Reference-guided assembly

Acknowledgements

 Dr. Mihai Pop: Professor, Computer Science, University of Maryland College Park

• Dr. Victoria Cepeda: Formerly PhD student, Pop lab

What is reference-guided assembly?

 An assembler with a strong assumption that genomes in your metagenome look a lot like those that are in a reference database.

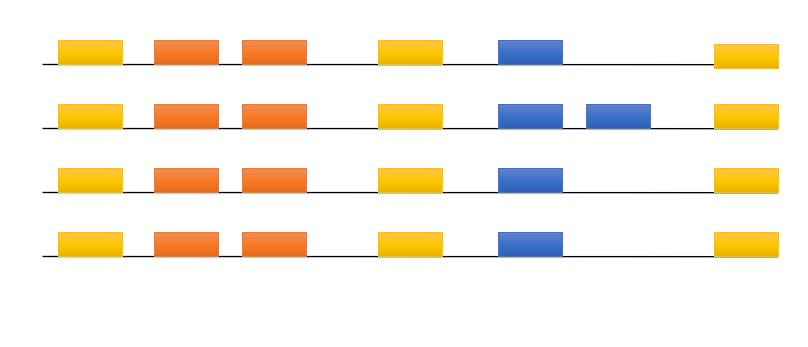
- If this is a reasonable assumption, proceed with caution:
 - Hint: rearrangements, horizonal gene transfer, and duplications are common!
- If this is not a reasonable assumption (viral genomes, soil samples),
 think de novo assembly:
 - Megahit
 - MetaSpades

Microbial genomes evolve over time

• The presence of two or more homologous sequences within a single genome might reflect the acquisition of DNA sequence from a foreign source rather than the duplication of a resident gene.

• Thus, since we do not know the origin a priori, we refer to these potential paralogs or xenologs as **synologs** (Lerat et al 2005).

Ubiquitous sequence fragments

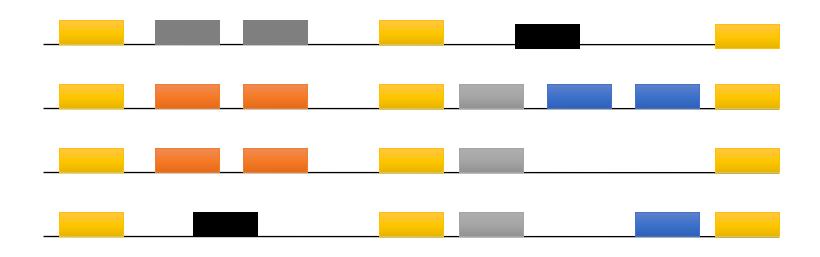


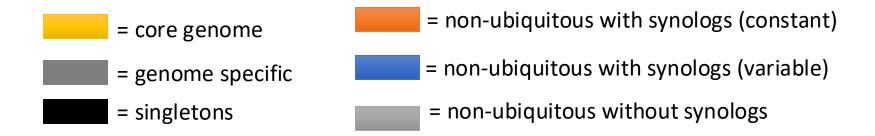
= ubiquitous with synologs (constant)

