

Devices for Biological Systems: On-Chip Horizontal Gene Transfer and 3D-Printed Microfluidic Applications

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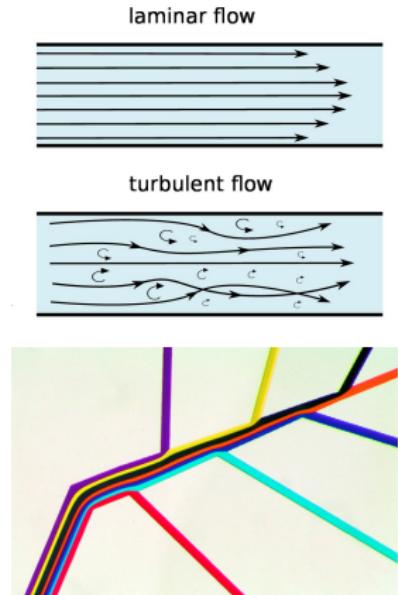
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December 10, 2015

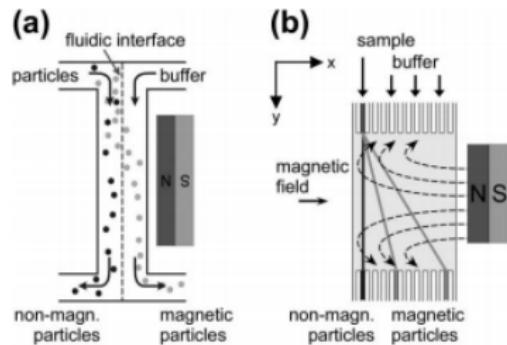
Overview

- ① Introduction of Microfluidics and applications to biological research
- ② Specific Aim 1 : Demonstrate a microfluidic approach for co-encapsulation of two strains of strep to observe single gene transfer events
- ③ Specific Aim 2: Develop an easy to use and fabricate oxygen control insert for a 24-well plate
- ④ Specific Aim 3: Develop a 3D printable micropipette and compare its performance to commercial versions
- ⑤ Conclusion and Thanks

Microfluidics

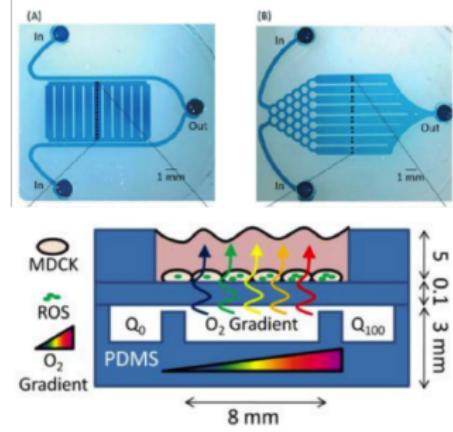
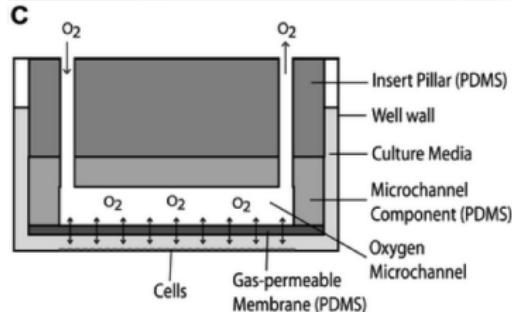
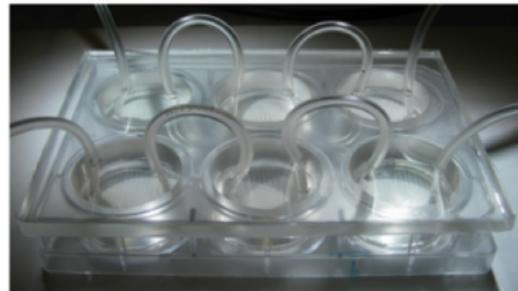


P.J.A. Kenis et al. *Science* 285 (1999): 83-85.



Pamme, *Lab Chip*. 2007

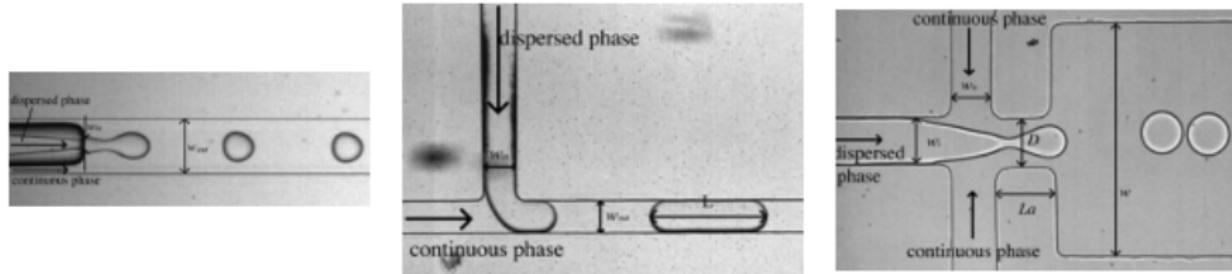
Rapid Diffusion Enables Oxygen Control



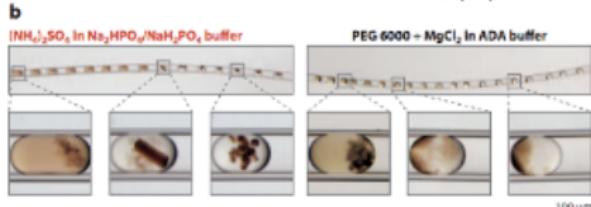
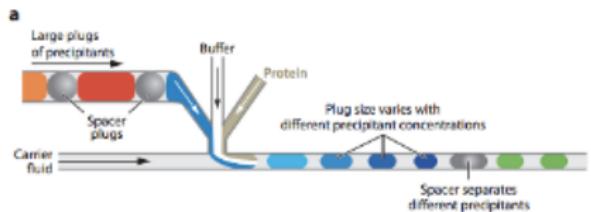
Lo et al. *Lab Chip*, 2010 10(18): 23942401

Oppegard et al. *Plos One* 2009

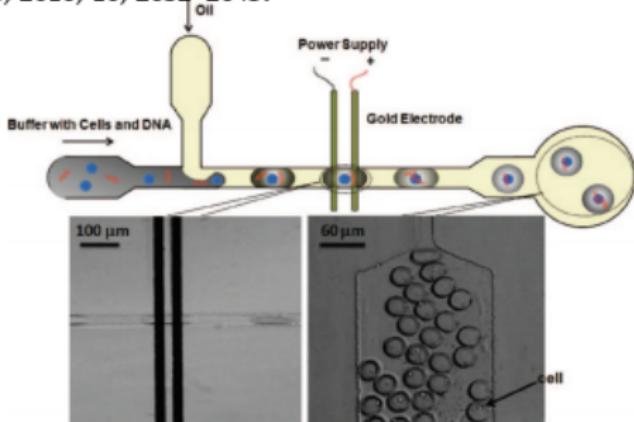
Droplet Microfluidics



Baroud et al. *Lab Chip*, 2010, 10, 2032-2045.



