Class 8: PCA Mini Project

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Side note before starting about scaling:

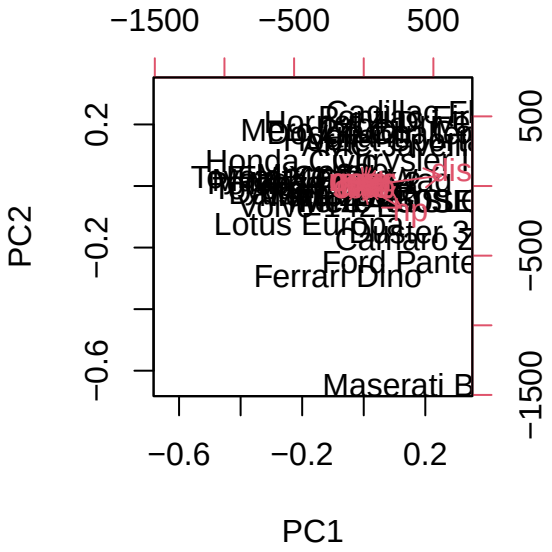
```
#Mean value of each column in mtcars dataset
apply(mtcars, 2, mean)
```

```
      mpg       cyl      disp        hp      drat        wt      qsec
20.090625  6.187500 230.721875 146.687500  3.596563  3.217250 17.848750
       vs        am      gear      carb
 0.437500  0.406250  3.687500  2.812500
```

```
#Spread of each column via standard deviation
apply(mtcars, 2, sd)
```

```
       mpg        cyl       disp         hp       drat         wt
 6.0269481  1.7859216 123.9386938 68.5628685  0.5346787  0.9784574
      qsec         vs         am       gear       carb
 1.7869432  0.5040161  0.4989909  0.7378041  1.6152000
```

```
pca <- prcomp(mtcars)
biplot(pca)
```



Without scaling, the columns are measured in different units and the pca analysis will be biased towards the units with higher counts/larger spread.

```
#Scale the data
mtscale <- scale(mtcars)
head(mtscale)
```

```
                    mpg        cyl        disp         hp       drat
Mazda RX4         0.1508848 -0.1049878 -0.57061982 -0.5350928  0.5675137
Mazda RX4 Wag     0.1508848 -0.1049878 -0.57061982 -0.5350928  0.5675137
Datsun 710        0.4495434 -1.2248578 -0.99018209 -0.7830405  0.4739996
Hornet 4 Drive    0.2172534 -0.1049878  0.22009369 -0.5350928 -0.9661175
Hornet Sportabout -0.2307345  1.0148821  1.04308123  0.4129422 -0.8351978
Valiant           -0.3302874 -0.1049878 -0.04616698 -0.6080186 -1.5646078
                          wt       qsec         vs         am       gear
Mazda RX4         -0.610399567 -0.7771651 -0.8680278  1.1899014  0.4235542
Mazda RX4 Wag     -0.349785269 -0.4637808 -0.8680278  1.1899014  0.4235542
Datsun 710        -0.917004624  0.4260068  1.1160357  1.1899014  0.4235542
Hornet 4 Drive    -0.002299538  0.8904872  1.1160357 -0.8141431 -0.9318192
Hornet Sportabout  0.227654255 -0.4637808 -0.8680278 -0.8141431 -0.9318192
Valiant            0.248094592  1.3269868  1.1160357 -0.8141431 -0.9318192
                         carb
Mazda RX4         0.7352031
```

```
Mazda RX4 Wag      0.7352031
Datsun 710        -1.1221521
Hornet 4 Drive    -1.1221521
Hornet Sportabout -0.5030337
Valiant           -1.1221521
```

```
#Now look at the mean and standard deviation of each column
round(apply(mtscale, 2, mean), 3)
```

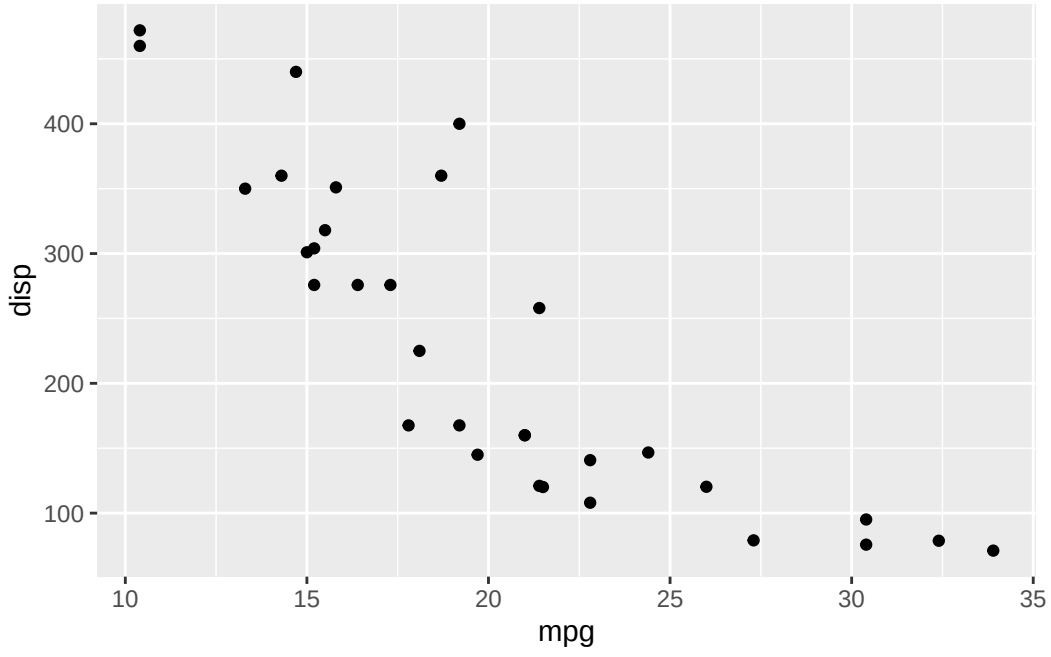
```
 mpg  cyl disp   hp drat   wt qsec   vs   am gear carb
   0    0    0    0    0    0    0    0    0    0    0
```

```
round(apply(mtscale, 2, sd), 3)
```

