



# Transplantation and Cellular Therapy

journal homepage: [www.tctjournal.org](http://www.tctjournal.org)



## American Society for Transplantation and Cellular Therapy Series: #5—Management of *Clostridioides difficile* Infection in Hematopoietic Cell Transplant Recipients

Carolyn D. Alonso<sup>1,2,\*</sup>, Gabriela Maron<sup>3</sup>, Mini Kamboj<sup>4</sup>, Paul A. Carpenter<sup>5</sup>, Arun Gurunathan<sup>6</sup>, Kathleen M. Mullane<sup>7</sup>, Erik R. Dubberke<sup>8</sup>

<sup>1</sup> Division of Infectious Diseases, Beth Israel Deaconess Medical Center, Boston, Massachusetts

<sup>2</sup> Harvard Medical School, Boston, Massachusetts

<sup>3</sup> Department of Infectious Diseases, St. Jude Children's Research Hospital, Memphis, Tennessee

<sup>4</sup> Division of Infectious Diseases, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, New York

<sup>5</sup> Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, Washington

<sup>6</sup> Seattle Children's Hospital, Seattle, Washington

<sup>7</sup> Section of Infectious Diseases and Global Health, University of Chicago Medicine, Chicago, Illinois

<sup>8</sup> Washington University School of Medicine, St. Louis, Missouri

### Article history:

Received 11 February 2022

Accepted 14 February 2022

### Key Word:

*Clostridioides difficile*  
Hematopoietic cell transplant  
Infection after transplant  
Fecal microbial transplant  
Graft-versus-host disease

### A B S T R A C T

The Practice Guidelines Committee of the American Society for Transplantation and Cellular Therapy partnered with its Transplant Infectious Disease Special Interest Group to update its 2009 compendium-style infectious disease guidelines for hematopoietic cell transplantation (HCT). A completely new approach was taken with the goal of better serving clinical providers by publishing each standalone topic in the infectious disease series as a concise format of frequently asked questions (FAQ), tables, and figures. Adult and pediatric infectious disease and HCT content experts developed and then answered FAQs and finalized topics with harmonized recommendations that were made by assigning an A through E strength of recommendation paired with a level of supporting evidence graded I through III. This fifth guideline in the series focuses on *Clostridioides difficile* infection with FAQs that address the prevalence, incidence, clinical features, colonization versus infection, clinical complications, diagnostic considerations, pharmacological therapies for episodic or recurrent infection, and the roles of prophylactic antibiotics, probiotics, and fecal microbiota transplantation.

© 2022 The American Society for Transplantation and Cellular Therapy. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

*Clostridioides difficile* infection (CDI) is the leading cause of infectious diarrhea among immunosuppressed hematopoietic cell transplant (HCT) recipients who are at an increased risk for the infection compared with other hospitalized populations because of iatrogenic immunosuppression, broad-spectrum antimicrobial exposure, and prolonged hospitalizations.

### EPIDEMIOLOGY AND COMPLICATIONS

**FAQ1: What are the major risk factors for CDI after allogeneic HCT and how do these compare to risk factors after autologous HCT?**

The first line of defense against CDI is a healthy microbiome. The second line of defense is the immune response against *C. difficile* and its toxins. As such, major risk factors for

CDI tend to be ubiquitous among HCT recipients and include antibiotic treatment (including fluoroquinolone prophylaxis), chemotherapeutic disruption to the bacterial microbiota and mucosa which may occur during conditioning [1], and compromised immunity, whether related to age, acuity of illness, or medical conditions such as graft-versus-host disease (GVHD). HCT studies have not consistently found additional risk factors beyond allo- (higher risk) versus auto-HCT and degree of immunosuppression [1–9].

**FAQ2: At what time point after HCT are most cases of CDI diagnosed?**

The incidence of CDI after allo HCT is 9% to 10% but was as high as 31% in one study and is consistently higher than that seen after auto HCT (5%–6%) [2,6,9–26]. CDI is diagnosed more frequently before engraftment versus after engraftment. After engraftment, the risk for CDI in allo-HCT is higher compared to

Financial disclosure: See Acknowledgments on page XXXX.

\*Correspondence and reprint requests: Carolyn D. Alonso, Beth Israel Deaconess Medical Center, 110 Francis Street LMOB GB, Boston, MA 02215.

E-mail address: [calonso@bidmc.harvard.edu](mailto:calonso@bidmc.harvard.edu) (C.D. Alonso).

<https://doi.org/10.1016/j.tct.2022.02.013>

2666-6367/© 2022 The American Society for Transplantation and Cellular Therapy. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

auto-HCT [27,28]; about half of all CDI cases in allo-HCT occur after engraftment [6,8,18,25,29].

**FAQ3: Are there secondary complications associated with CDI in the HCT population?**

As with other nontransplant populations, HCT recipients with CDI may experience direct complications related to CDI, including dehydration leading to acute kidney injury, toxic megacolon, bowel perforation, death [6,30]. CDI recurrence is a potential secondary complication after a primary CDI episode. Interestingly, most studies have not found a higher incidence of recurrent CDI compared to other patient populations [6,12,31–33].

CDI may also increase the risk for bacteremia with enteric organisms during HCT [34,35]. It is postulated that compromised gut mucosal integrity from chemotherapy and/or GVHD is contributory in the setting of immunocompromise. *C. difficile* toxins further impair colonic mucosal integrity, potentially facilitating translocation of gut bacteria into the bloodstream. GVHD has been found to be both a risk factor for, and a potential complication of, CDI in allo-HCT (Supplementary Table S1 [93–96]) [6,13,36,37]. GVHD and its treatment increase the risk for infection, which leads to antibiotic exposures, both of which increase the risk for CDI. CDI may increase the risk for GVHD because the damage caused by *C. difficile* toxins may expose host antigens to donor immune cells, inciting an immune response.

**CLINICAL FEATURES**

**FAQ4: What are the key clinical features associated with *C. difficile* infection in HCT?**

Like other patient populations, unexplained, new-onset diarrhea ( $\geq 3$  loose bowel movements in a 24-hour time period) or acute worsening of chronic diarrhea is the primary symptom and may be accompanied by abdominal cramping/pain, nausea, vomiting, and fever. More severe CDI is associated with elevated serum creatinine because of dehydration or sepsis. Scoring systems for severity of CDI in the general population have not been validated in HCT recipients [38–41]. For example, leukocytosis, used as a predictor of severity in the general population, may be absent in HCT recipients due to recent conditioning chemotherapy or underlying disease, and signs of inflammation or pain may be absent due to immunosuppressants.

**FAQ5: How can CDI be distinguished from other potential causes of diarrhea after HCT?**

CDI can coexist with noninfectious diarrhea, the latter includes medications (oral magnesium supplements, laxatives, oral contrast) or increasing the rate of enteral formula feeds as common examples. The relatively high frequency of post-transplantation diarrhea and *C. difficile* colonization (See FAQ6) can confound the interpretation of test results for CDI.

