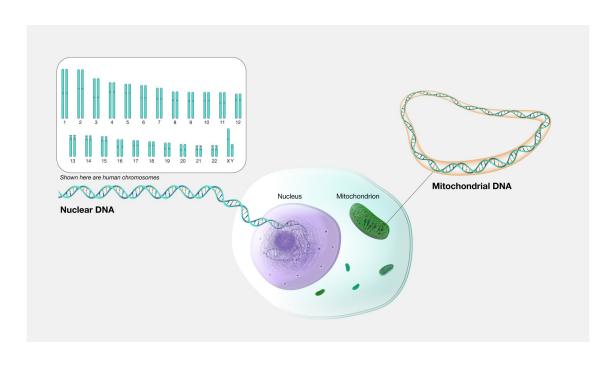
Genome and Sequencing Basics

Genome Biology (BIOL7263) 5Sept24

What is a genome?

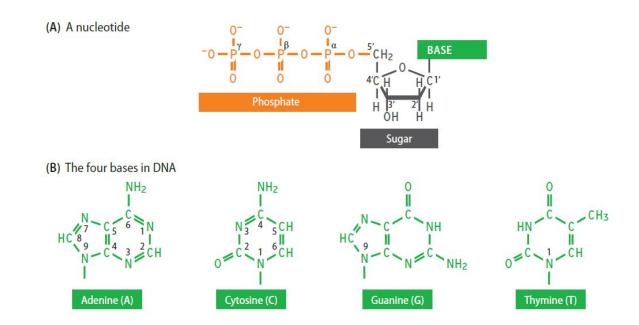
A <u>complete</u> set of <u>genetic</u> information.



What are genomes made of?

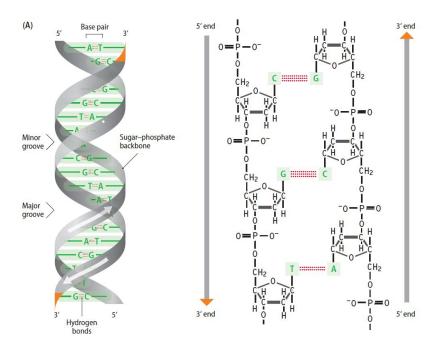
Nucleic Acids

- deoxyribonucleic acid (DNA) most genomes
- ribonucleic acid
 (RNA) some
 viruses



What are genomes made of?

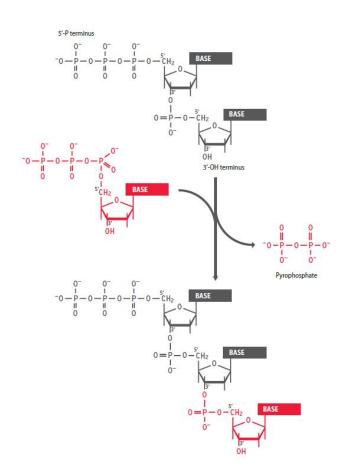
 Nucleotides are assembled into chains (polymers) that are 10s to 10⁶ units long.



What are genomes made of?

 Nucleotides are assembled into chains (polymers) that are 10s to 10⁶ units long.

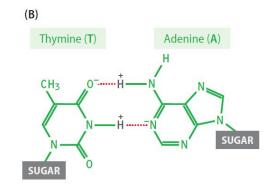
 Nucleotides are linked 5' to 3' through the formation of phosphodiester bonds.

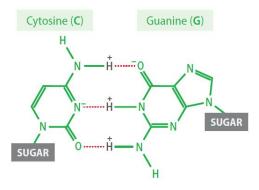


DNA is usually a double helix with specific base-pairing

 Strands held together by hydrogen-bonds.

- G-C have a stronger bond
 - High GC content can cause problems for variety of molecular biology techniques.

