

# DS 2021 Jelly Analyses

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```
knitr::opts_chunk$set(tidy.opts=list(width.cutoff=70), tidy=TRUE)
# code above ensures no text is cut off when knit
```

## Loading In Packages and Data

```
library(patchwork)
library(car)
library(performance)
library(DHARMA)
library(fitdistrplus)
library(gamlss)
library(tidyverse)

polyp_data <- read_csv("/Users/marabohm/Github/Jelly-Data/data/ds2021_polyp_data.csv")
```

## Cleaning the Data

```
polyp_clean <- polyp_data %>%
  rename(collection_day = "Data collection day", jar_code = "Jar Code (ex. E1)",
    treatment = Chemical, num_elongated = "Elongation?", num_ruffled = "Ruffled?",
    num_aseexual_buds = "Asexual Repro?", total_num = Total) %>%
  mutate(jar_code = as.factor(jar_code), treatment = as.factor(treatment),
    treatment = fct_relevel(treatment, "Control", "Caffeine", "Estradiol",
      "Combo")) %>%
  dplyr::select(collection_day, jar_code, treatment, num_elongated, num_ruffled,
    num_aseexual_buds, total_num)
```

## Polyp Elongation

### Step 1: Fitting Distributions for Elongation Data

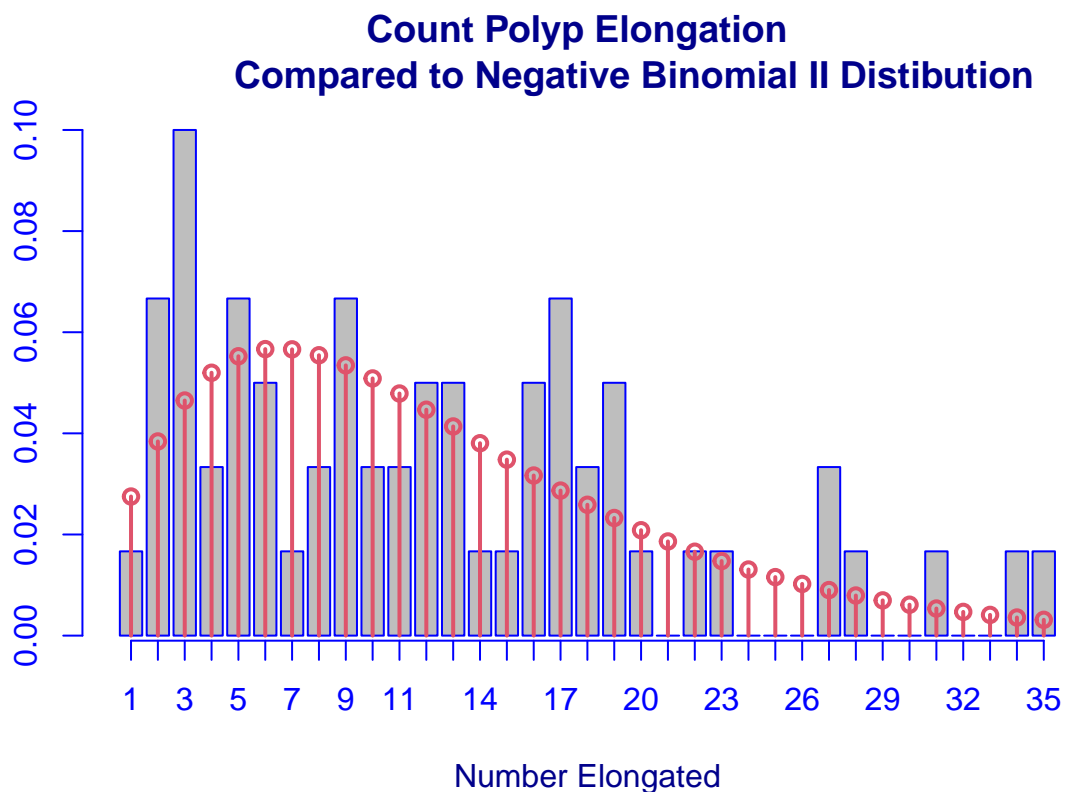
```
fitDist(num_elongated, data = polyp_clean, type = "counts", try.gamlss = T)
```

