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* **511**, 421–427 (2014).

83. Ripke, S. *et al.* Genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat. Genet.* **45**, 1150–1159 (2013).
84. Zhang, S.-Y. *et al.* Role of CACNA1C gene polymorphisms and protein expressions in the pathogenesis of schizophrenia: a case-control study in a Chinese population. *Neurol. Sci.* **38**, 1393–1403 (2017).
85. Takahashi, S., Glatt, S. J., Uchiyama, M., Faraone, S. V. & Tsuang, M. T. Meta-analysis of data from the Psychiatric Genomics Consortium and additional samples supports association of CACNA1C with risk for schizophrenia. *Schizophr. Res.* **168**, 429–433 (2015).
86. Porcelli, S. *et al.* CACNA1C gene and schizophrenia: a case-control and pharmacogenetic study. *Psychiatr. Genet.* **25**, 163–167 (2015).
87. Jiang, H. *et al.* Evaluating the association between CACNA1C rs1006737 and schizophrenia risk: A meta-analysis. *Asia Pac Psychiatry* **7**, 260–267 (2015).
88. Zheng, F. *et al.* Further evidence for genetic association of CACNA1C and schizophrenia: new risk loci in a Han Chinese population and a meta-analysis. *Schizophr. Res.* **152**, 105–110 (2014).
89. Hamshere, M. L. *et al.* Genome-wide significant associations in schizophrenia to ITIH3/4, CACNA1C and SDCCAG8, and extensive replication of associations reported by the Schizophrenia PGC. *Mol. Psychiatry* **18**, 708–712 (2013).
90. Guan, F. *et al.* MIR137 gene and target gene CACNA1C of miR-137 contribute to schizophrenia susceptibility in Han Chinese. *Schizophr. Res.* **152**, 97–104 (2014).

91. Mallas, E. *et al.* The impact of CACNA1C gene, and its epistasis with ZNF804A, on white matter microstructure in health, schizophrenia and bipolar disorder¹. *Genes Brain Behav.* **16**, 479–488 (2017).
92. Huang, L. *et al.* The impact of CACNA1C allelic variation on regional gray matter volume in Chinese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **171B**, 396–401 (2016).
93. Sumner, J. A. *et al.* Variation in CACNA1C is Associated with Amygdala Structure and Function in Adolescents. *J Child Adolesc Psychopharmacol* **25**, 701–710 (2015).
94. Lancaster, T. M., Foley, S., Tansey, K. E., Linden, D. E. J. & Caseras, X. CACNA1C risk variant is associated with increased amygdala volume. *Eur Arch Psychiatry Clin Neurosci* **266**, 269–275 (2016).
95. Woon, P. S. *et al.* CACNA1C genomewide supported psychosis genetic variation affects cortical brain white matter integrity in Chinese patients with schizophrenia. *J Clin Psychiatry* **75**, e1284-1290 (2014).
96. Wolf, C. *et al.* CACNA1C genotype explains interindividual differences in amygdala volume among patients with schizophrenia. *Eur Arch Psychiatry Clin Neurosci* **264**, 93–102 (2014).
97. Wang, F., McIntosh, A. M., He, Y., Gelernter, J. & Blumberg, H. P. The association of genetic variation in CACNA1C with structure and function of a frontotemporal system. *Bipolar Disord* **13**, 696–700 (2011).
98. Franke, B. *et al.* Genetic variation in CACNA1C, a gene associated with bipolar disorder, influences brainstem rather than gray matter volume in healthy individuals. *Biol. Psychiatry* **68**, 586–588 (2010).

99. Guan, F. *et al.* Evaluation of genetic susceptibility of common variants in CACNA1D with schizophrenia in Han Chinese. *Sci Rep* **5**, 12935 (2015).
100. Terwisscha van Scheltinga, A. F. *et al.* Association study of fibroblast growth factor genes and brain volumes in schizophrenic patients and healthy controls. *Psychiatr. Genet.* **24**, 283–284 (2014).
101. Liao, D.-L. *et al.* Association of muscarinic m1 receptor genetic polymorphisms with psychiatric symptoms and cognitive function in schizophrenic patients. *Neuropsychobiology* **48**, 72–76 (2003).
102. Cropley, V. L. *et al.* The effect of a muscarinic receptor 1 gene variant on grey matter volume in schizophrenia. *Psychiatry Res* **234**, 182–187 (2015).
103. Miyauchi, M. *et al.* Association of the Cholinergic Muscarinic M2 Receptor with Autonomic Nervous System Activity in Patients with Schizophrenia on High-Dose Antipsychotics. *Neuropsychobiology* **74**, 60–67 (2016).
104. Kishi, T. *et al.* Genetic association analysis of tagging SNPs in alpha4 and beta2 subunits of neuronal nicotinic acetylcholine receptor genes (CHRNA4 and CHRN2) with schizophrenia in the Japanese population. *J Neural Transm (Vienna)* **115**, 1457–1461 (2008).
105. De Luca, V., Voineskos, S., Wong, G. & Kennedy, J. L. Genetic interaction between alpha4 and beta2 subunits of high affinity nicotinic receptor: analysis in schizophrenia. *Exp Brain Res* **174**, 292–296 (2006).
106. Markett, S., Reuter, M., Montag, C. & Weber, B. The dopamine D2 receptor gene DRD2 and the nicotinic acetylcholine receptor gene CHRNA4 interact on striatal gray matter volume: evidence from a genetic imaging study. *Neuroimage* **64**, 167–172 (2013).

107. Hong, L. E. *et al.* A CHRNA5 allele related to nicotine addiction and schizophrenia. *Genes Brain Behav.* **10**, 530–535 (2011).
108. Di Giorgio, A. *et al.* DRD2/CHRNA5 interaction on prefrontal biology and physiology during working memory. *PLoS ONE* **9**, e95997 (2014).
109. Aberg, K. A. *et al.* A comprehensive family-based replication study of schizophrenia genes. *JAMA Psychiatry* **70**, 573–581 (2013).
110. Guan, F. *et al.* Two-stage replication of previous genome-wide association studies of AS3MT-CNNM2-NT5C2 gene cluster region in a large schizophrenia case-control sample from Han Chinese population. *Schizophr. Res.* **176**, 125–130 (2016).
111. Genome-wide association study identifies five new schizophrenia loci. *Nat. Genet.* **43**, 969–976 (2011).
112. Rose, E. J. *et al.* Effects of a novel schizophrenia risk variant rs7914558 at CNNM2 on brain structure and attributional style. *Br J Psychiatry* **204**, 115–121 (2014).
113. Ohi, K. *et al.* The impact of the genome-wide supported variant in the cyclin M2 gene on gray matter morphology in schizophrenia. *Behav Brain Funct* **9**, 40 (2013).
114. Gouvêa, E. S. *et al.* The role of the CNR1 gene in schizophrenia: a systematic review including unpublished data. *Rev Bras Psiquiatr* **39**, 160–171 (2017).
115. Bae, J. S. *et al.* Genetic association analysis of CNR1 and CNR2 polymorphisms with schizophrenia in a Korean population. *Psychiatr. Genet.* **24**, 225–229 (2014).
116. Yu, W. *et al.* CNR1 gene and risk of the metabolic syndrome in patients with schizophrenia. *J Clin Psychopharmacol* **33**, 186–192 (2013).

117. Ujike, H. *et al.* CNR1, central cannabinoid receptor gene, associated with susceptibility to hebephrenic schizophrenia. *Mol. Psychiatry* **7**, 515–518 (2002).
118. Tsai, S. J., Wang, Y. C. & Hong, C. J. Association study of a cannabinoid receptor gene (CNR1) polymorphism and schizophrenia. *Psychiatr. Genet.* **10**, 149–151 (2000).
119. Suárez-Pinilla, P. *et al.* Brain structural and clinical changes after first episode psychosis: Focus on cannabinoid receptor 1 polymorphisms. *Psychiatry Res* **233**, 112–119 (2015).
120. Ji, W. *et al.* CNTNAP2 is significantly associated with schizophrenia and major depression in the Han Chinese population. *Psychiatry Res* **207**, 225–228 (2013).
121. Clemm von Hohenberg, C. *et al.* CNTNAP2 polymorphisms and structural brain connectivity: a diffusion-tensor imaging study. *J Psychiatr Res* **47**, 1349–1356 (2013).
122. Uddén, J., Snijders, T. M., Fisher, S. E. & Hagoort, P. A common variant of the CNTNAP2 gene is associated with structural variation in the left superior occipital gyrus. *Brain Lang* **172**, 16–21 (2017).
123. Zhu, B. *et al.* Associations between the CNTNAP2 gene, dorsolateral prefrontal cortex, and cognitive performance on the Stroop task. *Neuroscience* **343**, 21–29 (2017).
124. Han, D. H. *et al.* Effects of brain-derived neurotrophic factor-catecholamine-O-methyltransferase gene interaction on schizophrenic symptoms. *Neuroreport* **19**, 1155–1158 (2008).
125. Matsuzaka, C. T. *et al.* Catechol-O-methyltransferase (COMT) polymorphisms modulate working memory in individuals with schizophrenia and healthy controls. *Rev Bras Psiquiatr* **39**, 302–308 (2017).

126. Mao, Q. *et al.* Association of catechol-O-methyltransferase Val(108/158) Met genetic polymorphism with schizophrenia, P50 sensory gating, and negative symptoms in a Chinese population. *Psychiatry Res* **242**, 271–276 (2016).
127. González-Castro, T. B. *et al.* The Role of a Catechol-O-Methyltransferase (COMT) Val158Met Genetic Polymorphism in Schizophrenia: A Systematic Review and Updated Meta-analysis on 32,816 Subjects. *Neuromolecular Med.* **18**, 216–231 (2016).
128. Higashiyama, R. *et al.* Association of copy number polymorphisms at the promoter and translated region of COMT with Japanese patients with schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **171B**, 447–457 (2016).
129. Behbahani, P., Kazemi-Nezhad, S. R., Foroughmand, A. M. & Ahmadi, L. Association study of single nucleotide polymorphism rs165599 of COMT gene, with schizophrenia and bipolar mood disorder in the south-west of Iran. *Mol Biol Res Commun* **4**, 67–72 (2015).
130. Vijayakumari, A. A. *et al.* Effect of polymorphisms of three genes mediating monoamine signalling on brain morphometry in schizophrenia and healthy subjects. *Clin Psychopharmacol Neurosci* **13**, 68–82 (2015).
131. Al-Asmary, S., Kadasah, S., Arfin, M., Tariq, M. & Al-Asmari, A. Genetic association of catechol-O-methyltransferase val(158)met polymorphism in Saudi schizophrenia patients. *Genet. Mol. Res.* **13**, 3079–3088 (2014).
132. Tsuchimine, S., Yasui-Furukori, N., Kaneda, A. & Kaneko, S. Differential effects of the catechol-O-methyltransferase Val158Met genotype on the cognitive function of schizophrenia patients and healthy Japanese individuals. *PLoS ONE* **8**, e76763 (2013).

133. Alfimova, M. V. *et al.* [The association of COMT and DRD2 gene polymorphisms with a cognitive ability to understand others in schizophrenic patients]. *Zh Nevrol Psikhiatr Im S S Korsakova* **113**, 50–56 (2013).
134. Liu, X. *et al.* Association study of polymorphisms in the alpha 7 nicotinic acetylcholine receptor subunit and catechol-o-methyl transferase genes with sensory gating in first-episode schizophrenia. *Psychiatry Res* **209**, 431–438 (2013).
135. Cordeiro, Q., Silva, R. T. da & Vallada, H. Association study between the rs165599 catechol-O-methyltransferase genetic polymorphism and schizophrenia in a Brazilian sample. *Arq Neuropsiquiatr* **70**, 913–916 (2012).
136. Bhakta, S. G., Zhang, J.-P. & Malhotra, A. K. The COMT Met158 allele and violence in schizophrenia: a meta-analysis. *Schizophr. Res.* **140**, 192–197 (2012).
137. Wright, G. E. B. *et al.* Association of MB-COMT polymorphisms with schizophrenia-susceptibility and symptom severity in an African cohort. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **39**, 163–169 (2012).
138. Kamath, V., Moberg, P. J., Gur, R. E., Doty, R. L. & Turetsky, B. I. Effects of the val(158)met catechol-O-methyltransferase gene polymorphism on olfactory processing in schizophrenia. *Behav. Neurosci.* **126**, 209–215 (2012).
139. Tee, S. F., Tang, P. Y. & Loh, H. C. COMT haplotype analyses in Malaysians with schizophrenia. *Psychiatry Res* **195**, 83–84 (2012).
140. Raznahan, A. *et al.* Catechol-o-methyl transferase (COMT) val158met polymorphism and adolescent cortical development in patients with childhood-onset schizophrenia, their non-psychotic siblings, and healthy controls. *Neuroimage* **57**, 1517–1523 (2011).

141. Kong, F., Peng, Z., Jiang, T. & Hong, X. [An association study of COMT gene polymorphisms with schizophrenia]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **28**, 208–211 (2011).
142. Boot, E. *et al.* Dopamine metabolism in adults with 22q11 deletion syndrome, with and without schizophrenia--relationship with COMT Val¹⁰⁸/¹⁵⁸Met polymorphism, gender and symptomatology. *J. Psychopharmacol. (Oxford)* **25**, 888–895 (2011).
143. Wan, C.-L., Zainal, N. Z., Lian, L.-H. & Mohamed, Z. Association of the functional polymorphism in the catechol-O-methyltransferase gene with schizophrenia in the three ethnic groups of the Malaysian population. *Psychiatry Res* **189**, 67–71 (2011).
144. Voisey, J. *et al.* HapMap tag-SNP analysis confirms a role for COMT in schizophrenia risk and reveals a novel association. *Eur. Psychiatry* **27**, 372–376 (2012).
145. Paweł, K. *et al.* [Family based association study of DRD1, DRD2, DRD3, DRD4, DAT, COMT gene polymorphism in schizophrenia]. *Psychiatr. Pol.* **44**, 405–413 (2010).
146. Nieratschker, V. *et al.* The catechol-O-methyl transferase (COMT) gene and its potential association with schizophrenia: findings from a large German case-control and family-based sample. *Schizophr. Res.* **122**, 24–30 (2010).
147. Sagud, M. *et al.* Catechol-O-methyl transferase and schizophrenia. *Psychiatr Danub* **22**, 270–274 (2010).
148. Costas, J. *et al.* Heterozygosity at catechol-O-methyltransferase Val158Met and schizophrenia: new data and meta-analysis. *J Psychiatr Res* **45**, 7–14 (2011).
149. Wang, Y., Fang, Y., Shen, Y. & Xu, Q. Analysis of association between the catechol-O-methyltransferase (COMT) gene and negative symptoms in chronic schizophrenia. *Psychiatry Res* **179**, 147–150 (2010).

