

Appendix S2. Model definitions, model fitting, and model evaluation.

Contents

1	Background	1
2	User inputs	4
3	Loading the fish data	4
4	Loading the covariates	5
5	Specifying the models in JAGS	6
5.1	Ricker model without covariates	6
5.2	Ricker model with covariates	9
6	Fitting the models	12
7	Model selection	14
8	Model diagnostics	15
8.1	Gelman & Rubin statistic	15
8.2	Autocorrelation	17
8.3	Effective sample sizes	18

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1 Background

This appendix describes how we fit the models and evaluated their relative performances. It demonstrates how to load the fish data and environmental covariates, specify the different models in the **JAGS** software, and fit each one.

All analyses require the R software (v3.5 or later) for data retrieval, data processing, and summarizing model results, and the JAGS software (v4.2.0) for Markov chain Monte Carlo (MCMC) simulation. Please note that some of the **R** code below may not work with older versions of **JAGS** due to some changes in the ways that arrays are handled.

We also need a few packages that are not included with the base installation of **R**, so we begin by installing them (if necessary) and then loading them.

```
if(!require("here")) {  
  install.packages("here")  
  library("here")  
}
```

```

if(!require("readr")) {
  install.packages("readr")
  library("readr")
}
if(!require("rjags")) {
  install.packages("rjags")
  library("rjags")
}
if(!require("loo")) {
  install.packages("loo")
  library("loo")
}
if(!require("knitr")) {
  install.packages("knitr")
  library("knitr")
}
if(!require("kableExtra")) {
  install.packages("kableExtra")
  library("kableExtra")
}
## set directory locations
datadir <- here("data")
jagsdir <- here("jags")

```

We also need a couple of helper functions.

```

## better round
Re2prec <- function(x, map = "round", prec = 1) {
  ## 'fun' can be "round", "floor", or "ceiling"
  ## 'prec' is nearest value
  ## (eg, 0.1 is to nearest tenth; 1 is to nearest integer)
  if(prec<=0) { stop("\n\"prec\" cannot be less than or equal to 0") }
  do.call(map,list(x/prec))*prec
}

## wrapper function to fit a JAGS model
fit_jags <- function(model, data, params, inits, ctrl, dir = jagsdir) {
  jm <- jags.model(file.path(jagsdir, model),
    data,
    inits,
    n.chains = ctrl$chains,
    n.adapt = 0,
    quiet = TRUE)

  adp <- FALSE
  while(!adp) {
    adp <- adapt(jm, n.iter = 1000)
  }
  update(jm, ctrl$burn, progress.bar = "none")
  return(coda.samples(jm, params, ctrl$length, ctrl$thin))
}

## inits function for base model
init_vals_AR <- function() {
  list(alpha = 5,

```

```

    beta_inv = exp(mean(ln_dat_esc, na.rm = TRUE)),
    pi_tau = 10,
    pi_eta = rep(1,A),
    pi_vec = matrix(c(0.05,0.5,0.4,0.05),
                     n_yrs-age_min, A,
                     byrow = TRUE),
    Rec_mu = log(1000),
    Rec_sig = 0.1,
    sigma_r = 0.5,
    sigma_s = 0.1,
    tot_ln_Rec = rep(log(1000), n_yrs - age_min),
    innov_1 = 0,
    phi = 0.5)
}

## inits function for cov models
init_vals_cov <- function() {
  list(alpha = 5,
        beta_inv = exp(mean(ln_dat_esc, na.rm = TRUE)),
        gamma = 0,
        pi_tau = 10,
        pi_eta = rep(1,A),
        pi_vec = matrix(c(0.05,0.5,0.4,0.05),
                         n_yrs-age_min, A,
                         byrow = TRUE),
        Rec_mu = log(1000),
        Rec_sig = 0.1,
        tot_ln_Rec = rep(log(1000), n_yrs - age_min),
        # phi = 0.5,
        innov_1 = 0)
}

## estimate LOOIC
looic <- function(jags_obj, mcmc_ctrl) {
  ## convert mcmc.list to matrix
  tmp_lp <- as.matrix(jags_obj)
  ## extract pointwise likelihoods
  tmp_lp <- tmp_lp[,grep("lp_", colnames(tmp_lp))]
  ## if numerical underflows, convert -Inf to 5% less than min(likelihood)
  if(any(is.infinite(tmp_lp))) {
    tmp_lp[is.infinite(tmp_lp)] <- NA
    tmp_min <- min(tmp_lp, na.rm = TRUE)
    tmp_lp[is.na(tmp_lp)] <- tmp_min * 1.05
  }
  ## effective sample size
  r_eff <- relative_eff(exp(tmp_lp),
                        chain_id = rep(seq(mcmc_ctrl$chains),
                                         each = mcmc_ctrl$length / mcmc_ctrl$thin))

  ## calculate LOOIC
  looic <- loo(tmp_lp, r_eff = r_eff)
  return(looic)
}

```

2 User inputs

We begin by supplying values for the minimum and maximum ages of spawning adults, plus some information for the model code and evaluation.

```
## min & max adult age classes
age_min <- 3
age_max <- 6

## file where to save JAGS model
fn_jags <- "Willamette_Chin_SR_flow_models_mainstem_JAGS.txt"

## upper threshold for Gelman & Rubin's potential scale reduction factor (Rhat).
Rhat_thresh <- 1.1
```