```
## |

##
## Family: c("NBII", "Negative Binomial type II")
## Fitting method: "nlminb"
##
## Call: gamlssML(formula = y, family = DIST[i])
##
## Mu Coefficients:
## [1] 2.504
## Sigma Coefficients:
## [1] 1.662
##
## Degrees of Freedom for the fit: 2 Residual Deg. of Freedom 58
## Global Deviance: 410.587
## AIC: 414.587
## SBC: 418.776
```

```
# best fit: Negative Binomial type II (AIC = 414.587)
```

```
# visualizing this fit
histDist(polyp_clean$num_elongated, "NBII", density = T, main = "Count Polyp Elongation
  Compared to Negative Binomial II Distribution",
  xlab = "Number Elongated")
```



## Step 2: Building the Model

```
mod_countwttotal_elongation <- gamlss(num_elongated ~ treatment * collection_day +
  re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = NBII(),
  method = RS(), data = polyp_clean, control = gamlss.control(n.cyc = 200))
```

## Step 3: Summarizing the Model

```
summary(mod_countwttotal_elongation)
```

```
## *****
## Family:  c("NBII", "Negative Binomial type II")
##
## Call:  gamlss(formula = num_elongated ~ treatment * collection_day +
##           re(random = ~1 | jar_code) + re(random = ~1 | total_num),
##           family = NBII(), data = polyp_clean, method = RS(),
##           control = gamlss.control(n.cyc = 200))
##
## Fitting method: RS()
##
## -----
## Mu link function:  log
## Mu Coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      1.09720    0.28047   3.912 0.000269 ***
## treatmentCaffeine -0.17935    0.38941  -0.461 0.647033
## treatmentEstradiol  0.03055    0.37398   0.082 0.935219
## treatmentCombination 0.11280    0.36307   0.311 0.757292
## collection_day      0.31578    0.07282   4.336 6.74e-05 ***
## treatmentCaffeine:collection_day 0.11929    0.09911   1.204 0.234253
## treatmentEstradiol:collection_day 0.10750    0.09556   1.125 0.265833
## treatmentCombination:collection_day 0.14295    0.09252   1.545 0.128455
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## -----
## Sigma link function:  log
## Sigma Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -1.4610    0.9872  -1.48   0.145
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## -----
## No. of observations in the fit:  60
## Degrees of Freedom for the fit:  8.35727
##      Residual Deg. of Freedom:  51.64273
##                               at cycle:  43
```

```
##
## Global Deviance:      316.6155
##           AIC:       333.33
##           SBC:       350.833
## *****
```

## Polyp Ruffling

### Step 1: Fitting Distributions for Ruffling Data

```
fitDist(num_ruffled, data = polyp_clean, type = "counts", try.gamlss = T)
# best fit: Negative Binomial type II (AIC = 444.109)
```

```
# model was having difficulty converging with NBII, so tried some
# other fits:
```

```
descdist(polyp_clean$num_ruffled)
```

```
# Cullen and Frey plot indicates Lognormal, Gamma and Weibull
# distributions may be a good fit, so we compare to each other using
# Generalized AIC:
```