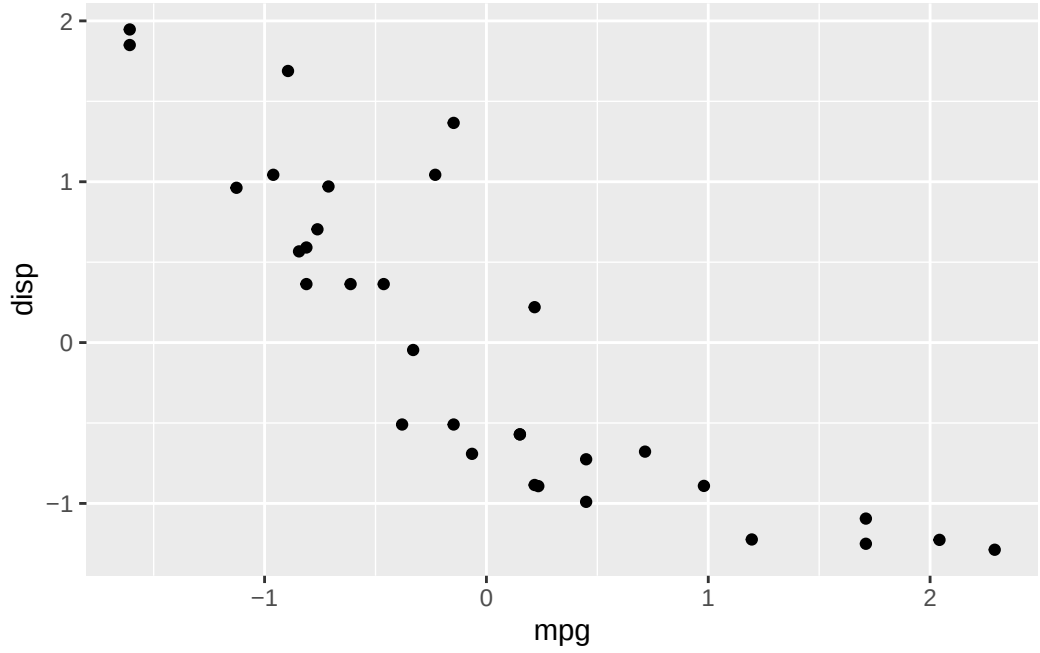
```
 mpg  cyl disp   hp drat   wt qsec   vs   am gear carb
   1    1    1    1    1    1    1    1    1    1    1
```

Let's plot to make sure the scaled data still has the same relationships. Plot 'mpg' vs 'disp' for both the original and scaled data

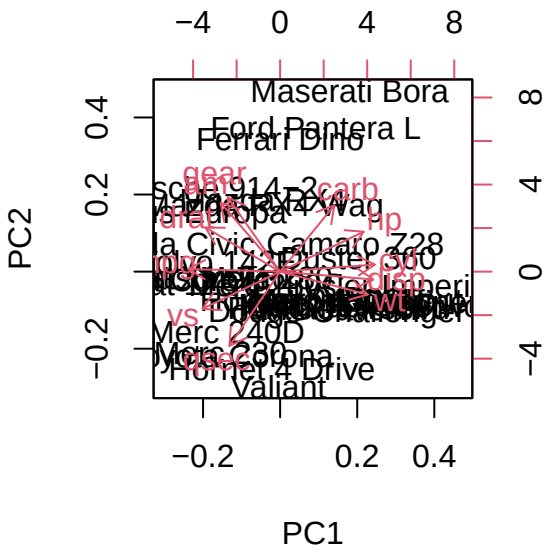
```
library(ggplot2)
ggplot(mtcars, aes(mpg, disp)) +
        geom_point()
```



```
ggplot(mtscale, aes(mpg, disp)) +
          geom_point()
```




```
pca2 <- prcomp(mtscale)
biplot(pca2)
```



Breast Cancer FNA Data

Preparing the data

First download the csv file from the class website and put it in the current working directory thru Finder, then read it into R

```
wisc.df <- read.csv("WisconsinCancer.csv",
                    row.names=1)
#head(wisc.df)
```

Omit the 'diagnosis' column and create a new dataset

```
wisc.data <- wisc.df[,-1]
```

Create a new vector that contains the diagnosis column from the original dataset.

```
diagnosis <- as.factor(wisc.df$diagnosis)
```

Q1: How many observations are in this dataset?

```
nrow(wisc.df)
```

```
[1] 569
```

Q2: How many of the observations have a malignant diagnosis?

```
table(wisc.df$diagnosis)
```

```
  B   M
357 212
```

There are 212 malignant diagnoses.