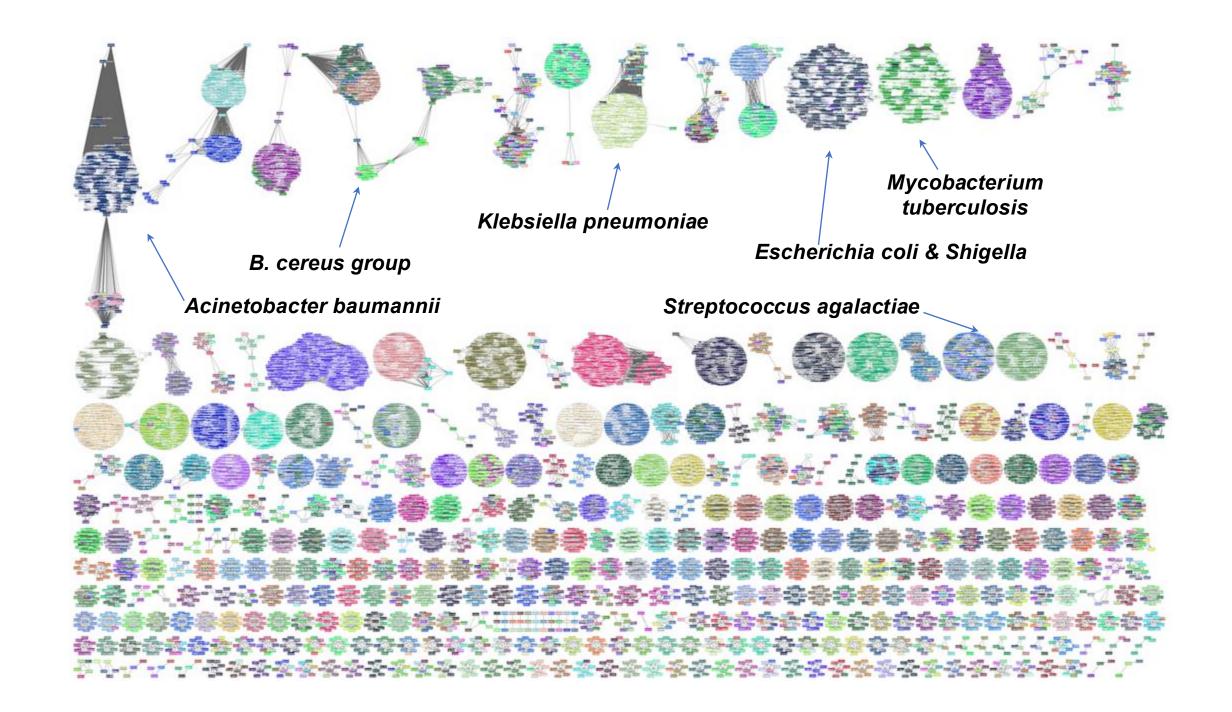
= core genome

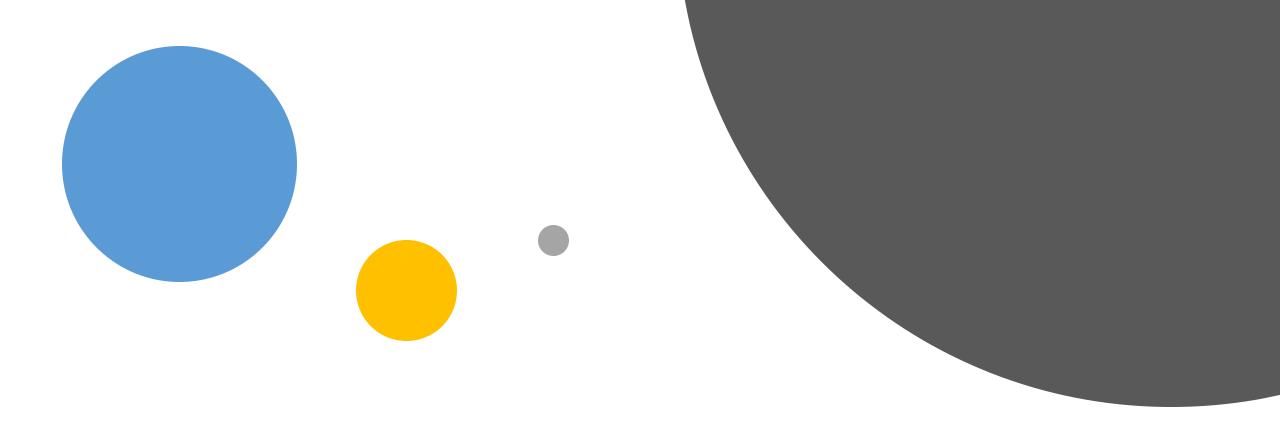
= ubiquitous with synologs (variable)

Non-Ubiquitous sequence fragments









Reference-guided metagenomic assembly

Ready, set, go!