Next we specify the names of five necessary data files containing the following information:

1. observed total number of adult spawners (escapement) by year;
2. observed age composition of adult spawners by year;
3. observed total harvest by year;
4. flow covariates by year;
5. metadata for flow covariates.

```
## 1. file with escapement data
## [n_yrs x 2] matrix of obs counts; 1st col is calendar yr
fn_esc <- "chin_esc.csv"

## 2. file with age comp data
## [n_yrs x (1+A)]; 1st col is calendar yr
fn_age <- "chin_agecomp.csv"

## 3. file with harvest data
## [n_yrs x 2] matrix of obs catch; 1st col is calendar yr
fn_harv <- "chin_harv.csv"

## 4. file with harvest data
## [n_yrs x 2] matrix of obs catch; 1st col is calendar yr
fn_cov <- "Willamette_Chin_SR_mainstem_flow_covariates.csv"

## 5. covariate metadata
cov_meta_file <- "chin_cov_metadata.csv"
```

3 Loading the fish data

Here we load in the first three data files and do some simple calculations and manipulations.

First the spawner data:

```
## escapement
dat_esc <- read.csv(file.path(datadir, fn_esc))
## use total counts
```

```

dat_esc <- dat_esc[dat_esc$group=="total",-1]
## years of data
dat_yrs <- dat_esc$year
## number of years of data
n_yrs <- length(dat_yrs)
## get first & last years
yr_first <- min(dat_yrs)
yr_last <- max(dat_yrs)
## log of escapement
ln_dat_esc <- log(dat_esc[,-1])

```

Next the age composition data:

```

## age comp data
dat_age <- read.csv(file.path(datadir, fn_age))
## drop first age_min rows; drop site & year col
dat_age <- dat_age[-(1:(age_min)), -1]
## num of age classes
A <- age_max-age_min+1
## total num of age obs by cal yr
dat_age[, "sum"] <- apply(dat_age, 1, sum)
## row indices for any years with no obs age comp
idx_NA_yrs <- which(dat_age$sum < A, TRUE)
if(length(idx_NA_yrs) > 0) {
  ## replace 0's in yrs w/o any obs with NA's
  dat_age[idx_NA_yrs, (1:A)] <- NA
  ## change total in yrs w/o any obs from 0 to A to help dmulti()
  dat_age[idx_NA_yrs, "sum"] <- A
}
## convert class
dat_age <- as.matrix(dat_age)

```

And then the harvest data:

```

## harvest
dat_harv <- read.csv(file.path(datadir, fn_harv))
## trim to correct years & drop year col
dat_harv <- dat_harv[dat_harv$year >= yr_first & dat_harv$year <= yr_last, -1]

```

4 Loading the covariates

Load the metadata file containing all of the specifications for the covariates to be used.

```

cov_meta <- read.csv(file.path(datadir, cov_meta_file), stringsAsFactors = FALSE)
cov_meta$code <- gsub("\\", "", cov_meta$code)
cov_meta$begin <- gsub("\\", "", cov_meta$begin)
cov_meta$end <- gsub("\\", "", cov_meta$end)

```

Load the saved covariates.

```

cov_flow <- read.csv(file.path(datadir, fn_cov))[, -1]
n_cov <- dim(cov_flow)[2]

```

5 Specifying the models in JAGS

Now we can specify the various models in JAGS. We fit a total of 4 different models, which we outline below, based on the 2 different process models with and without and covariates.

