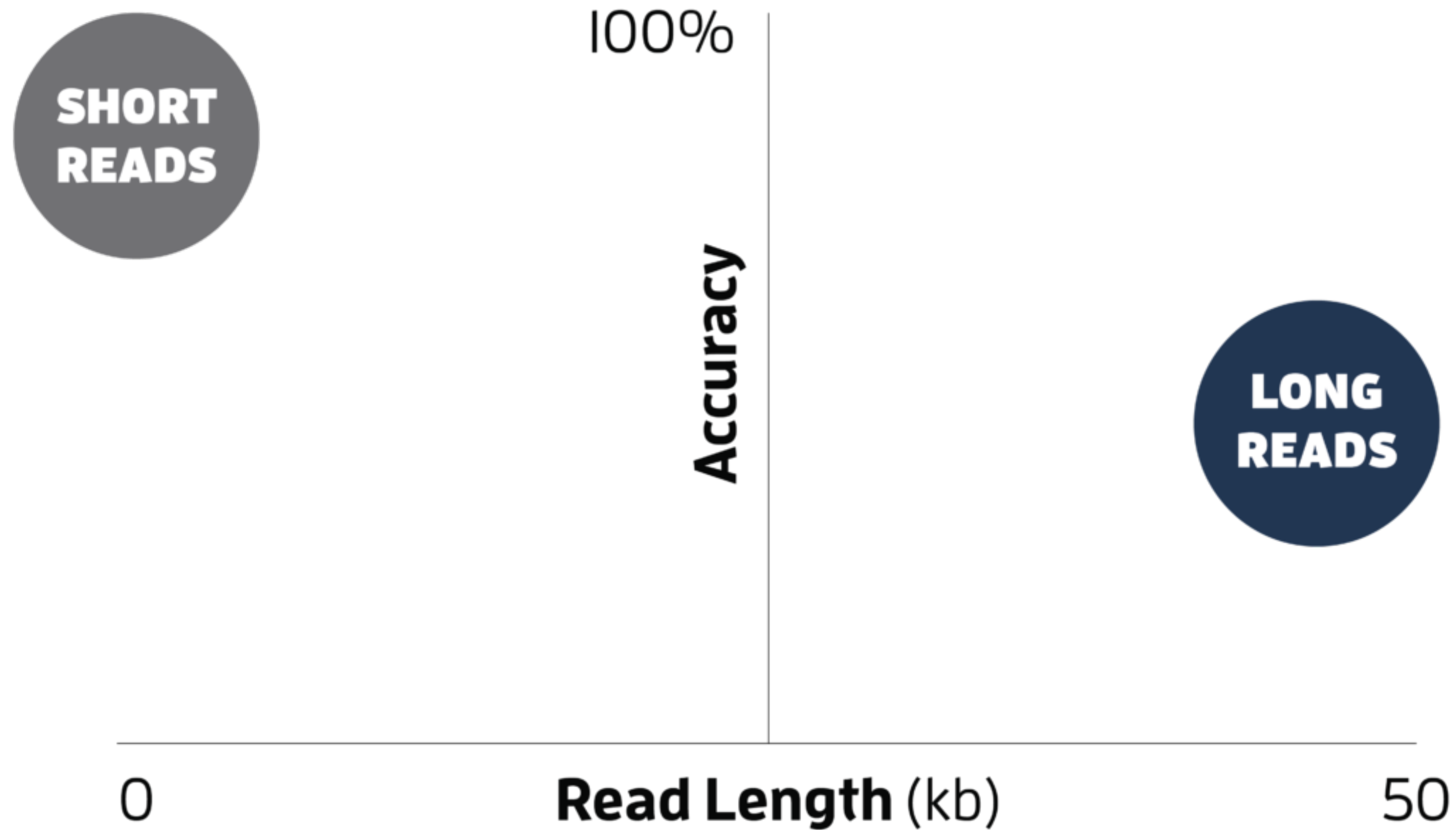
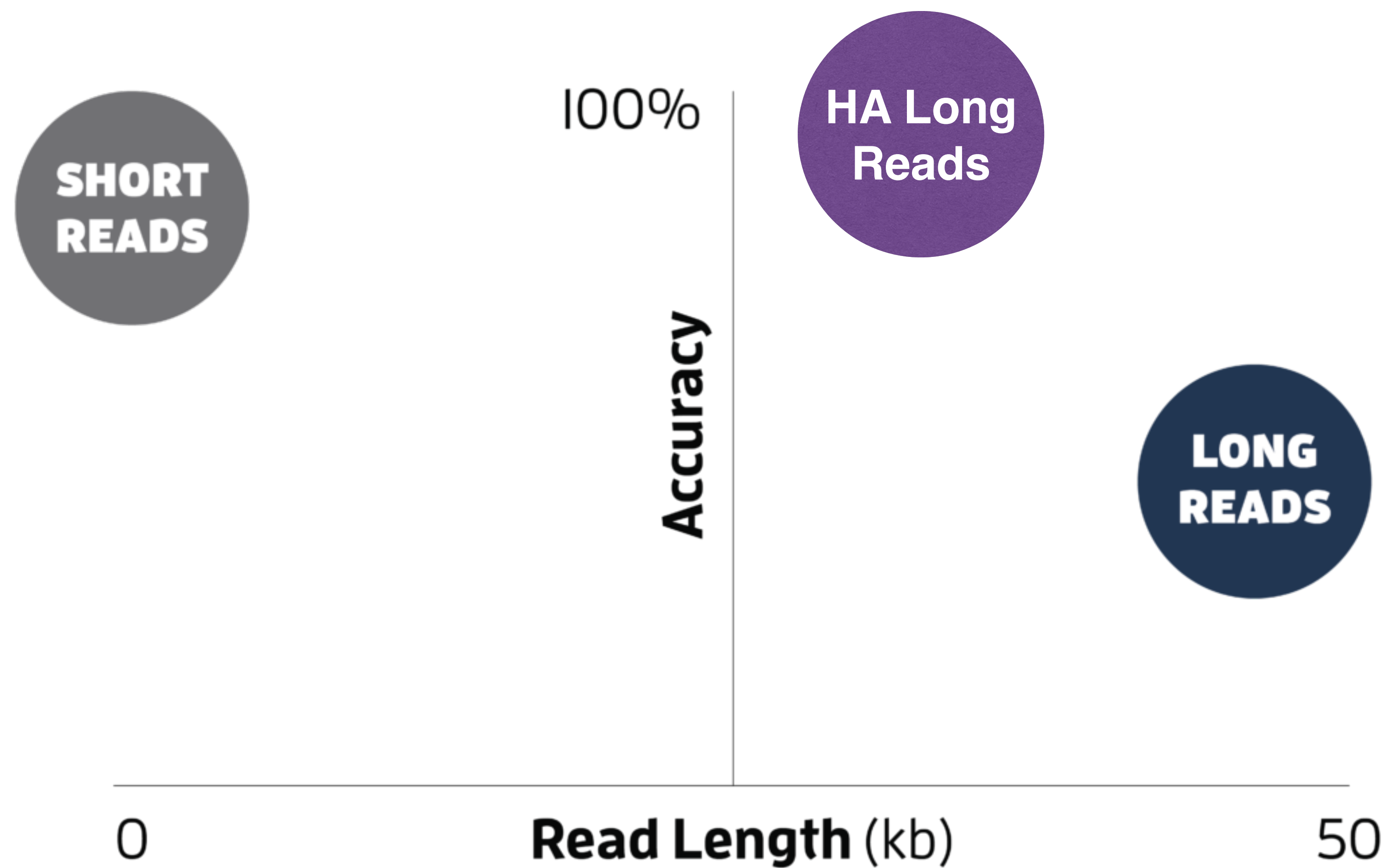


