

Appendix S2. Model definitions, model fitting, and model evaluation.

Supporting information for Scheuerell et al.

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1 Background

This appendix describes how we fit the models and evaluated their relative performances. It demonstrates how to load the fish data and environmental covariates, specify the different models in the **JAGS** software, and fit each one.

All analyses require the R software (v3.4.3 or later) for data retrieval, data processing, and summarizing model results, and the JAGS software (v4.2.0) for Markov chain Monte Carlo (MCMC) simulation. Please note that some of the **R** code below may not work with older versions of **JAGS** due to some changes in the ways that arrays are handled.

We also need a few packages that are not included with the base installation of **R**, so we begin by installing them (if necessary) and then loading them.

```
if(!require("here")) {  
  install.packages("here")  
  library("here")  
}
```

```

}
if(!require("readr")) {
  install.packages("readr")
  library("readr")
}
if(!require("rjags")) {
  install.packages("rjags")
  library("rjags")
}
if(!require("loo")) {
  install.packages("loo")
  library("loo")
}
## set directory locations
datadir <- here("data")
jagsdir <- here("jags")

```

We also need a couple of helper functions.

```

## better round
Re2prec <- function(x, fun = "round", prec = 1) {
  ## 'fun' can be "round", "floor", or "ceiling"
  ## 'prec' is nearest value
  ## (eg, 0.1 is to nearest tenth; 1 is to nearest integer)
  if(prec<=0) { stop("\\"prec\" cannot be less than or equal to 0") }
  do.call(map,list(x/prec))*prec
}

## wrapper function to fit a JAGS model
fit_jags <- function(model, data, params, inits, ctrl, dir = jagsdir) {
  jm <- jags.model(file.path(jagsdir, model),
    data,
    inits,
    n.chains = ctrl$chains,
    n.adapt = 0,
    quiet = TRUE)

  adp <- FALSE
  while(!adp) {
    adp <- adapt(jm, n.iter = 1000)
  }
  update(jm, ctrl$burn, progress.bar = "none")
  return(coda.samples(jm, params, ctrl$length, ctrl$thin))
}

## inits function for base model
init_vals_AR <- function() {
  list(alpha = 5,
    beta_inv = exp(mean(ln_dat_esc, na.rm = TRUE)),
    pi_tau = 10,
    pi_eta = rep(1,A),
    pi_vec = matrix(c(0.05,0.5,0.4,0.05),
      n_yrs-age_min, A,
      byrow = TRUE),
    Rec_mu = log(1000),

```

```

    Rec_sig = 0.1,
    sigma_r = 0.5,
    sigma_s = 0.1,
    tot_ln_Rec = rep(log(1000), n_yrs - age_min),
    innov_1 = 0,
    phi = 0.5)
}

## inits function for cov models
init_vals_cov <- function() {
  list(alpha = 5,
        beta_inv = exp(mean(ln_dat_esc, na.rm = TRUE)),
        gamma = 0,
        pi_tau = 10,
        pi_eta = rep(1,A),
        pi_vec = matrix(c(0.05,0.5,0.4,0.05),
                          n_yrs-age_min, A,
                          byrow = TRUE),
        Rec_mu = log(1000),
        Rec_sig = 0.1,
        tot_ln_Rec = rep(log(1000), n_yrs - age_min),
        # phi = 0.5,
        innov_1 = 0)
}

## estimate LOOIC
looic <- function(jags_obj, mcmc_ctrl) {
  ## convert mcmc.list to matrix
  tmp_lp <- as.matrix(jags_obj)
  ## extract pointwise likelihoods
  tmp_lp <- tmp_lp[,grepl("lp_", colnames(tmp_lp))]
  ## if numerical underflows, convert -Inf to 5% less than min(likelihood)
  if(any(is.infinite(tmp_lp))) {
    tmp_lp[is.infinite(tmp_lp)] <- NA
    tmp_min <- min(tmp_lp, na.rm = TRUE)
    tmp_lp[is.na(tmp_lp)] <- tmp_min * 1.05
  }
  ## effective sample size
  r_eff <- relative_eff(exp(tmp_lp),
                        chain_id = rep(seq(mcmc_ctrl$chains),
                                         each = mcmc_ctrl$length / mcmc_ctrl$thin))

  ## calculate LOOIC
  looic <- loo(tmp_lp, r_eff = r_eff)
  return(looic)
}

```

2 User inputs

We begin by supplying values for the minimum and maximum ages of spawning adults, plus some information for the model code and evaluation.

```
## min & max adult age classes
age_min <- 3
age_max <- 6

## file where to save JAGS model
fn_jags <- "Willamette_Chin_SR_flow_models_mainstem_JAGS.txt"

## upper threshold for Gelman & Rubin's potential scale reduction factor (Rhat).
Rhat_thresh <- 1.1
```

Next we specify the names of five necessary data files containing the following information:

1. observed total number of adult spawners (escapement) by year;
2. observed age composition of adult spawners by year;
3. observed total harvest by year;
4. flow covariates by year;
5. metadata for flow covariates.

```
## 1. file with escapement data
## [n_yrs x 2] matrix of obs counts; 1st col is calendar yr
fn_esc <- "chin_esc.csv"

## 2. file with age comp data
## [n_yrs x (1+A)]; 1st col is calendar yr
fn_age <- "chin_agecomp.csv"

## 3. file with harvest data
## [n_yrs x 2] matrix of obs catch; 1st col is calendar yr
fn_harv <- "chin_harv.csv"

## 4. file with harvest data
## [n_yrs x 2] matrix of obs catch; 1st col is calendar yr
fn_cov <- "Willamette_Chin_SR_mainstem_flow_covariates.csv"

## 5. covariate metadata
cov_meta_file <- "chin_cov_metadata.csv"
```

3 Loading the fish data

Here we load in the first three data files and do some simple calculations and manipulations.