150. Kang, C., Xu, X., Liu, H. & Yang, J. Association study of catechol-O-methyltransferase (COMT) gene Val158Met polymorphism with auditory P300 in Chinese Han patients with schizophrenia. *Psychiatry Res* **180**, 153–155 (2010).
151. Park, B. L. *et al.* Association analysis of COMT polymorphisms with schizophrenia and smooth pursuit eye movement abnormality. *J. Hum. Genet.* **54**, 709–712 (2009).
152. Gu, Y., Yun, L., Tian, Y. & Hu, Z. Association between COMT gene and Chinese male schizophrenic patients with violent behavior. *Med. Sci. Monit.* **15**, CR484-489 (2009).
153. Haraldsson, H. M. *et al.* COMT val(158)met genotype and smooth pursuit eye movements in schizophrenia. *Psychiatry Res* **169**, 173–175 (2009).
154. Neuhaus, A. H. *et al.* COMT Val 158 Met polymorphism is associated with cognitive flexibility in a signal discrimination task in schizophrenia. *Pharmacopsychiatry* **42**, 141–144 (2009).
155. Chien, Y.-L., Liu, C.-M., Fann, C. S.-J., Liu, Y.-L. & Hwu, H.-G. Association of the 3' region of COMT with schizophrenia in Taiwan. *J. Formos. Med. Assoc.* **108**, 301–309 (2009).
156. Hoenicka, J. *et al.* Gender-specific COMT Val158Met polymorphism association in Spanish schizophrenic patients. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 79–85 (2010).
157. Okochi, T. *et al.* Meta-analysis of association between genetic variants in COMT and schizophrenia: an update. *Schizophr. Res.* **110**, 140–148 (2009).
158. Mas, S. *et al.* [Effect of polymorphisms of the catechol-O-methyltransferase on schizophrenia risk in a Spanish population]. *Med Clin (Barc)* **131**, 761–764 (2008).

159. Kim, Y.-R., Kim, J. H., Kim, S. J., Lee, D. & Min, S. K. Catechol-O-methyltransferase Val158Met polymorphism in relation to aggressive schizophrenia in a Korean population. *Eur Neuropsychopharmacol* **18**, 820–825 (2008).
160. Quednow, B. B., Wagner, M., Mössner, R., Maier, W. & Kühn, K.-U. Sensorimotor gating of schizophrenia patients depends on Catechol O-methyltransferase Val158Met polymorphism. *Schizophr Bull* **36**, 341–346 (2010).
161. Haraldsson, H. M. *et al.* Catechol-O-methyltransferase Val 158 Met polymorphism and antisaccade eye movements in schizophrenia. *Schizophr Bull* **36**, 157–164 (2010).
162. Hong, J. P. *et al.* New functional single nucleotide polymorphism (Ala72Ser) in the COMT gene is associated with aggressive behavior in male schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **147B**, 658–660 (2008).
163. Muntjewerff, J.-W. *et al.* Polymorphisms in catechol-O-methyltransferase and methylenetetrahydrofolate reductase in relation to the risk of schizophrenia. *Eur Neuropsychopharmacol* **18**, 99–106 (2008).
164. Yu, R., Zhang, X.-N., Huang, X.-X., Ding, S.-P. & Li, J.-C. Association analysis of COMT polymorphisms and schizophrenia in a Chinese Han population: a case-control study. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **144B**, 570–573 (2007).
165. Barnett, J. H., Jones, P. B., Robbins, T. W. & Müller, U. Effects of the catechol-O-methyltransferase Val158Met polymorphism on executive function: a meta-analysis of the Wisconsin Card Sort Test in schizophrenia and healthy controls. *Mol. Psychiatry* **12**, 502–509 (2007).

166. Díez-Martín, J. *et al.* [COMT Val158Met polymorphism and schizophrenia in a series of Spanish patients]. *Med Clin (Barc)* **128**, 41–44 (2007).
167. Bassett, A. S., Caluseriu, O., Weksberg, R., Young, D. A. & Chow, E. W. C. Catechol-O-methyl transferase and expression of schizophrenia in 73 adults with 22q11 deletion syndrome. *Biol. Psychiatry* **61**, 1135–1140 (2007).
168. Golimbet, V. *et al.* Association study of COMT gene Val158Met polymorphism with auditory P300 and performance on neurocognitive tests in patients with schizophrenia and their relatives. *World J. Biol. Psychiatry* **7**, 238–245 (2006).
169. Nicodemus, K. K. *et al.* Evidence for statistical epistasis between catechol-O-methyltransferase (COMT) and polymorphisms in RGS4, G72 (DAOA), GRM3, and DISC1: influence on risk of schizophrenia. *Hum. Genet.* **120**, 889–906 (2007).
170. Han, D. H. *et al.* Effects of catechol-O-methyltransferase Val158Met polymorphism on the cognitive stability and aggression in the first-onset schizophrenic patients. *Neuroreport* **17**, 95–99 (2006).
171. Strous, R. D., Lapidus, R., Viglin, D., Kotler, M. & Lachman, H. M. Analysis of an association between the COMT polymorphism and clinical symptomatology in schizophrenia. *Neurosci. Lett.* **393**, 170–173 (2006).
172. Fan, J. B. *et al.* Family-based association studies of COMT gene polymorphisms and schizophrenia in the Chinese population. *Mol. Psychiatry* **7**, 446–447 (2002).
173. Lee, S.-G. *et al.* Association of Ala72Ser polymorphism with COMT enzyme activity and the risk of schizophrenia in Koreans. *Hum. Genet.* **116**, 319–328 (2005).

174. Thaker, G. K., Wonodi, I., Avila, M. T., Hong, L. E. & Stine, O. C. Catechol O-methyltransferase polymorphism and eye tracking in schizophrenia: a preliminary report. *Am J Psychiatry* **161**, 2320–2322 (2004).
175. Sazci, A. *et al.* Catechol-O-methyltransferase gene Val108/158Met polymorphism, and susceptibility to schizophrenia: association is more significant in women. *Brain Res. Mol. Brain Res.* **132**, 51–56 (2004).
176. Handoko, H. Y. *et al.* Separate and interacting effects within the catechol-O-methyltransferase (COMT) are associated with schizophrenia. *Mol. Psychiatry* **10**, 589–597 (2005).
177. Zammit, S. *et al.* Polymorphisms in the MAOA, MAOB, and COMT genes and aggressive behavior in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **128B**, 19–20 (2004).
178. Chen, X., Wang, X., O'Neill, A. F., Walsh, D. & Kendler, K. S. Variants in the catechol-o-methyltransferase (COMT) gene are associated with schizophrenia in Irish high-density families. *Mol. Psychiatry* **9**, 962–967 (2004).
179. Tsai, S.-J., Hong, C.-J., Liao, D.-L., Lai, I.-C. & Liou, Y.-J. Association study of a functional catechol-O-methyltransferase genetic polymorphism with age of onset, cognitive function, symptomatology and prognosis in chronic schizophrenia. *Neuropsychobiology* **49**, 196–200 (2004).
180. Palmatier, M. A. *et al.* COMT haplotypes suggest P2 promoter region relevance for schizophrenia. *Mol. Psychiatry* **9**, 859–870 (2004).

181. Goldberg, T. E. *et al.* Executive subprocesses in working memory: relationship to catechol-O-methyltransferase Val158Met genotype and schizophrenia. *Arch. Gen. Psychiatry* **60**, 889–896 (2003).
182. Wonodi, I., Stine, O. C., Mitchell, B. D., Buchanan, R. W. & Thaker, G. K. Association between Val108/158 Met polymorphism of the COMT gene and schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **120B**, 47–50 (2003).
183. Glatt, S. J., Faraone, S. V. & Tsuang, M. T. Association between a functional catechol O-methyltransferase gene polymorphism and schizophrenia: meta-analysis of case-control and family-based studies. *Am J Psychiatry* **160**, 469–476 (2003).
184. Rybakowski, J. K., Borkowska, A., Czerski, P. M. & Hauser, J. Eye movement disturbances in schizophrenia and a polymorphism of catechol-O-methyltransferase gene. *Psychiatry Res* **113**, 49–57 (2002).
185. Shifman, S. *et al.* A highly significant association between a COMT haplotype and schizophrenia. *Am. J. Hum. Genet.* **71**, 1296–1302 (2002).
186. Park, T.-W. *et al.* Functional catechol-O-methyltransferase gene polymorphism and susceptibility to schizophrenia. *Eur Neuropsychopharmacol* **12**, 299–303 (2002).
187. Herken, H. & Erdal, M. E. Catechol-O-methyltransferase gene polymorphism in schizophrenia: evidence for association between symptomatology and prognosis. *Psychiatr. Genet.* **11**, 105–109 (2001).
188. Liou, Y. J., Tsai, S. J., Hong, C. J., Wang, Y. C. & Lai, I. C. Association analysis of a functional catechol-o-methyltransferase gene polymorphism in schizophrenic patients in Taiwan. *Neuropsychobiology* **43**, 11–14 (2001).

189. Chen, C. H. *et al.* Systematic mutation analysis of the catechol O-methyltransferase gene as a candidate gene for schizophrenia. *Am J Psychiatry* **156**, 1273–1275 (1999).
190. Ohmori, O. *et al.* Association study of a functional catechol-O-methyltransferase gene polymorphism in Japanese schizophrenics. *Neurosci. Lett.* **243**, 109–112 (1998).
191. Karayiorgou, M. *et al.* Identification of sequence variants and analysis of the role of the catechol-O-methyl-transferase gene in schizophrenia susceptibility. *Biol. Psychiatry* **43**, 425–431 (1998).
192. Kunugi, H. *et al.* Catechol-O-methyltransferase polymorphisms and schizophrenia: a transmission disequilibrium study in multiply affected families. *Psychiatr. Genet.* **7**, 97–101 (1997).
193. Chen, C. H. *et al.* Identification of a BglI polymorphism of catechol-O-methyltransferase (COMT) gene, and association study with schizophrenia. *Am. J. Med. Genet.* **67**, 556–559 (1996).
194. Li, T. *et al.* Preferential transmission of the high activity allele of COMT in schizophrenia. *Psychiatr. Genet.* **6**, 131–133 (1996).
195. Bollettini, I. *et al.* Sexually divergent effect of COMT Val/met genotype on subcortical volumes in schizophrenia. *Brain Imaging Behav* (2017). doi:10.1007/s11682-017-9748-1
196. Shukla, A. A. *et al.* COMT val158met polymorphism and molecular alterations in the human dorsolateral prefrontal cortex: Differences in controls and in schizophrenia. *Schizophr. Res.* **173**, 94–100 (2016).
197. Poletti, S. *et al.* The COMT Val158Met polymorphism moderates the association between cognitive functions and white matter microstructure in schizophrenia. *Psychiatr. Genet.* **26**, 193–202 (2016).

198. Li, Y. *et al.* [Difference in brain surface area between first-episode familial and sporadic schizophrenia and its association with COMT gene polymorphisms]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **32**, 259–263 (2015).
199. Li, M.-L. *et al.* Morphological changes in gray matter volume correlate with catechol-O-methyl transferase gene Val158Met polymorphism in first-episode treatment-naïve patients with schizophrenia. *Neurosci Bull* **31**, 31–42 (2015).
200. Ehrlich, S. *et al.* The COMT Val108/158Met polymorphism and medial temporal lobe volumetry in patients with schizophrenia and healthy adults. *Neuroimage* **53**, 992–1000 (2010).
201. Honea, R. *et al.* Impact of interacting functional variants in COMT on regional gray matter volume in human brain. *Neuroimage* **45**, 44–51 (2009).
202. Zinkstok, J. *et al.* Genetic variation in COMT and PRODH is associated with brain anatomy in patients with schizophrenia. *Genes Brain Behav.* **7**, 61–69 (2008).
203. Ohnishi, T. *et al.* The association between the Val158Met polymorphism of the catechol-O-methyl transferase gene and morphological abnormalities of the brain in chronic schizophrenia. *Brain* **129**, 399–410 (2006).
204. Ho, B.-C., Wassink, T. H., O’Leary, D. S., Sheffield, V. C. & Andreasen, N. C. Catechol-O-methyl transferase Val158Met gene polymorphism in schizophrenia: working memory, frontal lobe MRI morphology and frontal cerebral blood flow. *Mol. Psychiatry* **10**, 229, 287–298 (2005).
205. Radua, J. *et al.* COMT Val158Met × SLC6A4 5-HTTLPR interaction impacts on gray matter volume of regions supporting emotion processing. *Soc Cogn Affect Neurosci* **9**, 1232–1238 (2014).

206. Cerasa, A. *et al.* Impact of catechol-O-methyltransferase Val(108/158) Met genotype on hippocampal and prefrontal gray matter volume. *Neuroreport* **19**, 405–408 (2008).
207. Tian, T. *et al.* Catechol-O-methyltransferase Val158Met polymorphism modulates gray matter volume and functional connectivity of the default mode network. *PLoS ONE* **8**, e78697 (2013).
208. Dutt, A. *et al.* COMT gene polymorphism and corpus callosum morphometry in preterm born adults. *Neuroimage* **54**, 148–153 (2011).
209. Tay, J. K. X., Tan, C. H., Chong, S.-A. & Tan, E.-C. Functional polymorphisms of the cytochrome P450 1A2 (CYP1A2) gene and prolonged QTc interval in schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **31**, 1297–1302 (2007).
210. Matsumoto, C. *et al.* Association study between functional polymorphisms in the cytochrome P450 1A2 and 2D6 genes and polydipsia in schizophrenia. *Neuromolecular Med.* **8**, 381–388 (2006).
211. Chu, C.-S. *et al.* The DAOA gene is associated with schizophrenia in the Taiwanese population. *Psychiatry Res* **252**, 201–207 (2017).
212. Sacchetti, E. *et al.* Schizophrenia susceptibility and NMDA-receptor mediated signalling: an association study involving 32 tagSNPs of DAO, DAOA, PPP3CC, and DTNBP1 genes. *BMC Med. Genet.* **14**, 33 (2013).
213. Chiesa, A. *et al.* DAOA variants and schizophrenia: influence on diagnosis and treatment outcomes. *Int J Psychiatry Clin Pract* **15**, 303–310 (2011).
214. Li, Y. *et al.* [Correlation of D-amino acid-oxidase gene polymorphism to schizophrenia]. *Nan Fang Yi Ke Da Xue Xue Bao* **30**, 2142–2144 (2010).