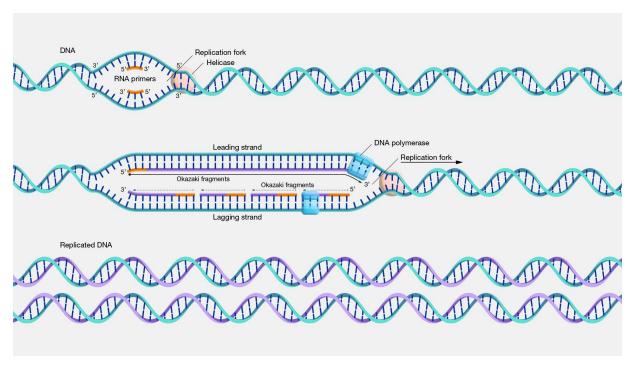




DNA is usually a double helix with specific base-pairing

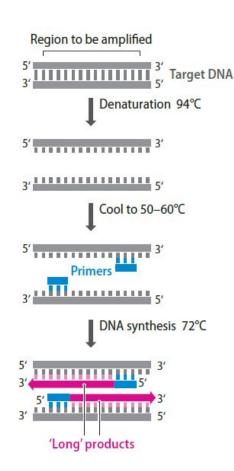
Each strand is entirely dependent of the sequence of the other strand.

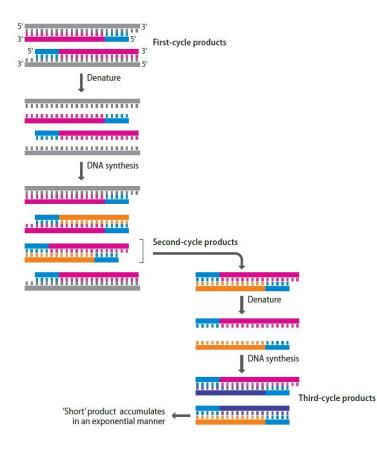
Each strand can serve as template for replication.



The structure of DNA allows for in vitro amplification

Polymerase chain reaction (PCR) is an essential component of most sequencing technologies

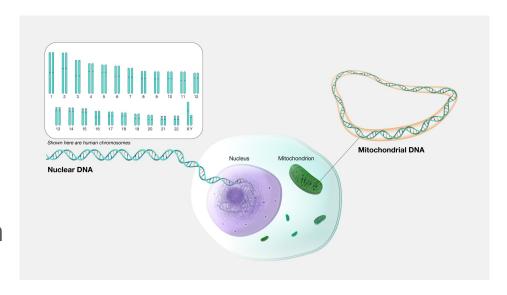




How many genomes?

All eukaryotes carry at least two genomes:

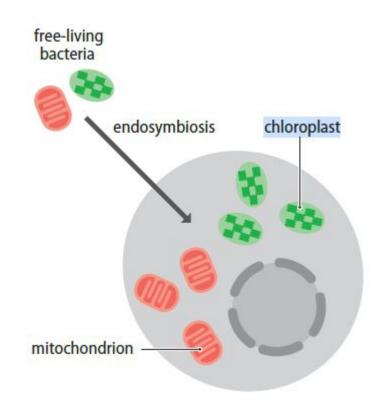
- Nuclear bi-parental inheritance
- Mitochondria maternally inherited - prokaryotic-like in structure



How many genomes?

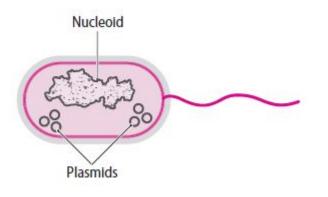
Plants have three genomes:

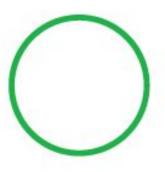
- Nuclear biparental inheritance
- Mitochondria maternally inherited - prokaryotic-like in structure
- chloroplast maternally inherited - prokaryotic-like in structure



How many genomes?

Prokaryotes





chromosome – located in nucleoid, carries essential genes



chromid – uses plasmid partitioning system, carries essential genes



plasmid – uses plasmid partitioning system, carries nonessential genes

Information is primarily encoded in the linear sequence of nucleotides:

Genes:

Portions of the genome the encode instructions to assemble proteins.

