DS 2021 Jelly Analyses

Mara Bohm and Naomi Lubkin

23/11/2021

```
knitr::opts_chunk$set(tidy.opts=list(width.cutoff=70), tidy=TRUE)
# code above ensures no text is cut off when knit
```

Loading In Packages and Data

```
library(patchwork)
library(car)
library(performance)
library(DHARMa)
library(fitdistrplus)
library(gamlss)
library(tidyverse)
polyp_data <- read_csv("/Users/marabohm/Github/Jelly-Data/data/ds2021_polyp_data.csv")
```

Cleaning the Data

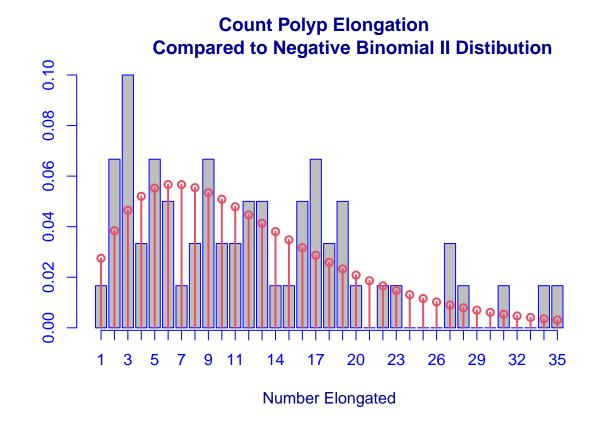
Polyp Elongation

Step 1: Fitting Distributions for Elongation Data

```
fitDist(num_elongated, data = polyp_clean, type = "counts", try.gamlss = T)
```

```
##
## Family: c("NBII", "Negative Binomial type II")
## Fitting method: "nlminb"
## Call: gamlssML(formula = y, family = DIST[i])
## Mu Coefficients:
## [1] 2.504
## Sigma Coefficients:
## [1]
       1.662
##
   Degrees of Freedom for the fit: 2 Residual Deg. of Freedom
                                                                  58
## Global Deviance:
                        410.587
##
               AIC:
                        414.587
##
               SBC:
                        418.776
# best fit: Negative Binomial type II (AIC = 414.587)
# visualizing this fit
histDist(polyp_clean$num_elongated, "NBII", density = T, main = "Count Polyp Elongation
         Compared to Negative Binomial II Distibution",
    xlab = "Number Elongated")
```

##



Step 2: Building the Model

```
mod_countwtotal_elongation <- gamlss(num_elongated ~ treatment * collection_day +
    re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = NBII(),
    method = RS(), data = polyp_clean, control = gamlss.control(n.cyc = 200))</pre>
```

Step 3: Summarizing the Model

```
summary(mod_countwtotal_elongation)
```

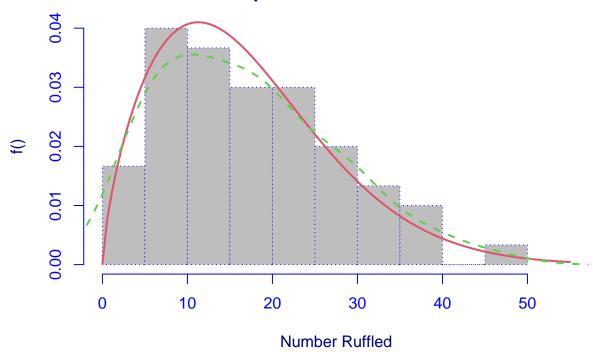
```
## Family: c("NBII", "Negative Binomial type II")
## Call: gamlss(formula = num_elongated ~ treatment * collection_day +
     re(random = ~1 | jar code) + re(random = ~1 | total num),
     family = NBII(), data = polyp_clean, method = RS(),
##
     control = gamlss.control(n.cyc = 200))
##
##
## Fitting method: RS()
##
## -----
## Mu link function: log
## Mu Coefficients:
                               Estimate Std. Error t value Pr(>|t|)
                               ## (Intercept)
## treatmentCaffeine
                               -0.17935
                                        0.38941 -0.461 0.647033
                               0.03055 0.37398
## treatmentEstradiol
                                               0.082 0.935219
## treatmentCombination
                               0.11280
                                        0.36307
                                                0.311 0.757292
## collection_day
                               0.31578 0.07282
                                                4.336 6.74e-05 ***
## treatmentCaffeine:collection_day
                               0.11929
                                        0.09911
                                                1.204 0.234253
## treatmentEstradiol:collection_day
                               0.10750
                                        0.09556
                                                1.125 0.265833
## treatmentCombination:collection day 0.14295
                                        0.09252
                                                1.545 0.128455
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## ------
## Sigma link function: log
## Sigma Coefficients:
           Estimate Std. Error t value Pr(>|t|)
## (Intercept) -1.4610 0.9872 -1.48 0.145
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## -----
## No. of observations in the fit: 60
## Degrees of Freedom for the fit: 8.35727
       Residual Deg. of Freedom: 51.64273
##
##
                   at cycle: 43
```