215. Müller, D. J., Zai, C. C., Shinkai, T., Strauss, J. & Kennedy, J. L. Association between the DAOA/G72 gene and bipolar disorder and meta-analyses in bipolar disorder and schizophrenia. *Bipolar Disord* **13**, 198–207 (2011).
216. Réthelyi, J. M. *et al.* Association study of NRG1, DTNBP1, RGS4, G72/G30, and PIP5K2A with schizophrenia and symptom severity in a Hungarian sample. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 792–801 (2010).
217. Ma, J. *et al.* Evidence for transmission disequilibrium at the DAOA gene locus in a schizophrenia family sample. *Neurosci. Lett.* **462**, 105–108 (2009).
218. Jönsson, E. G. *et al.* DTNBP1, NRG1, DAOA, DAO and GRM3 polymorphisms and schizophrenia: an association study. *Neuropsychobiology* **59**, 142–150 (2009).
219. Ohi, K. *et al.* Association study of the G72 gene with schizophrenia in a Japanese population: a multicenter study. *Schizophr. Res.* **109**, 80–85 (2009).
220. Shi, J. *et al.* Further evidence for an association of G72/G30 with schizophrenia in Chinese. *Schizophr. Res.* **107**, 324–326 (2009).
221. Opgen-Rhein, C. *et al.* Genetic variation in the DAOA gene complex: impact on susceptibility for schizophrenia and on cognitive performance. *Schizophr. Res.* **103**, 169–177 (2008).
222. Shin, H. D. *et al.* Association analysis of G72/G30 polymorphisms with schizophrenia in the Korean population. *Schizophr. Res.* **96**, 119–124 (2007).
223. Shinkai, T. *et al.* Association analyses of the DAOA/G30 and D-amino-acid oxidase genes in schizophrenia: further evidence for a role in schizophrenia. *Neuromolecular Med.* **9**, 169–177 (2007).

224. Corvin, A. *et al.* Evidence for association and epistasis at the DAOA/G30 and D-amino acid oxidase loci in an Irish schizophrenia sample. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **144B**, 949–953 (2007).
225. Yue, W. *et al.* Association of DAOA polymorphisms with schizophrenia and clinical symptoms or therapeutic effects. *Neurosci. Lett.* **416**, 96–100 (2007).
226. Li, D. & He, L. G72/G30 genes and schizophrenia: a systematic meta-analysis of association studies. *Genetics* **175**, 917–922 (2007).
227. Hong, C.-J., Hou, S.-J., Yen, F.-C., Liou, Y.-J. & Tsai, S.-J. Family-based association study between G72/G30 genetic polymorphism and schizophrenia. *Neuroreport* **17**, 1067–1069 (2006).
228. Goldberg, T. E. *et al.* The G72/G30 gene complex and cognitive abnormalities in schizophrenia. *Neuropsychopharmacology* **31**, 2022–2032 (2006).
229. Korostishevsky, M. *et al.* Transmission disequilibrium and haplotype analyses of the G72/G30 locus: suggestive linkage to schizophrenia in Palestinian Arabs living in the North of Israel. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141B**, 91–95 (2006).
230. Zou, F. *et al.* A family-based study of the association between the G72/G30 genes and schizophrenia in the Chinese population. *Schizophr. Res.* **73**, 257–261 (2005).
231. Wang, X. *et al.* Association of G72/G30 with schizophrenia in the Chinese population. *Biochem. Biophys. Res. Commun.* **319**, 1281–1286 (2004).
232. Addington, A. M. *et al.* Polymorphisms in the 13q33.2 gene G72/G30 are associated with childhood-onset schizophrenia and psychosis not otherwise specified. *Biol. Psychiatry* **55**, 976–980 (2004).

233. Schultz, C. C. *et al.* Reduced cortical thickness is associated with the glutamatergic regulatory gene risk variant DAOA Arg30Lys in schizophrenia. *Neuropsychopharmacology* **36**, 1747–1753 (2011).
234. Hartz, S. M. *et al.* G72 influences longitudinal change in frontal lobe volume in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 640–647 (2010).
235. Grant, A., Fathalli, F., Rouleau, G., Joober, R. & Flores, C. Association between schizophrenia and genetic variation in DCC: a case-control study. *Schizophr. Res.* **137**, 26–31 (2012).
236. Yan, P. *et al.* An Association Study Between Genetic Polymorphisms in Functional Regions of Five Genes and the Risk of Schizophrenia. *J. Mol. Neurosci.* **59**, 366–375 (2016).
237. Smeland, O. B. *et al.* Genetic Overlap Between Schizophrenia and Volumes of Hippocampus, Putamen, and Intracranial Volume Indicates Shared Molecular Genetic Mechanisms. *Schizophr Bull* (2017). doi:10.1093/schbul/sbx148
238. Meda, S. A. *et al.* Polymorphism of DCDC2 Reveals Differences in Cortical Morphology of Healthy Individuals-A Preliminary Voxel Based Morphometry Study. *Brain Imaging Behav* **2**, 21–26 (2008).
239. Jamadar, S. *et al.* Genetic influences of cortical gray matter in language-related regions in healthy controls and schizophrenia. *Schizophr. Res.* **129**, 141–148 (2011).
240. Xu, Y., Ren, J. & Ye, H. Association between variations in the disrupted in schizophrenia 1 gene and schizophrenia: A meta-analysis. *Gene* **651**, 94–99 (2018).

241. Mühle, C. *et al.* Additive sex-specific influence of common non-synonymous DISC1 variants on amygdala, basal ganglia, and white cortical surface area in healthy young adults. *Brain Struct Funct* **222**, 881–894 (2017).
242. He, B.-S. *et al.* Association of the DISC1 and NRG1 genetic polymorphisms with schizophrenia in a Chinese population. *Gene* **590**, 293–297 (2016).
243. Hu, G. *et al.* Association of schizophrenia with the rs821633 polymorphism in the DISC1 gene among Han Chinese. *Shanghai Arch Psychiatry* **27**, 348–355 (2015).
244. Luo, X. *et al.* Association study of DISC1 genetic variants with the risk of schizophrenia. *Psychiatr. Genet.* **26**, 132–135 (2016).
245. Arime, Y. *et al.* Effects of background mutations and single nucleotide polymorphisms (SNPs) on the Disc1 L100P behavioral phenotype associated with schizophrenia in mice. *Behav Brain Funct* **10**, 45 (2014).
246. Takahashi, T. *et al.* The Disrupted-in-Schizophrenia-1 Ser704Cys polymorphism and brain morphology in schizophrenia. *Psychiatry Res* **172**, 128–135 (2009).
247. Luo, X. *et al.* New findings support the association of DISC1 genetic variants with susceptibility to schizophrenia in the Han Chinese population. *Psychiatry Res* **228**, 966–968 (2015).
248. Gong, X. *et al.* A brain-wide association study of DISC1 genetic variants reveals a relationship with the structure and functional connectivity of the precuneus in schizophrenia. *Hum Brain Mapp* **35**, 5414–5430 (2014).

249. Ratta-Apha, W. *et al.* Association analysis of the DISC1 gene with schizophrenia in the Japanese population and DISC1 immunoreactivity in the postmortem brain. *Neurosci. Res.* **77**, 222–227 (2013).
250. Cao, F., Zhang, H., Feng, J., Gao, C. & Li, S. Association study of three microsatellite polymorphisms located in introns 1, 8, and 9 of DISC1 with schizophrenia in the Chinese Han population. *Genet Test Mol Biomarkers* **17**, 407–411 (2013).
251. Kinoshita, M. *et al.* Meta-analysis of association studies between DISC1 missense variants and schizophrenia in the Japanese population. *Schizophr. Res.* **141**, 271–273 (2012).
252. Wei, Q. *et al.* The effect of DISC1 on regional gray matter density of schizophrenia in Han Chinese population. *Neurosci. Lett.* **517**, 21–24 (2012).
253. Hotta, Y. *et al.* Association study between Disrupted-in-Schizophrenia-1 (DISC1) and Japanese patients with treatment-resistant schizophrenia (TRS). *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 636–639 (2011).
254. Norlelawati, A. T. *et al.* Disrupted-in-Schizophrenia-1 SNPs and Susceptibility to Schizophrenia: Evidence from Malaysia. *Psychiatry Investig* **12**, 103–111 (2015).
255. Ram Murthy, A. *et al.* Gender-specific association of TSNAX/DISC1 locus for schizophrenia and bipolar affective disorder in South Indian population. *J. Hum. Genet.* **57**, 523–530 (2012).
256. Mata, I. *et al.* Additive effect of NRG1 and DISC1 genes on lateral ventricle enlargement in first episode schizophrenia. *Neuroimage* **53**, 1016–1022 (2010).
257. Nakata, K. *et al.* DISC1 splice variants are upregulated in schizophrenia and associated with risk polymorphisms. *Proc. Natl. Acad. Sci. U.S.A.* **106**, 15873–15878 (2009).

258. Rastogi, A., Zai, C., Likhodi, O., Kennedy, J. L. & Wong, A. H. Genetic association and post-mortem brain mRNA analysis of DISC1 and related genes in schizophrenia. *Schizophr. Res.* **114**, 39–49 (2009).
259. Schumacher, J. *et al.* The DISC locus and schizophrenia: evidence from an association study in a central European sample and from a meta-analysis across different European populations. *Hum. Mol. Genet.* **18**, 2719–2727 (2009).
260. Saetre, P. *et al.* Association between a disrupted-in-schizophrenia 1 (DISC1) single nucleotide polymorphism and schizophrenia in a combined Scandinavian case-control sample. *Schizophr. Res.* **106**, 237–241 (2008).
261. Kim, H.-J. *et al.* Association study of polymorphisms between DISC1 and schizophrenia in a Korean population. *Neurosci. Lett.* **430**, 60–63 (2008).
262. Qu, M. *et al.* Positive association of the Disrupted-in-Schizophrenia-1 gene (DISC1) with schizophrenia in the Chinese Han population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **144B**, 266–270 (2007).
263. DeRosse, P. *et al.* Disrupted in schizophrenia 1 genotype and positive symptoms in schizophrenia. *Biol. Psychiatry* **61**, 1208–1210 (2007).
264. Lipska, B. K. *et al.* Expression of DISC1 binding partners is reduced in schizophrenia and associated with DISC1 SNPs. *Hum. Mol. Genet.* **15**, 1245–1258 (2006).
265. Zhang, F. *et al.* Genetic association between schizophrenia and the DISC1 gene in the Scottish population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141B**, 155–159 (2006).
266. Kockelkorn, T. T. J. P. *et al.* Association study of polymorphisms in the 5' upstream region of human DISC1 gene with schizophrenia. *Neurosci. Lett.* **368**, 41–45 (2004).

267. Hennah, W. *et al.* Haplotype transmission analysis provides evidence of association for DISC1 to schizophrenia and suggests sex-dependent effects. *Hum. Mol. Genet.* **12**, 3151–3159 (2003).
268. Trost, S. *et al.* DISC1 (disrupted-in-schizophrenia 1) is associated with cortical grey matter volumes in the human brain: a voxel-based morphometry (VBM) study. *J Psychiatr Res* **47**, 188–196 (2013).
269. Brauns, S. *et al.* DISC1 is associated with cortical thickness and neural efficiency. *Neuroimage* **57**, 1591–1600 (2011).
270. Di Giorgio, A. *et al.* Association of the SerCys DISC1 polymorphism with human hippocampal formation gray matter and function during memory encoding. *Eur. J. Neurosci.* **28**, 2129–2136 (2008).
271. Szeszko, P. R. *et al.* DISC1 is associated with prefrontal cortical gray matter and positive symptoms in schizophrenia. *Biol Psychol* **79**, 103–110 (2008).
272. Cannon, T. D. *et al.* Association of DISC1/TRAX haplotypes with schizophrenia, reduced prefrontal gray matter, and impaired short- and long-term memory. *Arch. Gen. Psychiatry* **62**, 1205–1213 (2005).
273. Callicott, J. H. *et al.* Variation in DISC1 affects hippocampal structure and function and increases risk for schizophrenia. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 8627–8632 (2005).
274. Proitsi, P. *et al.* Positional pathway screen of wnt signaling genes in schizophrenia: association with DKK4. *Biol. Psychiatry* **63**, 13–16 (2008).
275. Zhang, L.-H. *et al.* [Association of the schizophrenia susceptible gene DKK4 with brain volume in Chinese populations]. *Zool. Res.* **32**, 62–65 (2011).

276. Kristiansen, L. V., Beneyto, M., Haroutunian, V. & Meador-Woodruff, J. H. Changes in NMDA receptor subunits and interacting PSD proteins in dorsolateral prefrontal and anterior cingulate cortex indicate abnormal regional expression in schizophrenia. *Mol. Psychiatry* **11**, 737–747, 705 (2006).
277. Ingason, A. *et al.* Expression analysis in a rat psychosis model identifies novel candidate genes validated in a large case-control sample of schizophrenia. *Transl Psychiatry* **5**, e656 (2015).
278. Hibar, D. P. *et al.* Common genetic variants influence human subcortical brain structures. *Nature* **520**, 224–229 (2015).
279. Nkam, I. *et al.* Impact of DRD2/ANKK1 and COMT Polymorphisms on Attention and Cognitive Functions in Schizophrenia. *PLoS ONE* **12**, e0170147 (2017).
280. González-Castro, T. B. *et al.* The role of C957T, TaqI and Ser311Cys polymorphisms of the DRD2 gene in schizophrenia: systematic review and meta-analysis. *Behav Brain Funct* **12**, 29 (2016).
281. Vink, M. *et al.* DRD2 Schizophrenia-Risk Allele Is Associated With Impaired Striatal Functioning in Unaffected Siblings of Schizophrenia Patients. *Schizophr Bull* **42**, 843–850 (2016).
282. Cohen, O. S. *et al.* A splicing-regulatory polymorphism in DRD2 disrupts ZRANB2 binding, impairs cognitive functioning and increases risk for schizophrenia in six Han Chinese samples. *Mol. Psychiatry* **21**, 975–982 (2016).
283. Wang, Y. *et al.* The -141C Ins/Del and Taq1A polymorphism in the dopamine D2 receptor gene may confer susceptibility to schizophrenia in Asian populations. *J Clin Neurosci* **30**, 1–7 (2016).