 Gene expression is a multi-step process involving transcription and translation ("Central Dogma")

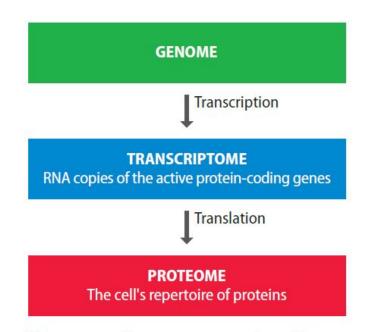
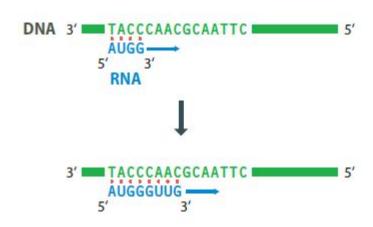


Figure 1.2 Genome expression. The genome specifies the transcriptome, and the transcriptome specifies the proteome.

Translation - DNA to RNA

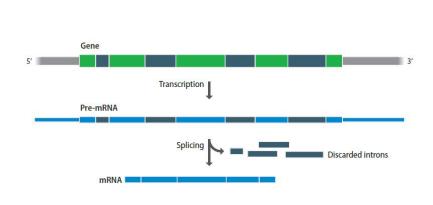


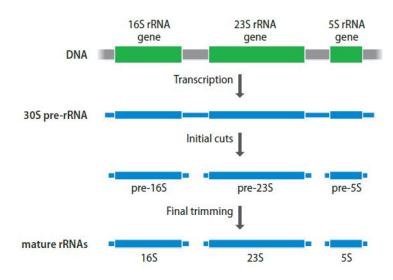
(A) A ribonucleotide

Translation - DNA to RNA

RNA is processed in different ways in different organisms and

organelles



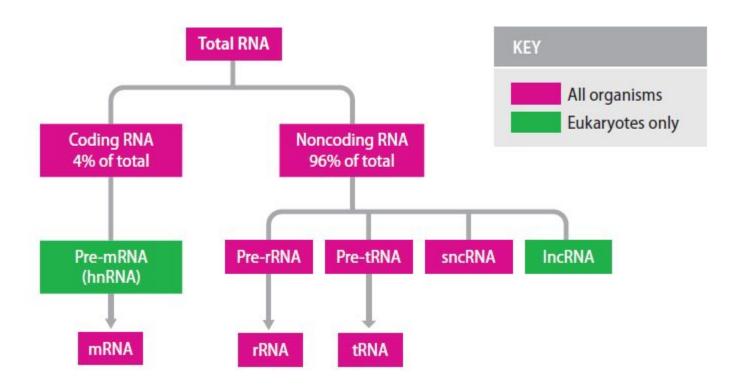


Eukaryotic mRNA

Prokaryotic rRNA

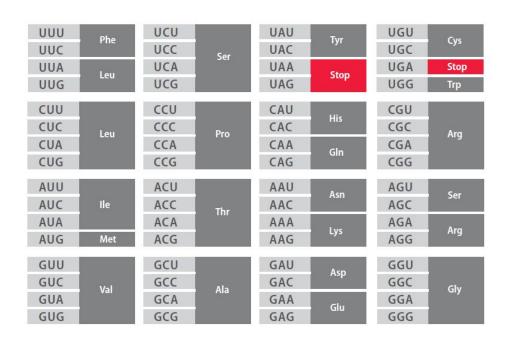
There are many types of RNAs in the cell

Protein coding mRNAs are relatively rare!!