Q3: How many variables/features in the data are suffixed with '_mean'?

```
length(grep("_mean", colnames(wisc.data)))
```

```
[1] 10
```

There are 10.

Prinicpal Component Analysis

Check the mean and standard deviation of the features to determine if the data should be scaled.

```
round(colMeans(wisc.data), 3)
```

radius_mean	texture_mean	perimeter_mean
14.127	19.290	91.969
area_mean	smoothness_mean	compactness_mean
654.889	0.096	0.104
concavity_mean	concave.points_mean	symmetry_mean
0.089	0.049	0.181
fractal_dimension_mean	radius_se	texture_se
0.063	0.405	1.217
perimeter_se	area_se	smoothness_se
2.866	40.337	0.007
compactness_se	concavity_se	concave.points_se
0.025	0.032	0.012
symmetry_se	fractal_dimension_se	radius_worst
0.021	0.004	16.269
texture_worst	perimeter_worst	area_worst
25.677	107.261	880.583
smoothness_worst	compactness_worst	concavity_worst
0.132	0.254	0.272
concave.points_worst	symmetry_worst	fractal_dimension_worst
0.115	0.290	0.084

```
round(apply(wisc.data, 2, sd), 3)
```

radius_mean	texture_mean	perimeter_mean
3.524	4.301	24.299
area_mean	smoothness_mean	compactness_mean
351.914	0.014	0.053
concavity_mean	concave.points_mean	symmetry_mean
0.080	0.039	0.027
fractal_dimension_mean	radius_se	texture_se
0.007	0.277	0.552
perimeter_se	area_se	smoothness_se
2.022	45.491	0.003
compactness_se	concavity_se	concave.points_se
0.018	0.030	0.006
symmetry_se	fractal_dimension_se	radius_worst
0.008	0.003	4.833
texture_worst	perimeter_worst	area_worst
6.146	33.603	569.357
smoothness_worst	compactness_worst	concavity_worst
0.023	0.157	0.209
concave.points_worst	symmetry_worst	fractal_dimension_worst

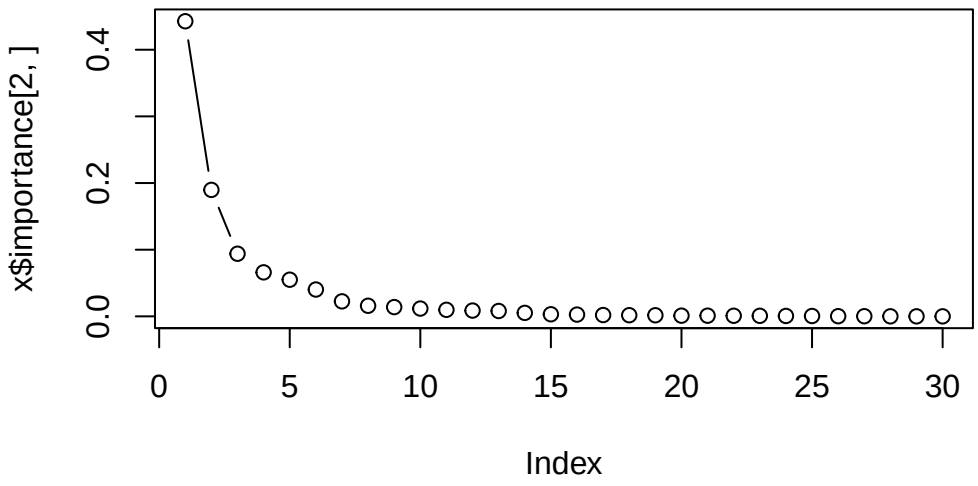
0.066 0.062 0.018

Perform PCA on wisc.data with scaling.

```
wisc.pr <- prcomp(wisc.data, scale=TRUE)
x <- summary(wisc.pr)
x$importance
```

```
                           PC1      PC2      PC3      PC4      PC5      PC6
Standard deviation     3.644394 2.385656 1.678675 1.407352 1.284029 1.098798
Proportion of Variance 0.442720 0.189710 0.093930 0.066020 0.054960 0.040250
Cumulative Proportion  0.442720 0.632430 0.726360 0.792390 0.847340 0.887590
                           PC7       PC8       PC9      PC10      PC11
Standard deviation     0.8217178 0.6903746 0.6456739 0.5921938 0.5421399
Proportion of Variance 0.0225100 0.0158900 0.0139000 0.0116900 0.0098000
Cumulative Proportion  0.9101000 0.9259800 0.9398800 0.9515700 0.9613700
                          PC12      PC13      PC14      PC15      PC16
Standard deviation     0.5110395 0.4912815 0.3962445 0.3068142 0.2826001
Proportion of Variance 0.0087100 0.0080500 0.0052300 0.0031400 0.0026600
Cumulative Proportion  0.9700700 0.9781200 0.9833500 0.9864900 0.9891500
                          PC17      PC18      PC19      PC20      PC21
Standard deviation     0.2437192 0.2293878 0.2224356 0.1765203 0.1731268
Proportion of Variance 0.0019800 0.0017500 0.0016500 0.0010400 0.0010000
Cumulative Proportion  0.9911300 0.9928800 0.9945300 0.9955700 0.9965700
                          PC22      PC23      PC24      PC25      PC26
Standard deviation     0.1656484 0.1560155 0.1343689 0.1244238 0.0904303
Proportion of Variance 0.0009100 0.0008100 0.0006000 0.0005200 0.0002700
Cumulative Proportion  0.9974900 0.9983000 0.9989000 0.9994200 0.9996900
                           PC27       PC28        PC29        PC30
Standard deviation     0.08306903 0.0398665 0.02736427 0.01153451
Proportion of Variance 0.00023000 0.0000500 0.00002000 0.00000000
Cumulative Proportion  0.99992000 0.9999700 1.00000000 1.00000000
```

```
#Plot the proportion of variance accounted for by each PC
plot(x$importance[2,], typ='b')
```



Q4: What proportion of the original variance is captured by the first PC?

