

MSc Computational Science  
joint programme UvA/VU



# MODELLING DIFFUSIVE SIGNALLING IN *ASPERGILLUS* SPP. GERMINATION INHIBITION



INTERMEDIATE PRESENTATION - MAY

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# OUTLINE



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Considerations for parameter fitting

Inducer assumptions

## 2 Inducer-inhibitor interactions

Overview

Independent induction/inhibition

Inducer-modulated inhibition threshold

Inhibitor-modulated carbon sensitivity

Signal attenuation

Threshold shift

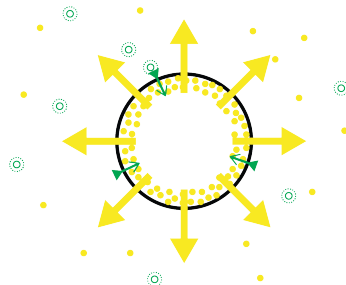
Feedback loop

# INTRODUCTION



## Considerations for parameter fitting

- » There are **two germination-determining mechanisms**:
  - auto-inhibition
  - carbon sensing
- » We have a model proposal for the **auto-inhibition**.
- » But the germination rate is also influenced by **carbon sensing**.
- » **Parameter fitting on germination rate data** requires the germination probability to be expressed in terms of both processes.



# INTRODUCTION



## Inducer assumptions

- » Carbon sources (glucose, amino acids) are **germination inducers** that bind to receptors in the cell wall.
- » Experiments have shown that pyrene [3] and 1-octen-3-ol [2] can easily pass through the hydrophobin layer, but these are **much more hydrophobic** molecules than glucose (positive  $\log K$ ).

carbon source	$\log K$
glucose	-3.2
arginine	-3.88
proline	-2.55
alanine	-0.6

Table: Octanol-water partition coefficient of different carbon sources.

- » Therefore, the rodlet layer **possibly slows down** the access of carbon signals to the receptor proteins.

# INTRODUCTION



## Inducer assumptions

- » Carbon signals are received by **G-proteins** and **RasA** proteins, which are potentially linked to the activation of the cAMP-PKA pathway upon break of dormancy.
- » These proteins are usually membrane-bound but can extend into the inner cell wall polysaccharide layer.
- » Despite their slower intake, the signal thresholds for activating the germination regime may be relatively low.

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# INDUCER-INHIBITOR INTERACTIONS



## Independent induction/inhibition

» The following types of interactions are possible:

- Independent induction/inhibition
- Inducer-modulated inhibition
  - Inducer shifts inhibition threshold (1)
  - Inducer increases inhibitor permeation (2)
- Inhibitor-modulated carbon sensitivity
  - Inhibitor attenuates signal strength (3)
  - Inhibitor shifts signal threshold (4)
- Feedback loop
  - (1) and (3)
  - (1) and (4)
  - (2) and (3)
  - (2) and (4)

1. Independent induction/inhibition



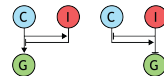
2. Inducer-mediated inhibition threshold



3. Inhibitor-mediated carbon sensitivity



4. Feedback loop



# INDUCER-INHIBITOR INTERACTIONS



## Independent induction/inhibition

- » In the simplest scenario, the two mechanisms can be viewed as **mutually independent**, causing germination when both conditions are fulfilled simultaneously:

$$c_{\text{in}} < c_T \quad \text{and} \quad s c_{\text{cs}} > s_T, \quad (1)$$

- » Like the inhibition threshold, the induction threshold  $s_T$  can be modelled by a normally distributed random variable  $\omega$ .
- » Since the inducer concentrations do not vary in the experiments of interest,  $c_{\text{cs}}$  can be absorbed into the signal strength  $s$ .
- » The combined germination probability is



$$P(\beta < \gamma \cap \omega < s) = \Phi\left(\frac{s - \mu_\omega}{\sigma_\omega}\right) \int_0^\infty \left[1 - \Phi\left(\frac{\beta(\xi) - \mu_\gamma}{\sigma_\gamma}\right)\right] \frac{1}{\sqrt{2\pi}\sigma_\xi} \exp\left(-\frac{(\xi - \mu_\xi)^2}{2\sigma_\xi^2}\right) d\xi. \quad (2)$$



# INDUCER-INHIBITOR INTERACTIONS



## Inducer-modulated inhibition threshold

- » In a different scenario, the germination starts solely upon depletion of the inhibitor, but **the inducer raises or lowers the threshold** at which the inhibitor is considered depleted.
- » As derived previously, the germination probability is

$$P(\beta(\xi) < \gamma) = \int_0^\infty \left[ 1 - \Phi\left(\frac{\beta(\xi) - \mu_\gamma}{\sigma_\gamma}\right) \right] \frac{1}{\sqrt{2\pi}\sigma_\xi} \exp\left(-\frac{(\xi - \mu_\xi)^2}{2\sigma_\xi^2}\right) d\xi, \quad (3)$$



where  $\gamma$  is a normally distributed random variable proportional to the signalling strength.