In general, the first step in evaluation of diarrhea should include a comprehensive review of medications and removal of any potentially offending agents. If the degree of diarrhea and associated symptoms and clinical parameters are within expectations based on the treatment(s) received, and the patient is clinically stable, it is reasonable to monitor the patient. If the degree of diarrhea or its associated symptoms are worse than would otherwise be expected, then testing for *C. difficile* is warranted (AIII) [38]. Other enteric pathogens (viral, bacterial, and parasitic) could also be considered based on host risk and exposures. Infectious diseases or gastroenterology consultation may be helpful in determining whether additional stool testing is warranted, particularly in cases where the presentation of CDI is atypical, or where the patient is not responding as expected

to CDI therapy. In these instances, alternative diagnoses such as gastrointestinal GVHD or infectious colitis may be considered, and additional work-up such as endoscopy with biopsies may be needed to solidify a diagnosis.

**DIAGNOSIS**

**FAQ6: How does colonization confound the diagnosis of *C. difficile* infection?**

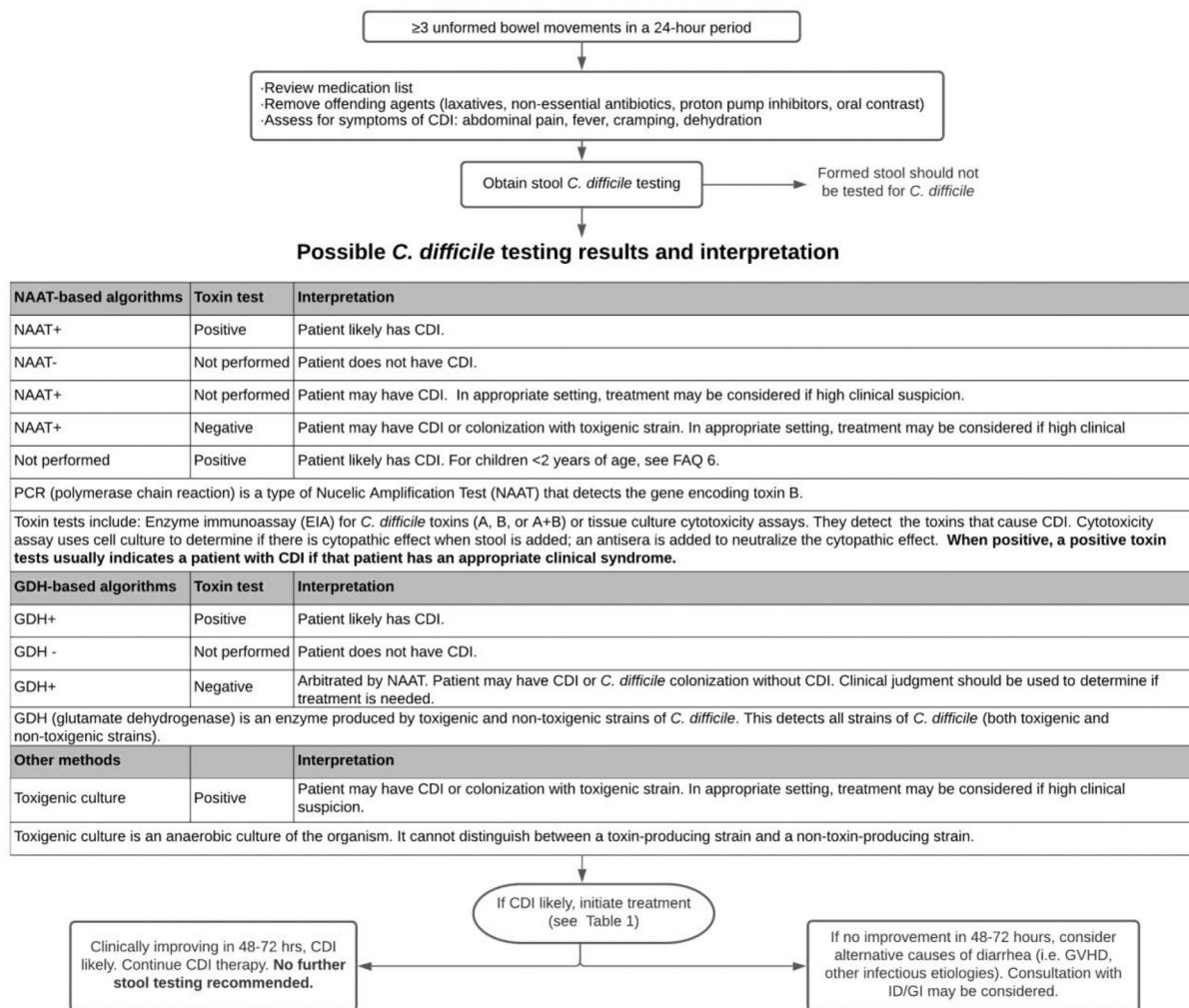
Colonization with toxigenic *C. difficile* has been detected in 11% to 39% of allo-HCT candidates before transplant. Colonization may increase the risk of early CDI, but relevant studies are confounded by use of nucleic acid amplification tests (NAATs; see FAQ7) as the diagnostic method and CDI being associated with conditioning regimens that are more likely to cause diarrhea. In other words, diarrhea may be from the conditioning regimen and NAAT detects pre-existing colonization [12]. Colonization in children is more common than in adults. Up to 50% of infants <1 year of age carry toxigenic strains of *C. difficile* [42–44] and *C. difficile* toxins can be found in the stool of asymptomatic infants [45,46]. Whether *C. difficile* can cause disease in children <2 years of age is unclear. In <1-year-olds, it is not recommended to routinely test for *C. difficile* (AIII) [38]. For 1- and 2-year-olds, testing should only be done after excluding other causes of diarrhea, when there is a high suspicion for CDI and should be limited to diarrheal stool specimens (AIII) [38].

**FAQ7: What is the optimal method for diagnosis of *C. difficile* infection in HCT?**

The optimal method to diagnose CDI has not been established, and clinicians typically do not have the ability to determine which diagnostic assays will be used. As such, it is important to be familiar with the diagnostics available, which one(s) are used at your facility, and their interpretation (Figure 1). There are 3 primary categories of commercially available tests for *C. difficile* used in the United States; NAATs (the most widely used being polymerase chain reaction), enzyme immunoassays (EIAs) for toxins A and B, and glutamate dehydrogenase (GDH) assays. Less frequently used in the United States, but at times more commonly used outside of the United States, are cytotoxicity cell assays and stool culture for *C. difficile*. Below we discuss characteristics consistent within a class of diagnostic assay, but the reader should be aware that differences may exist across manufacturers, platforms, and there can be inter-person variability in assay performance.

Current assays detect *C. difficile* toxins (toxin EIA and cytotoxicity cell assay) or the organism (GDH, NAAT, culture), but none are diagnostic for CDI. CDI is a clinical diagnosis based on presence of clinically significant diarrhea alongside other signs/symptoms of CDI *plus* temporally associated evidence of toxigenic *C. difficile* or its toxins in the stool. Simply detecting *C. difficile* or its toxins in stool without associated CDI symptoms does not indicate CDI because colonic *C. difficile* colonization most commonly is asymptomatic. Therefore testing for the presence of *C. difficile* in formed stools is not recommended. Risk factors for colonization are the same as those for CDI [47–49]. Interpreting a diagnostic assay result must take into account both patient and performance characteristics of the assay(s) used.