About 44% of the variance is captured by PC1.

Q5: How many PCs are required to describe at least 70% of the original variance?

Three principal components are required to describe at least 70% of the variance (PC3 cumulative proportion is ~72.64%).

Q6: How many PCs are required to describe at least 90% of the original variance?

Seven prinicpal components are required to describe at least 90% of the variance (PC7 cumulative proportion is ~91%).

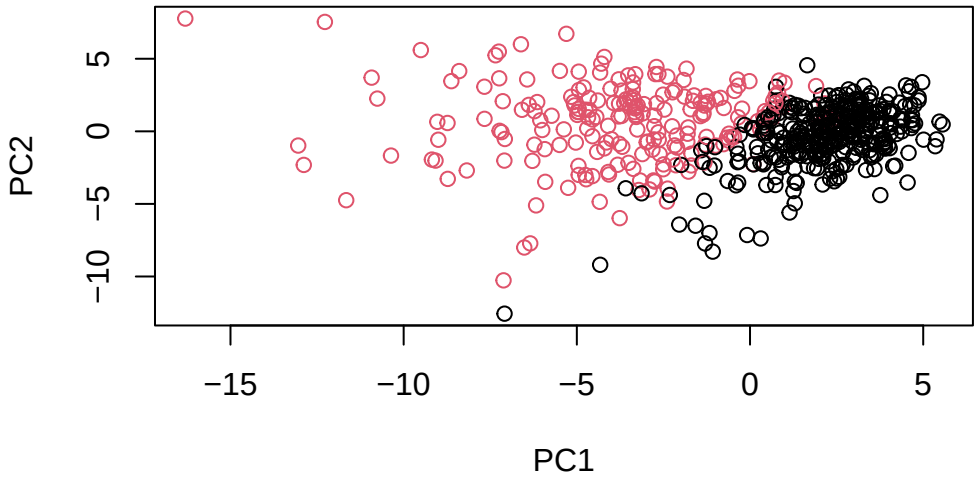
```
#biplot(wisc.pr)
```

Q7: What stands out to you about this biplot?

This plot is not understandable at all and is much too crowded to make any conclusions.

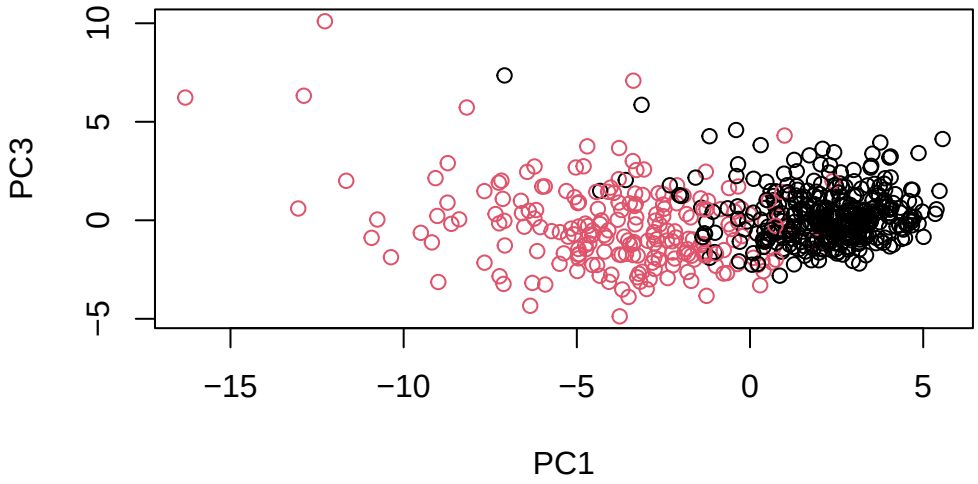
Plot the observations by PC1 and PC2:

```
#attributes(wisc.pr)
#head(wisc.pr)
plot(wisc.pr$x, col=diagnosis,
     xlab="PC1", ylab="PC2")
```



Plot the observations by PC1 and PC3:

```
plot(wisc.pr$x[,1], wisc.pr$x[,3], col=diagnosis,
     xlab="PC1", ylab="PC3")
```



Q8: What do you notice about the two previous plots?