```
b_NB2 <- histDist(polyp_clean$num_ruffled, "NBII", density = T)
```

```
b_LNO <- histDist(polyp_clean$num_ruffled, "LOGNO", density = T)
```

```
b_GA <- histDist(polyp_clean$num_ruffled, "GA", density = T)
```

```
b_WEI <- histDist(polyp_clean$num_ruffled, "WEI", density = T)
```

```
GAIC(b_LNO, b_GA, b_WEI, b_NB2)
```

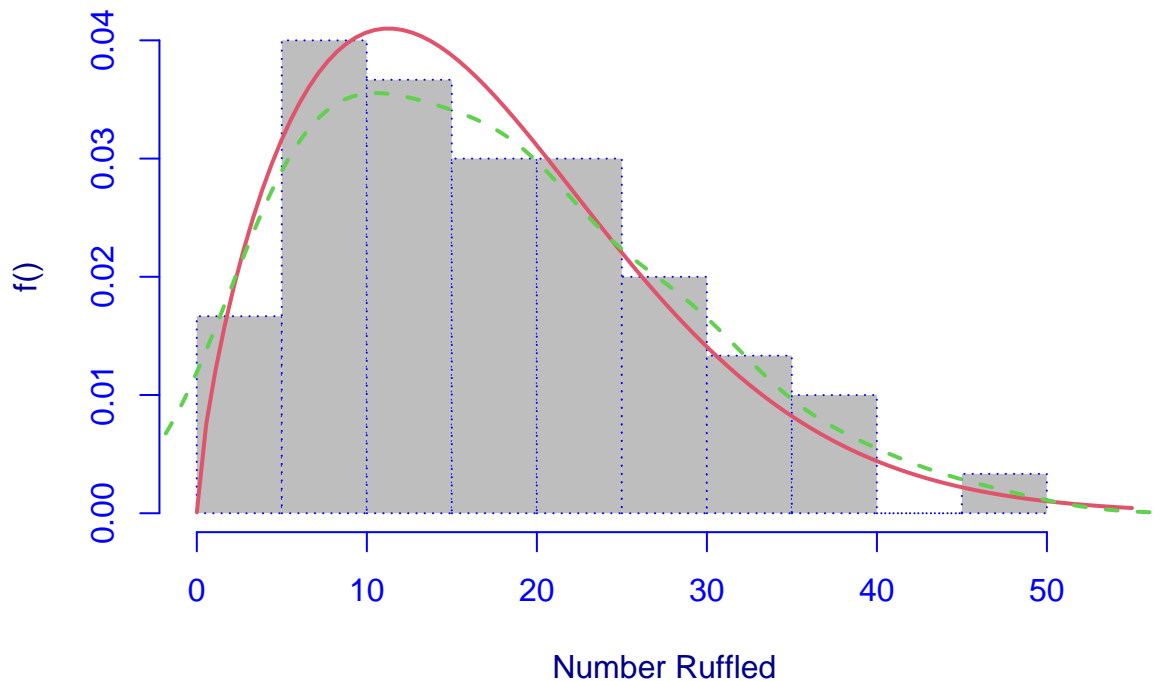
```
##           df           AIC
## b_WEI    2 442.8371
## b_NB2    2 444.1095
## b_GA     2 444.1204
## b_LNO    2 451.8062
```

```
# best fit: Weibull Distribution (AIC = 442.837)
```

```
# visualizing this fit
```

```
histDist(polyp_clean$num_ruffled, "WEI", density = T, main = "Count Polyp Ruffling
  Compared to Weibull Distribution",
  xlab = "Number Ruffled")
```

## Count Polyp Ruffling Compared to Weibull Distribution



```
##
## Family: c("WEI", "Weibull")
## Fitting method: "nlminb"
##
## Call: gamlssML(formula = polyp_clean$num_ruffled, family = "WEI")
##
## Mu Coefficients:
## [1] 2.947
## Sigma Coefficients:
## [1] 0.529
##
## Degrees of Freedom for the fit: 2 Residual Deg. of Freedom 58
## Global Deviance: 438.837
## AIC: 442.837
## SBC: 447.026
```

### Step 2: Building the Model

```
mod_countwttotal_ruffled <- gamlss(num_ruffled ~ treatment * collection_day +
  re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = WEI(),
  data = polyp_clean, control = gamlss.control(n.cyc = 60))
```

### Step 3: Summarizing the Model