In general, methods that detect the organism are more sensitive than methods that detect toxins, but methods that detect toxins are more specific for CDI. Currently in the United States, many laboratories use NAAT as a standalone test [50]. These are highly sensitive for detecting the gene that encodes *C. difficile* toxin production in stool. They have an excellent negative



**Figure 1.** Possible *C. difficile* testing results and interpretation.

predictive value for CDI (~99%); however, because of the high sensitivity, they have a poor positive predictive value for CDI (50% to 60%) [38]. GDH assays should not be used alone to determine who has CDI because bacteria (other than toxin-producing strains of *C. difficile*) that produce GDH may result in a positive assay. GDH assays are typically paired with a toxin EIA. GDH assays are very sensitive, and negative GDH assays have excellent negative predictive value for CDI (~99%) [38]. A positive GDH assay paired with a positive toxin EIA has very good positive predictive value for CDI (~85%) [38]. Most patients with a positive GDH but negative toxin EIA do not have CDI. Some laboratories will reflexively use a NAAT for stools that test positive for GDH and negative by toxin EIA. Approximately 50% of patients with a positive NAAT will have CDI, so clinical judgement is needed when determining which patients with a positive GDH, negative toxin EIA, and positive NAAT should receive treatment for CDI.

## PHARMACOLOGIC TREATMENT

**FAQ8: What are first-line treatments for an initial episode of CDI? (Table 1)**

### Table 1

Discontinuation of inciting antibiotic agent(s) as soon as possible should always be considered as their continued use has been shown to decrease clinical response and increase

recurrence rates (AII) [51,52]. Discontinue unnecessary proton-pump inhibitors when possible because some studies suggest an epidemiologic association between proton-pump inhibitor use and CDI risk (BIII) [53,54].

- Recommended first-line treatment is oral fidaxomicin, or vancomycin as an alternative agent if fidaxomicin is not available (AI) [38,55–58]. Although there are no randomized controlled trials specific to the stem cell transplant population, randomized trials in adults, which included patients with cancer, suggested higher cure and sustained response rates and fewer recurrences associated with fidaxomicin use [59]. Furthermore, when compared with oral vancomycin, fidaxomicin may cause less disruption to the gut microbiome, a factor that may influence the treatment decision-making process, particularly early during the transplant course (See FAQ 13) [60–62]. The choice of CDI treatment may be individualized based on shared informed-decision making between the provider and patient, taking into consideration factors such as the local epidemiology, the patient's risk for recurrent CDI, and other factors such as drug coverage benefits.
- Fidaxomicin is dosed at 200 mg by mouth twice per day for 10 days for adults (AI) [55,59]. The dosing for pediatric patients <6 years old is 16 mg/kg oral suspension twice

**Table 1**Treatment Options for *C. difficile* Infection in Adult and Pediatric Stem Cell Transplant Recipients

Agent	Ways Supplied	Indication/Dosing	Comment
Fidaxomicin	Oral tablet oral suspension (pediatrics)	First Episode CDI: Standard dosing (adults): 200 mg twice a day × 10 days Standard dosing (pediatrics): 16 mg/kg/dose (max 200 mg) twice a day for 10 days Recurrent CDI: Standard dosing (adults): 200 mg twice a day × 10 days Extended dosing (adults): 200 mg twice a day from days 1–5, then every other day from days 7–25 Tapered-pulsed (adults): 200 mg twice a day × 10 days, once a day for 7 days, every other day for 26 days (total 40 capsules) Recurrent CDI (pediatrics): 16 mg/kg/dose (max 200 mg) twice a day for 10 days	The package insert for fidaxomicin provides additional information regarding pediatric dosing according to body weight.
Vancomycin	Oral capsule oral reconstituted solution IV solution administered orally IV solution administered per rectum	First Episode CDI: Standard dosing (adults): 125 mg four times a day × 10 days Standard dosing (pediatrics): 10 mg/kg (125 mg max dose) by mouth every 6 hours × 10 days Recurrent CDI: Standard dosing (adults): 125 mg four times a day × 10 days (only if metronidazole used in prior episode) Tapered-pulsed (adults)* Recurrent CDI (pediatrics) <sup>†</sup> Fulminant CDI: Standard dosing (adults): Vancomycin 500 mg 4 times daily by mouth or by nasogastric tube (in addition to IV metronidazole 500 mg q8 hours) Standard dosing (pediatrics): 10 mg/kg (500 mg max dose) by mouth or by nasogastric tube every 6 hours (in addition to IV metronidazole 10 mg/kg/dose q8h, max 500 mg/dose)	Per rectum vancomycin (500 mg VAN in 100 mL NS via retention enema every 6 hours) can be considered in patients with fulminant disease.
Metronidazole	Oral tablet oral capsule IV	PO (adults): 500 mg every 8 hours (if VAN or FDX are not available) × 10 days IV (adults): (as an adjunctive agent for fulminant CDI in patients receiving VAN or FDX) 500 mg every 8 hours Pediatric dosing: 7.5 mg/kg/dose 3 times a day or 4 times a day (max 500 mg per dose) × 10 days IV (pediatrics): (as an adjunctive agent for fulminant CDI in patients receiving VAN or FDX) 10 mg/kg/dose q8h, max 500 mg/dose	Generally considered to be less efficacious than VAN or FDX. Used as an adjunct to oral vancomycin for fulminant CDI.
Adjunctive Therapies <sup>‡</sup>			
Bezlotoxumab	IV	Adults: Monoclonal antibody indicated for the prevention of recurrence in at-risk patients on CDI therapy	Supplied as a single 10 mg/kg dose [69]. Safety and efficacy have not been evaluated in pediatric patients; a study in children is currently recruiting subjects.
MRT	PO, endoscopically	No standardized dosing available	Has not been studied in larger randomized trials in immunocompromised patients. Potential risk of bacteremia associated with the product in immunosuppressed patients.

VAN indicates vancomycin; NS, normal saline; FDX, fidaxomicin; PO, per os.

\* Tapered and pulse regimen: one approach after completing a 10-day treatment course of 125 mg four times per day would be to decrease to once per day for 4 weeks, then 125 mg every other day for 2 weeks, and then 125 mg once every 3 days for 2 weeks.

<sup>†</sup> Vancomycin tapered regimen in pediatrics: 10 mg/kg (125 mg max dose) 4 times per day for 10–14 days, followed by 10 mg/kg 2 times per day for 7 days, then 10 mg/kg once per day for 7 days, then 10 mg/kg by mouth every 2 or 3 days for 2–8 weeks.<sup>‡</sup> Consideration for adjunctive therapies should be made in consultation with an infectious disease specialist.

daily (maximum 400 mg/d), and 200 mg twice per day for ≥6 years of age (AI) [63]. The phase 3 trials in adults comparing fidaxomicin to oral vancomycin for patients experiencing a first episode of CDI or a first recurrence found fidaxomicin to be noninferior to vancomycin for initial cure but superior for sustained clinical response (i.e., both initial cure plus no recurrence at 30 days after

treatment was stopped) [59,64]. Although not powered for efficacy, a safety study done in children found no difference for initial cure, but fidaxomicin was associated with a significantly better sustained clinical response compared to oral vancomycin [63].