First the spawner data:

```
## escapement
dat_esc <- read.csv(file.path(datadir, fn_esc))
## use total counts
dat_esc <- dat_esc[dat_esc$group=="total",-1]
## years of data
dat_yrs <- dat_esc$year
## number of years of data
n_yrs <- length(dat_yrs)
## get first & last years
```

```

yr_first <- min(dat_yrs)
yr_last <- max(dat_yrs)
## log of escapement
ln_dat_esc <- log(dat_esc[,-1])

```

Next the age composition data:

```

## age comp data
dat_age <- read.csv(file.path(datadir, fn_age))
## drop first age_min rows; drop site & year col
dat_age <- dat_age[-(1:(age_min)), -1]
## num of age classes
A <- age_max-age_min+1
## total num of age obs by cal yr
dat_age[, "sum"] <- apply(dat_age, 1, sum)
## row indices for any years with no obs age comp
idx_NA_yrs <- which(dat_age$sum < A, TRUE)
if(length(idx_NA_yrs) > 0) {
  ## replace 0's in yrs w/o any obs with NA's
  dat_age[idx_NA_yrs, (1:A)] <- NA
  ## change total in yrs w/o any obs from 0 to A to help dmulti()
  dat_age[idx_NA_yrs, "sum"] <- A
}
## convert class
dat_age <- as.matrix(dat_age)

```

And then the harvest data:

```

## harvest
dat_harv <- read.csv(file.path(datadir, fn_harv))
## trim to correct years & drop year col
dat_harv <- dat_harv[dat_harv$year >= yr_first & dat_harv$year <= yr_last, -1]

```

4 Loading the covariates

Load the metadata file containing all of the specifications for the covariates to be used.

```

cov_meta <- read.csv(file.path(datadir, cov_meta_file), stringsAsFactors = FALSE)
cov_meta$code <- gsub("\\", "", cov_meta$code)
cov_meta$begin <- gsub("\\", "", cov_meta$begin)
cov_meta$end <- gsub("\\", "", cov_meta$end)

```

Load the saved covariates.

```

cov_flow <- read.csv(file.path(datadir, fn_cov))[, -1]
n_cov <- dim(cov_flow)[2]

```

5 Specifying the models in JAGS

Now we can specify the various models in JAGS. We fit a total of 4 different models, which we outline below, based on the 2 different process models with and without and covariates.

