

cobalt

Covariate Balance Tables and Plots

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April 1, 2020

Introduction to cobalt

- Developed by Noah Greifer
 - PhD student, UNC Chapel Hill
- Standardized balance measures
 - different methods for conditioning
 - different packages available for conditioning (optmatch, MatchIt, CBPS, ebal, WeightIt, twang)
- Tables and beautiful plots
- Can be used with clustered data, multiply imputed data, continuous treatment



Review of Steps used in Pre-Processing

1. Estimate balancing scores (e.g., propensity scores)
2. Condition on balancing scores
 - Weighting
 - Stratification
 - Matching
3. Check the balance on covariates- compare balance before and after conditioning
4. Iterate
5. Estimate average treatment effect after conditioning

Balance Evaluation: Why?

Balance is crucial

- We are trying to emulate a randomized experiment
- Conditional on true propensity scores, distribution of observed covariates is independent of treatment (Austin, 2011)
- After conditioning on estimated propensity scores, need to check balance
- If balance is inadequate, effect estimate may be biased

Balance Evaluation

- Balance evaluation is very important but...
- Applied studies rarely report balance evaluation. Evaluation is often inappropriate (Greifer, 2017; Austin, 2009).
- Over 66% of applied studies that used propensity scores assessed balance using statistical tests (Thoemmes & Kim, 2011).
- However, statistical tests are not recommended for balance evaluation.
 - Interest in sample not population (Stuart, 2008; Stuart, 2010; Austin, 2011)
 - Reduced power (Imai et al., 2008)
 - `coba1t` does not provide statistical tests

Criteria

- Standardized difference in means (for continuous covariates)
- Variance ratios (for continuous)
- Raw difference in proportions (for binary)
- Love plots, density plots, bar plots

Standardized Difference in Means (Continuous)

Standardized difference in means: the estimate of mean difference (before/after conditioning) divided by standard deviation of the covariate from the unadjusted sample (Greifer, 2017; Rosenbaum & Rubin, 1983; Austin, 2009).

- Stuart (2008) recommended use of standard deviation from unadjusted sample even when checking balance after conditioning
 - Compare mean differences in unadjusted and adjusted sample- the denominator is the same
- sd in treated group (default for ATT), untreated group (ATU), or pooled sd (ATE)
- Threshold of .1 is recommended by Stuart et al. (2013).

Variance Ratio (Continuous)

- Use SMDs to compare center of distributions but also important to compare variance- spread of distributions (Greifer, 2017)
- **Variance ratio:** ratio of variances of continuous covariates in treated and untreated groups
 - Ratios closer to 1 indicate variances of the two groups are similar (Greifer, 2017)
 - Recommended thresholds are 0.5 and 2 (Rubin, 2001)
 - In `coba1t`, the larger variance is in the numerator

Weighted Variance Calculation

Formula used in `coba1t` to calculate variances after conditioning if weights are involved:

$$s_w^2 = \left(\frac{\sum_{i=1}^n w_i}{\left(\sum_{i=1}^n w_i \right)^2 - \sum_{i=1}^n w_i^2} \right) \sum_{i=1}^n w_i (x_i - \bar{x}_w)^2$$

Here w_i is weight for person i (from weighting or matching), x_i is value of covariate for person i and \bar{x}_w is the weighted mean of x within each treatment group.

This formula is recommended by Austin (2008) and Austin & Stuart (2015).

Raw Difference in Proportions (Binary)

- For binary covariates, raw differences in proportions between treated and untreated groups (before/ after conditioning) are used to evaluate balance (Greifer, 2017).
 - Already on the same scale
- No variance ratios- variance of binary variables derived from proportion so ratios do not provide new information (Greifer, 2017)

Interactions and Squared Terms

Assess balance on two-way interactions and squared terms (Rubin, 2001; Austin, 2009; Stuart, 2010)

- Interactions because joint distributions should be similar
- Comparing means of squared terms (for continuous predictors) equivalent to comparing variances of treatment and control group (Austin & Stuart, 2015)

Effective Sample Size

Effective Sample Size - "a measure of the sample size a non-weighted sample would have to have to achieve the same level of precision as the weighted sample" (Greifer, 2017; Ridgeway et al., 2016)

$$ESS = \frac{(\sum_{i=1}^n w_i)^2}{\sum_{i=1}^n w_i^2}$$

- Proportionally larger weights lead to:
 - Lower ESS
 - Larger variance of weighted mean
 - Loss of precision
- `cobalt` calculates ESS

cobalt demonstration

Libraries

```
# install.packages("cobalt")  
library(cobalt)
```

```
library(tidyverse)  
library(MatchIt)
```

Data

```
Algebra_dat <-  
  read_csv("8th-grade-Algebra-data.csv") %>%  
  mutate(Locale = factor(Locale,  
                          levels = c("R", "S", "U"),  
                          labels = c("Rural", "Suburban", "Urban")))  
  
Algebra_dat_org <- Algebra_dat
```

Formula

- There is a function in `cobalt` that takes in the outcome and a data frame or tibble containing the covariates and creates a formula based on that. It doesn't seem to have an easy way to add interaction terms or polynomial terms though.

```
# dataset with covariates  
covs <- Algebra_dat %>%  
  select(Math, SES, Locale)
```

```
f_lin <- f.build("D", covs)  
f_lin
```

```
## D ~ Math + SES + Locale  
## <environment: 0x7fb46c36b558>
```


Weighting: Calculations

Here, we are just estimating the propensity scores and calculating ATT weights.

```
# fitting propensity score model
ps_logit <- glm(f_lin, data = Algebra_dat, family = "binomial")

# estimating propensity scores
Algebra_dat$ps <- predict(ps_logit, type = "response")

# calculate the weights - ATT weighting by odds of treatment
Algebra_dat <- Algebra_dat %>%
  mutate(att_wt = D + (1 - D) * ps/(1-ps))
```