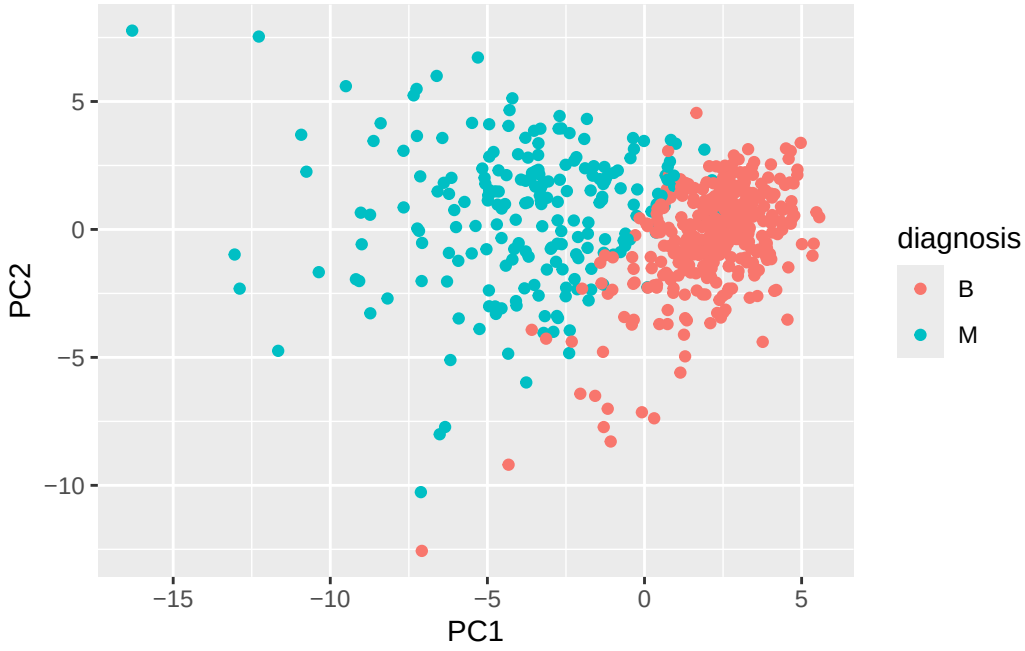
There is a cleaner more clear separation between clusters in the PC1 vs PC2 plot than the PC1 vs PC3 plot, indicating that PC2 explains more variance than PC3.

Use ggplot2 to make some nicer figures.

```
df <- as.data.frame(wisc.pr$x)
df$diagnosis <- diagnosis

library(ggplot2)

ggplot(df) +
  aes(PC1, PC2, col=diagnosis) +
  geom_point()
```



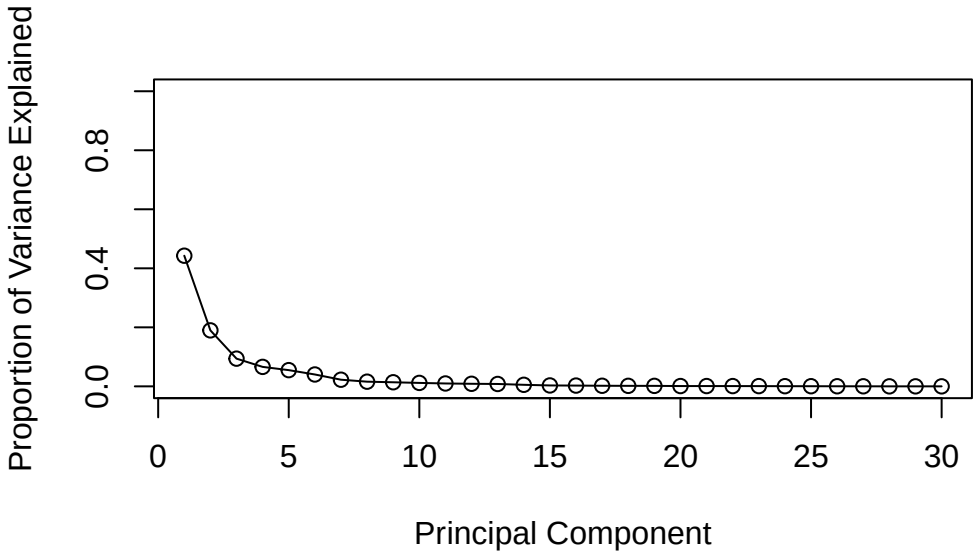
Variance explained

```
#Variance of each component
pr.var <- wisc.pr$sdev^2
head(pr.var)
```

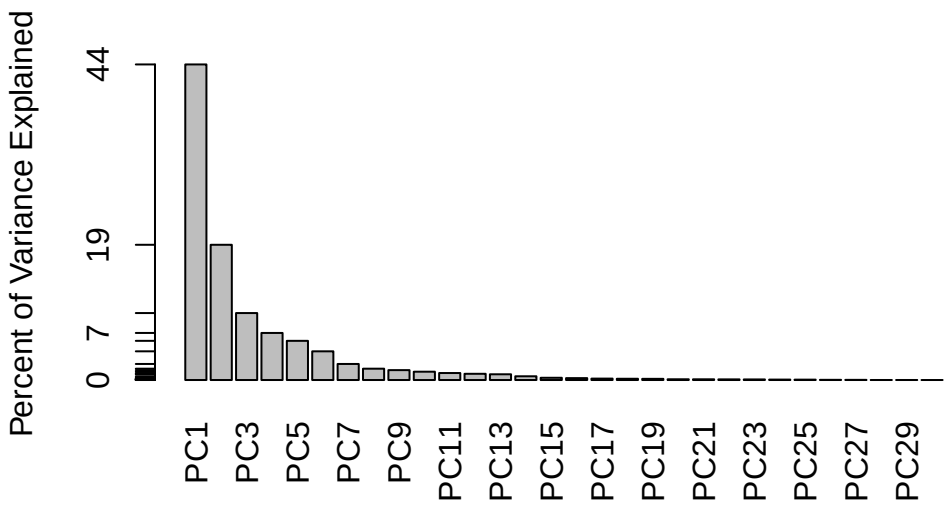
```
[1] 13.281608  5.691355  2.817949  1.980640  1.648731  1.207357
```

```
#Variance explained by each PC
pve <- pr.var/sum(pr.var)

plot(pve, xlab="Principal Component",
     ylab="Proportion of Variance Explained",
     ylim=c(0,1), type="o")
```




```
#Scree plot of the same data but in a bar plot with labels for each PC
barplot(pve, ylab="Percent of Variance Explained",
        names.arg=paste0("PC", 1:length(pve)), las=2, axes=FALSE)
axis(2, at=pve, labels=round(pve,2)*100) #Creates y-axis bar label showing
```