Li and Ismagilov, *Annu. Rev. Biophys.* 2010, 39:139-58



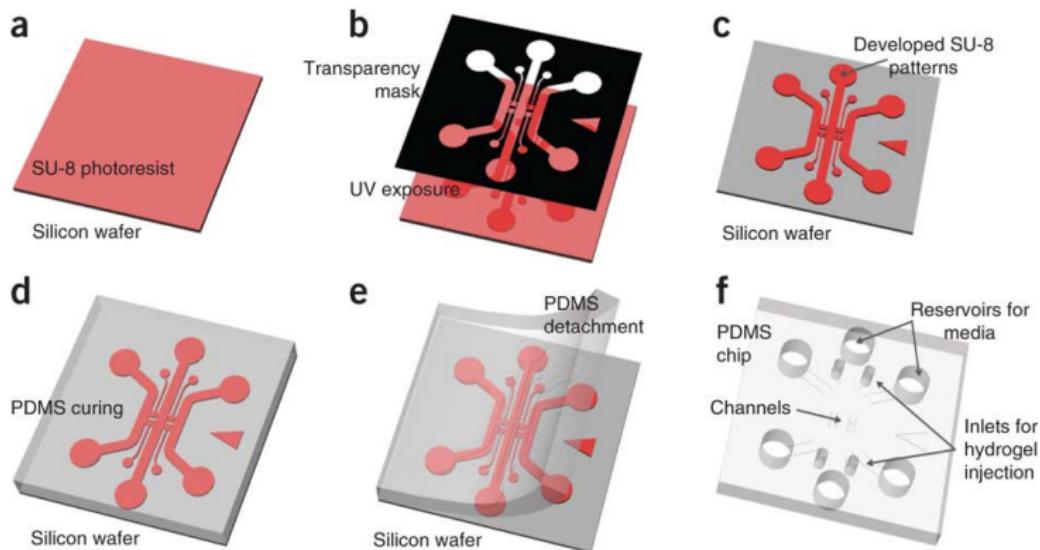
Zhan et al. *Anal. Chem.* 2009, 81, 2027-2031.

Droplet Microfluidics with cells

Mazutis *Nature Protocols* 8, 870891 (2013).

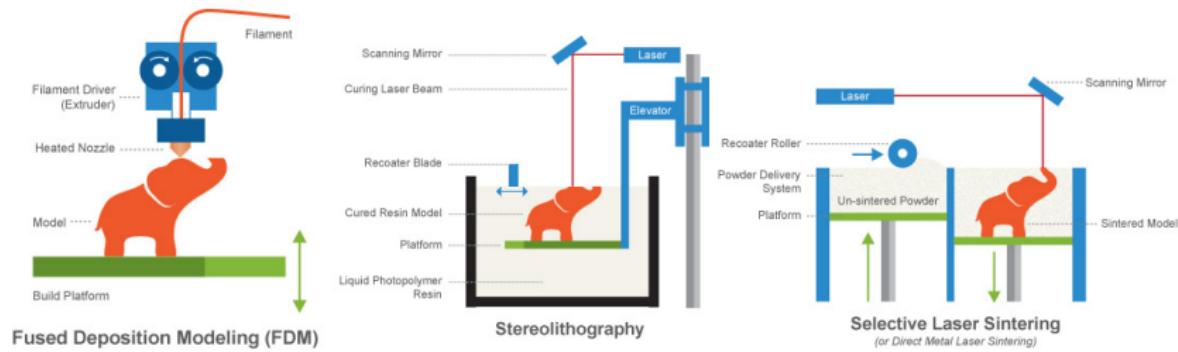


Fabrication of Microfluidic devices - Soft-Photolithography



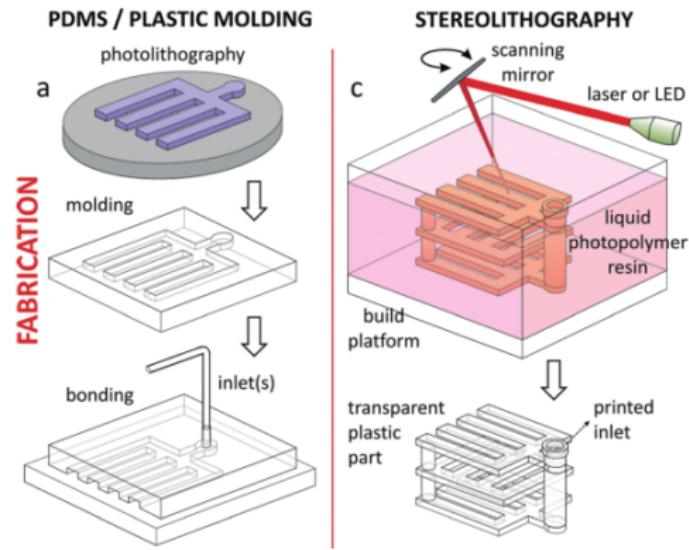
Shin et al. Nature Protocols 7, 12471259 (2012).