Maximizing Resolution with DADA2 and Long-read Amplicon Sequencing

Highly-Accurate Long Reads



Highly-Accurate Long Reads



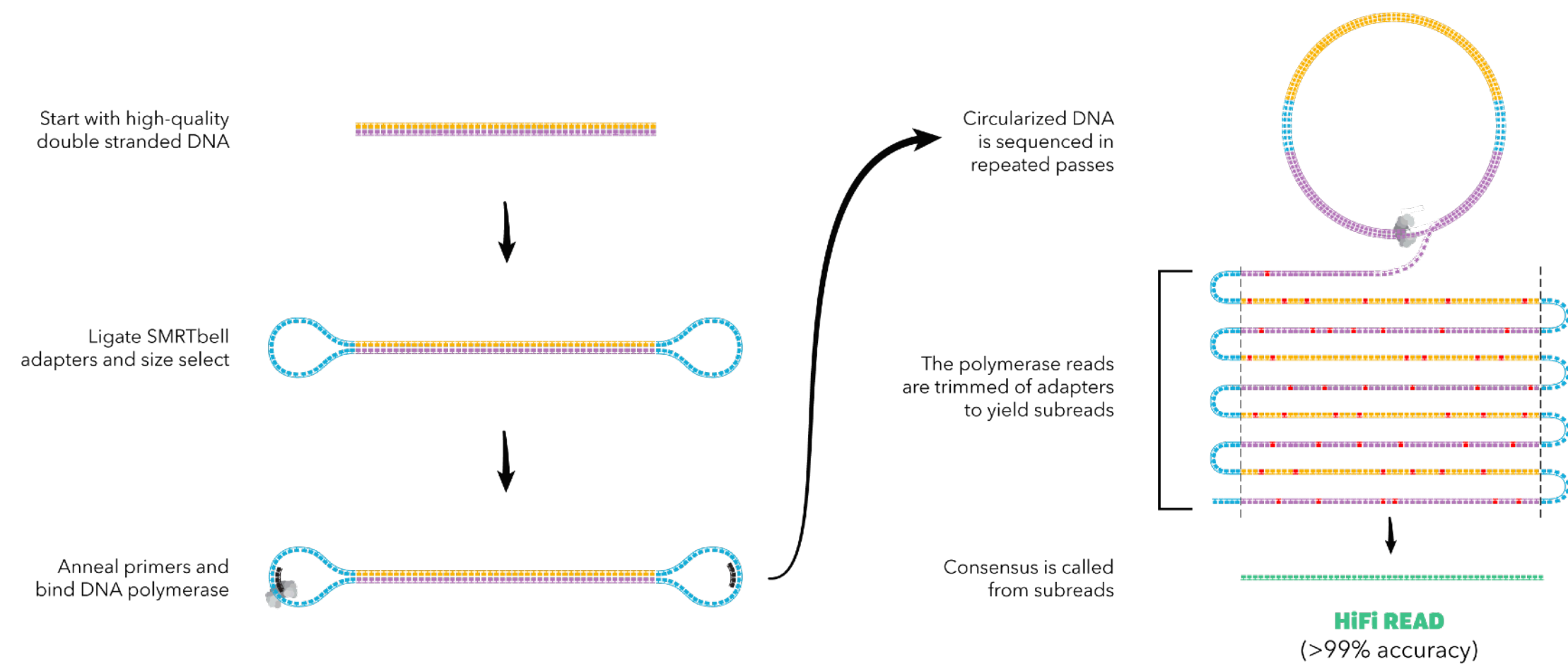
Highly-Accurate Long Reads

Strategy: Build Consensus from Multiple Reads of a Long DNA Molecule

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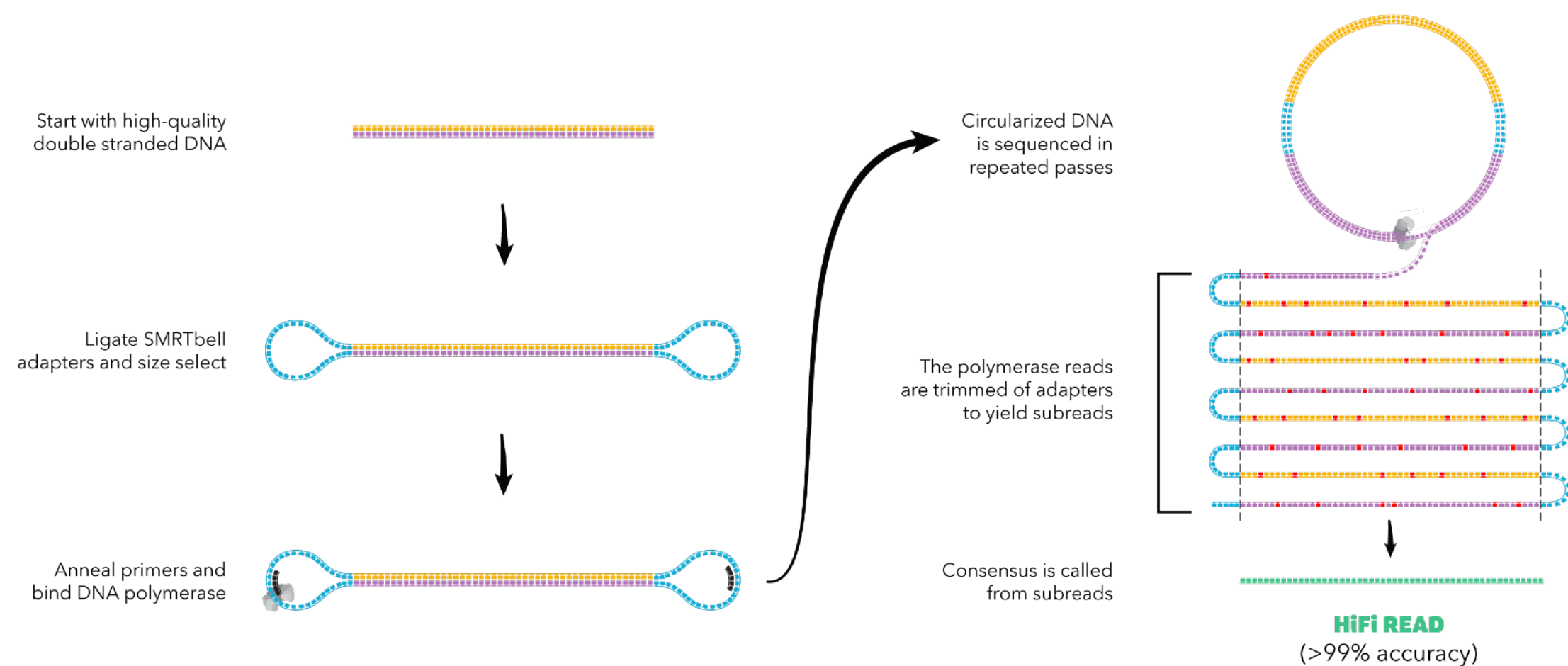
PacBio HiFi Sequencing



Highly-Accurate Long Reads

Strategy: Build Consensus from Multiple Reads of a Long DNA Molecule

PacBio HiFi Sequencing

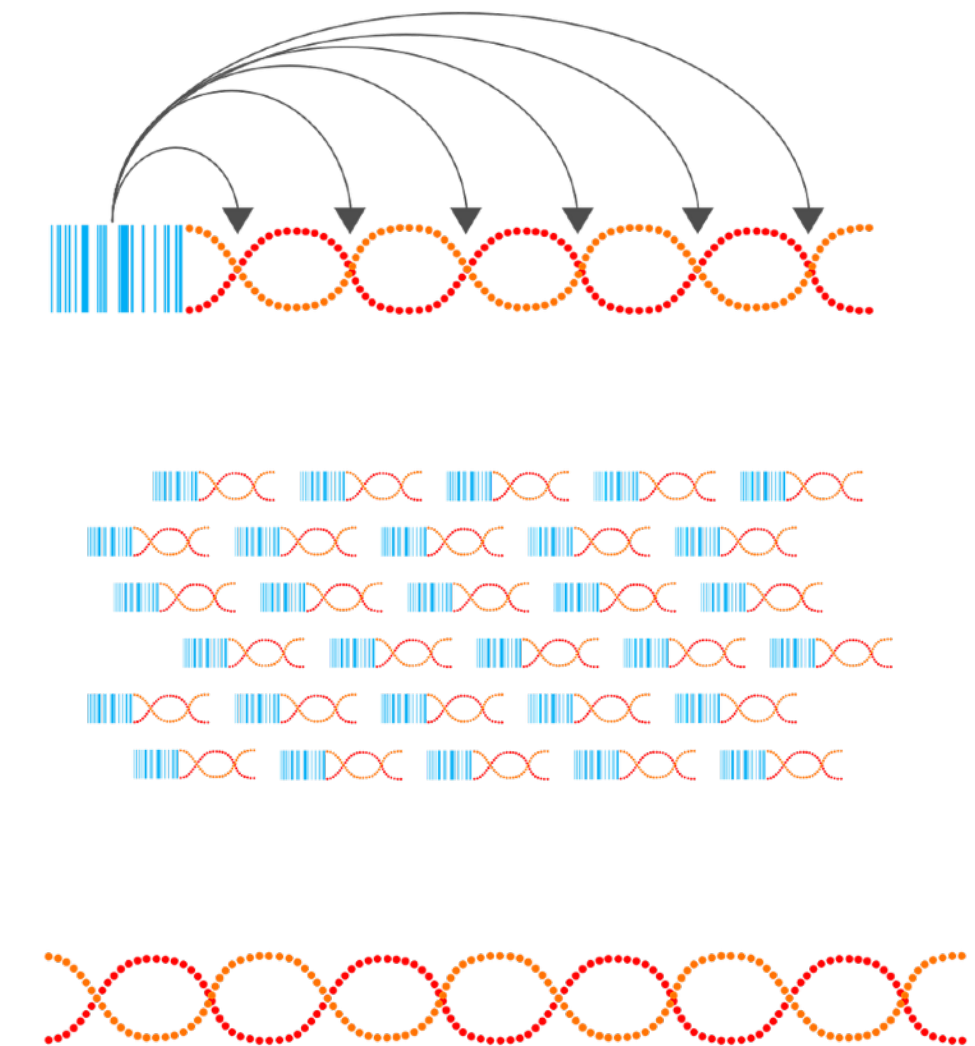


Element Biosciences Synthetic Long Reads (SLRs)

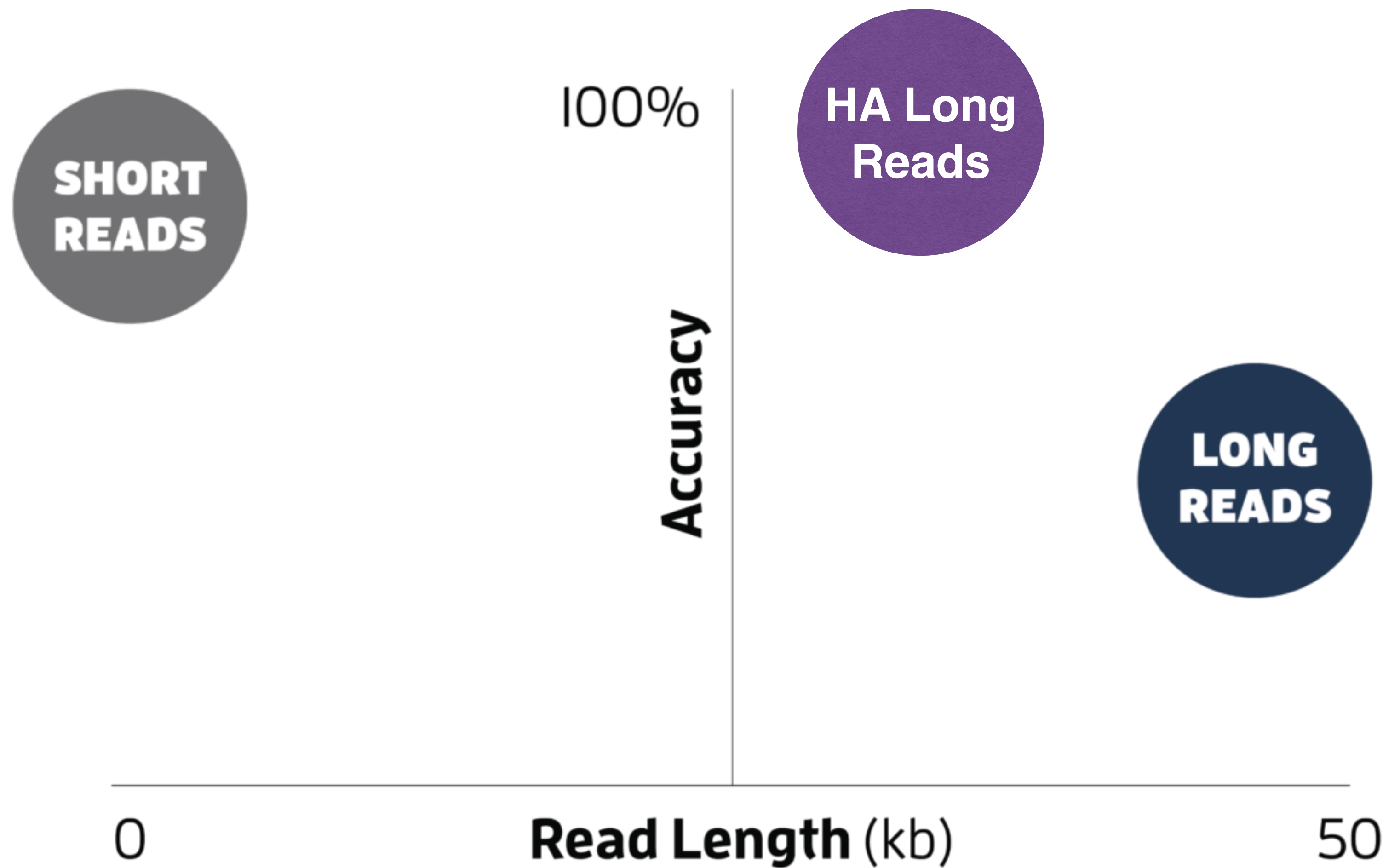
Distribute UMI throughout long DNA molecule