Weighting: Balance

Just a table with standardized difference in means after adjustment. `cobalt` normalizes the weights automatically.

```
bal.tab(f_lin,    # formula
        data = Algebra_dat, # data
        weights = "att_wt", # weights
        method = "weighting", # method
        m.threshold = .1) # threshold to judge
```

```

## Balance Measures
##
##           Type Diff.Adj           M.Threshold
## Math           Contin.    0.1390 Not Balanced, >0.1
## SES            Contin.    0.1829 Not Balanced, >0.1
## Locale_Rural    Binary   -0.0398   Balanced, <0.1
## Locale_Suburban Binary   -0.0288   Balanced, <0.1
## Locale_Urban    Binary    0.0686   Balanced, <0.1
##
## Balance tally for mean differences
##
##           count
## Balanced, <0.1      3
## Not Balanced, >0.1  2
##
## Variable with the greatest mean difference
##   Variable Diff.Adj           M.Threshold
##       SES    0.1829 Not Balanced, >0.1
##
## Effective sample sizes
##           Control Treated
## Unadjusted 390.000      610
## Adjusted   57.511      610

```

Weighting: Balance

Adding variance ratios and balance measures for unadjusted sample.

```
b_w1 <- bal.tab(f_lin,  
               data = Algebra_dat,  
               weights = "att_wt",  
               method = "weighting",  
               disp.v.ratio = TRUE, # ask for var ratios  
               un = TRUE) # asked for unadjusted sample stats  
  
b_w1
```

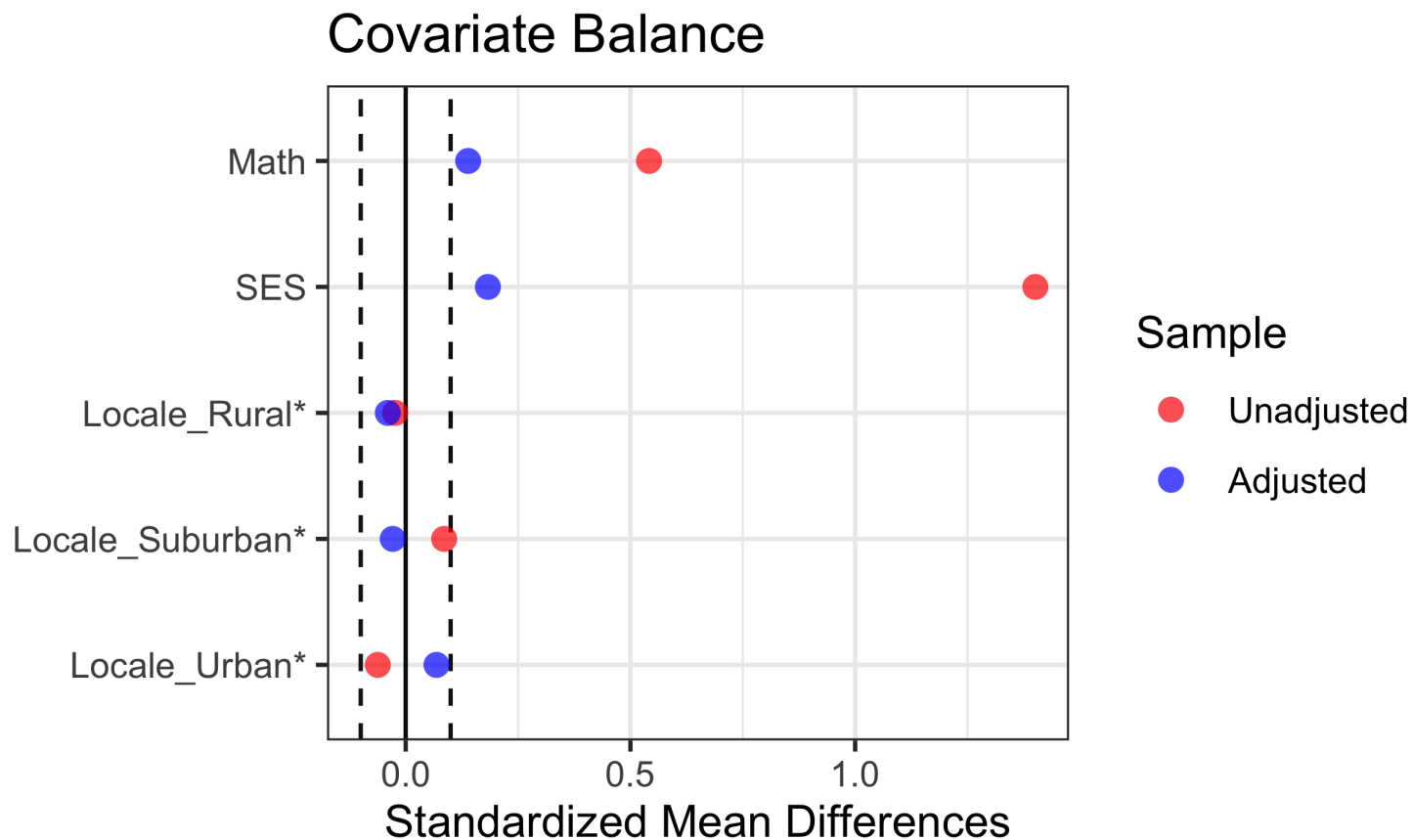
```
## Balance Measures
##              Type Diff.Un V.Ratio.Un Diff.Adj V.Ratio.Adj
## Math          Contin.  0.5415    1.0440    0.1390    0.9415
## SES            Contin.  1.4009    1.2191    0.1829    1.0423
## Locale_Rural   Binary -0.0235             -0.0398
## Locale_Suburban Binary  0.0855             -0.0288
## Locale_Urban   Binary -0.0620             0.0686
##
## Effective sample sizes
##              Control Treated
## Unadjusted 390.000    610
## Adjusted   57.511    610
```

Weighting: Love Plot

- Way to visualize results from balance evaluation
- Named after Dr. Thomas E. Love

```
love.plot(b_w1,    # add the table here
          threshold = .1, # threshold
          colors = c("red", "blue"), # change the colors
          size = 3, # size of dots
          alpha = .7, # transparency
          stars = "raw") + # change the x axis label
theme_bw()
```

