

*Evaluation of extending Proxy External Control Association Test  
(ProxECAT) to Poisson Regression*

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**Background**



**Objective**



**Methods**

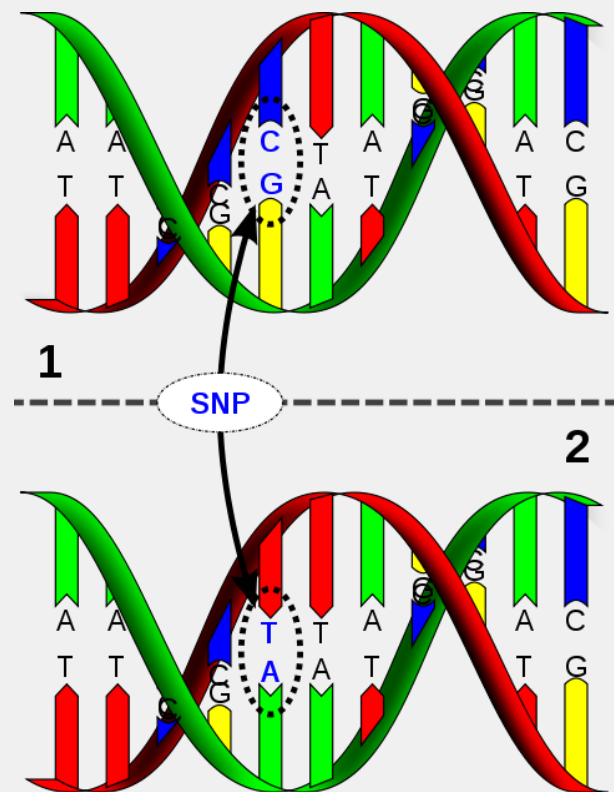


**Results**



**Future Work**

- Alleles: base pairs that differ
- Variants: location of the alleles
  - » Common Variants:  $MAF > 0.05$
  - » Rare Variants:  $MAF < 0.01$
- Minor Allele Frequency (MAF) =  
$$\frac{\text{total \# alternate alleles in the observed variant}}{\text{total \# of alternate alleles in a population}}$$



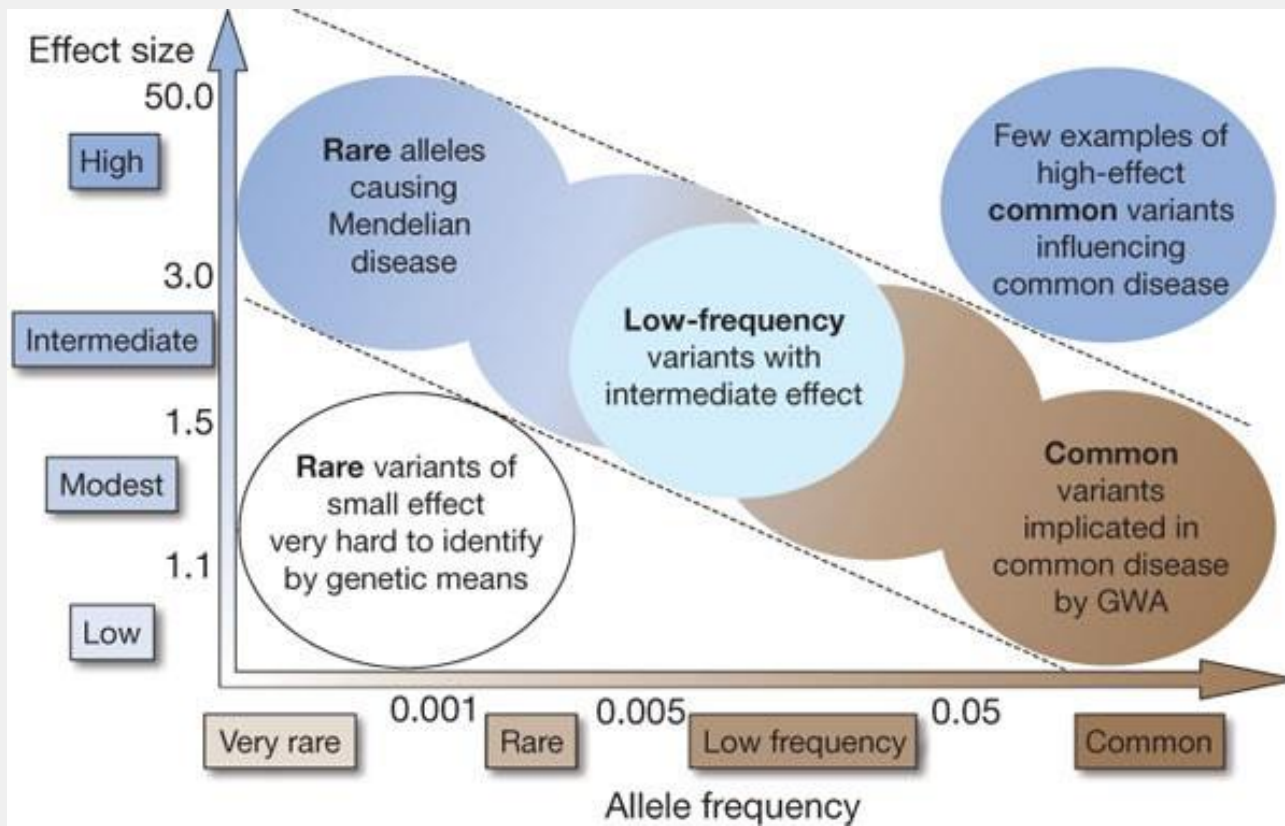
[https://isogg.org/wiki/Single-nucleotide\\_polymorphism](https://isogg.org/wiki/Single-nucleotide_polymorphism)