Short-read sequencing

Consensus assembly



Highly-Accurate Long Reads



Short Reads (Illumina)

Read length: **~250 nts**

Per-base error-rate: **0.1 - 0.5%**

Long Reads (PacBio, Oxford)

Read length: **1 - 200 kilobases**

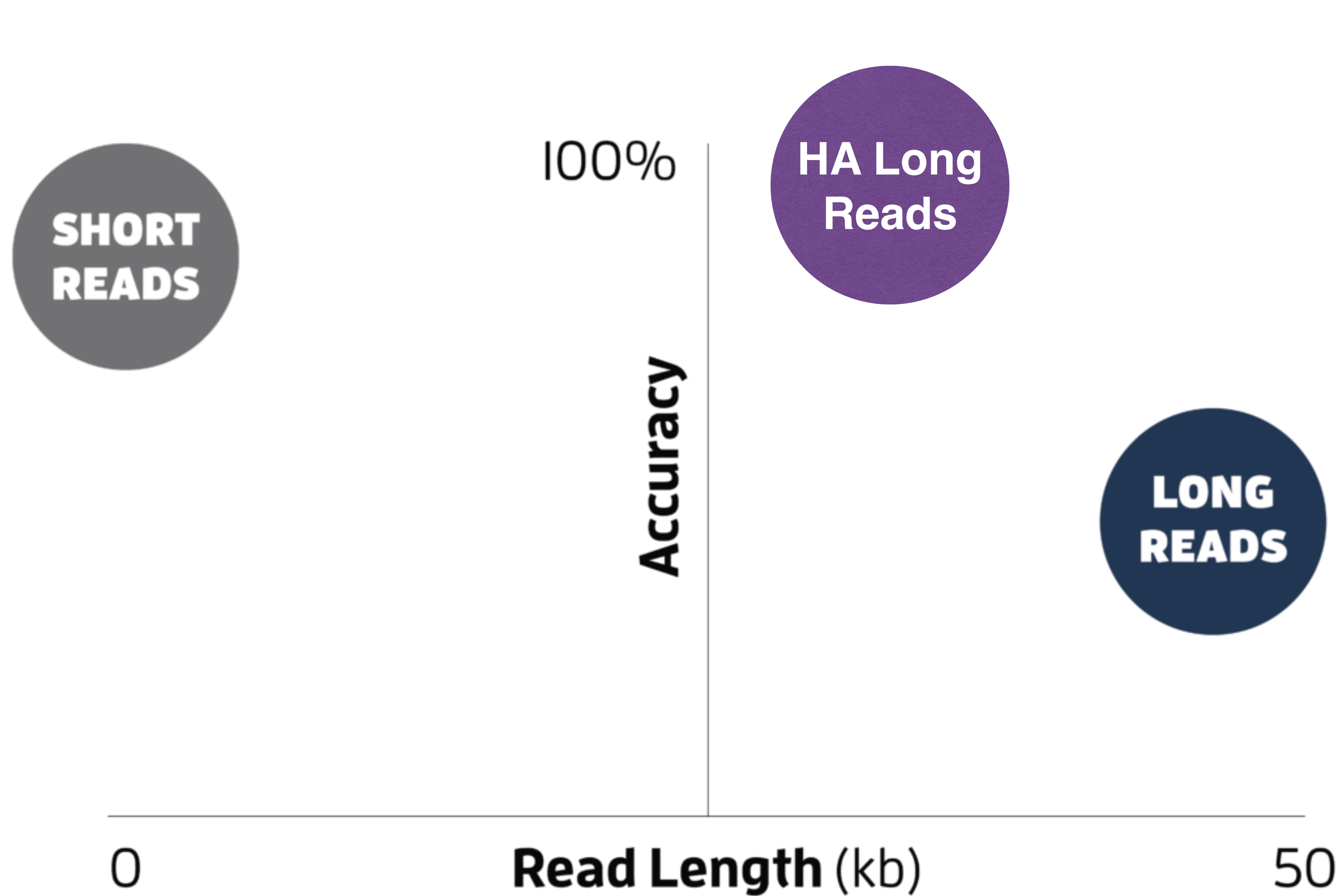
Per-base error-rate: **2 - 10%**

HA Long Reads (HiFi, LoopSeq)

Read length: **1 - 20 kilobases**

Per-base error-rate: **< 0.1%**

Highly-Accurate Long Reads



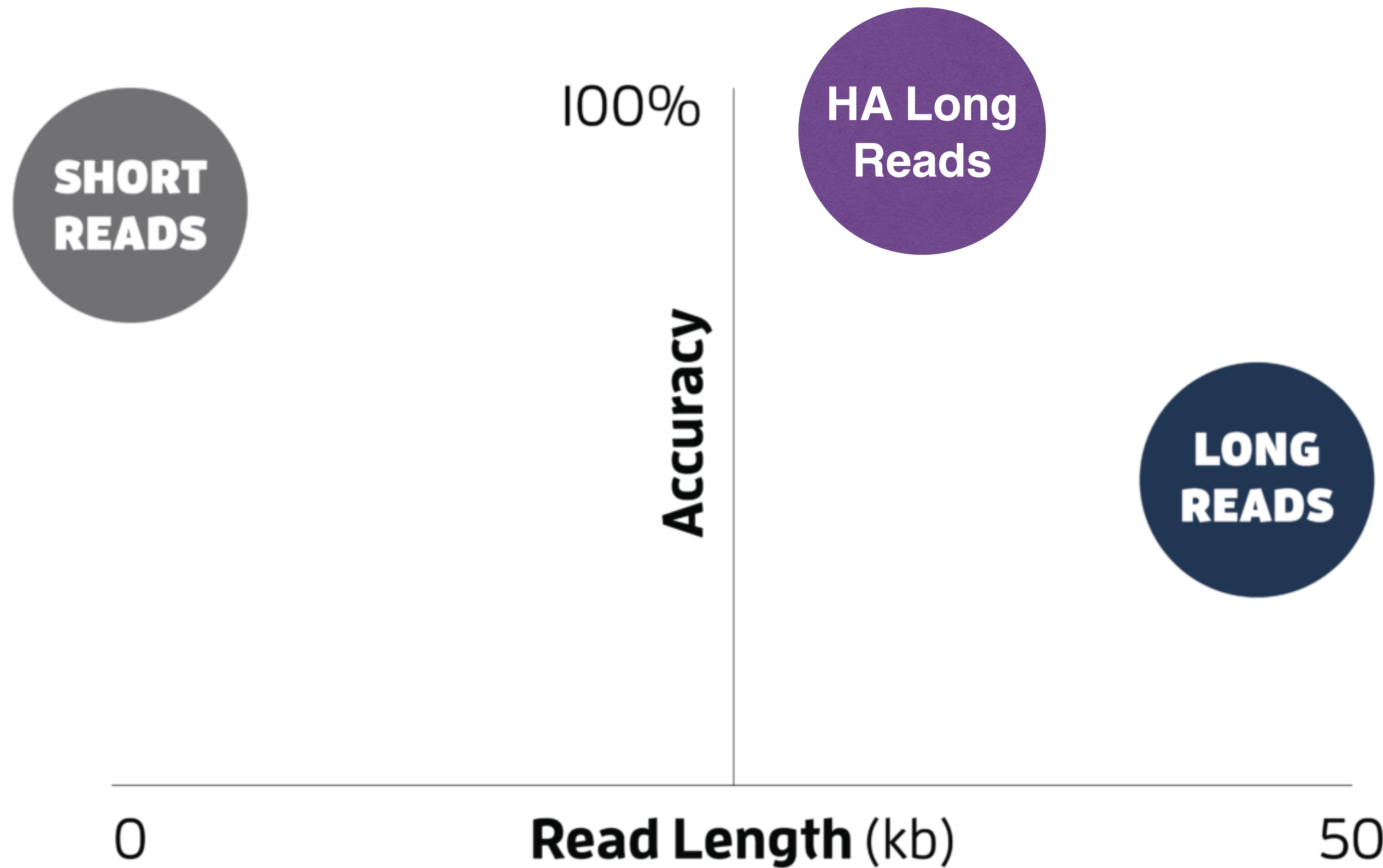
Highly-accurate long reads are effectively a new sequencing technology.

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Per-base error-rate: **< 0.1%**

Highly-Accurate Long Reads



Highly-accurate long reads are effectively a new sequencing technology.
But do they really work for amplicon sequencing?

HA Long Reads (HiFi, LoopSeq)

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Zymo Mock Community

8 Bacterial Strains at
nominally equal concentrations

- *Pseudomonas aeruginosa*
- *Escherichia coli*
- *Salmonella enterica*
- *Lactobacillus fermentum*
- *Enterococcus faecalis*
- *Staphylococcus aureus*
- *Listeria monocytogenes*
- *Bacillus subtilis*

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**PacBio
HiFi +
DADA2**

Full-Length, Full-Complement 16S

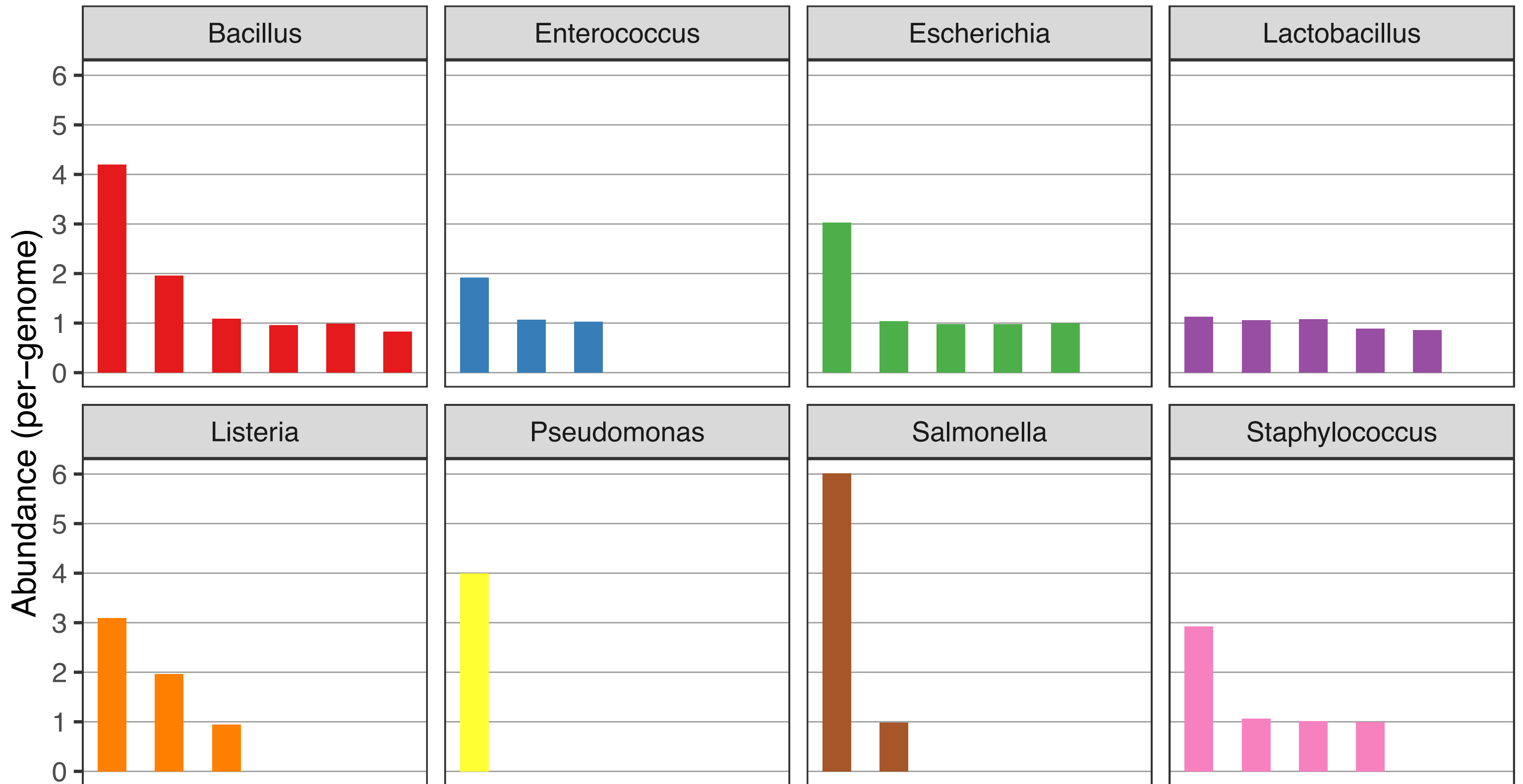
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**PacBio
HiFi +
DADA2**



