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1 Workflow

Virtual Screening Pipeline

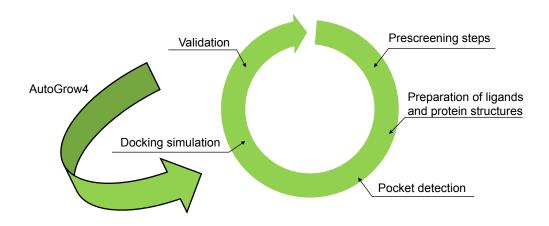


Figure 1: **Virtual Screening Pipeline.** It consists of the following steps: Prescreening steps, e.g. Toxicology Check, Preparation of ligands and protein structures with PyMOL by removing water molecules and converting to the correct file format, Pocket detection via Fpocket, Docking simulation with $AutoDock\ Vina\ using\ the\ Vinardo\ scoring\ function,\ using\ the\ results\ from\ that\ to\ assimilate\ new ligands\ using\ <math>AutoGrow\ 4$ to rerun the Docking simulation and lastly Validation using Molecular Dynamics Simulations with GROMACS.

2 Material and Methods

2.1 Structure and Binding Pocket Detection

The 6ZSL¹ structure from the RCSB PDB database was chosen for the detection of binding pockets. Fpocket [Guilloux et al., 2009] and Prankweb² [Jendele et al., 2019] were used to predict the binding pockets.

2.2 Test Dataset from ZINC15

A drug-like subset of 95,438,992 molecules was downloaded from the ZINC15-Database [Sterling and Irwin, 2015] for initial consideration.

2.3 Toxicology prediction using eToxPred

The machine learning based tool eToxPred [Pu et al., 2019] was used for toxicology prediction of the given molecules. Based on the information given by the paper the threshold was set to 0.58 for a substance to be considered toxic.

¹http://dx.doi.org/10.2210/pdb6zs1/pdb

²https://prankweb.cz/

2.4 Docking

For docking 5000 molecules out of the molecules that were predicted to be non-toxic were randomly chosen for docking. Docking was performed using *AutoDock Vina* [Forli et al., 2016] using the *Vinardo* scoring function [Quiroga and Villarreal, 2016].

2.5 PDB Preparation

The crystallized protein structures of NSP13 were downloaded as previously described at RCSB ³. Missing residues and atoms were added using the Python package 'pdbfixer' [Eastman et al., 2013].

2.6 Molecular Dynamics Simulation using *GROMACS*

GROMACS was used to validate the docking results by evaluating the ability of the top ligands to stay in the binding pocket [Bekker et al., 1993].

2.7 Visualization and Analysis of MD simulation results

Using the program *VMD* it is possible to look at the trajectory of the MD simulations of the different molecules [Humphrey et al., 1996].

2.8 Second Docking Step

Another Docking round again using *AutoDock Vina* with the *Vinardo* scoring function was performed. As per the task of this project a subset of the European Chemical Biology Database (ECBD)⁴, specifically the subset: https://ecbd.eu/compound/#lib{value='4'}.

2.9 Generating New Molecules

AutoGrow 4 can generate new ligands on the basis of ligands and a specified binding pocket which are likely to have a similar or even higher affinity than the ligands used as input [Spiegel and Durrant, 2020].

3 Results

3.1 General Docking Results ZINC15 dataset subset

The resulting binding energy of our 5000 starting molecules were loaded visualized in a histogram using R (see Figure ??). Most of the molecules have binding affinities between -2.5 and -10 $\left[\frac{kJ}{mol}\right]$. A small fraction of molecules has binding affinities below -10 $\left[\frac{kJ}{mol}\right]$.

3.2 Reranking our top 10 molecules based on the total protein-ligand interaction based on MD simulation results

Based on the MD simulations the ranking of the ligands amongst each other changed slightly even though the overall trend of the best ligands staying the best can be observed for most of them.

3.3 Visualizing pocket stability by calculating the RMSD between protein and ligand

While the best ligands mostly stayed the best even after reevaluation it could be observed that only the RMSD values for the top 3 ligands were particularly low.

³https://www.rcsb.org

⁴https://ecbd.eu

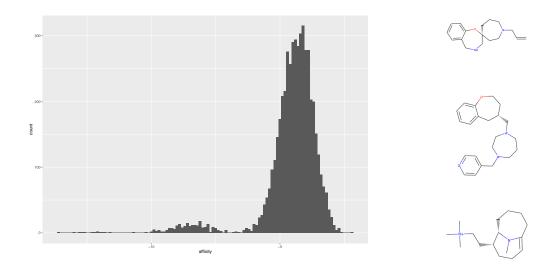


Figure 2: On the left: A histogram of the frequency-counts of the determined binding affinities in $[\frac{kJ}{mol}]$. On the right: Our top three candidates(ZINC000019015192, ZINC000095523345 and

ZINC000101042701) in descending order.

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