# Rare Variants

- Functional: alter a gene's function
- Non-Functional (Synonymous): have no effect on the gene's function



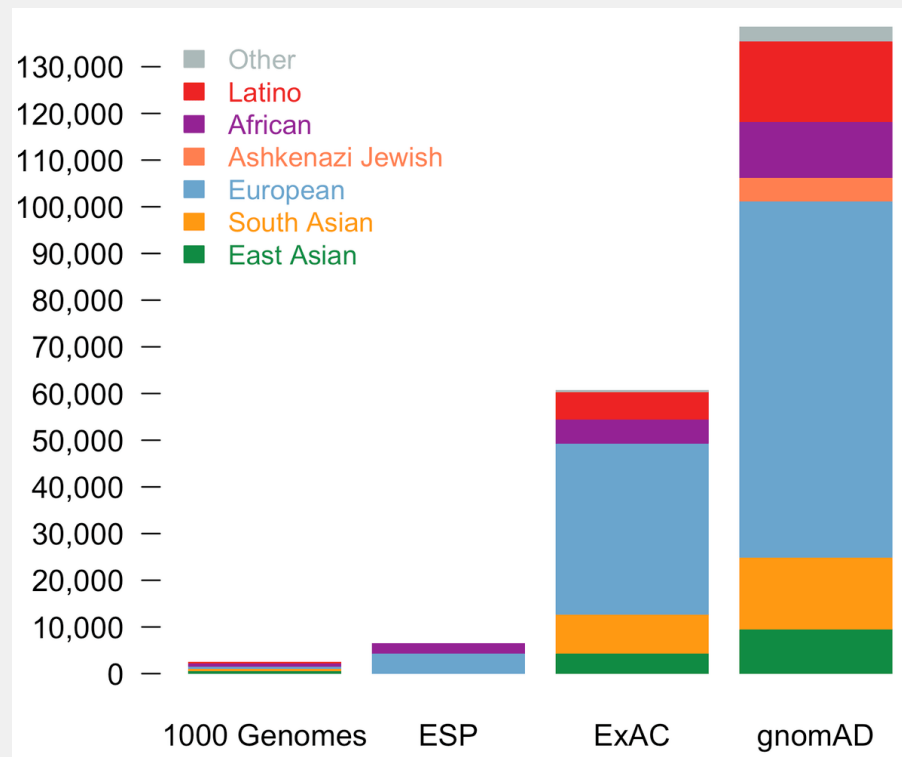
# Genome Wide Association Study (GWAS)



TA Manolio *et al. Nature* **461**, 747-753 (2009) doi:10.1038/nature08494

# External Controls for Rare Variant Association Test

- 10K to 100K sample sizes needed for adequate power
- Use external controls to increase power
- Public databases contain genetic summary data to be used as external controls
- Case control tests using external controls can be biased due to sequencing differences between cases and controls



<https://gnomad.broadinstitute.org/news/2017-02-the-genome-aggregation-database/>



# Proxy External Controls Association Test (ProxECAT)

- Can use very rare variants
  - Optimal when no or limited controls exist
  - Utilizes both functional and synonymous variants
- Singletons and Doubletons
  - Only requires external controls
  - synonymous variants are used as a “proxy” for how well rare variants are sequenced within a gene region