Full-length 16S Sequence Variants

Full-Length, Full-Complement 16S

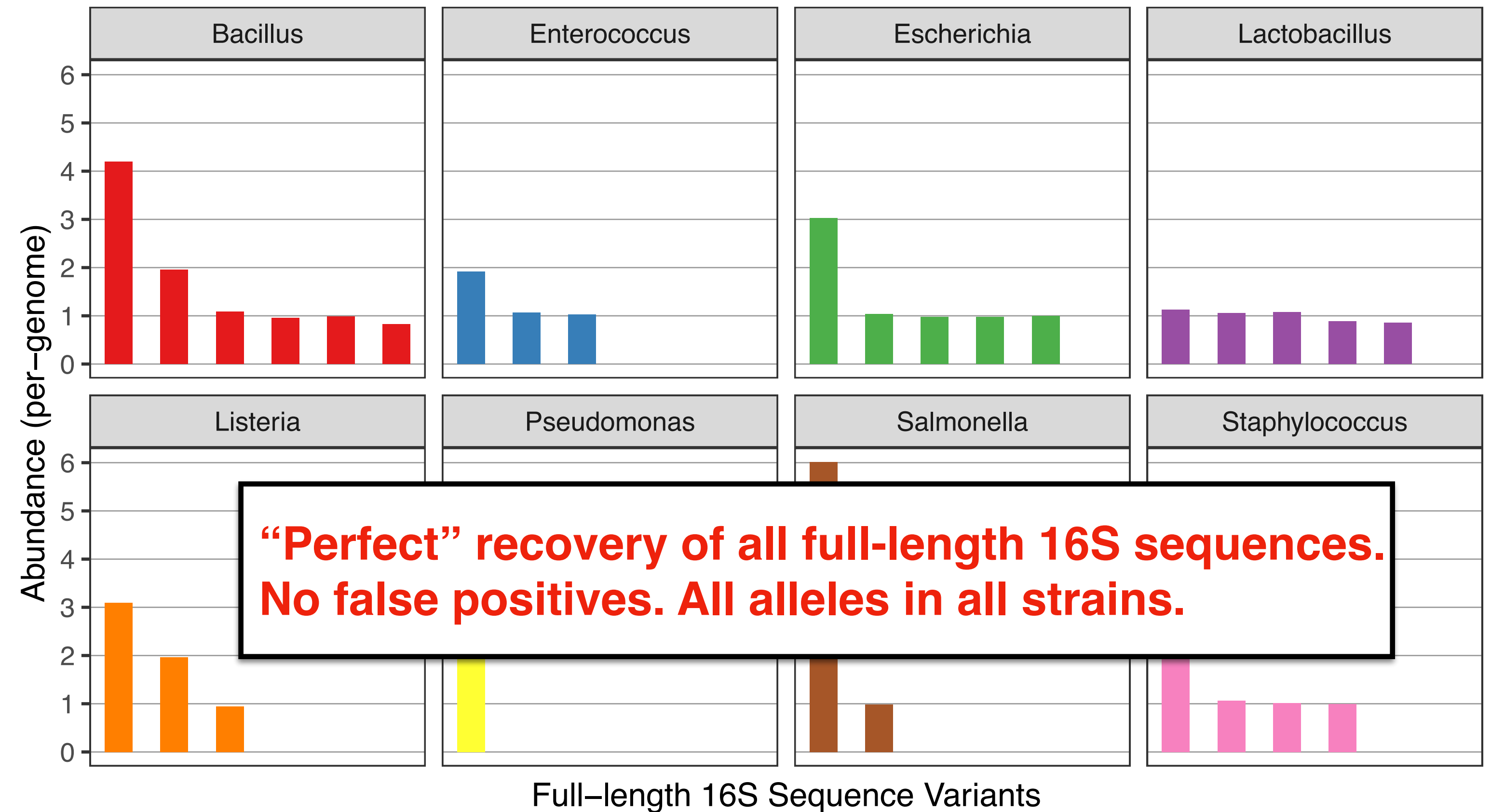
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**PacBio
HiFi +
DADA2**



Full-Length, Full-Complement 16S

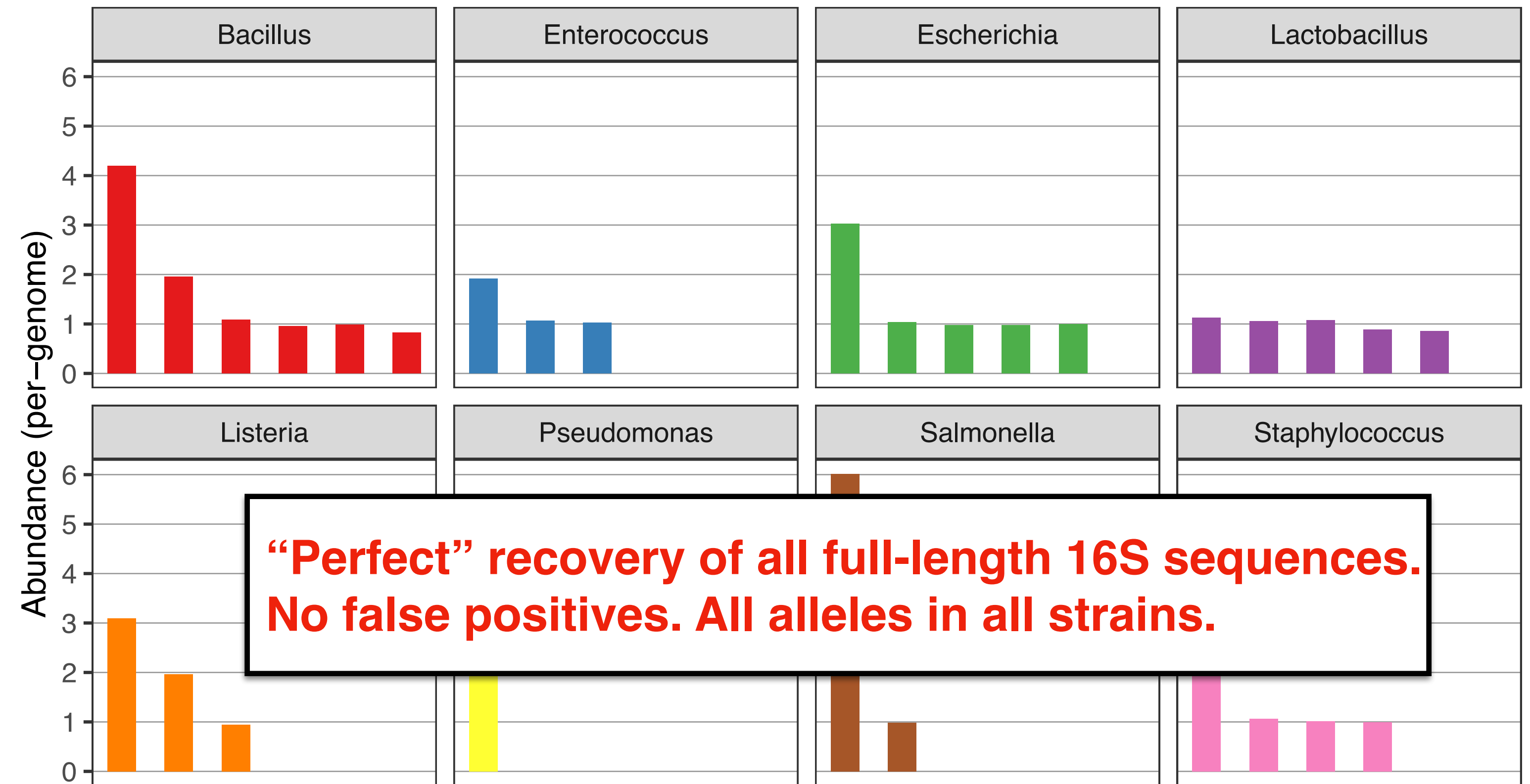
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**PacBio
HiFi +
DADA2***




**“Perfect” recovery of all full-length 16S sequences.
No false positives. All alleles in all strains.**

Full-length 16S Sequence Variants

*** Modestly modified workflow for long-read amplicon sequencing.**

PacBio HiFi

High-throughput amplicon sequencing of the full-length 16S rRNA gene with single-nucleotide resolution

Benjamin J Callahan , Joan Wong, Cheryl Heiner, Steve Oh, Casey M Theriot, Ajay S Gulati, Sarah K McGill, Michael K Dougherty

Nucleic Acids Research, Volume 47, Issue 18, 10 October 2019, Page e103,

<https://doi.org/10.1093/nar/gkz569>

Published: 03 July 2019 **Article history** ▼

LoopSeq

Ultra-accurate microbial amplicon sequencing with synthetic long reads


[Benjamin J. Callahan](#) , [Dmitry Grinevich](#), [Siddhartha Thakur](#), [Michael A. Balamotis](#) & [Tuval Ben Yehezkel](#)

[Microbiome](#) **9**, Article number: 130 (2021) | [Cite this article](#)

5717 Accesses | **11** Citations | **44** Altmetric | [Metrics](#)

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**Highly-accurate long reads
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sequencing technology.
But do they really
work for amplicon sequencing?**

Yes!
Multiple technologies
Out to 6+ kilo bases
>99.9% accuracy



We support long-read amplicon sequencing!



We support long-read amplicon sequencing!