Weighting: Love Plot



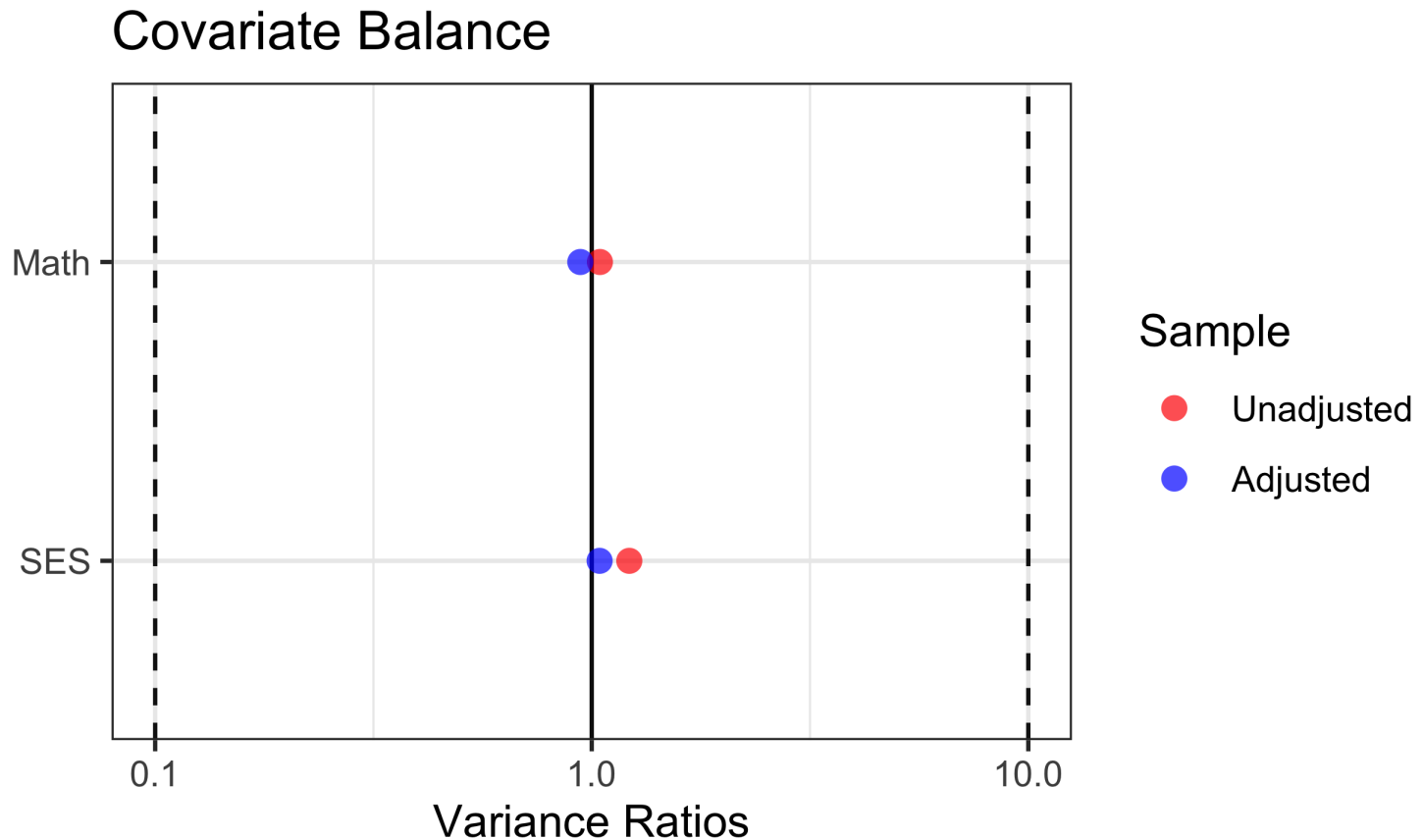
Weighting: Love Plot Default

```
love.plot(b_w1,  
          threshold = .1,  
          stars = "raw")
```


Weighting: Love Plot for Variance Ratios

```
love.plot(b_w1,  
          threshold = .1,  
          colors = c("red", "blue"),  
          size = 3,  
          alpha = .7,  
          stat = "variance.ratios", # just ask for var ratios here  
          stars = "raw") +  
theme_bw()
```

Weighting: Love Plot for Variance Ratios



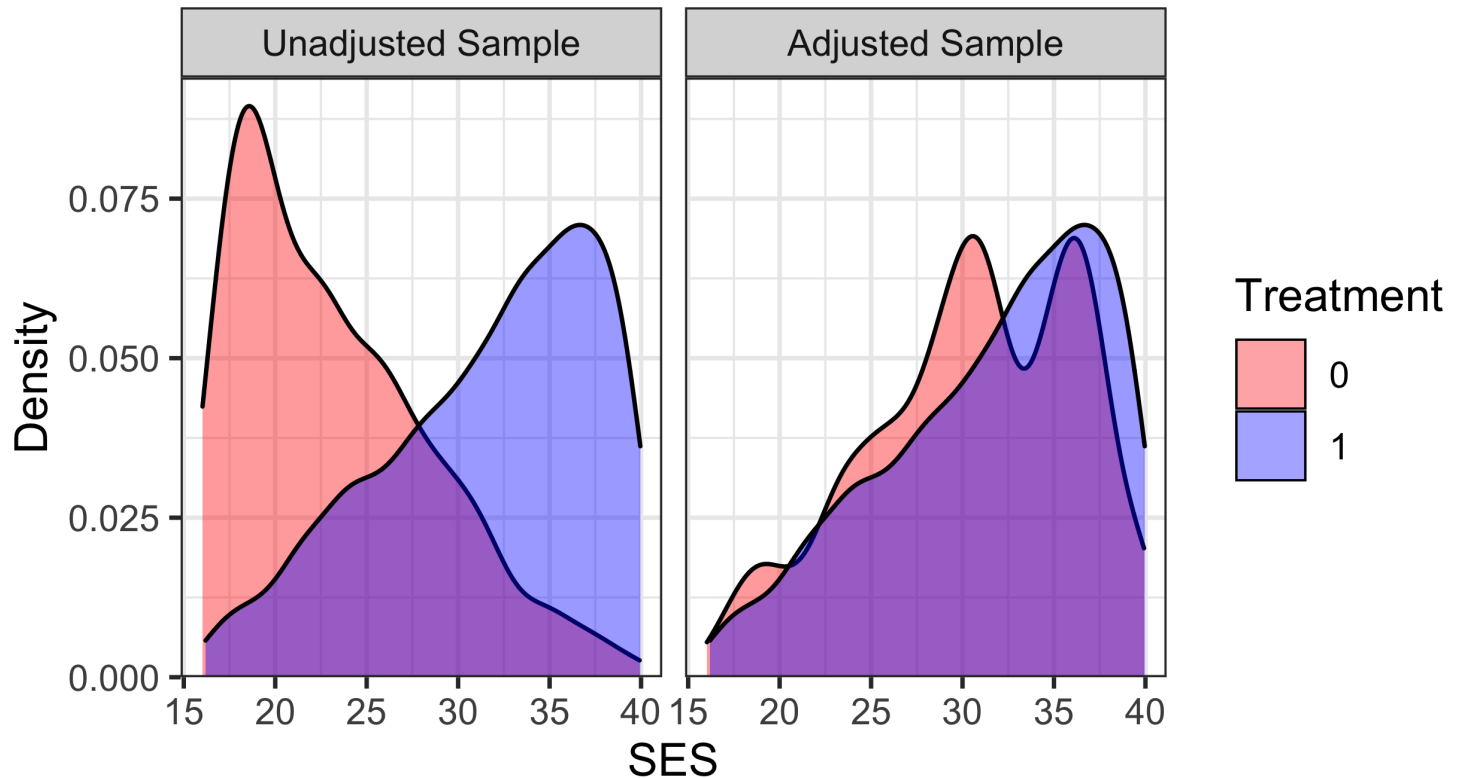
Weighting: Density Plot (SES)