284. Mahalaxmi, I., Sasikala, K., Arun, M. & Balachandar, V. Cytogenetics And Dopamine Receptor (Drd2) Gene Polymorphism In Schizophrenia Patients. *Value Health* **18**, A344 (2015).
285. Yao, J., Pan, Y., Ding, M., Pang, H. & Wang, B. Association between DRD2 (rs1799732 and rs1801028) and ANKK1 (rs1800497) polymorphisms and schizophrenia: a meta-analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **168B**, 1–13 (2015).
286. Liu, L. *et al.* The mRNA expression of DRD2, PI3KCB, and AKT1 in the blood of acute schizophrenia patients. *Psychiatry Res* **243**, 397–402 (2016).
287. Xiao, L. *et al.* Functional -141C Ins/Del polymorphism in the dopamine D2 receptor gene promoter and schizophrenia in a Chinese Han population. *J. Int. Med. Res.* **41**, 1171–1178 (2013).
288. Chien, Y.-L. *et al.* DRD2 haplotype associated with negative symptoms and sustained attention deficits in Han Chinese with schizophrenia in Taiwan. *J. Hum. Genet.* **58**, 229–232 (2013).
289. Fan, H. *et al.* An association study of DRD2 gene polymorphisms with schizophrenia in a Chinese Han population. *Neurosci. Lett.* **477**, 53–56 (2010).
290. Cordeiro, Q., Siqueira-Roberto, J., Zung, S. & Vallada, H. Association between the DRD2-141C Insertion/Deletion polymorphism and schizophrenia. *Arq Neuropsiquiatr* **67**, 191–194 (2009).
291. Betcheva, E. T. *et al.* Case-control association study of 59 candidate genes reveals the DRD2 SNP rs6277 (C957T) as the only susceptibility factor for schizophrenia in the Bulgarian population. *J. Hum. Genet.* **54**, 98–107 (2009).

292. Lafuente, A. *et al.* -141C Ins/Del polymorphism of the dopamine D2 receptor gene is associated with schizophrenia in a Spanish population. *Psychiatr. Genet.* **18**, 122–127 (2008).
293. Glatt, S. J. *et al.* Family-based association testing strongly implicates DRD2 as a risk gene for schizophrenia in Han Chinese from Taiwan. *Mol. Psychiatry* **14**, 885–893 (2009).
294. Parsons, M. J. *et al.* A dopamine D2 receptor gene-related polymorphism is associated with schizophrenia in a Spanish population isolate. *Psychiatr. Genet.* **17**, 159–163 (2007).
295. Hoenicka, J. *et al.* C957T DRD2 polymorphism is associated with schizophrenia in Spanish patients. *Acta Psychiatr Scand* **114**, 435–438 (2006).
296. Hänninen, K. *et al.* Association between the C957T polymorphism of the dopamine D2 receptor gene and schizophrenia. *Neurosci. Lett.* **407**, 195–198 (2006).
297. Kukreti, R. *et al.* Association of DRD2 gene variant with schizophrenia. *Neurosci. Lett.* **392**, 68–71 (2006).
298. Lawford, B. R. *et al.* The C/C genotype of the C957T polymorphism of the dopamine D2 receptor is associated with schizophrenia. *Schizophr. Res.* **73**, 31–37 (2005).
299. Dubertret, C. *et al.* The 3' region of the DRD2 gene is involved in genetic susceptibility to schizophrenia. *Schizophr. Res.* **67**, 75–85 (2004).
300. Jönsson, E. G. *et al.* Dopamine D2 receptor gene Ser311Cys variant and schizophrenia: association study and meta-analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **119B**, 28–34 (2003).
301. Chong, S.-A., Tan, E.-C., Tan, C. H., Mythily, null & Chan, Y. H. Polymorphisms of dopamine receptors and tardive dyskinesia among Chinese patients with schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **116B**, 51–54 (2003).

302. Schindler, K. M. *et al.* Association and linkage disequilibrium between a functional polymorphism of the dopamine-2 receptor gene and schizophrenia in a genetically homogeneous Portuguese population. *Mol. Psychiatry* **7**, 1002–1005 (2002).
303. Dubertret, C., Gorwood, P., Gouya, L., Deybach, J. C. & Adès, J. Association and excess of transmission of a DRD2 haplotype in a sample of French schizophrenic patients. *Schizophr. Res.* **49**, 203–212 (2001).
304. Hori, H., Ohmori, O., Shinkai, T., Kojima, H. & Nakamura, J. Association analysis between two functional dopamine D2 receptor gene polymorphisms and schizophrenia. *Am. J. Med. Genet.* **105**, 176–178 (2001).
305. Inada, T., Arinami, T. & Yagi, G. Association between a polymorphism in the promoter region of the dopamine D2 receptor gene and schizophrenia in Japanese subjects: replication and evaluation for antipsychotic-related features. *Int. J. Neuropsychopharmacol.* **2**, 181–186 (1999).
306. Breen, G. *et al.* -141 C del/ins polymorphism of the dopamine receptor 2 gene is associated with schizophrenia in a British population. *Am. J. Med. Genet.* **88**, 407–410 (1999).
307. Li, T. *et al.* Case-control, haplotype relative risk and transmission disequilibrium analysis of a dopamine D2 receptor functional promoter polymorphism in schizophrenia. *Schizophr. Res.* **32**, 87–92 (1998).
308. Kaneshima, M., Higa, T., Nakamoto, H. & Nagamine, M. An association study between the Cys311 variant of dopamine D2 receptor gene and schizophrenia in the Okinawan population. *Psychiatry Clin. Neurosci.* **51**, 379–381 (1997).

309. Arinami, T., Gao, M., Hamaguchi, H. & Toru, M. A functional polymorphism in the promoter region of the dopamine D2 receptor gene is associated with schizophrenia. *Hum. Mol. Genet.* **6**, 577–582 (1997).
310. Harano, M. Ser-311-Cys polymorphism of the dopamine D2 receptor gene and schizophrenia--an analysis of schizophrenic patients in Fukuoka. *Kurume Med J* **44**, 201–208 (1997).
311. Smith, C. T. *et al.* The impact of common dopamine D2 receptor gene polymorphisms on D2/3 receptor availability: C957T as a key determinant in putamen and ventral striatum. *Transl Psychiatry* **7**, e1091 (2017).
312. Lochman, J., Balcar, V. J., Šťastný, F. & Serý, O. Preliminary evidence for association between schizophrenia and polymorphisms in the regulatory Regions of the ADRA2A, DRD3 and SNAP-25 Genes. *Psychiatry Res* **205**, 7–12 (2013).
313. Urraca, N. *et al.* Association study of DRD3 gene in schizophrenia in Mexican sib-pairs. *Psychiatry Res* **190**, 367–368 (2011).
314. Tee, S., Tang, P. & Loh, H. Genetic Association Analysis of Dopamine DRD3 Ser9Gly Polymorphism and Schizophrenia in Malay Population. *Iran. J. Public Health* **40**, 6–10 (2011).
315. Nunokawa, A. *et al.* The dopamine D3 receptor (DRD3) gene and risk of schizophrenia: case-control studies and an updated meta-analysis. *Schizophr. Res.* **116**, 61–67 (2010).
316. Takahashi, T. *et al.* The association of genotypic combination of the DRD3 and BDNF polymorphisms on the adhesio interthalamica and medial temporal lobe structures. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **32**, 1236–1242 (2008).

317. Utsunomiya, K. *et al.* Genetic association between the dopamine D3 gene polymorphism (Ser9Gly) and schizophrenia in Japanese populations: evidence from a case-control study and meta-analysis. *Neurosci. Lett.* **444**, 161–165 (2008).
318. Lafuente, A. *et al.* -141C Ins/Del polymorphism of the dopamine D2 receptor gene is associated with schizophrenia in a Spanish population. *Psychiatr. Genet.* **18**, 122–127 (2008).
319. Ma, G. *et al.* The Ser9Gly polymorphism of the dopamine D3 receptor gene and risk of schizophrenia: an association study and a large meta-analysis. *Schizophr. Res.* **101**, 26–35 (2008).
320. Talkowski, M. E. *et al.* Novel, replicated associations between dopamine D3 receptor gene polymorphisms and schizophrenia in two independent samples. *Biol. Psychiatry* **60**, 570–577 (2006).
321. Aksenova, M. G. *et al.* [D3 dopamine receptor gene Ser9Gly polymorphism in Russian patients with schizophrenia]. *Zh Nevrol Psikhiatr Im S S Korsakova* **104**, 57–61 (2004).
322. Staddon, S. *et al.* Association between dopamine D3 receptor gene polymorphisms and schizophrenia in an isolate population. *Schizophr. Res.* **73**, 49–54 (2005).
323. Nimgaonkar, V. L. *et al.* Association study of schizophrenia and the dopamine D3 receptor gene locus in two independent samples. *Am. J. Med. Genet.* **67**, 505–514 (1996).
324. Tanaka, T. *et al.* Association study between schizophrenia and dopamine D3 receptor gene polymorphism. *Am. J. Med. Genet.* **67**, 366–368 (1996).
325. Shaikh, S. *et al.* Allelic association between a Ser-9-Gly polymorphism in the dopamine D3 receptor gene and schizophrenia. *Hum. Genet.* **97**, 714–719 (1996).

326. Asherson, P. *et al.* Linkage, association and mutational analysis of the dopamine D3 receptor gene in schizophrenia. *Mol. Psychiatry* **1**, 125–132 (1996).
327. Griffon, N. *et al.* Dopamine D3 receptor gene: organization, transcript variants, and polymorphism associated with schizophrenia. *Am. J. Med. Genet.* **67**, 63–70 (1996).
328. Kennedy, J. L. *et al.* Association study of dopamine D3 receptor gene and schizophrenia. *Am. J. Med. Genet.* **60**, 558–562 (1995).
329. Morell, R. T. Association between schizophrenia and homozygosity at the dopamine D3 receptor gene. *J. Med. Genet.* **30**, 708–709 (1993).
330. Crocq, M. A. *et al.* Association between schizophrenia and homozygosity at the dopamine D3 receptor gene. *J. Med. Genet.* **29**, 858–860 (1992).
331. Strohmaier, J. *et al.* A reappraisal of the association between Dysbindin (DTNBP1) and schizophrenia in a large combined case-control and family-based sample of German ancestry. *Schizophr. Res.* **118**, 98–105 (2010).
332. Sun, Y., Shen, Y. & Xu, Q. DTNBP1 gene is associated with some symptom factors of schizophrenia in Chinese Han nationality. *Chin. Med. Sci. J.* **25**, 85–89 (2010).
333. Vilella, E. *et al.* Association of schizophrenia with DTNBP1 but not with DAO, DAOA, NRG1 and RGS4 nor their genetic interaction. *J Psychiatr Res* **42**, 278–288 (2008).
334. Wessman, J. *et al.* Mixture model clustering of phenotype features reveals evidence for association of DTNBP1 to a specific subtype of schizophrenia. *Biol. Psychiatry* **66**, 990–996 (2009).

335. Thirunavukkarasu, P. *et al.* An exploratory association study of the influence of dysbindin and neuregulin polymorphisms on brain morphometry in patients with schizophrenia and healthy subjects from South India. *Asian J Psychiatr* **10**, 62–68 (2014).
336. Trost, S. *et al.* The DTNBP1 (dysbindin-1) gene variant rs2619522 is associated with variation of hippocampal and prefrontal grey matter volumes in humans. *Eur Arch Psychiatry Clin Neurosci* **263**, 53–63 (2013).
337. Zhang, B. *et al.* Common variants in SLC1A2 and schizophrenia: Association and cognitive function in patients with schizophrenia and healthy individuals. *Schizophr. Res.* **169**, 128–134 (2015).
338. Spangaro, M. *et al.* Cognitive dysfunction and glutamate reuptake: effect of EAAT2 polymorphism in schizophrenia. *Neurosci. Lett.* **522**, 151–155 (2012).
339. Deng, X. *et al.* Association study of polymorphisms in the excitatory amino acid transporter 2 gene (SLC1A2) with schizophrenia. *BMC Psychiatry* **4**, 21 (2004).
340. Poletti, S. *et al.* Effect of glutamate transporter EAAT2 gene variants and gray matter deficits on working memory in schizophrenia. *Eur. Psychiatry* **29**, 219–225 (2014).
341. Rao, W. *et al.* Association between forkhead-box P2 gene polymorphism and clinical symptoms in chronic schizophrenia in a Chinese population. *J Neural Transm (Vienna)* **124**, 891–897 (2017).
342. Tolosa, A. *et al.* FOXP2 gene and language impairment in schizophrenia: association and epigenetic studies. *BMC Med. Genet.* **11**, 114 (2010).
343. Sanjuan, J. *et al.* FOXP2 polymorphisms in patients with schizophrenia. *Schizophr. Res.* **73**, 253–256 (2005).