5.1 Ricker model without covariates

```
cat("

model {

  ##-----
  ## PRIORS
  ##-----
  ## alpha = exp(a) = intrinsic productivity
  alpha ~ dnorm(0,0.01) T(0,);
  mu_Rkr_a <- log(alpha);
  E_Rkr_a <- mu_Rkr_a + sigma_r/(2 - 2*phi^2);

  ## strength of dens depend
  beta_inv ~ dnorm(0, 1e-9) T(0,);
  beta <- 1/beta_inv;

  ## AR(1) coef for proc errors
  phi ~ dunif(-0.999,0.999);

  ## process variance for recruits model
  sigma_r ~ dnorm(0, 2e-2) T(0,);
  tau_r <- 1/sigma_r;

  ## innovation in first year
  innov_1 ~ dnorm(0,tau_r*(1-phi*phi));

  ## obs variance for spawners
  tau_s <- 1/sigma_s;
  sigma_s ~ dnorm(0, 0.001) T(0,);

  ## maturity schedule
  ## unif vec for Dirch prior
  theta <- c(2,20,20,1)
  ## hyper-mean for maturity
  pi_eta ~ ddirch(theta);
  ## hyper-prec for maturity
  pi_tau ~ dnorm(0, 0.01) T(0,);
  for(t in 1:(n_yrs-age_min)) { pi_vec[t,1:A] ~ ddirch(pi_eta*pi_tau) }

  ## unprojectable early recruits;
  ## hyper mean across all popns
  Rec_mu ~ dnorm(0,0.001);
  ## hyper SD across all popns
  Rec_sig ~ dunif(0,100);
  ## precision across all popns
```

```

Rec_tau <- pow(Rec_sig,-2);
## multipliers for unobservable total runs
  ttl_run_mu ~ dunif(1,5);
  ttl_run_tau ~ dunif(1,20);

## get total cal yr returns for first age_min yrs
for(i in 1:(age_min)) {
  ln_tot_Run[i] ~ dnorm(ttl_run_mu*Rec_mu,Rec_tau/ttl_run_tau);
  tot_Run[i] <- exp(ln_tot_Run[i]);
}

## estimated harvest rate
for(t in 1:n_yrs) { h_rate[t] ~ dunif(0,1) }

##-----
## LIKELIHOOD
##-----
## 1st brood yr requires different innovation
## predicted recruits in BY t
ln_Rkr_a[1] <- mu_Rkr_a;
E_ln_Rec[1] <- ln_Rkr_a[1] + ln_Sp[1] - beta*Sp[1] + phi*innov_1;
tot_ln_Rec[1] ~ dnorm(E_ln_Rec[1],tau_r);
res_ln_Rec[1] <- tot_ln_Rec[1] - E_ln_Rec[1];
## median of total recruits
tot_Rec[1] <- exp(tot_ln_Rec[1]);

## R/S
ln_RS[1] <- tot_ln_Rec[1] - ln_Sp[1];

## brood-yr recruits by age
for(a in 1:A) {
  Rec[1,a] <- tot_Rec[1] * pi_vec[1,a];
}

## brood years 2:(n_yrs-age_min)
for(t in 2:(n_yrs-age_min)) {
  ## predicted recruits in BY t
  ln_Rkr_a[t] <- mu_Rkr_a;
  E_ln_Rec[t] <- ln_Rkr_a[t] + ln_Sp[t] - beta*Sp[t] + phi*res_ln_Rec[t-1];
  tot_ln_Rec[t] ~ dnorm(E_ln_Rec[t],tau_r);
  res_ln_Rec[t] <- tot_ln_Rec[t] - E_ln_Rec[t];
  ## median of total recruits
  tot_Rec[t] <- exp(tot_ln_Rec[t]);
  ## R/S
  ln_RS[t] <- tot_ln_Rec[t] - ln_Sp[t];
  ## brood-yr recruits by age
  for(a in 1:A) {
    Rec[t,a] <- tot_Rec[t] * pi_vec[t,a];
  }
} ## end t loop over year

## get predicted calendar year returns by age
## matrix Run has dim [(n_yrs-age_min) x A]
## step 1: incomplete early broods

```

```

## first cal yr of this grp is first brood yr + age_min
for(i in 1:(age_max-age_min)) {
  ## projected recruits
  for(a in 1:i) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## imputed recruits
  for(a in (i+1):A) {
    lnRec[i,a] ~ dnorm(Rec_mu,Rec_tau);
    Run[i,a] <- exp(lnRec[i,a]);
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## step 2: info from complete broods
## first cal yr of this grp is first brood yr + age_max
for(i in A:(n_ysr-age_min)) {
  for(a in 1:A) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## get predicted calendar year spawners
## first cal yr is first brood yr
for(t in 1:n_ysr) {
  ## obs model for spawners
  # Sp[t] <- max(10,tot_Run[t] - dat_harv[t]);
  # est_harv[t] = h_rate[t] * tot_Run[t];
  # dat_harv[t] ~ dlnorm(log(est_harv[t]), 20);
  Sp[t] = tot_Run[t] - dat_harv[t];
  ln_Sp[t] <- log(Sp[t]);
  ln_dat_esc[t] ~ dnorm(ln_Sp[t], tau_s);
  lp_esc[t] <- logdensity.norm(ln_dat_esc[t],ln_Sp[t], tau_s);
}

} ## end model description