- Vancomycin is dosed at 125 mg by mouth 4 times per day for 10 days (AI) [39,65–67]. Intravenous (IV) vancomycin is



unable to treat CDI because of insufficient penetration into the colon when administered by IV.

- Oral metronidazole is no longer recommended as a first-line agent because it was found to be inferior to oral vancomycin in double blinded randomized trials in adults (DI) [39,67].
- In adults, bezlotoxumab (human monoclonal antibody against *C. difficile* toxin B), given as a single intravenous infusion (10 mg/kg) during active CDI treatment, may be used as an adjunct to standard antibacterial therapy for CDI to reduce the risk of recurrent CDI (rCDI) (BI). Two Phase III studies found a significant reduction in recurrent CDI among immunocompromised subjects receiving the infusion when compared to placebo [58,68,69]. In addition to patients with a history of immunocompromise (such as post-HCT), patients who are above the age of 65, who have severe CDI, or who have a history of prior CDI may further benefit from the addition of bezlotoxumab to standard of care antibiotics [68,70].
- For patients with fulminant CDI (i.e., hypotension or shock, ileus, or megacolon), a regimen of vancomycin 500 mg 4 times daily by mouth or by nasogastric tube in addition to intravenous metronidazole (500 mg every 8 hours) should be administered. For patients with an ileus, in whom there is no contraindication to rectal installation, consider adding a rectal instillation of vancomycin [38].

#### **FAQ9: What are the major treatment options for recurrent CDI?**

Recurrent CDI is defined as a new episode of symptom onset consistent with CDI and a positive assay after a successfully treated episode of CDI with an onset ranging from within days up to 4 to 12 weeks after cessation of treatment for the prior episode [38]. Consultation with an infectious diseases specialist should be considered for all patients with rCDI.

#### **First recurrence**

- Recommended treatments for a first episode of rCDI are a 10-day treatment course of oral fidaxomicin 200 mg twice per day (BI), or an extended course of fidaxomicin (200 mg twice per day for 1 to 5 days, then every other day for days 7 to 25) (BI) [38,56–58,64].
- An oral vancomycin taper-pulse is an acceptable alternative for a first recurrence (BII). Vancomycin tapers have not been standardized, so the optimal duration and taper strategy are not known. Typically, the daily number of doses of vancomycin are reduced over time down to once per day, followed by pulse dosing to every other day and once every 3 days. A retrospective, observational study suggests extending dosing to once every 3 days is associated with fewer recurrences than once every other day dosing [38,71,72].
- If not previously administered, bezlotoxumab, given as a single intravenous infusion (10 mg/kg) during active CDI treatment, may be used as an adjunct to standard antibacterial therapy for CDI to reduce the risk of rCDI (BI) [68,73]. There are no data on the efficacy and safety of additional doses of bezlotoxumab.

#### **Second or greater recurrence**

The optimal therapy for second or greater CDI recurrence has not been defined. There are a variety of approaches to subsequent rCDI, including:

- Standard dosing of fidaxomicin (200 mg twice per day for 10 days) [38,55–57] (CIII)
- Extended dosing of fidaxomicin (200 mg twice per day for 1 to 5 days, then every other day for days 7 to 25) [57] (CIII)
- Taper-pulse of fidaxomicin (for example: 200mg twice per day for 10 days, once per day for 7 days, then once every other day for 26 days [total 40 capsules]) [74] (CIII)
- Taper-pulse of oral vancomycin (CIII) [38,71,72]
- Microbiota restoration therapy (MRT)/fecal microbiota transplantation (FMT) to address associated gut dysbiosis (see FAQ10) (DIII)
- If not previously administered, bezlotoxumab, given as a single intravenous infusion (10 mg/kg) during active CDI treatment, may be used as an adjunct to standard antibacterial therapy for CDI to reduce the risk of rCDI (BI) [68,75]

#### **OTHER MANAGEMENT CONSIDERATIONS**

##### **FAQ10: Can MRT be administered after HCT and what are the potential associated risks?**

Although there are reports of successful intestinal MRT/FMT for rCDI in HCT recipients [76], there is insufficient safety and tolerability data in this context [77–79]. HCT recipients may be at increased risk for bacterial translocation from bacteria transferred with MRT, resulting in an invasive bloodstream infection, unanticipated immunologic consequences of MRT on GVHD, and procedure-associated risks [80]. With these uncertainties, MRT is not routinely recommended as a treatment option for rCDI in HCT recipients, especially neutropenic patients (DIII). Use of MRT/FMT in HCT needs to be individualized with a careful assessment of the risks and benefits. An infectious diseases consultation is recommended to determine whether this is appropriate for individual patients.

##### **FAQ11: What other supportive care can be considered?**

Probiotics have not been found to be helpful to prevent CDI or rCDI in well done studies [38]. In addition, bacteria and fungi found in probiotics can cause infection after HCT [81]. Because of the lack of benefit and potential risk, the use of probiotics are not recommended in HCT recipients for prevention of CDI (DIII). Addition of an anti-motility agent (e.g., loperamide) as an adjunct to specific antibacterial therapy for CDI may be safe, although no prospective or randomized studies are available (C-III) [38].

#### **SPECIAL CONSIDERATIONS**

##### **FAQ12: How should asymptomatic carriers of *C. difficile* be detected and managed?**

Whether patients should be screened for colonization on admission to the hospital to prevent transmission to other patients is an area of ongoing study. However, if a patient is found to be a carrier of *C. difficile* but without CDI, then contact precautions are recommended (BII). This is because carriers without active CDI can be a source of *C. difficile* transmission to other patients in the hospital [82–85].

The treatment of asymptomatic carriers is not recommended because the risks and benefits for this approach are not known (DIII). CDI treatment disrupts the microbiome and facilitates colonization and infection caused by other enteric organisms, such as vancomycin-resistant enterococcus, candida, or resistant Gram negatives. Microbiome disruption is also associated with worse outcomes after allo HCT [86,87].

### FAQ13: Is there a role for primary and secondary CDI prophylaxis in HCT?

Fidaxomicin 200 mg once per day was evaluated in a double-blinded randomized controlled adult study for primary prevention of CDI [88]. The study did not meet its primary composite endpoint, which was prophylaxis failure through 30 days after discontinuation of the study drug. Downsides to prophylaxis include promotion of resistance to fidaxomicin [89]. There are insufficient data to recommend fidaxomicin for CDI prophylaxis at this time: (CI).

Oral vancomycin as primary prophylaxis was shown to reduce CDI in one retrospective adult study [90], but this has not been validated in randomized controlled trials. Retrospective studies on secondary prophylaxis with oral vancomycin in non HCT settings have yielded mixed results. Oral vancomycin is highly disruptive to the microbiome, and microbiome disruption has been associated with worse outcomes after allogeneic HCT [86,87]. Oral vancomycin also facilitates colonization and infection caused by other enteric organisms (See FAQ12). There are insufficient data to recommend for the use of oral vancomycin for CDI prophylaxis at this time: (CII). Metronidazole should not be used for CDI prophylaxis because of lack of efficacy and risk of toxicity with extended use [91,92]: (EIII).