5.1 Ricker model without covariates

```
cat("

model {

  ##-----
  ## PRIORS
  ##-----
  ## alpha = exp(a) = intrinsic productivity
  alpha ~ dnorm(0,0.01) T(0,);
  mu_Rkr_a <- log(alpha);
  E_Rkr_a <- mu_Rkr_a + sigma_r/(2 - 2*phi^2);

  ## strength of dens depend
  beta_inv ~ dnorm(0, 1e-9) T(0,);
  beta <- 1/beta_inv;

  ## AR(1) coef for proc errors
  phi ~ dunif(-0.999,0.999);

  ## process variance for recruits model
  sigma_r ~ dnorm(0, 2e-2) T(0,);
  tau_r <- 1/sigma_r;

  ## innovation in first year
  innov_1 ~ dnorm(0,tau_r*(1-phi*phi));

  ## obs variance for spawners
  tau_s <- 1/sigma_s;
  sigma_s ~ dnorm(0, 0.001) T(0,);

  ## maturity schedule
  ## unif vec for Dirch prior
  theta <- c(2,20,20,1)
  ## hyper-mean for maturity
  pi_eta ~ ddirch(theta);
  ## hyper-prec for maturity
  pi_tau ~ dnorm(0, 0.01) T(0,);
  for(t in 1:(n_yrs-age_min)) { pi_vec[t,1:A] ~ ddirch(pi_eta*pi_tau) }

  ## unprojectable early recruits;
  ## hyper mean across all popns
  Rec_mu ~ dnorm(0,0.001);
  ## hyper SD across all popns
  Rec_sig ~ dunif(0,100);
  ## precision across all popns
```

```

Rec_tau <- pow(Rec_sig,-2);
## multipliers for unobservable total runs
  ttl_run_mu ~ dunif(1,5);
  ttl_run_tau ~ dunif(1,20);

## get total cal yr returns for first age_min yrs
for(i in 1:(age_min)) {
  ln_tot_Run[i] ~ dnorm(ttl_run_mu*Rec_mu,Rec_tau/ttl_run_tau);
  tot_Run[i] <- exp(ln_tot_Run[i]);
}

## estimated harvest rate
for(t in 1:n_yrs) { h_rate[t] ~ dunif(0,1) }

##-----
## LIKELIHOOD
##-----
## 1st brood yr requires different innovation
## predicted recruits in BY t
ln_Rkr_a[1] <- mu_Rkr_a;
E_ln_Rec[1] <- ln_Rkr_a[1] + ln_Sp[1] - beta*Sp[1] + phi*innov_1;
tot_ln_Rec[1] ~ dnorm(E_ln_Rec[1],tau_r);
res_ln_Rec[1] <- tot_ln_Rec[1] - E_ln_Rec[1];
## median of total recruits
tot_Rec[1] <- exp(tot_ln_Rec[1]);

## R/S
ln_RS[1] <- tot_ln_Rec[1] - ln_Sp[1];

## brood-yr recruits by age
for(a in 1:A) {
  Rec[1,a] <- tot_Rec[1] * pi_vec[1,a];
}

## brood years 2:(n_yrs-age_min)
for(t in 2:(n_yrs-age_min)) {
  ## predicted recruits in BY t
  ln_Rkr_a[t] <- mu_Rkr_a;
  E_ln_Rec[t] <- ln_Rkr_a[t] + ln_Sp[t] - beta*Sp[t] + phi*res_ln_Rec[t-1];
  tot_ln_Rec[t] ~ dnorm(E_ln_Rec[t],tau_r);
  res_ln_Rec[t] <- tot_ln_Rec[t] - E_ln_Rec[t];
  ## median of total recruits
  tot_Rec[t] <- exp(tot_ln_Rec[t]);
  ## R/S
  ln_RS[t] <- tot_ln_Rec[t] - ln_Sp[t];
  ## brood-yr recruits by age
  for(a in 1:A) {
    Rec[t,a] <- tot_Rec[t] * pi_vec[t,a];
  }
} ## end t loop over year

## get predicted calendar year returns by age
## matrix Run has dim [(n_yrs-age_min) x A]
## step 1: incomplete early broods

```

```

## first cal yr of this grp is first brood yr + age_min
for(i in 1:(age_max-age_min)) {
  ## projected recruits
  for(a in 1:i) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## imputed recruits
  for(a in (i+1):A) {
    lnRec[i,a] ~ dnorm(Rec_mu,Rec_tau);
    Run[i,a] <- exp(lnRec[i,a]);
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## step 2: info from complete broods
## first cal yr of this grp is first brood yr + age_max
for(i in A:(n_ysr-age_min)) {
  for(a in 1:A) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## get predicted calendar year spawners
## first cal yr is first brood yr
for(t in 1:n_ysr) {
  ## obs model for spawners
  # Sp[t] <- max(10,tot_Run[t] - dat_harv[t]);
  # est_harv[t] = h_rate[t] * tot_Run[t];
  # dat_harv[t] ~ dlnorm(log(est_harv[t]), 20);
  Sp[t] = tot_Run[t] - dat_harv[t];
  ln_Sp[t] <- log(Sp[t]);
  ln_dat_esc[t] ~ dnorm(ln_Sp[t], tau_s);
  lp_esc[t] <- logdensity.norm(ln_dat_esc[t],ln_Sp[t], tau_s);
}

} ## end model description