# ProxECAT Limitations

- Does not enable internal controls to be analyzed with external controls
- Cannot adjust for covariates such as sex, ancestry, or proportion of alternate variant reads or depth of coverage



# ★ Extend ProxECAT to a Poisson Regression

## Why?

- Regression can control for covariates
- Both internal and external controls can be evaluated together
- Rare allele counts are approximately distributed as a Poisson distribution

## How?

- Compare results from Poisson regression to ProxECAT and logistic regression

# Observational Units

Unit about which information is collected

In case control studies individuals is often the observational unit

Individual level data is hard to access



# ProxECAT

- Alternate alleles are used as the observational unit
- Alternate allele counts are modeled as a random sample of four independent Poisson distributions

$$X_1^f \sim \text{Poisson}(\lambda_1^f), X_0^f \sim \text{Poisson}(\lambda_0^f), X_1^p \sim \text{Poisson}(\lambda_1^p), \text{ and } X_0^p \sim \text{Poisson}(\lambda_0^p)$$



# ProxECAT Data Notation

		Predicted Functional Impact		Total
		Functional	Not Functional (Proxy)	
<b>Cases (Internal)</b>	$Y = 1$	$x_1^f$	$x_1^p$	$x_1$
<b>Controls (External)</b>	$Y = 0$	$x_0^f$	$x_0^p$	$x_0$
<b>Total</b>		$x^f$	$x^p$	$N$

$x$  – number of alternate alleles

$N$  – total number of alternate alleles in gene region



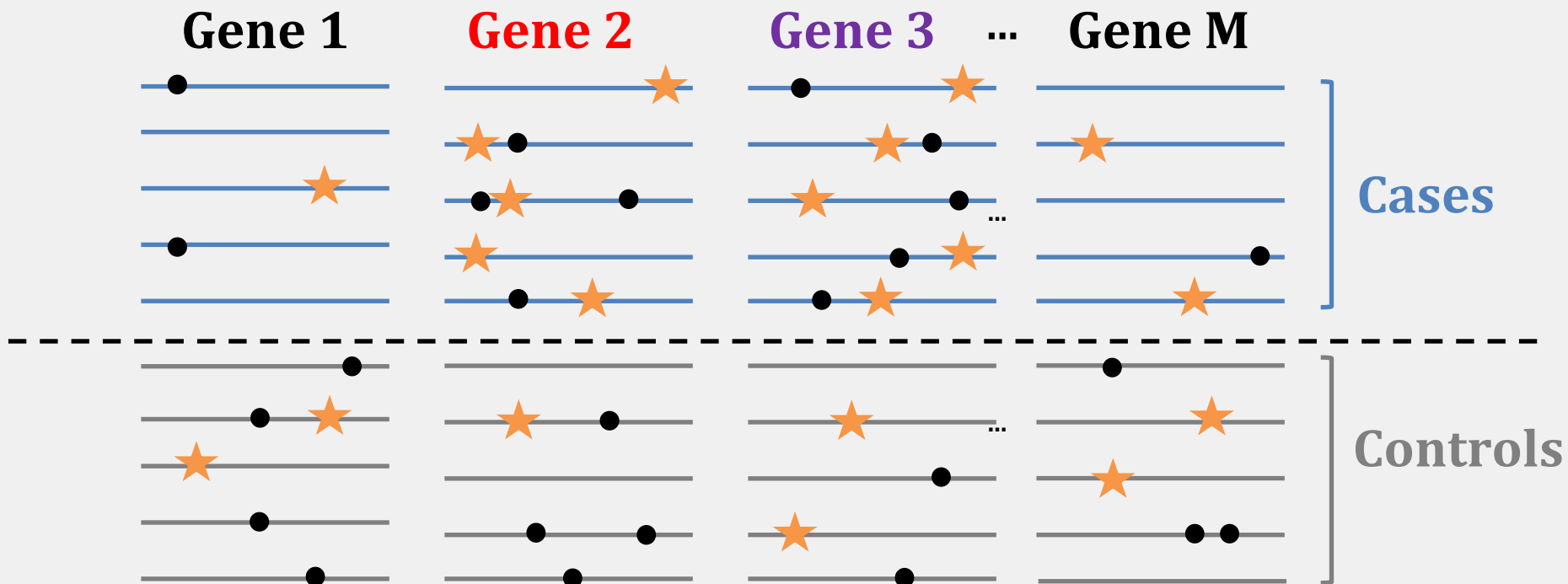
# ProxECAT

$$H_0: \frac{\lambda_{FUN,g^*,case}}{\lambda_{SYN,g^*,case}} = \frac{\lambda_{FUN,g^*,control}}{\lambda_{SYN,g^*,control}}$$