```
summary(mod_countwttotal_elongation)
```

```
## *****  
## Family: c("NBII", "Negative Binomial type II")  
##  
## Call: gamlss(formula = num_elongated ~ treatment * collection_day +  
## re(random = ~1 | jar_code) + re(random = ~1 | total_num),  
## family = NBII(), data = polyp_clean, method = RS(),  
## control = gamlss.control(n.cyc = 200))  
##  
## Fitting method: RS()  
##  
## -----  
## Mu link function: log  
## Mu Coefficients:  
##  
## Estimate Std. Error t value Pr(>|t|)  
## (Intercept) 1.09720 0.28047 3.912 0.000269 ***  
## treatmentCaffeine -0.17935 0.38941 -0.461 0.647033  
## treatmentEstradiol 0.03055 0.37398 0.082 0.935219  
## treatmentCombination 0.11280 0.36307 0.311 0.757292  
## collection_day 0.31578 0.07282 4.336 6.74e-05 ***  
## treatmentCaffeine:collection_day 0.11929 0.09911 1.204 0.234253  
## treatmentEstradiol:collection_day 0.10750 0.09556 1.125 0.265833  
## treatmentCombination:collection_day 0.14295 0.09252 1.545 0.128455  
## ---  
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1  
##  
## -----  
## Sigma link function: log  
## Sigma Coefficients:  
## Estimate Std. Error t value Pr(>|t|)  
## (Intercept) -1.4610 0.9872 -1.48 0.145  
##  
## -----  
## NOTE: Additive smoothing terms exist in the formulas:  
## i) Std. Error for smoothers are for the linear effect only.  
## ii) Std. Error for the linear terms maybe are not accurate.  
## -----  
## No. of observations in the fit: 60  
## Degrees of Freedom for the fit: 8.35727  
## Residual Deg. of Freedom: 51.64273  
## at cycle: 43  
##  
## Global Deviance: 316.6155  
## AIC: 333.33  
## SBC: 350.833  
## *****
```

# Polyp Asexual Budding

## Step 1: Fitting Distributions for Buds Data

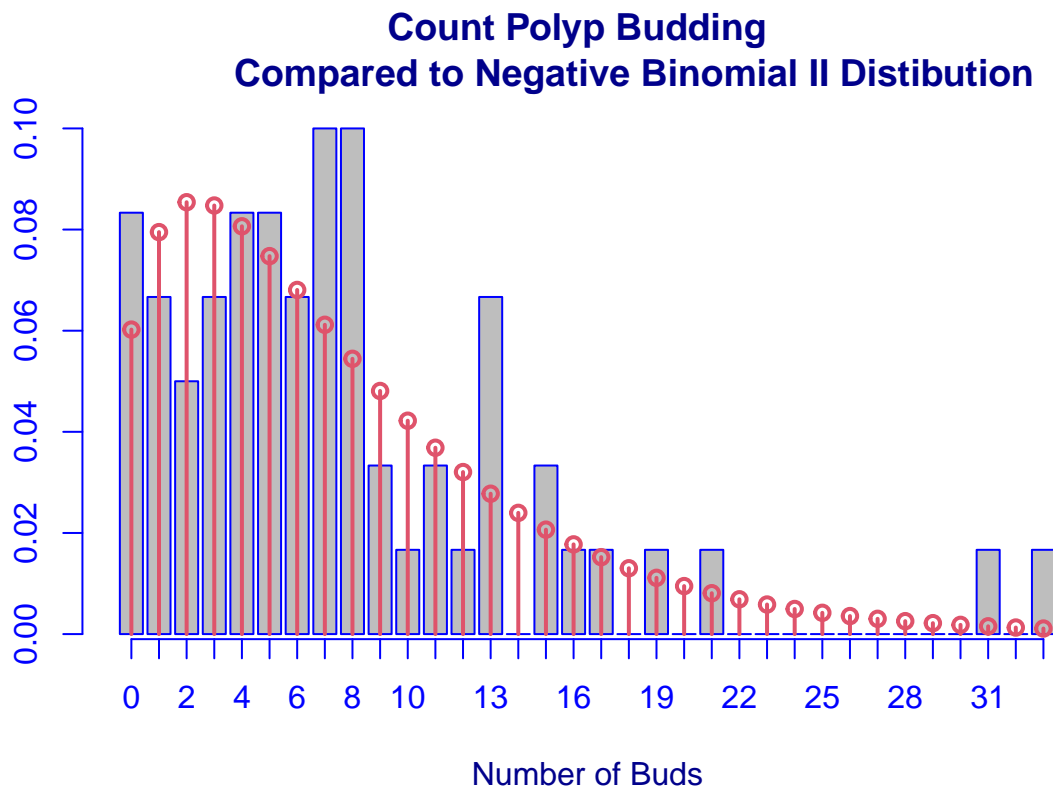
```
fitDist(num_asexual_buds, data = polyp_clean, type = "counts", try.gamlss = T)
```