```



```
", file=file.path(jagsdir, "IPM_RK_AR.txt"))
```

5.2 Ricker model with covariates

```
cat("

model {

  ##-----
  ## PRIORS
  ##-----
  ## alpha = exp(a) = intrinsic productivity
  alpha ~ dnorm(0,0.01) T(0,);
  mu_Rkr_a <- log(alpha);
  E_Rkr_a <- mu_Rkr_a + sigma_r/(2 - 2*phi^2);

  ## strength of dens depend
  beta_inv ~ dnorm(0, 1e-9) T(0,);
  beta <- 1/beta_inv;

  ## covariate effect
  gamma ~ dnorm(0,0.01)

  ## AR(1) coef for proc errors
  phi ~ dunif(-0.999,0.999);

  ## process variance for recruits model
  sigma_r ~ dnorm(0, 2e-2) T(0,);
  tau_r <- 1/sigma_r;

  ## innovation in first year
  innov_1 ~ dnorm(0,tau_r*(1-phi*phi));

  ## obs variance for spawners
  tau_s <- 1/sigma_s;
  sigma_s ~ dnorm(0, 0.001) T(0,);

  ## maturity schedule
  ## unif vec for Dirch prior
  theta <- c(2,20,20,1)
  ## hyper-mean for maturity
  pi_eta ~ ddirch(theta);
  ## hyper-prec for maturity
  pi_tau ~ dnorm(0, 0.01) T(0,);
  for(t in 1:(n_yrs-age_min)) { pi_vec[t,1:A] ~ ddirch(pi_eta*pi_tau) }

  ## unprojectable early recruits;
  ## hyper mean across all popns
  Rec_mu ~ dnorm(0,0.001);
  ## hyper SD across all popns
  Rec_sig ~ dunif(0,100);
  ## precision across all popns
  Rec_tau <- pow(Rec_sig,-2);
```

```

## multipliers for unobservable total runs
  ttl_run_mu ~ dunif(1,5);
  ttl_run_tau ~ dunif(1,20);

## get total cal yr returns for first age_min yrs
for(i in 1:(age_min)) {
  ln_tot_Run[i] ~ dnorm(ttl_run_mu*Rec_mu,Rec_tau/ttl_run_tau);
  tot_Run[i] <- exp(ln_tot_Run[i]);
}

## estimated harvest rate
for(t in 1:n_yrs) { h_rate[t] ~ dunif(0,1) }

##-----
## LIKELIHOOD
##-----
## 1st brood yr requires different innovation
## predicted recruits in BY t
covar[1] <- gamma * mod_cvrs[1];
ln_Rkr_a[1] <- mu_Rkr_a + covar[1];
E_ln_Rec[1] <- ln_Rkr_a[1] + ln_Sp[1] - beta*Sp[1] + phi*innov_1;
tot_ln_Rec[1] ~ dnorm(E_ln_Rec[1],tau_r);
res_ln_Rec[1] <- tot_ln_Rec[1] - E_ln_Rec[1];
## median of total recruits
tot_Rec[1] <- exp(tot_ln_Rec[1]);

## R/S
ln_RS[1] <- tot_ln_Rec[1] - ln_Sp[1];

## brood-yr recruits by age
for(a in 1:A) {
  Rec[1,a] <- tot_Rec[1] * pi_vec[1,a];
}

## brood years 2:(n_yrs-age_min)
for(t in 2:(n_yrs-age_min)) {
  ## predicted recruits in BY t
  covar[t] <- gamma * mod_cvrs[t];
  ln_Rkr_a[t] <- mu_Rkr_a + covar[t];
  E_ln_Rec[t] <- ln_Rkr_a[t] + ln_Sp[t] - beta*Sp[t] + phi*res_ln_Rec[t-1];
  tot_ln_Rec[t] ~ dnorm(E_ln_Rec[t],tau_r);
  res_ln_Rec[t] <- tot_ln_Rec[t] - E_ln_Rec[t];
  ## median of total recruits
  tot_Rec[t] <- exp(tot_ln_Rec[t]);
  ## R/S
  ln_RS[t] <- tot_ln_Rec[t] - ln_Sp[t];
  ## brood-yr recruits by age
  for(a in 1:A) {
    Rec[t,a] <- tot_Rec[t] * pi_vec[t,a];
  }
} ## end t loop over year

## get predicted calendar year returns by age
## matrix Run has dim [(n_yrs-age_min) x A]

```

```

## step 1: incomplete early broods
## first cal yr of this grp is first brood yr + age_min
for(i in 1:(age_max-age_min)) {
  ## projected recruits
  for(a in 1:i) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## imputed recruits
  for(a in (i+1):A) {
    lnRec[i,a] ~ dnorm(Rec_mu,Rec_tau);
    Run[i,a] <- exp(lnRec[i,a]);
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## step 2: info from complete broods
## first cal yr of this grp is first brood yr + age_max
for(i in A:(n_ysr-age_min)) {
  for(a in 1:A) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## get predicted calendar year spawners
## first cal yr is first brood yr
for(t in 1:n_ysr) {
  ## obs model for spawners
  # Sp[t] <- max(10,tot_Run[t] - dat_harv[t]);
  # est_harv[t] = h_rate[t] * tot_Run[t];
  # dat_harv[t] ~ dlnorm(log(est_harv[t]), 20);
  Sp[t] = tot_Run[t] - dat_harv[t];
  ln_Sp[t] <- log(Sp[t]);
  ln_dat_esc[t] ~ dnorm(ln_Sp[t], tau_s);
  lp_esc[t] <- logdensity.norm(ln_dat_esc[t],ln_Sp[t], tau_s);
}

} ## end model description

```

```
", file=file.path(jagsdir, "IPM_RK_cov_AR.txt"))
```

6 Fitting the models

Before fitting the model in JAGS, we need to specify:

1. the data and indices that go into the model;
2. the model parameters and states that we want JAGS to return;
3. the MCMC control parameters.

```
## 1. Data to pass to JAGS:
```

```
dat_jags <- list(dat_age = dat_age,
                ln_dat_esc = ln_dat_esc,
                dat_harv = dat_harv,
                A = A,
                age_min = age_min,
                age_max = age_max,
                n_yrs = n_yrs)
```

```
## 2. Model params/states for JAGS to return:
```

```
par_jags <- c("alpha", "E_Rkr_a", "ln_Rkr_a",
             "beta",
             "Sp", "Rec", "tot_ln_Rec", "ln_RS",
             "pi_eta", "pi_tau",
             "sigma_r", "sigma_s",
             "res_ln_Rec", "phi",
             "lp_age", "lp_esc")
```

```
## 3. MCMC control params:
```

```
mcmc_ctrl <- list(
  chains = 4,
  length = 1.25e5,
  burn = 5e4,
  thin = 100
)
```

```
## total number of MCMC samples after burnin
```

```
mcmc_samp <- mcmc_ctrl$length*mcmc_ctrl$chains/mcmc_ctrl$thin
```