344. Španiel, F. *et al.* Genetic variation in FOXP2 alters grey matter concentrations in schizophrenia patients. *Neurosci. Lett.* **493**, 131–135 (2011).
345. Hoogman, M. *et al.* Assessing the effects of common variation in the FOXP2 gene on human brain structure. *Front Hum Neurosci* **8**, 473 (2014).
346. Fatemi, S. H., Folsom, T. D., Rooney, R. J. & Thuras, P. D. Expression of GABAA $\alpha 2$ -, $\beta 1$ - and ϵ -receptors are altered significantly in the lateral cerebellum of subjects with schizophrenia, major depression and bipolar disorder. *Transl Psychiatry* **3**, e303–e303 (2013).
347. Zhu, B. *et al.* The GABRB1 gene is associated with thalamus volume and modulates the association between thalamus volume and intelligence. *Neuroimage* **102 Pt 2**, 756–763 (2014).
348. Tao, R. *et al.* GAD1 alternative transcripts and DNA methylation in human prefrontal cortex and hippocampus in brain development, schizophrenia. *Mol. Psychiatry* (2017).
doi:10.1038/mp.2017.105
349. Du, J. *et al.* Comprehensive analysis of polymorphisms throughout GAD1 gene: a family-based association study in schizophrenia. *J Neural Transm (Vienna)* **115**, 513–519 (2008).
350. Straub, R. E. *et al.* Allelic variation in GAD1 (GAD67) is associated with schizophrenia and influences cortical function and gene expression. *Mol. Psychiatry* **12**, 854–869 (2007).
351. Huang, H.-S. & Akbarian, S. GAD1 mRNA expression and DNA methylation in prefrontal cortex of subjects with schizophrenia. *PLoS ONE* **2**, e809 (2007).
352. Mitchell, A. C., Jiang, Y., Peter, C. & Akbarian, S. Transcriptional regulation of GAD1 GABA synthesis gene in the prefrontal cortex of subjects with schizophrenia. *Schizophr. Res.* **167**, 28–34 (2015).

353. Bharadwaj, R. *et al.* Conserved chromosome 2q31 conformations are associated with transcriptional regulation of GAD1 GABA synthesis enzyme and altered in prefrontal cortex of subjects with schizophrenia. *J. Neurosci.* **33**, 11839–11851 (2013).
354. Brauns, S. *et al.* Genetic variation in GAD1 is associated with cortical thickness in the parahippocampal gyrus. *J Psychiatr Res* **47**, 872–879 (2013).
355. Addington, A. M. *et al.* GAD1 (2q31.1), which encodes glutamic acid decarboxylase (GAD67), is associated with childhood-onset schizophrenia and cortical gray matter volume loss. *Mol. Psychiatry* **10**, 581–588 (2005).
356. Vawter, M. P. *et al.* Microarray screening of lymphocyte gene expression differences in a multiplex schizophrenia pedigree. *Schizophr. Res.* **67**, 41–52 (2004).
357. Chavarría-Siles, I. *et al.* Genes encoding heterotrimeric G-proteins are associated with gray matter volume variations in the medial frontal cortex. *Cereb. Cortex* **23**, 1025–1030 (2013).
358. Kunugi, H. *et al.* Association study of C825T polymorphism of the G-protein b3 subunit gene with schizophrenia and mood disorders. *J Neural Transm (Vienna)* **109**, 213–218 (2002).
359. Treutlein, J. *et al.* Dissection of phenotype reveals possible association between schizophrenia and Glutamate Receptor Delta 1 (GRID1) gene promoter. *Schizophrenia Research* **111**, 123–130 (2009).
360. Guo, S.-Z. *et al.* A case-control association study between the GRID1 gene and schizophrenia in the Chinese Northern Han population. *Schizophr. Res.* **93**, 385–390 (2007).
361. Begni, S. *et al.* Association between the G1001C polymorphism in the GRIN1 gene promoter region and schizophrenia. *Biol. Psychiatry* **53**, 617–619 (2003).

362. Sakurai, K., Toru, M., Yamakawa-Kobayashi, K. & Arinami, T. Mutation analysis of the N-methyl-D-aspartate receptor NR1 subunit gene (GRIN1) in schizophrenia. *Neurosci. Lett.* **296**, 168–170 (2000).
363. Nenadic, I. *et al.* Glutamate receptor delta 1 (GRID1) genetic variation and brain structure in schizophrenia. *Journal of Psychiatric Research* **46**, 1531–1539 (2012).
364. Liu, R. *et al.* Correlation of functional GRIN2A gene promoter polymorphisms with schizophrenia and serum D-serine levels. *Gene* **568**, 25–30 (2015).
365. Tang, J. *et al.* Significant linkage and association between a functional (GT)_n polymorphism in promoter of the N-methyl-D-aspartate receptor subunit gene (GRIN2A) and schizophrenia. *Neurosci. Lett.* **409**, 80–82 (2006).
366. Iwayama-Shigeno, Y. *et al.* Extended analyses support the association of a functional (GT)_n polymorphism in the GRIN2A promoter with Japanese schizophrenia. *Neurosci. Lett.* **378**, 102–105 (2005).
367. Itokawa, M. *et al.* A microsatellite repeat in the promoter of the N-methyl-D-aspartate receptor 2A subunit (GRIN2A) gene suppresses transcriptional activity and correlates with chronic outcome in schizophrenia. *Pharmacogenetics* **13**, 271–278 (2003).
368. Inoue, H. *et al.* Functional (GT)_n polymorphisms in promoter region of N-methyl-d-aspartate receptor 2A subunit (GRIN2A) gene affect hippocampal and amygdala volumes. *Genes Brain Behav.* **9**, 269–275 (2010).
369. Fatima, A. *et al.* Genome-Wide Supported Risk Variants in MIR137, CACNA1C, CSMD1, DRD2, and GRM3 Contribute to Schizophrenia Susceptibility in Pakistani Population. *Psychiatry Investig* **14**, 687–692 (2017).

370. Saini, S. M. *et al.* Meta-analysis supports GWAS-implicated link between GRM3 and schizophrenia risk. *Transl Psychiatry* **7**, e1196 (2017).
371. García-Bea, A., Bermudez, I., Harrison, P. J. & Lane, T. A. A group II metabotropic glutamate receptor 3 (mGlu3, GRM3) isoform implicated in schizophrenia interacts with canonical mGlu3 and reduces ligand binding. *J. Psychopharmacol. (Oxford)* **31**, 1519–1526 (2017).
372. Yang, X., Wang, G., Wang, Y. & Yue, X. Association of metabotropic glutamate receptor 3 gene polymorphisms with schizophrenia risk: evidence from a meta-analysis. *Neuropsychiatr Dis Treat* **11**, 823–833 (2015).
373. Bishop, J. R. *et al.* Pharmacogenetic associations of the type-3 metabotropic glutamate receptor (GRM3) gene with working memory and clinical symptom response to antipsychotics in first-episode schizophrenia. *Psychopharmacology (Berl.)* **232**, 145–154 (2015).
374. Mössner, R. *et al.* Further evidence for a functional role of the glutamate receptor gene GRM3 in schizophrenia. *Eur Neuropsychopharmacol* **18**, 768–772 (2008).
375. Schwab, S. G. *et al.* DNA sequence variants in the metabotropic glutamate receptor 3 and risk to schizophrenia: an association study. *Psychiatr. Genet.* **18**, 25–30 (2008).
376. Albalushi, T. *et al.* Replication study and meta-analysis of the genetic association of GRM3 gene polymorphisms with schizophrenia in a large Japanese case-control population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **147**, 392–396 (2008).
377. Chen, Q. *et al.* A case-control study of the relationship between the metabotropic glutamate receptor 3 gene and schizophrenia in the Chinese population. *Schizophr. Res.* **73**, 21–26 (2005).

378. Fujii, Y. *et al.* Positive associations of polymorphisms in the metabotropic glutamate receptor type 3 gene (GRM3) with schizophrenia. *Psychiatr. Genet.* **13**, 71–76 (2003).
379. Mounce, J. *et al.* Association of GRM3 polymorphism with white matter integrity in schizophrenia. *Schizophr. Res.* **155**, 8–14 (2014).
380. Devon, R. S. *et al.* The genomic organisation of the metabotropic glutamate receptor subtype 5 gene, and its association with schizophrenia. *Mol. Psychiatry* **6**, 311–314 (2001).
381. Matosin, N. *et al.* Effects of common GRM5 genetic variants on cognition, hippocampal volume and mGluR5 protein levels in schizophrenia. *Brain Imaging Behav* **12**, 509–517 (2018).
382. Hu, G. *et al.* The interaction of NOS1AP, DISC1, DAOA, and GSK3B confers susceptibility of early-onset schizophrenia in Chinese Han population. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **81**, 187–193 (2018).
383. Li, M. *et al.* Genetic association and identification of a functional SNP at GSK3 β for schizophrenia susceptibility. *Schizophr. Res.* **133**, 165–171 (2011).
384. Scassellati, C. *et al.* Association study of -1727 A/T, -50 C/T and (CAA)_n repeat GSK-3 β gene polymorphisms with schizophrenia. *Neuropsychobiology* **50**, 16–20 (2004).
385. Emamian, E. S., Hall, D., Birnbaum, M. J., Karayiorgou, M. & Gogos, J. A. Convergent evidence for impaired AKT1-GSK3 β signaling in schizophrenia. *Nat. Genet.* **36**, 131–137 (2004).
386. Kozlovsky, N. *et al.* Reduced GSK-3 β mRNA levels in postmortem dorsolateral prefrontal cortex of schizophrenic patients. *J Neural Transm (Vienna)* **111**, 1583–1592 (2004).

387. Blasi, G. *et al.* Association of GSK-3 β genetic variation with GSK-3 β expression, prefrontal cortical thickness, prefrontal physiology, and schizophrenia. *Am J Psychiatry* **170**, 868–876 (2013).
388. Benedetti, F. *et al.* Temporal lobe grey matter volume in schizophrenia is associated with a genetic polymorphism influencing glycogen synthase kinase 3- β activity. *Genes Brain Behav.* **9**, 365–371 (2010).
389. Metzger, W. S., Newton, J. E. & Paige, S. R. HLA-A1 and schizophrenia. *Biol. Psychiatry* **24**, 364–365 (1988).
390. Brucato, N., Guadalupe, T., Franke, B., Fisher, S. E. & Francks, C. A schizophrenia-associated HLA locus affects thalamus volume and asymmetry. *Brain Behav. Immun.* **46**, 311–318 (2015).
391. Shivakumar, V. *et al.* Influence of correlation between HLA-G polymorphism and Interleukin-6 (IL6) gene expression on the risk of schizophrenia. *Cytokine* (2017).
doi:10.1016/j.cyto.2017.11.016
392. Rajasekaran, A. *et al.* The impact of HLA-G 3' UTR variants and sHLA-G on risk and clinical correlates of schizophrenia. *Hum. Immunol.* **77**, 1166–1171 (2016).
393. Rajasekaran, A. *et al.* Association of HLA-G 14bp INS/DEL Polymorphism with brain morphology in Schizophrenia. *Molecular Cytogenetics* **7**, P43 (2014).
394. Sun, Q. *et al.* The promoter polymorphisms in HTR2A gene associated with schizophrenia in Chinese of Han ethnicity. *Psychiatry Res* **262**, 636–637 (2018).
395. Yildiz, S. H. *et al.* Association of schizophrenia with T102C (rs6313) and 1438 A/G (rs6311) polymorphisms of HTR2A gene. *Acta Neuropsychiatr* **25**, 342–348 (2013).

396. Ni, J. *et al.* T102C polymorphism of serotonin 2A type receptor gene confers susceptibility to (early onset) schizophrenia in Han Chinese: an association study and meta-analysis. *Asia Pac Psychiatry* **5**, 24–30 (2013).
397. Gu, L. *et al.* HTR2A-1438A/G polymorphism influences the risk of schizophrenia but not bipolar disorder or major depressive disorder: a meta-analysis. *J. Neurosci. Res.* **91**, 623–633 (2013).
398. Kim, Y.-K. & Yoon, H.-K. Effect of serotonin-related gene polymorphisms on pathogenesis and treatment response in Korean schizophrenic patients. *Behav. Genet.* **41**, 709–715 (2011).
399. Tee, S. F., Chow, T. J., Tang, P. Y. & Loh, H. C. Linkage of schizophrenia with TPH2 and 5-HTR2A gene polymorphisms in the Malay population. *Genet. Mol. Res.* **9**, 1274–1278 (2010).
400. Golimbet, V. E. *et al.* [Serotonin transporter gene polymorphism in families with schizophrenia]. *Zh Nevrol Psikhiatr Im S S Korsakova* **101**, 40–41 (2001).
401. Abdolmaleky, H. M., Faraone, S. V., Glatt, S. J. & Tsuang, M. T. Meta-analysis of association between the T102C polymorphism of the 5HT2a receptor gene and schizophrenia. *Schizophr. Res.* **67**, 53–62 (2004).
402. Mitiushina, N. G., Abramova, L. I., Kaleda, V. G. & Golimbet, V. E. [A-1438-G serotonin receptor type 2A (5-HTR2A) gene polymorphism in patients in patients with schizophrenia]. *Zh Nevrol Psikhiatr Im S S Korsakova* **103**, 43–46 (2003).
403. Inayama, Y. *et al.* Positive association between a DNA sequence variant in the serotonin 2A receptor gene and schizophrenia. *Am. J. Med. Genet.* **67**, 103–105 (1996).

404. Ikeda, M. *et al.* Positive association of the serotonin 5-HT₇ receptor gene with schizophrenia in a Japanese population. *Neuropsychopharmacology* **31**, 866–871 (2006).
405. Sprooten, E. *et al.* Common genetic variants and gene expression associated with white matter microstructure in the human brain. *Neuroimage* **97**, 252–261 (2014).
406. Li, J. *et al.* Genetic variations in the serotonergic system contribute to amygdala volume in humans. *Front Neuroanat* **9**, 129 (2015).
407. Zanardini, R. *et al.* Association between IL-1 β -511C/T and IL-1RA (86bp)_n repeats polymorphisms and schizophrenia. *J Psychiatr Res* **37**, 457–462 (2003).
408. Kapelski, P. *et al.* Association study of functional polymorphisms in interleukins and interleukin receptors genes: IL1A, IL1B, IL1RN, IL6, IL6R, IL10, IL10RA and TGFB1 in schizophrenia in Polish population. *Schizophr. Res.* **169**, 1–9 (2015).
409. Shibuya, M. *et al.* Interleukin 1 β gene and risk of schizophrenia: detailed case-control and family-based studies and an updated meta-analysis. *Hum Psychopharmacol* **29**, 31–37 (2014).
410. Yoshida, M. *et al.* Haplotypes in the expression quantitative trait locus of interleukin-1 β gene are associated with schizophrenia. *Schizophr. Res.* **140**, 185–191 (2012).
411. Shirts, B. H., Wood, J., Yolken, R. H. & Nimgaonkar, V. L. Association study of IL10, IL1 β , and IL1RN and schizophrenia using tag SNPs from a comprehensive database: suggestive association with rs16944 at IL1 β . *Schizophr. Res.* **88**, 235–244 (2006).
412. Meisenzahl, E. M. *et al.* Association of an interleukin-1 β genetic polymorphism with altered brain structure in patients with schizophrenia. *Am J Psychiatry* **158**, 1316–1319 (2001).