Fastq



**ASV Table or
Species Table (16S)**



Fastq
↓
ASV Table or
Species Table (16S)

We support long-read amplicon sequencing!

Long-read specific documentation

PacBio HiFi: <https://github.com/benjjneb/LRASmanuscript>

LoopSeq: <https://github.com/benjjneb/LoopManuscript>

Sequencing instrument



The graphic features a central image of the Revio sequencing instrument, a large black rectangular unit with a PacBio logo in the top left corner. The background is a gradient of blue and purple. Text is overlaid on the image in white. At the bottom, there are four columns, each with an icon, a title, and a description.

Revio system
Reveal more with accurate long-read sequencing at scale

Scale	Ease of use	Compute power	Affordability
2,500 human HiFi genomes per year	Only 500 ng DNA input and flexible run setup	Google DeepConsensus and more on board	~\$500 human HiFi genome

- Higher throughput
- Lower costs
- Binned quality scores

Sequencing instrument



Revio system
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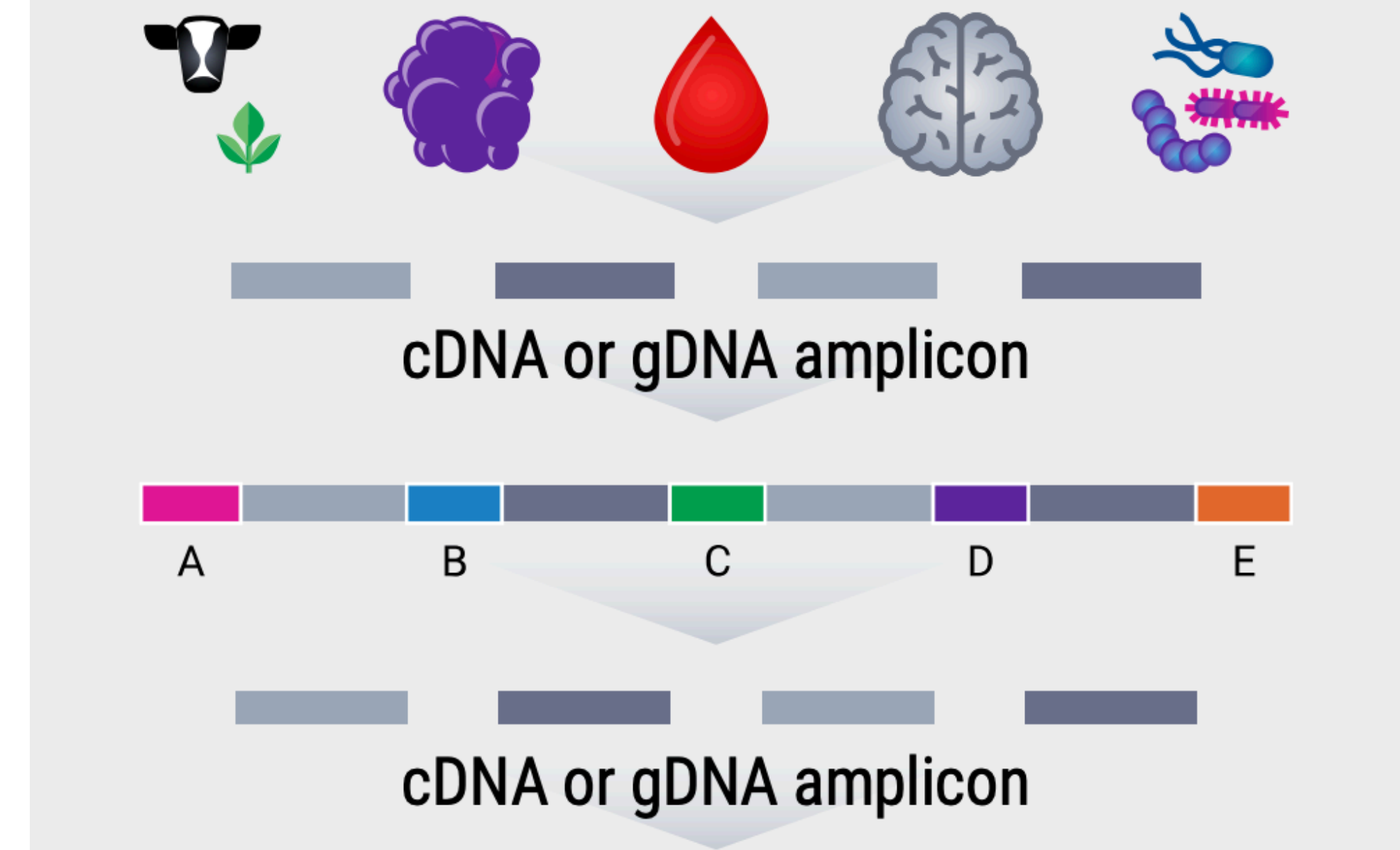
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Library preparation

How Kinnex works



- Higher throughput
- Lower costs
- Lower quality (16S)

Sequencing instrument

Revio system
Reveal more with accurate long-read sequencing at scale



The Revio system is a large, industrial-grade sequencing instrument with a dark, metallic finish and a prominent PacBio logo in the top left corner. It features a large, illuminated display area showing the system name and its capabilities.

Scale
2,500 human HiFi genomes per year

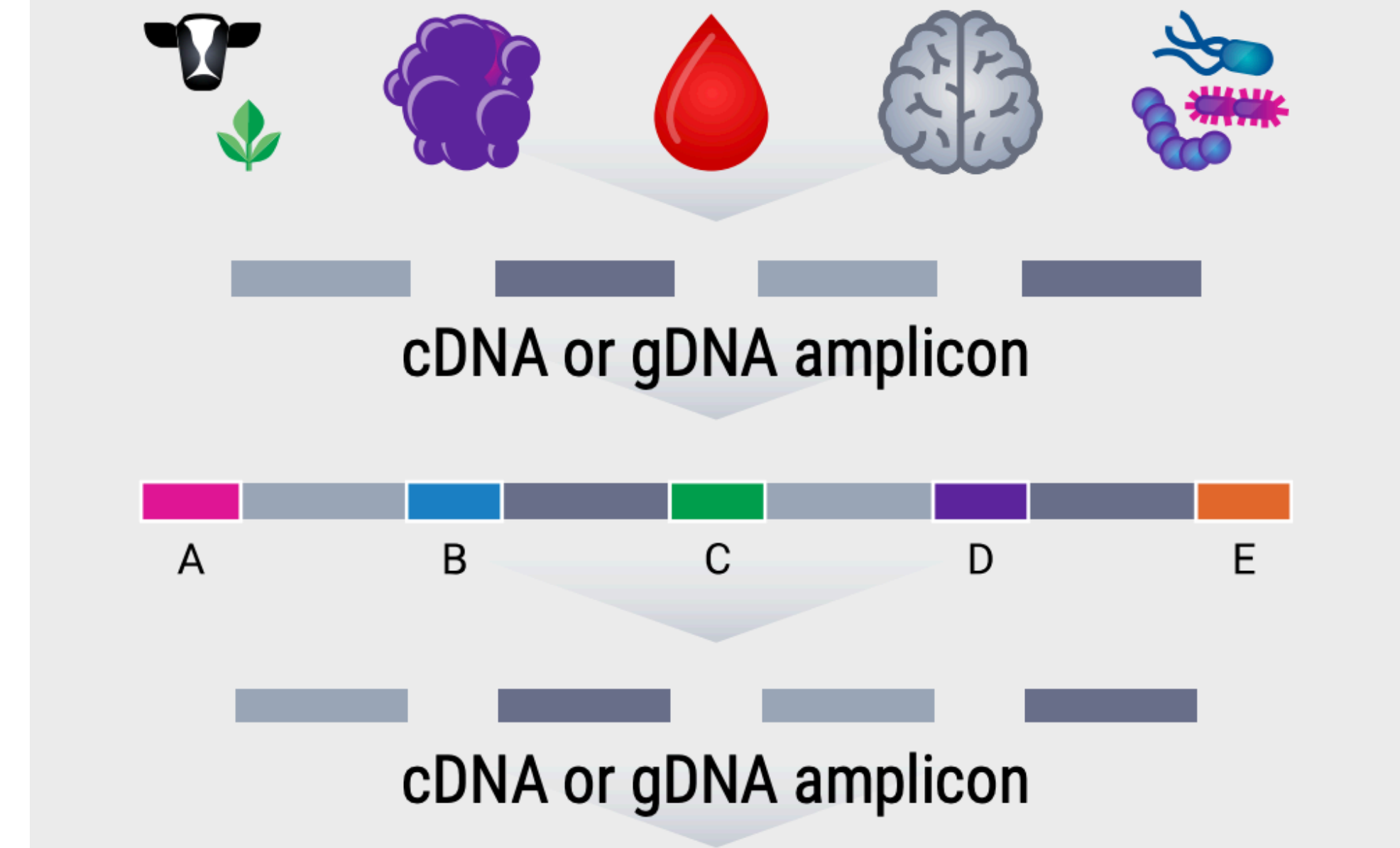
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How Kinnex works