Please note that the following code takes ~60 min to run on a quad-core machine with 3.5 GHz Intel processors.

```
## total number of models to fit
```

```
n_mods <- 1 + n_cov
```

```
## empty list for LOOIC values
```

```
LOOIC <- vector("list", n_mods)
```

```
## fit base model (if not already saved)
```

```
if(!file.exists(file.path(jagsdir, "fit_ricker_base.rds"))) {
  mod_fit <- fit_jags("IPM_RK_AR.txt", dat_jags, par_jags, init_vals_AR, mcmc_ctrl)
  ## save results to file
}
```

```

saveRDS(mod_fit, file.path(jagsdir, "fit_ricker_base.rds"))
## compute LOOIC
LOOIC[[1]] <- looic(mod_fit, mcmc_ctrl)
}

## fit models with covariates
par_jags <- c(par_jags, "gamma")
for(i in seq(n_mods-1)) {
  if(!file.exists(file.path(jagsdir, paste0("fit_ricker_cov_", i, ".rds")))) {
    dat_jags$mod_cvrs <- cov_flow[,i]
    mod_fit <- fit_jags("IPM_RK_cov_AR.txt", dat_jags, par_jags,
                       init_vals_cov, mcmc_ctrl)
    ## save results to file
    saveRDS(mod_fit, file.path(jagsdir, paste0("fit_ricker_cov_", i, ".rds")))
    ## compute LOOIC
    LOOIC[[i+1]] <- looic(mod_fit, mcmc_ctrl)
  }
}
if(!file.exists(file.path(jagsdir, "LOOIC_values.rds"))) {
  saveRDS(LOOIC, file.path(jagsdir, "LOOIC_values.rds"))
} else {
  LOOIC <- readRDS(file.path(jagsdir, "LOOIC_values.rds"))
}

```

6.0.0.1 Convergence checks

```

base_mod <- readRDS(file.path(jagsdir, "fit_ricker_base.rds"))

par_conv <- c("alpha", "beta",
             "sigma_r", "sigma_s",
             "pi_tau", paste0("pi_eta[", seq(A), "]"))

## Gelman-Rubin
gelman.diag(base_mod[,par_conv])

## Potential scale reduction factors:
##
##          Point est. Upper C.I.
## alpha          1.06      1.06
## beta           1.11      1.29
## sigma_r        1.07      1.14
## sigma_s        1.64      6.51
## pi_tau         1.00      1.00
## pi_eta[1]      1.00      1.00
## pi_eta[2]      1.00      1.00
## pi_eta[3]      1.00      1.00
## pi_eta[4]      1.00      1.01
##
## Multivariate psrf
##
## 1.37

## autocorrelation
t(round(autocorr.diag(base_mod[,par_conv],

```

```
lags = seq(mcmc_ctrl$thin, 4*mcmc_ctrl$thin, mcmc_ctrl$thin),
relative=FALSE), 2))
```

##	Lag 100	Lag 200	Lag 300	Lag 400
## alpha	0.10	0.06	0.06	0.09
## beta	0.27	0.23	0.21	0.21
## sigma_r	0.23	0.22	0.19	0.18
## sigma_s	0.38	0.27	0.21	0.18
## pi_tau	0.06	0.04	0.05	0.03
## pi_eta[1]	0.03	0.01	0.01	0.03
## pi_eta[2]	0.03	0.03	0.03	0.02
## pi_eta[3]	0.03	0.04	0.03	0.03
## pi_eta[4]	0.16	0.12	0.12	0.08

7 Model selection

Here is a table of LOOIC results as estimated with `loo()`.

```
## data frame of LOOIC values
tbl_LOOIC <- as.data.frame(round(compare(x = LOOIC), 1))
tbl_LOOIC$d_looic <- -2 * tbl_LOOIC$elpd_diff
tbl_LOOIC <- tbl_LOOIC[, c("p_loo", "se_p_loo", "looic", "se_looic", "d_looic")]
rownames(tbl_LOOIC) <- sub("model", "", rownames(tbl_LOOIC))
tbl_LOOIC <- tbl_LOOIC[order(as.numeric(rownames(tbl_LOOIC))),]
tbl_LOOIC <- data.frame(life_stage = c("base", cov_meta$life_stage),
  variable = c("NA", sub(" of 7-day mean", "", cov_meta$long_name)),
  begin = c("NA", cov_meta$begin),
  end = c("NA", cov_meta$end),
  lag = c(NA, cov_meta$lag_1),
  tbl_LOOIC)
saveRDS(tbl_LOOIC, file.path(jagsdir, "tbl_LOOIC.rds"))
## best model; need to subtract 1 from index to acct for base model
best_i <- which(tbl_LOOIC[, "looic"] == min(tbl_LOOIC[, "looic"])) - 1
best_fit <- readRDS(file.path(jagsdir, paste0("fit_ricker_cov_", best_i, ".rds")))
## table of LOOIC values
kable(tbl_LOOIC[order(tbl_LOOIC[, "looic"]),], "latex", booktabs = TRUE)
```