413. Zanardini, R. *et al.* Association between IL-1beta -511C/T and IL-1RA (86bp)n repeats polymorphisms and schizophrenia. *J Psychiatr Res* **37**, 457–462 (2003).
414. Ben Nejma, M. *et al.* A gender-specific association of interleukin 1 receptor antagonist polymorphism with schizophrenia susceptibility. *Acta Neuropsychiatr* **25**, 349–355 (2013).
415. Papiol, S. *et al.* Ventricular enlargement in schizophrenia is associated with a genetic polymorphism at the interleukin-1 receptor antagonist gene. *Neuroimage* **27**, 1002–1006 (2005).
416. Edwards, T. L. *et al.* Interaction between interleukin 3 and dystrobrevin-binding protein 1 in schizophrenia. *Schizophr. Res.* **106**, 208–217 (2008).
417. Chen, X. *et al.* Interleukin 3 and schizophrenia: the impact of sex and family history. *Mol. Psychiatry* **12**, 273–282 (2007).
418. Luo, X. *et al.* The interleukin 3 gene (IL3) contributes to human brain volume variation by regulating proliferation and survival of neural progenitors. *PLoS ONE* **7**, e50375 (2012).
419. Zakharyan, R. *et al.* Interleukin-6 promoter polymorphism and plasma levels in patients with schizophrenia. *Tissue Antigens* **80**, 136–142 (2012).
420. Baune, B. T. *et al.* Interleukin-6 gene (IL-6): a possible role in brain morphology in the healthy adult brain. *J Neuroinflammation* **9**, 125 (2012).
421. Voineskos, A. N. *et al.* A family-based association study of the myelin-associated glycoprotein and 2',3'-cyclic nucleotide 3'-phosphodiesterase genes with schizophrenia. *Psychiatr. Genet.* **18**, 143–146 (2008).
422. Wan, C. *et al.* Polymorphisms of myelin-associated glycoprotein gene are associated with schizophrenia in the Chinese Han population. *Neurosci. Lett.* **388**, 126–131 (2005).

423. Yang, Y. F. *et al.* Possible association of the MAG locus with schizophrenia in a Chinese Han cohort of family trios. *Schizophr. Res.* **75**, 11–19 (2005).
424. Felsky, D. *et al.* Myelin-associated glycoprotein gene and brain morphometry in schizophrenia. *Front Psychiatry* **3**, 40 (2012).
425. Camarena, B. *et al.* Monoamine oxidase a and B gene polymorphisms and negative and positive symptoms in schizophrenia. *ISRN Psychiatry* **2012**, 852949 (2012).
426. Chen, Y., Zhang, J., Zhang, L., Shen, Y. & Xu, Q. Effects of MAOA promoter methylation on susceptibility to paranoid schizophrenia. *Hum. Genet.* **131**, 1081–1087 (2012).
427. Sun, Y. *et al.* Study of a possible role of the monoamine oxidase A (MAOA) gene in paranoid schizophrenia among a Chinese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 104–111 (2012).
428. Wei, Y.-L., Li, C.-X., Li, S.-B., Liu, Y. & Hu, L. Association study of monoamine oxidase A/B genes and schizophrenia in Han Chinese. *Behav Brain Funct* **7**, 42 (2011).
429. Qiu, H. T. *et al.* Association between monoamine oxidase (MAO)-A gene variants and schizophrenia in a Chinese population. *Brain Res.* **1287**, 67–73 (2009).
430. Shi, Y. *et al.* [Association study of the polymorphisms of monoamine oxidase A genes with schizophrenia]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **24**, 457–459 (2007).
431. Jönsson, E. G. *et al.* Association between a promoter variant in the monoamine oxidase A gene and schizophrenia. *Schizophr. Res.* **61**, 31–37 (2003).
432. Cerasa, A. *et al.* MAO A VNTR polymorphism and amygdala volume in healthy subjects. *Psychiatry Res* **191**, 87–91 (2011).

433. The Schizophrenia Psychiatric Genome-Wide Association Study (GWAS) Consortium. Genome-wide association study identifies five new schizophrenia loci. *Nat Genet* **43**, 969–976 (2011).
434. Sudesh, R. *et al.* Minor allele C of rs12807809 polymorphism in NRGN contributes to the severity of psychosis in patients with Schizophrenia in South Indian population. *Neurosci. Lett.* **649**, 107–111 (2017).
435. Sakamoto, K. & Crowley, J. J. A comprehensive review of the genetic and biological evidence supports a role for MicroRNA-137 in the etiology of schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **177**, 242–256 (2018).
436. Lu, W. *et al.* Genetic association analysis of microRNA137 and its target complex 1 with schizophrenia in Han Chinese. *Sci Rep* **7**, 15084 (2017).
437. Sun, Y.-J. *et al.* Association between single nucleotide polymorphisms in MiR219-1 and MiR137 and susceptibility to schizophrenia in a Chinese population. *FEBS Open Bio* **5**, 774–778 (2015).
438. Cosgrove, D. *et al.* Effects of MiR-137 genetic risk score on brain volume and cortical measures in patients with schizophrenia and controls. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **177**, 369–376 (2018).
439. Kuswanto, C. N. *et al.* The impact of genome wide supported microRNA-137 (MIR137) risk variants on frontal and striatal white matter integrity, neurocognitive functioning, and negative symptoms in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **168B**, 317–326 (2015).

440. Liu, B. *et al.* The impact of MIR137 on dorsolateral prefrontal-hippocampal functional connectivity in healthy subjects. *Neuropsychopharmacology* **39**, 2153–2160 (2014).
441. Rai, V., Yadav, U., Kumar, P., Yadav, S. K. & Gupta, S. Methylenetetrahydrofolate reductase A1298C genetic variant & risk of schizophrenia: A meta-analysis. *Indian J. Med. Res.* **145**, 437–447 (2017).
442. Fryar-Williams, S. Fundamental Role of Methylenetetrahydrofolate Reductase 677 C → T Genotype and Flavin Compounds in Biochemical Phenotypes for Schizophrenia and Schizoaffective Psychosis. *Front Psychiatry* **7**, 172 (2016).
443. Yadav, U., Kumar, P., Gupta, S. & Rai, V. Role of MTHFR C677T gene polymorphism in the susceptibility of schizophrenia: An updated meta-analysis. *Asian J Psychiatr* **20**, 41–51 (2016).
444. Lajin, B., Alhaj Sakur, A., Michati, R. & Alachkar, A. Association between MTHFR C677T and A1298C, and MTRR A66G polymorphisms and susceptibility to schizophrenia in a Syrian study cohort. *Asian J Psychiatr* **5**, 144–149 (2012).
445. Saetre, P. *et al.* Methylenetetrahydrofolate reductase (MTHFR) C677T and A1298C polymorphisms and age of onset in schizophrenia: a combined analysis of independent samples. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **156**, 215–224 (2011).
446. Muntjewerff, J.-W. *et al.* Effects of season of birth and a common MTHFR gene variant on the risk of schizophrenia. *Eur Neuropsychopharmacol* **21**, 300–305 (2011).
447. Zhang, C. *et al.* Further evidence that methylenetetrahydrofolate reductase A1298C polymorphism is a risk factor for schizophrenia. *J Neural Transm (Vienna)* **117**, 1115–1117 (2010).

448. Yoshimi, A. *et al.* Gene-wide association study between the methylenetetrahydrofolate reductase gene (MTHFR) and schizophrenia in the Japanese population, with an updated meta-analysis on currently available data. *Schizophr. Res.* **124**, 216–222 (2010).
449. Vares, M. *et al.* Association between methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism and age of onset in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 610–618 (2010).
450. Roffman, J. L. *et al.* MTHFR 677C --> T genotype disrupts prefrontal function in schizophrenia through an interaction with COMT 158Val --> Met. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 17573–17578 (2008).
451. Mavros, M., Chiriță, V., Popescu, O. & Ferencz, B. [The genetic polymorphism of MTHFR gene in schizophrenia]. *Rev Med Chir Soc Med Nat Iasi* **112**, 76–82 (2008).
452. Lee, Y. S. *et al.* Serum homocysteine, folate level and methylenetetrahydrofolate reductase 677, 1298 gene polymorphism in Korean schizophrenic patients. *Neuroreport* **17**, 743–746 (2006).
453. Muntjewerff, J.-W. *et al.* Hyperhomocysteinemia, methylenetetrahydrofolate reductase 677TT genotype, and the risk for schizophrenia: a Dutch population based case-control study. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **135B**, 69–72 (2005).
454. Sazci, A., Ergül, E., Güzelhan, Y., Kaya, G. & Kara, I. Methylenetetrahydrofolate reductase gene polymorphisms in patients with schizophrenia. *Brain Res. Mol. Brain Res.* **117**, 104–107 (2003).
455. Joob, R. *et al.* Association between the methylenetetrahydrofolate reductase 677C-->T missense mutation and schizophrenia. *Mol. Psychiatry* **5**, 323–326 (2000).

456. Wei, J. & Hemmings, G. P. Allelic association of the MTHFR gene with schizophrenia. *Mol. Psychiatry* **4**, 115–116 (1999).
457. Roffman, J. L. *et al.* MTHFR 677C>T effects on anterior cingulate structure and function during response monitoring in schizophrenia: a preliminary study. *Brain Imaging Behav* **5**, 65–75 (2011).
458. Lee, Y. *et al.* Microduplications disrupting the MYT1L gene (2p25.3) are associated with schizophrenia. *Psychiatr. Genet.* **22**, 206–209 (2012).
459. Kepa, A. *et al.* Associations of the Intellectual Disability Gene MYT1L with Helix-Loop-Helix Gene Expression, Hippocampus Volume and Hippocampus Activation During Memory Retrieval. *Neuropsychopharmacology* **42**, 2516–2526 (2017).
460. Fanous, A. H. *et al.* Association between the 5q31.1 gene neurogenin1 and schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **144B**, 207–214 (2007).
461. Ho, B.-C. *et al.* Basic helix-loop-helix transcription factor NEUROG1 and schizophrenia: effects on illness susceptibility, MRI brain morphometry and cognitive abilities. *Schizophr. Res.* **106**, 192–199 (2008).
462. Ahmed, S. S. S. J., Akram Husain, R. S., Suresh Kumar, null & Ramakrishnan, V. Association Between NOS1 Gene Polymorphisms and Schizophrenia in Asian and Caucasian Populations: A Meta-Analysis. *Neuromolecular Med.* **19**, 452–461 (2017).
463. Li, Z. *et al.* Genome-wide association analysis identifies 30 new susceptibility loci for schizophrenia. *Nat. Genet.* **49**, 1576–1583 (2017).
464. Xu, F. *et al.* Association study of NOS1 gene polymorphisms with the risk of schizophrenia in Chinese Han origin. *Psychiatry Res* **246**, 844–845 (2016).

465. Zhang, Z. *et al.* Evidence for the contribution of NOS1 gene polymorphism (rs3782206) to prefrontal function in schizophrenia patients and healthy controls. *Neuropsychopharmacology* **40**, 1383–1394 (2015).
466. Wang, J. *et al.* [Association study of NOS1 gene polymorphisms and schizophrenia]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **29**, 459–463 (2012).
467. Cui, H. *et al.* A putative cis-acting polymorphism in the NOS1 gene is associated with schizophrenia and NOS1 immunoreactivity in the postmortem brain. *Schizophr. Res.* **121**, 172–178 (2010).
468. Liou, Y.-J., Tsai, S.-J., Hong, C.-J. & Liao, D.-L. Association analysis for the CA repeat polymorphism of the neuronal nitric oxide synthase (NOS1) gene and schizophrenia. *Schizophr. Res.* **65**, 57–59 (2003).
469. Shinkai, T., Ohmori, O., Hori, H. & Nakamura, J. Allelic association of the neuronal nitric oxide synthase (NOS1) gene with schizophrenia. *Mol. Psychiatry* **7**, 560–563 (2002).
470. Rose, E. J. *et al.* The NOS1 variant rs6490121 is associated with variation in prefrontal function and grey matter density in healthy individuals. *Neuroimage* **60**, 614–622 (2012).
471. Saiz, P. ASSOCIATION STUDY OF ENDOTHELIAL NITRIC OXIDE SYNTHASE (NOS3) GENE POLYMORPHISMS AND SCHIZOPHRENIA. *Schizophrenia Research* **102**, 199 (2008).
472. Zhang, B. *et al.* Association of the NOTCH4 Gene Polymorphism rs204993 with Schizophrenia in the Chinese Han Population. *Biomed Res Int* **2015**, 408096 (2015).
473. Su, L. *et al.* Association between the NOTCH4 gene rs3131296 polymorphism with schizophrenia risk in the Chinese Zhuang population and Chinese Han population. *Acta Neuropsychiatr* **26**, 240–245 (2014).

474. Shayevitz, C., Cohen, O. S., Faraone, S. V. & Glatt, S. J. A re-review of the association between the NOTCH4 locus and schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 477–483 (2012).
475. Liu, C.-M. *et al.* Association evidence of schizophrenia with distal genomic region of NOTCH4 in Taiwanese families. *Genes Brain Behav.* **6**, 497–502 (2007).
476. Glatt, S. J., Wang, R. S., Yeh, Y.-C., Tsuang, M. T. & Faraone, S. V. Five NOTCH4 polymorphisms show weak evidence for association with schizophrenia: evidence from meta-analyses. *Schizophr. Res.* **73**, 281–290 (2005).
477. Prasad, S. *et al.* Association analysis of NOTCH 4 polymorphisms with schizophrenia among two independent family based samples. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **131B**, 6–9 (2004).
478. Tochigi, M. *et al.* Association of six polymorphisms of the NOTCH4 gene with schizophrenia in the Japanese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **128B**, 37–40 (2004).
479. Kaneko, N. *et al.* Transmission disequilibrium test and haplotype analysis of the NOTCH4 gene in Japanese patients with schizophrenia. *Psychiatry Clin. Neurosci.* **58**, 199–205 (2004).
480. Luo, X. *et al.* NOTCH4 gene haplotype is associated with schizophrenia in African Americans. *Biol. Psychiatry* **55**, 112–117 (2004).
481. Takahashi, S. *et al.* Family-based association study of the NOTCH4 gene in schizophrenia using Japanese and Chinese samples. *Biol. Psychiatry* **54**, 129–135 (2003).
482. Fan, J. B. *et al.* A family-based and case-control association study of the NOTCH4 gene and schizophrenia. *Mol. Psychiatry* **7**, 100–103 (2002).