```



```
", file=file.path(jagsdir, "IPM_RK_AR.txt"))
```

5.2 Ricker model with covariates

```
cat("

model {

  ##-----
  ## PRIORS
  ##-----
  ## alpha = exp(a) = intrinsic productivity
  alpha ~ dnorm(0,0.01) T(0,);
  mu_Rkr_a <- log(alpha);
  E_Rkr_a <- mu_Rkr_a + sigma_r/(2 - 2*phi^2);

  ## strength of dens depend
  beta_inv ~ dnorm(0, 1e-9) T(0,);
  beta <- 1/beta_inv;

  ## covariate effect
  gamma ~ dnorm(0,0.01)

  ## AR(1) coef for proc errors
  # phi ~ dunif(-0.999,0.999);
  phi <- 0;

  ## process variance for recruits model
  sigma_r ~ dnorm(0, 2e-2) T(0,);
  tau_r <- 1/sigma_r;

  ## innovation in first year
  innov_1 ~ dnorm(0,tau_r*(1-phi*phi));

  ## obs variance for spawners
  tau_s <- 1/sigma_s;
  sigma_s ~ dnorm(0, 0.001) T(0,);

  ## maturity schedule
  ## unif vec for Dirch prior
  theta <- c(2,20,20,1)
  ## hyper-mean for maturity
  pi_eta ~ ddirch(theta);
  ## hyper-prec for maturity
  pi_tau ~ dnorm(0, 0.01) T(0,);
  for(t in 1:(n_yrs-age_min)) { pi_vec[t,1:A] ~ ddirch(pi_eta*pi_tau) }

  ## unprojectable early recruits;
  ## hyper mean across all popns
  Rec_mu ~ dnorm(0,0.001);
  ## hyper SD across all popns
  Rec_sig ~ dunif(0,100);
  ## precision across all popns
```

```

Rec_tau <- pow(Rec_sig,-2);
## multipliers for unobservable total runs
  ttl_run_mu ~ dunif(1,5);
  ttl_run_tau ~ dunif(1,20);

## get total cal yr returns for first age_min yrs
for(i in 1:(age_min)) {
  ln_tot_Run[i] ~ dnorm(ttl_run_mu*Rec_mu,Rec_tau/ttl_run_tau);
  tot_Run[i] <- exp(ln_tot_Run[i]);
}

## estimated harvest rate
for(t in 1:n_yrs) { h_rate[t] ~ dunif(0,1) }

##-----
## LIKELIHOOD
##-----
## 1st brood yr requires different innovation
## predicted recruits in BY t
covar[1] <- gamma * mod_cvrs[1];
ln_Rkr_a[1] <- mu_Rkr_a + covar[1];
E_ln_Rec[1] <- ln_Rkr_a[1] + ln_Sp[1] - beta*Sp[1] + phi*innov_1;
tot_ln_Rec[1] ~ dnorm(E_ln_Rec[1],tau_r);
res_ln_Rec[1] <- tot_ln_Rec[1] - E_ln_Rec[1];
## median of total recruits
tot_Rec[1] <- exp(tot_ln_Rec[1]);

## R/S
ln_RS[1] <- tot_ln_Rec[1] - ln_Sp[1];

## brood-yr recruits by age
for(a in 1:A) {
  Rec[1,a] <- tot_Rec[1] * pi_vec[1,a];
}

## brood years 2:(n_yrs-age_min)
for(t in 2:(n_yrs-age_min)) {
  ## predicted recruits in BY t
  covar[t] <- gamma * mod_cvrs[t];
  ln_Rkr_a[t] <- mu_Rkr_a + covar[t];
  E_ln_Rec[t] <- ln_Rkr_a[t] + ln_Sp[t] - beta*Sp[t] + phi*res_ln_Rec[t-1];
  tot_ln_Rec[t] ~ dnorm(E_ln_Rec[t],tau_r);
  res_ln_Rec[t] <- tot_ln_Rec[t] - E_ln_Rec[t];
  ## median of total recruits
  tot_Rec[t] <- exp(tot_ln_Rec[t]);
  ## R/S
  ln_RS[t] <- tot_ln_Rec[t] - ln_Sp[t];
  ## brood-yr recruits by age
  for(a in 1:A) {
    Rec[t,a] <- tot_Rec[t] * pi_vec[t,a];
  }
} ## end t loop over year

## get predicted calendar year returns by age

```

```

## matrix Run has dim [(n_yrs-age_min) x A]
## step 1: incomplete early broods
## first cal yr of this grp is first brood yr + age_min
for(i in 1:(age_max-age_min)) {
  ## projected recruits
  for(a in 1:i) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## imputed recruits
  for(a in (i+1):A) {
    lnRec[i,a] ~ dnorm(Rec_mu,Rec_tau);
    Run[i,a] <- exp(lnRec[i,a]);
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## step 2: info from complete broods
## first cal yr of this grp is first brood yr + age_max
for(i in A:(n_yrs-age_min)) {
  for(a in 1:A) {
    Run[i,a] <- Rec[i-a+1,a];
  }
  ## total run size
  tot_Run[i+age_min] <- sum(Run[i,1:A]);
  ## predicted age-prop vec for multinom
  for(a in 1:A) {
    age_v[i,a] <- Run[i,a] / tot_Run[i+age_min];
  }
  ## multinomial for age comp
  dat_age[i,1:A] ~ dmulti(age_v[i,1:A],dat_age[i,A+1]);
  lp_age[i] <- logdensity.multi(dat_age[i,1:A],age_v[i,1:A],dat_age[i,A+1]);
}

## get predicted calendar year spawners
## first cal yr is first brood yr
for(t in 1:n_yrs) {
  ## obs model for spawners
  # Sp[t] <- max(10,tot_Run[t] - dat_harv[t]);
  # est_harv[t] = h_rate[t] * tot_Run[t];
  # dat_harv[t] ~ dlnorm(log(est_harv[t]), 20);
  Sp[t] = tot_Run[t] - dat_harv[t];
  ln_Sp[t] <- log(Sp[t]);
  ln_dat_esc[t] ~ dnorm(ln_Sp[t], tau_s);
  lp_esc[t] <- logdensity.norm(ln_dat_esc[t],ln_Sp[t], tau_s);
}

```

```

} ## end model description

", file=file.path(jagsdir, "IPM_RK_cov_AR.txt"))

```

6 Fitting the models

Before fitting the model in JAGS, we need to specify:

1. the data and indices that go into the model;
2. the model parameters and states that we want JAGS to return;
3. the MCMC control parameters.

```

## 1. Data to pass to JAGS:
dat_jags <- list(dat_age = dat_age,
                ln_dat_esc = ln_dat_esc,
                dat_harv = dat_harv,
                A = A,
                age_min = age_min,
                age_max = age_max,
                n_yrs = n_yrs)

## 2. Model params/states for JAGS to return:
par_jags <- c("alpha", "E_Rkr_a", "mu_Rkr_a",
             "beta",
             "Sp", "Rec", "tot_ln_Rec", "ln_RS",
             "pi_eta", "pi_tau",
             "sigma_r", "sigma_s", "res_ln_Rec",
             "lp_age", "lp_esc")

## 3. MCMC control params:
mcmc_ctrl <- list(
  chains = 4,
  length = 1.25e5,
  burn = 5e4,
  thin = 100
)
## total number of MCMC samples after burnin
mcmc_samp <- mcmc_ctrl$length*mcmc_ctrl$chains/mcmc_ctrl$thin

```

Please note that the following code takes ~60 min to run on a quad-core machine with 3.5 GHz Intel processors.

```

## total number of models to fit
n_mods <- 1 + n_cov

## empty list for LOOIC values
LOOIC <- vector("list", n_mods)

## fit base model (if not already saved)
if(!file.exists(file.path(jagsdir, "fit_ricker_base.rds"))){
  mod_fit <- fit_jags("IPM_RK_AR.txt", dat_jags, par_jags, init_vals_AR, mcmc_ctrl)
  ## save results to file

```

```

saveRDS(mod_fit, file.path(jagsdir, "fit_ricker_base.rds"))
## compute LOOIC
LOOIC[[1]] <- looic(mod_fit, mcmc_ctrl)
}

## fit models with covariates
par_jags <- c(par_jags, "gamma")
for(i in seq(n_mods-1)) {
  if(!file.exists(file.path(jagsdir, paste0("fit_ricker_cov_", i, ".rds")))) {
    dat_jags$mod_cvrs <- cov_flow[,i]
    mod_fit <- fit_jags("IPM_RK_cov_AR.txt", dat_jags, par_jags,
                      init_vals_cov, mcmc_ctrl)
    ## save results to file
    saveRDS(mod_fit, file.path(jagsdir, paste0("fit_ricker_cov_", i, ".rds")))
    ## compute LOOIC
    LOOIC[[i+1]] <- looic(mod_fit, mcmc_ctrl)
  }
}
if(!file.exists(file.path(jagsdir, "LOOIC_values.rds"))) {
  saveRDS(LOOIC, file.path(jagsdir, "LOOIC_values.rds"))
} else {
  LOOIC <- readRDS(file.path(jagsdir, "LOOIC_values.rds"))
}

```

6.0.0.1 Convergence checks

```

base_mod <- readRDS(file.path(jagsdir, "fit_ricker_base.rds"))

par_conv <- c("alpha", "beta",
             "sigma_r", "sigma_s",
             "pi_tau", paste0("pi_eta[", seq(A), "]"))

## Gelman-Rubin
gelman.diag(base_mod[,par_conv])

## Potential scale reduction factors:
##
##          Point est. Upper C.I.
## alpha          1.06      1.06
## beta           1.11      1.29
## sigma_r        1.07      1.14
## sigma_s        1.64      6.51
## pi_tau         1.00      1.00
## pi_eta[1]      1.00      1.00
## pi_eta[2]      1.00      1.00
## pi_eta[3]      1.00      1.00
## pi_eta[4]      1.00      1.01
##
## Multivariate psrf
##
## 1.37

## autocorrelation
t(round(autocorr.diag(base_mod[,par_conv],

```

```

lags = seq(mcmc_ctrl$thin, 4*mcmc_ctrl$thin, mcmc_ctrl$thin),
relative=FALSE), 2))

##          Lag 100 Lag 200 Lag 300 Lag 400
## alpha         0.10   0.06   0.06   0.09
## beta          0.27   0.23   0.21   0.21
## sigma_r       0.23   0.22   0.19   0.18
## sigma_s       0.38   0.27   0.21   0.18
## pi_tau        0.06   0.04   0.05   0.03
## pi_eta[1]     0.03   0.01   0.01   0.03
## pi_eta[2]     0.03   0.03   0.03   0.02
## pi_eta[3]     0.03   0.04   0.03   0.03
## pi_eta[4]     0.16   0.12   0.12   0.08

