

## KELLY A. McGLYNN, Ph.D.

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### HIGHLIGHTS

- Motivated and determined scientist with 1 year post-PhD experience and **6 years total research experience in cancer epigenetics** (hematology). Broad technical skill set to adapt to project needs.
- Experienced with ***in vivo* oncology models** (mouse models of leukemia and normal hematopoiesis) and animal techniques, including harvesting of various tissues and **multicolor flow cytometry**. Performed **primary-cell assays** on murine bone marrow precursor cells.
- 6 years molecular cloning experience: designed and cloned 20+ mammalian and bacterial expression vectors customized with epitope tags, fluorescent or selectable markers. Designed a **CRISPR** experiment for a breast cancer cell line, generating a novel knockout cell line.
- Independently wrote a successful NIH pre-doctoral fellowship grant and submitted progress reports.
- Individually supervised, trained and provided mentorship to 11 undergraduate and junior graduate students. Able to collaborate and communicate across disciplines and looking forward to working on a team. Will thrive in a fast-paced environment!
- *Looking to permanently relocate!*

### EDUCATION & EXPERIENCE

#### **Postdoctoral Research** (December 2018 – Present)

Wilmot Cancer Center, University of Rochester

- *Drug Discovery* by University of California San Diego on Coursera (June 2019).
- *Data Science Specialization*, set of 10 courses on data analysis and programming in R (Johns Hopkins), *in progress*

#### **Ph.D. in Pharmacology** (October 2018)

University of Rochester Medical Center, Rochester, NY

- *Grants*: NIH F31 Predoctoral Fellowship, Trainee on NIH T32 Training Grant, Dean's Travel Award

#### **B.S. in Molecular Biology**, Minor in Chemistry (2012)

University of Wisconsin-La Crosse, La Crosse, WI

### POSTDOCTORAL RESEARCH

*Using a small molecule inhibitor of the H4K20 methyltransferase Suv420h2 to identify Suv420h2-regulated genes in breast cancer.* Project goals:

- Designed a CRISPR experiment to generate a knockout MCF7 breast cancer cell line. Tested transfection of single vs. multiple combined sgRNAs. Screened and confirmed knockout clones via genomic DNA sequencing, Western blot and T7 mismatch cleavage assay.
- Analyzed whether a small molecule compound could reduce surface PD-L1 (flow cytometry) in lung and breast cancer cell lines.

### GRADUATE RESEARCH

*Discovered a novel protein-protein interaction between a transcription factor and a chromatin remodeling complex*

- Developed an acute leukemia mouse model to identify protein-protein interactions in leukemic tissue. Performed pulldowns and mass spec, identifying a novel protein-protein interaction between the transcription factor EVI1 and multiple subunits of the chromatin remodeling complex SWI/SNF.
- Confirmed the interaction via co-IP in primary cells and cell lines, and employed ChIP-qPCR to confirm genomic co-localization at the transcription factor's DNA binding sites.

*Characterized a double knockout mouse model with a bone marrow failure phenotype*

- Initiated project to generate a tamoxifen-inducible double knockout of the homologous transcription factors *Prdm3* and *Prdm16*. Assisted with tamoxifen injections. Harvested bone marrow from long bones, prepared bone marrow touch prep and cytopsin slides, and analyzed myeloid precursor and mature bone marrow subsets via 7+-color flow cytometry.
- Established a cell culture functional assay (soft agar colony formation assay) in primary bone marrow cells to demonstrate that retroviral addback of the wild-type *Prdm3* gene, but not *Prdm3* with specific point mutations in its putative enzymatic domain, rescues the phenotype. Performed *in silico* structural analysis of the putative enzymatic domain to predict the potential consequences of point mutations on the ligand binding site.

*Analysis of mutations in bacterial porins in clinical outbreak strains of carbapenem-resistant Enterobacter aerogenes [Collaboration project]*

- Collaborated with a clinical microbiologist on a study of mutations in antibiotic-resistant bacteria from hospital patient isolates. Performed *in silico* structural analysis (PyMOL) and alignments of antibiotic importer proteins to predict the potential functional significance of point mutations for bacterial antibiotic resistance.

## TECHNICAL SKILLS

- **Molecular Biology:** Designed and engineered 20+ plasmid constructs during Ph.D.; mutagenesis, PCR, qPCR, CRISPR/Cas9 gene editing and screening. Contributed 7 plasmids to Addgene (June 2018), which have been requested 34 times.
- **Biochemistry:** Bacterial protein expression and purification, SDS-PAGE, Western blot, fluorescent DNA-binding assays, anisotropy/fluorescence polarization, ELISA-based enzyme activity assay, immunoprecipitation and protein-protein interaction studies.
- **Cell biology:** Cell culture functional assays (cell cycle, differentiation, hematopoietic colony formation), multicolor flow cytometry; culture of primary cells, adherent, and suspension cell lines; transfection; shRNA, lentiviral and retroviral infections.
- **Cancer & *in vivo* biology:** Isolation of bone marrow and splenocytes from mice, blood & bone marrow smears, mouse models of leukemia, CBC, isolation of primary murine thymocytes, spheroid culture.
- **Epigenetics:** ChIP-qPCR, histone methyltransferase assays, nucleosome reconstitution, *in vitro* SWI/SNF nucleosome remodeling assays.
- **Software:** Intermediate-level R and Python programming. Graphpad Prism, FlowJo, PyMOL, DNA/cloning software (Snapgene, SeqBuilder), mouse colony database software.

## PUBLICATIONS AND PRESENTATIONS

Malek, A., McGlynn, K., Taffner, S., Fine, L., Tesini, B., Wang, J., Mostafa, H., Petry, S., Perkins, A., Graman, P., *et al.* (2019). Next-Generation-Sequencing-Based Hospital Outbreak Investigation Yields Insight into *Klebsiella aerogenes* Population Structure and Determinants of Carbapenem Resistance and Pathogenicity. *Antimicrobial agents and chemotherapy* 63.

Ph.D. Thesis, available online: McGlynn K. *Cooperative and mechanistic roles of Mecom in hematopoiesis*. University of Rochester; 2018. <http://hdl.handle.net/1802/34859>.

McGlynn K., Sun R., Vonica A., Rudzinskas S., Zhang Y., and Perkins AS. *Prdm3* and *Prdm16* cooperatively maintain hematopoiesis with dependence on the PR domain. [*Submitted to Haematologica – impact factor 7.7*]

McGlynn K., Sun R., Vonica A., Rudzinskas S., Zhang Y., and Perkins AS. *Prdm3* and *Prdm16* contribute to hematopoietic stem cell regulation. (*Poster, national Keystone Symposium on Epigenetics & Cancer*) (2017)

McGlynn K., Zhang, Y., and Perkins, A. Characterization of *Prdm3*-containing protein complex in MLL leukemia. (*Poster, national EpiCypher conference on Clinical Frontiers in Epigenetics*) (2016)

McGlynn K., Zhang, Y., and Perkins, A. The Zinc Finger Transcription Factor Mds1-Evi1 Forms a Novel Protein Complex in MLL leukemia. (*Poster, national Keystone Symposium on Epigenetics & Cancer*) (2015)