Reference-guided genome assembly

- Reconstructing the original DNA sequence by aligning reads to a genome.
- Intuitively like a puzzle
- But we have the box!





Reference-guided metagenome assembly

- Reconstructing original DNA sequences aligning reads to a set of genomes.
- Intuitively like multiple puzzles
- But we need to find the boxes!





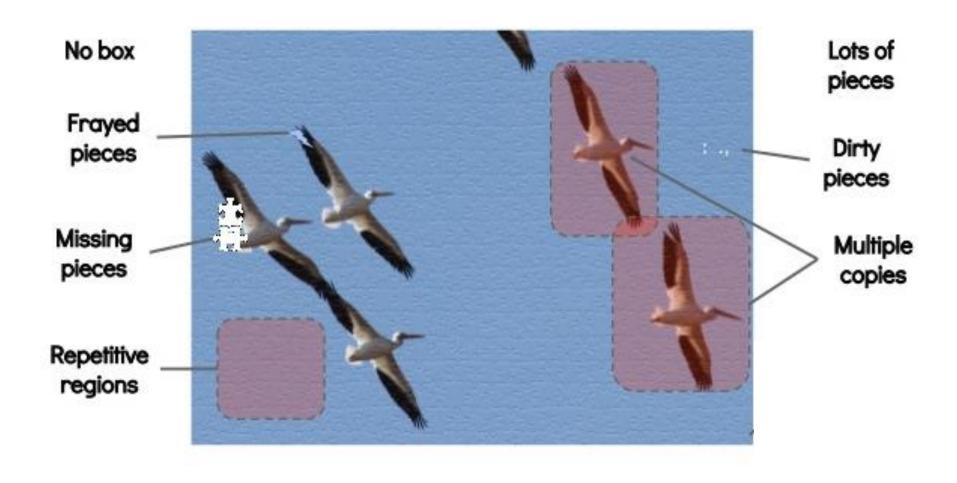
Reference-guided metagenome assembly

- Step 1: Find the puzzle boxes (reference selection)
- Step 2: Bin pieces into the right boxes (read mapping)
- Step 3: Solve each puzzle (assembly)