```

7 Model selection

Via `loo()` and `compare()` with full table of results. Note that `elpd_diff` will be negative (positive) if the expected predictive accuracy for the first (second) model is higher.

```

## table of LOOIC values
tbl_LOOIC <- as.data.frame(round(compare(x = LOOIC), 1))
tbl_LOOIC$d_looic <- -2 * tbl_LOOIC$elpd_diff
tbl_LOOIC <- tbl_LOOIC[, c("p_loo", "se_p_loo", "looic", "se_looic", "d_looic")]
rownames(tbl_LOOIC) <- sub("model", "", rownames(tbl_LOOIC))
tbl_LOOIC <- tbl_LOOIC[order(as.numeric(rownames(tbl_LOOIC))),]
tbl_LOOIC <- data.frame(life_stage = c("base", cov_meta$life_stage),
  variable = c("NA", sub(" of 7-day mean", "", cov_meta$long_name)),
  begin = c("NA", cov_meta$begin),
  end = c("NA", cov_meta$end),
  lag = c(NA, cov_meta$lag_1),
  tbl_LOOIC)
tbl_LOOIC[order(tbl_LOOIC[, "looic"]), ]

##      life_stage variable begin   end lag p_loo se_p_loo looic se_looic d_looic
## 29 2+ outmigrants   Range 02-01 04-30  2  53.1    3.8 335.0   63.8    0.0
## 22          rearing    Min 07-01 09-30  1  53.3    3.9 336.8   63.8    1.8
## 26 2+ outmigrants   Max 02-01 04-30  2  53.6    4.0 336.9   63.9    2.0
##  6      prespawn    Max 04-01 06-30  0  53.8    4.1 340.0   63.8    5.0
##  9      prespawn    Min 05-01 05-31  0  51.9    4.0 340.0   63.2    5.0
## 10      prespawn    Max 05-01 05-31  0  52.5    4.0 340.5   63.3    5.6
## 15 1+ outmigrants   Min 04-01 06-30  1  52.2    4.3 340.5   63.7    5.4
## 30 2+ outmigrants   Max 04-01 04-30  2  52.1    4.0 340.7   63.0    5.6
##  7      prespawn   Range 04-01 06-30  0  53.7    4.2 340.8   63.8    5.8
## 18 1+ outmigrants   Range 04-01 06-30  1  52.1    4.1 341.0   63.5    6.0
## 28 2+ outmigrants   Min 02-01 04-30  2  53.8    4.0 341.2   63.5    6.2
## 11      prespawn    Min 07-01 09-30  0  53.2    4.2 342.0   63.5    7.0
## 16 1+ outmigrants   Median 04-01 06-30  1  54.0    3.9 342.4   63.1    7.4
## 13      incubation    Max 11-01 03-31  0  54.3    4.2 342.5   64.0    7.4
## 33 2+ outmigrants   Range 04-01 04-30  2  53.5    4.0 342.6   63.3    7.6
##  5      prespawn   Median 04-01 06-30  0  53.6    4.0 342.8   63.1    7.8
## 14      incubation   Median 11-01 03-31  0  53.9    4.2 342.9   63.6    7.8
##  2      prespawn    Max 11-01 03-31 -1  53.6    4.3 343.3   63.8    8.4
## 24          rearing    Max 07-01 09-30  1  54.9    4.4 343.9   64.2    8.8

```

## 27	2+	outmigrants	Median	02-01	04-30	2	54.2	4.4	344.1	64.0	9.0
## 23		rearing	Median	07-01	09-30	1	55.1	4.6	344.2	64.7	9.2
## 25		rearing	Range	07-01	09-30	1	54.9	4.5	344.2	64.3	9.2
## 8		prespawn	Min	04-01	04-30	0	58.2	4.4	344.6	65.2	9.6
## 3		prespawn	Median	11-01	03-31	-1	54.7	4.6	344.7	64.3	9.8
## 12		prespawn	Median	07-01	09-30	0	54.0	4.5	344.8	63.8	9.8
## 32	2+	outmigrants	Min	04-01	04-30	2	54.4	4.3	345.2	63.7	10.2
## 31	2+	outmigrants	Median	04-01	04-30	2	53.4	4.3	345.8	63.3	10.8
## 17	1+	outmigrants	Max	04-01	06-30	1	55.4	4.6	346.1	64.4	11.0
## 20	1+	outmigrants	Max	05-01	05-31	1	54.9	4.7	346.2	64.2	11.2
## 4		prespawn	Min	04-01	06-30	0	55.3	4.3	346.3	63.8	11.4
## 19	1+	outmigrants	Min	04-01	04-30	1	56.0	4.3	346.6	64.0	11.6
## 21	1+	outmigrants	Min	05-01	05-31	1	56.4	4.7	348.6	64.3	13.6
## 1		base	NA	NA	NA	NA	67.6	9.1	374.5	61.1	39.4

```
## best model
best_i <- which(tbl_L00IC[, "looic"] == min(tbl_L00IC[, "looic"]))
best_fit <- readRDS(file.path(jagsdir, paste0("fit_ricker_cov_", best_i, ".rds")))
```