# INDUCER-INHIBITOR INTERACTIONS

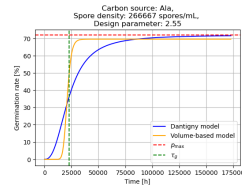
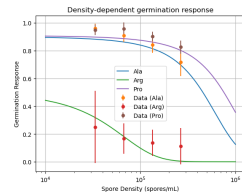


## Inducer-modulated inhibition threshold

- » Fitting this model to experimental data [1] yields

inducer	$P_{\text{eff}}$	$\mu_{\gamma}$	$\sigma_{\gamma}$
Ala	$4.4 \times 10^{-8} \text{ cm s}^{-1}$	$3.65572 \times 10^{-5}$	$2.84791 \times 10^{-5}$
Arg	$1.99 \times 10^{-8} \text{ cm s}^{-1}$	$1.03301 \times 10^{-20}$	$6.23048 \times 10^{-5}$
Pro	$6.36 \times 10^{-8} \text{ cm s}^{-1}$	$6.07702 \times 10^{-5}$	$4.57027 \times 10^{-5}$

- » The model fit is relatively aligned with the data and resembles the **time-dependent Dantigny curve**, but with a much steeper onset.
- » Here we assume that the carbon signal strength is **constant over time**.
- » The permeation constant changes slightly, so **permeation might be affected by the inducer** as well.



# INDUCER-INHIBITOR INTERACTIONS



## Inhibitor-modulated carbon sensitivity

- » Another possibility is that the **inhibitor suppresses the carbon signalling pathway**, which determines germination.
- » Once the inhibition is removed through diffusion, the cAMP-PKA pathway may receive stronger triggers that can switch the germination regime on.
- » The inhibitor may act on the carbon signalling pathway by
  - attenuating the signal strength,
  - shifting the signal threshold.



# INDUCER-INHIBITOR INTERACTIONS



## Inhibitor-modulated carbon sensitivity - **signal attenuation**

- » The inhibitor may attenuate the carbon signal strength by a Hill-type factor:

$$s(c_{\text{in}}) = s_{\text{max}} \frac{1}{1 + \left(\frac{c_{\text{in}}}{K_I}\right)^n}, \quad (4)$$

where  $K_I$  a half-saturation constant and  $n$  is a Hill exponent. The signal strength maximum,  $s_{\text{max}}$ , can be considered constant for simplicity (the threshold absorbs the variation).

- » The germination probability for a specific spore is thus

$$P(\omega < s \mid c_{\text{in}}) = \Phi\left(\frac{s(c_{\text{in}}) - \mu_\omega}{\sigma_\omega}\right). \quad (5)$$

# INDUCER-INHIBITOR INTERACTIONS



## Inhibitor-modulated carbon sensitivity - **signal attenuation**

» The two random variables in  $c_{\text{in}}$  are  $\xi$  and  $\psi$ . Therefore, the full CDF is

$$P(\omega < s) = \int_0^\infty \int_0^\infty \Phi \left( \frac{s_{\text{max}} \left[ 1 + \left( \frac{c_{\text{in}}(\xi, \psi)}{K_I} \right)^n \right]^{-1} - \mu_\omega}{\sigma_\omega} \right) d\xi d\psi, \quad (6)$$

where

$$c_{\text{in}}(\xi, \psi) = \left[ \phi + (1 - \phi) e^{-\frac{\xi}{\tau(1-\phi)}} \right] \psi. \quad (7)$$



## Inhibitor-modulated carbon sensitivity - **threshold shift**

- » In the second case, the threshold is linearly shifted from a randomly fluctuating baseline  $\omega_0$  by a factor  $k$ :

$$\omega(c_{\text{in}}) = \omega_0 + kc_{\text{in}}. \quad (8)$$

- » The condition for germination can then be rewritten from  $s > \omega_0 + kc_{\text{in}}$  to  $s - kc_{\text{in}} > \omega_0$ .
- » This yields the spore-specific germination probability:

$$P(\omega_0 < s - kc_{\text{in}} \mid c_{\text{in}}) = \Phi\left(\frac{s - kc_{\text{in}} - \mu_{\omega_0}}{\sigma_{\omega_0}}\right) \quad (9)$$

and the general probability

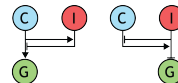
$$P(\omega < s) = \int_0^\infty \int_0^\infty \Phi\left(\frac{s - kc_{\text{in}} - \mu_{\omega_0}}{\sigma_{\omega_0}}\right) d\xi d\psi. \quad (10)$$

# INDUCER-INHIBITOR INTERACTIONS



## Feedback loop

- » Finally, a scenario may exist in which the inhibitory mechanism and the signalling pathway are **mutually dependent**.
- » For example, a **strengthened cAMP-PKA pathway** may gradually **lift the inhibition threshold**, while a **reduction in the inhibitor concentration** may **reinforce the carbon signalling**.
- » The germination-determining threshold may either be dependent on carbon signal or on the depletion of the inhibitor.
- » The ODEs for this systems are yet to be formulated.





- [1] Maryam Ijadpanahsaravi et al. "The impact of inter- and intra-species spore density on germination of the food spoilage fungus *Aspergillus niger*." In: *International journal of food microbiology* 410 (2023), p. 110495. URL: <https://api.semanticscholar.org/CorpusID:265268197>.
- [2] Kana Miyamoto et al. "Formation of 1-octen-3-ol from *Aspergillus flavus* conidia is accelerated after disruption of cells independently of Ppo oxygenases, and is not a main cause of inhibition of germination". In: *PeerJ* 2 (2014). URL: <https://api.semanticscholar.org/CorpusID:8514039>.
- [3] X. Wang et al. "The SC3 hydrophobin self-assembles into a membrane with distinct mass transfer properties." In: *Biophysical journal* 88 5 (2005), pp. 3434–43. URL: <https://api.semanticscholar.org/CorpusID:16739076>.