Polyp Ruffling

Step 1: Fitting Distributions for Ruffling Data

```
fitDist(num_ruffled, data = polyp_clean, type = "counts", try.gamlss = T)
# best fit: Negative Binomial type II (AIC = 444.109)
# model was having difficulty converging with NBII, so tried some
# other fits:
descdist(polyp_clean$num_ruffled)
# Cullen and Frey plot indicates Lognormal, Gamma and Weibull
# distributions may be a good fit, so we compare to each other using
# Generalized AIC:
b_NB2 <- histDist(polyp_clean$num_ruffled, "NBII", density = T)</pre>
b_lNO <- histDist(polyp_clean$num_ruffled, "LOGNO", density = T)</pre>
b_GA <- histDist(polyp_clean$num_ruffled, "GA", density = T)</pre>
b_WEI <- histDist(polyp_clean$num_ruffled, "WEI", density = T)</pre>
GAIC(b_1NO, b_GA, b_WEI, b_NB2)
         df
                 AIC
## b_WEI 2 442.8371
## b_NB2 2 444.1095
## b_GA 2 444.1204
## b_1NO 2 451.8062
# best fit: Weibull Distribution (AIC = 442.837)
# visualizing this fit
histDist(polyp_clean$num_ruffled, "WEI", density = T, main = "Count Polyp Ruffling
         Compared to Weibull Distribution",
    xlab = "Number Ruffled")
```

Count Polyp Ruffling Compared to Weibull Distribution



```
##
## Family: c("WEI", "Weibull")
## Fitting method: "nlminb"
## Call: gamlssML(formula = polyp_clean$num_ruffled, family = "WEI")
## Mu Coefficients:
## [1] 2.947
## Sigma Coefficients:
## [1] 0.529
##
## Degrees of Freedom for the fit: 2 Residual Deg. of Freedom
                                                                  58
## Global Deviance:
                        438.837
##
               AIC:
                        442.837
               SBC:
##
                        447.026
```

Step 2: Building the Model

```
mod_countwtotal_ruffled <- gamlss(num_ruffled ~ treatment * collection_day +
    re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = WEI(),
    data = polyp_clean, control = gamlss.control(n.cyc = 60))</pre>
```