Balance plots to evaluate similarities in univariate distributions of a covariate in treated and untreated groups. For continuous covariates, we look at density plots.

```
bal.plot(f_lin,  
        data = Algebra_dat,  
        weights = "att_wt",  
        method = "weighting",  
        var.name = "SES", # ask for a particular variable SES  
        which = "both", # ask for both unadjusted and adjusted samp  
        colors = c("red", "blue")) +  
theme_bw()
```

Weighting: Density Plot (SES)

Distributional Balance for "SES"

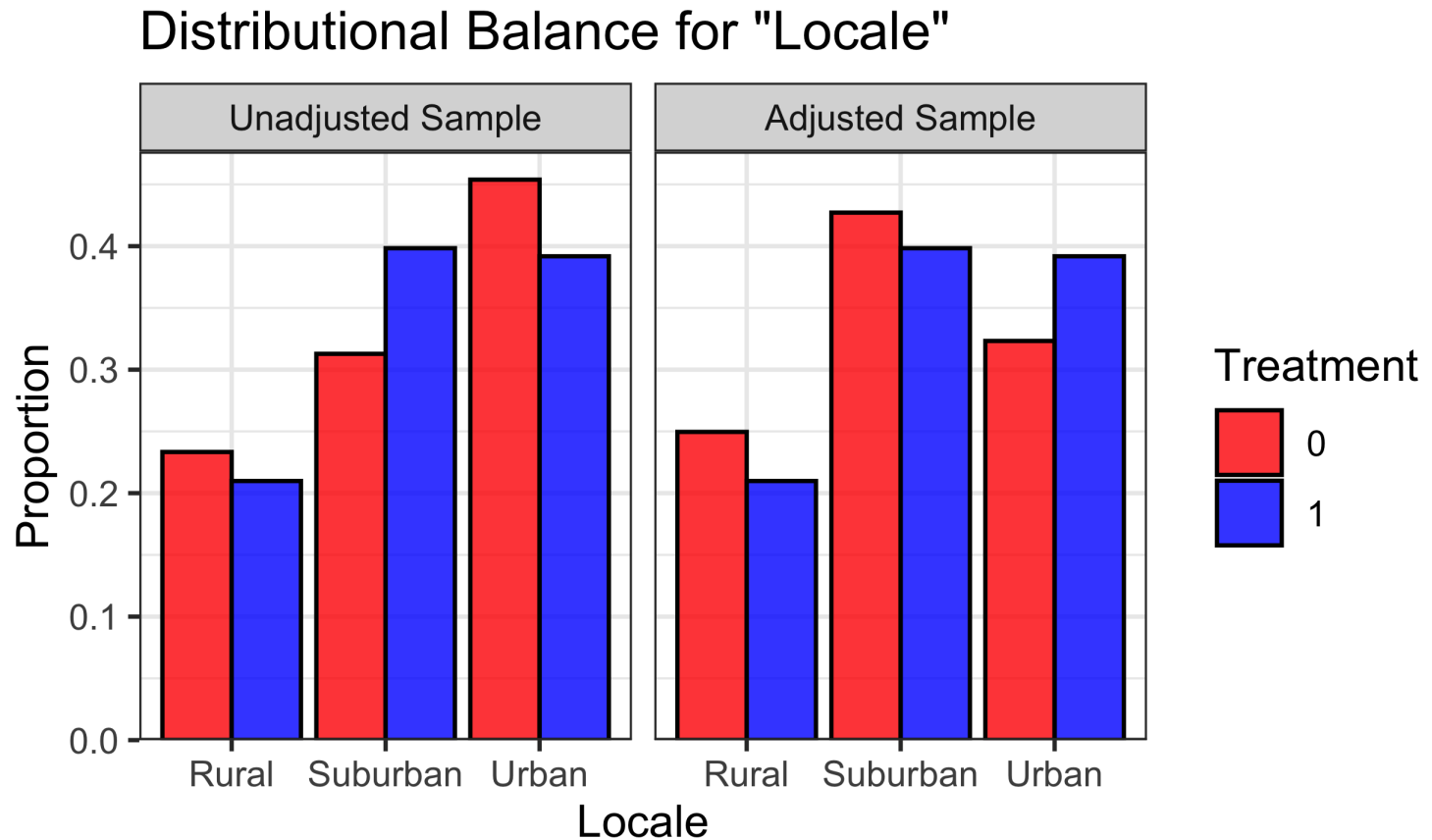


Weighting: Bar Plot (Locale)