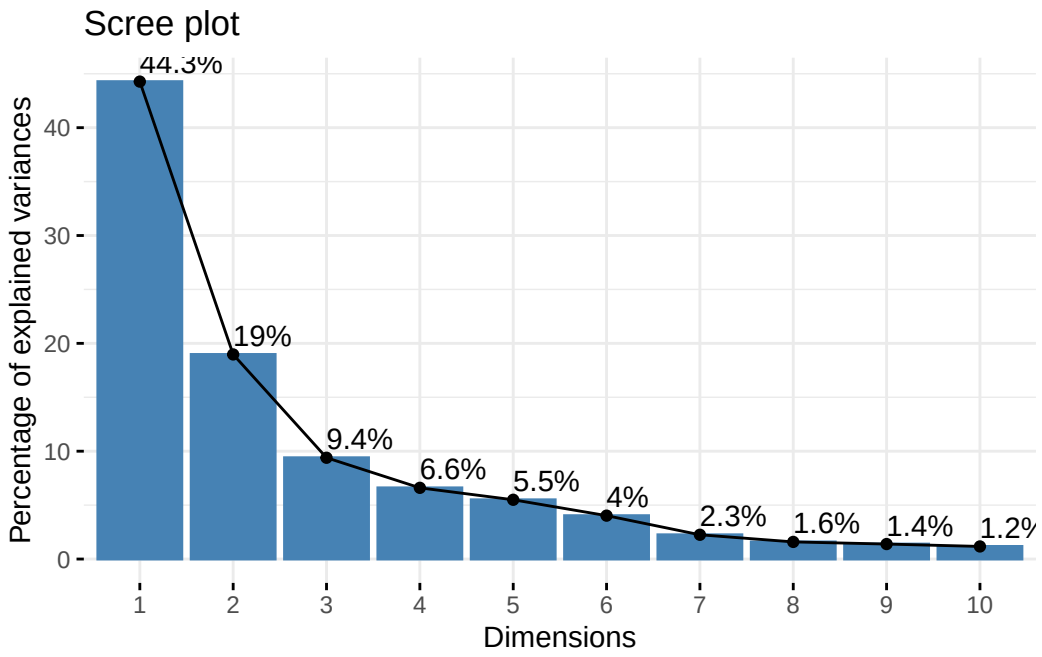



```
                                    #percentages rather than proportion


#ggplot based graph instead of base R bar plot
#install.packages("factoextra")
library(factoextra)
```

```
Welcome! Want to learn more? See two factoextra-related books at https://goo.gl/ve3WBa
```

```
fviz_eig(wisc.pr, addlabels = TRUE)
```



Communicating PCA results

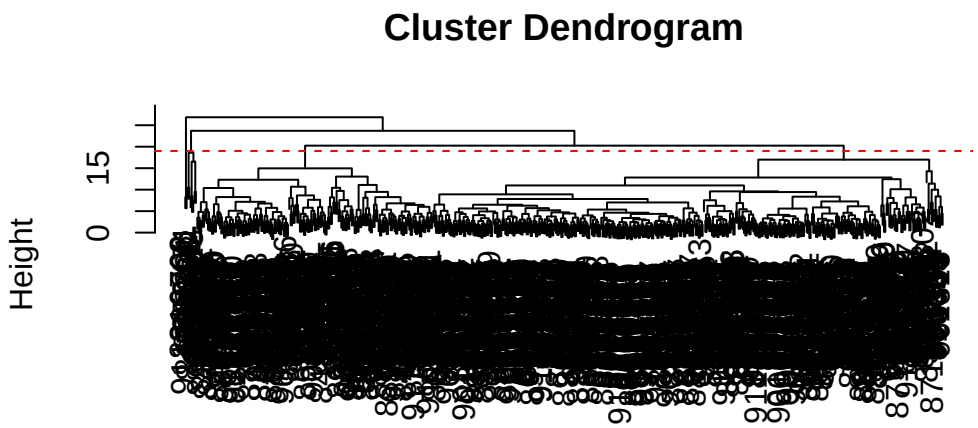
Q9: For the first PC, what is the component of the loading vector for the feature 'concave.points_mean'? This tells us how much this original feature contributes to the first PC.

```
wisc.pr$rotation["concave.points_mean",1]
```

```
[1] -0.2608538
```

Hierarchical clustering

```
data.scaled <- scale(wisc.data)
data.dist <- dist(data.scaled)
wisc.hclust <- hclust(data.dist)
plot(wisc.hclust)
abline(a=19, b=0, col="red", lty=2)
```



Q10: What's the height where this model has 4 clusters?