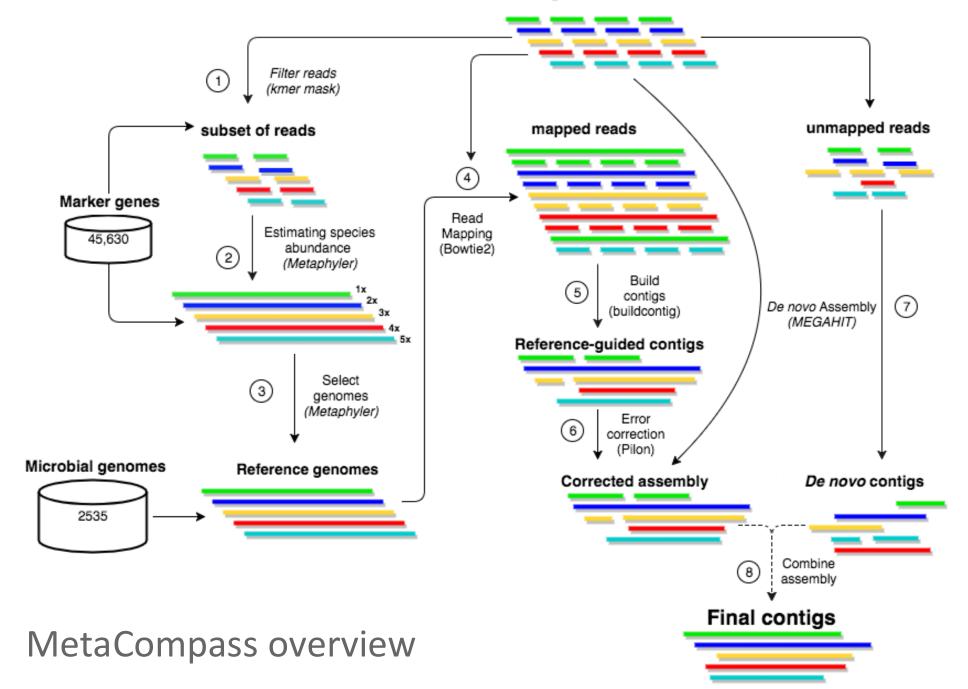


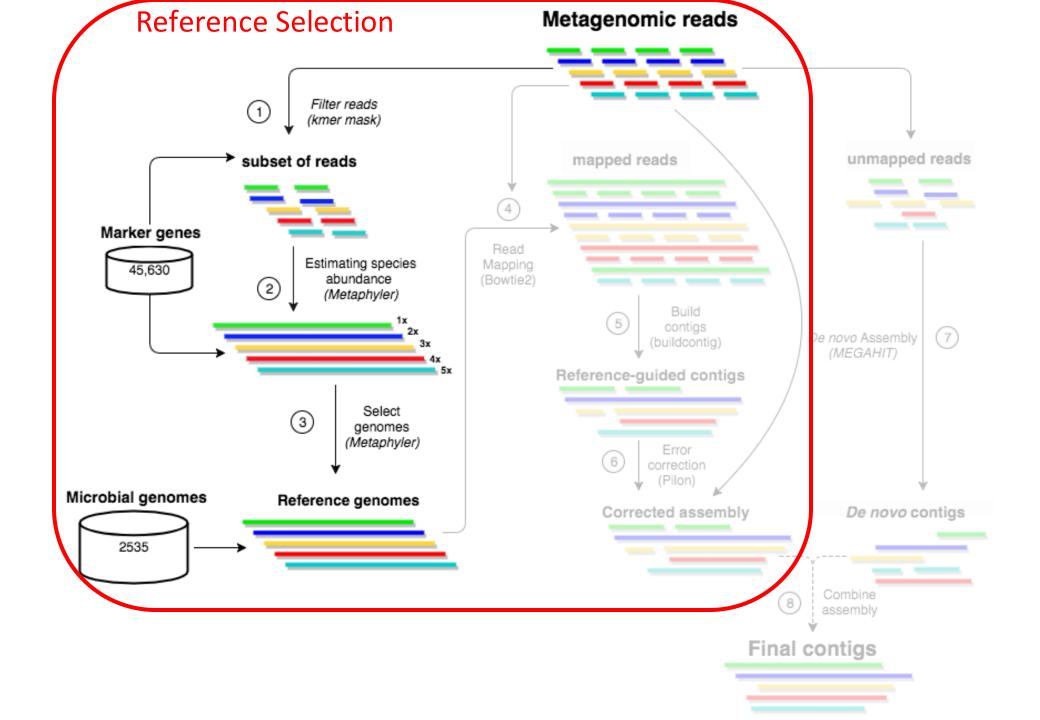


What makes a puzzle hard?