```
## |
## Lapack routine dgesv: system is exactly singular: U[1,1] = 0 |
## | |=====
## Lapack routine dgesv: system is exactly singular: U[2,2] = 0 |
## | |=====

##
## Family: c("NBII", "Negative Binomial type II")
## Fitting method: "nlminb"
##
## Call: gamlssML(formula = y, family = DIST[i])
##
## Mu Coefficients:
## [1] 2.041
## Sigma Coefficients:
## [1] 1.575
##
## Degrees of Freedom for the fit: 2 Residual Deg. of Freedom 58
## Global Deviance: 368.321
## AIC: 372.321
## SBC: 376.509
```

```
# best fit: Negative Binomial type II (AIC = 372.321)
```

```
# visualizing this fit
histDist(polyp_clean$num_asexual_buds, "NBII", density = T, main = "Count Polyp Budding
  Compared to Negative Binomial II Distribution",
  xlab = "Number of Buds")
```



## Step 2: Building the Model

```
mod_countwttotal_buds <- gamlss(num_asexual_buds ~ treatment * collection_day +
  re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = NBII(),
  data = polyp_clean, control = gamlss.control(n.cyc = 250))
```

## Step 3: Summarizing the Model

```
summary(mod_countwttotal_buds)
```

```
## *****
## Family: c("NBII", "Negative Binomial type II")
##
## Call: gamlss(formula = num_asexual_buds ~ treatment * collection_day +
##   re(random = ~1 | jar_code) + re(random = ~1 | total_num),
##   family = NBII(), data = polyp_clean, control = gamlss.control(n.cyc = 250))
##
##
## Fitting method: RS()
##
## -----
```