19

```
wisc.hclust.clusters <- cutree(wisc.hclust, k=4)
table(wisc.hclust.clusters, diagnosis)
```

```
                    diagnosis
wisc.hclust.clusters   B   M
                   1  12 165
                   2   2   5
                   3 343  40
                   4   0   2
```

Explore different methods to combine points in hierarchical clustering.

```
wisc.hclust.single <- hclust(data.dist, method="single")
wisc.hclust.cluster.single <- cutree(wisc.hclust.single, k=4)
table(wisc.hclust.cluster.single, diagnosis)
```

```
                          diagnosis
wisc.hclust.cluster.single   B   M
                         1 356 209
                         2   1   0
                         3   0   2
                         4   0   1
```

```
wisc.hclust.complete <- hclust(data.dist, method="complete")
wisc.hclust.cluster.complete <- cutree(wisc.hclust.complete, k=4)
table(wisc.hclust.cluster.complete, diagnosis)
```

```
                           diagnosis
wisc.hclust.cluster.complete   B   M
                           1  12 165
                           2   2   5
                           3 343  40
                           4   0   2
```

```
wisc.hclust.average <- hclust(data.dist, method="average")
wisc.hclust.cluster.average <- cutree(wisc.hclust.average, k=4)
table(wisc.hclust.cluster.average, diagnosis)
```

```
                           diagnosis
wisc.hclust.cluster.average   B   M
                           1 355 209
                           2   2   0
                           3   0   1
                           4   0   2
```

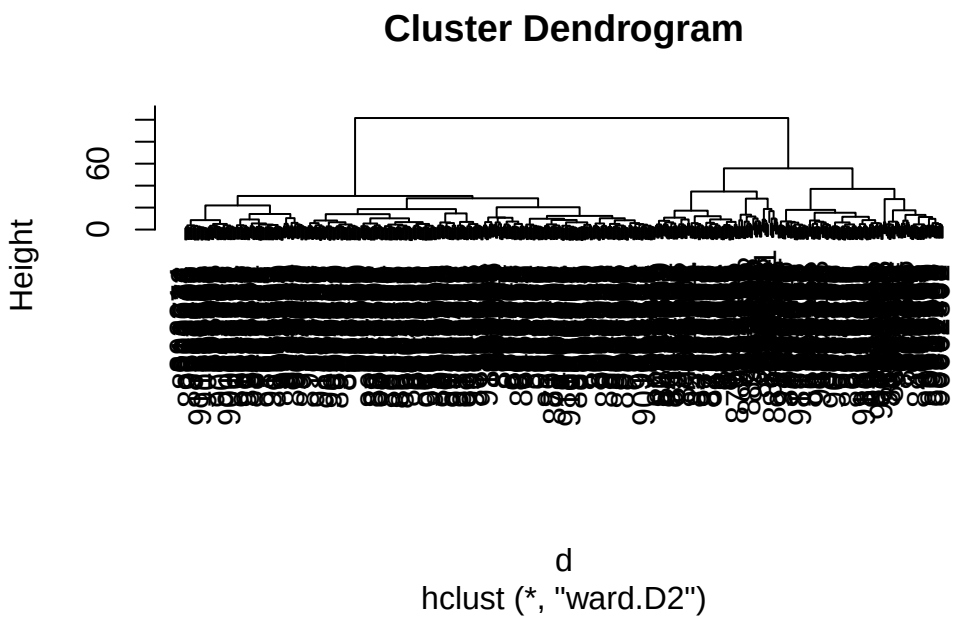
```
wisc.hclust.ward.D2 <- hclust(data.dist, method="ward.D2")
wisc.hclust.cluster.ward.D2 <- cutree(wisc.hclust.ward.D2, k=4)
table(wisc.hclust.cluster.ward.D2, diagnosis)
```

```
                           diagnosis
wisc.hclust.cluster.ward.D2   B   M
                           1   0 115
                           2   6  48
                           3 337  48
                           4  14   1
```

Q12: The "complete" method seems to give the best results compared to the expert diagnoses, giving less false positive/negative results but it is unclear whether this is a good thing or just fitting the data to match the experts conclusions.

Now use PCA results to cluster, using the "ward.D2" method

```
d<- dist(wisc.pr$x[,1:7])
hc <- hclust(d, method="ward.D2")
plot(hc)
```



Cut the tree to yield 2 clusters

```
grps <- cutree(hc, k=2)
table(grps)
```

```
grps
  1   2
216 353
```

Compare to the expert diagnoses of "M" vs "B"

```
table(diagnosis, grps)
```

```
         grps
diagnosis   1   2
        B  28 329
        M 188  24
```

Q13: How well does this model separate out the two diagnoses?

This separates out the two diagnoses fairly well, but it is saying there are 28 false benign and 24 false malignant diagnoses from the analysis done here.

```
table(diagnosis, wisc.hclust.clusters)
```