	life_stage	variable	begin	end	lag	p_loo	se_p_loo	looi	se_looi	d_looi
26	2+ outmigrants	Max	02-01	04-30	2	53.8	4.0	339.4	63.7	0.0
24	rearing	Max	07-01	09-30	1	53.9	4.4	342.2	64.1	2.8
25	rearing	Range	07-01	09-30	1	53.0	4.5	342.4	63.7	3.0
22	rearing	Min	07-01	09-30	1	55.6	4.4	342.6	64.4	3.2
8	prespaw	Min	04-01	04-30	0	55.3	4.1	342.7	63.9	3.2
10	prespaw	Max	05-01	05-31	0	53.3	4.3	343.0	63.5	3.6
20	1+ outmigrants	Max	05-01	05-31	1	52.9	4.2	343.0	63.3	3.6
1	base	NA	NA	NA	NA	53.4	4.4	343.6	63.6	4.2
33	2+ outmigrants	Range	04-01	04-30	2	54.0	4.2	344.0	63.3	4.6
3	prespaw	Median	11-01	03-31	-1	55.6	4.3	344.1	64.0	4.6
7	prespaw	Range	04-01	06-30	0	57.3	3.9	344.1	64.1	4.6
28	2+ outmigrants	Min	02-01	04-30	2	53.2	4.2	344.2	63.0	4.8
19	1+ outmigrants	Min	04-01	04-30	1	55.3	4.3	345.0	63.8	5.6
30	2+ outmigrants	Max	04-01	04-30	2	55.8	4.6	345.8	64.4	6.4
4	prespaw	Min	04-01	06-30	0	54.9	4.4	346.7	63.8	7.2
11	prespaw	Min	07-01	09-30	0	54.7	4.6	347.0	64.1	7.6
21	1+ outmigrants	Min	05-01	05-31	1	55.3	4.5	347.9	63.9	8.6
14	incubation	Median	11-01	03-31	0	79.4	1.5	403.0	55.3	63.6
18	1+ outmigrants	Range	04-01	06-30	1	84.3	1.3	416.4	52.4	77.0
27	2+ outmigrants	Median	02-01	04-30	2	87.8	2.0	421.6	51.0	82.2
17	1+ outmigrants	Max	04-01	06-30	1	86.9	2.7	421.8	49.0	82.4
13	incubation	Max	11-01	03-31	0	89.5	3.0	424.9	48.9	85.4
15	1+ outmigrants	Min	04-01	06-30	1	89.6	2.8	426.1	49.0	86.8
23	rearing	Median	07-01	09-30	1	90.4	2.6	426.2	50.1	86.8
9	prespaw	Min	05-01	05-31	0	90.5	3.1	428.7	48.8	89.4
12	prespaw	Median	07-01	09-30	0	92.4	2.3	430.5	51.7	91.0
29	2+ outmigrants	Range	02-01	04-30	2	97.4	3.8	435.9	52.0	96.6
5	prespaw	Median	04-01	06-30	0	93.8	3.1	448.4	47.1	109.0
6	prespaw	Max	04-01	06-30	0	101.1	6.5	450.1	55.8	110.6
16	1+ outmigrants	Median	04-01	06-30	1	98.7	3.5	459.3	48.7	120.0
31	2+ outmigrants	Median	04-01	04-30	2	107.2	9.4	467.0	58.7	127.6
2	prespaw	Max	11-01	03-31	-1	107.7	8.6	481.8	54.5	142.4
32	2+ outmigrants	Min	04-01	04-30	2	107.2	10.3	481.9	57.1	142.4

8 Model diagnostics

8.1 Gelman & Rubin statistic

Here is a table of the Gelman & Rubin statistics (R_{hat}) for the estimated parameters. Recall that we set an upper threshold of 1.1, so values larger than that deserve some additional inspection.

```
## params of interest
par_conv <- c("alpha","beta","gamma",
             "sigma_r","sigma_s",
             "pi_tau",paste0("pi_eta[",seq(A-1),"]"),
             paste0("Sp[",seq(n_yrs),"]"),
             paste0("tot_ln_Rec[",seq(n_yrs-age_min),"]"))
```

```

## Gelman-Rubin
gelman.diag(best_fit[,par_conv])

## Potential scale reduction factors:
##
##          Point est. Upper C.I.
## alpha          1.00      1.00
## beta           1.00      1.00
## gamma          1.00      1.00
## sigma_r        1.00      1.00
## sigma_s        1.01      1.01
## pi_tau         1.00      1.01
## pi_eta[1]      1.00      1.00
## pi_eta[2]      1.00      1.00
## pi_eta[3]      1.00      1.00
## Sp[1]          1.00      1.00
## Sp[2]          1.00      1.00
## Sp[3]          1.00      1.01
## Sp[4]          1.00      1.00
## Sp[5]          1.00      1.01
## Sp[6]          1.00      1.00
## Sp[7]          1.00      1.00
## Sp[8]          1.00      1.00
## Sp[9]          1.00      1.00
## Sp[10]         1.00      1.01
## Sp[11]         1.00      1.00
## Sp[12]         1.00      1.00
## Sp[13]         1.00      1.00
## Sp[14]         1.00      1.00
## Sp[15]         1.00      1.01
## Sp[16]         1.00      1.01
## Sp[17]         1.00      1.00
## Sp[18]         1.01      1.01
## Sp[19]         1.01      1.02
## tot_ln_Rec[1]  1.00      1.00
## tot_ln_Rec[2]  1.00      1.00
## tot_ln_Rec[3]  1.00      1.00
## tot_ln_Rec[4]  1.00      1.00
## tot_ln_Rec[5]  1.00      1.00
## tot_ln_Rec[6]  1.00      1.01
## tot_ln_Rec[7]  1.00      1.00
## tot_ln_Rec[8]  1.00      1.00
## tot_ln_Rec[9]  1.00      1.00
## tot_ln_Rec[10] 1.00      1.00
## tot_ln_Rec[11] 1.00      1.01
## tot_ln_Rec[12] 1.00      1.01
## tot_ln_Rec[13] 1.00      1.00
## tot_ln_Rec[14] 1.00      1.01
## tot_ln_Rec[15] 1.00      1.00
## tot_ln_Rec[16] 1.00      1.00
##
## Multivariate psrf
##
## 1.02