If the covariate is binary, `bal.plot` will create bar plots.

```
bal.plot(f_lin,  
         data = Algebra_dat,  
         weights = "att_wt",  
         method = "weighting",  
         var.name = "Locale",  
         which = "both",  
         alpha = .2,  
         position = "stack", #alpha and position don't get passed to  
         colors = c("red", "blue")) +  
theme_bw()
```

Weighting: Bar Plot (Locale)



Weighting: Int and Sq Terms

Balance should be evaluated for two-way interactions and squared terms of continuous covariates (Stuart, 2010). We can add `int = TRUE` and `poly = 2` as arguments. Note that the output presents variance ratios of squared terms which doesn't mean what it should.

```
b_w2 <- bal.tab(f_lin,
               data = Algebra_dat,
               weights = "att_wt",
               method = "weighting",
               int = TRUE, # add interaction terms
               poly = 2, # add sq terms
               disp.v.ratio = TRUE,
               un = TRUE)
```

```
b_w2
```

Balance Measures

##	Type	Diff.Un	V.Ratio.Un	Diff.Adj	V.Ratio.Adj
## Math	Contin.	0.5415	1.0440	0.1390	0.9415
## SES	Contin.	1.4009	1.2191	0.1829	1.0423
## Locale_Rural	Binary	-0.0235		-0.0398	
## Locale_Suburban	Binary	0.0855		-0.0288	
## Locale_Urban	Binary	-0.0620		0.0686	
## Math ²	Contin.	0.5315	1.1252	0.1364	0.9713
## SES ²	Contin.	1.3007	1.6729	0.1951	1.0783
## Math * SES	Contin.	1.5159	1.5552	0.2509	1.3351
## Math * Locale_Rural	Contin.	-0.0311	1.0174	-0.0897	0.8926
## Math * Locale_Suburban	Contin.	0.2058	1.2359	-0.0468	0.9861
## Math * Locale_Urban	Contin.	-0.0881	1.0437	0.1452	1.0940
## SES * Locale_Rural	Contin.	0.0886	1.6501	-0.1071	0.8389
## SES * Locale_Suburban	Contin.	0.3295	1.9538	-0.0389	1.0024
## SES * Locale_Urban	Contin.	0.1195	1.7533	0.1948	1.3097

##

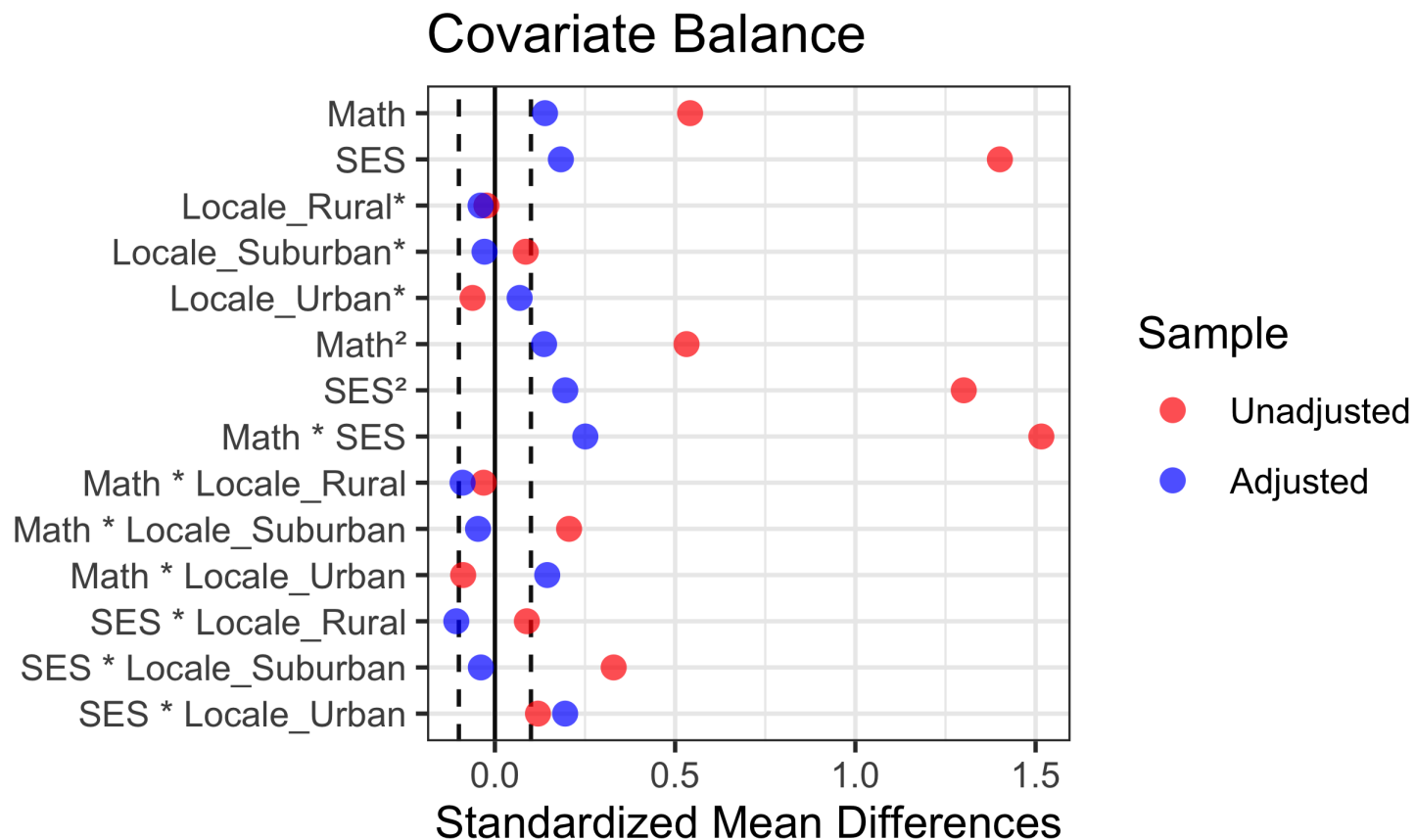
Effective sample sizes

##	Control	Treated
## Unadjusted	390.000	610
## Adjusted	57.511	610

Int & Sq Terms: Love Plot

```
love.plot(b_w2,  
          threshold = .1,  
          colors = c("red", "blue"),  
          size = 3,  
          alpha = .7,  
          stars = "raw") +  
theme_bw()
```

Int & Sq Terms: Love Plot



Iterate...