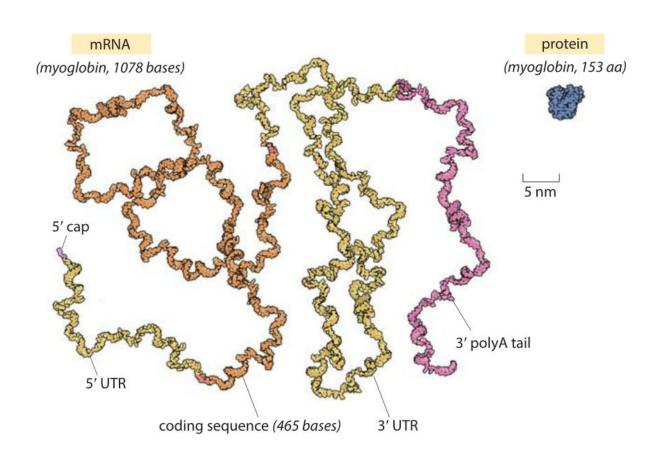


Information is primarily encoded in the linear sequence of nucleotides:

Genetic code is triplicate 3 nucleotides = 1 amino
 acid

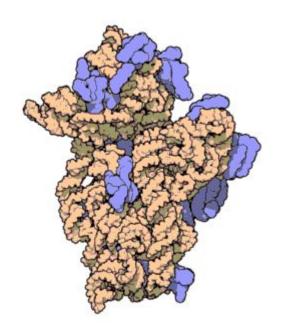


Transcripts are huge! Proteins are small



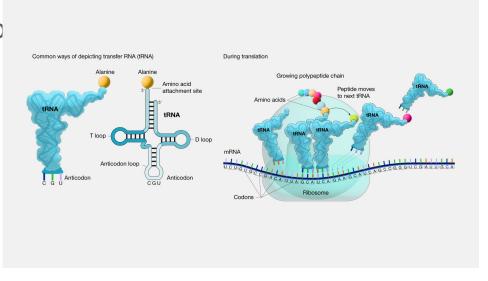
Information is primarily encoded in the linear sequence of nucleotides:

 rRNA - not translated structural and functional elements of the ribosome



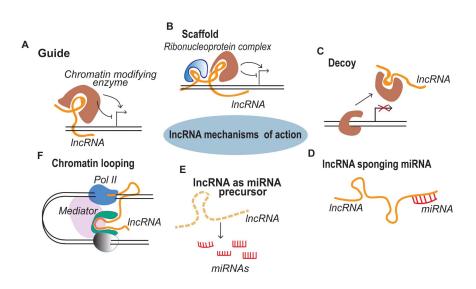
Information is primarily encoded in the linear sequence of nucleotides:

tRNA - not translated - adaption



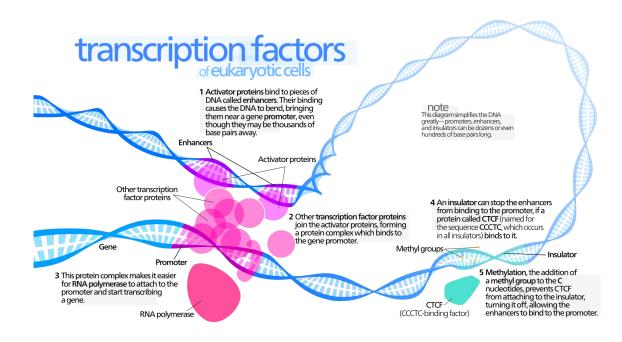
Information is primarily encoded in the linear sequence of nucleotides:

Noncoding RNAs incompletely understood roles
 regulating transcription,
 translation, and cellular
 physiology



Cis-regulatory elements

- Promoters
- Terminators
- Enhancers
- Repressors
- Insulators



Genome Sequencing

Three major approaches

- 1. Chain-termination (Sanger)
- 2. Short-read (Illumina)
- 3. Long-read (PacBio and Nanopore)

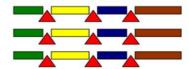
Genome Sequencing - terminology

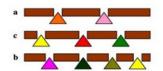
- Read A single sequence from one fragment in the sequencing library (one cluster, bead, etc.)
- Library A collection of DNA fragments that have been prepared to be sequenced
- Coverage number of reads spanning a region of the genome

Fred Sanger (1918-2013)

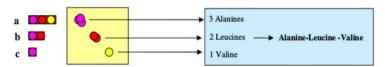
- Two-time nobel prize winner
- 1958 Protein sequencing
- 1980 DNA sequencing