```
## Mu link function: log
## Mu Coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -0.54561    0.44010  -1.240  0.22097
## treatmentCaffeine      0.79621    0.54887   1.451  0.15326
## treatmentEstradiol     0.17758    0.57850   0.307  0.76017
## treatmentCombination    1.66274    0.50878   3.268  0.00198 **
## collection_day         0.60862    0.10430   5.835  4.2e-07 ***
## treatmentCaffeine:collection_day -0.08091    0.13120  -0.617  0.54027
## treatmentEstradiol:collection_day  0.06207    0.13640   0.455  0.65108
## treatmentCombination:collection_day -0.26110    0.12315  -2.120  0.03908 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## -----
## Sigma link function: log
## Sigma Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -2.526    2.645  -0.955  0.344
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## -----
## No. of observations in the fit: 60
## Degrees of Freedom for the fit: 11.04057
##      Residual Deg. of Freedom: 48.95943
##              at cycle: 8
##
## Global Deviance:    271.739
##      AIC:          293.8201
##      SBC:          316.9429
## *****
```

## Polyp Total Number

### Step 1: Fitting Distributions for Total Data

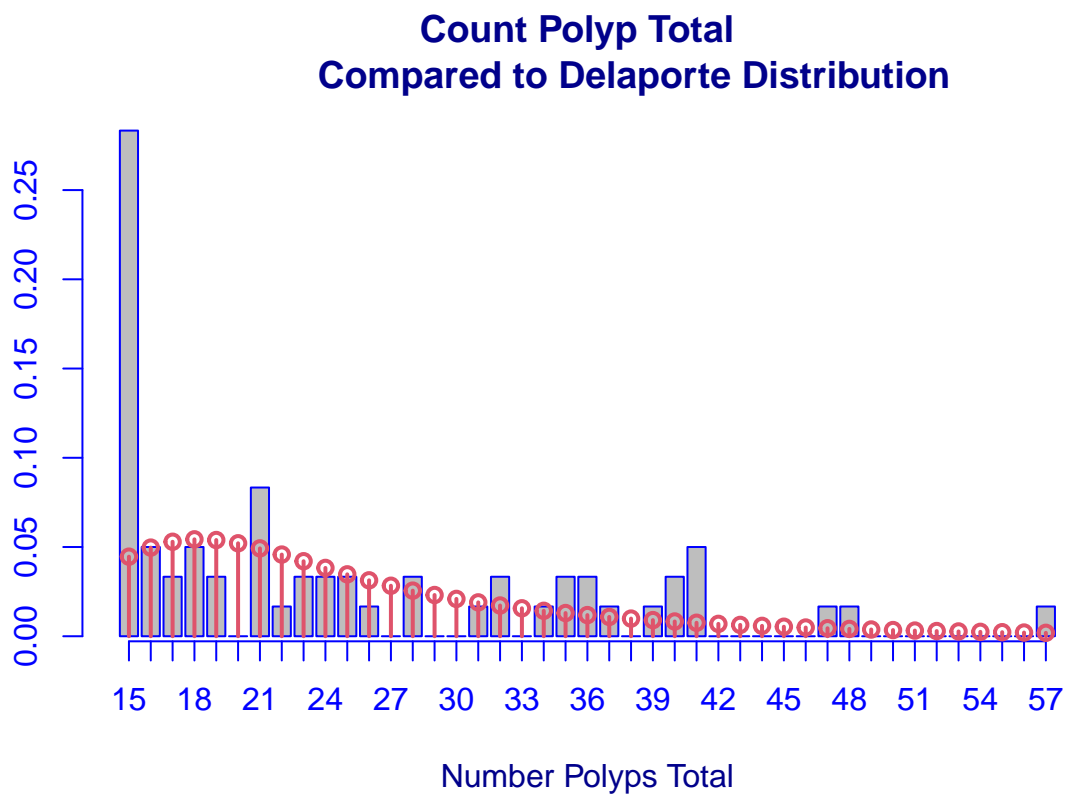
```
fitDist(total_num, data = polyp_clean, type = "counts", try.gamlss = T)
```

```
## |
## system is computationally singular: reciprocal condition number = 9.17006e-23
## |=====
##
## Family: c("DEL", "Delaporte")
## Fitting method: "nlminb"
##
## Call: gamlssML(formula = y, family = DIST[i])
##
```

```
## Mu Coefficients:
## [1] 3.208
## Sigma Coefficients:
## [1] 0.2898
## Nu Coefficients:
## [1] 0.5139
##
## Degrees of Freedom for the fit: 3 Residual Deg. of Freedom 57
## Global Deviance: 430.754
## AIC: 436.754
## SBC: 443.038
```

```
# best fit: delaporte (AIC = 436.754)
```

```
# visualizing this fit
histDist(polyp_clean$total_num, "DEL", density = T, main = "Count Polyp Total
Compared to Delaporte Distribution",
xlab = "Number Polyps Total")
```



## Step 2: Building the Model

```
mod_total <- gamlss(total_num ~ treatment * collection_day + re(random = ~1 |
  jar_code), family = DEL(), data = polyp_clean, control = gamlss.control(n.cyc = 60))
```

### Step 3: Summarizing the Model