3D-Printing - Additive Manufacturing

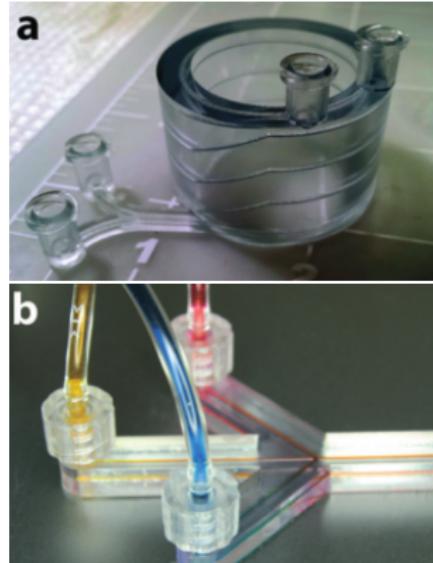


PrintSpace.com

3D-Printing as Alternative to Microfabrication

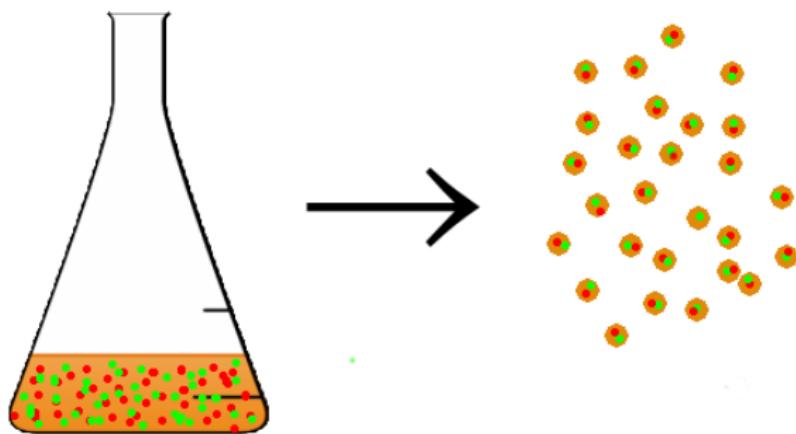


Au et al. Lab Chip. 2014



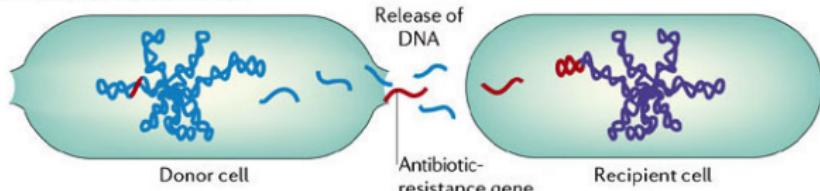
Au et al. Lab Chip. 2014

Tango Device - Co-Encapsulation and Observation of Gene Transfer

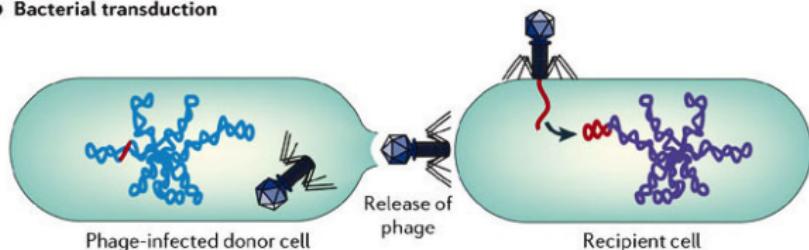


Horizontal Gene Transfer

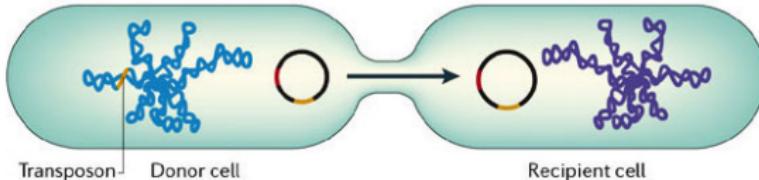
a Bacterial transformation



b Bacterial transduction

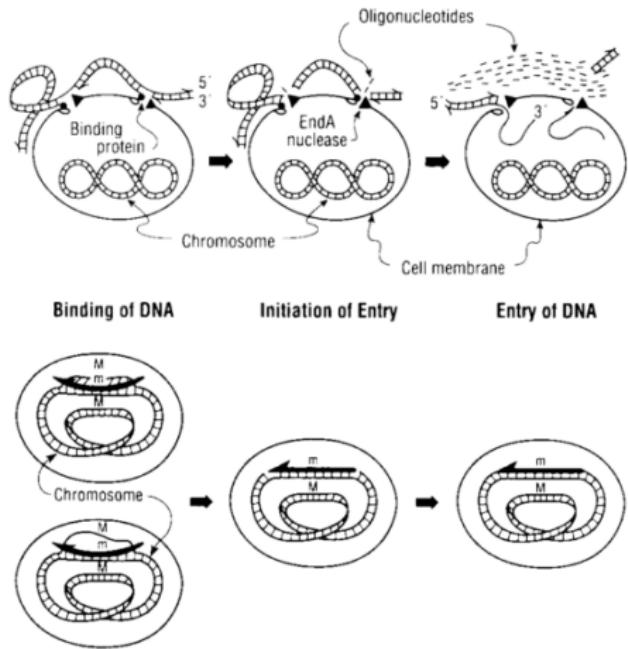


c Bacterial conjugation



Furuya and Lowy. *Nature Reviews Microbiology* 4, 36-45, 2006

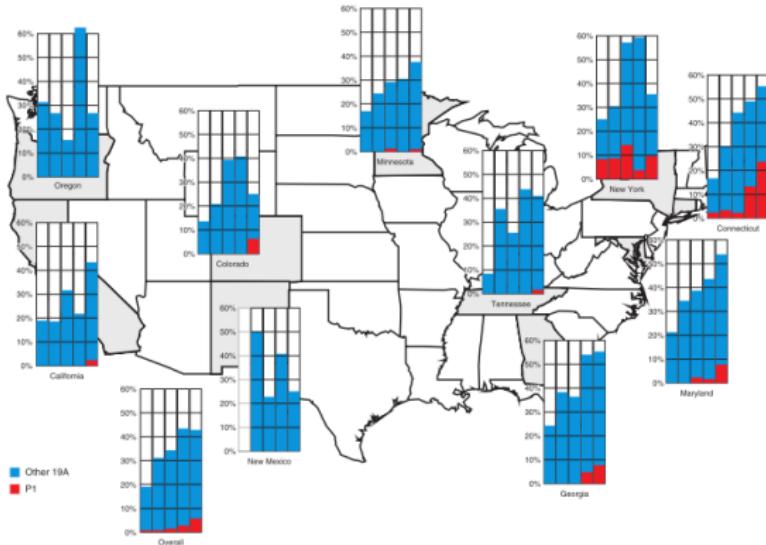
Uptake of Free DNA by a Competent Cell



Lacks. *MICROBIAL DRUG RESISTANCE* Volume 3, Number 4, 1997.

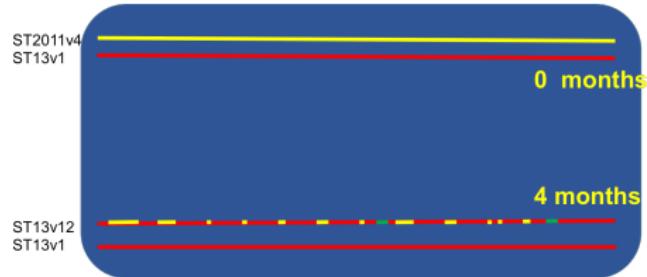
Impact of *S. pneumoniae*

- 14.5 million cases of serious disease, 826,000 deaths, in children less than 5 years
- pneumonia, meningitis and sepsis

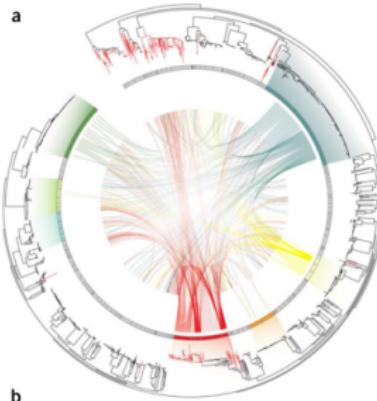


Golubchik. *Nature Genetics*. March 2012, Vol 44, Num 3.