If balance is not adequate, respecify propensity score model (e.g., add interactions, squared terms) and assess balance again.

Stratification: Calculation

```
# Quintiles based on treatment group
ATT_quint <- with(Algebra_dat, quantile(ps[D==1], seq(0,1,0.2)))

Algebra_dat$quintile <- cut(Algebra_dat$ps, ATT_quint,
                           labels = c("A", "B", "C", "D", "E"),
                           include.lowest = TRUE)
```

Stratification: Balance

Specify the subclass and method:

```
b_s1 <- bal.tab(f_lin,  
               data = Algebra_dat,  
               subclass = "quintile", # add the variable that defines  
               method = "subclassification", # specify method  
               disp.subclass = TRUE,  
               disp.v.ratio = TRUE,  
               un = TRUE)
```

```
b_s1
```

```

## Balance by subclass
## - - - Subclass A - - -
##           Type Diff.Adj V.Ratio.Adj
## Math      Contin.    0.1174      1.0884
## SES        Contin.    0.2457      1.0510
## Locale_Rural Binary    0.0172
## Locale_Suburban Binary    0.0130
## Locale_Urban Binary   -0.0302
##
## - - - Subclass B - - -
##           Type Diff.Adj V.Ratio.Adj
## Math      Contin.   -0.0692      1.0314
## SES        Contin.    0.1215      1.0080
## Locale_Rural Binary    0.0193
## Locale_Suburban Binary    0.0468
## Locale_Urban Binary   -0.0661
##
## - - - Subclass C - - -
##           Type Diff.Adj V.Ratio.Adj
## Math      Contin.    0.2728      0.7489
## SES        Contin.   -0.0194      0.6920
## Locale_Rural Binary   -0.0893
## Locale_Suburban Binary  -0.2013
## Locale_Urban Binary    0.2905
##
## - - - Subclass D - - -
##           Type Diff.Adj V.Ratio.Adj

```

Stratification: Across Subclasses

```
## Balance measures across subclasses
##
##           Type Diff.Un V.Ratio.Un Diff.Adj V.Ratio.Adj
## Math          Contin.  0.5415      1.0440   0.0612      1.0376
## SES            Contin.  1.4009      1.2191   0.1537      0.9329
## Locale_Rural    Binary -0.0235              -0.0603
## Locale_Suburban Binary  0.0855              -0.0212
## Locale_Urban    Binary -0.0620              0.0815
##
## Sample sizes by subclass
##           A    B    C    D    E    All
## Control  258   62   18    6    2   390
## Treated  122  122  122  122  122   610
## Total    380  184  140  128  124  1000
```

Stratification: Love Plot Fail

```
love.plot(b_s1)
```

```
## Error in is_not_null(facet): object 'facet' not found
```


Matching: Balance

Matching without replacement

- More treated than untreated units- discards treated units
- Depends on order of treated units
- Starts with treated units with highest propensity scores
 - Throws out those with lower- even though better match

```
Algebra_dat_org <- as.data.frame(Algebra_dat_org)

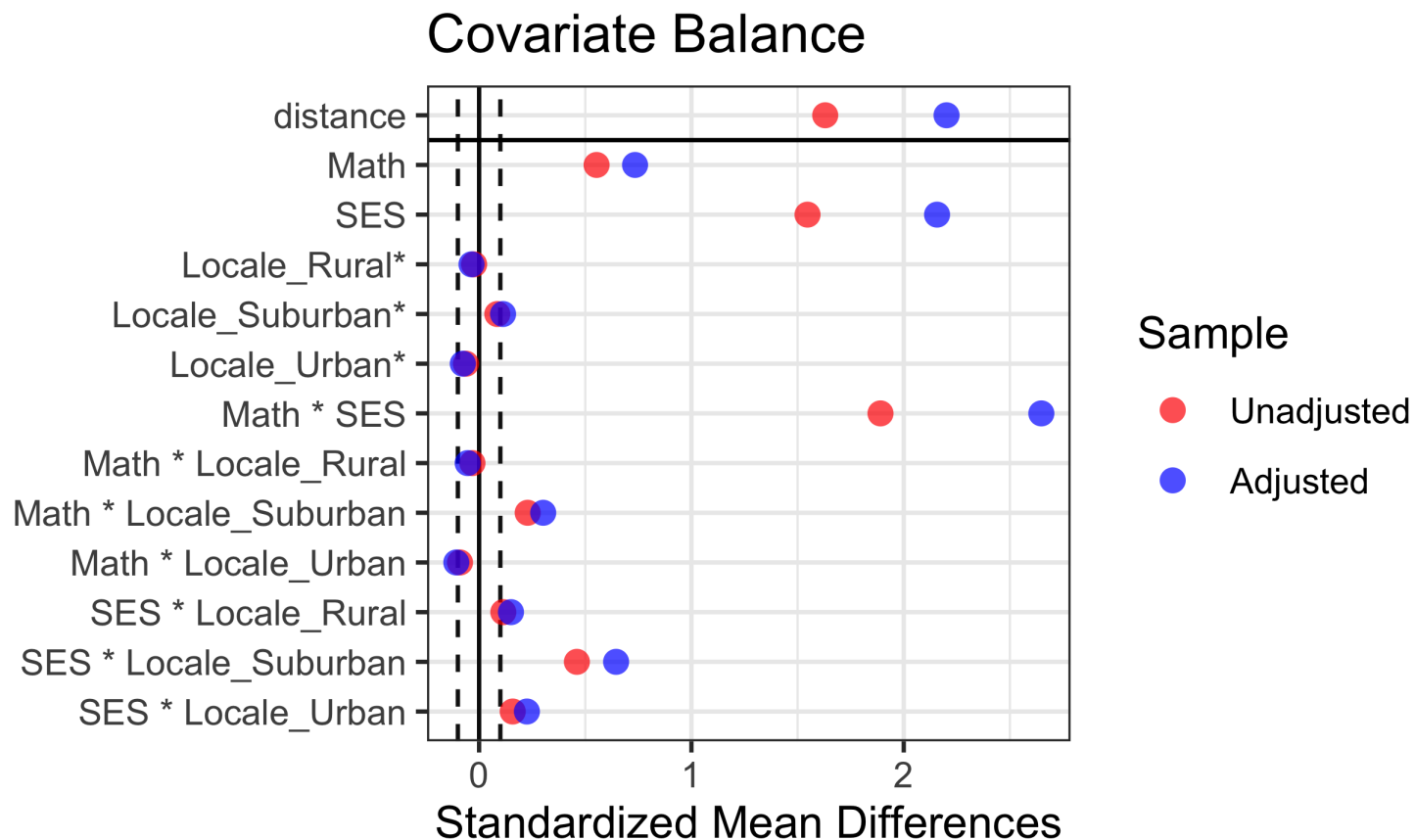
m_out <- matchit(D ~ Math + SES + Locale , data = Algebra_dat_org,
                 method = "nearest", distance = "logit")

b_m1 <- bal.tab(m_out,    # the output of matchit
               un = TRUE,
               int = TRUE)
```