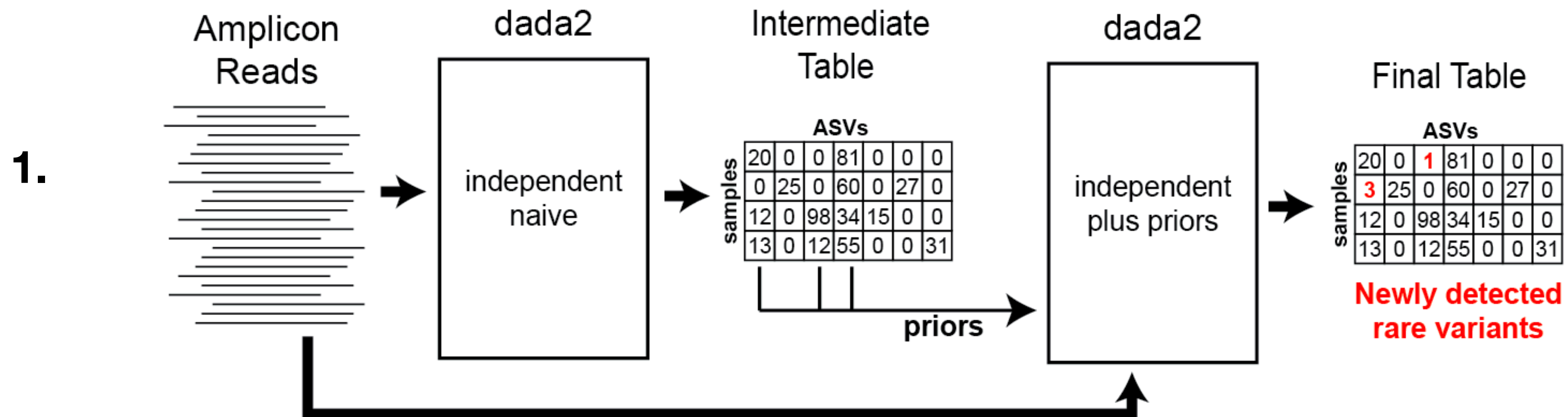


- **Higher throughput**
- Lower costs
- **Binned quality scores**

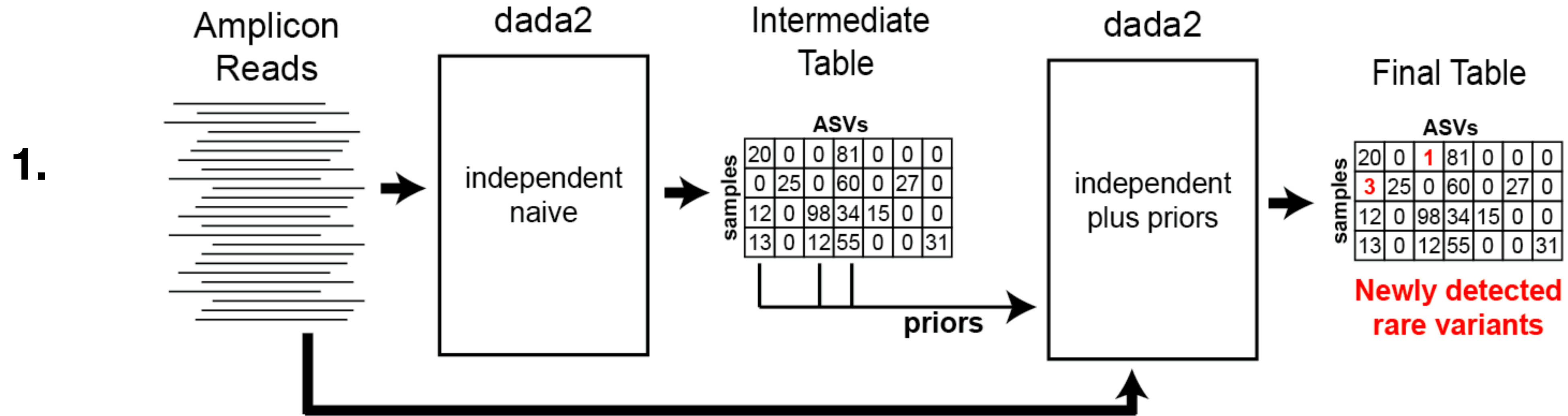
- Higher throughput
- Lower costs
- **Lower quality (16S)**

Updated methods and guidance for DADA2 workflow (in progress)

Pseudo-Pooling

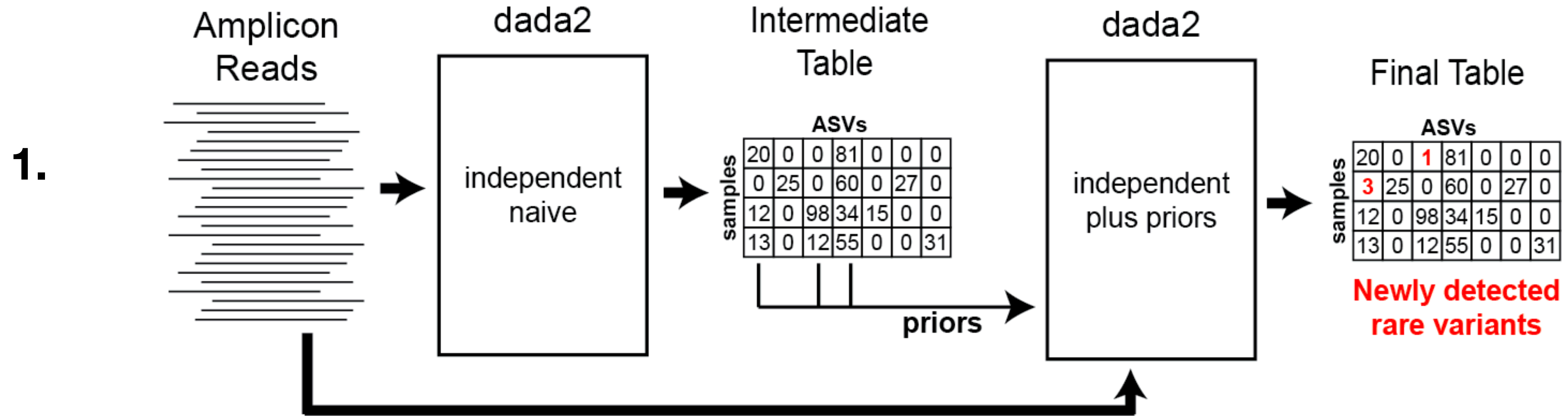


Pseudo-Pooling



2. DETECT_SINGLETONS=TRUE

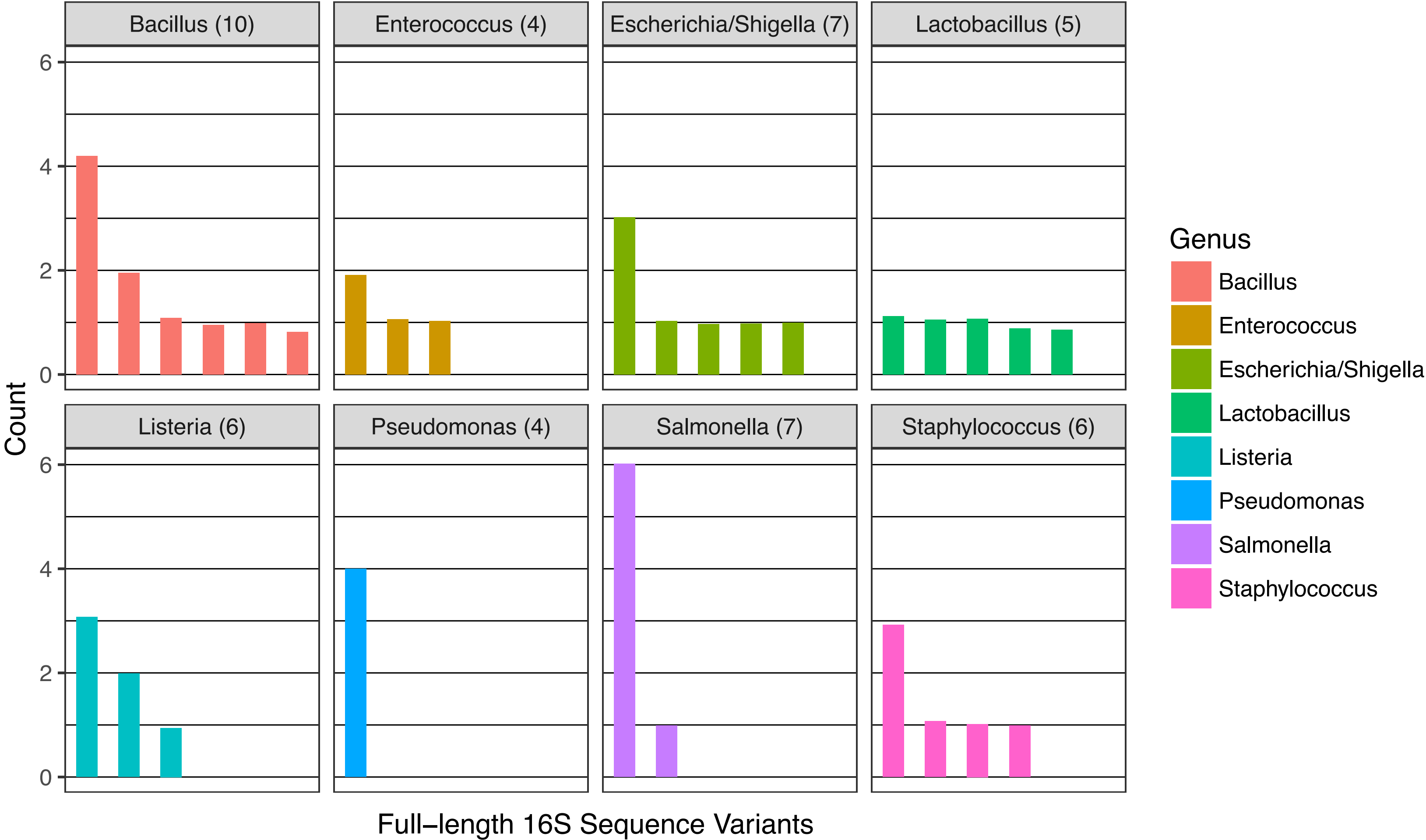
Pseudo-Pooling



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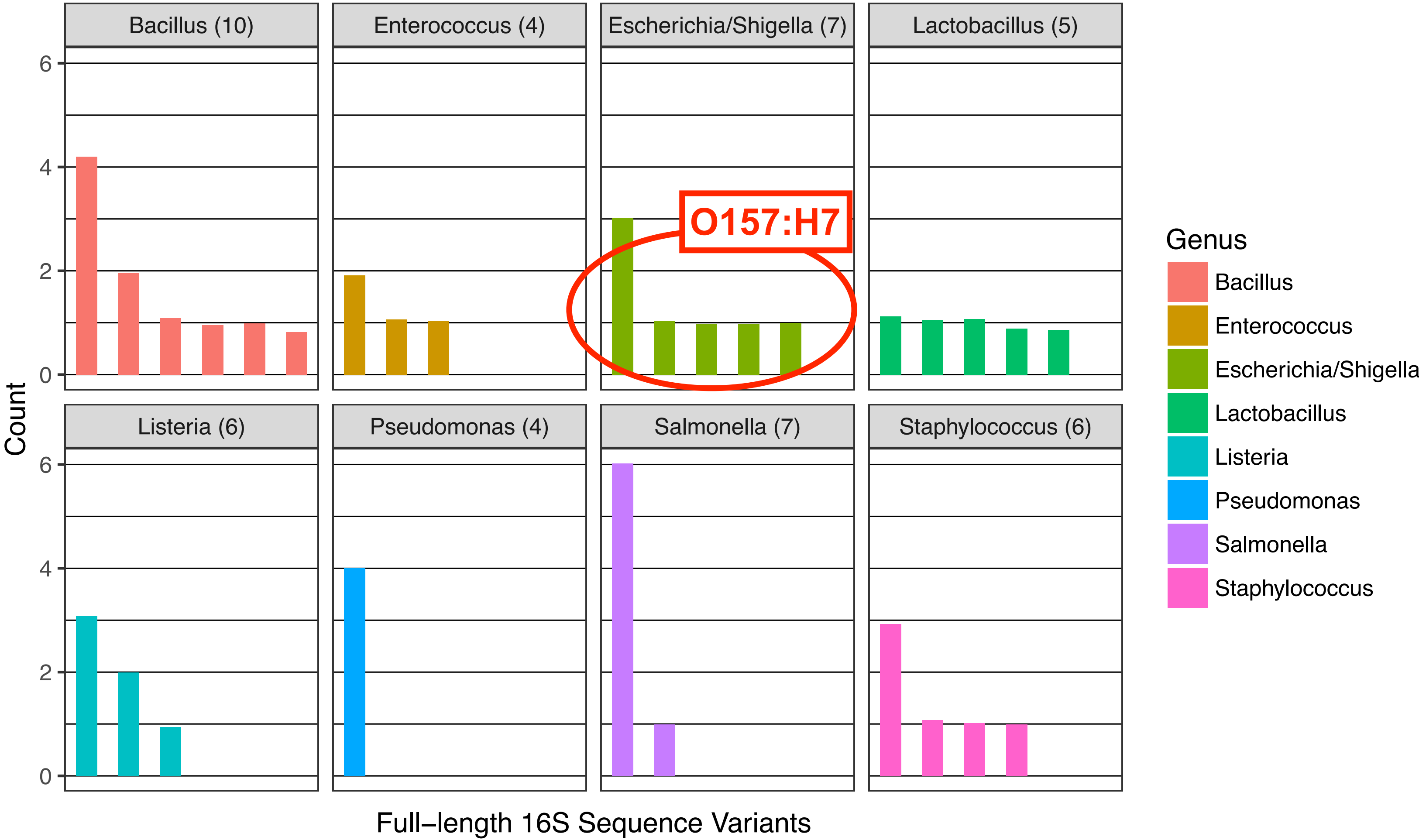
3. Long-read specific parameters.