In vivo



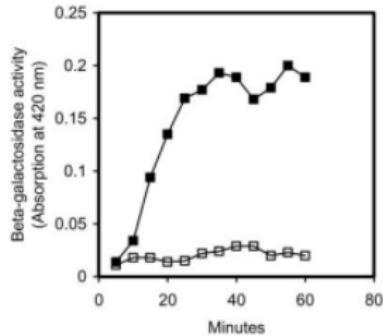
Hiller 2010 (Reproduced by Morrison)



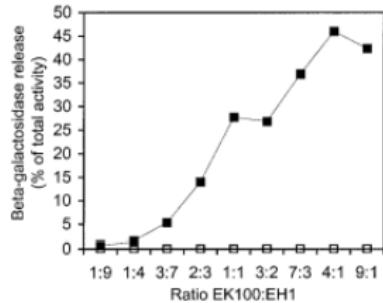
Chewapreecha. *Nature Genetics*. 2014, Vol 46, Num 3.

Competence is induced and involves attack on donor cell

- Competence is CSP concentration dependent, 1-10 ng, 10^7 cells/mL
- Fraction of cells lyse at critical CSP concentration
- Competence-induced cells lyse competence-deficient cells
- Lysis is dependent on activity of CbpD protein



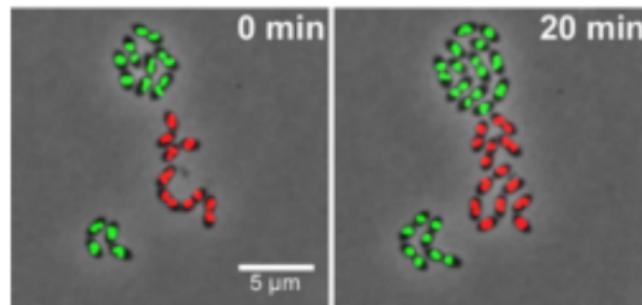
Steinmoen et al. PNAS 2002, Vol. 99, no. 11, 7681-7686



Steinmoen et al. J. Bacteriology, Dec. 2003, p. 71767183

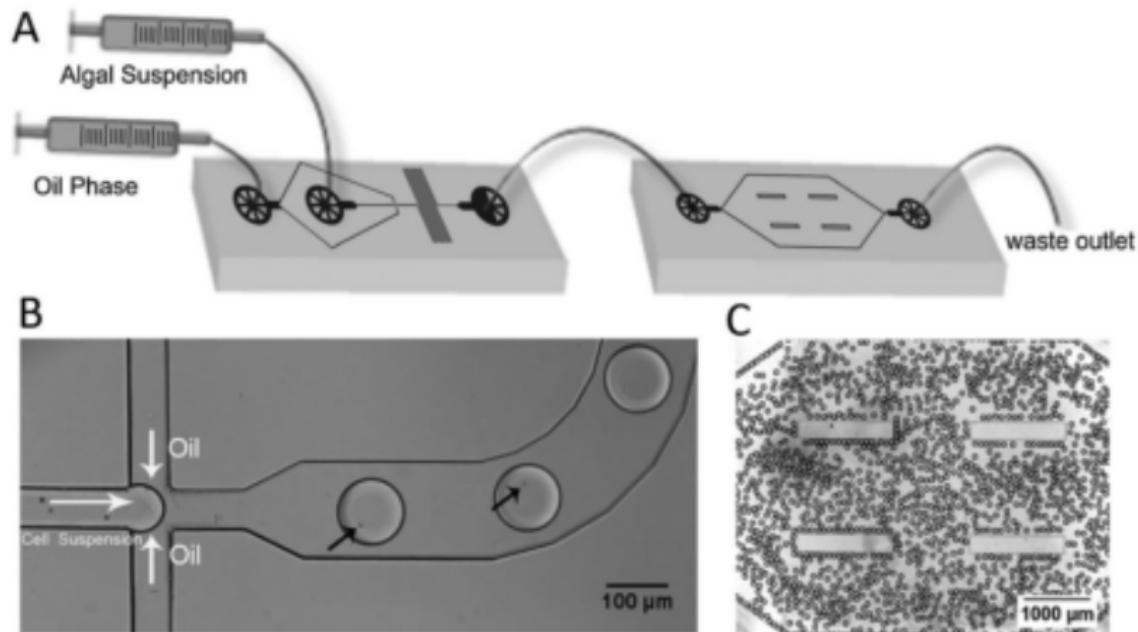
Aim: Co-encapsulation of two strains of *S. pneumoniae* to observe single gene transfer events

- Isolate two cells in a confined area to determine what happens in a single gene transfer
- Attack strain with inducible competence (Does not produce CSP), RFP labeled
- Non-competent victim strain (cannot sense CSP), GFP expressing
- Encapsulate attacker-victim pairs in Droplets



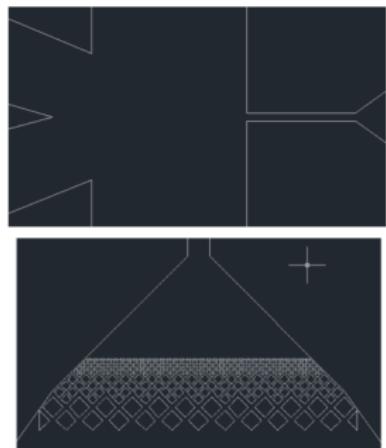
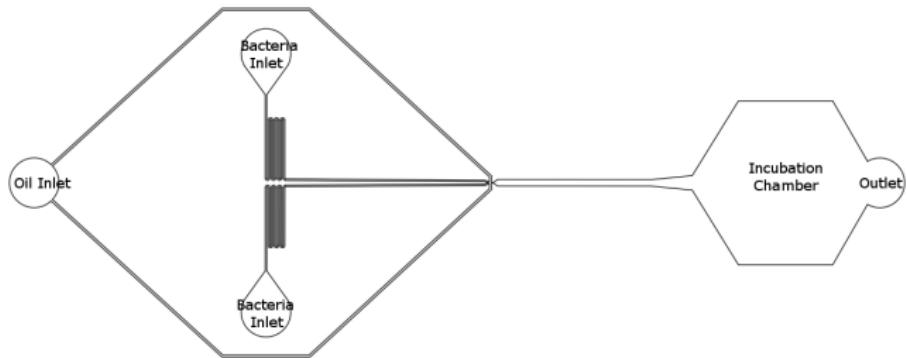
Kjos et al. *Journal of Bacteriology* 2015 Vol 197 No 5.