```


8.2 Autocorrelation

```
t(round(autocorr.diag(best_fit[,par_conv],
  lags = seq(mcmc_ctrl$thin, 3*mcmc_ctrl$thin, mcmc_ctrl$thin),
  relative=FALSE), 2))
```

##	Lag 100	Lag 200	Lag 300
## alpha	0.03	-0.03	-0.01
## beta	0.05	0.01	0.01
## gamma	0.02	-0.02	0.00
## sigma_r	0.00	-0.01	0.00
## sigma_s	0.26	0.11	0.07
## pi_tau	0.04	0.03	0.02
## pi_eta[1]	0.01	0.03	0.01
## pi_eta[2]	0.01	-0.01	0.00
## pi_eta[3]	-0.01	-0.01	0.01
## Sp[1]	0.01	-0.02	0.02
## Sp[2]	0.03	-0.01	-0.02
## Sp[3]	0.00	0.02	0.00
## Sp[4]	0.03	0.00	-0.03
## Sp[5]	0.11	-0.01	0.00
## Sp[6]	0.03	0.02	-0.01
## Sp[7]	0.10	0.05	0.01
## Sp[8]	0.05	0.03	0.02
## Sp[9]	0.10	-0.01	-0.01
## Sp[10]	0.12	0.02	-0.01
## Sp[11]	0.08	0.03	0.01
## Sp[12]	0.17	0.05	0.01
## Sp[13]	0.11	0.02	0.03
## Sp[14]	0.09	0.03	0.01
## Sp[15]	0.08	0.00	0.01
## Sp[16]	0.06	-0.01	0.02
## Sp[17]	0.03	0.02	0.00
## Sp[18]	0.09	0.03	0.03
## Sp[19]	0.05	0.02	-0.01
## tot_ln_Rec[1]	0.06	-0.02	-0.03
## tot_ln_Rec[2]	0.04	0.05	-0.01
## tot_ln_Rec[3]	0.02	0.01	0.03
## tot_ln_Rec[4]	0.07	0.00	0.00
## tot_ln_Rec[5]	0.10	0.02	-0.02
## tot_ln_Rec[6]	0.08	0.00	0.00
## tot_ln_Rec[7]	0.06	0.01	0.02
## tot_ln_Rec[8]	0.13	0.05	0.03
## tot_ln_Rec[9]	0.10	0.01	0.02
## tot_ln_Rec[10]	0.06	0.01	0.02
## tot_ln_Rec[11]	0.05	0.00	0.02
## tot_ln_Rec[12]	0.04	0.00	0.01
## tot_ln_Rec[13]	0.02	0.02	0.01
## tot_ln_Rec[14]	0.10	0.06	0.03
## tot_ln_Rec[15]	0.04	0.01	0.00
## tot_ln_Rec[16]	0.01	-0.03	0.00