8 Model diagnostics

8.1 Gelman & Rubin statistic

Here is a table of the Gelman & Rubin statistics (R_{hat}) for the estimated parameters. Recall that we set an upper threshold of 1.1, so values larger than that deserve some additional inspection.

```
## params of interest
par_conv <- c("alpha", "beta", "gamma",
             "sigma_r", "sigma_s",
             "pi_tau", paste0("pi_eta[", seq(A-1), "]"),
             paste0("Sp[", seq(n_yrs), "]"),
             paste0("tot_ln_Rec[", seq(n_yrs-age_min), "]"))

## Gelman-Rubin
gelman.diag(best_fit[, par_conv])

## Potential scale reduction factors:
##
##          Point est. Upper C.I.
## alpha          1.00      1.00
## beta           1.00      1.00
## gamma          1.00      1.00
## sigma_r        1.00      1.00
## sigma_s        1.00      1.00
## pi_tau         1.00      1.00
## pi_eta[1]      1.00      1.00
## pi_eta[2]      1.00      1.00
## pi_eta[3]      1.00      1.00
## Sp[1]          1.00      1.00
## Sp[2]          1.00      1.00
## Sp[3]          1.00      1.01
## Sp[4]          1.01      1.01
## Sp[5]          1.00      1.00
```

```

## Sp[6]          1.00      1.00
## Sp[7]          1.00      1.01
## Sp[8]          1.00      1.00
## Sp[9]          1.00      1.00
## Sp[10]         1.00      1.00
## Sp[11]         1.00      1.00
## Sp[12]         1.00      1.00
## Sp[13]         1.00      1.01
## Sp[14]         1.00      1.01
## Sp[15]         1.00      1.01
## Sp[16]         1.00      1.00
## Sp[17]         1.00      1.00
## Sp[18]         1.00      1.01
## Sp[19]         1.00      1.01
## tot_ln_Rec[1]  1.00      1.00
## tot_ln_Rec[2]  1.00      1.00
## tot_ln_Rec[3]  1.00      1.00
## tot_ln_Rec[4]  1.00      1.00
## tot_ln_Rec[5]  1.00      1.01
## tot_ln_Rec[6]  1.00      1.00
## tot_ln_Rec[7]  1.00      1.00
## tot_ln_Rec[8]  1.00      1.01
## tot_ln_Rec[9]  1.00      1.01
## tot_ln_Rec[10] 1.00      1.01
## tot_ln_Rec[11] 1.00      1.01
## tot_ln_Rec[12] 1.00      1.00
## tot_ln_Rec[13] 1.00      1.00
## tot_ln_Rec[14] 1.00      1.01
## tot_ln_Rec[15] 1.00      1.00
## tot_ln_Rec[16] 1.00      1.00
##
## Multivariate psrf
##
## 1.01

```

8.2 Autocorrelation

```

t(round(autocorr.diag(best_fit[,par_conv],
  lags = seq(mcmc_ctrl$thin, 3*mcmc_ctrl$thin, mcmc_ctrl$thin),
  relative=FALSE), 2))

##          Lag 100 Lag 200 Lag 300
## alpha      0.02  -0.01   0.03
## beta       0.03  -0.01   0.03
## gamma      0.01   0.01   0.00
## sigma_r    -0.01  -0.03   0.00
## sigma_s    0.27   0.11   0.05
## pi_tau     0.06   0.03   0.03
## pi_eta[1]   0.02   0.01  -0.01
## pi_eta[2]  -0.01   0.00   0.00
## pi_eta[3]  -0.01   0.00   0.00
## Sp[1]       0.04   0.02   0.02
## Sp[2]       0.03   0.02   0.01

```



```

## Sp[3]          -0.01  -0.01  -0.01
## Sp[4]          0.03  -0.01  0.02
## Sp[5]          0.05   0.02  0.01
## Sp[6]          0.09   0.04  0.02
## Sp[7]          0.05   0.03  0.01
## Sp[8]          0.09   0.00  0.01
## Sp[9]          0.11   0.03  0.01
## Sp[10]         0.17   0.06  0.02
## Sp[11]         0.12   0.04  0.00
## Sp[12]         0.18   0.07  0.02
## Sp[13]         0.13   0.02  0.01
## Sp[14]         0.06   0.01  0.00
## Sp[15]         0.07   0.01  0.00
## Sp[16]         0.05   0.00  0.00
## Sp[17]         0.07   0.04  0.02
## Sp[18]         0.09   0.04  0.01
## Sp[19]         0.07   0.03 -0.02
## tot_ln_Rec[1]  0.05   0.02  0.02
## tot_ln_Rec[2]  0.00   0.01  0.02
## tot_ln_Rec[3]  0.03  -0.01  0.00
## tot_ln_Rec[4]  0.08   0.00  0.01
## tot_ln_Rec[5]  0.12   0.05  0.01
## tot_ln_Rec[6]  0.13   0.03  0.01
## tot_ln_Rec[7]  0.08   0.03  0.00
## tot_ln_Rec[8]  0.17   0.06  0.01
## tot_ln_Rec[9]  0.11   0.02  0.01
## tot_ln_Rec[10] 0.05   0.03  0.01
## tot_ln_Rec[11] 0.06   0.00 -0.01
## tot_ln_Rec[12] 0.03  -0.02  0.01
## tot_ln_Rec[13] 0.04   0.03  0.01
## tot_ln_Rec[14] 0.11   0.05  0.02
## tot_ln_Rec[15] 0.05   0.03  0.01
## tot_ln_Rec[16] 0.02  -0.02  0.01