Step 3: Summarizing the Model

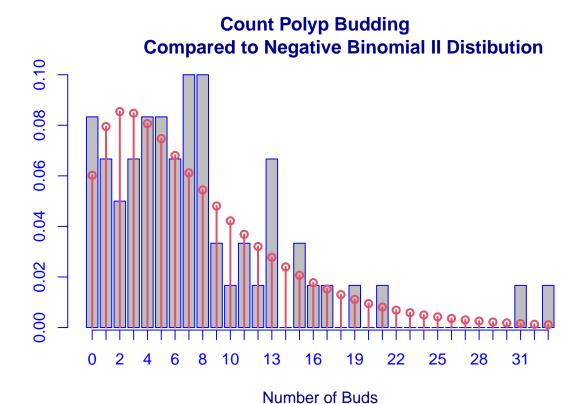
summary(mod_countwtotal_elongation)

```
## Family: c("NBII", "Negative Binomial type II")
##
## Call: gamlss(formula = num_elongated ~ treatment * collection_day +
##
     re(random = ~1 | jar_code) + re(random = ~1 | total_num),
##
     family = NBII(), data = polyp_clean, method = RS(),
##
     control = gamlss.control(n.cyc = 200))
##
## Fitting method: RS()
##
## -----
## Mu link function: log
## Mu Coefficients:
##
                               Estimate Std. Error t value Pr(>|t|)
                                1.09720 0.28047
## (Intercept)
                                                 3.912 0.000269 ***
                               -0.17935
                                         0.38941 -0.461 0.647033
## treatmentCaffeine
                                0.03055 0.37398 0.082 0.935219
## treatmentEstradiol
## treatmentCombination
                                0.11280 0.36307
                                                 0.311 0.757292
                                0.31578 0.07282
                                                 4.336 6.74e-05 ***
## collection_day
## treatmentCaffeine:collection_day 0.11929 0.09911 1.204 0.234253
## treatmentEstradiol:collection_day 0.10750 0.09556 1.125 0.265833
## treatmentCombination:collection_day 0.14295 0.09252
                                                 1.545 0.128455
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
## -----
## Sigma link function: log
## Sigma Coefficients:
           Estimate Std. Error t value Pr(>|t|)
## (Intercept) -1.4610 0.9872 -1.48 0.145
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## No. of observations in the fit: 60
## Degrees of Freedom for the fit: 8.35727
##
       Residual Deg. of Freedom: 51.64273
##
                    at cycle: 43
##
## Global Deviance:
                   316.6155
##
            AIC:
                   333.33
##
            SBC:
                   350.833
```

Polyp Asexual Budding

Step 1: Fitting Distributions for Buds Data

```
fitDist(num_asexual_buds, data = polyp_clean, type = "counts", try.gamlss = T)
##
##
    Lapack routine dgesv: system is exactly singular: U[1,1] = 0
##
    Lapack routine dgesv: system is exactly singular: U[2,2] = 0
##
##
                                                                                    |-----
##
## Family: c("NBII", "Negative Binomial type II")
## Fitting method: "nlminb"
## Call: gamlssML(formula = y, family = DIST[i])
##
## Mu Coefficients:
## [1] 2.041
## Sigma Coefficients:
## [1] 1.575
##
## Degrees of Freedom for the fit: 2 Residual Deg. of Freedom
## Global Deviance:
                       368.321
##
              AIC:
                       372.321
              SBC:
                       376.509
##
# best fit: Negative Binomial type II (AIC = 372.321)
# visualizing this fit
histDist(polyp_clean$num_asexual_buds, "NBII", density = T, main = "Count Polyp Budding
        Compared to Negative Binomial II Distibution",
  xlab = "Number of Buds")
```



Step 2: Building the Model

```
mod_countwtotal_buds <- gamlss(num_asexual_buds ~ treatment * collection_day +
    re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = NBII(),
    data = polyp_clean, control = gamlss.control(n.cyc = 250))</pre>
```

Step 3: Summarizing the Model

```
## Mu link function: log
## Mu Coefficients:
##
                               Estimate Std. Error t value Pr(>|t|)
                               -0.54561 0.44010 -1.240 0.22097
## (Intercept)
## treatmentCaffeine
                               0.79621
                                        0.54887
                                                1.451 0.15326
## treatmentEstradiol
                               0.17758 0.57850
                                               0.307 0.76017
## treatmentCombination
                               1.66274 0.50878
                                                3.268 0.00198 **
                               0.60862
                                                5.835 4.2e-07 ***
## collection day
                                        0.10430
## treatmentCaffeine:collection_day -0.08091 0.13120 -0.617 0.54027
## treatmentEstradiol:collection_day    0.06207    0.13640
                                                0.455 0.65108
## treatmentCombination:collection_day -0.26110
                                        0.12315 -2.120 0.03908 *
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
## -----
## Sigma link function: log
## Sigma Coefficients:
           Estimate Std. Error t value Pr(>|t|)
                     2.645 -0.955
## (Intercept) -2.526
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## -----
## No. of observations in the fit: 60
## Degrees of Freedom for the fit: 11.04057
       Residual Deg. of Freedom: 48.95943
##
                   at cycle: 8
## Global Deviance:
                   271.739
##
           AIC:
                   293.8201
           SBC:
                   316.9429
```