Sanger's degradation procedure for sequencing insulin



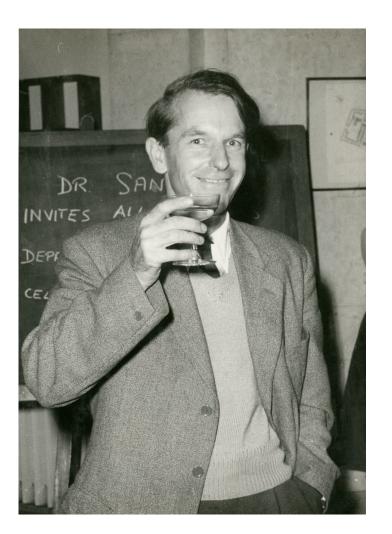


1. Various samples of the protein are broken into fragments by acids (when sequencing the final amino acids) or enzymes and acids (when sequencing the whole molecule) 2. Each fragment is further broken down with different acids-enzymes to work out the overlapping sections

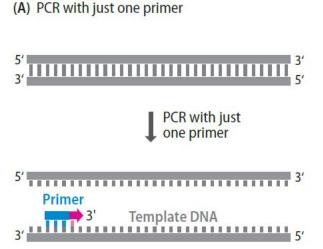


3. The overlapping fragments (a, b and c) are cut again and their constituent amino acids separated by paper chromatography

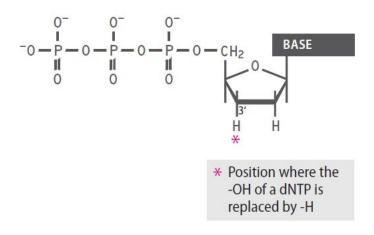
4. Based on the overlapping nature of the sub-fragments it is possible to work out the sequence of constituent amino acids



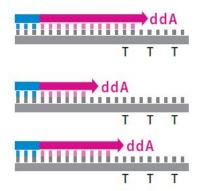
Chain-termination (Sanger)



(B) A dideoxynucleotide

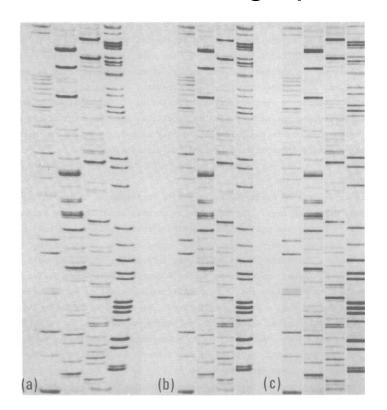


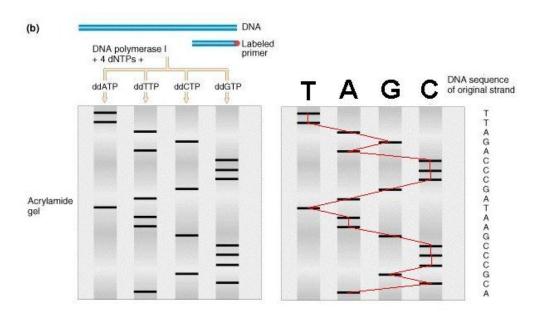
(C) Strand synthesis terminates when a ddNTP is added



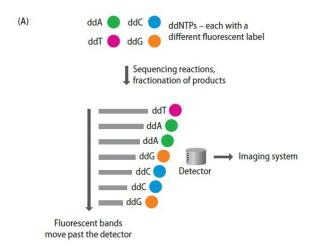
terminator nucleotide

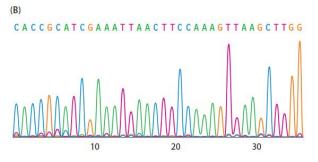
DNA autoradiograph



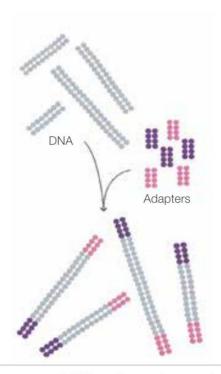


Chain-termination (Sanger) - Modern





- Read length: 700-1000 bp
- Throughput: 96 reads per run (~1 run per hour)
- Error rate: < 1/100,000 bases
- Cost: \$1 per read



Randomly fragment genomic DNA and ligate adapters to both ends of the fragments.