### ACKNOWLEDGMENTS

The authors thank Genovefa Papanicolaou, MD, and Pablo Okhuysen, MD, for their critical review of the manuscript.

*Financial disclosure:* None.

*Conflict of interest statement:* C.D.A. has received grant funding from Merck.

*Authorship statement:* Study conception and design: C.D.A, P.A.C, E.R.D; data collection: C.D.A, G.M, E.R.D, analysis and interpretation of results: C.D.A, G.M., M.K, P.A.C, A.G, K.M.M, E. R.D, draft manuscript preparation: C.D.A, G.M., E.R.D. All authors reviewed the results, participated in critical appraisal, and approved the final version of the manuscript.

### SUPPLEMENTARY MATERIALS

Supplementary material associated with this article can be found, in the online version, at [doi:10.1016/j.jct.2022.02.013](https://doi.org/10.1016/j.jct.2022.02.013).

### APPENDIX 1. GRADING OF STRENGTH OF RECOMMENDATION AND LEVEL OF EVIDENCE

	Grade	Supporting
FAQ1 to FAQ7 Recommendation		
If diarrhea or its associated symptoms are worse than would otherwise be expected, then testing for CDI is warranted.	AIII	[38]
In <1 year-olds, it is not recommended to routinely test for <i>C. difficile</i> .	AIII	[38]
For 1- and 2-year-olds, testing should be done only after excluding other causes of diarrhea, when there is a high suspicion for CDI, and should be limited to diarrheal stool specimens.	AIII	[38]
Discontinuation of inciting antibiotic agent(s) as soon as possible should always be considered as their continued use has been shown to decrease clinical response and increase recurrence rates.	AII	[51,52]
Discontinue unnecessary proton-pump inhibitors when possible.	BIII	[53,54]
FAQ8 Recommendation		
Recommended first-line treatment is oral fidaxomicin or vancomycin as an alternative agent if fidaxomicin is not available.	AI	[38,55-58,64]
	AI	[56,64]

(continued)

(Continued)

	Grade	Supporting
Fidaxomicin is dosed at 200 mg by mouth twice per day for 10 days for adults.		
Fidaxomicin dosing for pediatric patients <6 years old is 16 mg/kg oral suspension twice daily (maximum 400 mg/d) and 200 mg twice per day for ≥6 years of age.	AI	[63]
Vancomycin is dosed at 125 mg by mouth 4 times per day for 10 days.	AI	[39,65,66,97]
Oral metronidazole is no longer recommended as a first-line agent because it was found to be inferior to oral vancomycin in double-blinded randomized trials in adults.	DI	[39,67]
Bezlotoxumab, given as a single intravenous infusion (10 mg/kg) during active CDI treatment, may be used as an adjunct to standard antibacterial therapy for CDI to reduce the risk of rCDI.	BI	[58,68,69]
For fulminant CDI, a regiment of vancomycin 500 mg 4 times daily by mouth or by nasogastric tube in addition to intravenous metronidazole (500 mg every 8 hours) should be considered. Consider adding a rectal instillation of vancomycin if ileus present.	AII	[38]
FAQ9 to FAQ11 Recommendation		
For first episode of rCDI, recommended treatments are a 10-day treatment course of oral fidaxomicin 200 mg twice per day or an extended course of fidaxomicin (200 mg twice per day for 1-5 days, then every other day for days 7-25).	Grade BI	Supporting [38,56-58,64]
An acceptable alternative for first recurrence of CDI is an oral vancomycin taper-pulse.	BII	[38,71,72]
Optimal therapy for second or greater CDI recurrence has not been defined. In such cases, standard dosing of fidaxomicin could be used.	CIII	[38,55-57]
Optimal therapy for second or greater CDI recurrence has not been defined. In such cases, extended dosing of fidaxomicin could be used.	CIII	[57]
Optimal therapy for second or greater CDI recurrence has not been defined. In such cases, taper-pulse of fidaxomicin could be used.	CIII	[74]
Optimal therapy for second or greater CDI recurrence has not been defined. In such cases, taper-pulse of oral vancomycin could be used.	CIII	[38,71,72]
Routine use of MRT is not routinely recommended as a treatment option for rCDI in HCT recipients, especially neutropenic patients.	DIII	[77-80]
Probiotics are not routinely recommended for prevention of CDI in HCT recipients.	DIII	[81]
In patients with rCDI, bezlotoxumab may be used as an adjunct to standard antibacterial therapy for CDI to reduce the risk of rCDI.	BI	[68,73]
There are insufficient data to recommend for or against the addition of an anti-motility agent as an adjunct to specific antibacterial therapy for CDI.	CIII	[38]
FAQ12 to FAQ13 Recommendation		
Contact precautions are recommended for patients who test positive for <i>C. difficile</i> .	BII	[82-85]
Treatment of asymptomatic <i>C. difficile</i> carriers is not recommended.	DIII	[86,87]
There are insufficient data to recommend for or against the use of fidaxomicin for CDI prophylaxis in HCT recipients.	CI	[88,89]
There are insufficient data to recommend for or against the use of oral vancomycin for CDI prophylaxis in HCT recipients.	CII	[90]
Metronidazole should not be used for CDI prophylaxis due to lack of efficacy and risk of toxicity with extended use.	EIII	[91,92]