483. Wei, J. & Hemmings, G. P. The NOTCH4 locus is associated with susceptibility to schizophrenia. *Nat. Genet.* **25**, 376–377 (2000).
484. Wassink, T. H., Nopoulos, P., Pietila, J., Crowe, R. R. & Andreasen, N. C. NOTCH4 and the frontal lobe in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **118B**, 1–7 (2003).
485. Saito, A. *et al.* Association study of putative promoter polymorphisms in the neuroplastin gene and schizophrenia. *Neurosci. Lett.* **411**, 168–173 (2007).
486. Desrivières, S. *et al.* Single nucleotide polymorphism in the neuroplastin locus associates with cortical thickness and intellectual ability in adolescents. *Mol. Psychiatry* **20**, 263–274 (2015).
487. Yoosefee, S. *et al.* Association Between Neuregulin-1 Gene Variant (rs2439272) and Schizophrenia and Its Negative Symptoms in an Iranian Population. *Iran J Psychiatry* **11**, 147–153 (2016).
488. Réthelyi, J. M. *et al.* Association study of NRG1, DTNBP1, RGS4, G72/G30, and PIP5K2A with schizophrenia and symptom severity in a Hungarian sample. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 792–801 (2010).
489. Cho, Y. *et al.* Effects of genetic variations in NRG1 on cognitive domains in patients with schizophrenia and healthy individuals. *Psychiatr. Genet.* **25**, 147–154 (2015).
490. Kukshal, P. *et al.* Association study of neuregulin-1 gene polymorphisms in a North Indian schizophrenia sample. *Schizophr. Res.* **144**, 24–30 (2013).
491. Yang, J. Z. *et al.* Association study of neuregulin 1 gene with schizophrenia. *Mol. Psychiatry* **8**, 706–709 (2003).

492. Mohamad Shariati, S. A., Behmanesh, M. & Galehdari, H. A Study of the Association between SNP8NRG241930 in the 5' End of Neuroglin 1 Gene with Schizophrenia in a Group of Iranian Patients. *Cell J* **13**, 91–96 (2011).
493. Alaerts, M. *et al.* Support for NRG1 as a susceptibility factor for schizophrenia in a northern Swedish isolated population. *Arch. Gen. Psychiatry* **66**, 828–837 (2009).
494. Nicodemus, K. K. *et al.* A 5' promoter region SNP in NRG1 is associated with schizophrenia risk and type III isoform expression. *Mol. Psychiatry* **14**, 741–743 (2009).
495. Zhang, H. *et al.* [Association analysis of neuregulin 1 gene polymorphism with schizophrenia in Chinese Han population]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **26**, 16–20 (2009).
496. Munafò, M. R., Attwood, A. S. & Flint, J. Neuregulin 1 genotype and schizophrenia. *Schizophr Bull* **34**, 9–12 (2008).
497. Law, A. J. *et al.* Neuregulin 1 transcripts are differentially expressed in schizophrenia and regulated by 5' SNPs associated with the disease. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 6747–6752 (2006).
498. Kim, J. W. *et al.* Linkage and association of schizophrenia with genetic variations in the locus of neuregulin 1 in Korean population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141B**, 281–286 (2006).
499. Hong, C.-J. *et al.* Case-control and family-based association studies between the neuregulin 1 (Arg38Gln) polymorphism and schizophrenia. *Neurosci. Lett.* **366**, 158–161 (2004).
500. Yang, S.-A. Association between a Missense Polymorphism (rs3924999, Arg253Gln) of Neuregulin 1 and Schizophrenia in Korean Population. *Exp Neurobiol* **21**, 158–163 (2012).

501. Stefansson, H. *et al.* Association of neuregulin 1 with schizophrenia confirmed in a Scottish population. *Am. J. Hum. Genet.* **72**, 83–87 (2003).
502. Bousman, C. A. *et al.* Neuregulin-1 (NRG1) polymorphisms linked with psychosis transition are associated with enlarged lateral ventricles and white matter disruption in schizophrenia. *Psychol Med* **48**, 801–809 (2018).
503. Nickl-Jockschat, T. *et al.* A Neuregulin-1 schizophrenia susceptibility variant causes perihippocampal fiber tract anomalies in healthy young subjects. *Brain Behav* **4**, 215–226 (2014).
504. Tosato, S. *et al.* Is neuregulin 1 involved in determining cerebral volumes in schizophrenia? Preliminary results showing a decrease in superior temporal gyrus volume. *Neuropsychobiology* **65**, 119–125 (2012).
505. Mata, I. *et al.* A neuregulin 1 variant is associated with increased lateral ventricle volume in patients with first-episode schizophrenia. *Biol. Psychiatry* **65**, 535–540 (2009).
506. Winterer, G. *et al.* Association of 5' end neuregulin-1 (NRG1) gene variation with subcortical medial frontal microstructure in humans. *Neuroimage* **40**, 712–718 (2008).
507. McIntosh, A. M. *et al.* The effects of a neuregulin 1 variant on white matter density and integrity. *Mol. Psychiatry* **13**, 1054–1059 (2008).
508. Gruber, O. *et al.* Neuregulin-1 haplotype HAP(ICE) is associated with lower hippocampal volumes in schizophrenic patients and in non-affected family members. *J Psychiatr Res* **43**, 1–6 (2008).

509. Ohi, K. *et al.* Functional genetic variation at the NRG1 gene and schizophrenia: evidence from a gene-based case-control study and gene expression analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 405–413 (2012).
510. Donohoe, G. *et al.* A neuropsychological investigation of the genome wide associated schizophrenia risk variant NRG1 rs12807809. *Schizophr. Res.* **125**, 304–306 (2011).
511. Thong, J. Y. J. *et al.* Effects of the neurogranin variant rs12807809 on thalamocortical morphology in schizophrenia. *PLoS ONE* **8**, e85603 (2013).
512. Walton, E. *et al.* The impact of genome-wide supported schizophrenia risk variants in the neurogranin gene on brain structure and function. *PLoS ONE* **8**, e76815 (2013).
513. Ohi, K. *et al.* Impact of the genome wide supported NRG1 gene on anterior cingulate morphology in schizophrenia. *PLoS ONE* **7**, e29780 (2012).
514. Rose, E. J. *et al.* The effect of the neurogranin schizophrenia risk variant rs12807809 on brain structure and function. *Twin Res Hum Genet* **15**, 296–303 (2012).
515. Mühleisen, T. W. *et al.* Resequencing and follow-up of neurexin 1 (NRXN1) in schizophrenia patients. *Schizophr. Res.* **127**, 35–40 (2011).
516. Yue, W. *et al.* A case-control association study of NRXN1 polymorphisms with schizophrenia in Chinese Han population. *Behav Brain Funct* **7**, 7 (2011).
517. Voineskos, A. N. *et al.* Neurexin-1 and frontal lobe white matter: an overlapping intermediate phenotype for schizophrenia and autism spectrum disorders. *PLoS ONE* **6**, e20982 (2011).
518. Hattori, M. *et al.* Novel polymorphisms in the promoter region of the neurotrophin-3 gene and their associations with schizophrenia. *Am. J. Med. Genet.* **114**, 304–309 (2002).

519. Nanko, S. *et al.* Neurotrophin-3 gene polymorphism associated with schizophrenia. *Acta Psychiatr Scand* **89**, 390–392 (1994).
520. Kunugi, H. *et al.* Dinucleotide repeat polymorphism in the neurotrophin-3 gene and hippocampal volume in psychoses. *Schizophr. Res.* **37**, 271–273 (1999).
521. Austin, J. *et al.* The high affinity neurotensin receptor gene (NTSR1): comparative sequencing and association studies in schizophrenia. *Mol. Psychiatry* **5**, 552–557 (2000).
522. Li, J. *et al.* The NTSR1 gene modulates the association between hippocampal structure and working memory performance. *Neuroimage* **75**, 79–86 (2013).
523. Moens, L. N. *et al.* PCM1 and schizophrenia: a replication study in the Northern Swedish population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **153B**, 1240–1243 (2010).
524. Datta, S. R. *et al.* A threonine to isoleucine missense mutation in the pericentriolar material 1 gene is strongly associated with schizophrenia. *Mol. Psychiatry* **15**, 615–628 (2010).
525. Gurling, H. M. D. *et al.* Genetic association and brain morphology studies and the chromosome 8p22 pericentriolar material 1 (PCM1) gene in susceptibility to schizophrenia. *Arch. Gen. Psychiatry* **63**, 844–854 (2006).
526. Jungerius, B. J. *et al.* An association screen of myelin-related genes implicates the chromosome 22q11 PIK4CA gene in schizophrenia. *Mol. Psychiatry* **13**, 1060–1068 (2008).
527. Wang, Q. *et al.* Genome-wide association analysis with gray matter volume as a quantitative phenotype in first-episode treatment-naïve patients with schizophrenia. *PLoS ONE* **8**, e75083 (2013).
528. Kordi-Tamandani, D. M. & Mir, A. Relationship between phosphoinositide-3-kinase genetic polymorphism and schizophrenia. *Nord J Psychiatry* **70**, 272–275 (2016).

529. Hu, J.-X. *et al.* An association study between PPP1R1B gene and schizophrenia in the Chinese population. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **31**, 1303–1306 (2007).
530. Kunii, Y. *et al.* Altered DARPP-32 expression in the superior temporal gyrus in schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 1139–1143 (2011).
531. Meyer-Lindenberg, A. *et al.* Genetic evidence implicating DARPP-32 in human frontostriatal structure, function, and cognition. *J. Clin. Invest.* **117**, 672–682 (2007).
532. Persson, N., Persson, J., Lavebratt, C. & Fischer, H. Effects of DARPP-32 Genetic Variation on Prefrontal Cortex Volume and Episodic Memory Performance. *Front Neurosci* **11**, 244 (2017).
533. Rujescu, D. *et al.* Association study of a SNP coding for a M129V substitution in the prion protein in schizophrenia. *Schizophr. Res.* **62**, 289–291 (2003).
534. Rujescu, D. *et al.* Methionine homozygosity at codon 129 in the prion protein is associated with white matter reduction and enlargement of CSF compartments in healthy volunteers and schizophrenic patients. *Neuroimage* **15**, 200–206 (2002).
535. Guo, X., Tang, P., Yang, C. & Li, R. Proline dehydrogenase gene (PRODH) polymorphisms and schizophrenia susceptibility: a meta-analysis. *Metab Brain Dis* **33**, 89–97 (2018).
536. Ghasemvand, F., Omidinia, E., Salehi, Z. & Rahmanzadeh, S. Relationship between polymorphisms in the proline dehydrogenase gene and schizophrenia risk. *Genet. Mol. Res.* **14**, 11681–11691 (2015).
537. Willis, A., Bender, H. U., Steel, G. & Valle, D. PRODH variants and risk for schizophrenia. *Amino Acids* **35**, 673–679 (2008).

538. Li, T. *et al.* Evidence for association between novel polymorphisms in the PRODH gene and schizophrenia in a Chinese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **129B**, 13–15 (2004).
539. Fan, J.-B. *et al.* A family-based association study of T1945C polymorphism in the proline dehydrogenase gene and schizophrenia in the Chinese population. *Neurosci. Lett.* **338**, 252–254 (2003).
540. Ota, V. K. *et al.* PRODH polymorphisms, cortical volumes and thickness in schizophrenia. *PLoS ONE* **9**, e87686 (2014).
541. Ayalew, M. *et al.* Convergent functional genomics of schizophrenia: from comprehensive understanding to genetic risk prediction. *Mol. Psychiatry* **17**, 887–905 (2012).
542. Cheng, C.-Y. *et al.* The association of RAB18 gene polymorphism (rs3765133) with cerebellar volume in healthy adults. *Cerebellum* **13**, 616–622 (2014).
543. Shifman, S. *et al.* Genome-wide association identifies a common variant in the reelin gene that increases the risk of schizophrenia only in women. *PLoS Genet.* **4**, e28 (2008).
544. Chang, L.-H. *et al.* Association of RELN promoter SNPs with schizophrenia in the Chinese population. *Zool. Res.* **32**, 504–508 (2011).
545. Liu, X.-Y., Li, M., Yang, S.-Y., Su, B. & Yin, L.-D. [Association of RELN SNP rs7341475 with schizophrenia in the Chinese population]. *Zool. Res.* **32**, 499–503 (2011).
546. Li, W. *et al.* Association study of RELN polymorphisms with schizophrenia in Han Chinese population. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 1505–1511 (2011).
547. Li, M. *et al.* Analysis of common genetic variants identifies RELN as a risk gene for schizophrenia in Chinese population. *World J. Biol. Psychiatry* **14**, 91–99 (2013).

548. Galaktionova, D. I., Gareeva, A. E., Khusnutdinova, E. K. & Nasedkina, T. V. [The association of polymorphisms in SLC18A1, TPH1 and RELN genes with risk of paranoid schizophrenia]. *Mol. Biol. (Mosk.)* **48**, 629–639 (2014).
549. Li, W., Guo, X. & Xiao, S. Evaluating the relationship between reelin gene variants (rs7341475 and rs262355) and schizophrenia: A meta-analysis. *Neurosci. Lett.* **609**, 42–47 (2015).
550. Zhou, Z. *et al.* Identification of RELN variation p.Thr3192Ser in a Chinese family with schizophrenia. *Sci Rep* **6**, 24327 (2016).
551. Gregório, S. P. *et al.* Polymorphisms in genes involved in neurodevelopment may be associated with altered brain morphology in schizophrenia: preliminary evidence. *Psychiatry Res* **165**, 1–9 (2009).
552. Mirnics, K., Middleton, F. A., Stanwood, G. D., Lewis, D. A. & Levitt, P. Disease-specific changes in regulator of G-protein signaling 4 (RGS4) expression in schizophrenia. *Mol. Psychiatry* **6**, 293–301 (2001).
553. Chowdari, K. V. *et al.* Association and linkage analyses of RGS4 polymorphisms in schizophrenia. *Hum. Mol. Genet.* **11**, 1373–1380 (2002).
554. Morris, D. W. *et al.* Confirming RGS4 as a susceptibility gene for schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **125B**, 50–53 (2004).
555. Chen, X. *et al.* Regulator of G-protein signaling 4 (RGS4) gene is associated with schizophrenia in Irish high density families. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **129B**, 23–26 (2004).