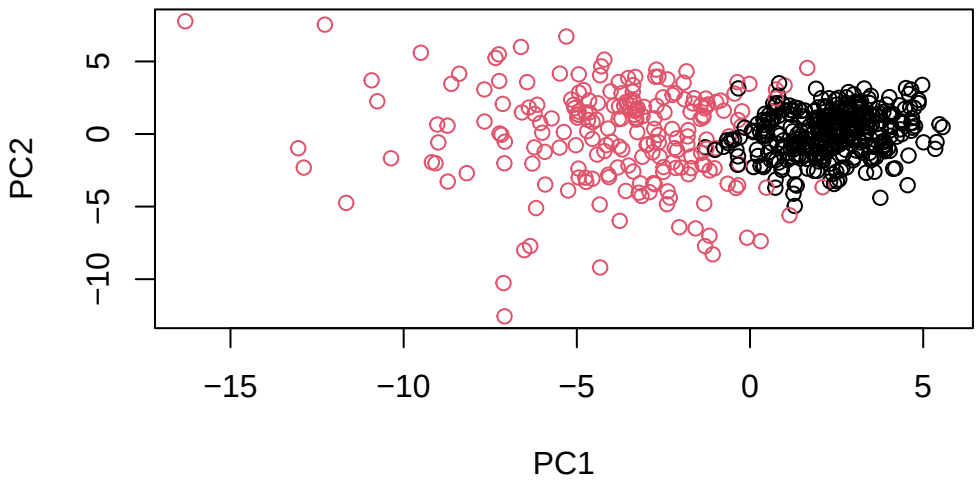
```
         wisc.hclust.clusters
diagnosis   1   2   3   4
        B  12   2 343   0
        M 165   5  40   2
```

Q14: How well does the earlier hierarchical clustering model (before PCA) do in terms of separating out the diagnoses?

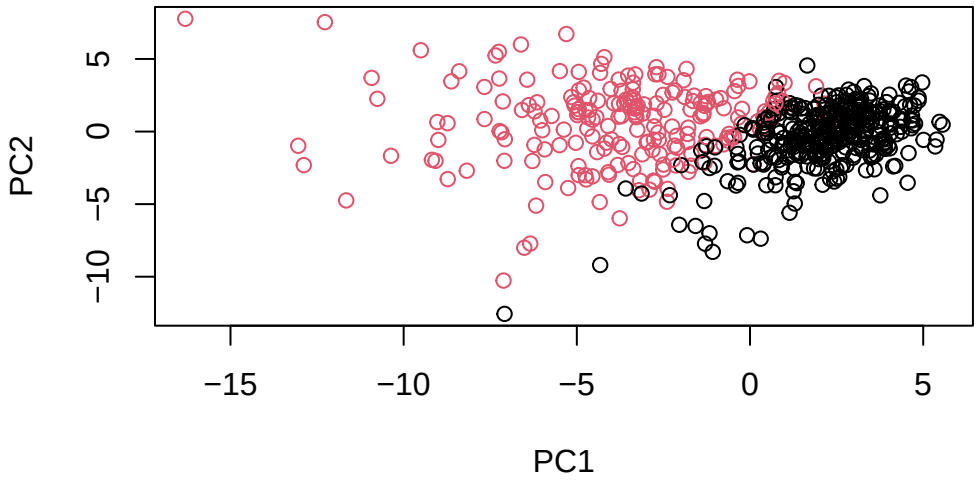
This model results in less false benign results compared to the expert diagnoses results. However, there are much more false malignant results in this model than in the PCA-based clustering model.

```
#plot(wisc.pr$x[,1:2], col=grps)

g <- as.factor(grps)
#levels(g)
g <- relevel(g,2)
#levels(g)
plot(wisc.pr$x[,1:2], col=g)
```



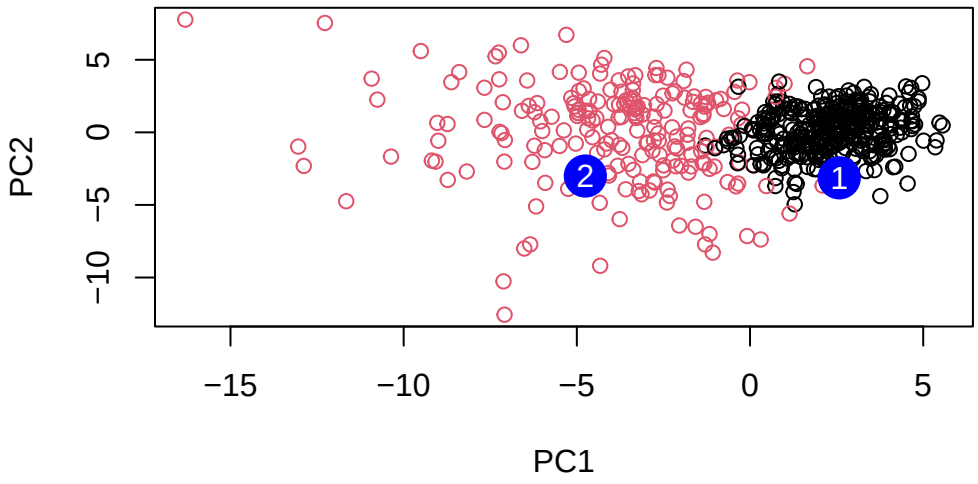

```
plot(wisc.pr$x[,1:2], col=diagnosis)
```



Prediction

```
url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
#npc

plot(wisc.pr$x[,1:2], col=g)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q16: Which of these new patient groups should we prioritize for follow up based on your results?

Since the numeral 1 was assigned to M (malignant) and the numeral 2 was assigned to B (benign) in this analysis, the patients in cluster 1 should be prioritized for follow-up since they have been both clustered into a malignant cluster by bioinformatics analysis and diagnosed by an expert as having malignant tissue.

```r
loadings <- wisc.pr$rotation

ggplot(loadings) +
  aes(abs(PC1), reorder(rownames(loadings), -PC1)) +
  geom_col()
```