## REFERENCES

- Kinnebrew MA, Lee YJ, Jenq RR, et al. Early *Clostridium difficile* infection during allogeneic hematopoietic stem cell transplantation. *PLoS One*. 2014;9(3):e90158.
- Carpenter PA, Papanicolaou G, Chemaly RF, Boeckh M, Savani BN. American Society for Transplantation and Cellular Therapy Infectious Disease Guidelines: preface to the series. *Transplant Cell Ther*. 2021;27:103–104.
- Schuster MG, Cleveland AA, Dubberke ER, et al. Infections in hematopoietic cell transplant recipients: results from the Organ Transplant Infection Project, a multicenter, prospective, cohort study. *Open Forum Infect Dis*. 2017;4(2):ofx050.
- Scardina TL, Kang Martinez E, Balasubramanian N, Fox-Geiman M, Smith SE, Parada JP. Evaluation of risk factors for *Clostridium difficile* infection in hematopoietic stem cell transplant recipients. *Pharmacotherapy*. 2017;37:420–428.
- Boyle NM, Magaret A, Stednick Z, et al. Evaluating risk factors for *Clostridium difficile* infection in adult and pediatric hematopoietic cell transplant recipients. *Antimicrob Resist Infect Control*. 2015;4:41.
- Alonso CD, Treadway SB, Hanna DB, et al. Epidemiology and outcomes of *Clostridium difficile* infections in hematopoietic stem cell transplant recipients. *Clin Infect Dis*. 2012;54:1053–1063.
- Misch EA, Safdar N. *Clostridioides difficile* infection in the stem cell transplant and hematologic malignancy population. *Infect Dis Clin North Am*. 2019;33:447–466.
- Lavallee C, Labbe AC, Talbot JD, et al. Risk factors for the development of *Clostridium difficile* infection in adult allogeneic hematopoietic stem cell transplant recipients: a single-center study in Quebec, Canada. *Transpl Infect Dis*. 2017;19(1):e12648.
- Arango JJ, Restrepo A, Schneider DL, et al. Incidence of *Clostridium difficile*-associated diarrhea before and after autologous peripheral blood stem cell transplantation for lymphoma and multiple myeloma. *Bone Marrow Transplant*. 2006;37:517–521.
- Zacharioudakis IM, Ziakas PD, Mylonakis E. *Clostridium difficile* infection in the hematopoietic unit: a meta-analysis of published studies. *Biol Blood Marrow Transplant*. 2014;20:1650–1654.
- Vehreschild MJ, Weitershausen D, Biehl LM, et al. *Clostridium difficile* infection in patients with acute myelogenous leukemia and in patients undergoing allogeneic stem cell transplantation: epidemiology and risk factor analysis. *Biol Blood Marrow Transplant*. 2014;20:823–828.
- Jain T, Crosswell C, Urdy-Cornejo V, et al. *Clostridium Difficile* colonization in hematopoietic stem cell transplant recipients: a prospective study of the epidemiology and outcomes involving toxigenic and nontoxigenic strains. *Biol Blood Marrow Transplant*. 2016;22:157–163.
- Trifilio SM, Pi J, Mehta J. Changing epidemiology of *Clostridium difficile*-associated disease during stem cell transplantation. *Biol Blood Marrow Transplant*. 2013;19:405–419.
- Alonso CD, Dufresne SF, Hanna DB, et al. *Clostridium difficile* infection after adult autologous stem cell transplantation: a multicenter study of epidemiology and risk factors. *Biol Blood Marrow Transplant*. 2013;19:1502–1508.
- Kamboj M, Son C, Cantu S, et al. Hospital-onset *Clostridium difficile* infection rates in persons with cancer or hematopoietic stem cell transplant: a C3IC network report. *Infect Control Hosp Epidemiol*. 2012;33:1162–1165.
- Willems L, Porcher R, Lafaurie M, et al. *Clostridium difficile* infection after allogeneic hematopoietic stem cell transplantation: incidence, risk factors, and outcome. *Biol Blood Marrow Transplant*. 2012;18:1295–1301.
- Chopra T, Chandrasekar P, Salimnia H, Heilbrun LK, Smith D, Alangaden GJ. Recent epidemiology of *Clostridium difficile* infection during hematopoietic stem cell transplantation. *Clin Transplant*. 2011;25(1):E82–E87.
- Alonso CD, Braun DA, Patel I, et al. A multicenter, retrospective, case-cohort study of the epidemiology and risk factors for *Clostridium difficile* infection among cord blood transplant recipients. *Transpl Infect Dis*. 2017;19(4).
- Hosokawa K, Takami A, Tsuji M, et al. Relative incidences and outcomes of *Clostridium difficile* infection following transplantation of unrelated cord blood, unrelated bone marrow, and related peripheral blood in adult patients: a single institute study. *Transpl Infect Dis*. 2014;16:412–420.
- Leung S, Metzger BS, Currie BP. Incidence of *Clostridium difficile* infection in patients with acute leukemia and lymphoma after allogeneic hematopoietic stem cell transplantation. *Infect Control Hosp Epidemiol*. 2010;31:313–315.
- Tomblyn M, Gordon L, Singhal S, et al. Rarity of toxigenic *Clostridium difficile* infections after hematopoietic stem cell transplantation: implications for symptomatic management of diarrhea. *Bone Marrow Transplant*. 2002;30:517–519.
- Barton T, Collis T, Stadtmauer E, Schuster M. Infectious complications the year after autologous bone marrow transplantation or peripheral stem cell transplantation for treatment of breast cancer. *Clin Infect Dis*. 2001;32:391–395.
- Avery R, Pohlman B, Adal K, et al. High prevalence of diarrhea but infrequency of documented *Clostridium difficile* in autologous peripheral blood progenitor cell transplant recipients. *Bone Marrow Transplant*. 2000;25:67–69.