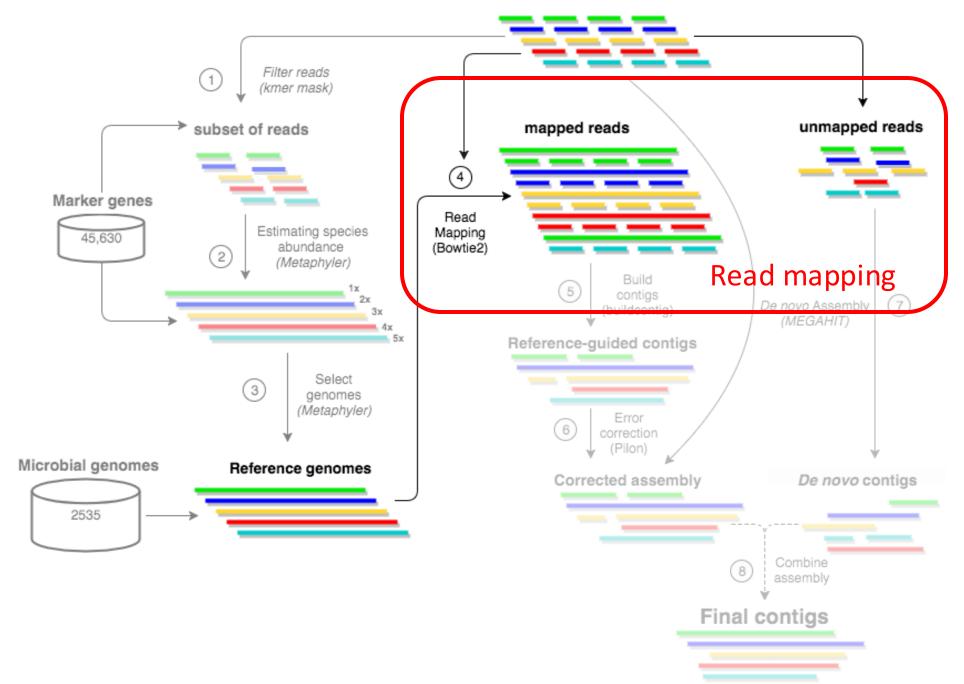


Metagenomic reads





Metagenomic reads

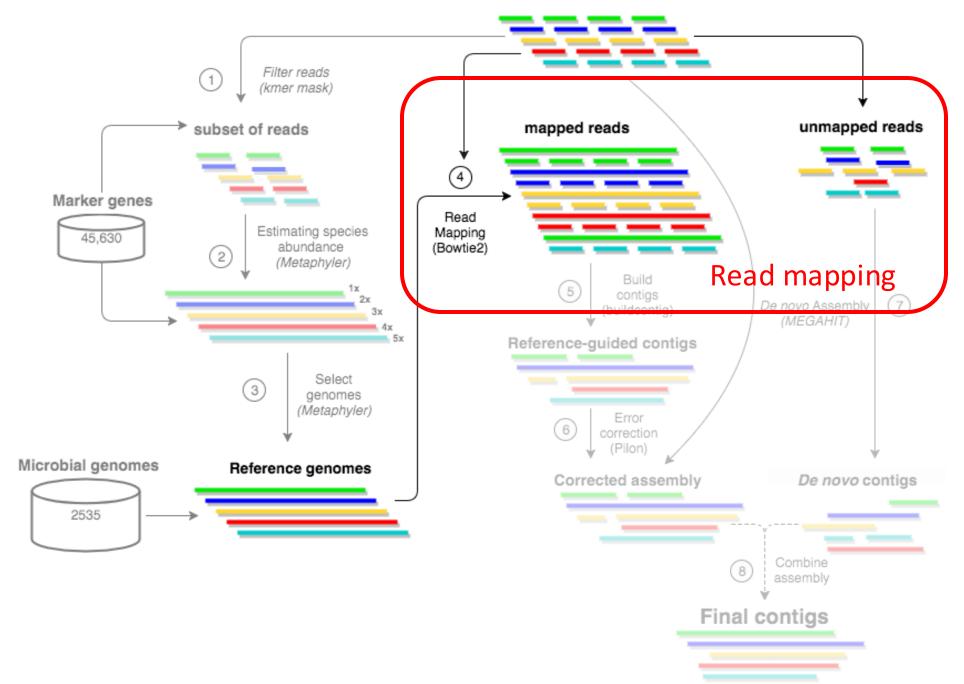


Metagenomic reads Filter reads (kmer mask) mapped reads unmapped reads subset of reads 4 Marker genes Read Estimating species Mapping 45,630 abundance (Bowtie2) (Metaphyler) Build (5) contigs 7 De novo Assembly (buildcontig) (MEGAHIT) Reference-guided contigs Select (3) genomes (Metaphyler) Error 6 correction (Pilon) Microbial genomes Reference genomes Corrected assembly De novo contigs 2535 Combine (8) assembly Metagenomic Final contigs assembly

Choose your own adventure: how shall we identify the reference genomes in our microbiomes?