Polyp Total Number

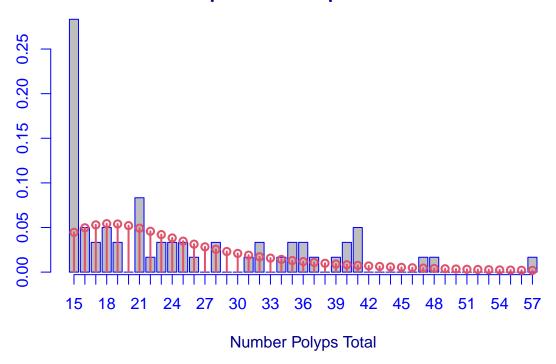
##

Step 1: Fitting Distributions for Total Data

Call: gamlssML(formula = y, family = DIST[i])

```
## Mu Coefficients:
## [1] 3.208
## Sigma Coefficients:
## [1]
       0.2898
## Nu Coefficients:
## [1] 0.5139
  Degrees of Freedom for the fit: 3 Residual Deg. of Freedom
##
## Global Deviance:
                        430.754
##
                        436.754
               AIC:
##
               SBC:
                        443.038
# best fit: delaporte (AIC = 436.754)
# visualizing this fit
histDist(polyp_clean$total_num, "DEL", density = T, main = "Count Polyp Total
         Compared to Delaporte Distribution",
   xlab = "Number Polyps Total")
```

Count Polyp Total Compared to Delaporte Distribution



Step 2: Building the Model

```
mod_total <- gamlss(total_num ~ treatment * collection_day + re(random = ~1 |
    jar_code), family = DEL(), data = polyp_clean, control = gamlss.control(n.cyc = 60))</pre>
```

Step 3: Summarizing the Model

```
summary(mod total)
## Family: c("DEL", "Delaporte")
## Call:
## gamlss(formula = total_num ~ treatment * collection_day + re(random = ~1 |
     jar_code), family = DEL(), data = polyp_clean, control = gamlss.control(n.cyc = 60))
##
##
## Fitting method: RS()
## -----
## Mu link function: log
## Mu Coefficients:
                             Estimate Std. Error t value Pr(>|t|)
##
                             ## (Intercept)
                            -0.011063 0.522953 -0.021
## treatmentCaffeine
                                                    0.9832
                             0.005354 0.469892 0.011
## treatmentEstradiol
                                                    0.9910
## treatmentCombination
                             0.048451 0.429514 0.113 0.9106
                             ## collection_day
## treatmentCaffeine:collection_day 0.050239 0.128599 0.391 0.6977
## treatmentEstradiol:collection_day -0.005437 0.111598 -0.049 0.9613
## treatmentCombination:collection_day 0.069381 0.105428
                                             0.658 0.5135
## ---
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
## -----
## Sigma link function: log
## Sigma Coefficients:
           Estimate Std. Error t value Pr(>|t|)
## (Intercept) -36.04 12909.94 -0.003 0.998
## -----
## Nu link function: logit
## Nu Coefficients:
     Estimate Std. Error t value Pr(>|t|)
## (Intercept) 0 12910 0 1
##
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms may not be reliable.
## -----
## No. of observations in the fit: 60
## Degrees of Freedom for the fit: 12.73762
      Residual Deg. of Freedom: 47.26238
##
##
                  at cycle: 5
##
## Global Deviance:
                 318.8629
##
          AIC:
                344.3381
```

SBC: 371.0151