- Bilgrami S, Feingold JM, Dorsky D, et al. Incidence and outcome of *Clostridium difficile* infection following autologous peripheral blood stem cell transplantation. *Bone Marrow Transplant*. 1999;23:1039–1042.
- Dubberke ER, Reske KA, Olsen MA, et al. Epidemiology and outcomes of *Clostridium difficile* infection in allogeneic hematopoietic cell and lung transplant recipients. *Transpl Infect Dis*. 2018;20(2):e12855.
- Ilett EE, Helleberg M, Reekie J, et al. Incidence rates and risk factors of *Clostridioides difficile* infection in solid organ and hematopoietic stem cell transplant recipients. *Open Forum Infect Dis*. 2019;6(4):ofz086.
- Agha A, Sehgal A, Lim MJ, et al. Peri-transplant *Clostridium difficile* infections in patients undergoing allogeneic hematopoietic progenitor cell transplant. *Am J Hematol*. 2016;91:291–294.
- Aldrete SD, Kraft CS, Magee MJ, et al. Risk factors and epidemiology of *Clostridium difficile* infection in hematopoietic stem cell transplant recipients during the peritransplant period. *Transpl Infect Dis*. 2017;19(1):e12649.
- Revolinski SL, Munoz-Price LS. *Clostridium difficile* in immunocompromised hosts: a review of epidemiology, risk factors, treatment, and prevention. *Clin Infect Dis*. 2019;68:2144–2153.
- Lingamaneni P, Katiyar V, Moturi K, et al. *Clostridium difficile* infection is independently associated with significantly worse outcomes in acute leukemia patients undergoing chemotherapy and/or hematopoietic stem cell transplant. *Blood*. 2020;136:13–14. Suppl 1.
- Bruminhent J, Wang ZX, Hu C, et al. *Clostridium difficile* colonization and disease in patients undergoing hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant*. 2014;20:1329–1334.
- Scappaticci GB, Perissinotti AJ, Nagel JL, Bixby DL, Marini BL. Risk factors and impact of *Clostridium difficile* recurrence on haematology patients. *J Antimicrob Chemother*. 2017;72:1488–1495.
- Mani S, Rybicki L, Jagadeesh D, Mossad SB. Risk factors for recurrent *Clostridium difficile* infection in allogeneic hematopoietic cell transplant recipients. *Bone Marrow Transplant*. 2016;51:713–717.
- Falcone M, Russo A, Iraci F, et al. Risk factors and outcomes for bloodstream infections secondary to *Clostridium difficile* infection. *Antimicrob Agents Chemother*. 2016;60:252–257.
- Thomas JA, Newman KC, Doshi S, Logan N, Musher DM. Bacteraemia from an unrecognized source (occult bacteraemia) occurring during *Clostridium difficile* infection. *Scand J Infect Dis*. 2011;43:269–274.
- Chakrabarti S, Lees A, Jones SG, Milligan DW. *Clostridium difficile* infection in allogeneic stem cell transplant recipients is associated with severe graft-versus-host disease and non-relapse mortality. *Bone Marrow Transplant*. 2000;26:871–876.
- Dubberke ER, Reske KA, Srivastava A, et al. *Clostridium difficile*-associated disease in allogeneic hematopoietic stem-cell transplant recipients: risk associations, protective associations, and outcomes. *Clin Transplant*. 2010;24:192–198.
- McDonald LC, Gerding DN, Johnson S, et al. Clinical practice guidelines for *Clostridium difficile* infection in adults and children: 2017 update by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). *Clin Infect Dis*. 2018;66(7):e1–e48.
- Zar FA, Bakkanagari SR, Moorthi KM, Davis MB. A comparison of vancomycin and metronidazole for the treatment of *Clostridium difficile*-associated diarrhea, stratified by disease severity. *Clin Infect Dis*. 2007;45:302–307.
- Belmares J, Gerding DN, Parada JP, Miskevics S, Weaver F, Johnson S. Outcome of metronidazole therapy for *Clostridium difficile* disease and correlation with a scoring system. *J Infect*. 2007;55:495–501.
- Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for *Clostridium difficile* infection. *Clin Microbiol Infect*. 2014;20(Suppl 2):1–26.
- Viscidi R, Willey S, Bartlett JG. Isolation rates and toxigenic potential of *Clostridium difficile* isolates from various patient populations. *Gastroenterology*. 1981;81:5–9.
- Tullus K, Aronsson B, Marcus S, Mollby R. Intestinal colonization with *Clostridium difficile* in infants up to 18 months of age. *Eur J Clin Microbiol Infect Dis*. 1989;8:390–393.
- Enoch DA, Butler MJ, Pai S, Aliyu SH, Karas JA. *Clostridium difficile* in children: colonisation and disease. *J Infect*. 2011;63:105–113.
- Donta ST, Myers MG. *Clostridium difficile* toxin in asymptomatic neonates. *J Pediatr*. 1982;100:431–434.
- Bolton RP, Tait SK, Dear PR, Losowsky MS. Asymptomatic neonatal colonization by *Clostridium difficile*. *Arch Dis Child*. 1984;59:466–472.
- Behar L, Chadwick D, Dunne A, et al. Toxigenic *Clostridium difficile* colonization among hospitalised adults; risk factors and impact on survival. *J Infect*. 2017;75:20–25.
- Kong LY, Dendukuri N, Schiller I, et al. Predictors of asymptomatic *Clostridium difficile* colonization on hospital admission. *Am J Infect Control*. 2015;43:248–253.
- Leekha S, Aronhalt KC, Sloan LM, Patel R, Orenstein R. Asymptomatic *Clostridium difficile* colonization in a tertiary care hospital: admission prevalence and risk factors. *Am J Infect Control*. 2013;41:390–393.
- Guh AY, Mu Y, Winston LG, et al. Trends in U.S. burden of *Clostridioides difficile* infection and outcomes. *N Engl J Med*. 2020;382:1320–1330.