$g^*$ —gene of interest

$\lambda$  — rate of rare alternate alleles per  $N$  cases or controls

- Controls for genetic bias in cases and controls



- Orange stars - rare, functional variants
- Dark circles - rare, synonymous variants

# Generalized Linear Models (GLM)

## ■ Assumptions

1. the observations are independent
2. the response variables follow a distribution from the exponential family
3. there exists a linear relationship between a transformation of the response variable and the predictor variables through a “link” function

$$g(\mu_i) = \beta_0 + \beta_1 x_{1i} + \cdots + \beta_p x_{pi}$$

$\mu_i$ : mean response for observation  $i$

$g(\mu_i)$ : link function

$x_{1i} \dots x_{pi}$ :  $p$  predictor variables for observation  $i$

$\beta_0$ : y-intercept

$\beta_1, \dots, \beta_p$ : regression estimates for each predictor variable  $p$



# Logistic Regression

- Data notation for ProxECAT can be observed as 2x2 Chi-square contingency table
- A 2x2 Chi-square contingency analysis is a specific case of logistic regression





# ProxECAT Implementation of a Logistic Regression

- Alternate alleles are used as the observational unit
- Assume
  1. There does not exist a high correlation between the predictor variables
  2. The distribution of the alternate alleles is binomial
  3. Logit link function



# Implementation of a Logistic Regression

$$\text{logit}(y_i) = \beta_0 + \beta_1 x_i^{\text{case}} + \beta_2 x_i^{\text{group}}$$

- $y_i$  - functional status of alternate allele; 1 is a functional allele and 0 is synonymous alleles
- $x_i^{\text{case}}$  - case status for the carrier of alternate allele
- $x_i^{\text{group}}$  - group status for the carrier of the alternate allele; internal or external
- $\beta_1$  - regression estimate for the association between genetic region and case status
- $\beta_2$  - covariate regression estimate

$$H_0: \beta_1 = 0$$

$$H_1: \beta_1 \neq 0$$

# Logistic Regression & Limitations of ProxECAT

- Can analyze both internal and external control data sets within the same test
- Can adjust for covariates e.g., internal/external, depth of coverage



# Poisson Regression

- counts of rare alleles at a genetic variant are approximately distributed as a Poisson distribution



# ProxECAT implementation of a Poisson Regression

- Genetic variants are used as the observational unit
  
- Assume
  1. Allele counts follow a Poisson distribution
  2. log link function



# Implementation of Poisson Regression for functional (left) and non-functional (right) variants

Association without bias:

small p-value

$$\log(w_i) = \gamma_0 + \gamma_1 x_i^{case}$$

$$H_0: \gamma_1 = 0$$

$$H_1: \gamma_1 \neq 0$$

high p-value

$$\log(z_i) = \alpha_0 + \alpha_1 x_i^{case}$$

$$H_0: \alpha_1 = 0$$

$$H_1: \alpha_1 \neq 0$$

- $w_i$  - positive integer of functional allele counts per gene
- $z_i$  - positive integer of non-functional allele counts per gene
- $x_i^{case}$  - case status for the carrier of the rare variant
- $\gamma_i, \alpha_i$  - regression estimates between genetic region and case status



# Implementation of Poisson Regression for Functional (left) and Non-Functional (right) Variants

Association with bias:

small p-value

$$\log(w_i) = \gamma_0 + \gamma_1 x_i^{case}$$

$$H_0: \gamma_1 = 0$$

$$H_1: \gamma_1 \neq 0$$

low p-value

$$\log(z_i) = \alpha_0 + \alpha_1 x_i^{case}$$

$$H_0: \alpha_1 = 0$$

$$H_1: \alpha_1 \neq 0$$

- $w_i$  - positive integer of functional allele counts per gene
- $z_i$  - positive integer of non-functional allele counts per gene
- $x_i^{case}$  - case status for the carrier of the rare variant
- $\gamma_i, \alpha_i$  - regression estimates between genetic region and case status



## Implementation of Poisson Regression with Functional Status and Case Status

$$\log(p_i) = \omega_0 + \omega_1 x_i^{fun} + \omega_2 x_i^{case} \quad H_0: \omega_1 = 0, \omega_2 = 0$$
$$H_1: \omega_1 = 0, \omega_2 \neq 0$$