Full-length 16S from Zymo mock community



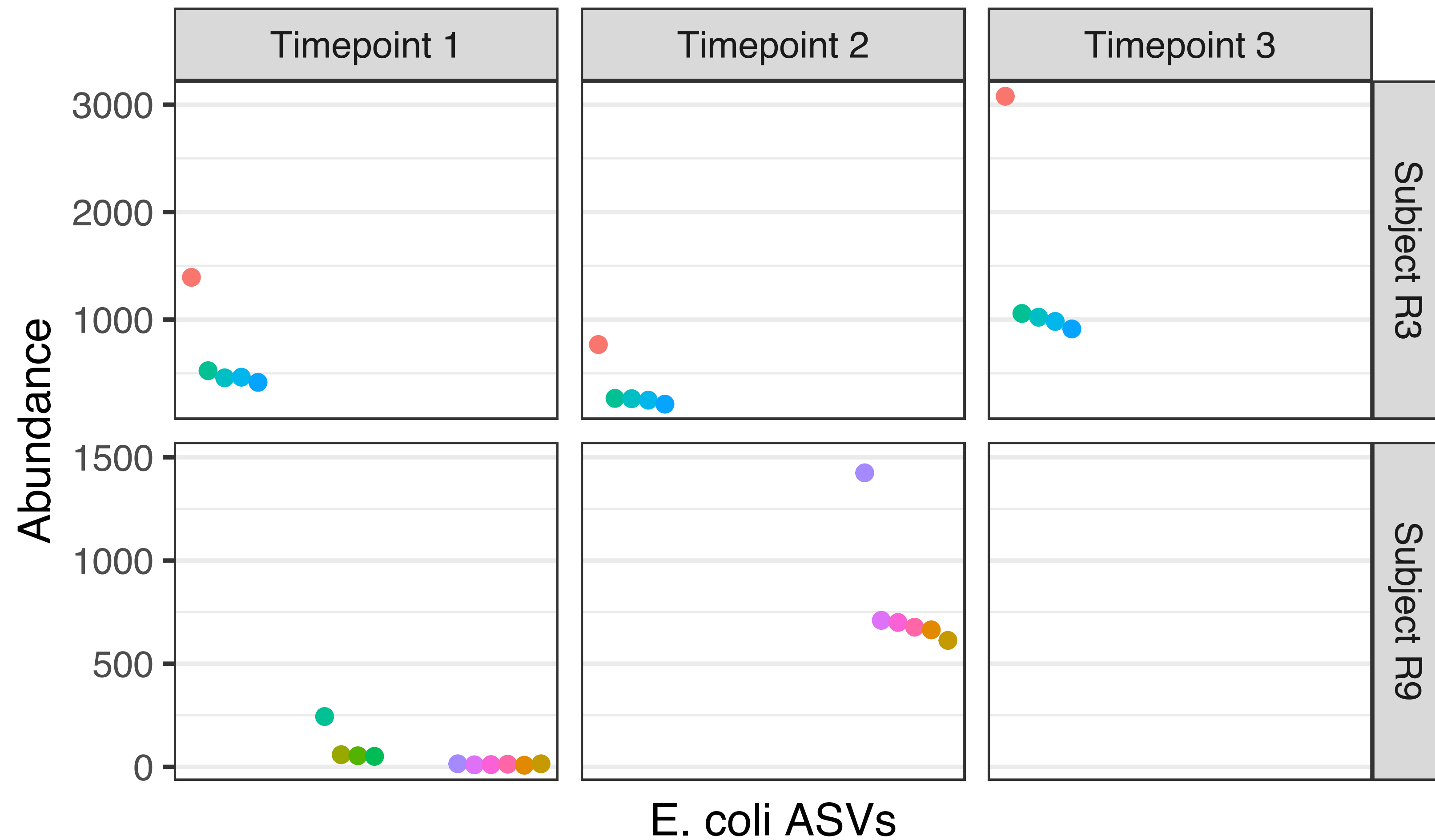
Sub-species bacterial profiling

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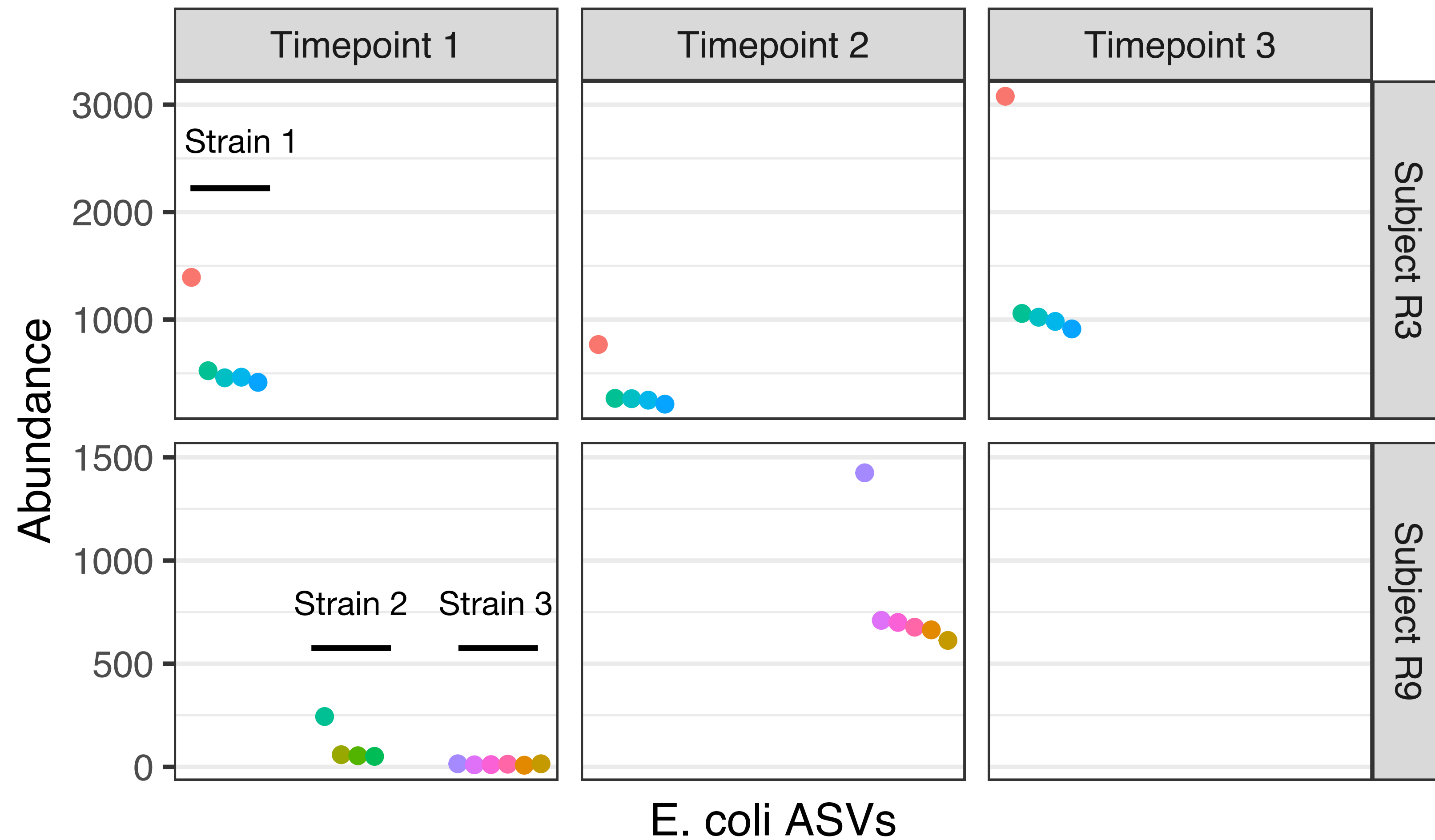
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Full-length 16S from human fecal samples