Matching: Love Plot

```
love.plot(b_m1,  
          threshold = .1,  
          colors = c("red", "blue"),  
          size = 3,  
          alpha = .7,  
          stars = "raw") +  
theme_bw()
```

Matching: Love Plot



Better Match

- Matching with replacement- match an untreated unit more than once
- Caliper- maximum tolerated difference- prevents matching with whatever nearest unit available

```
better_match <- matchit(D ~ Math + SES + Locale,  
                        data = Algebra_dat_org,  
                        distance = "logit",  
                        method = "nearest",  
                        replace = TRUE,  
                        caliper = 0.1)
```

Better Match: Balance Table

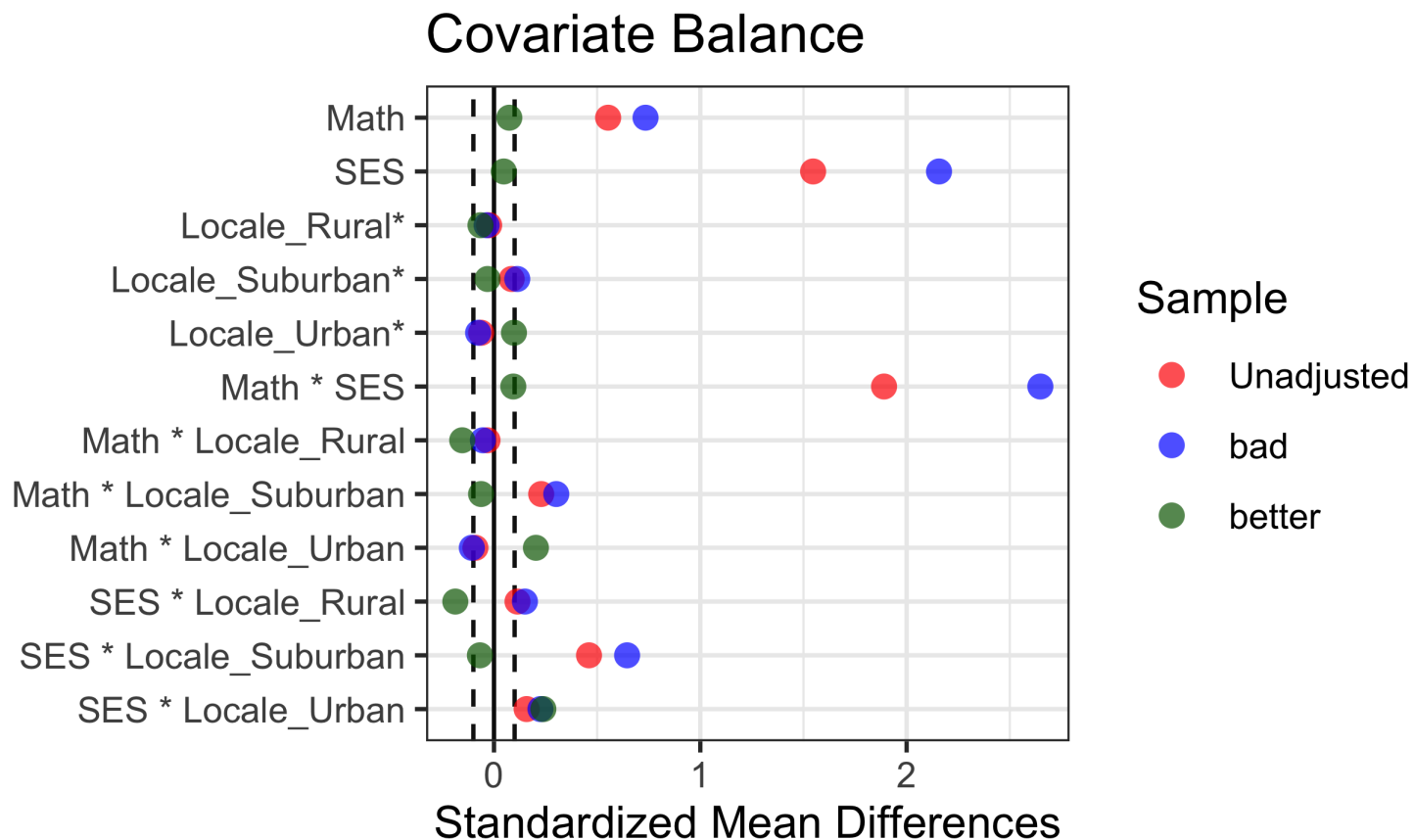
- We can compare the two different matching methods
- The `get.w` function will extract weights from `matchit` results
- Specify data frame with weights and specify method as "weighting"

```
b_m2 <- bal.tab(f_lin,
               data = Algebra_dat,
               weights = data.frame(      # create a data frame with weights
                 bad = get.w(m_out),
                 better = get.w(better_match)),
               method = "weighting",
               int = TRUE)
```

Better Match: Love Plot

```
love.plot(b_m2,  
          colors = c("red", "blue", "darkgreen"),  
          size = 3,  
          alpha = .7,  
          threshold = .1,  
          stars = "raw") +  
  theme_bw()
```