- $p_i$  - total allele count for each rare variant per gene
- $x_i^{fun}$  - functional status of the rare variant
- $x_i^{case}$  - case status for the carrier of the rare variant
- $\omega_1, \omega_2, \omega_3$  - regression estimates



# Implementation of Poisson Regression with Interaction

$$\log(p_i) = \omega_0 + \omega_1 x_i^{fun} + \omega_2 x_i^{case} + \omega_3 x_i^{int} \quad H_0: \omega_3 = 0$$
$$H_1: \omega_3 \neq 0$$

- $p_i$  - total allele count for each rare variant per gene
- $x_i^{fun}$  - functional status of the rare variant
- $x_i^{case}$  - case status for the carrier of the rare variant
- $x_i^{int}$  - interaction between functional status and case status
- $\omega_1, \omega_2, \omega_3$  - regression estimates

# Poisson Regression & Limitations of ProxECAT

- Can analyze both internal and external control data sets within the same test
- Can adjust for covariates e.g., internal/external



# Data

Simulations by Jessica Murphy



RARESim (Null, 2022) & Hapgen2

Non-Finnish European haplotypes from 1000 Genome Phase 3

Chromosome 19

12 genes

Simulated under the null hypothesis of no association



# Dataset

<b>Simulation Replicates</b>	100
<b>Genes</b>	12
<b>Total Sample Size</b>	22,000
<b>Internal Case Sample Size</b>	1,000
<b>Internal Control Sample Size</b>	1,000
<b>External Control Sample Size</b>	10,000 ; 10,000
<b>Rare Variants</b>	MAF < 0.01

# Type I Error Rate Comparisons

ProxECAT, Logistic , Poisson

3 main implementations of Poisson

- Functional variant model and non-functional variant model
- Model with functional status and case status
- Interaction model

Sample sizes

- Balanced (1,000 cases vs 1,000 controls)
- Unbalanced (1,000 cases vs 11,000 controls)

## Type I Error rates for Poisson and Logistic (1,000 cases vs 11,000 controls)

	ProxECAT	Logistic
ADGRE 2	0.02	0.06
ADGRE 3	0.10	0.06
ADGRE 5	0.03	0.04
CLEC17A	0.03	0.01
DDX39A	0.09	0.02
DNAJB1	0.03	0.02
GIPC1	0.02	0.02
NDUFB7	0.04	0.01
PKN1	0.06	0.08
PTGER1	0.07	0.01
TECR	0.05	0.00
ZNF333	0.06	0.03
Average	<b>0.050</b>	<b>0.030</b>

## Type I Error Rates for Poisson for Functional and Non-Functional Rare Variants (1,000 cases vs 11,000 controls)

	Functional Rare Variant Model	Non-Functional Rare Variant Model
ADGRE 2	1.00	1.00
ADGRE 3	1.00	1.00
ADGRE 5	1.00	1.00
CLEC17A	0.99	0.72
DDX39A	0.96	0.97
DNAJB1	1.00	1.00
GIPC1	1.00	0.99
NDUFB7	1.00	0.84
PKN1	1.00	1.00
PTGER1	0.98	0.92
TECR	0.90	0.99
ZNF333	1.00	0.99
Average	<b>0.986</b>	<b>0.952</b>



## Type I Error Rates for Poisson for Functional and Non-Functional Rare Variants (1,000 cases vs 1,000 controls)

	Functional Rare Variant Model	Non-Functional Rare Variant Model
ADGRE 2	0.04	0.02
ADGRE 3	0.03	0.04
ADGRE 5	0.02	0.00
CLEC17A	0.03	0.00
DDX39A	0.01	0.00
DNAJB1	0.07	0.03
GIPC1	0.01	0.04
NDUFB7	0.04	0.03
PKN1	0.04	0.03
PTGER1	0.03	0.00
TECR	0.00	0.03
ZNF333	0.04	0.03
Average	<b>0.030</b>	<b>0.021</b>





## Type I Error Rates for Poisson with Functional and Case Status (1,000 cases vs 11,000 controls)

	Functional Status	Case Status
ADGRE 2	0.72	1.00
ADGRE 3	0.66	1.00
ADGRE 5	0.75	1.00
CLEC17A	0.19	1.00
DDX39A	0.73	1.00
DNAJB1	0.84	1.00
GIPC1	0.67	1.00
NDUFB7	0.69	1.00
PKN1	0.87	1.00
PTGER1	0.55	1.00
TECR	0.84	1.00
ZNF333	0.68	1.00
Average	<b>0.682</b>	<b>1.00</b>