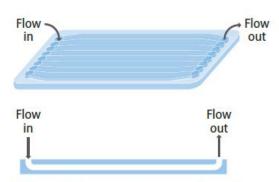
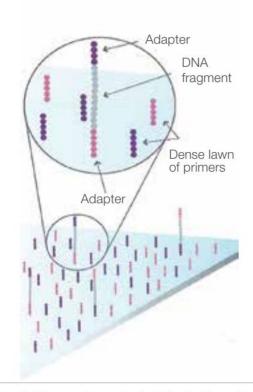
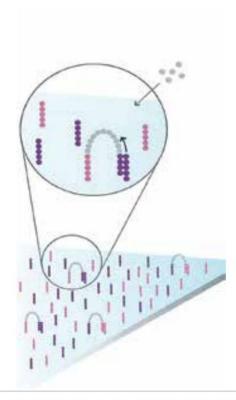


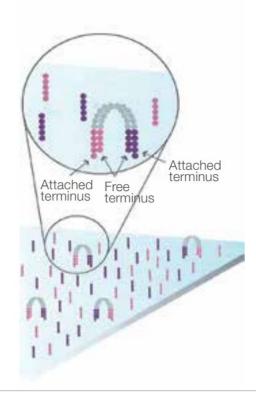
Figure 4.7 A typical flow cell used in DNA sequencing. The sequencing library is immobilized within the channels of the flow cell. To carry out the sequencing reactions, the necessary reagents flow through



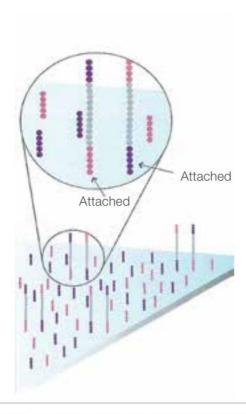
Bind single-stranded fragments randomly to the inside surface of the flow cell channels.



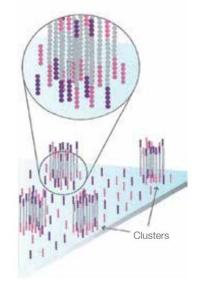
Add unlabeled nucleotides and enzyme to initiate solid-phase bridge amplification.



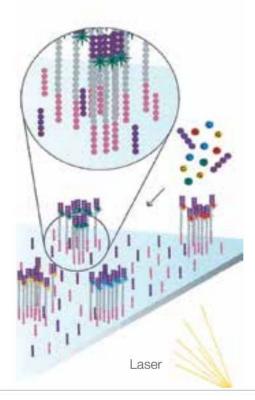
The enzyme incorporates nucleotides to build double-stranded bridges on the solid-phase substrate.

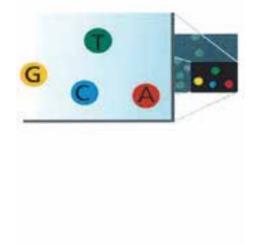


 Bridge PCR repeated 35x to create clusters of <u>identical</u> fragments



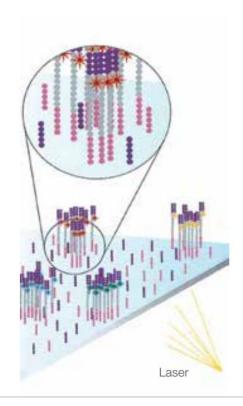
Denaturation leaves single-stranded templates anchored to the substrate.

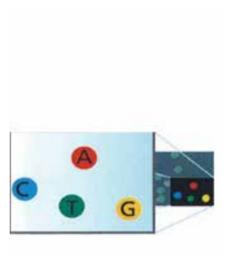




The first sequencing cycle begins by adding four labeled reversible terminators, primers, and DNA polymerase.