Inspiration - Algae Encapsulation Device



Pan et al. *Integr. Biol.*, 2011, 3, 10431051

Device Design



Droplet Generation

- Oil flowrate of $150 \mu\text{mL/h}$, aqueous $50 \mu\text{mL/h}$
- Neck width of $5 \mu\text{m}$
- Droplets of $5\text{-}10 \mu\text{m}$ are generated
- 5 femtoliter droplets

Beads in Droplets

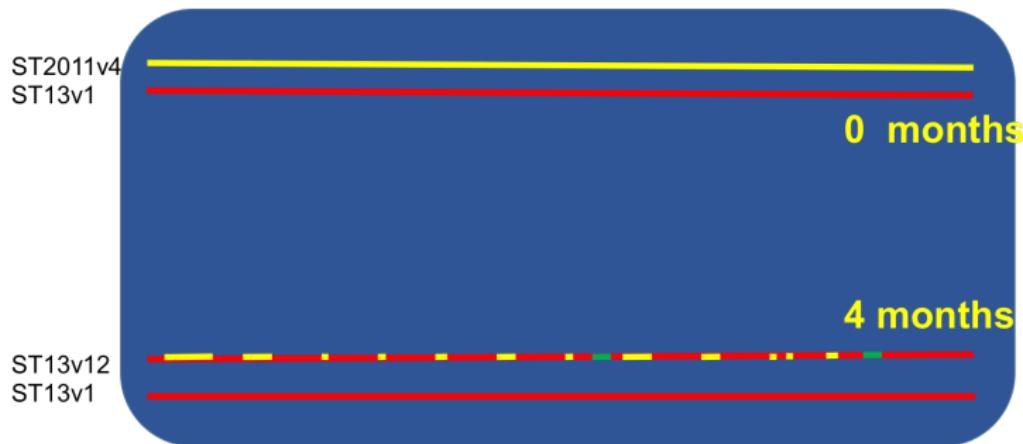
- Green and red fluorescent beads of $1 \mu\text{m}$ diameter
- 4×10^9 beads/ml
- 20% occupancy, 5% R&G

Full Experiment Procedure

- ① Encapsulate attacker and victim strains in droplets
- ② Incubate droplets in the device and observe attack
- ③ Break the emulsion, dilute and plate
- ④ Assess gene transfer with whole genome sequencing of remaining cultures

Expected Results

- Encapsulate cells and determine occupancy
- Image cells during incubation to observe attack
- Ultimately determine what can be expected from a single transfer event

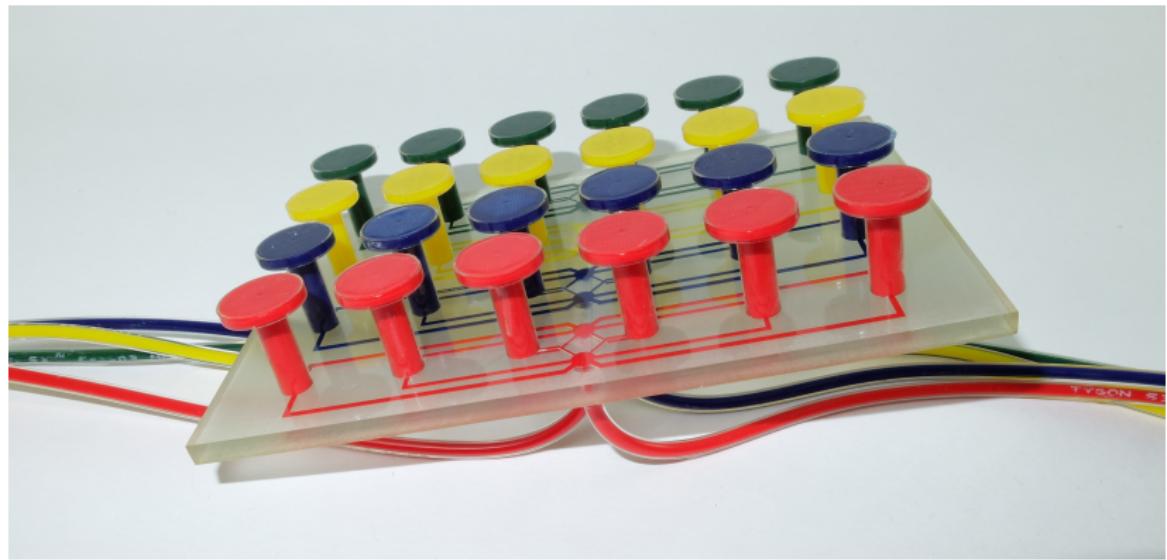


Hiller 2010 (Reproduced by Morrison)

Potential Difficulties

- Cells may not become competent due to encapsulation
- Cells may not physically meet
- Attack or transformation may require more than 2 cells
- Confinement may cause lysis of both cells

3D-Printed Oxygen Control Insert



Oxygen in Vivo Versus Oxygen in Research

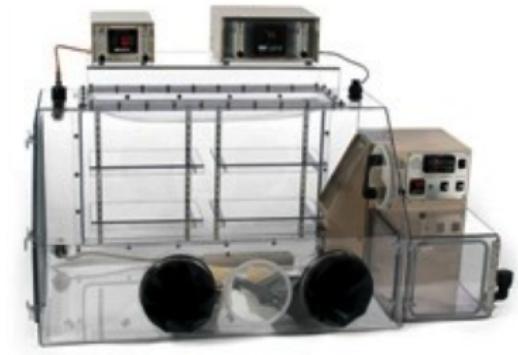
- Typically ignored, culture studied at 21%
- Equipment is inconvenient for oxygenating to multiple levels and is expensive
- Microfluidic oxygen devices must be expertly made and operated

Tissue	Physiologic oxygen (%)
Lung alveoli	13
Liver	10–13
Arterial blood	10–13
Venous blood	5
Bone marrow	0.5–7
Brain	0.5–7
Cartilage	1

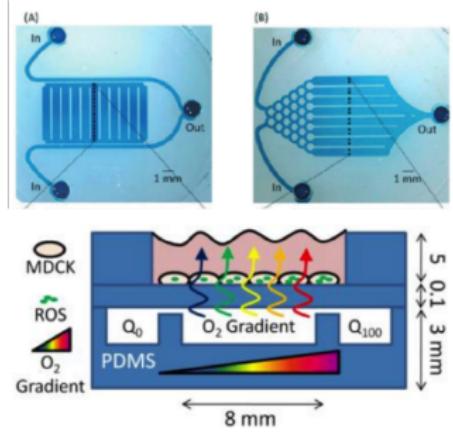
Brennan et al. *Lab Chip*, 2014, 14, 43054318