- 1. Universal marker gene-based approaches (MetaPhlan, etc)
- 2. MinHash based approaches (SourMash, etc)
- 3. Kmer + LCA based approaches (Kraken2, etc)

Metagenomic reads



STEP 2: Read mapping

Software

Open Access

Ultrafast and memory-efficient alignment of short DNA sequences to the human genome

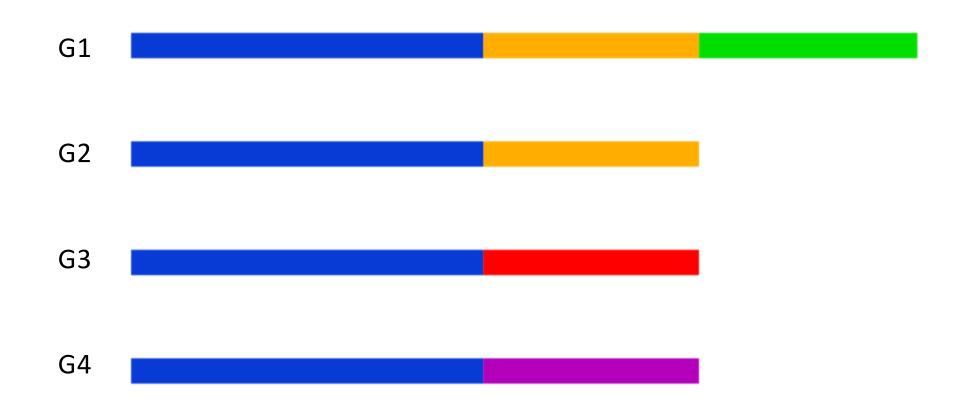
Ben Langmead ™, Cole Trapnell, Mihai Pop and Steven L Salzberg

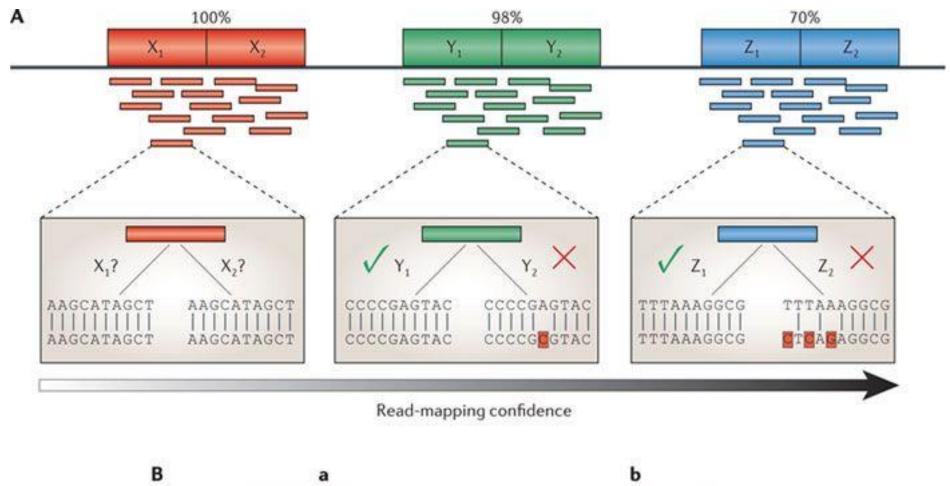
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STEP 2: Read mapping