8.3 Effective sample sizes

```
floor(effectiveSize(best_fit))
```

##	E_Rkr_a	Rec[1,1]	Rec[2,1]	Rec[3,1]	Rec[4,1]	Rec[5,1]
##	5000	4858	4177	3471	4306	4507
##	Rec[6,1]	Rec[7,1]	Rec[8,1]	Rec[9,1]	Rec[10,1]	Rec[11,1]
##	4493	4026	4052	4127	4229	4793
##	Rec[12,1]	Rec[13,1]	Rec[14,1]	Rec[15,1]	Rec[16,1]	Rec[1,2]
##	4320	4739	4521	4244	4933	4203
##	Rec[2,2]	Rec[3,2]	Rec[4,2]	Rec[5,2]	Rec[6,2]	Rec[7,2]
##	4733	3581	4385	4061	3836	4163
##	Rec[8,2]	Rec[9,2]	Rec[10,2]	Rec[11,2]	Rec[12,2]	Rec[13,2]
##	3702	3883	4267	4235	4947	4862
##	Rec[14,2]	Rec[15,2]	Rec[16,2]	Rec[1,3]	Rec[2,3]	Rec[3,3]
##	4082	4631	4845	4555	3420	4550
##	Rec[4,3]	Rec[5,3]	Rec[6,3]	Rec[7,3]	Rec[8,3]	Rec[9,3]
##	4467	4008	4410	3350	3825	4210
##	Rec[10,3]	Rec[11,3]	Rec[12,3]	Rec[13,3]	Rec[14,3]	Rec[15,3]
##	4351	4684	4606	4250	4678	4708
##	Rec[16,3]	Rec[1,4]	Rec[2,4]	Rec[3,4]	Rec[4,4]	Rec[5,4]
##	4881	5412	4636	4626	4881	4657
##	Rec[6,4]	Rec[7,4]	Rec[8,4]	Rec[9,4]	Rec[10,4]	Rec[11,4]
##	3518	4316	4219	4310	4816	4382
##	Rec[12,4]	Rec[13,4]	Rec[14,4]	Rec[15,4]	Rec[16,4]	Sp[1]
##	5211	1270	1505	2025	4268	5169
##	Sp[2]	Sp[3]	Sp[4]	Sp[5]	Sp[6]	Sp[7]
##	4848	4883	4803	4199	4656	3200
##	Sp[8]	Sp[9]	Sp[10]	Sp[11]	Sp[12]	Sp[13]
##	4347	4182	3759	4114	3458	3834
##	Sp[14]	Sp[15]	Sp[16]	Sp[17]	Sp[18]	Sp[19]
##	4038	4277	4602	4714	3967	4304
##	alpha	beta	gamma	ln_RS[1]	ln_RS[2]	ln_RS[3]
##	4980	4691	4835	4810	4635	4424
##	ln_RS[4]	ln_RS[5]	ln_RS[6]	ln_RS[7]	ln_RS[8]	ln_RS[9]
##	4971	4220	4552	3644	4638	4108
##	ln_RS[10]	ln_RS[11]	ln_RS[12]	ln_RS[13]	ln_RS[14]	ln_RS[15]
##	4369	4628	3634	3978	3689	4386
##	ln_RS[16]	ln_Rkr_a[1]	ln_Rkr_a[2]	ln_Rkr_a[3]	ln_Rkr_a[4]	ln_Rkr_a[5]
##	5396	4861	5402	5413	5412	4871
##	ln_Rkr_a[6]	ln_Rkr_a[7]	ln_Rkr_a[8]	ln_Rkr_a[9]	ln_Rkr_a[10]	ln_Rkr_a[11]
##	4870	5380	5393	4778	5403	5393
##	ln_Rkr_a[12]	ln_Rkr_a[13]	ln_Rkr_a[14]	ln_Rkr_a[15]	ln_Rkr_a[16]	lp_age[1]
##	4889	4781	4708	5404	5404	4846
##	lp_age[2]	lp_age[3]	lp_age[4]	lp_age[5]	lp_age[6]	lp_age[7]
##	4999	4893	4999	4999	5000	4792
##	lp_age[8]	lp_age[9]	lp_age[10]	lp_age[11]	lp_age[12]	lp_age[13]
##	5012	5224	5123	5176	4949	5000
##	lp_age[14]	lp_age[15]	lp_age[16]	lp_esc[1]	lp_esc[2]	lp_esc[3]
##	4999	4999	2307	3582	3620	3419
##	lp_esc[4]	lp_esc[5]	lp_esc[6]	lp_esc[7]	lp_esc[8]	lp_esc[9]
##	3567	4190	4049	3525	3348	3245
##	lp_esc[10]	lp_esc[11]	lp_esc[12]	lp_esc[13]	lp_esc[14]	lp_esc[15]
##	3419	3272	3283	3186	3079	3568

##	lp_esc[16]	lp_esc[17]	lp_esc[18]	lp_esc[19]	phi	pi_eta[1]
##	3549	3717	3269	3292	5345	5000
##	pi_eta[2]	pi_eta[3]	pi_eta[4]	pi_tau	res_ln_Rec[1]	res_ln_Rec[2]
##	4999	5000	1935	4601	5233	5000
##	res_ln_Rec[3]	res_ln_Rec[4]	res_ln_Rec[5]	res_ln_Rec[6]	res_ln_Rec[7]	res_ln_Rec[8]
##	5743	4891	4654	5000	5337	4957
##	res_ln_Rec[9]	res_ln_Rec[10]	res_ln_Rec[11]	res_ln_Rec[12]	res_ln_Rec[13]	res_ln_Rec[14]
##	5326	4893	4999	5484	4619	5000
##	res_ln_Rec[15]	res_ln_Rec[16]	sigma_r	sigma_s	tot_ln_Rec[1]	tot_ln_Rec[2]
##	5491	5485	4918	2744	5021	4442
##	tot_ln_Rec[3]	tot_ln_Rec[4]	tot_ln_Rec[5]	tot_ln_Rec[6]	tot_ln_Rec[7]	tot_ln_Rec[8]
##	4521	4316	4256	4268	4316	3658
##	tot_ln_Rec[9]	tot_ln_Rec[10]	tot_ln_Rec[11]	tot_ln_Rec[12]	tot_ln_Rec[13]	tot_ln_Rec[14]
##	4188	4597	4387	4722	4519	3807
##	tot_ln_Rec[15]	tot_ln_Rec[16]				
##	4854	5346				