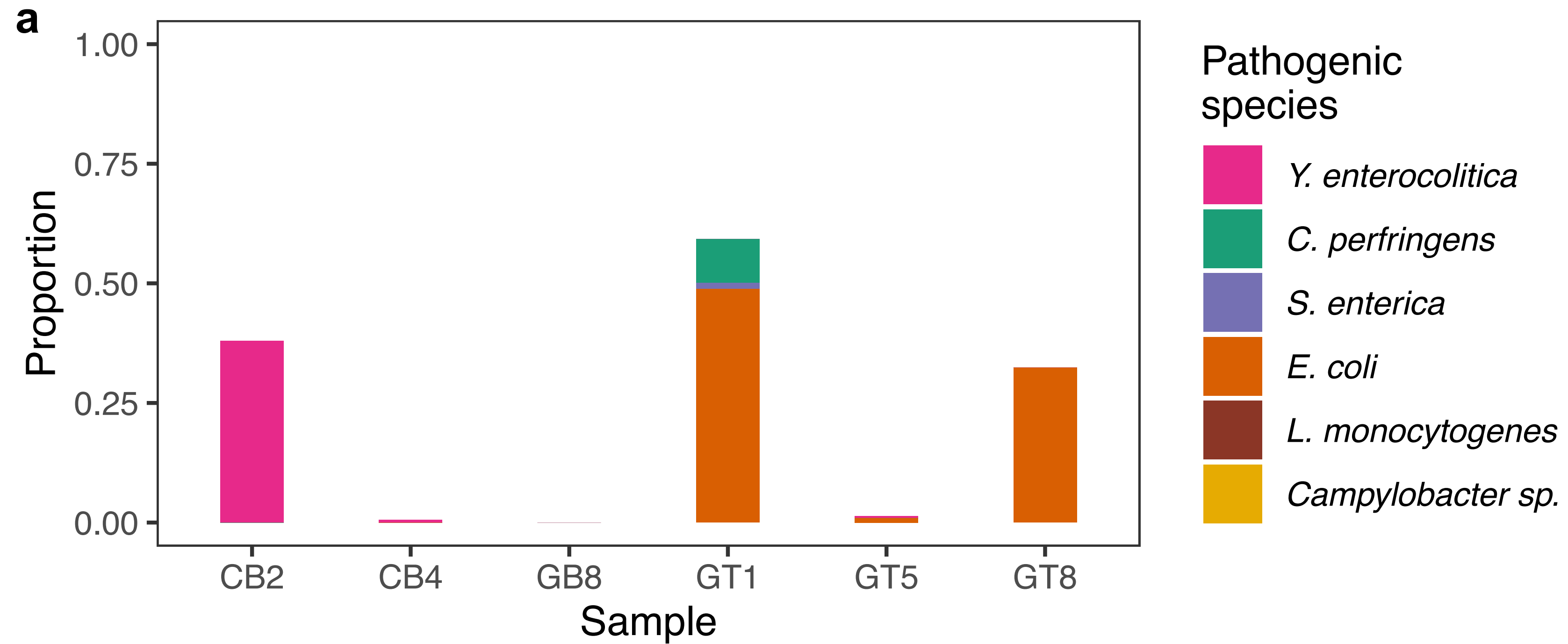
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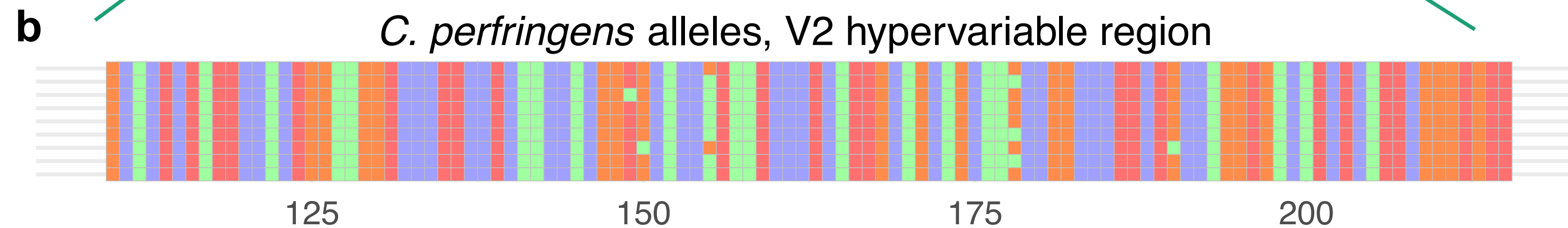
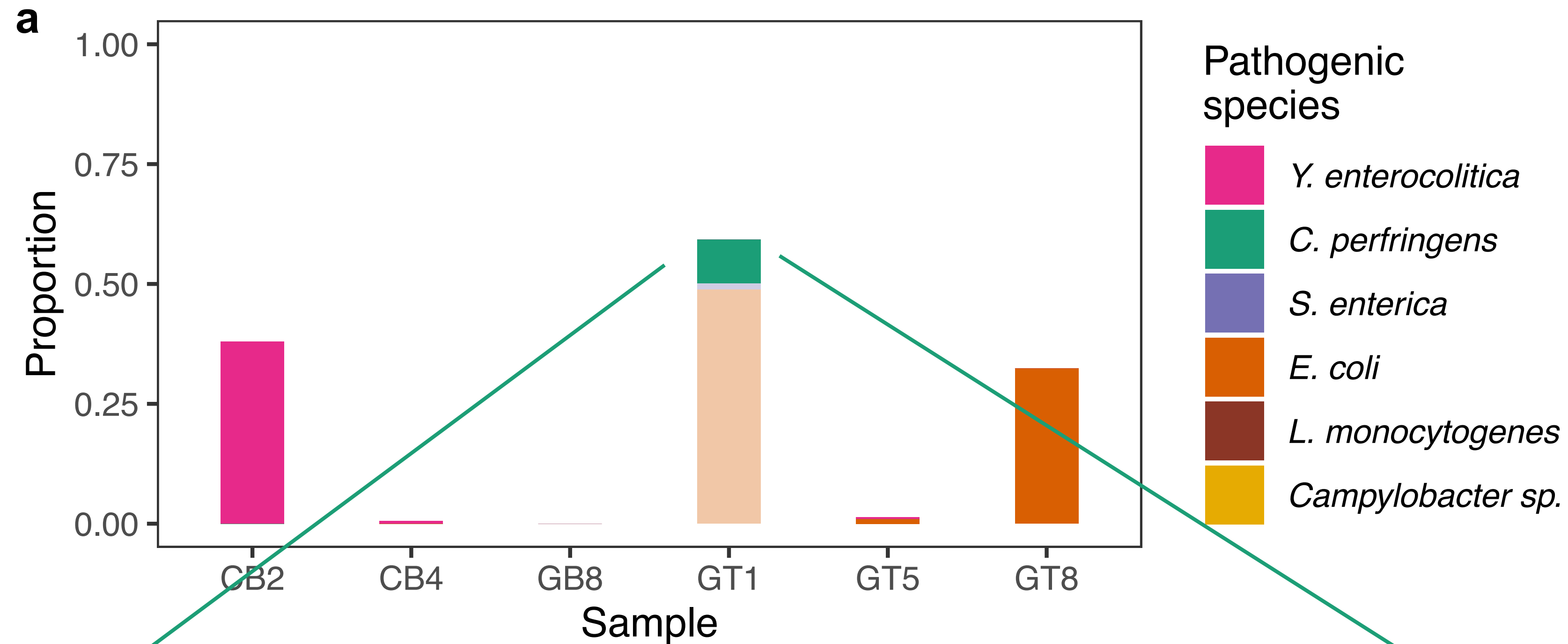
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Full-length 16S from retail meat samples



Sub-species bacterial profiling

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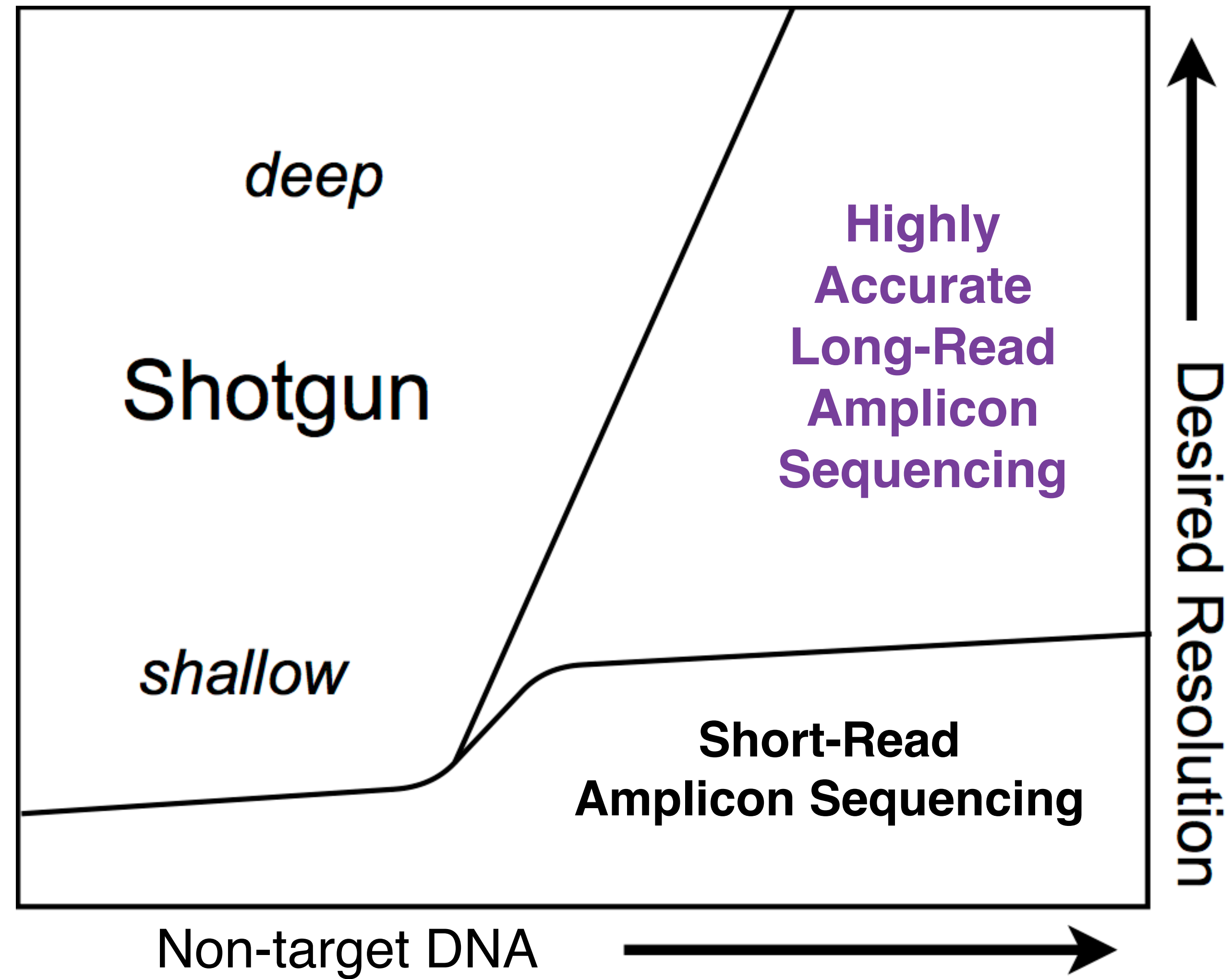


Full genomic complement of 16S alleles

Limitations

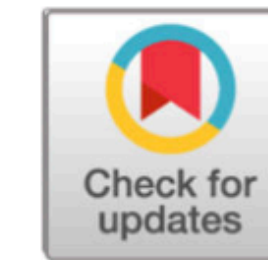
- PCR Biases
- PCR Length Scaling (past ~5 kilo bases)
- Cost-per-base
- Chimeras
- Tooling (especially sub-species assignment)
- ...

Highly Accurate Long-Read Amplicon Seq



Limitations

- PCR Biases
- PCR Length Scaling (past ~5 kilo bases)
- Cost-per-base (big improvements recently!)
- Chimeras
- **Tooling (especially sub-species assignment)**
- ...



8 | Computational Biology | Research Article

Serovar-level identification of bacterial foodborne pathogens from full-length 16S rRNA gene sequencing

Dmitry Grinevich,¹ Lyndy Harden,¹ Siddhartha Thakur,¹ Benjamin Callahan^{1,2}



Dmitry Grinevich



Sid Thakur and lab



National Institute of
Food and Agriculture

Bioinformatics

Susan Holmes
Joey McMurdie
Michael Rosen
Dmitry Grinevich

Applications

Dmitry Grinevich
Casey Theriot
Siddhartha Thakur



Technology (PacBio)

Joan Wong
Cheryl Heiner
Steve Oh



Technology (Loop Genomics)

Tuval Ben Yehezkel
Michael Balamotis