Better Match: Love Plot



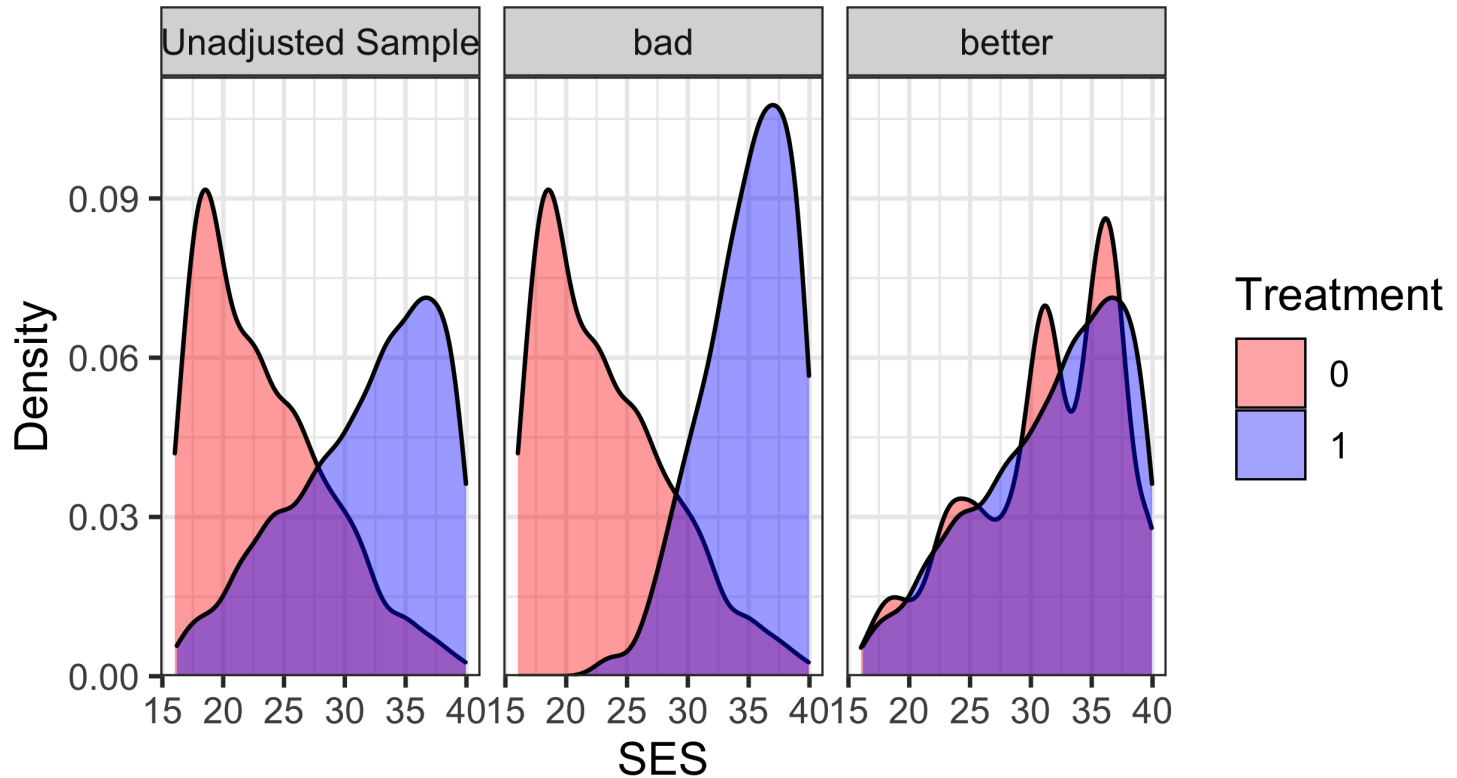
Better Match: Density Plots

Can compare the two matching methods in `bal.plot` too. Here creating density plots for SES.

```
bal.plot(f_lin,
        data = Algebra_dat,
        weights = data.frame( # create a data frame with weights
            bad = get.w(m_out),
            better = get.w(better_match)),
        method = "weighting",
        var.name = "SES", # specify var name
        colors = c("red", "blue"),
        which = "both",
        alpha = .2) + # alpha doesn't work
theme_bw()
```


Better Match: Density Plots

Distributional Balance for "SES"



Matching: Effective Sample Size

```
b_m2 <- bal.tab(f_lin,  
               data = Algebra_dat,  
               weights = data.frame(  
                 bad = get.w(m_out),  
                 better = get.w(better_match)),  
               method = "weighting",  
               int = TRUE)
```

b_m2

Matching: Effective Sample Size

```
## Balance Measures
##
##          Type Diff.bad Diff.better
## Math      Contin.    0.7345    0.0742
## SES        Contin.    2.1569    0.0475
## Locale_Rural Binary   -0.0359   -0.0656
## Locale_Suburban Binary    0.1128   -0.0311
## Locale_Urban Binary   -0.0769    0.0967
## Math * SES  Contin.    2.6481    0.0935
## Math * Locale_Rural Contin. -0.0540   -0.1540
## Math * Locale_Suburban Contin.  0.3019   -0.0625
## Math * Locale_Urban Contin. -0.1075    0.2032
## SES * Locale_Rural Contin.    0.1500   -0.1875
## SES * Locale_Suburban Contin.  0.6452   -0.0683
## SES * Locale_Urban Contin.  0.2251    0.2392
##
## Effective sample sizes
##          Control Treated
## All      390.000    610
## bad      390.000    390
## better   33.093    610
```

References

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Rubin, D. B. (2001). Using Propensity Scores to Help Design Observational Studies: Application to the Tobacco Litigation. *Health Services and Outcomes Research Methodology*, 2(3-4), 169–188. <http://doi.org/10.1023/A:1020363010465>

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<http://doi.org/10.1002/sim.3207>

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Thoemmes, F. J., & Kim, E. S. (2011). A Systematic Review of Propensity Score Methods in the Social Sciences. *Multivariate Behavioral Research*, 46(1), 90–118. <http://doi.org/10.1080/00273171.2011.540475>