## Type I Error Rates for Poisson with Functional and Case Status (1,000 cases vs 1,000 controls)

	Functional Status	Case Status
ADGRE 2	0.40	0.03
ADGRE 3	0.37	0.02
ADGRE 5	0.42	0.04
CLEC17A	0.02	0.01
DDX39A	0.26	0.03
DNAJB1	0.58	0.05
GIPC1	0.34	0.04
NDUFB7	0.53	0.04
PKN1	0.71	0.03
PTGER1	0.22	0.03
TECR	0.47	0.03
ZNF333	0.39	0.07
Average	<b>0.392</b>	<b>0.035</b>

## Type I Error Rates for Poisson with Interaction (1,000 cases vs 11,000 controls)

	Interaction between Functional and Case Status
ADGRE 2	0.02
ADGRE 3	0.06
ADGRE 5	0.02
CLEC17A	0.00
DDX39A	0.02
DNAJB1	0.03
GIPC1	0.01
NDUFB7	0.00
PKN1	0.05
PTGER1	0.03
TECR	0.01
ZNF333	0.02
Average	<b>0.022</b>



- ProxECAT and logistic regression both perform appropriately
- Poisson without interaction cannot account for the imbalance in cases and controls
- Interaction between functional allele status and case status shows the last Poisson model could be explored more

## Discussion

# Moving Forward

Standardize  
population to  
1,000 or 10,000

Different ratios  
of case and  
control samples

Incorporate ratio  
of functional to  
synonymous  
alleles

Additional  
distributions

Logistic  
regression



# Questions



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# References

- *Deoxyribonucleic Acid (DNA) Fact Sheet*. (2020, August 24). Retrieved from National Human Genome Research Institution. <https://www.genome.gov/about-genomics/fact-sheets/Deoxyribonucleic-Acid-Fact-Sheet>.
- Gonzalez, M. W., & Kann, M. G. (2012). Chapter 4: Protein interactions and disease. *PLoS computational biology*, 8(12), e1002819. <https://doi.org/10.1371/journal.pcbi.1002819>.
- “Single-Nucleotide Polymorphism.” Single-Nucleotide Polymorphism - ISOGG Wiki, 31 Jan. 2020, [https://isogg.org/wiki/Single-nucleotide\\_polymorphism](https://isogg.org/wiki/Single-nucleotide_polymorphism).
- Manolio, T., Collins, F., Cox, N. *et al.* Finding the missing heritability of complex diseases. *Nature* **461**, 747–753 (2009). <https://doi.org/10.1038/nature08494>
- Karczewski, Konrad, and Laurent Francioli. “The Genome Aggregation Database (GnomAD).” *The Genome Aggregation Database (GnomAD) | GnomAD News*, 27 Feb. 2017, <https://gnomad.broadinstitute.org/news/2017-02-the-genome-aggregation-database/>.
- Zuk O, Schaffner SF, Samocha K, Do R, Hechter E, Kathiresan S, et al. Searching for missing heritability: designing rare variant association studies. *Proc Natl Acad Sci U S A*. 2014;111(4):E455–64. Epub 2014/01/17. pmid:24443550; PubMed Central PMCID: PMCPMC3910587. <https://doi.org/10.1073/pnas.1322563111>.
- Barrett, J.C., Buxbaum, J.D., Cutler, D.J., Daly, M.J., Devlin, B., Gratten, J., Hurles, M.E., Kosmicki, J.A., Lander, E.S., MacArthur, D.G., Neale, B.M., Roeder, K., Visscher, P.M., & Wray, N.R. (2017). New mutations, old statistical challenges. *bioRxiv*.2. <https://www.biorxiv.org/content/10.1101/115964v3>.
- Dunn, P. K., & Smyth, G. K. (2018). *Generalized Linear Models With Examples in R*. New York: Springer Science+Business Media, LLC.
- Newsom, J. T. (1999-2007). *Lecture 21: Logistic Regression*. Retrieved from newsomj. <http://web.pdx.edu/~newsomj/pa551/lectur21.htm>.
- Null, M., Dupuis, J., Sheinidashtegol, P., Layer, R. M., Gignoux, C. R., & Hendricks, A. E. (2022). RAREsim: A simulation method for very rare genetic variants. *American journal of human genetics*, 109(4), 680–691. <https://doi.org/10.1016/j.ajhg.2022.02.009>
- Murphy, J. (2021, June 3). Common Controls Simulations.