Methods for Oxygen Control

Hypoxic Workstations



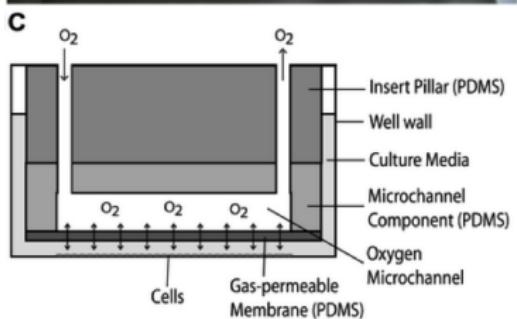
Microfluidic Devices



Lo et al. *Lab Chip*, 2010 10(18): 23942401

Aim: Develop an easy to use and fabricate oxygen control system for researchers

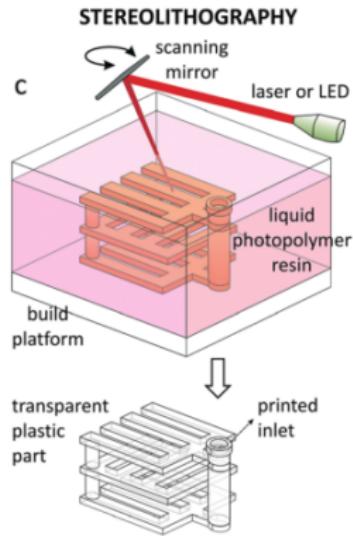
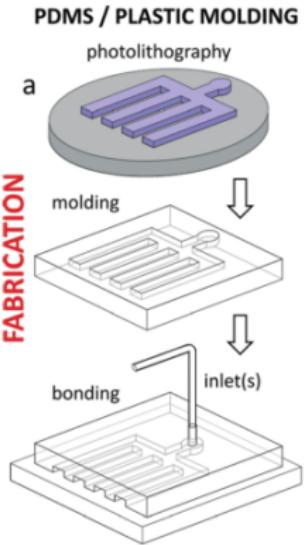
- Use Multi-well plate platform
- 3D-printing - outsource fabrication
- Integrate functionality



Oppegard et al. Plos One 2009

3D-Printing

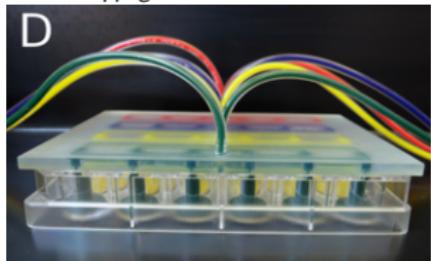
FABRICATION



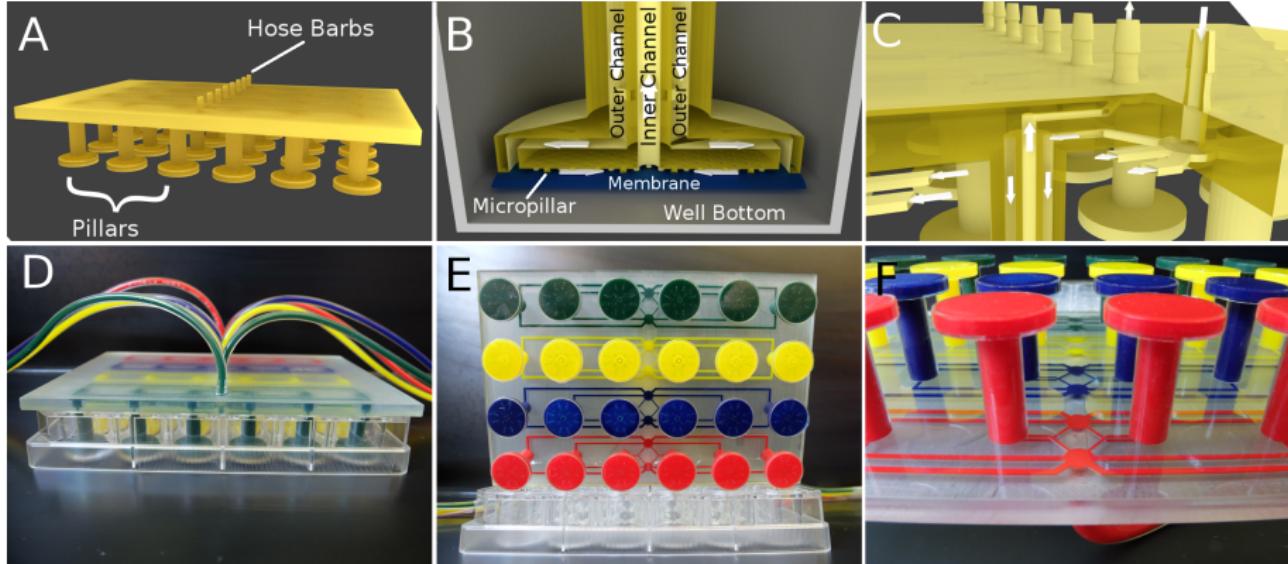
Au et al. Lab Chip. 2014



Oppegard et al. Plos One 2009



Insert Design

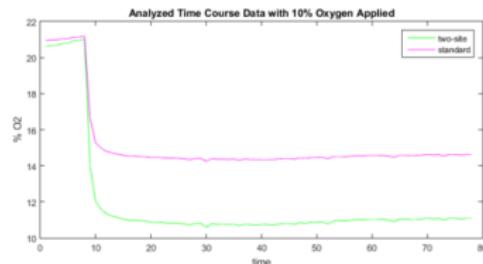
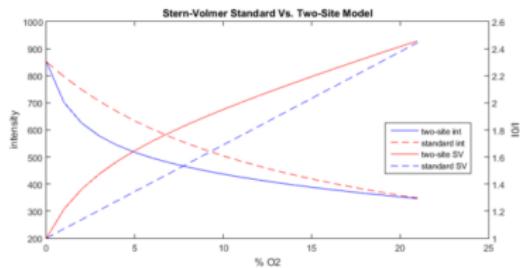
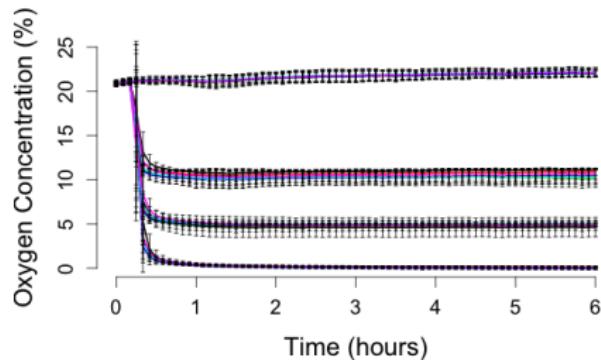