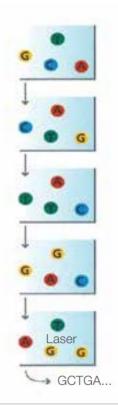
After laser excitation, the emitted fluorescence from each cluster is captured and the first base is identified.





The next cycle repeats the incorporation of four labeled reversible terminators, primers, and DNA polymerase.

After laser excitation, the image is captured as before, and the identity of the second base is recorded.



The sequencing cycles are repeated to determine the sequence of bases in a fragment, one base at a time.

Fragments can be sequenced from either end or both

- Fragment reads (come from fragment libraries)
 - Single read in one direction from a fragment

- Paired end reads (come from fragment libraries)
 - Two reads from opposite ends of the same fragment
 - Reads point towards each other



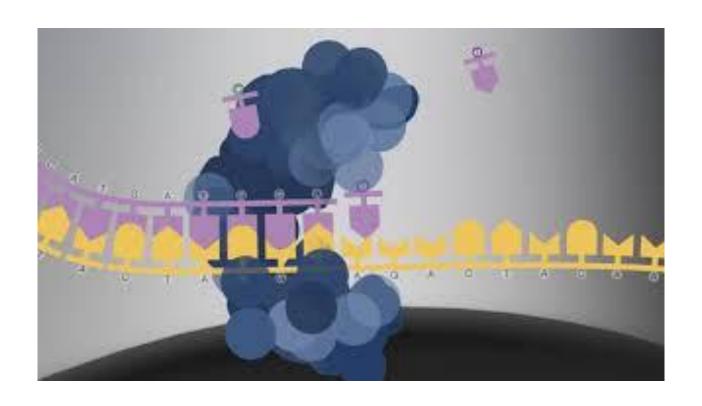
Height: 158.8 cm (62.5 in)

- Read length: 100-300 bp
- Throughput: 25 billion read pairs per run
- Error rate: ~ 1/100 1/1000 bases
- Cost: \$0.00000005 per read

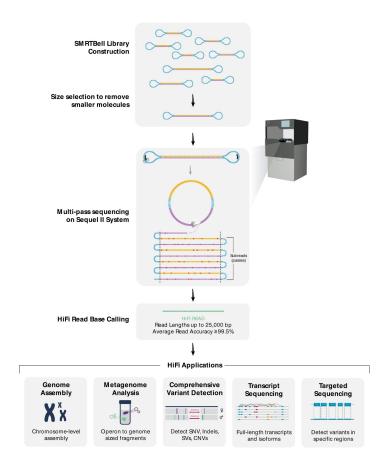
Depth: 86.4 cm (34 in)

Width: 93.3 cm (36.7 in)

Long-read sequencing - Pacific Bioscience



Long-read sequencing - Pacific Bioscience



- Read length: 10 20 kb
- Throughput: up to 4 million reads per run
- Error rate: ~ 1/1000 bases
- Cost: \$0.005 per read

Long-read sequencing - Oxford Nanopore

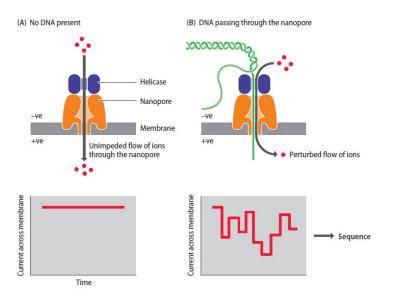
Figure 4.15 Nanopore sequencing.
(A) In the absence of DNA, the flow of ions through the nanopore is unimpeded and the electrical current across the

membrane is constant. (B) Passage of a polynucleotide through the nanopore perturbs the ion flow. Each nucleotide,

or combination of adjacent nucleotides,

perturbs the ion flow in a different way, resulting in fluctuations in the current from which the DNA sequence can be

deduced.