```
summary(mod_total)
```

```
## *****
## Family:  c("DEL", "Delaporte")
##
## Call:
## gamlss(formula = total_num ~ treatment * collection_day + re(random = ~1 |
##     jar_code), family = DEL(), data = polyp_clean, control = gamlss.control(n.cyc = 60))
##
## Fitting method: RS()
## -----
## Mu link function:  log
## Mu Coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      2.364608   0.356013   6.642 2.33e-08 ***
## treatmentCaffeine -0.011063   0.522953  -0.021  0.9832
## treatmentEstradiol  0.005354   0.469892   0.011  0.9910
## treatmentCombination 0.048451   0.429514   0.113  0.9106
## collection_day      0.223758   0.084778   2.639  0.0111 *
## treatmentCaffeine:collection_day 0.050239   0.128599   0.391  0.6977
## treatmentEstradiol:collection_day -0.005437   0.111598  -0.049  0.9613
## treatmentCombination:collection_day 0.069381   0.105428   0.658  0.5135
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## -----
## Sigma link function:  log
## Sigma Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -36.04   12909.94  -0.003   0.998
##
## -----
## Nu link function:  logit
## Nu Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)         0     12910         0         1
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms may not be reliable.
## -----
## No. of observations in the fit:  60
## Degrees of Freedom for the fit:  12.73762
## Residual Deg. of Freedom:  47.26238
## at cycle:  5
##
## Global Deviance:    318.8629
## AIC:                344.3381
```

```
##                      SBC:      371.0151
## *****
```

## Is There A Difference Between Caffeine and Combo in Ruffling?

### Step 1: Reordering Ruffling Data To Compare to Caffeine

```
ruffledcomparisons_data <- polyp_data %>%
  rename(collection_day = "Data collection day", jar_code = "Jar Code (ex. E1)",
    treatment = Chemical, num_elongated = "Elongation?", num_ruffled = "Ruffled?",
    num_aseexual_buds = "Asexual Repro?", total_num = Total) %>%
  mutate(jar_code = as.factor(jar_code), treatment = as.factor(treatment)) %>%
  dplyr::select(collection_day, jar_code, treatment, num_elongated, num_ruffled,
    num_aseexual_buds, total_num)
```

### Step 2: Building the Reordered Ruffled Model

```
mod_countwttotal_ruffled_reordered <- gamlss(num_ruffled ~ treatment * collection_day +
  re(random = ~1 | jar_code) + re(random = ~1 | total_num), family = WEI(),
  data = ruffledcomparisons_data, control = gamlss.control(n.cyc = 250))
```

### Step 3: Summarizing the Model

```
summary(mod_countwttotal_ruffled_reordered)
```

```
## *****
## Family:  c("WEI", "Weibull")
##
## Call:  gamlss(formula = num_ruffled ~ treatment * collection_day +
##   re(random = ~1 | jar_code) + re(random = ~1 | total_num),
##   family = WEI(), data = ruffledcomparisons_data,
##   control = gamlss.control(n.cyc = 250))
##
## Fitting method: RS()
##
## -----
## Mu link function:  log
## Mu Coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      1.703055   0.133629  12.745 < 2e-16 ***
## treatmentCombination  0.031960   0.204865   0.156  0.8767
## treatmentControl    -0.471997   0.207627  -2.273  0.0278 *
## treatmentEstradiol  -0.368531   0.204012  -1.806  0.0775 .
## collection_day       0.378951   0.039966   9.482 2.48e-12 ***
## treatmentCombination:collection_day  0.007854   0.063151   0.124  0.9016
## treatmentControl:collection_day  0.063279   0.064249   0.985  0.3299
## treatmentEstradiol:collection_day  0.054333   0.062840   0.865  0.3918
```

```

## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## -----
## Sigma link function:  log
## Sigma Coefficients:
##           Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.5480      0.1053   14.7  <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## -----
## NOTE: Additive smoothing terms exist in the formulas:
## i) Std. Error for smoothers are for the linear effect only.
## ii) Std. Error for the linear terms maybe are not accurate.
## -----
## No. of observations in the fit:  60
## Degrees of Freedom for the fit:  14.64746
##      Residual Deg. of Freedom:  45.35254
##                      at cycle:  3
##
## Global Deviance:      319.673
##           AIC:        348.9679
##           SBC:        379.6448
## *****

```