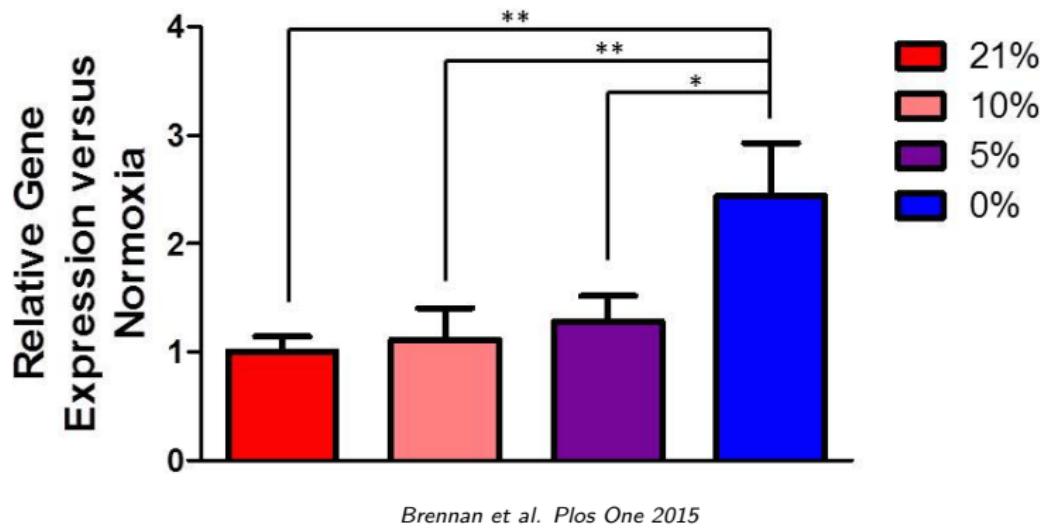
Brennan et al. Plos One 2015

Oxygen Performance - Stern Volmer Analysis

$$I_0/I = 1 + Ksv \cdot [Q]$$

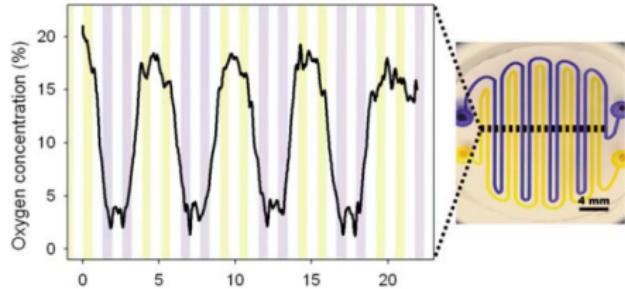
$$I/I_0 = \frac{f_1}{1 + Ksv_1 \cdot [Q]} + \frac{f_2}{1 + Ksv_2 \cdot [Q]}$$





Expanding and improving the device

- Membrane material
- Media exchange
- Patterning of oxygen in wells



Oppegard et al. Plos One 2009

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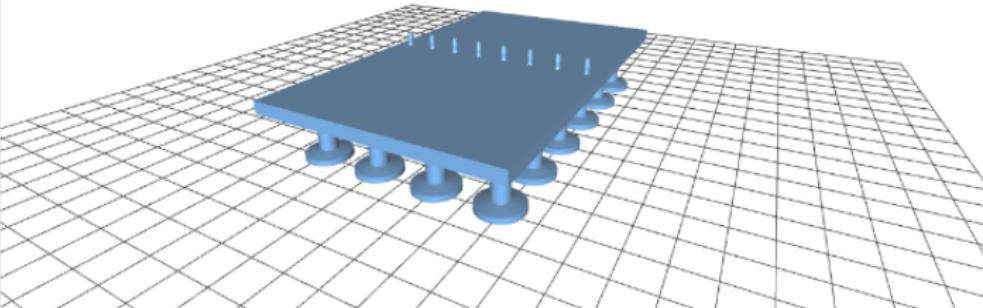
Code Issues 0 Pull requests 0 Wiki Pulse Graphs Settings

Branch: master 3d-printed-oxygen-control-insert / 24-well.stl Find file Copy path

mbrenn3 exported stl 456ae01 on Sep 9

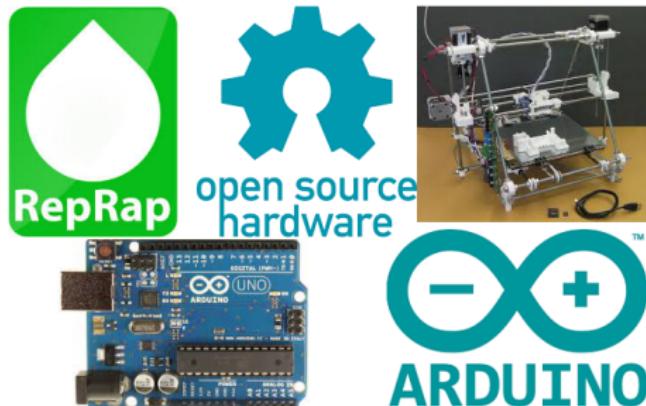
1 contributor

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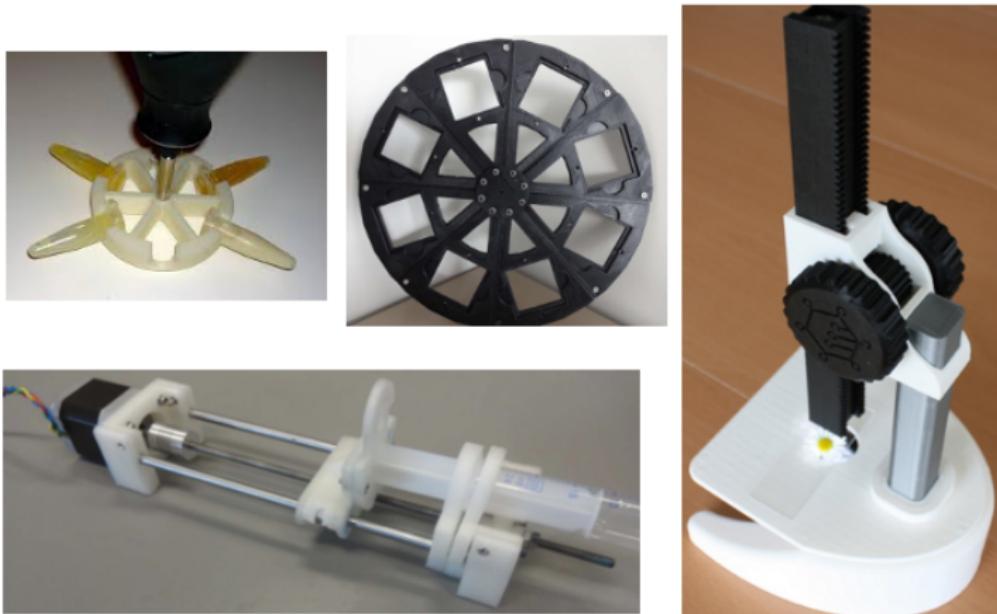