556. Zhang, F. *et al.* Association analysis of the RGS4 gene in Han Chinese and Scottish populations with schizophrenia. *Genes Brain Behav.* **4**, 444–448 (2005).
557. Guo, S. *et al.* RGS4 polymorphisms and risk of schizophrenia: an association study in Han Chinese plus meta-analysis. *Neurosci. Lett.* **406**, 122–127 (2006).
558. Chowdari, K. V. *et al.* Linkage disequilibrium patterns and functional analysis of RGS4 polymorphisms in relation to schizophrenia. *Schizophr Bull* **34**, 118–126 (2008).
559. So, H.-C. *et al.* An association study of RGS4 polymorphisms with clinical phenotypes of schizophrenia in a Chinese population. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **147B**, 77–85 (2008).
560. Bowden, N. A., Scott, R. J. & Tooney, P. A. Altered expression of regulator of G-protein signalling 4 (RGS4) mRNA in the superior temporal gyrus in schizophrenia. *Schizophr. Res.* **89**, 165–168 (2007).
561. Prasad, K. M. R. *et al.* Genetic polymorphisms of the RGS4 and dorsolateral prefrontal cortex morphometry among first episode schizophrenia patients. *Mol. Psychiatry* **10**, 213–219 (2005).
562. Buckholtz, J. W. *et al.* Allelic variation in RGS4 impacts functional and structural connectivity in the human brain. *J. Neurosci.* **27**, 1584–1593 (2007).
563. Bowden, N. A., Scott, R. J. & Tooney, P. A. Altered gene expression in the superior temporal gyrus in schizophrenia. *BMC Genomics* **9**, 199 (2008).
564. Zhu, B. *et al.* The SEMA5A gene is associated with hippocampal volume, and their interaction is associated with performance on Raven’s Progressive Matrices. *Neuroimage* **88**, 181–187 (2014).

565. Bonnet-Brilhault, F. *et al.* Serotonin transporter gene polymorphism and schizophrenia: an association study. *Biol. Psychiatry* **42**, 634–636 (1997).
566. Pae, C.-U. *et al.* Polymorphism of the serotonin transporter gene and symptomatic dimensions of schizophrenia in the Korean population. *Neuropsychobiology* **47**, 182–186 (2003).
567. Vijayan, N. N. *et al.* Evidence of association of serotonin transporter gene polymorphisms with schizophrenia in a South Indian population. *J. Hum. Genet.* **54**, 538–542 (2009).
568. Li, W. *et al.* Association of serotonin transporter gene (SLC6A4) polymorphisms with schizophrenia susceptibility and symptoms in a Chinese-Han population. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **44**, 290–295 (2013).
569. Pae, C.-U. *et al.* Tumor necrosis factor- α gene polymorphism at position -308 and schizophrenia in the Korean population. *Psychiatry Clin. Neurosci.* **57**, 399–403 (2003).
570. Hashimoto, R. *et al.* Association analysis of the -308G>A promoter polymorphism of the tumor necrosis factor α (TNF- α) gene in Japanese patients with schizophrenia. *J Neural Transm (Vienna)* **111**, 217–221 (2004).
571. Hänninen, K. *et al.* Tumor necrosis factor- α --G308A polymorphism in schizophrenia in a Finnish population. *Neurosci. Lett.* **385**, 76–81 (2005).
572. Morar, B. *et al.* Evaluation of association of SNPs in the TNF α gene region with schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **144B**, 318–324 (2007).
573. Kadasah, S., Arfin, M., Rizvi, S., Al-Asmari, M. & Al-Asmari, A. Tumor necrosis factor- α and - β genetic polymorphisms as a risk factor in Saudi patients with schizophrenia. *Neuropsychiatr Dis Treat* **13**, 1081–1088 (2017).

574. Baune, B. T. *et al.* Tumor necrosis factor gene variation predicts hippocampus volume in healthy individuals. *Biol. Psychiatry* **72**, 655–662 (2012).
575. Suchanek-Raif, R. *et al.* Association Study of Tumor Necrosis Factor Receptor 1 (TNFR1) Gene Polymorphisms with Schizophrenia in the Polish Population. *Mediators Inflamm.* **2017**, 6016023 (2017).
576. Stacey, D. *et al.* TNF receptors 1 and 2 exert distinct region-specific effects on striatal and hippocampal grey matter volumes (VBM) in healthy adults. *Genes Brain Behav.* **16**, 352–360 (2017).
577. Thabet, S. *et al.* Association of the Met-196-Arg variation of human tumor necrosis factor receptor 2 (TNFR2) with paranoid schizophrenia. *J. Mol. Neurosci.* **43**, 358–363 (2011).
578. Wassink, T. H., Crowe, R. R. & Andreasen, N. C. Tumor necrosis factor receptor-II: heritability and effect on brain morphology in schizophrenia. *Mol. Psychiatry* **5**, 678–682 (2000).
579. Chiu, H. J., Wang, Y. C., Chen, J. Y., Hong, C. J. & Tsai, S. J. Association study of the p53-gene Pro72Arg polymorphism in schizophrenia. *Psychiatry Res* **105**, 279–283 (2001).
580. Yang, Y. *et al.* Tumor suppressor gene TP53 is genetically associated with schizophrenia in the Chinese population. *Neurosci. Lett.* **369**, 126–131 (2004).
581. Ni, X. *et al.* Human p53 tumor suppressor gene (TP53) and schizophrenia: case-control and family studies. *Neurosci. Lett.* **388**, 173–178 (2005).
582. Lung, F.-W. *et al.* Association of DRD4 uVNTR and TP53 codon 72 polymorphisms with schizophrenia: a case-control study. *BMC Med. Genet.* **10**, 147 (2009).

583. Molina, V. *et al.* Convergent evidence of the contribution of TP53 genetic variation (Pro72Arg) to metabolic activity and white matter volume in the frontal lobe in schizophrenia patients. *Neuroimage* **56**, 45–51 (2011).
584. Watanabe, Y. *et al.* A two-stage case-control association study between the tryptophan hydroxylase 2 (TPH2) gene and schizophrenia in a Japanese population. *Schizophr. Res.* **137**, 264–266 (2012).
585. Yi, Z. *et al.* Common variants in the TPH2 promoter confer susceptibility to paranoid schizophrenia. *J. Mol. Neurosci.* **47**, 465–469 (2012).
586. Xu, X. M., Ding, M., Pang, H. & Wang, B. J. TPH2 gene polymorphisms in the regulatory region are associated with paranoid schizophrenia in Northern Han Chinese. *Genet. Mol. Res.* **13**, 1497–1507 (2014).
587. Zhang, X. *et al.* Association study of the DISC1/TRAX locus with schizophrenia in a Japanese population. *Schizophr. Res.* **79**, 175–180 (2005).
588. Cannon, T. D. *et al.* Association of DISC1/TRAX haplotypes with schizophrenia, reduced prefrontal gray matter, and impaired short- and long-term memory. *Arch. Gen. Psychiatry* **62**, 1205–1213 (2005).
589. Aleksic, B. *et al.* Analysis of the VAV3 as candidate gene for schizophrenia: evidences from voxel-based morphometry and mutation screening. *Schizophr Bull* **39**, 720–728 (2013).
590. Fulzele, S. & Pillai, A. Decreased VEGF mRNA expression in the dorsolateral prefrontal cortex of schizophrenia subjects. *Schizophr. Res.* **115**, 372–373 (2009).
591. Gao, K. *et al.* Association study of VEGFA polymorphisms with schizophrenia in Han Chinese population. *Neurosci. Lett.* **590**, 121–125 (2015).

592. Pillai, A. *et al.* Association of serum VEGF levels with prefrontal cortex volume in schizophrenia. *Mol. Psychiatry* **21**, 686–692 (2016).
593. Steinberg, S. *et al.* Common variants at VRK2 and TCF4 conferring risk of schizophrenia. *Hum. Mol. Genet.* **20**, 4076–4081 (2011).
594. Zhang, B. *et al.* Association of the VRK2 gene rs3732136 polymorphism with schizophrenia in a Northwest Chinese Han population. *Genet. Mol. Res.* **14**, 9404–9411 (2015).
595. Chang, H. *et al.* Further evidence of VRK2 rs2312147 associated with schizophrenia. *World J. Biol. Psychiatry* **17**, 457–466 (2016).
596. Sohn, H. *et al.* Effects of VRK2 (rs2312147) on white matter connectivity in patients with schizophrenia. *PLoS ONE* **9**, e103519 (2014).
597. Li, M. *et al.* Meta-analysis and brain imaging data support the involvement of VRK2 (rs2312147) in schizophrenia susceptibility. *Schizophr. Res.* **142**, 200–205 (2012).
598. Ikeda, M. *et al.* Identification of YWHAE, a gene encoding 14-3-3epsilon, as a possible susceptibility gene for schizophrenia. *Hum. Mol. Genet.* **17**, 3212–3222 (2008).
599. Kido, M. *et al.* The polymorphism of YWHAE, a gene encoding 14-3-3epsilon, and brain morphology in schizophrenia: a voxel-based morphometric study. *PLoS ONE* **9**, e103571 (2014).
600. Takahashi, T. *et al.* The polymorphism of YWHAE, a gene encoding 14-3-3epsilon, and orbitofrontal sulcogyral pattern in patients with schizophrenia and healthy subjects. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **51**, 166–171 (2014).
601. Chen, W.-Y. *et al.* Case-control study and transmission disequilibrium test provide consistent evidence for association between schizophrenia and genetic variation in the 22q11 gene ZDHC8. *Hum. Mol. Genet.* **13**, 2991–2995 (2004).

602. Mukai, J. *et al.* Evidence that the gene encoding ZDHHC8 contributes to the risk of schizophrenia. *Nat. Genet.* **36**, 725–731 (2004).
603. Faul, T. *et al.* ZDHHC8 as a candidate gene for schizophrenia: analysis of a putative functional intronic marker in case-control and family-based association studies. *BMC Psychiatry* **5**, 35 (2005).
604. Ota, V. K. *et al.* ZDHHC8 gene may play a role in cortical volumes of patients with schizophrenia. *Schizophr. Res.* **145**, 33–35 (2013).
605. O'Donovan, M. C. *et al.* Identification of loci associated with schizophrenia by genome-wide association and follow-up. *Nat. Genet.* **40**, 1053–1055 (2008).
606. Riley, B. *et al.* Replication of association between schizophrenia and ZNF804A in the Irish Case-Control Study of Schizophrenia sample. *Mol. Psychiatry* **15**, 29–37 (2010).
607. Li, M. *et al.* ZNF804A and schizophrenia susceptibility in Asian populations. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 794–802 (2012).
608. Sazci, A., Ozel, M. D., Ergul, E. & Yildiz, M. A polymerase chain reaction-restriction fragment length polymorphism method for screening ZNF804A gene polymorphism (rs1344706) in patients with schizophrenia: a significant association. *Genet Test Mol Biomarkers* **16**, 157–161 (2012).
609. Zhang, R. *et al.* Further evidence for the association of genetic variants of ZNF804A with schizophrenia and a meta-analysis for genome-wide significance variant rs1344706. *Schizophr. Res.* **141**, 40–47 (2012).
610. Schwab, S. G. *et al.* Association of rs1344706 in the ZNF804A gene with schizophrenia in a case/control sample from Indonesia. *Schizophr. Res.* **147**, 46–52 (2013).

611. Yang, Y. *et al.* Evaluation of the relationship between the ZNF804A single nucleotide polymorphism rs1344706 A/C variant and schizophrenia subtype in Han Chinese patients. *Int J Psychiatry Med* **45**, 269–278 (2013).
612. Zhu, M. *et al.* Association between rs1344706 of ZNF804A and schizophrenia: a meta-analysis. *Genomics Proteomics Bioinformatics* **12**, 292–296 (2014).
613. Donohoe, G. *et al.* ZNF804A risk allele is associated with relatively intact gray matter volume in patients with schizophrenia. *Neuroimage* **54**, 2132–2137 (2011).
614. Kuswanto, C. N. *et al.* Genome-wide supported psychosis risk variant in ZNF804A gene and impact on cortico-limbic WM integrity in schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **159B**, 255–262 (2012).
615. Sprooten, E. *et al.* An investigation of a genomewide supported psychosis variant in ZNF804A and white matter integrity in the human brain. *Magn Reson Imaging* **30**, 1373–1380 (2012).
616. Wassink, T. H. *et al.* Influence of ZNF804a on brain structure volumes and symptom severity in individuals with schizophrenia. *Arch. Gen. Psychiatry* **69**, 885–892 (2012).
617. Ikuta, T. *et al.* A schizophrenia risk gene, ZNF804A, is associated with brain white matter microstructure. *Schizophr. Res.* **155**, 15–20 (2014).
618. Schultz, C. C. *et al.* ZNF804A and cortical structure in schizophrenia: in vivo and postmortem studies. *Schizophr Bull* **40**, 532–541 (2014).
619. Nenadic, I. *et al.* ZNF804A genetic variation (rs1344706) affects brain grey but not white matter in schizophrenia and healthy subjects. *Psychol Med* **45**, 143–152 (2015).

620. Wei, Q. *et al.* Association of the ZNF804A gene polymorphism rs1344706 with white matter density changes in Chinese schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **36**, 122–127 (2012).
621. Mallas, E.-J. *et al.* Genome-wide discovered psychosis-risk gene ZNF804A impacts on white matter microstructure in health, schizophrenia and bipolar disorder. *PeerJ* **4**, e1570 (2016).