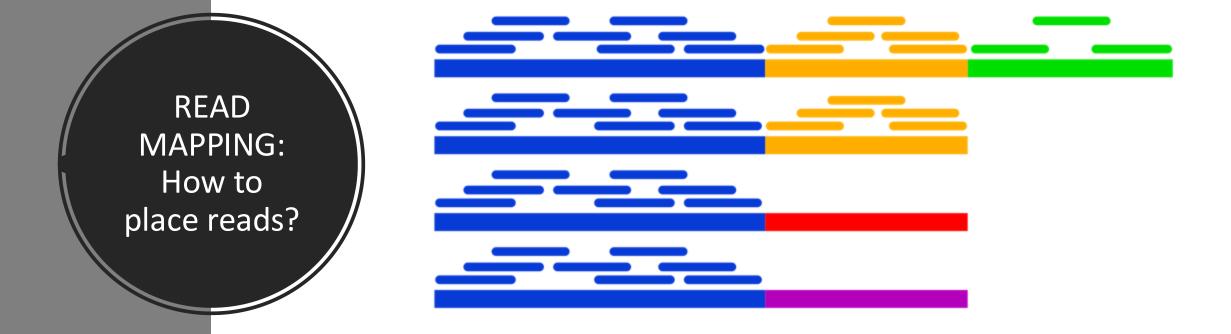


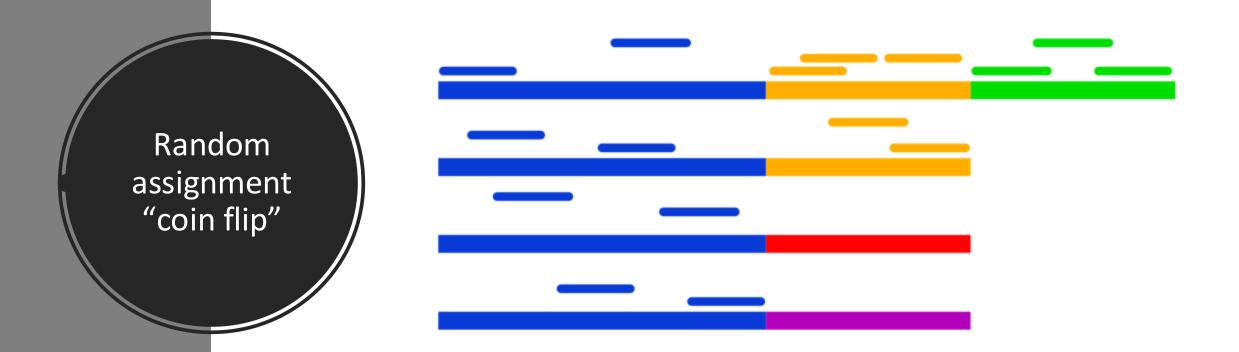


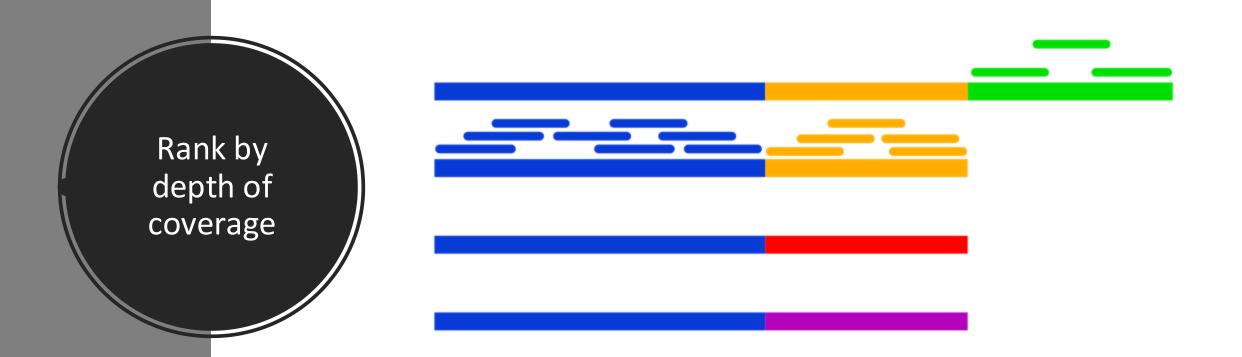


Choose your own adventure: how shall we map reads to the recruited genomes?