51. Garey KW, Sethi S, Yadav Y, DuPont HL. Meta-analysis to assess risk factors for recurrent *Clostridium difficile* infection. *J Hosp Infect.* 2008;70: 298–304.
52. Mullane KM, Miller MA, Weiss K, et al. Efficacy of fidaxomicin versus vancomycin as therapy for *Clostridium difficile* infection in individuals taking concomitant antibiotics for other concurrent infections. *Clin Infect Dis.* 2011;53:440–447.
53. Linsky A, Gupta K, Lawler EV, Fonda JR, Hermos JA. Proton pump inhibitors and risk for recurrent *Clostridium difficile* infection. *Arch Intern Med.* 2010;170:772–778.
54. Kim YG, Graham DY, Jang BI. Proton pump inhibitor use and recurrent *Clostridium difficile*-associated disease: a case-control analysis matched by propensity score. *J Clin Gastroenterol.* 2012;46:397–400.
55. Louie TJ, Miller MA, Mullane KM, et al. Fidaxomicin versus vancomycin for *Clostridium difficile* infection. *N Engl J Med.* 2011;364:422–431.
56. Cornely OA, Crook DW, Espósito R, et al. Fidaxomicin versus vancomycin for infection with *Clostridium difficile* in Europe, Canada, and the USA: a double-blind, non-inferiority, randomised controlled trial. *Lancet Infect Dis.* 2012;12:281–289.
57. Guery B, Menichetti F, Anttila VJ, et al. Extended-pulsed fidaxomicin versus vancomycin for *Clostridium difficile* infection in patients 60 years and older (EXTEND): a randomised, controlled, open-label, phase 3b/4 trial. *Lancet Infect Dis.* 2018;18:296–307.
58. Johnson S, Laverne V, Skinner AM, et al. Clinical practice guideline by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA): 2021 Focused Update Guidelines on Management of *Clostridioides difficile* Infection in Adults. *Clin Infect Dis.* 2021;73(5):e1029–e1044.
59. Cornely OA, Miller MA, Fantin B, Mullane K, Kean Y, Gorbach S. Resolution of *Clostridium difficile*-associated diarrhea in patients with cancer treated with fidaxomicin or vancomycin. *J Clin Oncol.* 2013;31:2493–2499.
60. Louie TJ, Cannon K, Byrne B, et al. Fidaxomicin preserves the intestinal microbiome during and after treatment of *Clostridium difficile* infection (CDI) and reduces both toxin reexpression and recurrence of CDI. *Clin Infect Dis.* 2012;55(Suppl 2):S132–S142.
61. Ajami NJ, Cope JL, Wong MC, Petrosino JF, Chesnel L. Impact of oral fidaxomicin administration on the intestinal microbiota and susceptibility to *Clostridium difficile* colonization in mice. *Antimicrob Agents Chemother.* 2018;62(5):e02112–e02117.
62. Yamaguchi T, Konishi H, Aoki K, Ishii Y, Chono K, Tateda K. The gut microbiome diversity of *Clostridioides difficile*-inoculated mice treated with vancomycin and fidaxomicin. *J Infect Chemother.* 2020;26:483–491.
63. Wolf J, Kalocsai K, Fortuny C, et al. Safety and efficacy of fidaxomicin and vancomycin in children and adolescents with *Clostridioides (Clostridium) difficile* infection: a phase 3, multicenter, randomized, single-blind clinical trial (SUNSHINE). *Clin Infect Dis.* 2020;71:2581–2588.
64. Louie TJ, Miller MA, Mullane KM, et al. Fidaxomicin versus vancomycin for *Clostridium difficile* infection. *N Engl J Med.* 2011;364:422–431.
65. Teasley DG, Gerding DN, Olson MM, et al. Prospective randomised trial of metronidazole versus vancomycin for *Clostridium-difficile*-associated diarrhoea and colitis. *Lancet.* 1983;2(8358):1043–1046.
66. Wenisch C, Parschall B, Hasenhundl M, Hirschl AM, Graninger W. Comparison of vancomycin, teicoplanin, metronidazole, and fusidic acid for the treatment of *Clostridium difficile*-associated diarrhea. *Clin Infect Dis.* 1996;22:813–818.
67. Johnson S, Louie TJ, Gerding DN, et al. Vancomycin, metronidazole, or tolevamer for *Clostridium difficile* infection: results from two multinational, randomized, controlled trials. *Clin Infect Dis.* 2014;59:345–354.
68. Wilcox MH, Gerding DN, Poxton IR, et al. Bezlotoxumab for prevention of recurrent *Clostridium difficile* infection. *N Engl J Med.* 2017;376:305–317.
69. Zinplava (bezlotoxumab) [prescribing information]. Whitehouse Station, NJ: Merck & Co, Inc.; 2016.
70. Prabhu VS, Dubberke ER, Dorr MB, et al. Cost-effectiveness of bezlotoxumab compared with placebo for the prevention of recurrent *Clostridium difficile* infection. *Clin Infect Dis.* 2018;66:355–362.
71. Sirbu BD, Soriano MM, Manzo C, Lum J, Gerding DN, Johnson S. Vancomycin taper and pulse regimen with careful follow-up for patients with recurrent *Clostridium difficile* infection. *Clin Infect Dis.* 2017;65:1396–1399.
72. McFarland LV, Elmer GW, Surawicz CM. Breaking the cycle: treatment strategies for 163 cases of recurrent *Clostridium difficile* disease. *Am J Gastroenterol.* 2002;97:1769–1775.
73. Gerding DN, Kelly CP, Rahav G, et al. Bezlotoxumab for prevention of recurrent *C. difficile* infection in patients at increased risk for recurrence. *Clin Infect Dis.* 2018;67:649–656.
74. Skinner AM, Tan X, Sirbu BD, Danziger LH, Gerding DN, Johnson S. A tapered-pulsed fidaxomicin regimen following treatment in patients with multiple *Clostridioides difficile* infection recurrences. *Clin Infect Dis.* 2021;73:1107–1109.
75. Gerding DN, Kelly CP, Rahav G, et al. Bezlotoxumab for prevention of recurrent *Clostridium difficile* infection in patients at increased risk for recurrence. *Clin Infect Dis.* 2018;67:649–656.
76. Kelly CR, Ihunnah C, Fischer M, et al. Fecal microbiota transplant for treatment of *Clostridium difficile* infection in immunocompromised patients. *Am J Gastroenterol.* 2014;109:1065–1071.
77. Webb BJ, Brunner A, Ford CD, Gazdik MA, Petersen FB, Hoda D. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection in hematopoietic stem cell transplant recipients. *Transpl Infect Dis.* 2016;18:628–633.
78. Battipaglia G, Malard F, Rubio MT, et al. Fecal microbiota transplantation before or after allogeneic hematopoietic transplantation in patients with hematologic malignancies carrying multidrug-resistance bacteria. *Haematologica.* 2019;104:1682–1688.
79. Bluestone H, Kronman MP, Suskind DL. Fecal microbiota transplantation for recurrent *Clostridium difficile* infections in pediatric hematopoietic stem cell transplant recipients. *J Pediatric Infect Dis Soc.* 2018;7(1):e6–e8.
80. DeFilipp Z, Bloom PP, Torres Soto M, et al. Drug-resistant *E. coli* bacteremia transmitted by fecal microbiota transplant. *N Engl J Med.* 2019;381:2043–2050.
81. Doron S, Snyderman DR. Risk and safety of probiotics. *Clin Infect Dis.* 2015;60 (Suppl 2):S129–S134.
82. Eyre DW, Griffiths D, Vaughan A, et al. Asymptomatic *Clostridium difficile* colonisation and onward transmission. *PLoS One.* 2013;8(11):e78445.
83. Kong LY, Eyre DW, Corbeil J, et al. *Clostridium difficile*: investigating transmission patterns between infected and colonized patients using whole genome sequencing. *Clin Infect Dis.* 2019;68:204–209.
84. Riggs MM, Sethi AK, Zabarsky TF, Eckstein EC, Jump RL, Donskey CJ. Asymptomatic carriers are a potential source for transmission of epidemic and non-epidemic *Clostridium difficile* strains among long-term care facility residents. *Clin Infect Dis.* 2007;45:992–998.
85. Kamboj M, Sheahan A, Sun J, et al. Transmission of *Clostridium difficile* during hospitalization for allogeneic stem cell transplant. *Infect Control Hosp Epidemiol.* 2016;37:8–15.
86. Fishbein SRS, Hink T, Reske KA, et al. Randomized controlled trial of oral vancomycin treatment in *Clostridioides difficile*-colonized patients. *mSphere.* 2021;6(1):e00936. –20.
87. Peled JU, Gomes ALC, Devlin SM, et al. Microbiota as predictor of mortality in allogeneic hematopoietic-cell transplantation. *N Engl J Med.* 2020;382:822–834.
88. Mullane KM, Winston DJ, Nooka A, et al. A randomized, placebo-controlled trial of fidaxomicin for prophylaxis of *Clostridium difficile*-associated diarrhea in adults undergoing hematopoietic stem cell transplantation. *Clin Infect Dis.* 2019;68:196–203.
89. Golan Y, Epstein L. Safety and efficacy of fidaxomicin in the treatment of *Clostridium difficile*-associated diarrhea. *Therap Adv Gastroenterol.* 2012;5:395–402.
90. Ganetsky A, Han JH, Hughes ME, et al. Oral vancomycin prophylaxis is highly effective in preventing *Clostridium difficile* infection in allogeneic hematopoietic cell transplant recipients. *Clin Infect Dis.* 2019;68:2003–2009.
91. Yamamoto T, Abe K, Anjiki H, Ishii T, Kuyama Y. Metronidazole-induced neurotoxicity developed in liver cirrhosis. *J Clin Med Res.* 2012;4:295–298.
92. Knorr JP, Javed I, Sahni N, Cankurtaran CZ, Ortiz JA. Metronidazole-induced encephalopathy in a patient with end-stage liver disease. *Case Reports Hepatol.* 2012;2012: 209258.
93. Amberg S, Kramer M, Schrötter P, et al. *Clostridium difficile* infections in patients with AML or MDS undergoing allogeneic hematopoietic stem cell transplantation identify high risk for adverse outcome. *Bone Marrow Transplant.* 2020;55:367–375.
94. Bhutani D, Jaiyeoba C, Kim S, et al. Relationship between *Clostridium difficile* infection and gastrointestinal graft versus host disease in recipients of allogeneic stem cell transplantation. *Bone Marrow Transplant.* 2019;54:164–167.
95. Dubberke ER, Reske KA, Olsen MA, et al. Risk for *Clostridium difficile* infection after allogeneic hematopoietic cell transplant remains elevated in the postengraftment period. *Transplant Direct.* 2017;3(4):e145.
96. Kamboj M, Xiao K, Kaltsas A, et al. *Clostridium difficile* infection after allogeneic hematopoietic stem cell transplant: strain diversity and outcomes associated with NAP1/027. *Biol Blood Marrow Transplant.* 2014;20:1626–1633.
97. Johnson S, Louie TJ, Gerding DN, et al. Vancomycin, metronidazole, or tolevamer for *Clostridium difficile* infection: results from two multinational, randomized, controlled trials. *Clin Infect Dis.* 2014;59:345–354.