Wireframe Surface Angle Solid

Open Design Scientific Tools



3D-Printed Scientific Tools



Thing:1483 Dremelfuge, Thing:26553 Filterwheel, Thing:77450 Microscope

Wijnen et al. 2014, Volume 9, Issue 9, e107216.

3D-Printed Pipettes

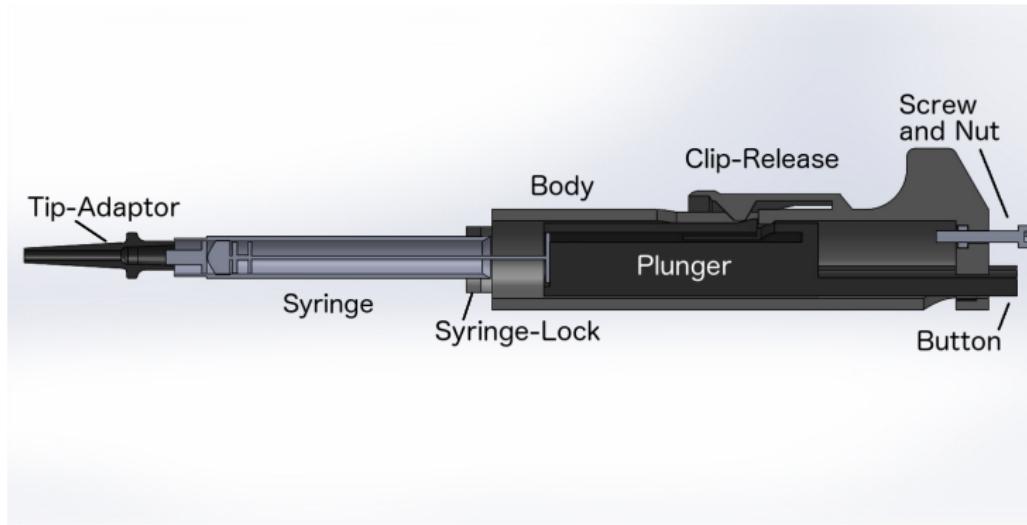


Thing:159052 Laboratory pipette, Thing:25519 Briopipette

Aim: Develop a 3D printable micropipette and compare to commercial versions

- Use a consumer grade printer
- Simple hardware
- Adjustable to desired volume a priori
- Comparable accuracy to a commercial pipette

Design of Pipette



Current Prototype

- Makerbot printable, \$5 USD including parts
- Uses syringe graduations to set volume
- Competitive with commercial pipettes in accuracy



Operating Procedure

- Compare displacement from bottom and top stop with graduations
- Adjust top stop with screw
- Re-check displacement
- Pipette

Preliminary Results

Table 1. Comparison of error between the printed and commercially produced pipette

Volume		Systematic error		Random error	
10 μL	printed	2.17 %	$\pm 0.217\mu\text{L}$	1.96%	$\pm 0.196\mu\text{L}$
	commercial	3%	$\pm 0.3\mu\text{L}$	1%	$\pm 0.1\mu\text{L}$
20 μL	printed	0.33%	$\pm 0.066\mu\text{L}$	1.18 %	$\pm 0.236\mu\text{L}$
	commercial	2.50%	$\pm 0.5\mu\text{L}$	0.70%	$\pm 0.14\mu\text{L}$
50 μL	printed	1.17%	$\pm 0.585\mu\text{L}$	0.50%	$\pm 0.25\mu\text{L}$
	commercial	1%	$\pm 0.5\mu\text{L}$	0.30%	$\pm 0.15\mu\text{L}$
200 μL	printed	2.52%	$\pm 5.04\mu\text{L}$	0.69%	$\pm 1.38\mu\text{L}$
	commercial	0.6%	$\pm 1.2\mu\text{L}$	0.20%	$\pm 0.4\mu\text{L}$

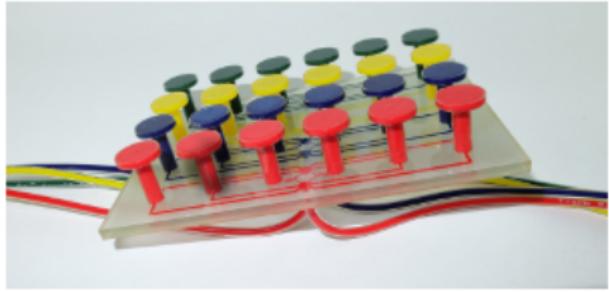
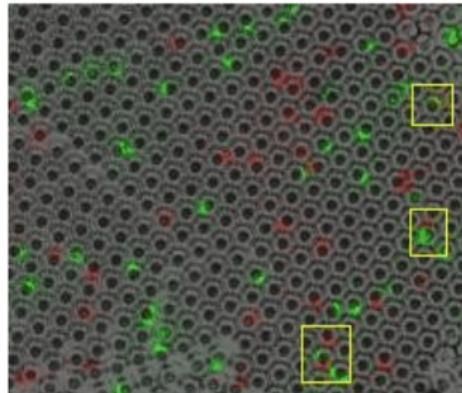
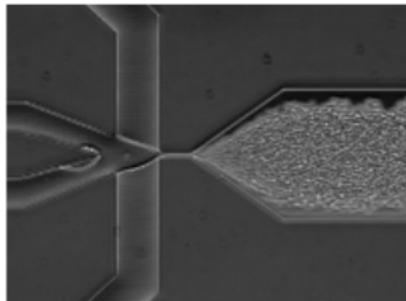
Error assessed for average measured volume by weight of water compared with published specifications of commercial pipette.

Future Approach

- Add features such as ejector for tips
- Push-push operation
- improve user friendliness



Wrap-Up



Acknowledgments

Thank You

- Dr. Morrison - Gene Transfer on a Chip
- Megan - Bioverification PCR
- Fahad - CAD
- Dr. Eddington - Advisor
- Committee

Thank You