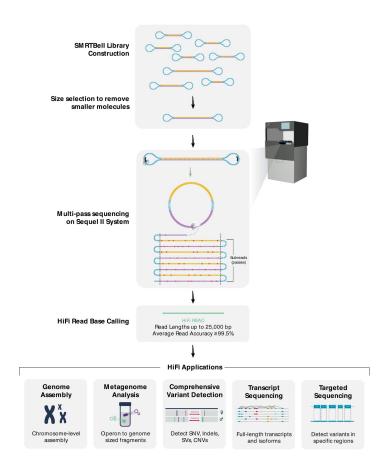


Five nucleotides located in the pore

Membrane

Figure 4.16 More than one nucleotide is present in the nanopore at a single time.

Long-read sequencing - Oxford Nanopore



- Read length: >1 Mbs
- Throughput: 7-12 million reads
- Error rate: ~ 1/10 bases
- Cost: \$0.005 per read

Sequencing data - FastQ file

- ► Simple format for raw unaligned sequencing reads
- ▶ Paired-end sequencing: two FASTQ files or one interleaved file
- ▶ Quality encoded in ASCII characters with decimal codes 33-126
 - ▶ ASCII code of "A" is 65, the corresponding quality is Q = 65 33 = 32

- ► Beware: multiple quality scores were in use!
 - Sanger, Solexa, Illumina 1.3+
 - See https://en.wikipedia.org/wiki/FASTQ_format for details

FastQ - Read name info

Illumina sequence identifiers [edit]

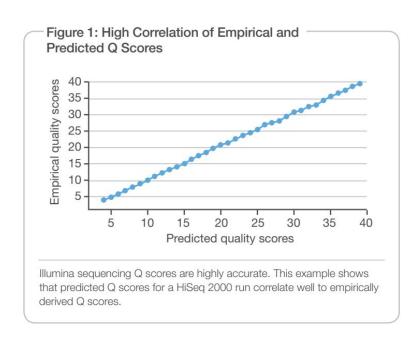
Sequences from the Illumina software use a systematic identifier:

@HWUSI-EAS100R:6:73:941:1973#0/1

HWUSI-EAS100R	the unique instrument name
6	flowcell lane
73	tile number within the flowcell lane
941	'x'-coordinate of the cluster within the tile
1973	'y'-coordinate of the cluster within the tile
#0	index number for a multiplexed sample (0 for no indexing)
/1	the member of a pair, /1 or /2 (paired-end or mate-pair reads only)

FastQ - Quality Info - Phred scores

 Metrics produced by assessing the signal from the sequencing instrument



https://www.illumina.com/documents/products/technotes/technote_Q-Scores.pdf

FastQ - Quality Info

0 1.000	a (
1 0.794	
2 0.631	
3 0.501	
4 0.398	
5 0.316	
6 0.251	
7 0.199	
8 0.158	
9 0.126	
10 0.100	
11 0.079	
12 0.063	
13 0.050	
14 0.040	
15 0.032	

47	@ERR007731.739 IL16_2979:6:1:9:1684/1 ← Read name CTTGACGACTTGAAAAATGACGAAATCACTAAAAAACGTGAAAAATGAGAAATG ← Sequence +
	BBCBCBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBB
	@ERR007731.740 IL16_2979:6:1:9:1419/1
02	AAAAAAAAGATGTCATCAGCACATCAGAAAAGAAGGCAACTTTAAAACTTTTC
•	+
	BBABB/ABABABABBBBBAAA>@B@BBAA@4AAA>.>BAA@779:AAA@A

1	16	0.025
2	17	0.020
3	18	0.016
4	19	0.013
5	20	0.010
6	21	0.008
7	22	0.006
8	23	0.005
9	24	0.004
:	25	0.003
;	26	0.002
<	27	0.002
=	28	0.001
>	29	0.001
?	30	0.001
@	31	0.0008
Α	32	0.0006
В	33	0.0005
С	34	0.0004

https://en.wikipedia.org/wiki/Phred_quality_score

Let's start the <u>genomics adventure</u> and take a look a some typical short reads!