- 1. All: map all reads to every equally good mapping location
- 1. Random: randomly assign reads amongst equally good mapping location
- 1. Depth: genome with highest depth of coverage takes all of the reads that map to it
- 1. Breadth: genome with highest breadth of coverage takes all the reads that map to it





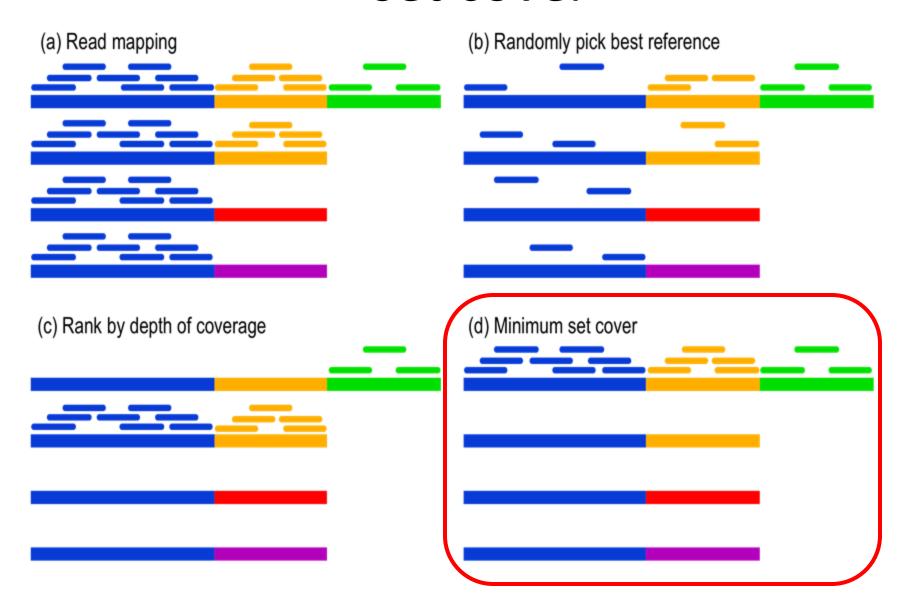




Minimum set cover?

- Given a set of elements U { 1, 2, ..., n } and a collection S of m sets whose union equals the universe, the set cover problem is to identify the smallest sub-collection of S whose union equals the universe.
- For example, consider the universe U = { 1, 2, 3, 4, 5 } and the collection of sets S = { { 1, 2, 3 }, { 2, 4 }, { 3, 4 }, { 4, 5 } } the union of S is U.
- The minimum set cover is the smallest number of m sets that cover U: { 1, 2, 3 }, { 4, 5 } }
- For reference selection, it's the smallest number of genomes that cover all of the input reads.

Read mapping selection: Minimum set cover



STEP 3: Building the contigs

```
Reads TGCACGGATG TGCATGCACG

TTAATGCACG TG-ATGCATG

TGGATTAATG TGGATG-ATG

TGGATTCATGCATGGATGCATGCACG Reference

TGGATTAATGCACGGATG-ATGCACTGCACG Contig
```

Min. depth of coverage:2 Min. length:10

STEP 4: de novo assembly

Assembly unmapped reads to reference



De novo assembly using MEGAHIT

Evaluation Datasets

- Dataset 1: Synthetic dataset, Shakya et. al.
- Dataset 2: Down-sampled Dataset 1(low coverage)
- Dataset 3: 2,077 samples from HMP2

Results - Dataset 1

 Mixture of 64 bacterial and archaeal species (Shakya et al., 2013)

 109 million reads with mean insert size 206 bp and 100 bp read length

Easier to evaluate assembly since the truth is known

Results – Assembly Statistics

Method	No. Contigs	Longest Contig (bp)	Median genome recovery	Mismatches (Per 1Mbp)	Misassemblies (Per 1Mbp)
MetaCompass	18,766	7,057,109	100%	61.9	1.9
IDBA-UD	22,355	991,792	98%	98.6	6.3
MEGAHIT	35,351	1,151,857	99%	66.5	2.5
metaSPAdes	21,424	1,438,235	99%	97.1	2.3