```

8.3 Effective sample sizes

```

floor(effectiveSize(best_fit))

```

```

##      E_Rkr_a      Rec[1,1]      Rec[2,1]      Rec[3,1]      Rec[4,1]      Rec[5,1]
##      5228          4763          4869          3526          4656          4485
##      Rec[6,1]      Rec[7,1]      Rec[8,1]      Rec[9,1]      Rec[10,1]     Rec[11,1]
##      4619          3878          4238          3769          4484          5141
##      Rec[12,1]     Rec[13,1]     Rec[14,1]     Rec[15,1]     Rec[16,1]     Rec[1,2]
##      4482          4993          4656          4483          4900          4519
##      Rec[2,2]      Rec[3,2]      Rec[4,2]      Rec[5,2]      Rec[6,2]      Rec[7,2]
##      4088          4476          4312          3877          3400          3703
##      Rec[8,2]      Rec[9,2]      Rec[10,2]     Rec[11,2]     Rec[12,2]     Rec[13,2]
##      3313          3892          4462          4446          4513          4140
##      Rec[14,2]     Rec[15,2]     Rec[16,2]     Rec[1,3]      Rec[2,3]      Rec[3,3]
##      3989          4036          5000          4017          4472          4292
##      Rec[4,3]      Rec[5,3]      Rec[6,3]      Rec[7,3]      Rec[8,3]      Rec[9,3]
##      3858          3494          3661          3291          3920          4612
##      Rec[10,3]     Rec[11,3]     Rec[12,3]     Rec[13,3]     Rec[14,3]     Rec[15,3]

```

##	4898	4352	4225	4166	4501	4424
##	Rec[16,3]	Rec[1,4]	Rec[2,4]	Rec[3,4]	Rec[4,4]	Rec[5,4]
##	4755	4412	4885	4793	4393	4346
##	Rec[6,4]	Rec[7,4]	Rec[8,4]	Rec[9,4]	Rec[10,4]	Rec[11,4]
##	3684	3649	4703	4472	4601	4093
##	Rec[12,4]	Rec[13,4]	Rec[14,4]	Rec[15,4]	Rec[16,4]	Sp[1]
##	4531	952	1450	2229	4134	4469
##	Sp[2]	Sp[3]	Sp[4]	Sp[5]	Sp[6]	Sp[7]
##	4611	5000	4889	4600	3809	4445
##	Sp[8]	Sp[9]	Sp[10]	Sp[11]	Sp[12]	Sp[13]
##	4095	3843	3373	3664	3183	3844
##	Sp[14]	Sp[15]	Sp[16]	Sp[17]	Sp[18]	Sp[19]
##	4471	4479	4475	3971	3957	4090
##	alpha	beta	gamma	ln_RS[1]	ln_RS[2]	ln_RS[3]
##	4823	4657	4897	4381	4808	5171
##	ln_RS[4]	ln_RS[5]	ln_RS[6]	ln_RS[7]	ln_RS[8]	ln_RS[9]
##	4579	4283	3510	3628	4032	3708
##	ln_RS[10]	ln_RS[11]	ln_RS[12]	ln_RS[13]	ln_RS[14]	ln_RS[15]
##	3610	3794	3478	3856	4181	4865
##	ln_RS[16]	lp_age[1]	lp_age[2]	lp_age[3]	lp_age[4]	lp_age[5]
##	4777	4945	4949	4831	4814	5000
##	lp_age[6]	lp_age[7]	lp_age[8]	lp_age[9]	lp_age[10]	lp_age[11]
##	4962	4801	4911	5000	5107	4912
##	lp_age[12]	lp_age[13]	lp_age[14]	lp_age[15]	lp_age[16]	lp_esc[1]
##	5136	4720	5124	4883	1847	3443
##	lp_esc[2]	lp_esc[3]	lp_esc[4]	lp_esc[5]	lp_esc[6]	lp_esc[7]
##	3683	3859	3079	3239	3113	2958
##	lp_esc[8]	lp_esc[9]	lp_esc[10]	lp_esc[11]	lp_esc[12]	lp_esc[13]
##	2941	3113	3217	3246	3274	3282
##	lp_esc[14]	lp_esc[15]	lp_esc[16]	lp_esc[17]	lp_esc[18]	lp_esc[19]
##	3250	3532	3273	3266	3158	3709
##	mu_Rkr_a	pi_eta[1]	pi_eta[2]	pi_eta[3]	pi_eta[4]	pi_tau
##	5121	4831	4981	5000	1892	4371
##	res_ln_Rec[1]	res_ln_Rec[2]	res_ln_Rec[3]	res_ln_Rec[4]	res_ln_Rec[5]	res_ln_Rec[6]
##	4766	4714	4999	4901	5577	4200
##	res_ln_Rec[7]	res_ln_Rec[8]	res_ln_Rec[9]	res_ln_Rec[10]	res_ln_Rec[11]	res_ln_Rec[12]
##	4999	4988	4204	5000	4745	4898
##	res_ln_Rec[13]	res_ln_Rec[14]	res_ln_Rec[15]	res_ln_Rec[16]	sigma_r	sigma_s
##	4905	4745	4673	4542	5251	2691
##	tot_ln_Rec[1]	tot_ln_Rec[2]	tot_ln_Rec[3]	tot_ln_Rec[4]	tot_ln_Rec[5]	tot_ln_Rec[6]
##	4353	4872	4852	4448	3513	3829
##	tot_ln_Rec[7]	tot_ln_Rec[8]	tot_ln_Rec[9]	tot_ln_Rec[10]	tot_ln_Rec[11]	tot_ln_Rec[12]
##	4148	3414	4021	4470	4813	4817
##	tot_ln_Rec[13]	tot_ln_Rec[14]	tot_ln_Rec[15]	tot_ln_Rec[16]		
##	4401	3523	4235	5270		